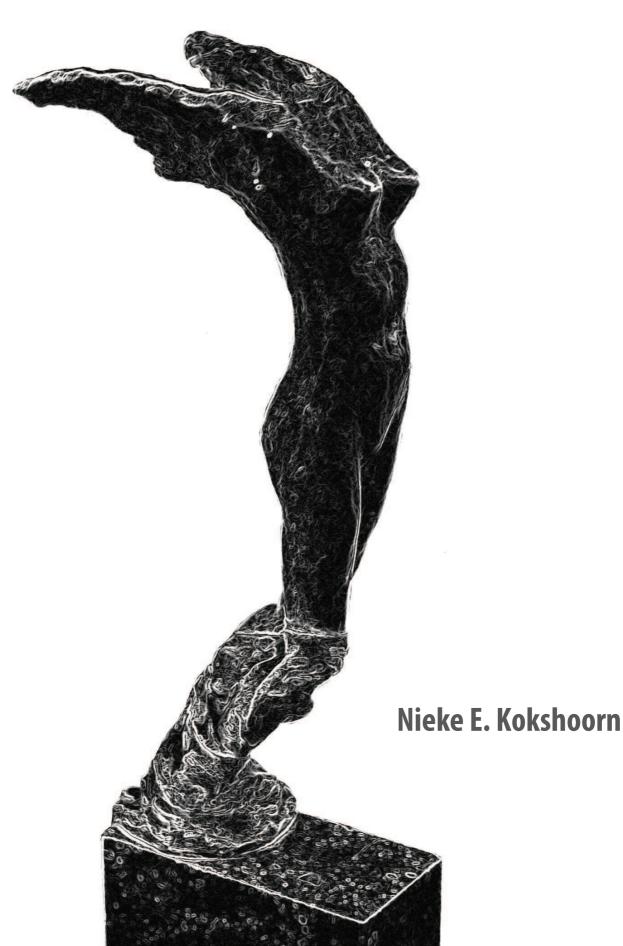
Hypopituitarism Clinical assessment in different conditions



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Voor Papa en Mama, Aan mijn lieve Oma

Hypopituitarism Clinical assessment in different conditions

Proefschrift

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Contents

Chapter 1	General introduction and outline of this thesis	9
Chapter 2	Clinical review: Hypopituitarism following trau- matic brain injury – the prevalence is affected by the use of different dynamic tests and different nor- mal values <i>Eur J Endocrinol. 2010 Jan;162(1): 11–18</i>	37
Chapter 3	Low prevalence of hypopituitarism after traumatic brain injury – a multi-center study <i>Eur J Endocrinol. 2011 Aug;165(2):225–31.</i>	75
Chapter 4	The use of an early postoperative CRH test to assess adrenal function after transsphenoidal surgery for pituitary adenomas <i>Pituitary . 2011 Sep 9. [Epub ahead of print]</i>	95
Chapter 5	Low incidence of adrenal insufficiency after transsphenoidal surgery in patients with acromegaly: a long-term follow-up study <i>J Clin Endocrinol Metab. 2011 Jul;96(7):E1163–70.</i>	119
Chapter 6	Pituitary dysfunction in adult patients after cranial radiotherapy: a systematic review and meta-analysis <i>J Clin Endocrinol Metab. 2011 Aug;96(8):2330–40</i>	141
Chapter 7	Growth hormone replacement therapy in elderly growth hormone deficient patients: a systematic review <i>Eur J Endocrinol. 2011 May;164(5):657–65.</i>	171

Contents

Chapter 8	General discussion and summary	197
Chapter 9	Nederlandse samenvatting	219
Curriculum Vitae		235
List of publications		239
Notes		243

Chapter 1

General introduction and outline of this thesis

Contents

I.	General introduction		11
II.	The p	ituitary gland	12
	Anatomy and Physiology		
	1.	Regulation and secretion of growth hormone and IGF-I	13
	2.	Regulation of ACTH and cortisol secretion	16
	3.	Regulation and secretion of thyroid hormone,	
		gonadotropins and prolactin	17
III. Pituitary insufficiency		18	
	Growth hormone deficiency		19
	Corticotropin deficiency		
	Thyrotropin deficiency		23
	Gonadotropin deficiency		23
	Prola	ctin deficiency	24
IV.	Outli	ne of this thesis	25
	The evaluation of pituitary function in patients after traumatic		
	brain	injury	25
	Dyna	mic tests of pituitary function in other pituitary diseases	26
	Treat	ment of GH deficiency	27

I. General introduction

The pituitary gland is the master regulator of the endocrine system. Different pathophysiological conditions can affect the function of the pituitary gland and, consequently, endocrine homeostasis. The evaluation of pituitary function is therefore complex and the different tools that have become available to evaluate pituitary function only provide limited information of different aspects of hormone secretion. In this thesis, several difficulties encountered in establishing a diagnosis of pituitary insufficiency are studied in different pathophysiological conditions.

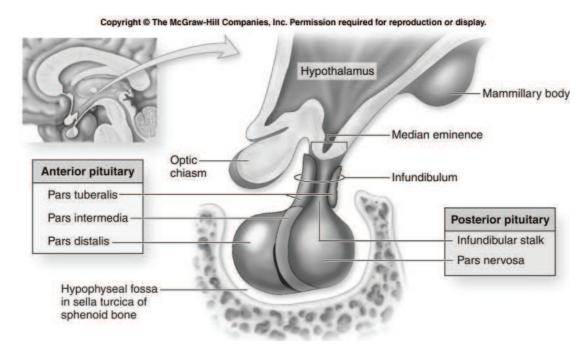


Figure 1. The pituitary gland

II. The pituitary gland

Anatomy and Physiology

The pituitary gland is a small gland located at the base of the skull in a socket of sphenoid bone, called the sella turcica. The gland consists of two lobes, the anterior lobe (or adenohypophysis; 80%), and a posterior lobe (neurohypophysis; 20%) (Figure 1).

Together with the hypothalamus, the pituitary controls the function of different endocrine glands (*i.e.* thyroid, adrenal and reproductive glands) (1). The hypothalamus receives signals from upper corticol inputs and the environment (such as light and temperature) and, in turn, delivers signals to the pituitary gland (*i.e.* regulating the endocrine system). Hormones released by the pituitary gland influence the endocrine systems in the body and also have a feedback on the hypothalamus.

The communication between the hypothalamus and anterior pituitary is via the portal system that runs through the pituitary stalk. Hormones released by the hypothalamus are delivered to the anterior pituitary through these vessels and reach the anterior lobe through a dense capillary network. The communication with the posterior gland is via axons and nerve terminals of larger neurons that originate from within the hypothalamus. The hormones produced in these neurons, arginine-vasopressin and oxytocin, are released directly from the posterior pituitary into the systemic circulation.

The anterior pituitary is controlled by specific hypothalamic hormones: thyrotropin releasing hormone (TRH), gonadotropin releasing hormone (GnRH), corticotropin releasing hormone (CRH), growth hormone releasing hormone (GHRH), and somatostatin, that bind specific transmembrane receptors expressed in different anterior pituitary cells. These anterior pituitary cells are classified by their specific secretory products: somatotrophs (GH-secreting cells, expressing the GHRH and somatostatin receptor; 50%), lactotrophs (PRL-secreting cells, expressing the prolactin receptor; 10–25%), corticotrophs (cells secreting ACTH, expressing the CRH receptor; 15–20%), thyrotrophs (cells secreting TSH, expressing the TRH receptor; 10%), and gonadotrophs (LH and FSH secreting cells, expressing the GnRH receptor; 10–15%) (2).

1. Regulation and secretion of growth hormone and IGF-I

The regulation of growth hormone (GH) secretion is complex and involves many stimulatory and inhibitory hypothalamic peptides. However, the two most important components are growth hormone-releasing hormone (GHRH), which stimulates the somatotrophic cells, and somatostatin (SST) which inhibits GH release (2). The secretion of GH is also affected by factors such as nutrition (increased in fasting, stimulated by high protein meals and inhibit by hyperglycemia and leptin), other hormones (stimulated by estrogens and inhibited by glucocorticoid excess), neuropeptides, neurotransmitters and opiates (2–4).

The secretion of GH is pulsatile with undetectable serum GH levels between the pulses. In normal subjects the 24-hour profile of plasma GH levels consists of stable low levels interrupted by bursts of secretion (Figure 2 and 3). The major determinant of GH secretion in humans is sleep. GH secretion is lower in elderly and obese subjects and there are sex-specific differences in GH pulse amplitude and mass (5;6). The ageassociated changes in the GH profile include a reduction in GH secretory burst frequency, the half life of endogenous GH and the daily secretory rate (7). In obese subjects decreased GH concentrations result from both diminished pulsatile GH secretion and accelerated metabolic clearance (4;8).

In the liver, GH stimulates the production of insulin-like growth factor (IGF-I). The primary function of GH is promotion of linear growth in children by acting directly and indirectly (via the synthesis of IGF-I which mediates most of the peripheral actions of GH) on the epiphyseal plates of long bones. Whereas GH and IGF-I have synergistic effects on linear and organ growth by their control of mitogenesis and apoptosis and on glomerular filtration rate, their metabolic actions are opposing: GH stimulates lipolysis and reduces insulin sensitivity, IGF-I is anti-lipolytic and ameliorates insulin sensitivity (9;10).

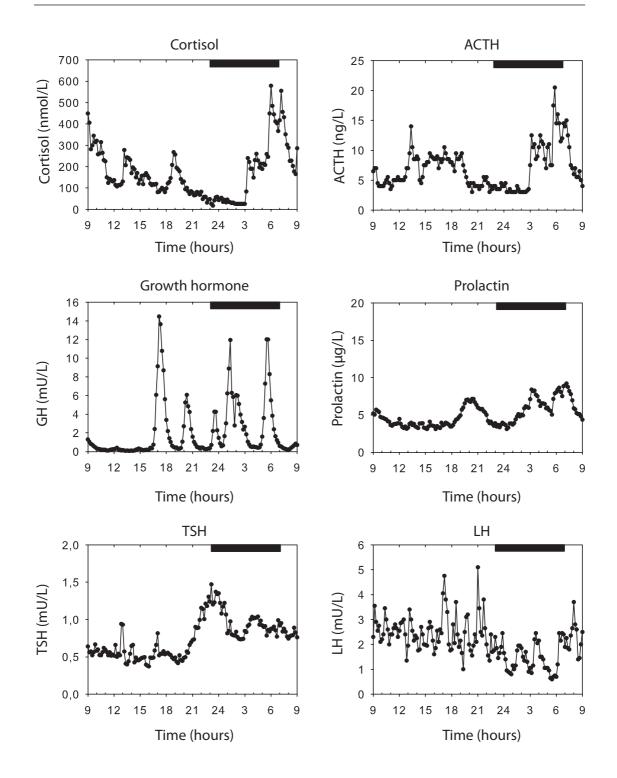


Figure 2. Plasma hormone concentration profiles of a 33 year-old healthy female volunteer. Blood samples were taken at 10 min intervals during 24 hours. The black bar in the top of the panels indicate the period with lights off. Note the diurnal and the pulsatile characteristics of each hormone. (*The figure was provided by Dr. F. Roelfsema, Leiden University.*)

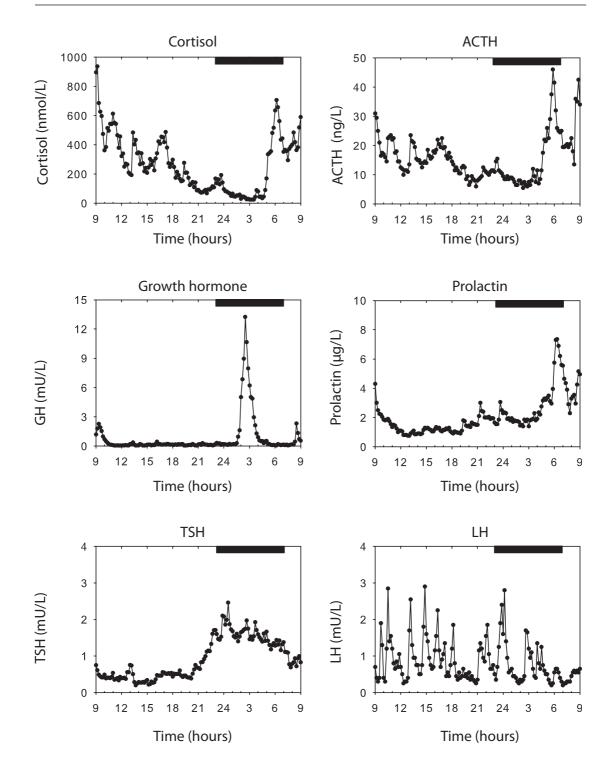


Figure 3. Plasma hormone concentration profiles of a 37 year-old healthy male volunteer. Blood samples were taken at 10 min intervals during 24 hours. The black bar in the top of the panels indicate the period with lights off. Note the diurnal and the pulsatile characteristics of each hormone. (*The figure was provided by Dr. F. Roelfsema, Leiden University.*)

2. Regulation of ACTH and cortisol secretion

The secretion of corticotropin (ACTH) and cortisol is regulated by hormonal interactions between the hypothalamus, pituitary, and adrenal glands. The secretion of hypothalamic corticotropin-releasing hormone (CRH) is regulated mainly by hippocampal neurons that express both receptors for cortisol, the mineralocorticoid- and glucocorticoid receptor. In addition the secretion is influenced by the circadian pacemaker and stress (2;11). CRH regulates the secretion of ACTH by the pituitary gland, which is potentiated by arginine-vasopressin. Subsequently ACTH binds to its receptor on the adrenal cortex to stimulate the secretion of cortisol and other steroids. The negative feedback loop is completed by the inhibitory effect of cortisol on CRH and ACTH synthesis and secretion (11).

Pulsatile secretion and circadian rhythm

ACTH is secreted in brief episodic bursts resulting in a diurnal rhythm of ACTH secretion with a concordant diurnal secretion of cortisol from the adrenal cortex (12;13). Plasma ACTH and serum cortisol concentrations are highest early in the morning at time of awakening. During the day plasma cortisol levels fall resulting in low levels in the late afternoon and evening with a nadir one or two hours after sleep onset (Figure 2 and 3) (11;14;15).

Stress-induced secretion

The HPA axis is activated both by physical and psychological stressors, resulting in increased plasma ACTH and cortisol concentrations. Physical stressors include severe trauma, like burns (16;17), or illnesses, major surgery (18;19), but also hypoglycemia (20;21), hypotension, exercise (22), and cold exposure (23).

Negative feedback inhibition by glucocorticoids

Both endogenous and exogenous glucocorticoids have a negative feedback on ACTH secretion which occurs at both the hypothalamic (CRH suppression) and pituitary (ACTH suppression) levels. This leads to atrophy of the adrenal glands resulting in loss of cortisol secretory capacity (24). The degree of probably depends upon the dose, potency and duration of action of the glucocorticoid, and the time of its administration (25–29). The shorter the interval between the administration of glucocorticoid and the normal early morning peak of ACTH secretion, the greater the suppressive effect of the glucocorticoid. The duration of suppression is increased by higher doses and longeracting glucocorticoids. After withdrawal of chronic administration of high doses of glucocorticoid, suppression of the hypothalamic-pituitaryadrenal axis may persist for weeks but may even persist for many years.

3. Regulation and secretion of thyroid hormone, gonadotropins and prolactin

The hypothalamus-pituitary thyroidal axis regulates the production of thyroid hormone by the thyroid gland. The hypothalamus produces thyroptropin releasing hormone (TRH) which stimulates the pituitary gland to secrete thyrotropin (TSH). TSH stimulates the synthesis of the thyroid hormones (thyroxine (T_4) and triiodothyronine (T_3)) by binding to the TSH receptors on the thyroid cells. The response of TSH to TRH is, in turn, modulated by the circulating concentrations of T_3 and T_4 . High serum levels of T_3 and T_4 inhibit and low levels stimulate TSH synthesis (2).

The reproductive axis is controlled by periodic pulsatile release of the hypothalamic gonadotropin-releasing hormone (GnRH). GnRH stimulates the pituitary to secrete the gonadotropins, luteinizing hormone (LH) and follicle-stimulating hormone (FSH). The production of steroid hormones (including estradiol, progesterone and testosterone), as well as other factors such as inhibin, activin and insulin-like growth factor-I (IGF-I) are induced by the gonadotropins. The circulating sex steroids have a positive as well as negative feedback on GnRH and thereby also influence LH and FSH concentrations.

The function of LH in men is to stimulate testosterone production from interstitial cells of the testes (Leydig cells) and FSH is required for spermatogenesis. In women, LH is critical for ovulation and maintenance of the corpus luteum, whereas FSH promotes follicular development (2).

The main physiological role of prolactin (PRL) is for nursing. The hypothalamic control of PRL secretion is predominantly inhibitory, and dopamine is the most important inhibitory factor. TRH is a potent prolactin-releasing factor (2).

III. Pituitary insufficiency

Hypopituitarism refers to decreased secretion of pituitary hormones, which can result from diseases of the pituitary gland and/or the hypothalamus, which cause diminished secretion of hypothalamic releasing hormones, thereby reducing secretion of the corresponding pituitary hormones. Pituitary insufficiency can be congenital (which will not be further addressed in this thesis) or acquired.

Pituitary adenomas and its treatment

A common cause of pituitary dysfunction is the presence of a pituitary adenoma. Macro-adenomas (> 10 mm) can be associated with pituitary insufficiency, with one or more anterior pituitary hormone deficiencies (30-32). In the presence of macroadenomas, hypopituitarism may result from compression of the rest of the pituitary and/or compression of the portal vessels in the pituitary stalk, secondary to either the expanding tumor mass or directly by increased intra-sellar pressure (30). Conversely, reduction of tumor mass by surgery and/or medication relieves the pressure and may restore pituitary function.

In pituitary surgery the surgeon attempts to preserve the adjacent normal pituitary tissue. However, if the surgeon is not able to visually distinguish the normal pituitary tissue from the adenoma, the normal tissue may be damaged, resulting in pituitary deficiency (33;34).

Radiation of pituitary adenomas, usually to prevent regrowth of residual tissue after surgery or to control excessive GH or ACTH secretion, also exposes the nonadenomatous pituitary and the hypothalamus to irradiation resulting in pituitary insufficiency (35;36). Not only patients with pituitary tumors but also patients treated with radiotherapy for suprasellar lesions, primary brain tumors, nasopharyngeal tumors, head and neck tumors, or hematological malignancies (*i.e.* acute lymphoblastic leukemia (ALL)) are at risk for developing pituitary hormone deficiencies if the hypothalamus and/or the pituitary have been exposed to radiation (37–39).

Traumatic brain injury

In recent years, several studies have reported a high prevalence of pituitary insufficiency ranging from 15–90% in patients who experienced traumatic brain injury (TBI) (40–51). Large neuropathological series demonstrate pituitary as well as hypothalamic lesions after TBI (52). Infarction is believed to be the cause of posttraumatic hypopituitarism, found at post mortem in 26% to 86% of patients who died after TBI. Possible mechanisms for post-traumatic infarction include compression of the pituitary gland caused by changes in intracranial pressure resulting from cerebral edema, hemorrhage or skull fracture, hypoxia, or direct damage to the gland itself (53;54).

The diagnosis of hypopituitarism, defined as deficient secretion of one or more pituitary hormones secondary to pituitary or hypothalamic disease, is made by documenting subnormal secretion of these pituitary hormones under defined (*i.e.* controlled) circumstances. Since there is a variable pattern of hormone deficiencies among patients with hypopituitarism, each pituitary hormone must be tested separately. For the evaluation of each axis basal serum hormones levels, but also dynamic testing is available.

Growth hormone deficiency

Clinical consequences

Growth hormone deficiency (GHD) in adults is characterized by increased body fat and decreased lean body mass, decreased bone mass and increased fracture rate, impaired cardiac function and reduced muscle strength (55–57). Adult patients with GHD also share a number of characteristics of the metabolic syndrome, including hypertension, abdominal obesity, insulin resistance and dyslipidemia (58). In addition, quality of life is impaired, with reduction in physical and mental energy, increased anxiety, and dissatisfaction with body image and poor memory (2;57;59;60). Replacement therapy with growth hormone (rhGH) was associated with apparent benefits, particularly in terms of body composition, bone mass, muscle strength, cardiac function and quality of life (61).

Diagnosis

Because of the pulsatile nature of GH secretion, basal serum GH levels are not useful to assess the GH-IGF-I axis, although basal serum IGF-I levels below the reference ranges are indicative for GHD in the presence of two or more other insufficiencies (62;63). Normal IGF-I concentrations, however, do not exclude the diagnosis of GHD, as IGF-I levels are within the normal reference range in about one third of patients with GHD, especially in elderly subjects (64–66). Therefore, the use of dynamic testing is mandatory for the evaluation of GH secretory reserve.

Different stimulation tests are available (*i.e.* insulin tolerance test (ITT), and stimulation tests with glucagon, GHRH, GHRH-arginine, or GHRH-GHRP6). However the preferred test for evaluation of this axis still remains the insulin tolerance test (63;67). With the administration of insulin a hypoglycemia is induced which is a very strong physiological stimulator of the stress response. Hypoglycemia activates the hypothalamus to secrete GHRH resulting in stimulation of GH by the somatotropic cells of the pituitary gland. A peak GH response below 3 µg/L indicates a severe GHD (63;67). During ITT simultaneous assessment of the hypothalamus-pituitary-adrenal (HPA) axis is possible. Important contra-indications to perform the ITT are coronary insufficiency and/or epilepsy. To assess the GH axis in these patients, alternative provocative tests for GH secretion must be used with adapted appropriate cut-offs. The combined administration of arginine and GHRH is the most frequently used alternative GH stimulation test (66;68;69). GHRH and arginine both have a stimulatory effect on the pituitary gland (70;71). When given simultaneously they enhance their effect resulting in a secretion of GH. A bolus dose of GHRH (1 µg/kg body weight) is given intravenously at baseline, immediately followed by an intravenous infusion of arginine (0.5 gr/kg body weight (to a maximum) of 30 gr)) for 30 minutes. Measurements of GH are done 30, 45, 60 and 90 minutes after infusion. Recently, cut-off values adjusted for both body mass index (BMI) and age have been published (72).

Pitfalls

Several factors play a role when testing the GH secretion reserve, such as age, gender, BMI, other hormones and insulin sensitivity. Obese subjects have a blunted GH response to any provocative stimulus (8;73;74). There is an estrogen-related difference in GH axis activity: GH secreted per burst

greater and 24-hour GH release pattern is less orderly in women than men (75). Finally, GH secretion decreases with increasing age. Therefore, these factors should be considered when defining the diagnostic cut-off points in the assessment of GHD (72;76).

Corticotropin deficiency

Clinical consequences

ACTH deficiency leads to adrenocortical insufficiency, characterized by decreased secretion of cortisol. Normal corticotroph function is mandatory for adequate increase of cortisol concentrations in case of stress. However, to maintain sufficient cortisol concentrations, normal basal secretion of ACTH is necessary.

Hypocortisolism can be secondary to either adrenal gland destruction (primary adrenal insufficiency, mostly auto-immune adrenalitis or tuberculous adrenalitis) or to ACTH deficiency (secondary or central adrenal insufficiency) (77).

Diagnosis

Similar to the secretion of GH, the secretion of ACTH is pulsatile with circadian variation resulting in a circadian rhythm of cortisol secretion. Therefore, it is necessary to evaluate basal serum cortisol secretion in the early morning, during fasting. When cortisol concentrations are lower than, or exceed, a certain threshold (< 100 nmol/L or > 500 nmol/L) the likelihood of the presence or absence of adrenal insufficiency is very high, or negligible, respectively. In these cases stimulatory tests are not necessary (78). In all other condition, a dynamic test is mandatory.

The initial and most convenient test to evaluate the function of the HPA axis is the plasma cortisol response to synthetic ACTH (Synacthen test) (79;80). The test is performed by administering a bolus of 1 or 250 µg of cosyntropin intramuscularly or intravenously with measurements of serum cortisol 30 and 60 minutes thereafter. A serum cortisol concentration of \geq 500–550 nmol/L is considered a normal response. However, this test does not discriminate between the different causes of adrenal insufficiency, and a normal test response does not exclude mild secondary forms of adrenal insufficiency (81–84).

Other tests, that directly evaluate pituitary reserve are also available (insulin induced hypoglycemia, metyrapone administration, or CRH stimulation). Also in the assessment of the HPA axis (similar to the GH-IGF-I axis) the ITT still remains the golden standard (85;86). Insulin induced hypoglycemia results in stress which actives the entire HPA axis providing proof for adequate hypothalamic (CRH) and pituitary (ACTH) function. In healthy subjects serum cortisol levels will increase above 550 nmol/L if adequate hypoglycemia is achieved (glucose 2.2 mmol/L or lower). Stimulation with metyrapone is an alternative test to assess the HPA axis. The adrenal enzyme 11-\u00df-hydroxylase (CYP11B1) that catalyzes the conversion of 11-deoxycortisol to cortisol, is inhibited by metyrapone, resulting in a reduction of cortisol secretion. Administration of metyrapone will thus result in activation of the HPA axis, an increase in ACTH secretion and consequently an increase in adrenal steroidogenesis up to 11-deoxycortisol. An 11-deoxycortisol concentration above 200 nmol/L in the presence of suppressed cortisol levels (below 100 nmol/L) is then indicative for central adrenal insufficiency (87–89). This test can be performed as a prolonged and short overnight version, depending on the number of dosages of metyrapone given. It appears, however, that the ACTH stimulus of a single dose of metyrapone is comparable to that of an insulin tolerance test (89).

Since the 1980's ovine CRH is used for the evaluation of the HPA axis, mainly to discriminate between pituitary or adrenal causes of Cushing's syndrome (90-94). However, in recent years the CRH test is more often used to assess secondary adrenal insufficiency (95;96). Administration of an intravenous bolus of ovine CRH results in pituitary ACTH secretion resulting in cortisol secretion by the adrenal glands. In healthy subjects a 1 μ g/kg i.v. CRH bolus results in a peak ACTH response within 15 min and a peak cortisol response within 30–60 min. A peak cortisol of 550 nmol/L or higher is considered to be a sufficient reaction. The CRH test however, is inferior to the ITT and metyrapone test (97).

Pitfalls

The use of exogenous corticosteroids can suppress the HPA axis. Therefore, in case of exogenous glucocorticoid use, a reliable evaluation of the HPA axis can not be performed within 6 weeks after withdrawal of the steroid but might even be disturbed many months thereafter. Contraceptives in females should also be stopped for at least 6 weeks because of the effects of hormonal agents on cortisol binding globulin (CBG) levels (98;99).

Thyrotropin deficiency

Clinical manifestation

The symptoms and signs associated with thyrotropin (TSH) deficiency are similar to those of primary hypothyroidism but usually are less severe, as there often is some residual thyrotropin secretion. In addition, TSH deficiency is almost always part of complete anterior pituitary hormone deficiency because thyreotroph secretion is the most resistant to insufficiency. Tiredness, cold intolerance, weight gain, constipation, dry skin, and hair loss are common features.

Diagnosis

TSH deficiency is diagnosed by low or normal serum TSH concentrations in the presence of low serum free thyroxine (fT_4) level. Measurement of serum fT_3 is not of additional value but may be low or normal. Thyrotropinreleasing hormone (TRH) can be used to assess TSH secretion. However, the response to TRH varies widely among individuals. Therefore it is not possible to discriminate between a normal and abnormal response in the majority of cases and TRH has not been incorporated into routine clinical practice of the evaluation of TSH deficiency (100).

Gonadotropin deficiency

Clinical manifestations

The clinical features of gonadotropin deficiency are determined by gender and the age of development. The physical examination in men with recent onset hypogonadism will usually be normal. However, in longstanding hypogonadism diminished facial and body hair, gynaecomastia, and small, weak testes can be present. Libido may be reduced and the ability to achieve and maintain an erection may be compromised. Patients can also complain of nonspecific symptoms, such as tiredness, reduced muscle strength, reduced exercise capacity, but also emotional lability and depression. The symptoms in men are nonspecific and therefore may not become evident for many years, particularly if fertility is not an issue (77).

In women gonadotropin deficiency leads to menstrual disturbances (*i.e.* oligomenorrhea or amenorrhea) and therefore often earlier diagnosed compared to men (2).

Diagnosis

The diagnosis in women is straightforward. In premenopausal women secondary amenorrhea with low levels of estradiol and low or nomal levels of gonadotropins will confirm the diagnosis. Whereas, in postmenopausal women FSH and LH will be (undetectably) low.

Low or normal gonadotropin levels combined with serum testosterone levels below the reference range, corrected for age are sufficient to confirm the diagnosis. Because of the great circadian variation randomly found decreased testosterone levels should be repeated in the early morning (between 8–9:00 AM).

Prolactin deficiency

Clinical manifestations

Mild hyperprolactinaemia (up to 5 times the upper limit of normal) is common in patients with hypopituitarism. A pituitary mass with supra-sellar extension may compress the stalk resulting in decreased dopaminergic inhibition of prolactin secretion.

Raised prolactine levels effects pulsatile secretion of gonadtropins resulting in hypogonadism. Galacthorrhea can also be present. Prolactin deficiency almost invariably results from lactotroph deficiency secondary to hypothalamic damage as a result of irradiation and or surgery.

Diagnosis

The diagnosis of prolactin deficiency is straightforward using commercially available assays with gender adjusted reference ranges for the determination of prolactin concentrations. However, unless it is in the postpartum period, there are no clinical implications.

