

Neuroprotection by the Endogenous Cannabinoid Anandamide and Arvanil against *In Vivo* Excitotoxicity in the Rat: Role of Vanilloid Receptors and Lipoxygenases

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Type 1 vanilloid receptors (VR₁) have been identified recently in the brain, in which they serve as yet primarily undetermined purposes. The endocannabinoid anandamide (AEA) and some of its oxidative metabolites are ligands for VR₁, and AEA has been shown to afford protection against ouabain-induced *in vivo* excitotoxicity, in a manner that is only in part dependent on the type 1 cannabinoid (CB₁) receptor. In the present study, we assessed whether VR₁ is involved in neuroprotection by AEA and by arvanil, a hydrolysis-stable AEA analog that is a ligand for both VR₁ and CB₁. Furthermore, we assessed the putative involvement of lipoxygenase metabolites of AEA in conveying neuroprotection. Using HPLC and gas chromatography/mass spectroscopy, we demonstrated that rat brain and blood cells converted AEA into 12-hydroxy-*N*-arachidoylethanolamine (12-HAEA) and 15-hydroxy-*N*-arachidonoyl ethanolamine (15-HAEA) and that this conversion was blocked by addition of the lipoxygenase inhibitor nordihydroguaiaretic acid. Using magnetic resonance imaging we show the following: (1) pretreatment with the reduced 12-lipoxygenase metabolite of AEA, 12-HAEA, attenuated cytotoxic edema formation in a CB₁ receptor-independent manner in the acute phase after intracranial injection of the Na⁺/K⁺-ATPase inhibitor ouabain; (2) the reduced 15-lipoxygenase metabolite, 15-HAEA, enhanced the neuroprotective effect of AEA in the acute phase; (3) modulation of VR₁, as tested using arvanil, the VR₁ agonist capsaicin, and the antagonist capsazepine, leads to neuroprotective effects in this model, and arvanil is a potent neuroprotectant, acting at both CB₁ and VR₁; and (4) the *in vivo* neuroprotective effects of AEA are mediated by CB₁ but not by lipoxygenase metabolites or VR₁.

Key words: arvanil; anandamide; cannabinoid; vanilloid; CNS; excitotoxicity; ouabain; neuroprotection; neurodegeneration

Introduction

The eicosanoid anandamide [*N*-arachidonoyl ethanolamine (AEA)] mimics the pharmacological actions of Δ⁹-tetrahydrocannabinol (THC), the main psychoactive compound in marijuana, and was the first endocannabinoid to be discovered (Devane et al., 1992) (for review, see Di Marzo et al., 1998). THC, AEA, and the other endocannabinoid 2-arachidonoylglycerol (Mechoulam et al., 1995) exert neuroprotection in several models of neuronal injury (Nagayama et al., 1999; Panikashvili et al., 2001; van der Stelt et al., 2001a,b) (for review, see Mechoulam et

al., 2002), and mice with a defective type 1 cannabinoid (CB₁) receptor gene appear to be more susceptible to injury after stroke (Parmentier-Batteur et al., 2002). At variance with the protection observed with AEA in the late phase (7 d) after induction of excitotoxicity (van der Stelt et al., 2001b) and unlike the effect of THC (van der Stelt et al., 2001a), the protection afforded by the endocannabinoid in the early phase (15 min) was not sensitive to the CB₁ receptor antagonist SR141716A. This suggests that AEA or its metabolites may convey neuroprotection via other molecular targets in addition to the CB₁ receptor. Indeed, both AEA and 2-arachidonoylglycerol exert, *in vitro*, neuroprotective effects that are not always attenuated by a cannabinoid CB₁ antagonist (Sinor et al., 2000).

AEA is also a full agonist at the type 1 vanilloid receptor (VR₁) (Zygmunt et al., 1999; Smart et al., 2000), which is the target of capsaicin, the pungent principle in hot chili peppers. The VR₁ is a nonselective cation channel expressed in sensory fibers, in which it acts as a ligand-, proton-, and heat-activated integrator of nociceptive stimuli (Szallasi and Blumberg, 1999) and whose presence in the brain has been also established (Mezey et al., 2000; Sanchez et al., 2001; Szabo et al., 2002). Because, in the CNS, VR₁ is unlikely to be gated by noxious heat and low pH as in the peripheral nervous system, endogenous ligands for this receptor,

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such as AEA, *N*-arachidonoyldopamine (Huang et al., 2002), and lipoxygenase products of both arachidonic acid (AA) (Hwang et al., 2000) and AEA (Craib et al., 2001) might exist in the brain. These compounds, once produced under inflammatory conditions, might activate VR₁ with subsequent calcium influx (Szalasi and Blumberg, 1999), glutamate release (Marinelli et al., 2002), and substantial contribution to neuronal excitotoxicity. Conversely, exogenous compounds capable of quickly desensitizing VR₁, such as many VR₁ agonists, including AEA, might exert neuroprotective actions by preventing VR₁ activation by endogenous stimuli, and this mechanism might underlie the previously reported anticonvulsant effect of capsaicin (Dib and Falchi, 1996).

In the current study, we investigated the role of VR₁ and lipoxygenase products in the neuroprotection elicited *in vivo* by AEA. To this aim, we assessed the following: (1) the formation of lipoxygenase metabolites of AEA in rat blood and brain; (2) their effects in our *in vivo* model of neurodegeneration; (3) the effect of VR₁ stimulation using capsaicin and arvanil, a “hybrid” VR₁/CB₁ agonist; and (4) the effects of the VR₁ antagonist capsazepine and of the lipoxygenase inhibitor nordihydroguaiaretic acid (NDGA) on AEA-induced neuroprotection.

Materials and Methods

Brain tissue preparation. Male Wistar (200–300 gm) rats were killed by an overdose of pentobarbital and decapitated, after which their forebrains were rapidly isolated. Blood was collected in PBS containing Na₃-citrate (0.15 M). Some of the animals were transcardially perfused with PBS. Brains were minced with a razor blade and collected in ice-cold PBS (10 gm of tissue/100 ml of PBS).

Anandamide incubations. Brain tissue or blood cells were preincubated at 37°C for 10 min before a 20 min incubation with ionophore A23187 (10 μM), Ca²⁺ (1 mM), and 40 μM substrate. In the case of AEA, 100 μM PMSF was included in the incubation mixture. Reaction was terminated by addition of precooled chloroform/methanol (2:1) to extract lipids according to the Bligh and Dyer method (Bligh and Dyer, 1959).

HPLC analysis and product identification. Reversed-phase HPLC chromatograms were obtained using a Hewlett-Packard (Palo Alto, CA) HP1090 liquid chromatograph equipped with an HP1040A diode array detector with a detection limit <40 pmol (for conjugated dienes) and with an HP79994A analytical workstation. HPLC analysis was performed on a Cosmosil 5C18-AR (5 mm; 250 × 4 mm inner diameter; Nacal Tesque, Kyoto, Japan) column, using a methanol/water mixture (80:20) as the eluent at a flow rate of 1 ml/min. Compounds were collected and dried under N₂ flow and silylated with a *bis*(trimethylsilyl)fluoroacetamide for 15 min at room temperature. Gas liquid chromatography/mass spectroscopy (GC/MS) analysis was performed by injecting the samples on column (25 m HT-5 SGE column, 0.1 mm film thickness; SGE, Austin, TX). The column temperature was held at 70°C for 3 min and then allowed to rise 240°C at 40°C/min, followed by an increase of 16°C/min up to 330°C. Mass spectrometry was performed with a Fisons Instruments MD 800 mass detector. Mass spectra were recorded under electron impact with an ionization energy of 70 eV.

Animal model. Neonatal Wistar rats (U:Wu/Cpb; 7–8 d old) were anesthetized with ether and immobilized in a stereotaxic frame. A small burr hole was drilled in the cranium over the left hemisphere, 2.5 mm lateral of bregma. A 1 μl syringe was lowered into the left striatum to a depth of 4.0 mm. Ouabain (0.5 μl, 1 mM; Sigma-Aldrich, Zwijndrecht, The Netherlands) was injected at a rate of 0.125 μl/min using a micro-drive. After injection, the needle was left *in situ* for 2 min to avoid leakage of injection fluid from the needle tract. Body temperature was maintained at 37°C using a water-filled heating pad and an infrared heating lamp. Animals, needed for magnetic resonance imaging (MRI) study, were then positioned in the magnet, and anesthesia was continued using a mixture of halothane (0.4–1%) in N₂O–O₂.

Pharmacological treatments. Arvanil was synthesized as described previously (Melck et al., 1999). 12-Hydroxy-*N*-arachidoylethanolamine

(12-HAEA) and 15-hydroxy-*N*-arachidoylethanolamine (15-HAEA) were synthesized, purified, and characterized as described previously (van Zadelhoff et al., 1998). AEA was obtained from Biomol (Heerhugowaard, The Netherlands), SR141716A from Sanofi Recherche (Montpellier, France), capsaicin and capsazepine from Tocris (Köln, Germany), and NDGA from Sigma-Aldrich. Animals used in the MRI study were treated intraperitoneally with arvanil (0.1 or 1 mg/kg; *n* = 5 and *n* = 6, respectively), arvanil (1 mg/kg) plus SR141716A (1 and 3 mg/kg; *n* = 5 and *n* = 4, respectively), arvanil (1 mg/kg) plus capsazepine (5 and 10 mg/kg; *n* = 4 and 10, respectively), capsaicin (1 mg/kg; *n* = 5), AEA (1 mg/kg) plus 15-HAEA (7 mg/kg; *n* = 7), NDGA (10 mg/kg; *n* = 6) 5 min before AEA (10 mg/kg), NDGA (10 mg/kg; *n* = 6), AEA (10 mg/kg) plus capsazepine (10 mg/kg; *n* = 5), capsazepine (10 mg/kg; *n* = 5), 12-HAEA (3 mg/kg; *n* = 6), or 12-HAEA (3 mg/kg) plus SR141716A (3 mg/kg; *n* = 5). Control animals received vehicle alone (*n* = 16). 15-HAEA was tested at a dose likely to produce a molar ratio with AEA (7:1) similar to the one previously found *in vitro* to inhibit AEA degradation (van der Stelt et al., 2002b). Because 12-HAEA and 15-HAEA are only very weak agonists at VR₁ and 15-HAEA is only a weak ligand at CB₁, we did not test the effect of capsazepine on the two compounds, nor of SR141716A on 15-HAEA or of 15-HAEA alone (Hwang et al., 2000; van der Stelt et al., 2002b). Because AEA is a very weak functional agonist at CB₂ receptors (Bayewitch et al., 1995; Gonsiorek et al., 2000) and arvanil has little if any affinity for these receptors (Melck et al., 1999; Di Marzo et al., 2002), no experiment with CB₂ antagonists were performed. All drugs were dissolved in 1 ml/kg body weight 18:1:1 v/v PBS/Tween 80/ethanol, 30 min before toxin injection. There was no difference in body weight and growth rate between any of the groups. The Utrecht University Animal Experiment Ethical Committee approved all protocols.