IV. Outline of this thesis

Pituitary insufficiency in the presence of a pituitary macroadenoma or after pituitary irradiation is frequently reported. In addition, pituitary insufficiency is increasingly reported after traumatic head injuries. The correct evaluation and interpretation, however, of the pituitary axes, and consequently, the potential therapeutical consequences are a matter of controversies. The studies reported in this thesis aim to provide better insight into the complexity of different endocrine tests used for the evaluation of possible pituitary insufficiency and in the treatment of patients with pituitary insufficiency.

The evaluation of pituitary function in patients after traumatic brain injury

Traumatic brain injury (TBI) has emerged as an important cause of hypopituitarism. However, considerable variations in the prevalence of hypopituitarism are reported. These variations can partly be explained by the severity of trauma and timing of hormonal evaluation, but may also be dependent on endocrine tests and criteria used for diagnosis of hypopituitarism. Therefore, in **chapter 2**, we performed a systematic review of the literature to critically compare pituitary function tests, and definitions of hypopituitarism in studies that assessed the long-term outcome of TBI on pituitary function.

Because of the great variation in prevalence rates reported and the great variation in endocrine assessments used, we decided to perform a cross-sectional study in the Netherlands of a large cohort of 112 TBI patients evaluated after long-term follow-up. We assessed the prevalence of pituitary insufficiency in our own large cohort of TBI patients using a standardized endocrine evaluation, described in **chapter 3**. In these patients, we also evaluated quality of life (QoL) using different QoL questionnaires.

Dynamic tests of pituitary function in other pituitary diseases

Pituitary adenomas and their treatment (*i.e.* surgery and/or radiotherapy) are also causes for pituitary insufficiency. Pituitary insufficiency is a complication that can be attributed to the tumor itself (compression) but also to the surgical approach and/or subsequent radiotherapeutical intervention. Therefore, accurate assessment of pituitary function is critical for appropriate management of patients with pituitary adenoma after surgery with or without irradiation. For many years, all patients in our hospital underwent a CRH stimulation test for the evaluation of the HPA axis shortly after pituitary surgery. In **chapter 4**, we describe a retrospective study that evaluated the clinical applicability of the CRH test directly after TS in our center.

The ITT, however, is considered the golden standard test for the evaluation of the HPA axis. In **chapter 5**, we describe a study on the long-term prevalence of adrenal insufficiency after transsphenoidal surgery for growth-hormone secreting pituitary adenomas using the ITT and CRH test in the majority of the patients. The reason for this evaluation was a recently published study that reported a remarkably high prevalence of adrenal insufficiency after surgical and/or medical treatment without postoperative radiotherapy in patients treated for acromegaly. Therefore, in our study, we evaluated the prevalence and incidence rate of adrenal insufficiency in 91 consecutive patients during long-term follow-up after successful transsphenoidal surgery for acromegaly.

In addition to patients with pituitary tumors, patients with nonpituitary intracranial and or nasopharyngeal tumors are treated by radiotherapy, in which the pituitary gland is involved in the radiation field. These patients are also at risk for pituitary insufficiency. To assess the prevalence of possible pituitary insufficiencies we performed a systemic literature search and meta-analysis focusing on the prevalence of pituitary dysfunction in adult patients treated with radiotherapy for nonpituitary tumors, which is described in **chapter 6**.

Treatment of GH deficiency

When growth hormone deficiency is diagnosed, the therapeutical consequences should be carefully evaluated, especially in certain conditions like obesity and during senescence where GH secretion overlaps with a GH deficient state. With increasing age, but also increasing BMI, GH secretion decreases. Therefore, the effects of treatment with rhGH in obesity and in the elderly diagnosed with GHD might be different.

Therefore, in **chapter 7**, we performed a structured review, to critically assess the available literature in order to evaluate the available evidence for treatments of elderly patients with GHD.

References

- 1. Berne R, Levy NM, Koeppen BM, Stanton A. Physiology. 5th ed. 2011
- 2. Greenspan FS, Strewler GJ. Basic & Clinical Endocrinology. 5th ed. 1997
- 3. Giustina A, Veldhuis JD 1998 Pathophysiology of the neuroregulation of growth hormone secretion in experimental animals and the human. Endocr Rev 19:717–797
- 4. Hartman ML, Veldhuis JD, Thorner MO 1993 Normal control of growth hormone secretion. Horm Res 40:37–47
- Pincus SM, Gevers EF, Robinson IC, van den BG, Roelfsema F, Hartman ML, Veldhuis JD 1996 Females secrete growth hormone with more process irregularity than males in both humans and rats. Am J Physiol 270:E107–E115
- 6. Veldhuis JD, Liem AY, South S, Weltman A, Weltman J, Clemmons DA, Abbott R, Mulligan T, Johnson ML, Pincus S, . 1995 Differential impact of age, sex steroid hormones, and obesity on basal versus pulsatile growth hormone secretion in men as assessed in an ultrasensitive chemiluminescence assay. J Clin Endocrinol Metab 80:3209–3222
- 7. Iranmanesh A, Lizarralde G, Veldhuis JD 1991 Age and relative adiposity are specific negative determinants of the frequency and amplitude of growth hormone (GH) secretory bursts and the half-life of endogenous GH in healthy men. J Clin Endocrinol Metab 73:1081–1088
- Veldhuis JD, Iranmanesh A, Ho KK, Waters MJ, Johnson ML, Lizarralde G 1991 Dual defects in pulsatile growth hormone secretion and clearance subserve the hyposomatotropism of obesity in man. J Clin Endocrinol Metab 72:51–59
- 9. Rosenfeld RG, Roberts CT. The IGF System. 1999
- 10. Le Roith D, Bondy C, Yakar S, Liu JL, Butler A 2001 The somatomedin hypothesis: 2001. Endocr Rev 22:53–74
- 11. Gwinup G, Johnson B 1975 Clinical testing of the hypothalamic-pituitary-adrenocortical system in states of hypo- and hypercortisolism. Metabolism 24:777–791
- Debono M, Ghobadi C, Rostami-Hodjegan A, Huatan H, Campbell MJ, Newell-Price J, Darzy K, Merke DP, Arlt W, Ross RJ 2009 Modified-release hydrocortisone to provide circadian cortisol profiles. J Clin Endocrinol Metab 94:1548–1554
- Weitzman ED, Fukushima D, Nogeire C, Roffwarg H, Gallagher TF, Hellman L 1971 Twenty-four hour pattern of the episodic secretion of cortisol in normal subjects. J Clin Endocrinol Metab 33:14–22

- 14. Gallagher TF, Yoshida K, Roffwarg HD, Fukushima DK, Weitzman ED, Hellman L 1973 ACTH and cortisol secretory patterns in man. J Clin Endocrinol Metab 36:1058–1068
- 15. Veldhuis JD, Iranmanesh A, Johnson ML, Lizarralde G 1990 Twenty-four-hour rhythms in plasma concentrations of adenohypophyseal hormones are generated by distinct amplitude and/or frequency modulation of underlying pituitary secretory bursts. J Clin Endocrinol Metab 71:1616–1623
- 16. Vaughan GM, Becker RA, Allen JP, Goodwin CW, Jr., Pruitt BA, Jr., Mason AD, Jr. 1982 Cortisol and corticotrophin in burned patients. J Trauma 22:263–273
- 17. Jeffries MK, Vance ML 1992 Growth hormone and cortisol secretion in patients with burn injury. J Burn Care Rehabil 13:391–395
- Udelsman R, Norton JA, Jelenich SE, Goldstein DS, Linehan WM, Loriaux DL, Chrousos GP 1987 Responses of the hypothalamic-pituitary-adrenal and renin-angiotensin axes and the sympathetic system during controlled surgical and anesthetic stress. J Clin Endocrinol Metab 64:986–994
- Calogero AE, Norton JA, Sheppard BC, Listwak SJ, Cromack DT, Wall R, Jensen RT, Chrousos GP 1992 Pulsatile activation of the hypothalamic-pituitary-adrenal axis during major surgery. Metabolism 41:839–845
- Watabe T, Tanaka K, Kumagae M, Itoh S, Takeda F, Morio K, Hasegawa M, Horiuchi T, Miyabe S, Shimizu N 1987 Hormonal responses to insulin-induced hypoglycemia in man. J Clin Endocrinol Metab 65:1187–1191
- 21. Fish HR, Chernow B, O'Brian JT 1986 Endocrine and neurophysiologic responses of the pituitary to insulin-induced hypoglycemia: a review. Metabolism 35:763–780
- Luger A, Deuster PA, Kyle SB, Gallucci WT, Montgomery LC, Gold PW, Loriaux DL, Chrousos GP 1987 Acute hypothalamic-pituitary-adrenal responses to the stress of treadmill exercise. Physiologic adaptations to physical training. N Engl J Med 316:1309–1315
- Streeten DH, Anderson GH, Jr., Dalakos TG, Seeley D, Mallov JS, Eusebio R, Sunderlin FS, Badawy SZ, King RB 1984 Normal and abnormal function of the hypothalamic-pituitaryadrenocortical system in man. Endocr Rev 5:371–394
- 24. Keller-Wood M, Shinsako J, Dallman MF 1984 Interaction between stimulus intensity and corticosteroid feedback in control of ACTH. Am J Physiol 247:E489–E494
- 25. Schlaghecke R, Kornely E, Santen RT, Ridderskamp P 1992 The effect of long-term glucocorticoid therapy on pituitary-adrenal responses to exogenous corticotropin-releasing hormone. N Engl J Med 326:226–230
- 26. Livanou T, Ferriman D, James VH 1967 Recovery of hypothalamo-pituitary-adrenal function after corticosteroid therapy. Lancet 2:856–859
- Harrison BD, Rees LH, Cayton RM, Nabarro JD 1982 Recovery of hypothalamo-pituitaryadrenal function in asthmatics whose oral steroids have been stopped or reduced. Clin Endocrinol (Oxf) 17:109–118

- 28. Streck WF, Lockwood DH 1979 Pituitary adrenal recovery following short-term suppression with corticosteroids. Am J Med 66:910–914
- 29. Melby JC 1974 Drug spotlight program: systemic corticosteroid therapy: pharmacology and endocrinologic considerations. Ann Intern Med 81:505–512
- 30. Arafah BM, Prunty D, Ybarra J, Hlavin ML, Selman WR 2000 The dominant role of increased intrasellar pressure in the pathogenesis of hypopituitarism, hyperprolactinemia, and headaches in patients with pituitary adenomas. J Clin Endocrinol Metab 85:1789–1793
- Fainstein DP, Guitelman M, Artese R, Fiszledjer L, Chervin A, Vitale NM, Stalldecker G, De M, V, Cornalo D, Alfieri A, Susana M, Gil M 2004 Retrospective multicentric study of pituitary incidentalomas. Pituitary 7:145–148
- Feldkamp J, Santen R, Harms E, Aulich A, Modder U, Scherbaum WA 1999 Incidentally discovered pituitary lesions: high frequency of macroadenomas and hormone-secreting adenomas – results of a prospective study. Clin Endocrinol (Oxf) 51:109–113
- Gondim JA, Almeida JP, Albuquerque LA, Schops M, Gomes E, Ferraz T, Sobreira W, Kretzmann MT Endoscopic endonasal approach for pituitary adenoma: surgical complications in 301 patients. Pituitary 2011 Jun;14(2):174-83
- Tabaee A, Anand VK, Barron Y, Hiltzik DH, Brown SM, Kacker A, Mazumdar M, Schwartz TH 2009 Endoscopic pituitary surgery: a systematic review and meta-analysis. J Neurosurg 111:545–554
- 35. Ayuk J, Stewart PM 2009 Mortality following pituitary radiotherapy. Pituitary 12:35–39
- 36. Fernandez A, Brada M, Zabuliene L, Karavitaki N, Wass JA 2009 Radiation-induced hypopituitarism. Endocr Relat Cancer 16:733–772
- 37. Bhandare N, Kennedy L, Morris CG, Malyapa R, Mendenhall WM 2006 1099: Hypopituitarism After Radiation Therapy for Extracranial Head and Neck Cancers. International Journal of Radiation Oncology Biology Physics 66:S187–S188
- Darzy KH, Shalet SM 2009 Hypopituitarism following Radiotherapy Revisited. Endocr Dev 15:1–24
- 39. Shalet SM 1993 Radiation and pituitary dysfunction. N Engl J Med 328:131–133
- 40. Agha A, Rogers B, Sherlock M, O'Kelly P, Tormey W, Phillips J, Thompson CJ 2004 Anterior pituitary dysfunction in survivors of traumatic brain injury. J Clin Endocrinol Metab 89:4929–4936
- Bondanelli M, Ambrosio MR, Cavazzini L, Bertocchi A, Zatelli MC, Carli A, Valle D, Basaglia N, Uberti EC 2007 Anterior pituitary function may predict functional and cognitive outcome in patients with traumatic brain injury undergoing rehabilitation. J Neurotrauma 24:1687–1697
- 42. Bushnik T, Englander J, Katznelson L 2007 Fatigue after TBI: association with neuroendocrine abnormalities. Brain Inj 21:559–566

- Herrmann BL, Rehder J, Kahlke S, Wiedemayer H, Doerfler A, Ischebeck W, Laumer R, Forsting M, Stolke D, Mann K 2006 Hypopituitarism following severe traumatic brain injury. Exp Clin Endocrinol Diabetes 114:316–321
- 44. Kelly DF, Gonzalo IT, Cohan P, Berman N, Swerdloff R, Wang C 2000 Hypopituitarism following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a preliminary report. J Neurosurg 93:743–752
- 45. Klose M, Juul A, Poulsgaard L, Kosteljanetz M, Brennum J, Feldt-Rasmussen U 2007 Prevalence and predictive factors of post-traumatic hypopituitarism. Clin Endocrinol (Oxf) 67:193–201
- 46. Leal-Cerro A, Flores JM, Rincon M, Murillo F, Pujol M, Garcia-Pesquera F, Dieguez C, Casanueva FF 2005 Prevalence of hypopituitarism and growth hormone deficiency in adults long-term after severe traumatic brain injury. Clin Endocrinol (Oxf) 62:525–532
- Lieberman SA, Oberoi AL, Gilkison CR, Masel BE, Urban RJ 2001 Prevalence of neuroendocrine dysfunction in patients recovering from traumatic brain injury. J Clin Endocrinol Metab 86:2752–2756
- 48. Popovic V, Pekic S, Pavlovic D, Maric N, Jasovic-Gasic M, Djurovic B, Medic SM, Zivkovic V, Stojanovic M, Doknic M, Milic N, Djurovic M, Dieguez C, Casanueva FF 2004 Hypopituitarism as a consequence of traumatic brain injury (TBI) and its possible relation with cognitive disabilities and mental distress. J Endocrinol Invest 27:1048–1054
- Schneider HJ, Schneider M, Saller B, Petersenn S, Uhr M, Husemann B, von RF, Stalla GK 2006 Prevalence of anterior pituitary insufficiency 3 and 12 months after traumatic brain injury. Eur J Endocrinol 154:259–265
- 50. Tanriverdi F, Senyurek H, Unluhizarci K, Selcuklu A, Casanueva FF, Kelestimur F 2006 High risk of hypopituitarism after traumatic brain injury: a prospective investigation of anterior pituitary function in the acute phase and 12 months after trauma. J Clin Endocrinol Metab 91:2105–2111
- 51. Wachter D, Gundling K, Oertel MF, Stracke H, Boker DK 2009 Pituitary insufficiency after traumatic brain injury. J Clin Neurosci 16:202–208
- 52. Schneider HJ, Kreitschmann-Andermahr I, Ghigo E, Stalla GK, Agha A 2007 Hypothalamopituitary dysfunction following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a systematic review. JAMA 298:1429–1438
- Urban RJ, Harris P, Masel B 2005 Anterior hypopituitarism following traumatic brain injury. Brain Inj 19:349–358
- 54. Agha A, Thompson CJ 2006 Anterior pituitary dysfunction following traumatic brain injury (TBI). Clin Endocrinol (Oxf) 64:481–488
- de Boer H, Blok GJ, Van der Veen, V 1995 Clinical aspects of growth hormone deficiency in adults. Endocr Rev 16:63–86

- 56. Brabant G, Poll EM, Jonsson P, Polydorou D, Kreitschmann-Andermahr I 2009 Etiology, baseline characteristics, and biochemical diagnosis of GH deficiency in the adult: are there regional variations? Eur J Endocrinol 161 Suppl 1:S25–S31
- 57. Carroll PV, Christ ER, Bengtsson BA, Carlsson L, Christiansen JS, Clemmons D, Hintz R, Ho K, Laron Z, Sizonenko P, Sonksen PH, Tanaka T, Thorne M 1998 Growth hormone deficiency in adulthood and the effects of growth hormone replacement: a review. Growth Hormone Research Society Scientific Committee. J Clin Endocrinol Metab 83:382–395
- 58. Attanasio AF, Mo D, Erfurth EM, Tan M, Ho KY, Kleinberg D, Zimmermann AG, Chanson P 2010 Prevalence of metabolic syndrome in adult hypopituitary growth hormone (GH)-deficient patients before and after GH replacement. J Clin Endocrinol Metab 95:74–81
- 59. Burman P, Deijen JB 1998 Quality of life and cognitive function in patients with pituitary insufficiency. Psychother Psychosom 67:154–167
- 60. Rosen T, Wiren L, Wilhelmsen L, Wiklund I, Bengtsson BA 1994 Decreased psychological wellbeing in adult patients with growth hormone deficiency. Clin Endocrinol (Oxf) 40:111–116
- 61. Abs R, Mattsson AF, Bengtsson BA, Feldt-Rasmussen U, Goth MI, Koltowska-Haggstrom M, Monson JP, Verhelst J, Wilton P 2005 Isolated growth hormone (GH) deficiency in adult patients: baseline clinical characteristics and responses to GH replacement in comparison with hypopituitary patients. A sub-analysis of the KIMS database. Growth Horm IGF Res 15:349–359
- 62. Aimaretti G, Corneli G, Rovere S, Granata R, Baldelli R, Grottoli S, Ghigo E 2004 Insulin-like growth factor I levels and the diagnosis of adult growth hormone deficiency. Horm Res 62 Suppl 1:26–33
- 63. Ghigo E, Aimaretti G, Corneli G 2008 Diagnosis of adult GH deficiency. Growth Horm IGF Res 18:1–16
- 64. Aimaretti G, Corneli G, Baldelli R, Di SC, Gasco V, Durante C, Ausiello L, Rovere S, Grottoli S, Tamburrano G, Ghigo E 2003 Diagnostic reliability of a single IGF-I measurement in 237 adults with total anterior hypopituitarism and severe GH deficiency. Clin Endocrinol (Oxf) 59:56–61
- 65. Aimaretti G, Corneli G, Di SC, Baldelli R, Gasco V, Rovere S, Migliaretti G, Colao A, Tamburrano G, Lombardi G, Ghigo E, Camanni F 2005 Different degrees of GH deficiency evidenced by GHRH-arginine test and IGF-I levels in adults with pituritary disease. J Endocrinol Invest 28:247–252
- 66. Ghigo E, Aimaretti G, Arvat E, Camanni F 2001 Growth hormone-releasing hormone combined with arginine or growth hormone secretagogues for the diagnosis of growth hormone deficiency in adults. Endocrine 15:29–38
- 67. Consensus guidelines for the diagnosis and treatment of adults with growth hormone deficiency: summary statement of the Growth Hormone Research Society Workshop on Adult Growth Hormone Deficiency 1998 J Clin Endocrinol Metab 83:379–381

- 68. Aimaretti G, Corneli G, Razzore P, Bellone S, Baffoni C, Arvat E, Camanni F, Ghigo E 1998 Comparison between insulin-induced hypoglycemia and growth hormone (GH)-releasing hormone + arginine as provocative tests for the diagnosis of GH deficiency in adults. J Clin Endocrinol Metab 83:1615–1618
- Biller BM, Samuels MH, Zagar A, Cook DM, Arafah BM, Bonert V, Stavrou S, Kleinberg DL, Chipman JJ, Hartman ML 2002 Sensitivity and specificity of six tests for the diagnosis of adult GH deficiency. J Clin Endocrinol Metab 87:2067–2079
- Alba-Roth J, Muller OA, Schopohl J, von WK 1988 Arginine stimulates growth hormone secretion by suppressing endogenous somatostatin secretion. J Clin Endocrinol Metab 67:1186–1189
- Shibasaki T, Hotta M, Masuda A, Imaki T, Obara N, Demura H, Ling N, Shizume K 1985 Plasma GH responses to GHRH and insulin-induced hypoglycemia in man. J Clin Endocrinol Metab 60:1265–1267
- 72. Colao A, Di SC, Savastano S, Rota F, Savanelli MC, Aimaretti G, Lombardi G 2009 A reappraisal of diagnosing GH deficiency in adults: role of gender, age, waist circumference, and body mass index. J Clin Endocrinol Metab 94:4414–4422
- Kopelman PG, Noonan K, Goulton R, Forrest AJ 1985 Impaired growth hormone response to growth hormone releasing factor and insulin-hypoglycaemia in obesity. Clin Endocrinol (Oxf) 23:87–94
- 74. Scacchi M, Pincelli Al, Cavagnini F 1999 Growth hormone in obesity. Int J Obes Relat Metab Disord 23:260–271
- 75. Veldhuis JD, Roelfsema F, Keenan DM, Pincus S 2011 Gender, age, body mass index, and IGF-I individually and jointly determine distinct GH dynamics: analyses in one hundred healthy adults. J Clin Endocrinol Metab 96:115–121
- 76. Qu XD, Gaw G, I, Al Sayed MY, Cohan P, Christenson PD, Swerdloff RS, Kelly DF, Wang C 2005 Influence of body mass index and gender on growth hormone (GH) responses to GH-releasing hormone plus arginine and insulin tolerance tests. J Clin Endocrinol Metab 90:1563–1569
- 77. van Aken MO, Lamberts SW 2005 Diagnosis and treatment of hypopituitarism: an update. Pituitary 8:183–191
- 78. Arlt W, Allolio B 2003 Adrenal insufficiency. Lancet 361:1881–1893
- 79. Clayton RN 1996 Short Synacthen test versus insulin stress test for assessment of the hypothalamo [correction of hypothalmo]-pituitary--adrenal axis: controversy revisited. Clin Endocrinol (Oxf) 44:147–149
- 80. Grinspoon SK, Biller BM 1994 Clinical review 62: Laboratory assessment of adrenal insufficiency. J Clin Endocrinol Metab 79:923–931
- Streeten DH 1999 What test for hypothalamic-pituitary-adrenocortical insufficiency? Lancet 354:179–180

- 82. Nye EJ, Grice JE, Hockings GI, Strakosch CR, Crosbie GV, Walters MM, Jackson RV 1999 Comparison of adrenocorticotropin (ACTH) stimulation tests and insulin hypoglycemia in normal humans: low dose, standard high dose, and 8-hour ACTH-(1–24) infusion tests. J Clin Endocrinol Metab 84:3648–3655
- Mayenknecht J, Diederich S, Bahr V, Plockinger U, Oelkers W 1998 Comparison of low and high dose corticotropin stimulation tests in patients with pituitary disease. J Clin Endocrinol Metab 83:1558–1562
- 84. Abdu TA, Elhadd TA, Neary R, Clayton RN 1999 Comparison of the low dose short synacthen test (1 microg), the conventional dose short synacthen test (250 microg), and the insulin tolerance test for assessment of the hypothalamo-pituitary-adrenal axis in patients with pituitary disease. J Clin Endocrinol Metab 84:838–843
- 85. Stewart PM, Clark PM, Sheppard MC 1998 Comparison of the short ACTH stimulation test with the insulin tolerance/glucagon test. Clin Endocrinol (Oxf) 48:124–126
- 86. Mukherjee JJ, de Castro JJ, Kaltsas G, Afshar F, Grossman AB, Wass JA, Besser GM 1997 A comparison of the insulin tolerance/glucagon test with the short ACTH stimulation test in the assessment of the hypothalamo-pituitary-adrenal axis in the early post-operative period after hypophysectomy. Clin Endocrinol (Oxf) 47:51–60
- Fiad TM, Kirby JM, Cunningham SK, McKenna TJ 1994 The overnight single-dose metyrapone test is a simple and reliable index of the hypothalamic-pituitary-adrenal axis. Clin Endocrinol (Oxf) 40:603–609
- Courtney CH, McAllister AS, McCance DR, Hadden DR, Leslie H, Sheridan B, Atkinson AB 2000 The insulin hypoglycaemia and overnight metyrapone tests in the assessment of the hypothalamic-pituitary-adrenal axis following pituitary surgery. Clin Endocrinol (Oxf) 53:309–312
- 89. Staub JJ, Noelpp B, Girard J, Baumann JB, Graf S, Ratcliffe JG 1979 The short metyrapone test: comparison of the plasma ACTH response to metyrapone and insulin-induced hypoglycaemia. Clin Endocrinol (Oxf) 10:595–601
- 90. de Lange WE, Sluiter WJ, Pratt JJ, Wolf RF, Doorenbos H 1988 Is the ovine CRF test an aid in the evaluation of patients with secondary adrenal insufficiency? Neth J Med 32:72–78
- 91. Hermus AR, Pieters GF, Smals AG, Kloppenborg PW 1985 Corticotropin-releasing factor, a new aid in the diagnosis of pituitary-adrenal disorders? Neth J Med 28:2–5
- 92. Pieters GF, Hermus AR, Smals AG, Bartelink AK, Benraad TJ, Kloppenborg PW 1983 Responsiveness of the hypophyseal-adrenocortical axis to corticotropin-releasing factor in pituitary-dependent Cushing's disease. J Clin Endocrinol Metab 57:513–516
- 93. Schulte HM, Chrousos GP, Avgerinos P, Oldfield EH, Gold PW, Cutler GB, Jr., Loriaux DL 1984 The corticotropin-releasing hormone stimulation test: a possible aid in the evaluation of patients with adrenal insufficiency. J Clin Endocrinol Metab 58:1064–1067

- 94. Stalla GK, Losa M, Oeckler R, Muller OA, von WK 1988 Insulin hypoglycemia test and releasing hormone (corticotropin-releasing hormone and growth hormone-releasing hormone) stimulation in patients with pituitary failure of different origin. Horm Res 29:191–196
- 95. Dullaart RP, Pasterkamp SH, Beentjes JA, Sluiter WJ 1999 Evaluation of adrenal function in patients with hypothalamic and pituitary disorders: comparison of serum cortisol, urinary free cortisol and the human-corticotrophin releasing hormone test with the insulin tolerance test. Clin Endocrinol (Oxf) 50:465–471
- 96. Tsukada T, Nakai Y, Koh T, Tsujii S, Inada M, Nishikawa M, Shinoda H, Kawai I, Takezawa N, Imura H 1984 Plasma adrenocorticotropin and cortisol responses to ovine corticotropinreleasing factor in patients with adrenocortical insufficiency due to hypothalamic and pituitary disorders. J Clin Endocrinol Metab 58:758–760
- 97. Dickstein G 2003 The assessment of the hypothalamo-pituitary-adrenal axis in pituitary disease: are there short cuts? J Endocrinol Invest 26:25–30
- 98. Kirschbaum C, Kudielka BM, Gaab J, Schommer NC, Hellhammer DH 1999 Impact of gender, menstrual cycle phase, and oral contraceptives on the activity of the hypothalamus-pituitaryadrenal axis. Psychosom Med 61:154–162
- Kumsta R, Entringer S, Hellhammer DH, Wust S 2007 Cortisol and ACTH responses to psychosocial stress are modulated by corticosteroid binding globulin levels. Psychoneuroendocrinology 32:1153–1157
- 100. Faglia G 1998 The clinical impact of the thyrotropin-releasing hormone test. Thyroid 8:903–908

Chapter 2

Clinical review: Hypopituitarism following traumatic brain injury – the prevalence is affected by the use of different dynamic tests and different normal values

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Abstract

Objective: Traumatic brain injury (TBI) has emerged as an important cause of hypopituitarism. However, considerable variations in the prevalence of hypopituitarism are reported. These can partly be explained by severity of trauma and timing of hormonal evaluation, but may also be dependent on endocrine tests and criteria used for diagnosis of hypopituitarism.

Methods: Systematic review of studies reporting prevalence of hypopituitarism in adults \geq 1 year after TBI focusing on used (dynamic) tests and biochemical criteria.

Results: We included data from 14 studies with a total of 931 patients. There was considerable variation in definition of hypopituitarism. Overall, reported prevalences of severe GH deficiency varied between 2 and 39%. Prevalences were 8–20% using the GHRH-arginine test (cut-off < 9 μ g/L), 11–39% using the glucagon test (cut-off 1–5 μ g/L), 2% using the GHRH test (no cut-off) and 15–18% using the insulin tolerance test (ITT) (cut-off < 3 μ g/L).

Overall, the reported prevalence of secondary adrenal insufficiency had a broad range from 0 to 60%. This prevalence was 0–60% with basal cortisol (cut-off < 220 or < 440 nmol/L), 7–19% using the ACTH test and 5% with the ITT as first test (cut-off < 500 or < 550 nmol/L). Secondary hypothyroidism was present in 0–19% (free T_4) or 5–15% (TRH stimulation). Secondary hypogonadism was present in 0–29%.

Conclusion: The reported variations in the prevalence rates of hypopituitarism after TBI are in part caused by differences in definitions, endocrine assessments of hypopituitarism and confounding factors. These methodological issues prohibit simple generalizations of results of original studies on TBI-associated hypopituitarism in the perspective of meta-analyses or reviews.