MRI experiments. MRI was performed on a 4.7 T Varian horizontal bore spectrometer. Resonance frequency excitation and signal detection were accomplished by means of a Helmholtz volume coil (9 cm diameter) and an inductively coupled surface coil (2 cm diameter), respectively. A single-scan diffusion-trace MRI-sequence [four *b* values, 100–1300 sec/mm²; repetition time (TR), 3 sec; echo time (TE), 100 msec] was used to generate quantified images of tissue water trace apparent diffusion coefficient. Diffusion-trace- and T₂-weighted imaging (TE of 18, 40, 62, and 84 msec; TR of 2 sec; number of transients, 2) were performed in all animals (2.2 × 2.2 cm field of view; 64 × 64 data matrix), starting at *t* = 15 min after injection on day 0 and were repeated 1 week later. As expected, at this early time point, no changes in T₂-weighted MRI were detected. Both the T₂-weighted and the diffusion-weighted (DW) datasets consisted of seven consecutive 1.5-mm-thick slices, with 0 mm slice gap. To minimize interference at the slice boundaries, slices were acquired in alternating order (1, 3, 5, 7, 2, 4, 6), thus maximizing the time between excitation of two neighboring slices. For the diffusion-weighted imaging, we used a double spin-echo pulse sequence with four pairs of bipolar gradients with specific predetermined signs in each of the three orthogonal directions as published recently (De Graaf et al., 2001). The combination of gradient directions leads to cancellation of all off-diagonal tensor elements, thus effectively measuring the trace of the diffusion tensor. This provides unambiguous and rotationally invariant apparent diffusion coefficient (ADC) values in one experiment, circumventing the need for three separate experiments. For each *b* value, two scans were averaged. The total scan time for acquisition of seven slices, with four *b* values and two averages, was 17 min.

Data analysis. ADC and T₂ maps were generated by monoexponential fitting using the Interactive Data Language software package. Parametric images were analyzed in anatomic regions of interest using the Interactive Data Language software package. Pixels in the ipsilateral hemisphere were considered pathological if their ADC or T₂ value differed more than twice the SD of the mean value in the contralateral hemisphere. The ventricles were segmented out in the average ADC and T₂ measurements. The lesion volume per slice was calculated by multiplying the lesion area (number of pathological pixels × field-of-view in cm²/number of points acquired per image) by the slice thickness. The total lesion volume was obtained by summation of the lesion volumes for all slices. The absence of a slice gap makes interpolation of lesion areas between slices unneces-

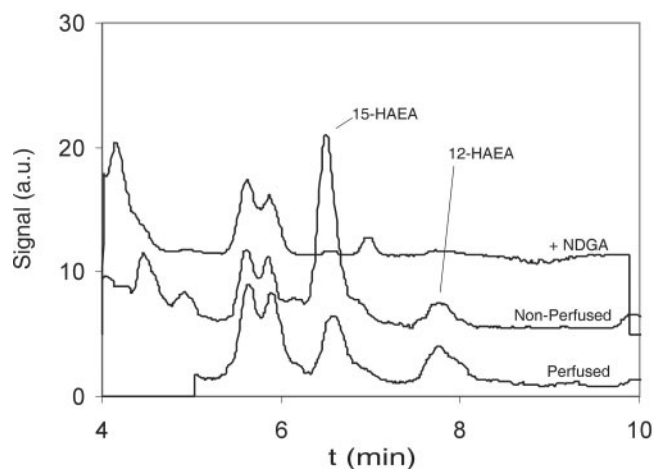


Figure 1. Reversed-phase HPLC profiles ($\lambda = 236$ nm) of lipid extracts of anandamide incubations with rat brain homogenates using a C18-column with methanol/water (80:20 v/v) as eluent. Top trace, Anandamide incubation with rat brain and NDGA, a nonselective lipoxygenase inhibitor. Middle trace, Anandamide incubation with nonperfused rat brain. Bottom trace, Anandamide incubation with PBS perfused rat brain. a.u., Arbitrary units.

sary, reducing systematic errors to within-slice “averaging” of signal intensity. Statistical analysis was performed using SPSS 10.0 (SPSS, Chicago, IL). Differences between groups were analyzed using Student’s *t* test; reported *p* values correspond to two-tailed significance.

Results

Lipoxygenase metabolism of anandamide

Reversed-phase HPLC analyses of the lipid extracts of AEA incubations with rat brain homogenates were performed to determine whether AEA metabolites were formed. The reversed-phase HPLC analyses revealed that several products eluted with an absorption at $\lambda = 236$ nm, which were absent in control incubations without AEA. These peaks were not present when NDGA, a nonselective lipoxygenase inhibitor, was included in the incubation (Fig. 1). Two peaks eluted at the same retention times as the reduced lipoxygenase products of AEA, 15-HAEA (TR of 6.4 min) and 12-HAEA (TR of 7.9 min), respectively. The UV spectra of these materials demonstrated an absorption maximum at 236 nm and were similar to the spectra of standards of 12-HAEA and 15-HAEA. The peaks were collected, derivatized, and subjected to GC/MS analysis. The mass spectra confirmed that the peaks eluting at 6.4 and 7.9 min represented 15-HAEA and 12-HAEA, respectively. AEA incubations with brains from rats that were transcardially perfused with PBS before dissection demonstrated a similar elution profile, although the ratio of 15-HAEA to 12-HAEA was changed (Fig. 1). This suggested that at least a part of the oxygenated metabolites, especially 15-HAEA, were produced by lipoxygenases expressed in blood cells. AEA incubations with rat blood cells demonstrated that 15-HAEA and 12-HAEA could be formed (data not shown). The cellular origin of the lipoxygenase activity was not investigated.

Role of lipoxygenase metabolites in neuroprotection after anandamide treatment

We wanted to investigate whether the lipoxygenase metabolites of AEA detected here might mediate the non-CB₁-mediated acute neuroprotective effect of AEA in our rat model of *in vivo* excitotoxicity (van der Stelt et al., 2001b; Veldhuis et al., 2003). In these experiments, we used reduced lipoxygenase products of AEA because the hydroperoxy derivatives will be rapidly degraded *in vivo*. In our model, excitotoxicity is induced by phar-

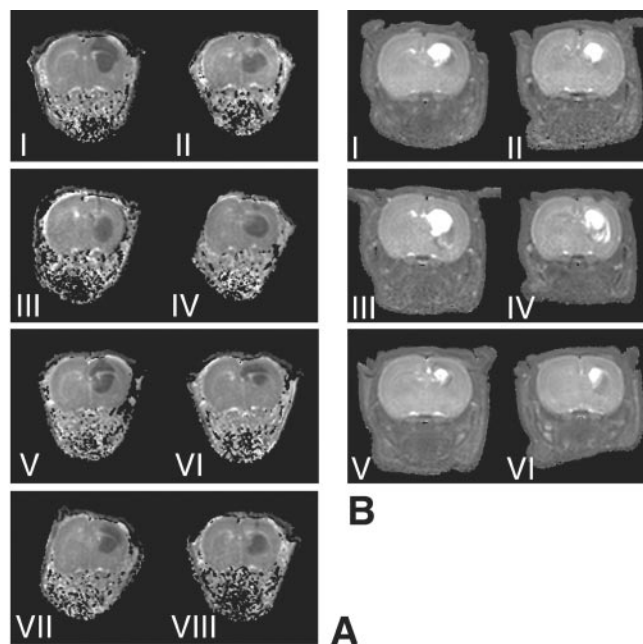


Figure 2. *A*, Typical parametric ADC maps of rat brain, calculated from diffusion-weighted MRI data acquired 15 min after ouabain injection. *B*, Parametric T_2 maps of the corresponding slices of the same animals, calculated from T_2 -weighted MRI data acquired 1 week later. The treatments are as follows: *I*, vehicle; *II*, AEA (1 mg/kg) plus 15-HAEA; *III*, 12-HAEA; *IV*, 12-HAEA plus SR141716A; *V*, capsazepine; *VI*, AEA (10 mg/kg) plus capsazepine; *VII*, NDGA; and *VIII*, AEA (10 mg/kg) plus NDGA.