Introduction

In recent years, an increasing number of studies have reported the presence of pituitary insufficiency in patients who experienced traumatic brain injury (TBI) (1–14). The prevalence of pituitary insufficiency after TBI appeared to be unexpectedly high (15;16). Remarkably, the prevalence rates varied considerably among the different studies, ranging from 15 to even 90% of the patients.

Several factors influence the prevalence of hypopituitarism after TBI. First, the time interval between TBI and the assessment of pituitary function, since hormone alterations mimicking pituitary insufficiency are prevalent in the early post-traumatic period. Second, the type and severity of the brain injury affects the prevalence of hypopituitarism, because persistent pituitary insufficiency is only frequent after severe TBI (7;15). Third, endocrine tests, assays, and criteria for the diagnosis of hypopituitarism differ between the studies. Although many reviews have addressed TBI-related hypopituitarism, a detailed comparison of these methodological issues between the different studies has not been performed for each pituitary axis.

We hypothesized that these methodological differences may have contributed, at least in part, to the discrepancies in prevalence rates of hypopituitarism after TBI, reported by the different studies. Therefore, the aim of this study was to critically compare the pituitary function tests, and definitions of hypopituitarism between studies that assessed the long term outcome of TBI on pituitary function.

Patients and methods

Search strategy

We performed a search in PubMed, EMBASE, Web of Science, and the Cochrane database, for all published studies on the association between TBI and hypopituitarism. The following search strategy was used: (*Traumatic Brain Injury OR Traumatic brain injuries*) AND (traumatic OR trauma) AND (Hypopituitarism OR Hypopituitar* OR Hypothalamus Hypophysis System OR "Hypothalamopituitary dysfunction" OR "pituitary dysfunction" OR Hypothalamo-Hypophyseal System OR Pituitary Gland OR Hypophysis).

In addition, the references of relevant articles were checked for additional articles. The search was performed on 23 March 2009. Only original articles were included. We used the following exclusion criteria: pediatric or adolescent population, publications concerning pituitary testing < 12 months after injury (a median of 12 months was accepted), and articles that evaluated pituitary insufficiency after subarachnoidal bleeding (SAB).

Data review

The following data were extracted from each study: 1) age and gender, 2) the endocrine tests used for assessment of each pituitary axis, 3) definitions used for pituitary insufficiency for each pituitary axis, 4) hormone assays, 5) reference values provided in the manuscript, and 6) use of control populations. Tables were constructed per pituitary axis. These tables are added as supplemental data files. The growth hormone (GH)-IGF-I axis (Table 2), the pituitary-adrenal axis (Table 3), the pituitary-thyroidal axis (Table 4), the pituitary-gonadal axis (Table 5), and prolactin (Table 6).

Results

We identified 278 articles, of which 218 were excluded on the basis of title and abstract. Of the remaining 60 articles, 46 were reviews. Finally, 14 original studies were included with a total of 931 patients. Details of these studies are summarized in Table 1. The number of patients evaluated by the different studies varied between 22 and 105.

	Year of		Time of testing post TBI	Trauma severity	γ	Any pituitary
Study	publication	No. of patients	[months (median)]	(GCS)	BMI (kg/m²)	deficiency (%)
Kelly <i>et al</i> .(6)	2000	22	3–276 (median 26)	3-15	25.1±6.5	37
Lieberman <i>et al.</i> (9)	2001	70	1–276 (median 13)	3–15	NR	69
Bondanelli <i>et al.</i> (3)	2004	50	12–64	84% GCS ≤ 8 3-15	24.6±0.4	54
Aqha <i>et al.</i> (1)	2004	102	6–36 (median 17)	54% GCS ≤ 8 3-13	NR	28
				56% GCS ≤ 8		
Popovic <i>et al.</i> (10)	2004	67	12–264	3-13	24.8±0.5	34
Aimaretti et al.(2)	2005	70	12	3-15	23.8±0.4	23
				21% GCS ≤ 8		
Leal-Cerro <i>et al.</i> (8)	2005	66	>12	8 VI	25.2±3.0 (n=44)	25
Schneider <i>et al.</i> (11)	2006	70	12	3-15	23.8±3.2	36
Tanriverdi <i>et al.</i> (12)	2006	52	12	3–15	NR	51
				25% GCS ≤ 8		
Herrmann <i>et al.</i> (5)	2006	76	5-47	8 8	25.8±4.2	24
Bushnik <i>et al.</i> (4)	2007	64	> 12 months	NR	NR	90
Klose <i>et al.</i> (7)	2007	104	10–27 (median 13)	3-15	25*	15
				38% GCS ≤ 8	(17–39)	
Tanriverdi <i>et al.</i> (14)	2008	30	36	3-15	NR	30
				16.7% GCS ≤ 8		
Wachter <i>et al.</i> (13)	2009	55	NR	3–15	NR	25
				17% GCS ≤ 8		
Total No. of patients:		931				

 Table 1.
 Studies on TBI and pituitary deficiency

42

CHAPTER 2

The GH-IGF-I axis

The prevalence of GH deficiency (GHD) ranged between 2 and 66% (severe GHD 39%; Figures 1 and 2 and Suppl Table 2). The presence of GHD was associated with higher body mass index (BMI) values in some of the studies (Figure 1). In addition to basal serum GH and IGF-I values, all studies used a dynamic test to assess GH secretory reserve. However, different dynamic tests were used.

Three studies (196/931=21% of all patients) used the combined GHRHarginine test as the first screening. The criterion for severe GHD was a peak GH level < 9.0 μ g/L in all three, which was not adjusted for BMI. Prevalence rates of severe GHD varied between 8 and 20% (weighted mean 12%) (2;3;5). Schneider *et al.* (11) also used the GHRH-arginine test, but only in a subset of the patients (those with abnormal serum cortisol levels, n=32); the prevalence of GHD in this study was 10%.

Two studies (112/931=12% of all patients) used an insulin tolerance test (ITT) as the primary screening test (6;7). The criterion for severe GHD was a peak GH response < 3 μ g/L in both, and the prevalence of GHD was comparable (18 and 15% respectively; weighted mean 16%).

Of the eight remaining studies, three used a stimulation test with glucagon (n=209) (1;4;9) with prevalence rates for severe GHD between 11 and 39% (weighted mean 20%). The cut-off values differed considerably and varied between 1 and 5 μ g/L between these studies. Just one study used a stimulation test with GHRH only (number of patients not recorded) reporting a GHD prevalence of 2% (13). Two studies (n=119) used the combined GHRH-GHRP6 test with a prevalence of 15 and 33% respectively (weighted mean 21%) (10;12). The cut-off values were similar (GH < 10 μ g/L) within these studies, and were derived from another report (17).

Finally, two studies used a combination of these tests (8;14). For instance, Agha *et al.* (1) used a glucagon stimulation test for the initial screening in 102 subjects, and in case of incomplete GH response, they used an ITT (n=14) or combined GHRH plus arginine test (n=4) to confirm GHD.

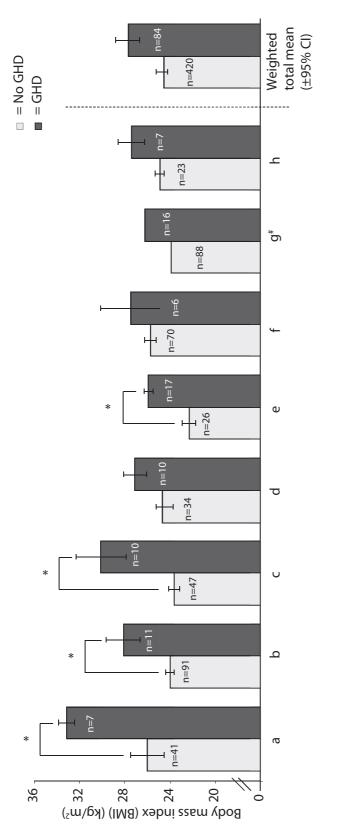
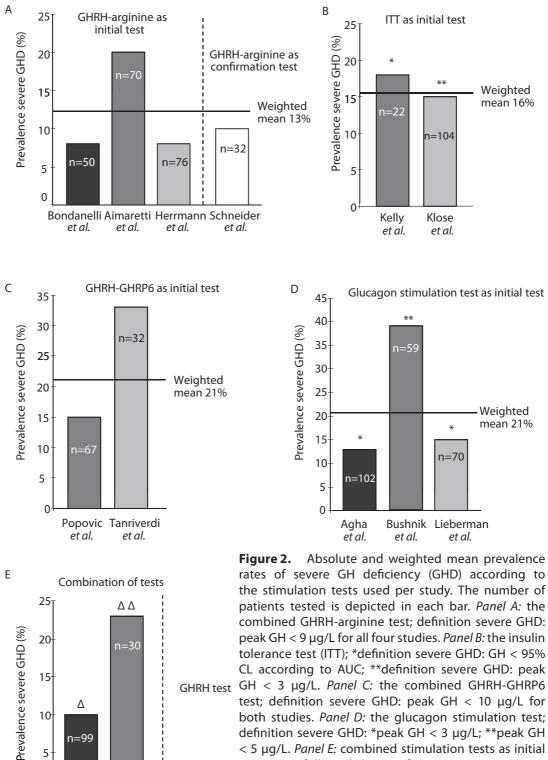


Figure 1. Body mass index in patients diagnosed with versus those without GH deficiency (data available only in 8 out of the 16 studies). a, Lieberman etal. 9); b, Agha etal.(1); c, Popovic etal. (10); d, Leal-Cerro etal. (8); e, Tanriverdi et al.(12); f, Herrmann etal.(5); g, Klose etal.(7); h, Tanriverdi etal.(14); i, weighted total mean (mean±95% CI: no GHD 24.5 (24.2–24.9) versus GHD 27.7 (26.7–28.8) kg/m²). GHD, GH deficiency; BMI, body mass index; *P<0.05. *Data reported as median with range; not included in the total weighted mean.



10

5

0

n=99

et al.

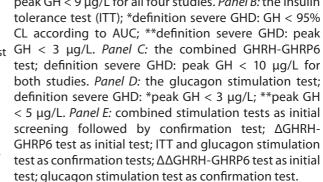
Leal-Cerro Tanriverdi

et al.

n=55

Wachter

et al.



45

The Pituitary-Adrenal axis

The prevalence of secondary adrenal insufficiency deficiency ranged from 0 to 60% between the studies (Figure 3, Suppl Table 3).

Four studies (251/931=27%) of all patients) only measured basal morning fasting serum cortisol and/or ACTH levels (2-4;10), resulting in prevalence rates between 0 and 60% (weighted mean 15%). The criteria for pituitary-adrenal insufficiency differed between three studies (cortisol < 220–440 nmol/L), and were not reported in the fourth study (10). The study reporting the highest prevalence of 60% used a cut-off value of 440 nmol/L (4).

Four studies (145/931=12% of all patients) used an ACTH stimulation test (Synacthen; either with 1 or 250 μ g). However, only one study performed this test in all patients and the prevalence of ACTH deficiency was 7% (9). In the other three studies, only a subset of the patients (those with subnormal basal cortisol levels) underwent stimulation with ACTH. The prevalence in these studies varied between 7 and 19% (weighted mean 10%) (11;12;14).

One study (55/931=6% of all patients) used nonstimulated cortisol values between 1600 and 2000 h (reference values 63–339 nmol/L), which was followed by a corticotrope releasing hormone (CRH) test only in those with values below this reference range, or in those who responded confirmatory to a specific questionnaire (13).

In the remaining five studies (403/931=43% of all patients), the ITT was used in 169 patients as a primary test (n=112) resulting in a prevalence of 5% in both studies (6;7), or as a confirmation test in a subset of the patients. Two studies measured basal serum cortisol levels and used ITT as a confirmation test (prevalence of 3 and 11% respectively) (5;8). One study assessed primarily with a glucagon stimulation test (n=102), and used the ITT and ACTH tests to confirm ACTH deficiency (prevalence 13%) (1). The criteria for a normal cortisol response to hypoglycemia were a peak cortisol level > 550 nmol/L in one (8), and > 500 nmol/L in three other studies (1;5;7). The fifth study used a control group of 18 healthy subjects to define normal cortisol responses to ITT (cortisol response < 95% confidence limit according to the obtained area under the curve) (6). The CRH test was used in only one study and did not report the number of patients (13).

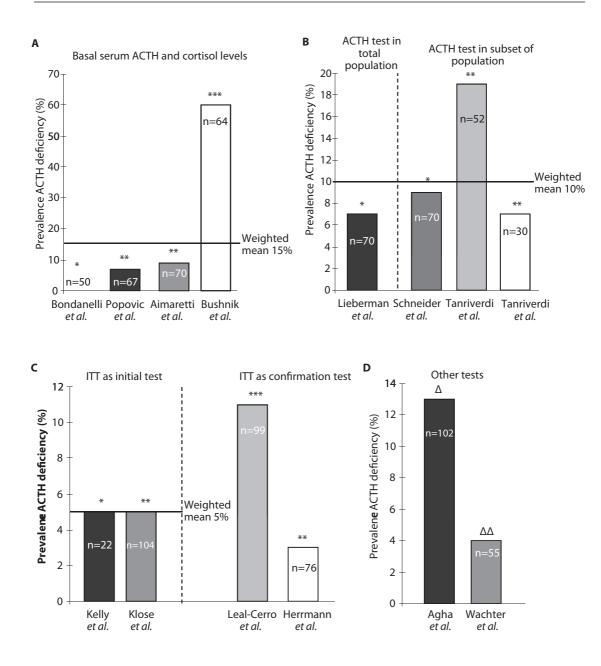


Figure 3. Absolute and weighted mean prevalence rates of corticotropin (HPA axis) deficiency according to the stimulation test used per study. The number of patients tested is depicted in each bar. *Panel A*: basal cortisol concentrations only using different cut-off levels; *cut-off level: NR; **cut-off level: cortisol < 220 nmol/L; ***cut-off level: cortisol < 440 nmol/L. *Panel B*: the ACTH stimulation test *using 250 µg ACTH and peak cortisol < 500 nmol/L; **using 1 µg ACTH and peak cortisol < 550 mol/L. *Panel C*: the insulin tolerance test (ITT); *peak cortisol < 95% CL according to AUC; **peak cortisol < 500 nmol/L; ***peak cortisol < 550 mol/L. *Panel D*: other stimulation tests: Δ the glucagon stimulation test; $\Delta\Delta$ CRH test.NR, not reported.

The Hypothalamus-Pituitary-Thyroid axis

The prevalence of hypothalamus-pituitary-thyroid axis deficiency ranged from 0 to 19% between the studies (Suppl Table 4).

The criteria for TSH deficiency were different. Nine studies used basal free thyroxine (fT_4) and TSH levels only. Within these studies, the cut-off value for decreased fT_4 varied between 8 and 12 pmol/L (2;5;7;11;14;18;19). In two studies, reference values were not reported (4;10), one of which (Bushnik *et al.*) reported the highest prevalence of secondary hypothyroidism.

The thyroid-releasing hormone (TRH) stimulation test was used in five studies, using i.v. doses of 200 (13) and 500 μ g (5;6;8;9). The criterion for a normal response differed considerably: a TSH peak response > 7 mIU/L, a TSH peak between 5 and 30 mIU/L, or were not reported (12;13).

The prevalence rates between the studies that only measured basal fT_4 levels varied between 0 and 19% (weighted mean 5%) (1–5;7;10;11;14), and between 5 and 15% (weighted mean 8%) in those that also used TRH (6;8;9;12;13).

The Hypothalamus-Pituitary-Gonadal axis

The hypothalamus-pituitary-gonadal axis deficiency ranged from 0 to 29% (weighted mean 13%) between the studies (Table 5). Basal LH and FSH were measured in all but one study (4). Basal estradiol (E_2 in women) was measured in 9 studies, and the menstrual history was recorded in 10 out of 14 studies. Testosterone (in men) was measured in all studies. In four studies, a GnRH stimulation test was performed in a subset of the patients (6;8;9;13). The criterion for a normal test response differed between the studies (Suppl Table 5). The definition of secondary hypogonadism was mainly based on basal testosterone (in men) and E_2 concentrations (in women) below the reference ranges, in the presence of decreased or normal LH and FSH levels. A subset of the studies also incorporated the GnRH test result (see above) and menstrual cycle abnormalities in premenopausal females.

Prolactin

The prevalence of abnormal serum PRL concentrations ranged from 0 to 16% (Suppl Table 6). Abnormal PRL secretion was defined as hyperprolactinemia (8/14 studies) (1;2;5;7;9;11;12;14), hypoprolactinemia

(one study) (6), or both (3). In accordance, prevalence rates were between 3 and 12% using the definition of hyperprolactinemia, 0% using the definition of hypoprolactinemia, and 16% using the combination of both. Out of the 14 studies, 10 measured basal serum PRL concentrations only (1–3;5;7;9–12;14). Three studies also used a TRH test (doses 100 and 500 μ g respectively) (6;8;13). Prevalence rates were not reported in two of these (8;13) and were 0% in the third (6).

Discussion

This review demonstrates that the endocrine evaluations and definitions of hypopituitarism differ considerably among the studies that have assessed TBI-related hypopituitarism. From the existing literature, the notion emerges that most of the tests that are currently used to establish the diagnosis of hypopituitarism in general, and GHD in specific, are not validated sufficiently regarding cut-off values, reproducibility, and dependence on confounding factors in TBI patients.

In general, there are hardly any data on reproducibility of tests or dependence on confounding factors in TBI patients. One factor that comes forward in the current review is the potential effect of increased BMI, which in general is associated with decreased GH responses to GH stimulation tests. Therefore, increased BMI may result in an inadvertently higher incidence rate of GHD, if the cut-off values for normal GH responses to GH stimulation tests are not adapted according to BMI. All these methodological issues limit the applicability of the individual studies, *i.e.* the decision whether the study results are valid for patients to whom the results are generalizable but who are subjected to a different endocrine diagnostic assessment than the original study population. Moreover, these methodological limitations prohibit simple generalizations of the results from the perspective of a meta-analysis or a review.

The question arises whether post-traumatic hypopituitarism, especially GHD, has been overdiagnosed on the basis of the older cross-sectional studies. Consensus guidelines for the evaluation of adult GHD state that different dynamic tests can be used to diagnose GHD, including the ITT, the glucagon stimulation test, the combined GHRH-arginine test, and the combined GHRH-GHRP6 test (20). The present assessment, however, documents a higher prevalence of GHD for the glucagon stimulation test and the combined GHRH-GHRP6 test, compared to the results of the combined GHRH-arginine test and the ITT. With the exception of the ITT (which was used in only 12%

of the patients (6;7)), the outcome of each test varied greatly (Figure 2) using different (glucagon stimulation test) or similar cut-off levels (other test). In addition, the studies that used two dynamic tests to assess GH reserve revealed a lower prevalence of GHD than the studies with only one test. Moreover, the results of GH stimulation tests are confounded by BMI, with higher BMI being associated with decreased GH responses. Although BMI-adjusted reference values have been reported (21), none of the studies on TBI-associated GHD reports adjusted their cut-off values for BMI. Moreover, the data indicate that BMI tends to be higher in the TBI patients with GHD (Figure 1). Finally, an important aspect is that most patients had only GHD or one additional pituitary hormone deficiency. Therefore, one test may not be sufficiently reliable and the use of two tests would increase the confidence in the diagnosis, although only if the two tests yield concordant results. However, the application of two tests may introduce an even greater uncertainty in case of discordant results. This discrepancy has been documented for instance in GHD in irradiated patients, in whom the attenuation in GH responses to the ITT was greater compared with the combined GHRH-arginine test (22). These factors impose major problems for an accurate assessment of GHD in these patients. Therefore, these methodological issues have contributed to the suspicion that GHD is probably over diagnosed in the older crosssectional studies.

We observed similar variations in test results of the pituitary-adrenal axis (Figure 3). The use of different tests with different cut-off values resulted in prevalence rates that varied between 5 and 19%. ACTH deficiency can be diagnosed by measuring basal early morning cortisol levels: values below 100 nmol/L are indicative of ACTH deficiency, whereas cortisol values above 500 nmol/L essentially exclude ACTH deficiency. The ACTH stimulation test is reliable in diagnosing clinically significant adrenal insufficiency in patients who are at risk (23-25). ACTH stimulation tests, however, are not fully reliable in excluding the presence of mild secondary adrenal insufficiency (26). The ITT still remains the golden standard, and has the advantage that ACTH/cortisol and GH secretory reserve can be assessed simultaneously. If an ITT is contraindicated, a CRH test can be alternatively used (27). The effect of the initial choice for a specific stimulation test on the variation in outcome of adrenal insufficiency and GHD based on the available data after TBI is illustrated in Figure 4.

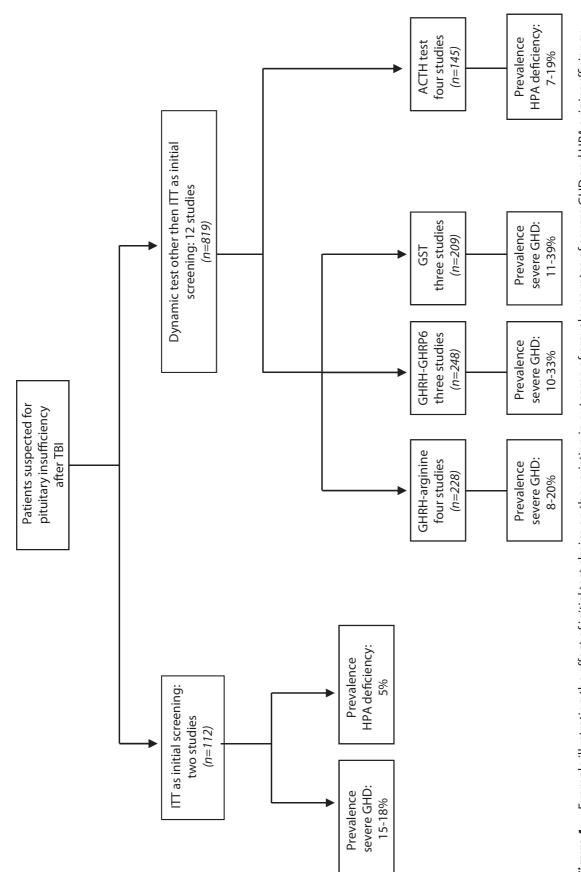


Figure 4. Example illustrating the effect of initial test choice on the variation in outcome of prevalence rates of severe GHD and HPA axis insufficiency.

CHAPTER 2

The diagnosis of secondary hypothyroidism is usually made based on fT₄ values. However, basal fT₄ levels show a relatively small intraindividual variability, although inter-individual variability is large (28). As a consequence, a diagnosis of possible secondary hypothyroidism may not be straightforward, since fT_4 levels within the normal reference range can reflect hypothyroidism in one patient but euthyroidism in another patient. Basal TSH levels are also of limited help for the diagnosis of secondary hypothyrodism, since normal or even increased levels of TSH can be found (29). In addition, a TRH test is of limited value because patients with central hypothyroidism may show different patterns of TSH responses to TRH, with absent or exaggerated responses, which considerably overlap with those found in healthy volunteers. Moreover, the magnitude of the TSH peak is proportional to the injected TRH dose, is higher in women, and tends to decline with age (30). In accordance, the prevalence rates were not affected by the use of TRH stimulation. In analogy, the interpretation of the GnRH test is complex, and individual responses vary greatly in both adults and children (31). In men, it is sufficient to measure non-stimulated LH, FSH, and testosterone concentrations. In premenopausal women, the evaluation of the menstrual cycle is a prerequisite, whereas in postmenopausal women, the absence of increased LH and FSH levels almost invariably indicates hypogonadotropic hypogonadism.

Analytical factors will most likely also have affected the different outcomes of the studies. For instance, the GH and cortisol assays varied between studies, and it is known that the between-laboratory performance of the GH assay is not very good. Moreover, most were not validated sufficiently regarding normal cut-off values, reproducibility, and dependence on confounding factors even in a 'normal' population. None of these tests have been validated in TBI patients at all.

The time point of evaluation may also influence outcome; therefore, we focused only on studies in the chronic phase after TBI, *i.e.* one year after the trauma. Studies that analyzed patients with a median duration of 12 months after TBI, however, were also included. Thus, part of these patients was assessed within 12 months after TBI. In general, the transient effects of TBI mimicking pituitary insufficiency are almost exclusively reported only within the first six months after TBI (15). Therefore, it is unlikely that the pituitary results of the studies with a median duration of follow-up of 12 months of TBI are caused by the transient effects of

TBI. This is supported by similar results of additional analyses of the remaining studies, which included only patients with a follow-up of more than 12 months after TBI. Lastly, the underlying mechanisms of TBI-related hypopituitarism have not been resolved. It is unclear to that extent hypothalamic versus pituitary damage is present in TBI patients with hypopituitarism and what impact these processes may have on endocrine tests.

Recently, many clinical reviews have summarized the studies on pituitary insufficiency after TBI (15;16). These studies concluded that hypopituitarism is a common complication of TBI and might contribute to morbidity and poor recovery after brain injury (16). However, these reviews did not take into account the variability in diagnostic strategies and definitions of pituitary insufficiency. These discrepancies, in addition to differences in inclusion and exclusion criteria, limit the possibility to compare the results of studies on TBI. We agree with Klose and Feldt-Rasmussen that future studies should be designed to ensure a high diagnostic robustness for proper identification of reliable predictors, as the results may be highly dependent on diagnostic pitfalls (15).

In conclusion, the reported prevalence rates of pituitary insufficiency after TBI vary considerably, which is associated with major differences in endocrine and analytical methods of assessment and definitions used for hypopituitarism. This does not only apply to the case of TBIrelated hypopituitarism, but most likely also to hypopituitarism caused by pituitary diseases. The same caution with respect to the evaluation of pituitary function should be considered in pituitary diseases, because the diagnosis of definitive hypopituitarism remains a challenge in clinical endocrinology. In pituitary pathology, definitive data on robust accuracy of basal or dynamic hormonal tests are incomplete.

		Basal	Basal		GHRH-	Other	Criterion	Criterion GH deficiency		
	No. of	serum serum	serum		arginine	dynamic			Assays	Outcome
Study	patients	НD	I-191	Ħ	test	test	НЭ	IGF-I	GH / IGF-I	(%)
Kelly <i>et al.</i> (6)	22	+	+	+ (n=22)	1	I	GH < 95% CL according to AUC ^a	16–24 yr: 182–780 μg/L 25–39 yr: 114–492 μg/L 40–54 yr: 90–360 μg/L	IFA/RIA*	18 ^b
Lieberman <i>et al.</i> (9)	70	+	+	I	I	+ ^c (GST; n=48)	Peak GH < 3 μg/L	NR	IRMA/RIA*	15
Bondanelli <i>et al.</i> (3)	20	+	+	I	+ (n=50)	I	Peak GH < 16.5 μg/L Severe GHD = GH < 9 μg/L Partial GHD = GH 9–16.5 μg/L	20–30 yr : 135–485 μg/L IRMA/RIA Partial:28 31–40 yr: 120–397 μg/L Severe:8 41–50 yr: 113–306 μg/L 51–60 yr: 100–250μg/L >60 yr: 92–229 μg/L	IRMA/RIA	Partial:28 Severe:8
Agha <i>et al.</i> (1)	102 ^d	+	+	+ (n=14)	+ (n=4)	+° (GST; n=102)	GST: peak GH < 5 μg/L ITT: peak GH < 5 μg/L GHRH-arg: peak GH < 9 μg/L	IGF-I SDS: ln(IGF-I) – [5.9 –(0.0146xage in yrs)]/0.272	IRMA/ IRMA	1

Supplemental data **Table 2.** GH-IGF-I Axis

No. ofserumarginitedynamicCriterion GH deficiencyAssayspatientsGHIGF1ITTtest GH GH $IGF-1$ $GH/IGF-1$ $GH/IGF-1$ 67 +++Severe GHD=Normal levels matched RM/ RIA 67 +++Severe GHD=Normal levels matched RM/ RIA 67 +++Severe GHD=Normal levels matched RM/ RIA 1 70+++Severe GHD=Normal levels matched RM/ RIA 1 70++++Severe GHD= $11-35$ nmo//. $11-35$ nmo//. 1 70++++- $11-35$ nmo//. $11-35$ nmo//. 1 70+++ $100-200$ µg/L $11-35$ nmo//. $11-35$ nmo//. 1 70+++ $1-35$ nmo//. $100-200$ µg/L $11-35$ nmo//. 1 70++++ $--165 µg/L100-200 µg/L100-200 µg/L110-3510-35 µg/L10-35 µg/L10-35 µg/L10-35 µg/L10-35 µg/L170+++---110-3210-36 µg/L10-30 µg/L10-36 µg/L10-36 µg/L110-3210-32 µg/L10-36 µg/L$	Popovic										
patientsGFITtest GH IT $IGF-I$ $GH/IGF-I$ 67 ++++Severe GHD =Normal levels matched $RMA \ RIA$ 67 ++++Severe GHD =Normal levels matched $RMA \ RIA$ 70 ++++Severe GHD =Normal levels matched $RMA \ RIA$ 70 +++-Severe GHD =Normal levels matched $RMA \ RIA$ 70 +++-Normal levels matched $RMA \ RIA$ 70 +++Normal levels matched $RMA \ RIA$ 70 ++++ $-$ Normal levels matched $RMA \ RIA$ 70 ++++ $-$ Normal levels matched $RMA \ RIA$ 70 ++++ $ 70$ ++++ $ 70$ +++ $ 70$ +++ $ 70$ +++ $ 70$ +++ $ -$	Popovic	No. of		serum		arginine	dynamic	Criterion	ו GH deficiency	Assays	Outcome
67 + + - + + Esere GHD Normal levels matched IRMA/ RIA 1 2 1 1 2 1 3 1 3 1 3 1 3 1 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3	Popovic	patients		IGF-I	Ξ	test	test	НЭ	IGF-I	GH / IGF-I	(%)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1011- +-	67	+	+	I	I	+	Severe GHD =	Normal levels matched	IRMA/ RIA	Partial: 30
ti 70 + + - + + - + + $^{-67}$ $^{-10.0}_{-20.0}$ $^{-10.3}_{-10.2}$ $^{-10.3}_{-10.3}$ $^{-10.3}_{-10.4}$ ro 99 + + + + + + + + + + $^{-1}$ $^{-16.5}_{-10.5}$ $^{-10.5}_{-10.4}$ $^{-10.3}_{-10.4}$ $^{-10.4}_{-10.4}$ $^{$	er al.(10)						(GHRH-	peak GH < 10.0	for age and sex		Severe: 15
ti 70 + + + - + + - + + \cdot Peak GH 25 th centile age-related IRMA/RIA ro 99 + + + + + + \cdot Peak GH 25 th contal limits Severe GHD = peak GH 0. pg/L normal limits Peak GH 0. pg/L 10 pg/L (±2505) g/L IRMA/ GHRP6; G5T and ITT: GH 0. dg/L (±2505) g/L IRMA/ di 52 + + + $\frac{1}{2}$ GH 0. g/L (±2505) g/L IRMA/ di 52 + + + $\frac{1}{2}$ GH 0. g/L Age-dependent 5D5 th CLA/CLA di 52 + + + $\frac{1}{2}$ GH 0. g/L Age-dependent 5D5 th CLA/CLA di 52 + + + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA di 52 + + + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA di 52 + + + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA di 52 + + + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA GHRP6; G5 th GHRP6; GFI + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA GHRP6; GFI + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA GHRP6; GFI + $\frac{1}{2}$ GH 0. g/L ITT: GH Age-dependent 5D5 th CLA/CLA							GHRP6;	hg/L hg/L	11–35 nmol/L		
ti 70 + + - + - + - + - Pak GH 25 th centile age-related IRMA/RIA $(n=70)$ $< 16.5 \mu g/L$ normal limits $(n=70)$ $< 16.5 \mu g/L$ normal limits $(n=70)$ $< 16.5 \mu g/L$ normal limits $(n=30)$ $(n=30)$ $(n=30)$ $(GHRH - GHRPG: IGF-I < 200 \mu g/L$ IRMA $(n=30)$ $(GHRH - GHR - GHD = 10 \mu g/L$ $(=2500)$ $(=2500)$ $(=2500)$ $(=260)$							n=67)	чп = реак чп < 10.0–20.0 µg/L			
ro 99 + + + + + + $^{(n=70)}$ $< 16.5 \mu g/L$ normal limits Severe GHD = $peak GH < 9.0 \mu g/L$ peak GH < 9.0 $\mu g/L$ (n=35) (GHRH- peak GH < 10 $\mu g/L$ (EF-I < 200 $\mu g/L$ IRMA* (n=32) 3 $\mu g/L$ (GHRH- peak GH < 10 $\mu g/L$ (±25D5) (RMA* GHRP6; G5T and ITT: GH n=44) 3 $\mu g/L$ (GHRH- peak GH < 10 $\mu g/L$ (±25D5) (CIA/CIA di 52 + + + - + + GH < 10 $\mu g/L$ Age-dependent SD5 ^h CLA/CIA di 52 + + + + GH < 10 $\mu g/L$ (GF-I < 84 $\mu g/L$ IRMA* GHRP6; GF-I < 200 $\mu g/L$ (GF-I < 84 $\mu g/L$ IRMA*	Aimaretti	70	+	+	I	+	I	Peak GH	25 th centile age-related	IRMA/RIA	Partial: 39
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	<i>et al.</i> (2)					(n=70)		< 16.5 µg/L Severe GHD =	normal limits		Severe: 20
$ \begin{array}{lcccccccccccccccccccccccccccccccccccc$								реак GH < 9.0 µg/L			
er 70 + + - + + - + $^{(n=35)}$ (GHRH- peak GH $\leq 10 \mu g/L$ ($\pm 2SDS$) IRMA* GHRP6; GST and ITT: GH n=44) n=46) $n=6$ ng/L IGF-I < 84 $\mu g/L$ IRMA/ IRMA* GHRP6; n=50)	Leal-Cerro	66	+	+	+	+	ъ +	GHRH-GHRP6:	IGF-I < 200 µg/L	IRMA/	10
70 + + + - + $^{+9}$ - GH c ST and ITT: GH 70 + + + - + $^{+9}$ - GH < 9.0 µg/L Age-dependent SDS ^h CLA/CLA (n=32) 52 + + + GH < 10 µg/L IGF-I < 84 µg/L IRMA/ (GHRH- GH RP6; n=52)	<i>et al.</i> (8)				(n=35)		(GHRH-	peak GH ≤ 10 μg/L	(±2SDS)	IRMA*	
70 + + - + 9 - GH < 9.0 µg/L Age-dependent SDS ^h CLA/CLA (n=32) 52 + + + 9 - 9 GH < 10 µg/L 1 IGF-I < 84 µg/L 1 IRMA' GHRP6; n=50)						U	GHRP6; GST	- GST and ITT: GH			
$\begin{array}{cccccccccccccccccccccccccccccccccccc$							n=44)	3 µg/L			
52 + + + GH < 10 μg/L IGF-I < 84 μg/L IRMA/ (GHRH- GHRP6; n=52)	Schneider	70	+	+	I	6+	Ι	GH < 9.0 µg/L	Age-dependent SDS ^h	CLA/CLA	10
52 + + + GH < 10 μg/L IGF-I < 84 μg/L IRMA/ (GHRH- GHRP6; n=52)	<i>et al.</i> (11)					(n=32)					
(GHRH- GHRP6; n=52)	Tanriverdi	52	+	+	I	I	+	GH < 10 μg/L	IGF-I < 84 μg/L	IRMA/	33
GHRP6; n=52)	et al.(12)						(GHRH-			IRMA*	
n=52)							GHRP6;				
							n=52)				

Table 2. Continued

		Basal	Basal		GHRH-	Other				
	No. of	serum	serum serum		arginine	dynamic	Criterion	Criterion GH deficiency	Assays	Outcome
Study	patients	НÐ	I-191	E	test	test	НЭ	IGF-I	GH / IGF-I	(%)
Herrmann	76	+	+	+	+	I	GHRH-arg:	16–24 yr: 182–780 µg/L	CLA/IRMA	8
<i>et al.</i> (5)				(u=7)	(n=76)		GH < 9 µg/L	25–39yr: 114–492 µg/L		
							ITT: GH peak	40–54 yr: 90–360 µg/L		
							<3 µg/L	≥ 55 yr: 71–290 µg/L		
Bushnik	64	+	+	I	I	+	Severe GHD =	IGF-l age-corrected	NR	Partial: 66
<i>et al.</i> (4)						(GST; n=59)	(GST; n=59) ⁱ peak GH < 3 μg/L	normal levels		Severe: 39
							Moderate: peak			
							GH 3 –9.9 µg/L			
Klose <i>et al.</i>	104	+	+	+	+	I	Severe GHD =	IGF-I SDS ^k	FIA/RIA	15
(2)				(06=u)	(n=14)		ITT: peak			
							GH< 3 µg/L			
							GHRH-arg:			
							GH< 9 µg/L			
							Partial GHD: =			
							ITT: peak GH			
							5 µg/L ≤ GH			
							≥ 3 μg/L			
							GHRH-arg: peak			
							16.5 µg/L ≤ GH			
							≥ 9 μg/L			
Tanriverdi	30	+	+	I	I	+	GRHR-GHRP6:	18–30 yr: 197–476 µg/L	IRMA/	23
<i>et al.</i> (14)						(GRHR-	severe GHD = GH	31–40 yr: 100–494 µg/L	IRMA*	
						GHRP6	< 10 µg/L	41–70 yr: 101–303 µg/L		
						n-30.6ST	GST: GH			
							< 1.18 µg/L			
						n=/)"				

lable 2.	Continued	Basal	Basal		GHRH-	Other						
	No. of	serum	serum		arginine	dynamic		Criterio	Criterion GH deficiency	iency	Assays	Outcome
Study	patients	НÐ	I-191	E	test	test	•	ВН		I-JDI	GH / IGF-I	(%)
Wachter <i>et al.</i> (13)	55	+	+	1	1	+ (GHRH; n=NR) ^m	NR		Age adjusted: 116–270 μg/L	sted: µg/L	NR	5
ACS, Autom growth horr hormone pl insulin toler a Normal val b 34 patients c GST in n=4 d Cut-off valu d Cut-off valu b 34 patients wi f Patients wi GST: 0.03 m c GST: 0.03 m b Calculated i Patients wi m Hormonal reported ho reported ho reported ho	ACS, Automated chemiluminescence system; AUC, area under the curve; CL, confidence limit; CLA, chemiluminometric as growth hormone deficiency; GHI, growth hormone releasing hormone; GHRH-GHR hormone plus growth hormone releasing peptide 6; GST, glucagon stimulation test; IGF-I, insulin-like growth factor I; IR ^I insulin tolerance test; NR, not reported; RIA, radio immunometric assay Normal values defined by a group of healthy subjects (n=18) * So are a sufficiency; GHI (n=14) * Grooff values defined by a group of healthy subjects (n=18) * Grooff values defined by a group of healthy subjects (n=18) * Grooff values defined by a group of healthy subjects (n=18) * Grooff values defined by 31 healthy control subjects * All patients underwent GST; patients with GHRH-GHRP (n=44, glucagon stimulation test (n=44) * All patients underwent GST; patients with GHRH-GHRP (n=44, glucagon stimulation test (n=44) * All patients underwent GST; patients with GHRH-GHRP (n=44, glucagon stimulation test (n=44) * Calculated according to Brabant <i>et al.</i> 2003(31) * GST: 0.03 mg kg-1 max 1 mg intramuscular * Calculated according to Brabant <i>et al.</i> 2003(31) * GST: 0.03 mg kg-1 max 1 mg intramuscular * Calculated by Juul <i>et al.</i> 1994(32) * Galculated by Juul <i>et al.</i> 1994(32) * Hormonal stimulation tests. GHRH: 100 µg * Calculated by Juul <i>et al.</i> 1994(32) * Mormonal stimulation tests were performed if abnormalities in basal hormone screening or if patients answered 's * After acid-ethanol extraction * After acid-ethanol extraction * After acid-ethanol extraction	minescenk rcy; GHI, g rmone rel not repor y a group d been tes rwent L-dk 35T; patier G Brabant <i>et</i> mg intrar d healthy 1994(32) evels of G evels of G rests were nts under ction	ce system rowth hor easing pe rted; RIA, r of health, ited < 12 r opa test hy control nts with al nuscular controls a controls a r af.203(3) nuscular controls a r af.203(3) nuscular controls a r af.203(3) nuscular controls a	; AUC, ar rmone ir radio im y subjec months p nunder ist to tes ist to tes is	rea under the curve; nsufficiency; GRHR, <u>G</u> (GST, glucagon stimu imunometric assay its (n=18) post TBI (7,5,5 month post TBI (7,5,5 month in results under went d GHRH-GHRP6 (n=44, st appropriatness of st appropriatness of st appropriaties (n=44, grapher (n=18) in test, ghr (n=19, n) onormalities in basa i tests. GHRH: 100 µg	e curve; CL, c GRHR, grow on stimulatik : assay 5 months) 5 months) 6 (n=44,gluc ness of cut-o ness of cut-o ry testing ry testing ry testing in basal hor : 100 µg	onfidence li th hormone on test; IGF- agon stimu off values underwent mone scree	imit; CLA, c e releasing -l, insulin-li lation test GST (n=7) ening or if	hemilumino hormone; GF ke growth fa st (n=4) (n=44) and I ⁻ patients an	metric assay; HRH-GHRP6 te ctor l; IRMA, i TT (n=35) swered 'yes' t	ACS, Automated chemiluminescence system; AUC, area under the curve; CL, confidence limit; CLA, chemiluminometric assay; FIA, fluoroimmunoassay; GHD, growth hormone deficiency; GHI, growth hormone insufficiency; GRHR, growth hormone; GHRH-GHRP6 test, growth hormone releasing profite 6: GST, glucagon stimulation test; IGF-1, insulin-like growth factor 1; IRMA, immunoradiometric assay; ITT, insulin tolerance test; NR, not reported; RN, radio immunometric assay • Normal values defined by a group of healthy subjects (i=18) • Somal values defined by a group of healthy subjects (i=18) • CGST in n=48; n=20 undervent L-dopa test • Cuch of values defined by 31 healthy control subjects • All paients with GFI-1 in lower range were tested < 12 months joint (i=14) or GHRH-arg test (n=4) • Cuch of values defined by 31 healthy control subjects • All paients undervent GST; patients with abnormal results undervent ITT (n=14) or GHRH-arg test (n=4) • Cuch of values defined by 31 healthy control subjects • All patients undervent GST; patients with abnormal results undervent ITT (n=15) • Sale baselity, controls undervent GHRH-arg test to test appropriatness of cut-off values • Calculated according to Brabant <i>et al.</i> 2003(31) • CGST on mdervent GHRH-arg test to test appropriatness of cut-off values • Calculated according to Brabant <i>et al.</i> 2003(31) • Calculated according to Brabant <i>et al.</i> 1994(32) • Calculated by Juul <i>et al.</i> 1994(32) • Alter acid-ethanol extraction • After acid-ethanol extraction	oassay; GHD, one releasing ric assay; ITT, onnaire. Not

			Basal				Other		Assay	
	No. of	No. of Basal serum serum	serum		ACTH		dynamic	Criterion for ACTH-	cortisol/	Outcome
Study	patients	s cortisol	ACTH	CRH test	test	ΤT	test	deficiency	ACTH	(%)
Kelly <i>et al.</i>	22	+	+	1	I	+	1	Cortisol < 95% Cl according	RIA/IRMA	5
Lieberman	70	+	I	I	+ 1	- (77=U)	I	Peak cortisol < 500 nmol/L	MSA/-	7
Bondanelli	50	+	+	I	- (0/=u)	I	I	Cortisol and ACTH below	RIA/IRMA	0
et al. (3)								reported reference range (cortisol: 220 – 700 nmol/L ACTH: 1.5–11.5 pmol/L)		
Agha <i>et al.</i> (1)	102 °	+	+	I	+ (n=8) ^d	+ (n=15)	+ (GST; n=102)	Failing 2 tests: GST: peak cortisol < 450 nmol/L ITT: peak cortisol < 500 nmol/L SST: peak cortisol	FIA/IRMA	13
Popovic et al.(10)	67	+	I	I	I	I	I	<500 nmol/L NR	RIA/-	٢
Aimaretti <i>et al.</i> (2)	70	+	I	I	I	I	I	Cortisol < 220 nmol/L 24h urinary free cortisol < 30 µg/24 h	RIA/-	σ

Table 3. Hypothalamic-Pituitary Adrenal Axis

			Basal				Other		Assav	
	No. of	No. of Basal serum serum	serum	CRH	ACTH		dynamic	Criterion for ACTH-	cortisol/	Outcome
Study	patients	cortisol	ACTH	test	test	ΤTI	test	deficiency	ACTH	(%)
Leal-Cerro et al.(8)	66	+	+	1	I	+ ^f (n=NR)	1	Basal cortisol < 200 nmol/L Peak cortisol < 550 nmol/L Peak ACTH < 6.6 pmol/L	FIA / IRMA	1
Schneider <i>et al.</i> (11)	70	+	I	I	+ (n=33) ^g	I	I	Cortisol < 500 nmol/L	ECLIA/-	6
Tanriverdi <i>et al.</i> (12)	52	+	+	I	+ (n=12) ^h	I	I	Basal cortisol < 193 nmol/L ACTH test: Cortisol < 552 nmol/L	RIA/IRMA	19
Herrmann <i>et al.</i> (5)	76	+	+	Ι	I	+ (n=7) ⁱ	I	Cortisol and ACTH below reported reference range (cortisol 180–640 nmol/L; ACTH 3.74– 11.44 pmol/L) ITT: Peak cortisol < 500 nmol/L	IA/CL	m
Bushnik <i>et al.</i> (4)	64	+	I	I	I	I	I	Fasting serum cortisol < 440 nmol/L	NR	60
Klose <i>et al.</i> (7)	104	+	+	I	+ + + (n=14) ^k (n=90)	+ (06=u)	I	Peak cortisol < 500 nmol/L	ECLIA /-	Ŋ
Tanriverdi <i>et al.</i> (14)	30	+	+	I	+ (n=3) ¹	I	I	Basal cortisol < 193 nmol/L ACTH test: Cortisol < 550 nmol/L	RIA/IRMA	2

CHAPTER 2

60

Table 3. Continued

	me	~		netric st; IA, s; NR, s; NN,
	Outcome	(%)	4	iluminor Lation te cion assay stionnaire
Assay	cortisol/	ACTH	NR	CLA, chem agon stimu tic separat ol/L ecific ques
	Criterion for ACTH-	deficiency	Cortisol below reported reference range (cortisol (16:00 – 20:00 h): 63 – 339 nmol/L)	ACS, Automated chemiluminescence system; ACTH, adrenal corticotope hormone; AUC, area under the curve; CL, confidence limit; CLA, chemiluminometric assay; IRH, munuoassay; IRMA, immunoradiometric assay; ITT, insulin tolerance test; MEIA, microparticle enzyme immunoassay; GST, glucagon stimulation test; IA, fluoroimmunoassay; IRMA, immunoradiometric assay; TT, insulin tolerance test; MEIA, microparticle enzyme immunoassay; MSA, magnetic separation assays; NR, not reported; RIA, radio immunometric assay; STT, short synacthen stimulation test ACTH test: 250 ug cosyntropin • ACTH test: 250 ug Synacthen • Curcle fix vith subnormal response to GST (n=23) underwent ITT (n=15) or SST (n=8) • ACTH test: 250 ug Synacthen • Patients with subnormal basal serum cortisol and ACTH values underwent ITT; not reported how many patients cortisol < 200nmol/L. • ACTH test: 250 ug Synacthen • Patients with subnormal basal serum cortisol and ACTH values underwent ITT; not reported how many patients cortisol < 200nmol/L. • ACTH test: 1.05 Synacthen • ACTH test: 2.00 ug Synacthen • ACTH test: 1.05 Synacthen • ACTH test: 1.06 Synacthen • ACTH test: 1.06 Synacthen • ACTH test: 1.06 Synacthen • ACTH test: 1.06 Synacthen • ACTH test: 2.06 ug Synacthen • ACTH test: 1.06 Synacthen • ACTH test: 2.06 ug
Other	dynamic	test	I	e; AUC, area u mmunoassay; microparticle, ist =8) ot reported h =7) =7) ne screening ne screening cotrope releas
		E	I	: hormon scence ir t; MEIA, r or SST (n ent ITT; n ng ng ng ng cortic
	ACTH	test	I	rticotrope emilumini erance tes then stimu s underwe itary testi es in basa es in basa
	CRH	test	+ + n=NR)	adrenal co electroch insulin tol nort synac cts (n=18) .cTH value CTH value rwent pitu onormaliti i tests; CRH
Basal	serum	ACTH	+	em; ACTH, ne; ECLIA, assay; ITT, assay; SST, sh lthy subject rol subject [(n=23) ur f (n=23) ur tisol and A ls all unde ls all unde rrmed if al
	No. of Basal serum serum	cortisol	+	ACS, Automated chemiluminescence system; ACTH, adrenal corticotrope hormone; AUC, area under the curv assay; CRH, corticotrope releasing hormone; ECLIA, electrochemiluminiscence immunoassay; IRMA, immunoradiometric assay; STT, insulin tolerance test; MEIA, microparticle enzyme immunor treported; RIA, radio immunometric assay; SST, short synacthen stimulation test ^a Normal values defined by a group of healthy subjects (n=18) ^b ACTH test: 250 µg cosyntropin ^c Cut-off values defined by 31 healthy control subjects ^d ACTH test: 250 µg Synacthen ^e Patients with subnormal response to GST (n=23) underwent ITT (n=15) or SST (n=8) ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT; not reported how many pati ^g ACTH test: 250 µg Synacthen ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^g ACTH test: 250 µg Synacthen ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^g ACTH test: 1 µg Synacthen ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^f Patients with subnormal basal serum cortisol and ACTH values underwent ITT (n=7) ^f ACTH test: 1 µg Synacthen ^f ACTH test: 1
	No. of	patients	55	ACS, Automated chemiluminesc assay; CRH, corticotrope releasi Immunoassay; IRMA, immunora not reported; RIA, radio immuno a Normal values defined by a gro b ACTH test: 250 µg Synacthen cCut-off values defined by 31 he. dACTH test: 250 µg Synacthen e Patients with subnormal basal. 9ACTH test: 1 µg Synacthen hACTH test: 250 µg Synacthen f Patients with subnormal basal. 9ACTH test: 250 µg Synacthen hACTH test: 250 µg Synacthen hACTH test: 250 µg Synacthen n hACTH test: 250 µg Synacthen reported how many patients un reported how many patients un
		Study	Wachter <i>et al.</i> (13)	ACS, Automa assay; CRH, c Immunoassa not reported ^a Normal valu ^b ACTH test: 2 ^c Cut-off valu ^d ACTH test: 2 ^d ACTH test: 2 ^h ACTH test: 1 ¹ Patients with ^f ACTH test: 1 ^m Hormonal reported hov

				Basal				
	No. of	Basal	Basal	serum			Assay	Outcome
Study	patients	serum fT	serum fT $_3$ serum fT $_4$	TSH	TRH	Criterion TH deficiency	TSH/FT₄	(%)
Kelly <i>et al.</i> (6)	22	1	+	+	+	TSH < 95% CL according to AUC ^b	ICL/ RIA	5
					(n=22) ^a			
Lieberman	70	I	+	+	+	TSH: 0.49–4.7 mIU/L	MPIA/	15
et al.(9)					(n=27) ^c	fT₄: 9.7 – 23.8 pmol/L Insufficient ↑ TSH (5–30 mlU/L); peak TSH not within 60 min	MPIA	
Bondanelli <i>et al.</i> (3)	50	I	+	+	I	Low serum fT $_4$ with normal or low serum TSH	ACS/ACS	10
						TSH: 0.4–4.2 mIU/L fT _a : 10.3–19.4 pmol/L		
Agha <i>et al.</i> (1)	102 ^d	I	+	+	I	Low fT ₄ without elevation TSH TSH: 0.5 –4.2 mlU/L fT ₄ : 8–21 pmol/L	FIA/FIA	-
Popovic <i>et al.</i> (10)	67	I	+	+	I	NR	IRMA/RIA	4
Aimaretti <i>et al.</i> (2)	70	I	+	+	I	${ m fT}_4$ < 10.29 pmol/L with normal or low TSH	IRMA/RIA	Q
Leal-Cerro <i>et al.</i> (8)	66	I	+	+	+ (n=NR) ^e	fT₄ ≤ 7.74 pmol/L with normal or low TSH TRH test: TSH peak ≤ 7 mlU/L	RIA/RIA	Q

CHAPTER 2

Table 4. Hypothalamus Pituitary Thyroid Axis

			Basal	Basal				
	No. of	Basal	serum	serum			Assay	Outcome
Study	patients	serum fT	fT₄	TSH	TRH	Criterion TH deficiency	TSH/FT4	(%)
Schneider et al.(11)	70	1	+	+	I	${ m fT}_4$ < 11.97 pmol/L; TSH not elevated	ECLIA/ ECLIA	4
Tanriverdi <i>et al.</i> (12)	52	+	+	+	+ (n=NR) ^f	fT ₄ < 10.3 pmol/L without appropriate elevation TSH	fT₃, fT₄: RIA TSH: IRMA	9
Herrmann <i>et al.</i> (5)	76	+	+	+	I	Low serum fT ₄ without appropriate elevation in serum TSH TSH: 0.3–3.0 mIU/L fT ₄ : 10–25 pmol/L	IA/IA	m
Bushnik <i>et al.</i> (4)	64	I	+	+	I	Low serum fT $_4$ with low or normal TSH Normal reference values: NR	NR	19
Klose <i>et al.</i> (7)	1049	I	+	+	I	Subnormal fT $_4$ (<12 pmol/L) with inappropriate low TSH	ECLIA/ ECLIA	7
Tanriverdi <i>et al.</i> (14)	30	+	+	+	I	fT ₄ < 10.3 pmol/L without appropriate elevation TSH	fT₃, fT₄: RIA TSH: IRMA	0

	No. of	Basal	Basal serum	Basal serum			Assay	Outcome
Study	patients	serum fT	fT_	TSH	TRH	Criterion TH deficiency	TSH/FT4	(%)
Wachter <i>et al.</i> (13)	55	1	+	+	+ (n=NR) ^h	 + Criterion for TRH test: NR (n=NR)^h fT₄: 12–22 pmol/L TSH: NR 	NR	9
ACS, Automated chemiluminescence system; AUC, area under the curve; CL, confide Fluoroimmunoassay; fT ₃ : free triiodothyronine; fT ₄ , free thyroxine; IRMA, immuno radiome enzyme immunoassay; NR, not reported; RIA, radio immunoassay; TRH, thyroid releasing ho ^a TRH test: 500 µg ^b Normal values defined by a group of healthy subjects (n=18) ^b Normal TSH and fT ₄ values underwent TRH stimulation test (n=27); 0.5 mg TRH; IV, TSH a ^d Cut-off values defined by 31 healthy control subjects ^e TRH test: 500 µg Protirelin ^f Not reported how many patients underwent TRH testing and how the test was performed ^g 30 age- and BMI-matched healthy controls all underwent pituitary testing ^h Hormonal stimulation tests were performed if abnormalities in basal hormone screen reported how many patients underwent stimaltionsts. TRH: 200 µg	d chemilumir ssay; fT₃: free bassay; NR, no g defined by a g and fT₄ values lefined by 31 t protirelin w many patie l-matched he ulation tests any patients u	rescence systuriodothyroni it reported; RI/ it reported; RI/ underwent TF healthy contro ints underwen althy controls were perform underwent stir	em; AUC, ar ine; fT ₄ , free t A, radio immu Y subjects (n RH stimulatio I subjects all underwer ed if abnorm maltionsts. TF	ea under ti thyroxine; IR unoassay; TR n=18) in test (n=27 and how th nt pituitary t natlities in k 3H: 200 μg	ne curve; MA, immu H, thyroid); 0.5 mg Tl); 0.5 mg Tl); 0.5 mg Tl); 0.5 mg ne test was esting asal horm	ACS. Automated chemiluminescence system; AUC, area under the curve; CL, confidence limit; ECL, Electrochemiluminiscence immunoassay; FlA, Fluoroimmunoassay; fT ₃ ; free triiodothyronine; fT ₄ , free thyroxine; IRMA, immuno radiometric assay; ICL, Immunochemoluminiscense; MPIA, microparticle enzyme immunoassay; NR, not reported; RIA, radio immunoassay; TRH, thyroid releasing hormone; TSH, thyroid stimulating hormone • TBH1est: 500 µg • Normal values defined by a group of healthy subjects (n=18) • Cut-off values defined by a group of healthy subjects (n=18) • Cut-off values defined by a group of healthy subjects (n=18) • Cut-off values defined by a group of healthy control subjects • TBH test: 500 µg • Cut-off values defined by 31 healthy control subjects • Cut-off values defined by 31 healthy control subjects • Cut-off values defined by • TBH test: 500 µg Protirelin • Cut-off values defined by • TBH test: 500 µg Protirelin • Nor reported how many patients underwent TRH testing and how the test was performed • 30 age- and BMI-matched healthy controls all underwent pituitary testing • Hormonal stimulation tests were performed if abnormatilities in basal hormone screening or if patients answered 'yes' to specific questionnaire. Not reported how many patients underwent stimaltionsts. TRH: 200 µg	miniscense; MPIA miniscense; MPIA hormone to specific ques	unoassay; FIA , microparticl stionnaire. No

			Basal	Basal		GnRH			Out-
	No. of	Basal	serum	serum	Menstrual	Stimula-		Assay	come
Study	patients	patients serum LH	FSH	E ₂ /T	history	tion test	Criterion for hypogonadism	LH/FSH/T/E ₂	(%)
Kelly et al. (6) 22	22	+	+	+	+	+	LH and FSH < 95% CL according to AUC ^a	LH, FSH: FIA	23
						(n=18)	F: increase LH <10 IU/L after GnRH	T: RIA	
							administration	E ₂ : direct assay	
							T: 10.3 – 36.2 nmol/L	1	
							${\sf E}_2$: (early follicular phase) 73.4–550.7 pmol/L		
Lieberman	70	+	I	+ L	+	9 4	T below reported reference range	T: RIA	0
<i>et al.</i> (9)						(n=NR)	(9.7 – 30.5 nmol/L)	LH: MEIA	
							LH below reported reference rage		
							(M= 2-12 IU/L)		
							Normal response GnRH test:		
							M: peak LH <12 IU/L and increase in		
							TH < 7 IU/L		
							F: peak LH < 10 IU/L and increase in		
							LH < 5 IU/L		