macologically inhibiting the Na⁺/K⁺-ATPase using ouabain. The resulting acute cellular swelling (i.e., cytotoxic edema) is conveniently monitored using diffusion-weighted MRI. ADC maps of tissue water, calculated from DW-MRI datasets, showed hypointense regions with reduced ADC values ($0.64 \pm 0.006 \times 10^{-3}$ mm²/sec) in the ipsilateral hemisphere of all animals. In the contralateral hemisphere, normal ADC values ($1.13 \pm 0.01 \times 10^{-3}$ mm²/sec) were measured (Fig. 2*A*). We showed previously that administration of AEA itself dose-dependently reduces lesion volume in this model (van der Stelt et al., 2001b). The reduced 12-lipoxygenase metabolite of AEA, 12-HAEA, attenuated the volume of cytotoxic edematous tissue in the acute phase after injection of ouabain ($p < 0.05$) (Figs. 2*A*, 3). The CB₁ antagonist SR141716A was unable to block this effect, indicating that the CB₁ receptor was not involved in the protection afforded by 12-HAEA (Figs. 2*A*, 3). The reduced 15-lipoxygenase metabolite of AEA, 15-HAEA [which was shown previously to be only a weak ligand at CB₁ but to inhibit the enzymatic hydrolysis of AEA (van der Stelt et al., 2002b)], enhanced the protection afforded by a low dose (1 mg/kg) of AEA [from 37.2 mm³ (van der Stelt et al., 2001b) to 26.6 mm³ ($p < 0.05$); (Figs. 2*A*, 3)]. T_2 maps calculated from T_2 -weighted images acquired 7 d later showed normal T_2 values (71.9 ± 0.04 msec) in the contralateral hemisphere of all animals. The ouabain-injected hemisphere showed hypointensities and hyperintensities (Fig. 2*B*). Both types of T_2 abnormalities indicate pathological change. Hyperintensities correlate to regions of vasogenic edema and tissue loss, whereas hypointensities correspond to regions displaying reactive gliosis (Veldhuis et al., 2003). Calculation of total lesion volumes based on both hyperintensities and hypointensities indicated that neither the protection afforded by 12-HAEA nor the effect of 15-HAEA lasted until day 7 (Figs. 2*B*, 3). The dose of 12-HAEA tested significantly reduced lesion volume in the acute phase,

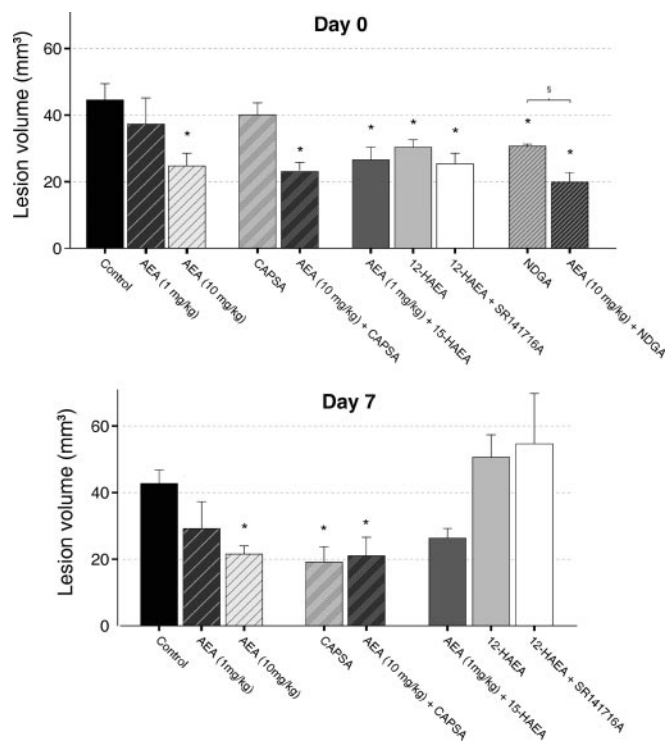


Figure 3. Lesion volumes as determined from ADC maps on day 0 and T₂ maps on day 7. The effects of several combinations of AEA, AEA metabolites, CB₁ and VR₁ antagonists, and a lipoxygenase inhibitor on lesion volume are shown. For comparison, the effect of AEA alone is also shown. This effect was determined in a previous study (van der Stelt et al., 2001b). Error bars represent means \pm SE. * p < 0.05 versus control; S_p < 0.05 for AEA plus NDGA versus AEA alone. See legend to Figure 2 for all of the doses. CAPSA, Capsazepine (10 mg/kg).

obviating the need to use a higher dose at that time point. In the late phase, higher doses of this compound were not investigated because the neuroprotective effect of AEA in this phase is blocked by SR141716A.

These data might suggest that the SR141716A-insensitive, early neuroprotective effect observed previously with AEA in the same *in vivo* model (van der Stelt et al., 2001b) may have been attributable to the formation of 12-HAEA and 15-HAEA. To test this hypothesis, the nonselective lipoxygenase inhibitor NDGA was injected 5 min before AEA. NDGA did not reverse the AEA-induced reduction in lesion volume (Figs. 2A, 3). In contrast, pretreatment of NDGA enhanced the neuroprotective effect of AEA (p < 0.05) (Figs. 2A, 3), resulting in a 51% smaller lesion volume compared with control animals (p < 0.05) (Figs. 2A, 3).

Effects of VR₁ modulation on ouabain-induced neurodegeneration

To investigate the possible involvement of VR₁ in AEA-induced neuroprotection, we performed three types of experiments. First, we tested the neuroprotective effect of arvanil, a synthetic AEA analog, which is metabolically more stable than AEA and activates both CB₁ and, more potently, VR₁ receptors but not CB₂ receptors (Melck et al., 1999; Di Marzo et al., 2001b). Next, we tested the effect of capsaicin, a selective VR₁ agonist. And finally, we tested the effect of the VR₁ antagonist capsazepine on AEA-induced neuroprotection.