			Ddbdl	basal					Cur
	No. of	Basal	serum	serum	Menstrual	Stimula-		Assay	come
Study	patients	serum LH	FSH	E_2/T	history	tion test	Criterion for hypogonadism	LH/FSH/T/E ₂	(%)
Bondanelli	50	+	+	+	NR	I	LH below reported reference range:	LH, FSH: CLIA	14
<i>et al.</i> (3)							F: follicular phase 2.5–10 IU/L; menopausal	T, E_2 : RIA	
							40-104 IU/L		
							M: 1–10 IU/L		
							FSH below reported reference range:		
							F: follicular phase 2.5–10 IU/L; menopausal		
							34-96 IU/L		
							M: 1–7 IU/L		
							T below reported reference range		
							(10.1–34.7 nmol/L)		
							E, below reported reference range (follicular		
							phase 74–555 pmol/L; menopausal		
							<180pmol/L)		
Agha <i>et al.</i> (1)	102℃	+	+	+	+	I	M: low T; inappropriately low LH and FSH	LH, FSH: FIA	12
							Premenopausal F: amenorrhea with low E_2		
							without \uparrow LH and FSH		
							Postmenopausal F: LH and FSH in range of		
							premenopausal		
Popovic <i>et al.</i>	67	+	+	T+	+	I	NR	LH, FSH: IRMA	6
(10)								T: RIA	
Aimaretti	70	+	+	+	+	I	M : T < 12.1 nmol/L with low or normal FSH	LH, FSH: IRMA	11
<i>et al.</i> (2)							and LH	T,E_2 : RIA	
							Premenopausal F: menstrual disturbances,		
							low $E_2 < 73.4$ pmol/L with normal or low LH		
							and FSH		

Table 5. Continued

66

CHAPTER 2

			Basal	Basal		GnRH			Out-
	No. of	Basal	serum	serum	Menstrual	Stimula-		Assay	come
Study	patients	serum LH	FSH	E,/T	history	tion test	Criterion for hypogonadism	LH/FSH/T/E ₃	(%)
Leal-Cerro	66	+	+	+	+	+	M: T < ref val with normal LH and FSH	T: RIA	29
<i>et al.</i> (8)						(n=NR)	F: menstrual disorder or amenorrhea after TBI, $\mbox{ E}_{2}$, LH, FSH: FIA $\mbox{ E}_{2}$ <0.08 nmol/L normal or low LH and FSH	E ₂ , LH, FSH: FIA	
Schneider <i>et al.</i> (11)	70	+	+	+	+	I	M: T < 12.1 nmol/ without ↑ LH and FSH Premenopausal F = no menses Postmenopausal F = low FSH, LH	All: ECL	29
Tanriverdi <i>et al.</i> (12)	52	+	+	+	NR	I	M: T < reference values; normal or JLH and FSH	T: RIA E ₂ : CLIA	8
							Premenopausal F: $E_2 < 40.4 \text{ pmol/L}$; inappropriate low LH and FSH Postmenopausal F: LH and FSH in rage of premenopausal women	LH, FSH: RIA	
Herrmann <i>et al.</i> (5)	76	+	+	+	+	I	M: ↓T; inappropriately low LH and FSH Premenopausal F: amenorrhea ↓E ₂ without ↑LH and FSH Postmenopausal F: LH and FSH in premenopausal range	All: IA	17
Bushnik <i>et al.</i> (4)	64	I	I	÷	I	I	M: testosterone deficiency: T < 9.0 nmol/L	NR	17
Klose <i>et al.</i> (7)	104 ^d	+	+	° +	+	I	M: T < 10 nmol/L; inappropriate low LH; inhibin B and SHBG Premenopausal F: amenorrhoea/ oligomenorrhoe; low E_2 , LH,FSH Postmenopausal F: inappropriate low LH,FSH for age	All: ECL	Ν

Basal Basal Basal Basal Basal Serum Mon.	Table 5.	Table 5. Continued								
No. of Basal Serum Menstrual Stimulation Menstrual Stimulation Messal Stable Construction Messal Stable Messal Construction Messal Stable Messal Mess				Basal	Basal		GnRH			Out-
StudySerue IIF3HE,Thistorytiotorytiotorytinterion for hypogonadismH/F3H/TE, $\sqrt{6}$ Tam/werdi30++++-M:T < normal for LIH, F5HF4 $\sqrt{6}$ Tam/werdi30+++-M:T < normal for LIH, F5HF5:CLIAF7 $\sqrt{6}$ Vachter55++T+TTTTTTTVachter55++T+TTTTTTTVachter55++T+TTTTTTTTVachter55++T+TT<		No. of	Basal	serum	serum	Menstrual	Stimula-		Assay	come
Tarriverdi30+++-M:T < normal (total T < 4.6 nmol/L;	Study	patients	serum LH	FSH	E_2/T	history	tion test	Criterion for hypogonadism	LH/FSH/T/E ₂	(%)
era/(14) free T < 11.5pg/ml); normal or LLH, FSH Premenopausal F: E, < 40.4 pmo/LL; LH, FSH: RIA Premenopausal F: E, < 40.4 pmo/LL; LH, FSH: RIA Premenopausal F: E, < 40.4 pmo/LL; LH, FSH: RIA Premenopausal F: E, < 40.4 pmo/LL; LH, FSH: RIA Premenopausal F: E, < 40.4 pmo/LL; PH, FSH: RIA Premenopausal F: E, < 40.4 pmo/LL; PH, FSH: RIA Premenopausal F: E, < 40.4 pmo/LL; PH,	Tanriverdi	30	+	+	+	+	1	M: T < normal (total T < 4.6 nmol/L;	T: RIA	0
Imagnopriate low LH, FSH Imagnopriate low LH, FSH in rage of Postmenopausal F: LH, FSH in rage of Postmenopausal Y: LH, Intel Postmenosasy, FS, Postmenopausal Y: Legendor, ECI, electrochemiluminiscence F, Fondie T, Intel Postmeno E, HDH, Intel Pizing hormone: LHRH, Intel:// Anomali response LHRH, Intere:// Anomali response contered Phopopaditeri	et al.(14)							free T < 11.5pg/ml); normal or ↓LH, FSH Premenopausal F: E ₂ < 40.4 pmol/L;	E ₂ : CLIA LH, FSH: RIA	
Wachter 55 + T+ - T+ + T Pelow reported reference range NR 13 et d.(13) (13) (1-27.7) mo/L) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7) (1-27.7)								inappropriate low LH, FSH Postmenopausal F: LH, FSH in rage of		
Wachter 55 + T+ + T below reported reference range NR 13 et d.(13) (1)								premenopausal women		
eral.(13) (n=NR) ^f (9.7–27.7 mmo/L) E ₂ below reported reference range: (follicular phase 48–709 pmo/L1; midcycle peak 316–1828 pmo/L1; luteal phase 162–775 pmo/L2; postmenopausal < 128 pmo/L1) Abnormal response LHRH test: NR Abnormal response LHRH test: NR Abnormal response LHRH test: NR Abnormal response LHRH, luteinizing hormone; HHR, luteinizing hormone; A, Immunoassay; E ₂ setadiol; ECL, electrochemiluminiscence; F, female; FIA, fluoroimmunoassay; F5H, follicle stimulating hormone; GHD, gonadotrope releasing hormone; JA, Immunoassay; IRA, assays: NR, not reported rAA, ratio immunomerici cassay; T, testosterone e f, female; FIA, microparticle enzyme immunoassay; K ₂ setadiol; ECL, electrochemiluminiscence; G, female; FIA, fluoroimmunoassay; T, testosterone assays apoup of healthy subjects (n=18) 0.00 and all set defined by 3 group of healthy subjects (n=18) 0.00 age- and BMI-matchealthy control subjects (n=18) 0.00 age- and BMI-matchealthy cont	Wachter	55	+	+	+	I	÷	T below reported reference range	NR	13
E ₂ below reported reference range: (follicular phase 48–709 pmo//L; luteal phase 162–775 pmo//L; postmenopausal < 128 pmo//L) Albnormal response LHRH test: NR AcS, automated chemiluminescence system; AUC, area under the curve; CL, confidence limit; CLIA, chemiluminometric immunoassay; F34, follicula simularing homone; GHD, growth homone; LHRH test: NR f. female; F1A, fluoroimmunoassay; F34, follicle stimulating homone; GHD, growth homone; M, male; MEIA, microparticle enzyme immunoassay; MSA, magnetic separation assays; NR, not reported; RIA, radio immunometric assay; T, testosterone assays; NR, not reported; RIA, radio immunometric assay; T, testosterone assays; NR, not reported; RIA, radio immunometric assay; T, testosterone assays; OF and testosterone levels or menstrual history underwent GnRH; only LH measurements for the abard lestosterone levels or menstrual history underwent GnRH; only LH measurements assays; SIR, actic assay; T, testosterone assay; SIR, actic assay; SIR, actic assay; T, testosterone assay; SIR, actic assay; SIR, actic assay; SIR, actic assay; SIR, actic assay; SIR, actic assay; SIR, actic assay; SIR, actic assay; SIR, actic questionnaire, soft equetion assection assay; SIR, testosterone events of inhibin B and SHBG in case of superced hypogonadism in men, evaluation was repeated with measurement of inhibin B and SHBC inderwent stimulation tests were performed if abnormatifies in bas	<i>et al.</i> (13)						(n=NR) ^f	(9.7– 27.7 nmol/L)		
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pmol/L; postmenopausal < 128 pmol/L) Abnormal response LHRH test: NR Abnormal response LHRH test: NR F, female; FIA, fluoroimmunoassay; FSH, follicle stimulating hormone; GHD, growth hormone deficiency; GnRH, gonadotrope releasing hormone; IA, Immunoassay; IRMA, immunoradiometric assay; LHA, Iuteinizing hormone; GHD, growth hormone; M, male; MEIA, microparticle enzyme immunoassay; MSA, magnetic separation assays; NR, not reported; RIA, radio immonmetric assay; T, testosterone • Normal values defined by a group of healthy subjects (n=18) • Tote of fuees defined by 31 healthy subjects (n=18) • Tote off values defined by 31 healthy control subjects • age - and BMT-matched healthy control subjects • In case of subsected hypogonadism in men, evaluation was repeated with measurements • In case of subsected hypogonadism in men, evaluation was repeated with measurement of inhibin B and SHBG • In case of subsects LHRH: 100µg								316–1828 pmol/L; luteal phase 162–775		
Abnormal response LHRH test: NR ACS, automated chemiluminescence system; AUC, area under the curve; CL, confidence limit; CLIA, chemiluminometric immunoassay; E ₃ , estradiol; ECL, electrochemiluminiscence; F, female; FIA, fluoroimmunoassay; FSH, follicle stimulating hormone; GHD, growth hormone deficiency; GnRH, gonadotrope releasing hormone; IA, Immunoassay; IRMA, immunoradiometric assay; LH, luteinizing hormone; LHRH, luteinizing hormone; GHD, growth hormone; M, male; MEIA, microparticle enzyme immunoassay; RMA, immunoradiometric assay; LH, luteinizing hormone; LHRH, luteinizing hormone; IA, male; MEIA, microparticle enzyme immunoassay; MSA, magnetic separation assays; NR, not reported; RIA, radio immunometric assay; T, testosterone • Normal values defined by a group of healthy subjects (n=18) • Normal values defined by 31 healthy cubjects (n=18) • Cut-off values defined by 31 healthy control subjects • Cut-off values defined by 31 healthy control subjects • Cut-off values defined by 31 healthy control subjects • 130 age- and BMI-match values and BMI-methy controls all underwent • In case of suspected hypogonadism in men, evaluation was repeated with measurement of inhibin B and SHBG • In case of suspected hypogonadism in men, evaluation was repeated with measurement of inhibin B and SHBG • In case of suspection tests were performed if abnormatities in basal hormone screening or if patients answered 'yes' to specific questionnaire. Not reported how many patients underwent stimulation tests; LHRH: 100 µg								pmol/L; postmenopausal < 128 pmol/L)		
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	ACS, automati F, female; FIA immunoradio assays; NR, no assays; NR	ed chemilumin , fluoroimmur metric assay; Ll t reported; RIA, abnormalities s defined by 31 3MI-matched h, pected hypogr mulation tests maltions tests;	lescence system noassay: FSH, fc H, luteinizing hc group of health in basal testost healthy controls ealthy controls anadism in men were performe LHRH: 100 µg	r, AUC, area ollicle stimu ormone; LHf metric assay ry subjects (r subjects all underwe , evaluation d if abnorm.	under the c llating horr RH, luteinizii 7, T, testoste n=18) s or menstru in t pituitary i was repeat atlities in ba	urve; CL, confid none; GHD, gru ng hormone rel rone ial history unde testing ed with measur isal hormone sc	ence limit; C owth hormo easing horm rwent GnRH; rwent of inh :reening or if	LIA, chemiluminometric immunoassay; E ₂ estradiol; ECI one deficiency; GnRH, gonadotrope releasing hormon ione; M, male; MEIA, microparticle enzyme immunoassa ; only LH measurements ; only LH measurements ibin B and SHBG f patients answered 'yes' to specific questionnaire. Not	., electrochemilum ne; IA, Immunoas; iy; MSA, magnetic ; reported how man	iniscence; ay; IRMA, eparation y patients

CHAPTER 2

		Basal				
	No. of	serum			Assay	Assay Outcome
Study	patients	PRL	TRH	Criterion for abnormal PRL secretion	PRL	(%)
Kelly <i>et al.</i> (6)	22	1	+ (n= NR)	Hypoprolactinemia: PRL < 95% CL according to AUC ^a	RIA	0
Lieberman <i>et al.</i> (9)	70	+	I	Hyperprolactinemia: M: > 565 pmol/L F: > 1160 pmol/L	MSA	0
Bondanelli <i>et al.</i> (3)	50	+	1	Hyperprolactinemie = PRL above reported reference range Hypoprolactinemia = PRL below reported reference range (M: 87 – 696 pmol/L; F: 174 – 1043 pmol/L)	ACS	16
Agha <i>et al.</i> (1)	102 ^b	+	I	Hyperprolactinemia = basal level PRL > than the locally derived normal assay reference range (M: 83–414 mlU/L; F: 90–523 mlU/L)	FIA	12
Popovic <i>et al.</i> (10)	67	+	I	N	IRMA	12
Aimaretti et al.(2)	70	+	I	Hyperprolactinemia: NR	IRMA	9
Leal-Cerro <i>et al.</i> (8)	66	+	+ ^c (n=NR)	NR	NR	NR
Schneider <i>et al.</i> (11)	70	+	I	Hyperprolactinemia: PRL > 1087 pmol	ECLIA	NR

Table 6. Prolactin secretion

Table 6. Co	Continued					
		Basal				
	No. of	serum			Assay	Assay Outcome
Study	patients	PRL	TRH	Criterion for abnormal PRL secretion	PRL	(%)
Tanriverdi <i>et al.</i> (12)	52	+	1	Hyperprolactinemia: basal level greater than the normal reference range (M: 87 –1783 pmol; premenopausal F: 122–1261 pmol; postmenopausal F:78–870 pmol)	RIA	ω
Herrmann <i>et al.</i> (5)	76	+	I	Hyperprolactinemia: PRL above reported reference range (M:870 pmol; F: <1087 pmol)	ΡI	ſ
Bushnik <i>et al.</i> (4)	64	I	I	1	I	I
Klose <i>et al.</i> (7)	104 ^d	+	I	Hyperprolactinemia: PRL > 510 mlU/L	ECLIA	NR
Tanriverdi <i>et al.</i> (14)	30	+	I	Hyperprolactinemia: basal level greater than the normal reference range (M: 87 –1783 pmol; premenopausal F: 122–1261 pmol; postmenopausal F: 1.8–20 ng/ml)	RIA	NR
Wachter <i>et al.</i> (13)	55	+	+ ^e (n=NR)	NR	NR	NR
AUC, area under the curve; ACS, automated chemilumine Fluoroimmunoassay; IA, Immunoassay; IRMA, immunoradiometric assay; MSA, magnetic separation a hormone ^a Normal values defined by a group of healthy subjects (n=18) ^b Cut-off values defined by 31 healthy control subjects ^c TRH test: 500 ug ^d 30 age- and BMI-matched healthy controls all underwent pit ^e Hormonal stimulation tests were performed if abnormalit reported how many patients underwent stimulation tests. TR	der the curvé assay; IA, Imn radiometric a s defined by a defined by 31 ug MI-matched h mulation test nany patients	 ACS, auto nunoassay; ASA, π group of he group of he healthy cont nealthy conti were perfixion underwent 	mated cherr nagnetic sepa althy subject itrol subjects rols all under ormed if abr stimulation t	AUC, area under the curve; ACS, automated chemiluminescence system; CL, confidence limit; ECLIA, Electrochemiluminiscence immunoassay; FIA, Fluoroimmunoassay; IA, Immunoassay; IRMA, immunoradiometric assay; MSA, magnetic separation assays; NR, not reported; PRL, prolactin; RIA, radio immunometric assay; TRH, thyroid releasing hormone • Normal values defined by a group of healthy subjects (n=18) • Cut-off values defined by 31 healthy control subjects • Cut-off values defined by 31 healthy control subjects • TRH test: 500 ug • Hormonal stimulation tests were performed if abnormalities in basal hormone screening or if patients answered 'yes' to specific questionnaire. Not reported how many patients underwent stimulation tests. TRH: 100 μg	rce immu ; TRH, thyi cific quest	noassay; FIA, oid releasing ionnaire. Not

References

- 1. Agha A, Rogers B, Sherlock M, O'Kelly P, Tormey W, Phillips J *et al*. Anterior pituitary dysfunction in survivors of traumatic brain injury. J Clin Endocrinol Metab 2004 89 4929–4936
- Aimaretti G, Ambrosio MR, Di SC, Gasperi M, Cannavo S, Scaroni C *et al.* Residual pituitary function after brain injury-induced hypopituitarism: a prospective 12-month study. J Clin Endocrinol Metab 2005 90 6085–6092
- 3. Bondanelli M, De ML, Ambrosio MR, Monesi M, Valle D, Zatelli MC *et al*. Occurrence of pituitary dysfunction following traumatic brain injury. J Neurotrauma 2004 21 685–696
- 4. Bushnik T, Englander J & Katznelson L. Fatigue after TBI: association with neuroendocrine abnormalities. Brain Inj 2007 21 559–566
- 5. Herrmann BL, Rehder J, Kahlke S, Wiedemayer H, Doerfler A, Ischebeck W *et al.* Hypopituitarism following severe traumatic brain injury. Exp Clin Endocrinol Diabetes 2006 114 316–321
- Kelly DF, Gonzalo IT, Cohan P, Berman N, Swerdloff R & Wang C. Hypopituitarism following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a preliminary report. J Neurosurg 2000 93 743–752
- 7. Klose M, Juul A, Poulsgaard L, Kosteljanetz M, Brennum J & Feldt-Rasmussen U. Prevalence and predictive factors of post-traumatic hypopituitarism. Clin Endocrinol (Oxf) 2007 67 193–201
- 8. Leal-Cerro A, Flores JM, Rincon M, Murillo F, Pujol M, Garcia-Pesquera F *et al.* Prevalence of hypopituitarism and growth hormone deficiency in adults long-term after severe traumatic brain injury. Clin Endocrinol (Oxf) 2005 62 525–532
- Lieberman SA, Oberoi AL, Gilkison CR, Masel BE & Urban RJ. Prevalence of neuroendocrine dysfunction in patients recovering from traumatic brain injury. J Clin Endocrinol Metab 2001 86 2752–2756
- 10. Popovic V, Pekic S, Pavlovic D, Maric N, Jasovic-Gasic M, Djurovic B *et al*. Hypopituitarism as a consequence of traumatic brain injury (TBI) and its possible relation with cognitive disabilities and mental distress. J Endocrinol Invest 2004 27 1048–1054
- Schneider HJ, Schneider M, Saller B, Petersenn S, Uhr M, Husemann B *et al.* Prevalence of anterior pituitary insufficiency 3 and 12 months after traumatic brain injury. Eur J Endocrinol 2006 154 259–265
- 12. Tanriverdi F, Senyurek H, Unluhizarci K, Selcuklu A, Casanueva FF & Kelestimur F. High risk of hypopituitarism after traumatic brain injury: a prospective investigation of anterior pituitary

function in the acute phase and 12 months after trauma. J Clin Endocrinol Metab 2006 91 2105–2111

- 13. Wachter D, Gundling K, Oertel MF, Stracke H & Boker DK. Pituitary insufficiency after traumatic brain injury. J Clin Neurosci 2009 16 202–208
- 14. Tanriverdi F, Ulutabanca H, Unluhizarci K, Selcuklu A, Casanueva FF & Kelestimur F. Three years prospective investigation of anterior pituitary function after traumatic brain injury: a pilot study. Clin Endocrinol (Oxf) 2008 68 573–579
- 15. Klose M & Feldt-Rasmussen U. Does the type and severity of brain injury predict hypothalamopituitary dysfunction? Does post-traumatic hypopituitarism predict worse outcome? Pituitary 2008 11 255–261
- Schneider HJ, Kreitschmann-Andermahr I, Ghigo E, Stalla GK & Agha A. Hypothalamopituitary dysfunction following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a systematic review. JAMA 2007 298 1429–1438
- Popovic V, Leal A, Micic D, Koppeschaar HP, Torres E, Paramo C *et al.* GH-releasing hormone and GH-releasing peptide-6 for diagnostic testing in GH-deficient adults. Lancet 2000 356 1137–1142
- Agha A, Phillips J & Thompson CJ. Hypopituitarism following traumatic brain injury (TBI). Br Neurosurg 2007 21 210–216
- 19. Bondanelli M, Ambrosio MR, Cavazzini L, Bertocchi A, Zatelli MC, Carli A *et al*. Anterior pituitary function may predict functional and cognitive outcome in patients with traumatic brain injury undergoing rehabilitation. J Neurotrauma 2007 24 1687–1697
- 20. Ghigo E, Aimaretti G & Corneli G. Diagnosis of adult GH deficiency. Growth Horm IGF Res 2008 18 1–16
- 21. Corneli G, Di SC, Baldelli R, Rovere S, Gasco V, Croce CG *et al.* The cut-off limits of the GH response to GH-releasing hormone-arginine test related to body mass index. Eur J Endocrinol 2005 153 257–264
- 22. Darzy KH, Pezzoli SS, Thorner MO & Shalet SM. Cranial irradiation and growth hormone neurosecretory dysfunction: a critical appraisal. J Clin Endocrinol Metab 2007 92 1666–1672
- 23. Agha A, Tomlinson JW, Clark PM, Holder G & Stewart PM. The long-term predictive accuracy of the short synacthen (corticotropin) stimulation test for assessment of the hypothalamic-pituitary-adrenal axis. J Clin Endocrinol Metab 2006 91 43–47
- Clayton RN. Short Synacthen test versus insulin stress test for assessment of the hypothalamo [correction of hypothalmo]-pituitary--adrenal axis: controversy revisited. Clin Endocrinol (Oxf) 1996 44 147-149
- 25. Stewart PM, Corrie J, Seckl JR, Edwards CR & Padfield PL. A rational approach for assessing the hypothalamo-pituitary-adrenal axis. Lancet 1988 1 1208–1210

- 26. Giordano R, Picu A, Bonelli L, Balbo M, Berardelli R, Marinazzo E *et al.* Hypothalamus-pituitaryadrenal axis evaluation in patients with hypothalamo-pituitary disorders: comparison of different provocative tests. Clin Endocrinol (Oxf) 2008 68 935–941
- Sesmilo G, Halperin I & Puig-Domingo M. Endocrine evaluation of patients after brain injury: what else is needed to define specific clinical recommendations? Hormones (Athens) 2007 6 132–137
- 28. Andersen S, Bruun NH, Pedersen KM & Laurberg P. Biologic variation is important for interpretation of thyroid function tests. Thyroid 2003 13 1069-1078
- 29. Adriaanse R, Romijn JA, Endert E & Wiersinga WM. The nocturnal thyroid-stimulating hormone surge is absent in overt, present in mild primary and equivocal in central hypothyroidism. Acta Endocrinol (Copenh) 1992 126 206–212
- 30. Faglia G. The clinical impact of the thyrotropin-releasing hormone test. Thyroid 1998 8 903–908
- 31. Westwood ME, Butler GE, McLellan AC & Barth JH. The combined pituitary function test in children: an evaluation of the clinical usefulness of TRH and LHRH stimulation tests through a retrospective analysis of one hundred and twenty six cases. Clin Endocrinol (Oxf) 2000 52 727–733

Chapter 3

Low prevalence of hypopituitarism after traumatic brain injury – a multi-center study

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Abstract

Objective: Hypopituitarism after traumatic brain injury (TBI) is considered to be a prevalent condition. However, prevalence rates differ considerably among reported studies, due to differences in definitions, endocrine assessments of hypopituitarism, and confounding factors, like timing of evaluation and the severity of the trauma.

Aim: To evaluate the prevalence of hypopituitarism in a large cohort of TBI patients after long-term follow-up using a standardized endocrine evaluation.

Study design: Cross-sectional study

Patients and Methods: We included 112 patients with TBI, hospitalized for at least 3 days and a duration of follow-up > 1 yr after TBI from 5 (neurosurgical) referral centers. Evaluation of pituitary function included fasting morning hormone measurements and insulin tolerance test (ITT n=90) or, when contraindicated, ACTH-stimulation and/or CRH-stimulation test and a GHRH-arginine test (n=22). Clinical evaluation included quality of life questionnaires.

Results: We studied 112 patients (75 males), with median age 48 yr, and mean BMI 26.7 \pm 4.8 kg/m². Mean duration of hospitalization was 11 days (3–105) and 33% had a severe trauma (Glasgow Coma Scale < 9) after TBI. The mean duration of follow-up was 4 (1–12) years. Hypopituitarism was diagnosed in 5.4% (6/112) of patients: severe GH deficiency (n=4), hypogonadism (n=1), adrenal insufficiency (n=2). Patients diagnosed with pituitary insufficiency had significantly higher BMI (*P* =0.002).

Conclusion: In this study, the prevalence of hypopituitarism during long-term follow-up after TBI was low. Prospective studies are urgently needed to find reliable predictive tools for the identification of patients with a significant pre-test likelihood for hypopituitarism after TBI.

Introduction

Traumatic brain injury (TBI) is common and an important cause of death, especially among adolescents in developed countries. The last decade, pituitary insufficiency has emerged as an important sequel following TBI, potentially influencing short- and long-term morbidity. After TBI, many patients experience persistent, invalidating complaints that resemble those observed in patients with hypopituitarism, such as impaired cognition, depression, fatigue and impaired quality of life (QoL) (1–4). Consequently, pituitary insufficiency following TBI may contribute to the problems reported by these patients (4). This condition is important to identify since it can be treated by hormone replacement therapy resulting in improved QoL (3;5).

However, the actual prevalence of hypopituitarism after TBI in an unselected population is subject for debate. The available cohort studies studying the prevalence of pituitary insufficiency report percentages ranging from 15 to 90% (6–18). There are several explanations for this remarkably wide range in reported prevalences, including differences in inclusion criteria, differences in duration of follow-up since TBI (short *vs* long term follow-up) and the use of different tests, different assays and different cut-off values (19).

Therefore, the aim of our study was to evaluate the prevalence of hypopituitarism in a large cohort of TBI-patients after long-term followup, using standardized endocrine evaluation including golden standard tests. The secondary aim was to assess QoL and the contribution of hypopituitarism on QoL.

Patients and methods

Study protocol

We performed a multicenter study in 5 hospitals across the Netherlands (Leiden University Medical Center; Academic Medical Center, Amsterdam, St. Elisabeth Hospital, Tilburg; Isala Clinics, Zwolle; Medical Spectrum Twente, Enschede). Eligible patients were selected from electronic registries of the departments of neurology using the following inclusion criteria: confirmed diagnosis of TBI and hospitalization for at least three days for head injury at least one year prior to endocrine evaluation (to exclude possible hormone alterations mimicking pituitary insufficiency in the early post trauma period), age 18-70 yrs. Exclusion criteria were: medical or psychological problems (not related to TBI) that could disturb interpretation of results, including drug- or alcohol abuse, previously known hypothalamic- or pituitary dysfunction or history of cranial irradiation or pregnancy. Details on trauma severity were derived from the medical records. The Glasgow Coma Scale (GCS) at hospitalisation defined trauma severity. A GCS 13-15 indicates mild trauma, between 9-12 moderate trauma, and < 9 severe trauma (20;21). Ethical approval was obtained by the Medical Ethics Committees of all centers and all patients gave written informed consent.

Patients

A total of 2350 potential patients were retrieved from the electronic databases that had been diagnosed with TBI. The electronic patient records of these patients were retrieved in the departments of neurology of all participating hospitals. However, 1960 patients did not meet the abovementioned inclusion criteria and were excluded. The remaining 390 patients were invited to participate, of whom 278 patients could not be included for various reasons: not willing to participate without giving any reason, not meeting the inclusion criteria (either 2 days of hospitalization, drug or alcohol abuse, or medication that could not be stopped) or were loss to follow-up. Ultimately, we included a total of 112 patients in the study (Figure 1).