Unlike THC, AEA, and 12-HAEA, arvanil (1 mg/kg) did not reduce lesion size on DW-MRI in the acute phase after injection of ouabain (38 ± 4.2 mm³; $p \gg 0.05$ vs control) (data not shown), although it was dose-dependently effective at 7 d (Figs. 4,

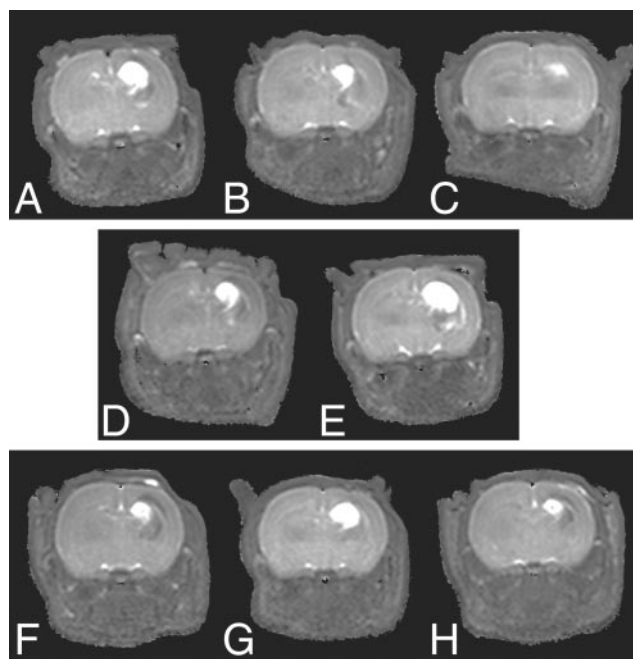


Figure 4. Typical parametric T₂ maps calculated from T₂-weighted MRI data acquired 7 d after injection of ouabain. The treatments are as follows: A, vehicle; B, arvanil (0.1 mg/kg); C, arvanil (1 mg/kg); D, arvanil (1 mg/kg) plus SR141716A (1 mg/kg); E, arvanil (1 mg/kg) plus SR141716A (3 mg/kg); F, arvanil (1 mg/kg) plus capsazepine (5 mg/kg); G, arvanil (1 mg/kg) plus capsazepine (10 mg/kg); and H, capsaicin (1 mg/kg).

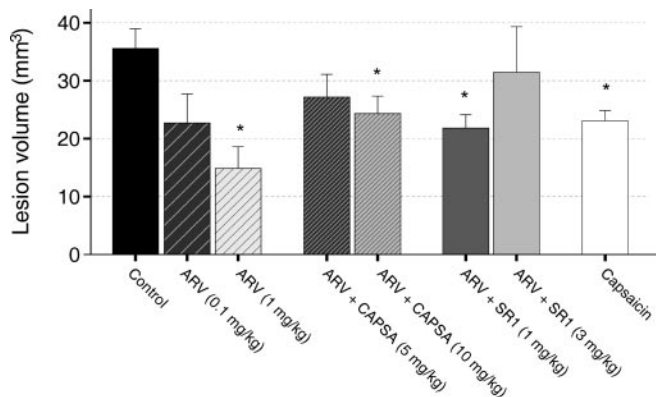


Figure 5. Lesion volumes as determined from T₂ maps acquired 7 d after ouabain injection. Error bars represent means \pm SE. * p < 0.05 versus control. See legend to Figure 4 for all of the doses. ARV, Arvanil; CAPSA, capsazepine; SR1, SR141716A.

5). This effect was mimicked by capsaicin at 1 mg/kg. Noteworthy, arvanil was more potent than AEA, capsaicin, and THC after 7 d. The 1 mg/kg dose was the highest tolerated dose of capsaicin for neonatal animals. With arvanil, previous data in adult rats (Di Marzo et al., 2001a) and mice (Di Marzo et al., 2000), obtained after intraperitoneal or intravenous administrations of 10 mg/kg of the substance, respectively, showed that arvanil is much better tolerated and less toxic than capsaicin (whose highest tolerated dose was 1 mg/kg in both cases).

To investigate which receptor between CB₁ and VR₁ mediates arvanil-induced neuroprotection in the late phase, the CB₁ receptor antagonist SR141716A or the VR₁ antagonist capsazepine were coinjected with arvanil (1 mg/kg) into neonatal rats. SR141716A dose-dependently reduced the neuroprotective effect of arvanil, suggesting the involvement of CB₁ receptors. Interest-

ingly, capsazepine was also able to reduce the effect of arvanil, indicating the possible involvement of VR₁ (Figs. 4, 5). Capsazepine at 10 mg/kg did not completely block the effect of arvanil, but higher doses could not be tested because of the limited solubility of capsazepine in the vehicle solution. However, this dose of the antagonist was shown previously to be sufficient to entirely block capsaicin-induced (1 mg/kg) hypolocomotor effects in adult rats but has no effect on movement impairment induced by arvanil (1 mg/kg) (Di Marzo et al., 2001a). The finding that arvanil and capsaicin were able to attenuate ouabain-induced neurodegeneration in neonatal rats in a capsazepine-dependent manner suggests that VR₁ stimulation can afford neuroprotection in this *in vivo* model. To test whether VR₁ could also be involved in the neuroprotective effects of AEA, capsazepine (10 mg/kg) was coinjected with AEA (10 mg/kg). However, capsazepine was unable to block the neuroprotective effect of AEA in both the acute and the late phase (Figs. 2, 3), arguing against a VR₁-mediated mechanism for AEA. Importantly, VR₁ antagonism by capsazepine (10 mg/kg) alone also reduced brain injury at 7 d (Figs. 2, 3).

Discussion

The endocannabinoid system is likely to be exploited for the development of therapeutics against acute neurodegeneration (Nagayama et al., 1999; Panikashvili et al., 2001; van der Stelt et al., 2001b) (for review, see van der Stelt et al., 2002a). AEA affords neuroprotection, both *in vitro* and *in vivo*, which is only in part mediated by the CB₁ receptor (Sinor et al., 2000; van der Stelt et al., 2001b). This suggests that other targets of AEA or its metabolites may contribute to the observed effects *in vivo*. In this study, we assessed a possible role for VR₁ and/or lipoxygenase metabolism in the neuroprotection induced by AEA against ouabain-induced *in vivo* excitotoxicity. The main findings are as follows: (1) the 12-lipoxygenase metabolite of AEA, 12-HAEA, attenuated cytotoxic edema in a CB₁ receptor-independent manner in the acute phase after ouabain injection; (2) the 15-lipoxygenase metabolite, 15-HAEA, enhanced the neuroprotective effect of AEA in the acute phase; (3) modulation of VR₁ leads to neuroprotective effects in our model, and arvanil is a potent neuroprotectant, acting at both cannabinoid and vanilloid receptors; and (4) the acute *in vivo* neuroprotective effects of AEA are not mediated by either lipoxygenase metabolism or VR₁.

Neuroprotection by lipoxygenase products of anandamide

We showed that rat blood and perfused (blood-free) rat brain convert exogenous AEA into 12-HAEA and 15-HAEA in an NDGA-sensitive manner. Our findings are in line with previous studies using rat brain (Miyamoto et al., 1987) or human blood (Edgemond et al., 1998). Endogenous lipoxygenase metabolites of AEA have never been detected, but recent data suggest that they may mediate some of the actions ascribed to AEA (Kagaya et al., 2002). For example, the VR₁-mediated contractile action of AEA in guinea pig bronchus may be attributable, at least in part, to some of its oxygenated metabolites (Craib et al., 2001). Here, we showed that 12-HAEA was able to mimic the AEA-induced reduction of cytotoxic edema in the acute phase after ouabain injection, in a CB₁-independent manner. The 12-HAEA-induced reduction in cellular swelling is likely to occur also independently of the activation of VR₁, because 12-HAEA, unlike its hydroperoxy homolog, is not an agonist at VR₁ (Hwang et al., 2000). Furthermore, VR₁-stimulation by arvanil did not mimic the reduction of cellular swelling in the early phase (see below). 15-HAEA was able to enhance the neuroprotective effect of AEA. Because 15-HAEA has been shown to competitively inhibit the

hydrolysis of AEA by FAAH (van der Stelt et al., 2002b), an “entourage” effect might be responsible for the neuroprotection afforded by this compound (for a definition of “entourage” effect, see Mechoulam et al., 1998). However, neither 12-HAEA nor 15-HAEA afforded long-lasting protection in our model, although, in contrast to the effect of 12-HAEA, the effect of 15-HAEA almost reached significance also on day 7 ($p = 0.07$) (Figs. 2, 3).

At the moment, it is not clear which molecular targets are responsible for the observed effects of 12-HAEA in the acute phase. However, pretreatment with the nonselective lipoxygenase inhibitor NDGA did not reverse the AEA-induced reduction in cytotoxic edema (Fig. 2), and, in contrast, it enhanced the effect of AEA ($p < 0.05$). These results indicate that *in vivo* metabolism catalyzed by lipoxygenases, either directly on AEA or on AA produced from its hydrolysis (Willoughby et al., 1997; Adams et al., 1998), is unlikely to contribute to the AEA-induced reduction of cytotoxic edema. Indeed, application of NDGA alone also reduced the volume of cytotoxic edema by 25% ($p < 0.05$) (Fig. 2). This may indicate that endogenous lipoxygenase activity contributes to the neurotoxic events, although an alternative explanation might be that NDGA exerts its protective actions via non-specific antioxidant activity (Shishido et al., 2001). Altogether, these data indicate that endogenous lipoxygenase activity does not contribute to the neuroprotective actions of AEA.