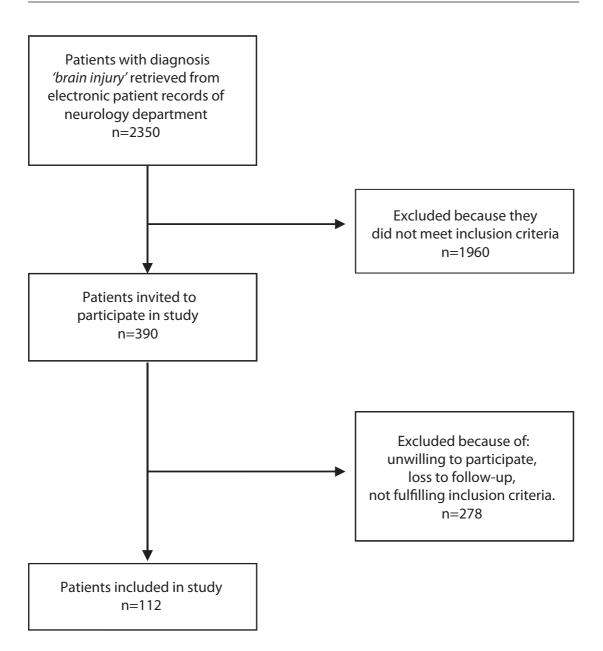


Figure 1. Flow chart of inclusion of patients

Endocrine evaluation

Blood was sampled for assessment of basal and stimulated hormone concentrations between 08.00 and 09.00h A.M. after an overnight fast. All patients rested 30 min prior to testing after insertion of an indwelling catheter in a large forearm vein. Baseline samples were drawn for analyses of cortisol, free thyroxine (fT_4), TSH, testosterone (men) estradiol (E_2 ; women), LH, FSH, prolactin (PRL), GH and IGF-I. Oral contraceptives were discontinued for at least 6 weeks before testing.

The hypothalamic-pitutiary adrenal (HPA) and GH-IGF-I axes were evaluated with an insulin tolerance test (ITT), unless contraindicated, or alternatively by ACTH/CRH and GHRH-stimulation tests. An ACTH-test (1 or 250µg Synacthen iv, Novartis Pharma BV, Arnhem, the Netherlands), with measurement of cortisol at T = -5, 30 and 60 min, was performed routinely in all patients prior to the ITT to ensure sufficient adrenal function. ITT was performed by administering soluble insulin intravenously (0.10 U/kg, Actrapid, Novo, Alphen aan den Rijn, The Netherlands) to induce hypoglycaemia (glucose < 2.2 mmol/L). Cortisol, ACTH, GH and glucose levels were measured at T = -15, 0, 15, 30, 45, 60and 90 min. Peak values of GH of $3 \mu g/L$ and cortisol of >500 nmol/L were considered to reflect sufficient pituitary GH and ACTH function. If ITT was contraindicated, a GHRH-arginine test was conducted to evaluate GH secretory reserve. Patients received 1 µg/kg GHRH (Ferring BV, Hoofddorp, the Netherlands) and 500 mg/kg arginine with a maximum of 30 gr. GH levels were measured at T = -15, 0, 30, 45, 60, 75 and 90 min. BMI adjusted cut-off values of 11.5 μ g/L (< 25 kg/m²), 8.0 μ g/L $(25-30 \text{ kg/m}^2)$, and $4.2 \mu \text{g/L}$ (> 30 kg/m^2) were used (22). For the evaluation of the HPA axis when ITT was contraindicated, the response to ACTHstimulation was considered and an additional CRH-stimulation test was performed in selected cases (Table 2).

Assays

GH was measured in participating centers using in-house assays. The measurement of GH has been harmonized in the Netherlands (23) and in all centers, GH was calibrated against the WHO-IRP 98/574 (1 μ g/L = 3.0 mU/L). IGF-I measurement was centralized at the Department of Clinical Chemistry, Sahlgrenska University Hospital, Göteborg, Sweden using a chemiluminescence immunoassay (DPC, Immulite 2500 system, Siemens Healthcare Diagnostics, Deerfield, IL, USA). The intra- and inter-assay coefficients of variation (CVs) were 4 and 11%. Reference values based on Brabant *et al.* (24) were used. With these IGF-I values IGF-I SD scores were calculated.

The participating centers used the following in house assays and cut-off values:

Leiden University Medical Center, Leiden: Cortisol, fT₄, TSH, LH, FSH and prolactine blood levels were measured by electrochemoluminescent

immunoassay (ECLIA), using a Modular E170, (Roche Diagnostics, Mannheim, Germany). The maximal inter-assay coefficient of variation was 5.0%. ACTH, GH and IGF-I were determined by immunolimunimetric assay using an Immulite 2500 (Siemens Healthcare Diagnostics, Deerfield, IL, USA). The maximal inter-assay coefficient of variation was between 5.0 and 10.0%. Glucose levels were measured using a Modular P800 (Roche Diagnostics Mannheim, Germany) (CV is 3%). For measurement of estradiol levels a radioimmunoassay (RIA, Orion Diagnostica, Espoo, Finland) was used (CV is 6% at 70 pmol/L). The estradiol detection limit was 20 pmol/L. Testosterone was measured using a RIA (Siemens Healthcare Diagnostics, Deerfield IL, USA). (CV is 20% at 1.0 nmol/L and 12% at 14 nmol/L) The detection limit was 0.2 nmol/L.

Academic Medical Center, Amsterdam: Plasma LH, TSH and FSH were analysed by an automated assay on the E170 of Roche (Roche, Mannheim, Germany). The maximal intra- and inter-assay variations were < 5%. Plasma fT₄, PRL and GH were analyzed by fluoroimmunoassay (Delfia, Perkin Elmer, Waltham, MA, USA) using the Delfia 1232 Fluorometer (Perkin Elmer). The maximal intra- and inter-assay CVs were 5.1 and 6.8% for fT4, 3.4 and 5.3% for PRL, and 3.8 and 6.2% for GH, respectively. Testosteron was analysed by an in-house RIA. The maximal intra- and inter-assay CVs were 11.8 and 12.8% respectively. Cortisol was analysed by chemoluminiscence assay using the Immulite 2000 (Siemens, Healthcare Diagnostics). The maximal intra- and inter-assay CVs were 5.5 and 8.3% respectively. E₂ was measured by RIA (Siemens Healthcare Diagnostics). The intra- and inter-assay CVs were < 20% (low level) and maximal at 8.6% (medium level).

St. Elisabeth Hospital, Tilburg and Isala Clinics, Zwolle: Plasma TSH, fT_4 , PRL, LH, FSH, testosterone and E_2 were analyzed by ECLIA (Modular Analytics E170, Roche, GmbH). The maximal intra- and inter-assay CVs as specified by the manufacturer were as follows: TSH, 3.0 and 7.2%; fT_4 , 2.0 and 4.8%; PRL, 1.7 and 2.0%; LH, 1.2 and 2.2%; FSH, 2.8 and 4.5%; testosterone, 2.8 and 3.2%; and E_2 , 3.6 and 3.9%. GH was analyzed by a solid-phase, two-site chemiluminescent immunometric assay (Immulite 2000, Siemens Healthcare Diagnostics). Intra- and inter-assay CVs given by the manufacturer were 4.2 and 6.6% respectively.

Medical Spectrum Twente, Enschede: Plasma GH, LH, FSH, PRL, testosterone, and E_2 levels were analyzed by solid-phase, two-site

chemiluminescent immunoassays (Immulite 2000, Siemens Healthcare Diagnostics). The maximal intra- and inter-assay CVs were as follows: GH, 4.2 and 6.6%; LH, 3.6 and 6.7%; FSH, 2.9 and 4.1%; PRL, 3.6 and 7.4%; testosterone, 10.0 and 10.3%, and E_2 , 7.8 and 11.0%. Cortisol was analyzed by a solid-phase, competitive chemiluminescent immunoassay (Immulite 2000, Siemens). Intra- and inter-assay CVs were 7.4 and 9.4% respectively. Plasma TSH and fT_4 were analysed by ECLIA (Modular Analytics E170, Roche, GmbH). Intra- and inter-assay CVs were: 3.0 and 7.2% for TSH and 2.0 and 3.6% for fT_4 .

Quality of life assessment

To assess QoL the following questionnaires were used:

Hospital Anxiety and Depression Scale (HADS) – The HADS questionnaire consists of 14 items pertaining to anxiety and depression, measured on a four-point scale. The scores for the two subscales anxiety and depression range from 0-21 and the total score from 0-42. A high score indicates more severe anxiety or depression (25).

Nottingham Health Profile (NHP) – The NHP questionnaire features 38 yes/no questions subdivided in six subscales, *i.e.* energy, pain, emotional reaction, sleep, physical ability and social isolation. Scores of the subscales are valued in a range from 0–100. The total score is the mean of all subscales. A high score indicates a worse QoL (26;27).

Multidimensional fatigue index (MFI-20) – The MFI-20 questionnaire contains 20 statements to assess fatigue, measured on a five-point scale. The scores of the five subscales general fatigue, physical fatigue, reduced activity, reduced motivation and mental fatigue vary from 0 to 20. A high score indicates more fatigue experienced (28).

Short Form-36 (SF-36) – The SF-36 consists of 36 statements or questions evaluating general well-being during the previous 30 days. Scores of the nine subscales physical functioning, social functioning, role limitations due to physical problems, role limitations due to emotional problems, mental health, vitality, pain, general health perception and health change are expressed in a 0–100 scale. Higher scores indicate a better QoL (29;30).

Statistics

Data were analyzed using PASW Statistics version 17.0.2 (SPSS Inc., Chicago, IL, USA). All data were presented as mean±SD,

unless mentioned otherwise. The analysis comprised the comparison of the results between patients with and without pituitary insufficiency.

Groups were compared using an independent-samples t-test. A χ^2 -test was used in case of categorical data. To analyse QoL the groups were compared using univariate analysis of variance (ANOVA) with gender and GCS as fixed factors and age as covariate when appropriate. Factors influencing QoL were explored using a Pearson correlation. A P-value of <0.05 was considered to be statistically significant.

Results

Patient demographics

We included 112 patients (75 males) with a median age of 48 (range 19–69) years (Table 1). Patients were evaluated 1–12 years after trauma (median 3 years). The median duration of hospitalization after TBI had been 11 (3–105) days. BMI was 25 (18–43) kg/m². The causes of TBI had been traffic accidents (51%), fall (38%), violence (5%), and sport- or work related accidents (6%), respectively. A total of 36 patients (32%) had been diagnosed with a severe trauma and 56% of the patients (n=60) had a mild trauma, in 4 patients the GCS was not clear from the medical records.

Table 1. Baseline characteristics

	TBI Patients	
	(n=112)	
Gender (M/F)	75/37	
Age (years)	48 (19–69)	
BMI (kg/m ²)	26.7 ± 4.8	
GCS:		
Mild (%)	57%	
Moderate – Severe (%)	43%	
Time since TBI (years)	4.2 ± 3.3	
Duration of hospitalization (days)	11 (3–105)	

BMI, body mass index; F, female; M, male; TBI, traumatic brain injury Data are presented as mean±SD or median (range).

Endocrine evaluation

Any pituitary insufficiency was diagnosed in only 6/112 patients, resulting in a prevalence rate of 5.4%. Patients with and without pituitary insufficiency were comparable in age and gender, but in patients diagnosed with pituitary insufficiency BMI was significantly higher (P = 0.02). Trauma severity, the duration of follow-up, and duration of hospitalization were not different between the two groups.

GH-IGF-I axis: The ITT was used in 80% of the patients (90/112) for the evaluation of GH secretory reserve (Figure 2). Because of contraindications (epilepsy (n=6), ischemic heart disease or rhythm disorders (n=3), other (n=13)), the remaining patients were tested using combined GHRH-arginine stimulation. Severe growth hormone deficiency (GHD) was diagnosed in 3.6% of the patients (2M/2F, Table 2).

HPA axis: At baseline, all patients initially were screened with basal morning cortisol levels and a 1 or 250μ g ACTH-test to evaluate adrenal function. Subsequently, 90 patients were tested by ITT (Figure 2). In the remaining 22 patients the HPA axis was assessed by the results of basal cortisol and the ACTH-test. In addition, 2 patients (diagnosed with other pituitary insufficiencies) were tested also by a 100 µg CRH test.

ACTH deficiency was diagnosed in 1.8% of patients (2/112) by insufficient cortisol responses during ITT (Table 2).

Gonadal axis: Hypogonadism was diagnosed only in one male patient (0.9%).

Thyroid axis: We did not diagnose any patient with thyroid insufficiency.

Quality of life

There were differences in QoL between patients diagnosed with and without pituitary insufficiency. Patients with pituitary insufficiency scored worse on almost all subscales of the QoL questionnaires. More specifically, they scored significantly worse on the subscale 'Depression' of the HADS (P = 0.05), on the subscale 'Social isolation' of the NHP (P = 0.02), on the subscale 'Reduced activity' of the MFI-20 (P = 0.027) and on the subscale 'General health perception' of the SF-36 (P = 0.016) (data not shown).

					Time since	Dvnam	Dvnamic test			Peak	
Patient		Age	BMI	gcs	TBI			IGF-I	Peak GH	cortisol	
no.	Sex	(years)	(kg/m²)	score	(years)	GH axis	HPA axis	SDS	(hg /L)	(nmol/L)	(nmol/L) Deficiency
-	Σ	65	32.2	m	ſ	L	L	-1.0	4.0	425	Cortisol
2	Σ	64	29.7	7	6	GHRH-arg	ACTH,CRH	-2.4	2.8	508	GH
ŝ	Σ	41	32.8	ŝ	4	GHRH-arg	ACTH	-0.2	9.9	590	Testosterone
4	Σ	27	23.5	ŝ	10	GHRH-arg	ACTH, CRH	-0.7	9.4	757	GH
S	ш	28	29	15	-	Ħ	E	1.2	1.9	790	GH
9	ш	23	32.3	14	Ŷ	μ	ΤΠ	-0.6	2.4	395	GH, Cortisol
BMI, body	mass in	dex; CRH, cc	orticotropin n	eleasing ho	ormone; F, femë	ale; GCS, Glasgov	w Coma Scale; Gł	H, growth	hormone; GF	HRH, growth	BMI, body mass index; CRH, corticotropin releasing hormone; F, female; GCS, Glasgow Coma Scale; GH, growth hormone; GHRH, growth hormone releasing

 Table 2.
 Characteristics of patients diagnosed with any pituitary insufficiency

hormone; HPA, hypothalamic pituitary adrenal axis; ITT, insulin tolerance test; M male

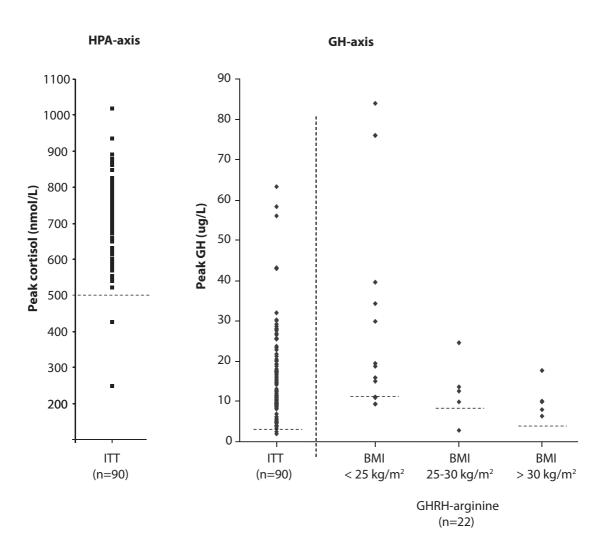


Figure 2. Test results of the stimulation of GH and cortisol secretory reserves during ITT or combined GHRH-arginine test. The dotted horizontal line represents the cut-off value used to define an insufficient response.

Discussion

This study demonstrates that the prevalence of hypopituitarism after TBI in a large patient cohort after long-term follow-up is low. Using a standardized evaluation that included the gold standard test for the evaluation of GH and cortisol secretory reserves in the majority of the patients, we found a prevalence of only 5.4% of any pituitary insufficiency.

This prevalence of hypopituitarism is much lower compared with the prevalence rates reported in the majority of the previous studies (15–90%) (6–18). This might be explained by the use of different endocrine tests and cut-off values (19). For example, comparable low prevalences of hypopituitarism was found in another study that also used the ITT for screening (15). In addition, when using the combined GHRH-arginine test without BMI-adjusted cut-off values the prevalence of severe GHD varied between 8 and 20%(19). A higher BMI is associated with a decreased GH response to GH stimulation tests (22). If BMI-adjusted cut-off values are not used, a higher proportion of patients will be classified as GHD. In addition, age adjusted cut-off values have recently been reported for the GHRH-arginine test (31).

Differences in the duration of follow-up between TBI and endocrine assessment may also play an important role. Hormone alterations mimicking pituitary insufficiency can be present in the acute phase after trauma. In general, these transient effects are almost exclusively reported only within the first six months after TBI (15;32). Therefore, assessment of the function of pituitary axes within this timeframe may result in higher prevalence rates of hypopituitarism. To avoid this bias we decided to assess patients at least one year after the trauma, as suggested in the consensus guidelines for the evaluation and diagnosis of patients with possible GHD (33). In addition to the time interval between TBI and endocrine assessment, the severity of trauma may affect the prevalence rate of pituitary insufficiency (15;34). As shown by Klose *et al.* (34), increased trauma severity increases the risk of pituitary insufficiency. This may result in higher prevalence rates when patients with a more severe degree of trauma are included. Conversely, prevalence rates of hypopituitarism may decrease when patients with only minor traumas are included (35).

It is important to note that in our study, only a minority of the screened patients fulfilled our inclusion criteria, of which 28.7% participated. Therefore, by definition, we investigated a pre-selected cohort, which may have affected the results, and, therefore, our conclusions cannot simply be extrapolated to all TBI patients. However, we were able to evaluate the most important clinical characteristics in the majority of the patients (79%) who did not participate and found no differences in age during TBI, gender, trauma severity and duration of hospitalization when compared to those that finally did participate (data not shown). This makes a possible bias as a result of pre-selection less likely.

Thus, according to our results, pituitary insufficiency may be a rare complication of TBI in patients evaluated at least one year after TBI. Intriguingly, comparable low prevalence rates were found in another study that also used the ITT to evaluate cortisol and GH secretory reserve (32). However, it should be taken into account that there is a high incidence of TBI in the population probably translating in still a high prevalence of posttraumatic hypopituitarism on a populationbased level. Besides pre-selections of patients, the use of different tests with different cut-off values has contributed to the differences and large variations in the prevalence rates found in previous studies (19). Our results accentuate that we urgently need consensus for a more uniform and protocol endocrine evaluation after TBI. More importantly, we urgently need prospective studies to find reliable predictors that enable the identification of patients with a significant pre-test likelihood for hypopituitarism. This is of paramount importance, because the presence of pituitary failure, even in a small proportion of patients, is potentially treatable, may be lifesaving, and is likely to significantly ameliorate quality of life (3;5).

References

- Fleminger S & Ponsford J. Long term outcome after traumatic brain injury. BMJ 2005 331 1419–1420
- van Aken MO & Lamberts SW. Diagnosis and treatment of hypopituitarism: an update. Pituitary 2005 8 183–191
- 3. Blair JC. Prevalence, natural history and consequences of posttraumatic hypopituitarism: a case for endocrine surveillance. Br J Neurosurg 2010 24 10–17
- Klose M, Watt T, Brennum J & Feldt-Rasmussen U, Posttraumatic hypopituitarism is associated with an unfavorable body composition and lipid profile, and decreased quality of life 12 months after injury. J Clin Endocrinol Metab 2007 92 3861–3868
- 5. Maric NP, Doknic M, Pavlovic D, Pekic S, Stojanovic M, Jasovic-Gasic M *et al.* Psychiatric and neuropsychological changes in growth hormone-deficient patients after traumatic brain injury in response to growth hormone therapy. J Endocrinol Invest 2010 33 770–775
- 6 Lieberman SA, Oberoi AL, Gilkison CR, Masel BE & Urban RJ. Prevalence of neuroendocrine dysfunction in patients recovering from traumatic brain injury. J Clin Endocrinol Metab 2001 86 2752–2756
- Kelly DF, Gonzalo IT, Cohan P, Berman N, Swerdloff R & Wang C. Hypopituitarism following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a preliminary report. J Neurosurg 2000 93 743–752
- 8. Bondanelli M, De Marinis L, Ambrosio MR, Monesi M, Valle D, Zatelli MC *et al.* Occurrence of pituitary dysfunction following traumatic brain injury. J Neurotrauma 2004 21 685–696
- 9. Agha A, Rogers B, Sherlock M, O'Kelly P, Tormey W, Phillips J *et al*. Anterior pituitary dysfunction in survivors of traumatic brain injury. J Clin Endocrinol Metab 2004 89 4929–4936
- 10. Popovic V, Pekic S, Pavlovic D, Maric N, Jasovic-Gasic M, Djurovic B *et al*. Hypopituitarism as a consequence of traumatic brain injury (TBI) and its possible relation with cognitive disabilities and mental distress. J Endocrinol Invest 2004 27 1048–1054
- 11. Leal-Cerro A, Flores JM, Rincon M, Murillo F, Pujol M, Garcia-Pesquera F *et al.* Prevalence of hypopituitarism and growth hormone deficiency in adults long-term after severe traumatic brain injury. Clin Endocrinol (Oxf) 2005 62 525–532
- Schneider HJ, Schneider M, Saller B, Petersenn S, Uhr M, Husemann B *et al.* Prevalence of anterior pituitary insufficiency 3 and 12 months after traumatic brain injury. Eur J Endocrinol 2006 154 259–265

- Tanriverdi F, Senyurek H, Unluhizarci K, Selcuklu A, Casanueva FF & Kelestimur F. High risk of hypopituitarism after traumatic brain injury: a prospective investigation of anterior pituitary function in the acute phase and 12 months after trauma. J Clin Endocrinol Metab 2006 91 2105-2111
- 14. Herrmann BL, Rehder J, Kahlke S, Wiedemayer H, Doerfler A, Ischebeck W *et al.* Hypopituitarism following severe traumatic brain injury. Exp Clin Endocrinol Diabetes 2006 114 316–321
- 15. Klose M, Juul A, Poulsgaard L, Kosteljanetz M, Brennum J & Feldt-Rasmussen U. Prevalence and predictive factors of post-traumatic hypopituitarism. Clin Endocrinol (Oxf) 2007 67 193-201
- 16. Bushnik T, Englander J & Katznelson L. Fatigue after TBI: association with neuroendocrine abnormalities. Brain Inj 2007 21 559–566
- 17. Wachter D, Gundling K, Oertel MF, Stracke H & Boker DK. Pituitary insufficiency after traumatic brain injury. J Clin Neurosci 2009 16 202–208
- Tanriverdi F, Ulutabanca H, Unluhizarci K, Selcuklu A, Casanueva FF & Kelestimur F. Three years prospective investigation of anterior pituitary function after traumatic brain injury: a pilot study. Clin Endocrinol (Oxf) 2008 68 573–579
- Kokshoorn NE, Wassenaar MJ, Biermasz NR, Roelfsema F, Smit JW, Romijn JA *et al.* Hypopituitarism following traumatic brain injury: prevalence is affected by the use of different dynamic tests and different normal values. Eur J Endocrinol 2010 162 11–18
- 20. Sternbach GL. The Glasgow coma scale. J Emerg Med 2000 19 67–71
- 21. Teasdale G & Jennett B. Assessment of coma and impaired consciousness. A practical scale. Lancet 1974 2 81–84
- 22. Corneli G, Di Somma C, Baldelli R, Rovere S, Gasco V, Croce CG *et al*. The cut-off limits of the GH response to GH-releasing hormone-arginine test related to body mass index. Eur J Endocrinol 2005 153 257–264
- Ross HA. Reporting growth hormone assay results in terms of one consensus recombinant standard preparation offers less than optimal reduction of between-method variation. Clin Chem Lab Med 2008 46 1334–1335
- 24. Brabant G, von zur Mühlen A, Wuster C, Ranke MB, Kratzsch J, Kiess W *et al.* Serum insulin-like growth factor I reference values for an automated chemiluminescence immunoassay system: results from a multicenter study. Horm Res 2003 60 53–60
- 25. Zigmond AS & Snaith RP. The hospital anxiety and depression scale. Acta Psychiatr Scand 1983 67 361–370
- Hunt SM & McEwen J. The development of a subjective health indicator. Sociol Health Illn 1980
 2 231–246
- 27. Hunt SM, McKenna SP, McEwen J, Backett EM, Williams J & Papp E. A quantitative approach to perceived health status: a validation study. J Epidemiol Community Health 1980 34 281–286
- 28. Smets EM, Garssen B, Bonke B & De Haes JC. The Multidimensional Fatigue Inventory (MFI) psychometric qualities of an instrument to assess fatigue. J Psychosom Res 1995 39 315–325

- 29. Brazier JE, Harper R, Jones NM, O'Cathain A, Thomas KJ, Usherwood T *et al.* Validating the SF-36 health survey questionnaire: new outcome measure for primary care. BMJ 1992 305 160–164
- 30. Ware JE, Jr. & Sherbourne CD. The MOS 36-item short-form health survey (SF-36). I. Conceptual framework and item selection. Med Care 1992 30 473–483
- 31. Colao A, Di Somma C, Savastano S, Rota F, Savanelli MC, Aimaretti G *et al.* A reappraisal of diagnosing GH deficiency in adults: role of gender, age, waist circumference, and body mass index. J Clin Endocrinol Metab 2009 94 4414–4422
- 32. Klose M, Juul A, Struck J, Morgenthaler NG, Kosteljanetz M & Feldt-Rasmussen U. Acute and long-term pituitary insufficiency in traumatic brain injury: a prospective single-centre study. Clin Endocrinol (Oxf) 2007 67 598–606
- 33. Ho KK. Consensus guidelines for the diagnosis and treatment of adults with GH deficiency II: a statement of the GH Research Society in association with the European Society for Pediatric Endocrinology, Lawson Wilkins Society, European Society of Endocrinology, Japan Endocrine Society, and Endocrine Society of Australia. Eur J Endocrinol 2007 157 695–700
- 34. Klose M & Feldt-Rasmussen U. Does the type and severity of brain injury predict hypothalamo-pituitary dysfunction? Does post-traumatic hypopituitarism predict worse outcome? Pituitary 2008 11 255–261
- 35'. van der Eerden AW, Twickler MT, Sweep FC, Beems T, Hendricks HT, Hermus AR *et al.* Should anterior pituitary function be tested during follow-up of all patients presenting at the emergency department because of traumatic brain injury? Eur J Endocrinol 2010 162 19–28

Chapter 4

The use of an early postoperative CRH test to assess adrenal function after transsphenoidal surgery for pituitary adenomas

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Abstract

Purpose: Transsphenoidal surgery (TS) is the treatment of choice for many pituitary tumors. Because TS may cause pituitary insufficiency in some of these patients, early postoperative assessment of pituitary function is essential for appropriate endocrine management. The aim of our study was to evaluate the clinical relevance of the CRH stimulation test in assessing postoperative pituitary-adrenal function.

Methods: We performed a retrospective analysis of 144 patients treated by TS between January 1990 and November 2009, in whom a CRH test and a second stimulation test was performed to assess adrenal function during follow-up. Patients with Cushing's disease were excluded. Hydrocortisone substitution was started if peak cortisol levels were < 550nmol/L.

Results: The cortisol response was insufficient in 42 (29%) and sufficient in 102 patients at the postoperative CRH test. Thirteen of 42 (30%) demonstrated a normal cortisol response during a second cortisol stimulation test. In 75 of the 102 patients with a sufficient response to CRH repeat testing revealed an insufficient cortisol response in 14 patients (14%). All but one had concomitant pituitary hormone deficits. There were no cases of adrenal crises during follow-up. Additional pituitary insufficiency was significantly more present (P < 0.001) in the group of patients with an abnormal response to CRH directly after surgery.

Conclusion: In this study a substitution strategy of hydrocortisone guided by the postoperative cortisol response to CRH appeared safe and did not result in any case of adrenal crises. However, the early postoperative CRH test does not reliably predict adrenal function after TS for pituitary adenomas in all patients, and retesting should be strongly considered.

Introduction

Transsphenoidal surgery (TS) is the treatment of choice for many pituitary tumors. TS may result in (additional) pituitary insufficiency in some of these patients (1–3). Therefore, accurate assessment of pituitary function is essential for appropriate management of postoperative patients after TS. In this respect, evaluation of the pituitary-adrenal axis is clinically relevant to assess the need for hydrocortisone replacement therapy at discharge.

The insulin tolerance test (ITT) is considered to be the gold standard for the evaluation of secondary adrenal insufficiency (4;5). Because there are contraindications for ITT in some patients, the CRH test, the metyrapone test or the ACTH stimulation test can be used as alternative dynamic tests to assess adrenal function (6-8). However, there is no international consensus for postoperative testing after pituitary surgery. We performed a structured literature search for articles that 1) evaluated the postoperative strategy for evaluation of adrenal function and 2) use of the CRH test to evaluate the pituitary-adrenal axis in postoperative patients after TS for pituitary adenomas, excluding manuscripts on patients with Cushing's disease. However, specific data on this topic are hardly available. Moreover, studies that compared CRH test and other dynamic test in other situations (*i.e.* in patients with (suspected) hypothalamic-pituitary insufficiency not specifically related to surgery) reported contradictory results (8;9). Therefore, at our center we developed a strategy for evaluation of patients after pituitary surgery in 1990 using the CRH test as the first postoperative test.

The aim of the present study was to assess the clinical relevance of the CRH stimulation test, as a part of this evaluation strategy, in assessing pituitary-adrenal function after TS. We performed a retrospective analysis of all patients treated by TS between January 1990 and November 2009, in whom a CRH test and a second stimulation test was performed to assess adrenal function during follow-up in non-Cushing patients.

Patients and methods

Study design

We performed a retrospective chart review of all patients, who had been treated by TS in the Leiden University Medical Center between January 1990 (when human CRH (hCRH) became available for routine clinical use) and November 2009. Patients with available data on a postsurgical CRH test, who also had a second (confirmation) test of adrenal function during follow-up were included. We excluded patients on high dose glucocorticoids, reoperation, postoperative cranial radiotherapy, and patients treated by TS for Cushing's disease.

The Medical Ethics Committee of our hospital declared that no formal ethical approval and written informed consent was needed for this anonymous retrospective chart review.

Endocrine assessment

According to the postoperative protocol, which has been implemented in our hospital, the pituitary-adrenal axis is assessed by CRH test, 7–10 days after surgery. The CRH test is performed after an overnight fast, after withdrawal of hydrocortisone for 24 hours, using 100 µg CRH (Corticoliberine, Ferring Farmaceuticals Hoofddorp, the Netherlands). Venous blood samples for measurement of ACTH and cortisol concentrations are collected at -15, 0, 15, 30, 45 and 60 minutes after infusion. A peak plasma cortisol of \geq 550 nmol/L is considered to reflect a normal response (10;11).

In case of insufficient cortisol responses to CRH, hydrocortisone is prescribed (20 mg/day, divided in 3 doses). During follow-up, the treating endocrinologist decided on re-testing of the adrenal function. For the assessment of the HPA axis during follow-up either basal serum cortisol levels or a stimulation test was used. The ITT was performed after an overnight fast by intravenous administration of insulin (0.10 U/kg, Actrapid, Novo Nordisk Farma, Bagsvaerd, Denmark) to induce adequate hypoglycemia, defined as nadir glucose levels < 2.2 mmol/L. Blood was collected for measurement of cortisol, ACTH and GH at -15, 0, 15, 30, 45, 60, 90 and 120 minutes after i.v. administration of insulin. Peak values of GH > 9 mU/L (corresponding with 3 μ g/L) and cortisol of \geq 550 nmol/L were considered to reflect normal pituitary function of GH and ACTH secretion (4;12–15).

For the ACTH test 1 µg Synacthen (Novartis Pharma, Arnhem, The Netherlands) was administered i.v. and cortisol levels were measured at -15, 0 and 30 minutes after infusion. A peak cortisol value of \geq 550 nmol/L was considered to reflect normal adrenal reserve (16–18). In addition, a basal serum cortisol concentration of > 550 nmol/L was considered to reflect normal adrenal function (9).

In some patients a metyrapone test was used as a second test to assess pituitary adrenal function. Metyrapone (30 mg/kg, Metopiron, Novartis Pharma B.V., Arnhem, the Netherlands) was administered orally at midnight. The next morning postabsorptive blood samples were obtained for measurement of 11-deoxycortisol, cortisol and ACTH levels. A cutoff value for 11-deoxycortisol of 200 nmol/L was used to define normal adrenal function (6;19;20).

Assays

Between 1986 and 1994, a fluorescence energy-transfer immunoassay Syva-Advance (Syva Company, Palo Alto, CA) was used, with an interassay variation coefficient of 3.6 – 6.1% and a detection limit of 50 nmol/L. From 1994, cortisol was measured by fluorescencepolarization assay on a TDx (Abbott Laboratories, Abbott Park, IL). The interassay variation coefficient is 5–6% above 500 nmol/L and amounts to 12% under 200 nmol/L. The detection limit is 20 nmol/L. The methods correlated well with each other, and therfore no correction factors were introduced for follow-up of patients. ACTH was determined by immunolimunimetric assay using an Immulite 2500 (Siemens Healthcare Diagnostics, Deerfield, IL, USA). The maximal inter-assay coefficient of variation (CV) was between 5.0 and 10.0%. During the insulin tolerance test glucose levels were measured using a Modular P800 (Roche Diagnostics, Mannheim, Germany).

For the measurement of 11-deoxycortisol a radioimmunoassay (RIA) of Diasource (previously Biosource Europe, Nivelles, Belgium) was used. CV was approximately 11%.

Free T_4 , TSH, LH, FSH and prolactine blood levels were measured by electrochemoluminescent immunoassay (ECLIA), using a Modular E170, (Roche Diagnostics, Mannheim, Germany). The maximal inter-assay CV for these hormones was 5.0%. ACTH, GH and IGF-I were determined by immunolimunimetric assay using an Immulite 2500 (Siemens Healthcare Diagnostics, Deerfield, IL, USA). The maximal inter-assay CV was between 5.0 and 10.0%. Glucose levels were measured using a Modular P800 (Roche Diagnostics Mannheim, Germany) (CV is 3%). For measurement of estradiol levels a RIA (Orion Diagnostica, Espoo, Finland) was used (CV is 6% at 70 pmol/L). The estradiol detection limit was 20 pmol/L. Testosterone was measured using a RIA (Siemens Healthcare Diagnostics, Deerfield IL, USA). (CV is 20% at 1.0 nmol/L and 12% at 14 nmol/L). The detection limit was 0.2 nmol/L.

Statistical analysis

PAWS for Windows version 17.0 (SPSS Inc. Chicago, IL) was used to perform data analysis. Data were presented as mean \pm SD unless otherwise mentioned. To evaluate the difference between peak cortisol of the direct postsurgical CRH test and the confirmation test during followup we used a paired t-test. A χ^2 -test was used to evaluate the difference in prevalence of additional pituitary insufficiency in patients diagnosed with or without adrenal insufficiency based on the CRH stimulation test. The level of significance was set at $P \leq 0.05$.

Results

Patients

Between January 1990 and November 2009, 291 patients were treated by TS for non-functioning pituitary adenomas; NFA (n=160), GHproducing adenomas (n=96), prolactinomas (n=16) or other pituitary tumors (n=19) (Figure 1). A CRH test directly following surgery was not performed in 82 patients for several reasons (pituitary insufficiency prior to surgery n=29, follow-up in outpatient clinic n=11, use of corticosteroids surrounding surgery n=5, other stimulation test directly after surgery n=7, other n=30). Consequently, a CRH test was performed in 209 postoperative patients after TS. In 65 of these 209 patients, there was no additional adrenal test performed in follow-up between TS and referral for postoperative radiotherapy (n=24), repeat surgery (n=5), or death of the patient (n=10), or due to follow-up in another hospital (n=17) and lost to follow-up (n=9). Therefore, 144 patients were finally included in this study. Baseline characteristics of these 144 patients are presented in Table 1.

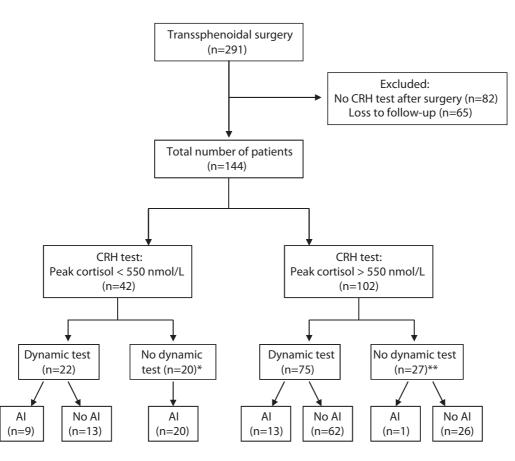


Figure 1. Flow-chart of patient selection and follow-up. AI = adrenal insufficiency. *pre-existent panhypopituitarism before or immediately after surgery (n=12), pre-existent isolated severe adrenal insufficiency before surgery (n=4) or very low basal serum cortisol concentrations (mean 10 nmol/L) during follow-up after surgery (n=4).

**basal serum cortisol levels > 550 nmol/L (n=12), normal urine cortisol levels (n=3), short followup between repeated surgery or additional radiotherapy (n=2), and follow-up < 1 year (n=2) or unspecified reasons (n=7), basal serum cortisol < 110 nmol/L (n=1)

Baseline characteristics	Number of patients (n=144)
Gender (M/F)	71/73
Age (years)	50 (15–83)
Diagnosis (n):	
NFA	70
Acromegaly	63
Prolactinoma	6
Other pituitary tumors	5
Time between CRH test and confirmation test (months)	25.5 (2days*–219 months)
Confirmation test (n=97):	
ITT	55
CRH	16
ACTH stimulation test	21
Metyrapone test	5

* Basal serum cortisol was low, however CRH test peak cortisol 0.61; 2 days after CRH test a metyrapone test was performed

Patients with a decreased postoperative cortisol response to CRH (n=42)

The peak levels of cortisol during the postoperative CRH test classified 42 of the 144 patients with pituitary-adrenal insufficiency (peak cortisol < 550 nmol/L) (Figure 1). In 22 of these 42 patients with a median peak cortisol response to CRH of 480 (30-547) nmol/L, a second stimulation test was performed during follow-up: ITT (n=8), ACTH stimulation test (n=8), CRH stimulation test (n=5) and metyrapone test (n=1). These confirmation tests were performed with a median interval of 27.5 (1–139) months after the initial postoperative CRH test. Based on this repeat test, 9 of these 22 (41%) patients had persistent adrenal insufficiency [median initial cortisol response 356 (30-547) nmol/L; median cortisol response confirmation test 219 (3-514) nmol/L}, who received hydrocortisone (HC) replacement and 13 (59%) with a normal response, in whom HC was discontinued. In these 13 patients, the median peak cortisol level to postoperative CRH stimulation was 480 (340-543) nmol/L, whereas the median peak cortisol level during the second test were 672 (570–890) nmol/L (P < 0.001). The clinical characteristics of these patients are detailed in Table 2. Based on the results of the CRH test, four patients did not receive HC directly after surgery, or only if necessary. In two of these patients (Table 3; patient 2 and 8) the physician defined the HPA axis as normal based on the peak cortisol of the CRH test (540 and 543 nmol/L respectively). No clinical events were reported.

In 20 of these 42 patients with a median CRH-stimulated cortisol concentration of 194 (6–510) nmol/L, no additional stimulation test of adrenal function was performed during follow-up, but basal morning cortisol levels after the withdrawal of hydrocortisone for 18-24 h were used to assess the axis. Persistent adrenal insufficiency was considered to be present in these 20 patients because of pre-existent panhypopituitarism before or immediately after surgery (n=12), pre-existent isolated severe adrenal insufficiency before surgery (n=4) or very low basal serum cortisol concentrations (mean 10 nmol/L) during follow-up after surgery (n=4). Accordingly, all these patients received hydrocortisone supplementation directly after the post surgical CRH test until now.

Patients with a normal postoperative cortisol response to CRH (n=102)

The peak levels of cortisol during the postoperative CRH test classified 102 of the 144 patients with normal pituitary-adrenal function (peak cortisol > 550 nmol/L) (Figure 1). In 75 of these 102 patients, adrenal

function was assessed during follow-up using a second stimulation test and by basal postabsorptive cortisol levels only in the other 27 patients. These 27 patients were not subjected to a second stimulation test because of basal serum cortisol levels > 550 nmol/L (n=12), normal urine cortisol levels (n=3), short follow-up between repeated surgery or additional radiotherapy (n=2), and follow-up < 1 yr (n=2) or unspecified reasons (n=7). One patient returned within three months after surgery with complaints and basal postabsorptive serum cortisol levels of 90 nmol/L and HC was started without additional stimulation test. The ITT was used in 49 of the 75 patients, the CRH test in 11 patients, the ACTH stimulation test in 11, and the metyrapone test in four patients. A normal response to these tests was found in 62 patients. However, 13 patients had an insufficient adrenal response to these tests. With the inclusion of the patient with very low basal serum cortisol levels (see above), 14 patients were classified as adrenal insufficient (Table 3). Thirteen of these 14 patients had been diagnosed with any other additional pituitary insufficiencies and 8 of these patients (57%) had panhypopituitarism. Six patients already received HC directly after surgery. None of these 14 patients experienced any clinical event related to cortisol deficiency.

Prevalence of additional pituitary insufficiency

A total of 73 patients had additional pituitary insufficiency. The prevalence of additional pituitary insufficiency was significantly higher in patients diagnosed with an insufficient CRH stimulation test after surgery compared to patients with a normal test result (any hypopituitarism P < 0.001; GHD P < 0.001; TSH deficiency P < 0.001; LH/FSH deficiency P = 0.001).

cortisol Diagnosiscortisol (MmO/L)curgety (y/n)Endion (years)cortisol testNFA 410 y y y x x x NFA 410 y x x x x x Acromegaly 540 n x x x x Acromegaly 540 n x x x x Acromegaly 480 n x x x x Acromegaly 480 n x x x x Prolactinoma 480 y x x x x NFA 440 y x x x x NFA 469 y x x x x NFA 469 y x x x x	Ag	Age at time of		CHR test Peak	HC after		Confir-	Peak			Clinical
DiagnosiImolul(yn)(yn)testImolulNFA410y3 muthsACTH890NFA410y3.8CRH790Acromegaly540n3.8CRH790Acromegaly480n4.9171780Acromegaly480y8.9171780Prolactinoma480y8.9171750NFA440y1 muthCRH710NFA469y7 muths164710	Gen- surgery	Σ		cortisol	Surgery	Follow-up	mation	cortisol	Other defi-		event
NFA 410 y 3 muths ACTH 890 Acromegaly 540 n 3.8 CRH 790 Acromegaly 480 n 4.9 ITT 780 Prolactinoma 480 y 8.9 ITT 750 NFA 440 y 100th CRH 710 NFA 469 y 7muths ITT 695	(yrs)	_	Diagnosis	(nmol/L)	(n/y)	(years)	test	(nmol/L)	ciencies	Follow-up	(u/k)
Acromegaly 540 n 3.8 CRH 790 Acromegaly 480 n 4.9 171 780 Acromegaly 480 y 8.9 177 780 Prolactinoma 480 y 8.9 177 780 NFA 440 y 8.9 171 750 NFA 440 y 710 750 NFA 469 y 710 696	53		NFA	410	~	3 mnths	ACTH	890	LH/FSH, GHD	LH/FSH, GHD No optimal reaction CRH test, HC	c
Acromegaly 540 n 3.8 CRH 790 Acromegaly 480 n 4.9 1T 780 Prolactinoma 480 y 8.9 1T 780 Prolactinoma 480 y 8.9 1T 750 NFA 440 y 1mth CRH 710 NFA 469 y 7mths 1T 696										discontinued before commitmention test. After 3 years ITT peak cortisol 860 nmol/L	
Acromegaly 480 n 4.9 ITT 780 Prolactinoma 480 y 8.9 ITT 750 NFA 440 y 8.9 ITT 750 NFA 460 y 1 mnth CRH 710 NFA 469 y 7 mnths ITT 696	48		Acromegaly	540	c	3.8	CRH	062	None	After CRH test HPA axis defined as normal, no HC	۲
Prolactinoma 480 y 8.9 ITT 750 NFA 440 y 1 muth CRH 710 NFA 469 y 7 muths ITT 696	23		Acromegaly	480	c	4.9	E	780	None	After CRH test another CRH test and 24h urine still not sufficient.; ITT after 4 years insufficient	c
440 y 1 mnth CRH 710 469 y 7 mnths ITT 696	26		Prolactinoma		>	8 0.	E	750	ō	HC until next outpatient appoint- ment; 5 yrs loss to follow-up; recur- rence of prolactinoma treatment with Dostinex. Did not use HC. 8 yrs after surgery 2 sufficient ACTH tests and 1 yr later normal ITT	c
NFA 469 y 7 mnths ITT 696	53		NFA	440	>	1 mnth	CRH	710	GHD	HC discontinued after CRH test. (9 yrs after surgery RT)	c
	62		NFA	469	>	7 mnths	E	696	GHD	After 4 months ACTH peak cortisol 580 therefore stop HC. 3 months later ITT	c

 Table 2.
 Patients incorrectly diagnosed with adrenal insufficiency based on the CRH test directly after surgery

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Gen- surgery der (vrs)		corticol	Surgery	Follow-up	mation	00:4000			,
						COLLISOI	Other defi-		event
	Diagnosis	(nmol/L)	(u/k)	(years)	test	(nmol/L)	ciencies	Follow-up	(n/y)
M 42	Acromegaly	530	i.n.	11.6	ACTH	618	None	Based on CRH only HC if necessary.	C
								Shortly after CRH test ACTH test with normal cortisol response; no HC ne-	
								cessary. 11 yrs later another ACTH test	
F 42	NFA	482	Х	1.3	CRH	606	None	6 months after surgery 1st ACTH test;	۲
								insufficient cortisol response; follo-	
								wed by 4 ACTH tests within 6 months.	
								All insufficient cortisol response. Fol-	
								lowed by a CRH tests with a sufficient	
								response.	
F 39	Acromegaly	500	Х	4.1	TTI	570	None	4 months after surgery 2nd CRH test	c
								still insufficient cortisol response;	
								continue HC. 1 yr later ITT; peak cor-	
								tisol 520 nmol/L. 3 yrs later 2nd ITT	
								sufficient stop HC	

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time of		CHK test Peak	HC after		Confir-	Peak			Clinical
Gen- surgery		cortisol	Surgery	Follow-up	mation	cortisol	Other defi-		event
(yrs)	Diagnosis	(nmol/L)	(u/k)	(years)	test	(nmol/L)	ciencies	Follow-up	(u/k)
55	NFA 750 n 2 m	750	۲	2 months	ΤΠ	512	Panhypopit	Panhypopit Before surgery panhypopituitarism; ITT after 2 months insufficiënt	с
50	NFA	690	c	1.7	L	510	Panhypopit	Panhypopit Not all results known when patient left hospital. Appeared to be TSH- and GH deficient. Outpatient follow-up 1st ITT sufficient; 2nd ITT insufficient	۲
19	NFA	780	c	6.5	E	480	Panhypopit	Panhypopit After surgery gonadotrophic deficien- cy; ITT 2002 sufficient; ITT insufficiënt; corticotrope and GH deficiency; start suppletion	۲
50	NFA	640	>	1.8	Ē	410	Panhypopit	Panhypopit After surgery HC ; follow-up after 1 year Metyrapone test insufficient; 1 yr later ITT still insufficient	۲
53	NFA	1100	>	2 months	Mety- rapone test	06	TSH, GHD	Before surgery TSH deficiency and hypocortisolism; Start HC after sur- gery; 2 mnths after surgery ACTH and Metyrapone test both insufficient	c
60	NFA	600	>	6 months	E	200	Panhypopit	Panhypopit Received HC before surgery; continued after surgery; follow-up after 6 months	۲

	Clinical	event	(n/y)	suf- n	. 2 yrs		ł GH n H; 2 yrs	hypo- 2TH		outpa- n		sol n	l mety-	insuf- n	1 by
			Follow-up	Panhypopit 1 yr after surgery new CRH test suf-	ficient. Complaints of tiredness. 2 yrs	later Metyrapone insufficiënt.	Panhypopit After surgery gonadotropic and GH deficiency. After 3 yrs start rhGH; 2 yrs	later low cortisol 24h urine and hypo- thyroidism. Euthyreotic state ACTH	test insufficiënt start HC	Cardial problems; follow-up in outpa-	tient clinic	Based on low basal serum cortisol	levels start HC; 2 days after CRH mety- rapone test insufficient.	Gonadotrope, Several years no complaints no insuf-	ficiency; 2002 ITT GHD followed by
		Other defi-	ciencies	Panhypopit			Panhypopit			None		TSH, DI		Gonadotrope	GHD
	Peak	cortisol	(nmol/L)	190	159		170			120	84.9	06	79.5	06	107
	Confir-	mation	test	Mety-	rapone	test	ACTH 1 ug			Mety-	rapone test	Mety-	rapone test	Mety-	rapone
		Follow-up	(years)	3.4			4.3			3 months		2 days		7	
	HC after	Surgery	(u/k)	С			c			c		>		c	
CHR test	Peak	cortisol	(nmol/L)	670			670			620		610		770	
			Diagnosis	NFA 670 n			NFA			NFA		NFA		NFA	
Age at		Gen- surgery	der (yrs)	57			54			78		77		50	
		Gen-	der	ш			Σ			ш		ш		ш	
				7			œ			6		10		11	

		Age at		CHR test							
		time of		Peak	HC after		Confir-	Peak			Clinical
	Gen-	Gen- surgery		cortisol	Surgery	cortisol Surgery Follow-up	mation	cortisol	Other defi-		event
	der	(yrs)	Diagnosis (nmol/L)	(nmol/L)	(n/n)	(years)	test	(nmol/L)	ciencies	Follow-up	(n/n)
12	ш	67	NFA	600	Х	1.4	TTI	4	Panhypopit	Panhypopit Directly post surgery probably pan-	c
										hypopit because of lesion/rupture pituitary stalk. No basal ACTH pro-	
										duction with stimulation after CRH;	
										Start HC	
13	Σ	38	NFA	797	L	3 months	Basal	06	Panhypopit	Panhypopit Before surgery panhypopit; CRH test	c
							serum			normal. 3 months after surgery com-	
							cortisol			plaints with low basal serum cortisol.	
										Start HC	
14	Σ	39	NFA	660	Х	5 months	ACTH-	300	GHD,	Before surgery panhypopituitarism.	c
							test		gonadotrope	gonadotrope, Probably HC during CRH test after	
									D	surgery.	

GHD, growth hormone deficiency.

Discussion

This study evaluated the postoperative response of cortisol to CRH stimulation in a large cohort of patients after TS for pituitary adenomas compared with the adrenal function assessed during postoperative follow-up. The second adrenal function test documented a normal cortisol response in 31% of the patients with a decreased cortisol response to CRH stimulation directly after surgery. Conversely, the second adrenal stimulation test documented an insufficient cortisol response in 14% of the patients with a normal cortisol response to direct postoperative CRH stimulation. Therefore, the postoperative CRH test does not reliably predict adrenal function after TS for pituitary adenomas in all patients. Nonetheless, our substitution strategy of hydrocortisone guided by the postoperative cortisol responses to CRH did not result in any case of adrenal crises in our patients.

Although CRH stimulation has been incorporated in the diagnostic procedures of ACTH dependent Cushing's syndrome (21–23), reports on the use of CRH stimulation to assess cortisol dependency after transsphenoidal surgery for other pituitary adenomas are scarce. We found three publications that assessed pituitary function using CRH, but these were not specifically in patients after transsphenoidal surgery (8;9). Dullaart *et al.* (9) and Schmidt *et al.* (8) compared the CRH test with basal serum cortisol levels and found no higher diagnostic applicability of the CRH test to basal morning cortisol levels. In contrast, Maghnie *et al.* concluded that the CRH test provided better results than the short Synacthen test (SST) and low-dose short Synacthen test (LDSST), and that CRH may be useful in patients who have a contraindication for ITT (6).

In the current study, the postoperative CRH stimulation test classified 42 of the 144 patients with hypocortisolism. However, 13 of these patients had sufficient adrenal function during follow-up. There are several explanations for these discrepant results. They may be related to differences in cut-off values of the different tests. Regularly accepted

cut-off values (500-550 nmol/L) have been defined for the ITT, which still remains the gold standard test for the assessment of the HPA axis. For the CRH test, some authors have proposed different cut-off values for peak cortisol responses. For example, Schmidt *et al.*(8) reported an optimal peak cortisol cut-off of < 377 nmol/L, yielding a 96% specificity, but poor sensitivity of 76% for the diagnosis of adrenal insufficiency (8). A sensitivity of 100% was reached using a peak cortisol levels of 514 nmol/L (with a specificity of 32%), and 100% specificity with peak cortisol levels of 349 nmol/L (sensitivity 66%). Dullaart et al. found that a peak cortisol value of 420 nmol/L reflected 100% specificity, but 100% sensitivity for the CRH test was only reached using a peak cortisol of 615 nmol/L. Because in our center the CRH test is used as a screening test for hypocortisolism after TS to identify those patients that require hydrocortisone supplementation, we applied a generally accepted stringent criterion of 550 nmol/L. The data indicate that this choice for a higher sensitivity of the CRH test is at the expense of a lower specificity. In other words, using this strategy a higher proportion of patients will be incorrectly diagnosed with adrenal insufficiency. Based on the available literature the use of a cut-off levels of peak cortisol of 514 nmol/L would have resulted in 4/13 patients which would not have been diagnosed with adrenal insufficiency, but with the criteria suggested by Dullaart *et al.* even more patients would have had discrepant results (8;9).

Recovery of preoperative adrenal insufficiency following TS has been described previously (24;25). In a recent study that compared the ITT response at 3 and 12 months after TS, recovery of adrenal function was demonstrated within the first year (26). In agreement, we found a normal function of the HPA axis in eight patients within the first year after surgery who were initially diagnosed as being adrenal insufficient, indicating the necessity of an extensive follow-up in patients after surgery within one year.

In the current study, the postoperative CRH test classified 102 of the 144 patients as having a normal functioning of the HPA axis based on the post-operative CRH test. Fourteen percent of these patients later proved to have hypocortisolism by a second test. These discrepant test results can be potentially life-threatening because these patients are at risk for adrenal crises. It is possible that additional pituitary insufficiencies affected pituitary-adrenal function. Growth hormone and thyroid hormone deficiency can influence these test results. Growth hormone replacement

therapy in patients with GH deficiency may also play an important role because of the influence of GH on the cortisol metabolism. Growth hormone stimulates 11- β hydroxysteroid dehydrogenase (11 β HSD-1), leading to increased cortisol-cortisone interconversion (27). The use of GH replacement therapy in GH deficient patients may therefore unmask cortisol deficiency (28;29). This may also be the case in some of our patients, because their adrenal insufficiency became clear after start of rhGH therapy. Despite all the confounding factors none of our patients had a clinical event.

In conclusion, the CRH test can be safely used to guide hydrocortisone substitution after TS. Nonetheless, the cortisol response to this test cannot reliably predict adrenal function in all patients during longer follow-up after TS. We therefore recommend to perform a second test of pituitary adrenal function during longer follow-up, e.g. 3–6 months after surgery (see Figure 2). This approach is not required in patients with an impaired postoperative cortisol response to CRH, who have multiple pituitary insufficiencies.