Role of VR₁ modulation in neuroprotection by AEA and arvanil

VR₁ is a ligand-gated nonselective cation channel, activation of which results in nonselective cation influx, Ca²⁺ influx, membrane depolarization, and glutamate release (Szallasi and Blumberg, 1999). During acute neurodegenerative insults, such as ischemia or trauma, these mechanisms are likely to be detrimental. In fact, capsaicin treatment is sometimes used to degenerate peripheral nerves. However, VR₁ is easily desensitized by its agonists, and desensitization might lead to neuroprotection if VR₁ contributes in any way to the neuronal injury during excitotoxicity. In fact, capsaicin, the prototypic VR₁ agonist, has been shown previously to inhibit Tween 80-induced convulsions *in vivo* (Dib and Falchi, 1996). We report here for the first time a dose-dependent protection afforded by a hybrid CB₁/VR₁ ligand, arvanil, and by capsaicin. The neuroprotective effect of arvanil was mediated by activation of both CB₁ and VR₁ receptors and was more potent than that of capsaicin, AEA, and THC. The higher efficacy of arvanil may be attributable to a synergistic action on two signaling pathways, i.e., (1) stimulation of the CB₁ receptor and (2) rapid desensitization of VR₁, which might lead to a reduction in calcium influx and glutamate release. The latter, tentative mechanism is supported by the following data. (1) VR₁ is present in the CNS (Mezey et al., 2000; Sanchez et al., 2001; Szabo et al., 2002), including in the striatum (Hajos et al., 1988). (2) In the PNS, desensitization is a standard response of VR₁ to stimulation by exogenously administered agonists (Szallasi and Blumberg, 1999; Sanchez et al., 2001). Desensitization is known to take place on time scales relevant to the current experiments [e.g., within 30 min in sensory nerve terminals of guinea pigs (Maggi, 1995) or in the rat heart (McLean et al., 2002)]. (3) Although little is known about the pharmacology of VR₁ in the brain, preliminary observations of repeated capsaicin injections showed that capsaicin-induced stimulation of CNS neurons could be blocked by capsaicin pretreatment, suggesting that desensitization may also occur centrally (Hajos et al., 1987). (4) Desensitization by agonists is prevented by the simultaneous presence of an antagonist (Jung et al., 1998, 1999; Liu et al., 2001),

a phenomenon also observed in our study after cotreatment of arvanil with capsazepine. (5) In the current study, blocking VR₁ using capsazepine also afforded protection. Thus, we propose that both CB₁ activation and VR₁ desensitization may have contributed to the effect of arvanil. Interestingly, similar results have been reported previously for the cytostatic treatment of human breast cancer cells. In those experiments, the effect of AEA was mediated via CB₁ receptors, whereas arvanil exerted a more potent effect that was antagonized by both SR141716A and capsazepine (Melck et al., 1999).

VR₁ activation is unlikely to be deleterious to the CNS in general, but we argue that this might be the case in a setting of CNS injury. For example, in ischemia, the inflammatory mediator bradykinin and the prototypic proinflammatory cytokine tumor necrosis factor- α may sensitize VR₁ (Nicol et al., 1997; Shin et al., 2002; Sugiura et al., 2002). Sensitization of VR₁ by ligand binding, mildly acidic pH, or inflammatory mediators may result in activation of the receptor by temperatures well within the physiologic range (Tominaga et al., 1998; Vyklicky et al., 1998). Additionally, VR₁ stimulation results in release of substance P, which is a proinflammatory neuropeptide (Martin et al., 1992; Annunziata et al., 2002). If these conditions occur during ouabain-induced excitotoxicity, VR₁ desensitization by capsaicin and arvanil or VR₁ antagonism by capsazepine may explain the neuroprotective effects of these substances.

However, we cannot exclude the possibility that other mechanisms may have contributed to the protection afforded by arvanil, capsaicin, or capsazepine. For example, both capsaicin and its dihydroderivate nordihydrocapsiate have similar, potent anti-inflammatory properties *in vivo* via nuclear factor- κ B inhibition, possibly via a non-VR₁-mediated mechanism (Sancho et al., 2002). Furthermore, capsaicin has been shown to stimulate the biosynthesis of endocannabinoids (Di Marzo et al., 2001a), which may afford neuroprotection. Alternatively, the presence in the CNS of as yet undefined CB_n receptors, sensitive to capsaicin, capsazepine, SR141716A, arvanil, and AEA has been suggested (Brooks et al., 2002; Hajos and Freund, 2002), which might play a role in arvanil-mediated neuroprotection.

In our model, we observed non-CB₁-dependent effects of AEA directly after ouabain injection and CB₁-dependent effects after 7 d. In contrast, the neuroprotection by arvanil was evident at 7 d and was not manifested by a reduction in cytotoxic edema in the acute phase. These observations, as well as the finding that the action of AEA was not counteracted by capsazepine, used at a dose that was effective against a much more potent VR₁ agonist (arvanil), strongly suggest that the neuroprotective effects of the endocannabinoid are not attributable to VR₁ stimulation. Therefore, it can be concluded that, although the late effect of AEA is entirely mediated by CB₁ receptors, the early action of this compound is attributable to a mechanism yet to be identified.

Conclusions

AEA protects rat brain from ouabain-induced excitotoxicity. This effect is inhibited by CB₁ but not VR₁ antagonists and is not mediated by lipoxygenase metabolites. Arvanil is more potent than AEA, and both CB₁ and VR₁ are involved in neuroprotection after arvanil treatment. We suggest that both VR₁ and CB₁ receptors might be valuable targets for therapy and drug discovery in protection against brain injury and that arvanil represents a promising subject for future studies aiming at developing a monotherapy that targets both receptors simultaneously.

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