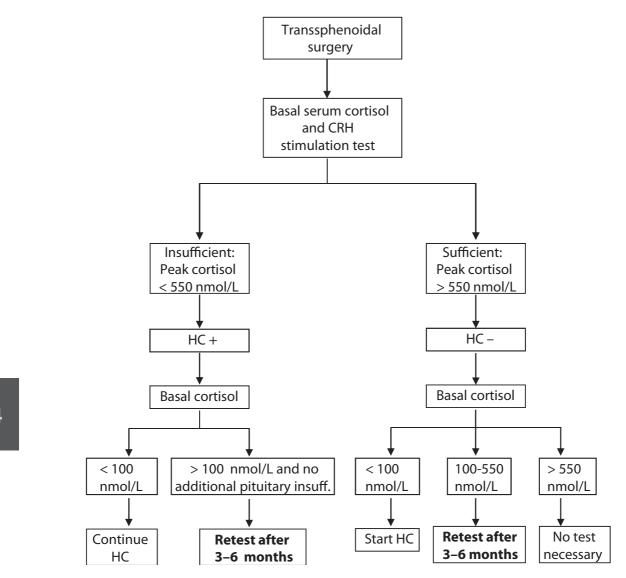


Figure 2. Proposed algorithm for the postoperative follow-up of adrenal function in non ACTH dependent pituitary disease (HC; hydrocortisone)

References

- 1. Gondim JA: Endoscopic endonasal transsphenoidal surgery: surgical results of 228 pituitary adenomas treated in a pituitary center. 2010
- Tabaee A, Anand VK, Barron Y, Hiltzik DH, Brown SM, Kacker A, Mazumdar M, Schwartz TH: Endoscopic pituitary surgery: a systematic review and meta-analysis. J Neurosurg 2009;111:545–554
- Gondim JA, Almeida JP, Albuquerque LA, Schops M, Gomes E, Ferraz T, Sobreira W, Kretzmann MT: Endoscopic endonasal approach for pituitary adenoma: surgical complications in 301 patients. Pituitary 2011;14:174–183
- 4. Fish HR, Chernow B, O'Brian JT: Endocrine and neurophysiologic responses of the pituitary to insulin-induced hypoglycemia: a review. Metabolism 1986;35:763–780
- Grinspoon SK, Biller BM: Clinical review 62: Laboratory assessment of adrenal insufficiency. J Clin Endocrinol Metab 1994;79:923–931
- Courtney CH, McAllister AS, McCance DR, Hadden DR, Leslie H, Sheridan B, Atkinson AB: The insulin hypoglycaemia and overnight metyrapone tests in the assessment of the hypothalamic-pituitary-adrenal axis following pituitary surgery. Clin Endocrinol (Oxf) 2000;53:309–312
- Maghnie M: Evaluation of adrenal function in patients with growth hormone deficiency and hypothalamic-pituitary disorders: comparison between insulin-induced hypoglycemia, lowdose ACTH, standard ACTH and CRH stimulation tests. 2005
- Schmidt IL, Lahner H, Mann K, Petersenn S: Diagnosis of adrenal insufficiency: Evaluation of the corticotropin-releasing hormone test and Basal serum cortisol in comparison to the insulin tolerance test in patients with hypothalamic-pituitary-adrenal disease. J Clin Endocrinol Metab 2003;88:4193–4198
- Dullaart RP, Pasterkamp SH, Beentjes JA, Sluiter WJ: Evaluation of adrenal function in patients with hypothalamic and pituitary disorders: comparison of serum cortisol, urinary free cortisol and the human-corticotrophin releasing hormone test with the insulin tolerance test. Clin Endocrinol (Oxf) 1999;50:465–471
- Lytras N, Grossman A, Perry L, Tomlin S, Wass JA, Coy DH, Schally AV, Rees LH, Besser GM: Corticotrophin releasing factor: responses in normal subjects and patients with disorders of the hypothalamus and pituitary. Clin Endocrinol (Oxf) 1984;20:71–84

- Trainer PJ, Faria M, Newell-Price J, Browne P, Kopelman P, Coy DH, Besser GM, Grossman AB: A comparison of the effects of human and ovine corticotropin-releasing hormone on the pituitary-adrenal axis. J Clin Endocrinol Metab 1995;80:412–417
- 12. Arlt W: Adrenal insufficiency. Clin Med 2008;8:211–215
- 13. Dorin RI, Qualls CR, Crapo LM: Diagnosis of adrenal insufficiency. Ann Intern Med 5-8-2003;139:194–204
- 14. Erturk E, Jaffe CA, Barkan AL: Evaluation of the integrity of the hypothalamic-pituitary-adrenal axis by insulin hypoglycemia test. J Clin Endocrinol Metab 1998;83:2350–2354
- 15. Jacobs HS, Nabarro JD: Tests of hypothalamic-pituitary-adrenal function in man. Q J Med 1969;38:475–491
- Abdu TA, Elhadd TA, Neary R, Clayton RN: Comparison of the low dose short synacthen test (1 microg), the conventional dose short synacthen test (250 microg), and the insulin tolerance test for assessment of the hypothalamo-pituitary-adrenal axis in patients with pituitary disease. J Clin Endocrinol Metab 1999;84:838–843
- Mayenknecht J, Diederich S, Bahr V, Plockinger U, Oelkers W: Comparison of low and high dose corticotropin stimulation tests in patients with pituitary disease. J Clin Endocrinol Metab 1998;83:1558–1562
- Nye EJ, Grice JE, Hockings GI, Strakosch CR, Crosbie GV, Walters MM, Jackson RV: Comparison of adrenocorticotropin (ACTH) stimulation tests and insulin hypoglycemia in normal humans: low dose, standard high dose, and 8-hour ACTH-(1–24) infusion tests. J Clin Endocrinol Metab 1999;84:3648–3655
- 19. Fiad TM, Kirby JM, Cunningham SK, McKenna TJ: The overnight single-dose metyrapone test is a simple and reliable index of the hypothalamic-pituitary-adrenal axis. Clin Endocrinol (Oxf) 1994;40:603–609
- 20. Staub JJ, Noelpp B, Girard J, Baumann JB, Graf S, Ratcliffe JG: The short metyrapone test: comparison of the plasma ACTH response to metyrapone and insulin-induced hypoglycaemia. Clin Endocrinol (Oxf) 1979;10:595–601
- 21. Arnaldi G, Angeli A, Atkinson AB, Bertagna X, Cavagnini F, Chrousos GP, Fava GA, Findling JW, Gaillard RC, Grossman AB, Kola B, Lacroix A, Mancini T, Mantero F, Newell-Price J, Nieman LK, Sonino N, Vance ML, Giustina A, Boscaro M: Diagnosis and complications of Cushing's syndrome: a consensus statement. J Clin Endocrinol Metab 2003;88:5593–5602
- 22. Lindsay JR, Nieman LK: Differential diagnosis and imaging in Cushing's syndrome. Endocrinol Metab Clin North Am 2005;34:403–21, x
- 23. Newell-Price J, Bertagna X, Grossman AB, Nieman LK: Cushing's syndrome. Lancet 13-5-2006;367:1605–1617
- 24. Arafah BM: Reversible hypopituitarism in patients with large nonfunctioning pituitary adenomas. J Clin Endocrinol Metab 1986;62:1173–1179

- 25. Webb SM, Rigla M, Wagner A, Oliver B, Bartumeus F: Recovery of hypopituitarism after neurosurgical treatment of pituitary adenomas. J Clin Endocrinol Metab 1999;84:3696–3700
- 26. Berg C, Meinel T, Lahner H, Mann K, Petersenn S: Recovery of pituitary function in the latepostoperative phase after pituitary surgery: results of dynamic testing in patients with pituitary disease by insulin tolerance test 3 and 12 months after surgery. Eur J Endocrinol 2010;162:853–859
- 27. Moore JS, Monson JP, Kaltsas G, Putignano P, Wood PJ, Sheppard MC, Besser GM, Taylor NF, Stewart PM: Modulation of 11beta-hydroxysteroid dehydrogenase isozymes by growth hormone and insulin-like growth factor: in vivo and in vitro studies. J Clin Endocrinol Metab 1999;84:4172–4177
- 28. Giavoli C, Libe R, Corbetta S, Ferrante E, Lania A, Arosio M, Spada A, Beck-Peccoz P: Effect of recombinant human growth hormone (GH) replacement on the hypothalamic-pituitaryadrenal axis in adult GH-deficient patients. J Clin Endocrinol Metab 2004;89:5397–5401
- 29. Rodriguez-Arnao J, Perry L, Besser GM, Ross RJ: Growth hormone treatment in hypopituitary GH deficient adults reduces circulating cortisol levels during hydrocortisone replacement therapy. Clin Endocrinol (Oxf) 1996;45:33–37

Chapter 5

Low incidence of adrenal insufficiency after transsphenoidal surgery in patients with acromegaly: a long-term follow-up study

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Abstract

Context: The long-term prevalence of adrenal insufficiency after transsphenoidal surgery for GH secreting pituitary adenomas is unknown. However, recently a single study reported a high prevalence of adrenal insufficiency in acromegalic patients after surgical and/or medical treatment without postoperative radiotherapy.

Objective: The objective of the study was to assess the prevalence and incidence rates of adrenal insufficiency in consecutive patients during long-term follow-up after successful transsphenoidal surgery for acromegaly.

Design: In 91 consecutive patients in remission after transsphenoidal surgery only, we retrospectively reviewed insulin tolerance tests, CRH stimulation tests, metyrapone tests, and ACTH stimulation tests used to assess corticotrope function.

Results: Early postoperatively, insufficient adrenal function was observed in 16 patients (18%), which was transient in eight and irreversible in eight other patients in the first year of postoperative follow-up. Therefore, after the first year, the prevalence of adrenal insufficiency was 9%. Late, newonset adrenal insufficiency developed in only three patients 13, 18, and 24 yr after surgery. The incidence rate of late adrenal insufficiency after successful surgery was 2/1000 person-years. After long-term follow-up, a median of 8.1 (1–31 yr), the prevalence of secondary adrenal insufficiency was 12% in patients in remission after surgery for acromegaly.

Conclusion: The prevalence of adrenal insufficiency 1 yr after surgery was 9%, whereas during prolonged follow-up, the incidence rate of adrenal insufficiency was only 2/1000 person-years in patients in remission after surgery. Therefore, development of late-onset adrenal insufficiency is a very infrequent complication in patients with acromegaly in remission after transsphenoidal surgery only.

Introduction

Acromegaly is a chronic disabling disease caused by a GH-producing pituitary adenoma. Transsphenoidal surgery is the curative treatment of choice and somatostatin analogs or radiotherapy is given as needed (1). Hypopituitarism, requiring replacement therapy, can be present postoperatively as a result of surgical or additional radiotherapy. Pituitary irradiation induces hypopituitarism in 50%-75% of the patients after 10–20 yr of follow-up (2). The prevalence of late-onset hypopituitarism in surgically treated patients is not precisely known, but it is generally considered to occur infrequently. The 2009 guidelines on management of acromegaly state that pituitary function should be assessed three months after surgery and that if a dynamic evaluation reveals normal function, there is no need for repeated dynamic function tests unless a patient receives radiotherapy or has clinical symptoms of hypopituitarism (1). Recently, however, Ronchi et al. (3) evaluated adrenal function using the low-dose 1 µg ACTH stimulation test in 36 patients with acromegaly treated by surgery with or without somatostatin analog treatment or by primary medical treatment with somatostatin analogs. A cut-off value for cortisol of 500 nmol/L was used to demonstrate normal adrenal function. They reported a high prevalence of adrenal insufficiency in 32% of patients after a median duration of follow-up of 6 yr after surgery and eventually somatostatin analog treatment. The authors concluded that hypothalamic-pituitary-adrenal (HPA) axis function may worsen over time and should be carefully monitored by dynamic testing in all acromegalic patients, independently from the type of treatment. This recommendation has obvious implications for the long-term management of nonirradiated patients with acromegaly (3). However, the high prevalence of HPA axis insufficiency in surgically treated acromegalic patients is not yet confirmed by others. Therefore, the aim of the present study was to evaluate the prevalence of adrenal insufficiency during long-term follow-up in an unselected cohort of consecutive patients in remission of GH excess by transsphenoidal surgery in our hospital during the period of 1979–2003.

Patients and Methods

Patients

For this study, all 164 consecutive patients, diagnosed with acromegaly and treated at the Leiden University Medical Center, a tertiary referral center, by transsphenoidal surgery between 1979 and 2003 were reviewed. For the purpose of this study, we excluded patients, who were additionally treated by radiotherapy (n=59) as well as patients who had persistent active disease after surgery (n=14). Consequently, 91 patients were included. The diagnosis of acromegaly was based on clinical characteristics and confirmed by insufficient suppression of GH levels after an oral glucose tolerance test. All patients had careful preoperative and postoperative biochemical evaluation. Criteria for cure were serum GH less than 2.5 μ g/L, normal glucose-suppressed GH levels (<1.25 μ g/L for the RIA and $<0.38 \ \mu g/L$ for the immunofluorometric assay), and normal IGF-I values for age. During postoperative follow-up, serum GH, glucose-suppressed GH levels, and IGF-I values were measured at yearly intervals. The surgical results of the complete cohort have been reported previously (4). Data regarding clinical and biochemical characteristics, treatment, and pituitary function were available from all patients. HPA axis function was routinely studied early postoperatively (7–10 days postoperatively), using the CRH test or the insulin tolerance test (ITT). Thereafter nonstimulated morning cortisol measurements were performed at yearly follow-up visits, and dynamic tests to assess corticotrope function were performed at increasing nonstandardized follow-up intervals. The Medical Ethical Committee approved the analysis of treatment results in patients with acromegaly, and no informed consent was required for this retrospective analysis.

Methods

We retrospectively evaluated HPA axis function in the total, unselected cohort of patients in remission after surgery to exclude a potential selection bias. None of the patients received pharmacological treatment

for acromegaly. We reviewed all available dynamic tests in our database performed to evaluate corticotrope function. We considered the ITT as the gold standard test. If ITT results were not available, other stimulation tests like the CRH, ACTH, and metyrapone tests were evaluated. In addition, basal morning cortisol values were collected. Patients were considered to have adrenal insufficiency if they had biochemically confirmed insufficiency (see below). All patients had an endocrine assessment every year. The use of hormone stimulation tests changed during the follow-up period. Initially, ITT was used early postoperatively and during follow-up. After the clinical introduction of the CRH and GHRH test, the ITT lost its leading position in the screening for somatotrope and corticotrope deficiencies for obvious reasons. From 1990 onward, according to protocol, the CRH test was performed early postoperatively to assess whether corticotrope function was sufficient to discharge patients without hydrocortisone replacement, and confirmation tests were performed at the outpatient department.

The metyrapone test was used for follow-up assessment in patients with contraindication for ITT. In recent years, 1 μ g ACTH tests were performed to screen for corticotrope deficiency in late follow-up. However, the ITT remained the gold standard for confirmation of adrenal insufficiency, especially if other test revealed borderline results (in patients without a contraindication for ITT).

Evaluation of HPA axis

An insulin tolerance test (insulin 0.1 IE/kg, Actrapid; Novo Nordisk, Bagsvaerd, Denmark) was administered i.v. in the postabsorptive state between 0800 and 0900 h to induce hypoglycemia (< 2.2 mmol/L). Cortisol was measured at -15, 0, 15, 30, 45, 60, 90, and 120 min. A cutoff value of cortisol greater than 550 nmol/L was used to define normal function of the HPA axis (5–9). An ACTH stimulation test (ACTH 1µg Synacthen[®]; Novartis Pharma B.V., Arnhem, The Netherlands) was administered i.v. between 0800 and 0900 h after blood samples had been taken at -15 and 0 min for measurement of cortisol values. The response of cortisol to ACTH was assessed in a single blood sample obtained 30 min after ACTH injection. A cut-off value of cortisol greater than 550 nmol/L was used to define normal adrenal function (10–12).

CRH test (CRH 100 μ g; Ferring B.V., Hoofddorp, The Netherlands) was administered in the postabsorptive state between 0800 and 0900 h.

Cortisol and ACTH were measured at -15, -5, 15, 30, 45, and 60 min. A cut-off value for cortisol of greater than 550 nmol/Liter was used to define normal function (13, 14). A Metyrapone test (metyrapone 30 mg/ kg, Metopiron; Novartis Pharma B.V., Arnhem, The Netherlands) was administered at midnight. The next morning postabsorptive blood samples were obtained for measurement of 11-deoxycortisol, cortisol, and ACTH levels. A cut-off value for 11-deoxycortisol of 200 nmol/L was used to define normal adrenal function (15–17).

For morning cortisol, blood was sampled between 0800 and 0900 h for assessment of cortisol values. A cut-off value of cortisol greater than 500 nmol/L was used to define normal function only in case dynamic tests were not available. Premenopausal women were tested after stopping estrogen replacement for 3 months.

Assays

Cortisol was measured between 1978 and 1986 by in-house RIA with an interassay coefficient of variation (CV) of 10% and with a detection limit of 5 nmol/L. Between 1986 and 1994, cortisol was measured by fluorescence energy-transfer immunoassay (Syva-Advance; Syva Co., Palo Alto, CA) with an interassay variation coefficient of 3.6-6.1% and a detection limit of 0.05 µmol/L. From 1994, cortisol was measured by fluorescence polarization assay on a TDx (Abbott Laboratories, Abbott Park, IL). The interassay variation coefficient is 5–6% above 0.50 µmol/L and amounts to 12% under 20 nmol/L. The detection limit was 2 nmol/L. Before 1993 GH was measured by RIA (Biolab; Serona, Coissins, Switzerland), calibrated against World Health Organization international reference preparation 66/21 (detection limit: 0.5 mU/L, with an interassay CV less than 5%; for the conversion of micrograms per liter to milliunits per liter, multiply by 2). After 1993 serum GH concentration was measured with a sensitive time-resolved fluoroimmunoassay (Wallac, Turku, Finland). The assay is specific for 22 kDa GH. The standard was recombinant human GH (Genotropin; Pharmacia & Upjohn, Uppsala, Sweden), which was calibrated against the World Health Organization First International Reference Preparation 80/505 (to convert milliunits per liter to micrograms per liter, divide by 2.6) (18). The limit of detection (defined as the value 2 SD above the mean value of the zero standard) was 0.03 mU/L (0.0115 μ g/L). The intraassay CV varied from 1.6 to 8.4% in the assay range 0.26–47 mU/L, with corresponding interassay CV of 2.0–9.9%. Until 2005 serum IGF-I concentrations were determined by RIA (Incstar, Stillwater, MN) with a detection limit of 1.5 nmol/L and an interassay CV below 11%. IGF-I is expressed as SD scores for ageand gender-related normal levels determined in the same laboratory (18). From 2005 onward, serum IGF-I concentration (nanograms per milliliter) was measured with an immunometric technique on the Immulite 2500 system (Diagnostic Products Corp., Los Angeles, CA). The intraassay CV was 5.0 and 7.5% at mean serum concentrations of 8 and 75 nmol/L, respectively. The IGF-I concentration was expressed as SD score, using the λ - μ - σ smoothed reference curves based on measurements in 906 healthy individuals (19).

Statistical analysis

All results are shown as mean±SD. Descriptive statistics were calculated. Student's t-tests were used when appropriate. P < 0.05 was considered to be statistically significant. Duration of follow-up in person-years was calculated for all patients as time between surgery until July 1, 2010, if patients were followed-up in our center, until date of secondary treatment in case of a recurrence, until last visit if patients were lost to follow-up, or until date of death in case patients had died. Incidence rates were calculated using number of cases divided over person-years of follow-up. Analyses were performed by SPSS package (version 16.0.2, 2008; SPSS, Chicago, IL).

Results

Baseline characteristics

We included 91 consecutive patients, 49 male and 42 female patients cured by transsphenoidal adenomectomy for a GH producing pituitary adenoma (Table 1). The mean age at the time of surgery was 46.8 \pm 12.3 yr (range 18–76 yr). The mean disease duration before surgery was 9.0 \pm 8.0 yr. Twenty-eight patients had a microadenoma (31%), 55 had a noninvasive macroadenoma (60%), and eight had an invasive macroadenoma (9%). Mean GH preoperative concentrations and IGF-I SD scores decreased significantly after surgery (*P* < 0.001, Table 1). Mean GH concentrations were 0.8 \pm 1.0 µg/L, and IGF-I SD scores were 0.8 \pm 2.1 at the end of follow-up.

Immediate postoperative assessment of adrenal function

Seven to 10 days after surgery, assessment of adrenal function was performed by CRH test (49%), ITT (43%), or by basal cortisol level in a minority of patients (2%). Three patients (3%) were not retested postoperatively because they had been cortisol dependent preoperatively. Data were missing in 2%. Sufficient adrenal function was observed in 36 of 44 patients according to the results of the CRH test, 35 of 39 patients according to ITT, and one of two patients according to basal cortisol. Thus, early postoperative adrenal insufficiency was observed in 16 patients (18%) including the three patients with preoperative secondary insufficiency. Hydrocortisone replacement was prescribed to 11 patients.

	Patients (n=91)
M/F	49/42
Age at TNS (yrs)	46.8 ± 12.3 (range: 18–76)
Disease duration (yrs)	9.0 ± 8.0
Tumorclass (n(%))	
Class 1 – Microadenoma	28 (31%)
Class 2 – Non-invasive macroadenoma	55 (60%)
Class 3 – Invasive macroadenoma	8 (9%)
Preoperative GH (μg/L)	23.1 ± 27.1
Postoperative GH (μg/L)	0.9 ± 0.8
Follow-up GH (μg/L)	0.8 ± 1.0
Preoperative IGF-I SD	7.4 ± 4.2
Postoperative IGF-I SD	0.9 ± 1.9
Follow-up IGF-I SD	0.8 ± 2.1
Follow-up	
Pituitary deficiencies	
TSH	8 (9%)
LH/FSH	11 (12%)
GH	8 (14%)*

Table 1. Baseline characteristics and follow-up characteristics of patients with acromegaly cured by transsphenoidal surgery

F, female; M, male; TNS, transnasosphenoidal surgery

Data are shown as mean \pm SD unless mentioned otherwise. Significant decrease following surgery (P <0.001) for both GH and IGF-I SD. No difference between postoperative and follow-up concentations (P=ns). *Assessed in 58 patients by ITT.

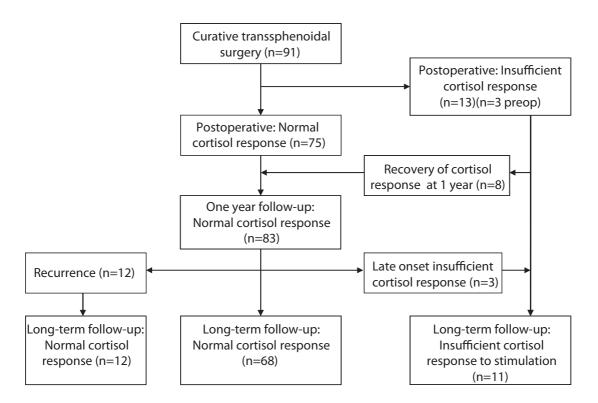


Figure 1. Flowchart of corticotrope function during long-term follow-up

Adrenal function 1 yr after surgery

At 1 yr postoperatively, the prevalence of adrenal insufficiency was 9% (eight of 91), three patients being diagnosed preoperatively and five patients diagnosed early postoperatively. These eight patients received hydrocortisone replacement therapy. Adrenal insufficiency was diagnosed by insufficient response to ITT (n=3), CRH (n=3), metyrapone test (n=1), or low basal cortisol of 20 nmol/L (n=1).

In the eight other patients with an early postoperative insufficient cortisol response to dynamic testing (CRH or ITT), retesting within the first year revealed normal adrenal function. The results of postoperative tests and follow-up tests in these patients are detailed in Table 2.

Adrenal function during prolonged follow-up

During prolonged follow-up, 262 ITT, 110 CRH tests, and 67 ACTH tests were performed in the patients cured by transsphenoidal surgery. Twelve patients with initial cure developed a recurrence of acromegaly (13%) a median of 8.7 yr (range 1.2–24.6 yr) after surgery. Consequently, 79 patients were in long-term cure after surgery only. Patients with

a recurrence of acromegaly were followed up until the date of the secondary treatment for the present analysis. At late evaluation (n=91), a median of 8.1 yr (1-31 yr) postoperatively, corticotrope function was assessed by ITT (67%), CRH (7%), ACTH (10%), or basal cortisol levels greater than 0.50 µmol/Liter (6%), whereas there were no data available in 10% of patients due to death (n=3) or loss to follow-up (n=6). During long-term follow-up of 1489 person-years, lateonset adrenal insufficiency developed in three patients (3%) 13, 18, and 24 yr postoperatively. The incidence rate for new-onset adrenal insufficiency was 2/1000 personyears. The clinical characteristics of these patients with late-onset adrenal insufficiency and their presenting symptoms of late adrenal deficiency are detailed in Table 3. All three patients had been treated for a noninvasive macroadenoma. Two patients had complaints of tiredness, dizziness, and/or general malaise. The third patient presented with unexplained hypoglycemias. Due to high age, no ITT was performed, but the ACTH test revealed insufficient response of cortisol. All patients improved clinically after replacement therapy. Thus, at the end of followup, adrenal insufficiency was present in 12% (11 of 91) of the patients in remission after transsphenoidal surgery. All these patients required hydrocortisone replacement therapy.

at Postopera- (mol/L)cortisol cortisolFollow-upyr)tive test(µmol/L)Postoperative follow-up*testCRH0.260.34Temporary hydrocortisone replacementCRHTTT0.160.42No Hydrocortisone replacement. Test and basal cortisol considered adequate.basal cortisolTTT0.130.47No Hydrocortisone replacement. Test and basal cortisone replacement.ITTCRH0.320.48No Hydrocortisone replacementITTCRH0.320.48No Hydrocortisone replacementITTCRH0.320.48No Hydrocortisone replacementITTCRH0.300.50Temporary hydrocortisone replacementITTCRH0.370.50Temporary hydrocortisone replacementITTCRH0.370.50Temporary hydrocortisone replacementITTCRH0.370.52Temporary hydrocortisone replacementITTCRH0.430.53Hydrocortisone replacementITTCRH0.370.52Temporary hydrocortisone replacementITTCRH0.370.53Hydrocortisone replacementITTCRH0.430.53Hydrocortisone replacementITTCRH0.370.53Hydrocortisone replacementITTCRH0.430.53Hydrocortisone replacementITTCRH0.430.53Hydrocortisone replacementITTCRH0.430.53Hydrocortisone replace				Basal	Peak			Basal	Peak
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CRH 0.37 0.52 Temporary hydrocortisone replacement ITT CRH 0.43 0.53 Hydrocortisone if needed. ACTH	ш	39	LT	0.30	0.50	Temporary hydrocortisone replacement	Ξ	0.15	0.57
CRH 0.43 0.53 Hvdrocortisone if needed. ACTH	Z	52	CRH	0.37	0.52	Temporary hydrocortisone replacement	Ħ	0.25	0.71
	Μ	42	CRH	0.43	0.53	Hydrocortisone if needed.	ACTH	0.42	0.62

CHAPTER 5

							Test d	Test diagnose		
			Postop-	basal	РеаК				1	pituitary
Gen-	Gen- Age at Tumor- erative Cortisol	Tumor-	erative	Cortisol	cortisol		Stimulation		Start HC deficien-	deficien-
der	TSS (yr)	TSS (yr) class*	test	(J/lomd)	(hmol/L)	(µmol/L) Clinical follow-up	test	Results	after (yr) cies	cies
Σ	33	7	E	0.09	0.55	Tiredness, malaise, chest pain, paresthesias	Metyrapone	11-DOC 44.3 cort 0.13	24	None
ш	43	7	Ħ	0.42	0.62	Headache, tiredness, astenia Yearly follow-up: basal cortisol 0.02 nmol/L	#		13	GH-TSH
ш	67	7	ΤΠ	0.34	0.60	DM II Unexplained hypoglycemia	ACTH -test**	ACTH -test** Basal cort 0.16 Peak cort 0.47	18	GH-LH/ FSH

Characteristics of patients cured for acromegaly with late onset adrenocortical insufficiency Table 3.

F, female; FSH, follicle stimulating hormone; GH, growth hormone; HC, hydrocortisone; ITT, insulin tolerance test; LH, luteinizing hormone; M, male; TSS, transsphenoidal surgery; TSH, thyroid stimulating hormone

* Tumorclass 2 – non-invasive macroadenoma

#Based on low basal serum cortisol no stimulation test was performed.

** Due to high age at, no ITT was performed to confirm adrenal insufficiency

Discussion

The present study documents that new-onset adrenal insufficiency after successful surgical treatment for acromegaly in follow-up is not frequently observed. The prevalence of adrenal insufficiency was 9% 1 yr after surgery. In our well-characterized cohort of consecutive patients in remission after transsphenoidal surgery, the incidence rate of new-onset late adrenal insufficiency was only 2/1000 person-years during a long-term clinical follow-up. In accordance with the study by Ronchi *et al.* (3), our study demonstrates that HPA axis function may worsen over time, but adrenal insufficiency is an infrequent complication in patients in remission of acromegaly after surgery.

The discrepancies in the prevalence of adrenal insufficiency between the current study and the study by Ronchi *et al.* (3) may be explained by differences in study design and study population. In the study by Ronchi *et al.*, 16 of 36 patients had neuroradiological evidence of residual postoperative tumor remnants, and 16 were treated by somatostatin analogs. In addition, the authors used the low-dose ACTH stimulation test, which may lead to a false-negative response in 10% of healthy subjects (20).

Furthermore, Ronchi *et al.* used a cut-off value for cortisol of 500 nmol/L for the evaluation of the HPA axis, whereas we used mainly the ITT and CRH test with a cut-off value of 550 nmol/L. The ITT is generally regarded as the gold standard (7;21). Alternatively, the differences between the two studies may also be caused by patient selection and differences in surgical techniques. Ronchi *et al.* (3) observed a remarkably high prevalence of adrenal insufficiency in 32% of the patients treated by surgery and/or somatostatin analogs for acromegaly. However, only 16% of their patients required substitution therapy with hydrocortisone, in agreement with our data.

Potential caveats in our study include changed cortisol binding globulin (CBG) levels in acromegaly and the presence of postoperative GH deficiency. However, studies on the effect of GH on serum CBG concentration are controversial. Some authors reported that GH administration in hypopituitary patients decrease CBG levels by approximately 20% (22–24), but other studies in larger cohorts did not observe a difference in CBG levels during GH treatment (25;26). Data on CBG concentrations in patients with active acromegaly are also scarce. One study investigated the effect of pegvisomant treatment on cortisol metabolism (27). These authors did not observe any change in serum CBG concentrations, although the majority of the patients reached normal IGF-I levels. Collectively these data indicate that the effect of GH on serum CBG levels is not unequivocal, especially in GH-deficient patients, and an increase in CBG concentrations after GH normalization in acromegaly has not been demonstrated.

GH has a strong impact on cortisol metabolism by its action on 11β-hydroxysteroid dehydrogenase, leading to increased cortisolcortisone interconversion (28). For instance, GH replacement therapy in GH-deficient patients may unmask cortisol deficiency (23;26). In our study, untreated GH deficiency in the presence of a normal corticotrope function was present in only three of 58 patients (5%) with available GH measurements during ITT (data not shown), suggesting that it is unlikely that this significantly affected our results. Patients underwent surgery on a low-dose dexamethasone scheme, which may have influenced the test results, leading to overestimation of adrenal insufficiency shortly after surgery. Indeed, in eight patients with suboptimal cortisol response to ITT or CRH postoperatively, adrenal function normalized within 1 yr. This observation is in accordance with a recent study that compared the ITT response at 3 and 12 months postoperatively (29). In that study, cortisol peak responses increased by 17% and adrenal function had recovered in four of 20 patients with an insufficient cortisol peak response directly after surgery.

Recovery of preoperative adrenal insufficiency after transsphenoidal surgery has been described previously (30;31). Therefore, early postoperative testing may not reflect the definitive outcome of adrenal function. The outcome of these tests can be influenced by incipient GH or thyroid hormone deficiency, as discussed above (32). This observation emphasizes the importance of repeated dynamic tests also in patients with early postoperative insufficient response to adrenal function tests. After 1 yr, less frequent control of dynamic pituitary function may suffice in those patients with a confirmed normal adrenal function. A recent meta analysis of 12 studies on the diagnostic value of basal cortisol values using summary receiver-operating characteristic curves showed an area under the curve of 0.79 (95% confidence interval 0.75–0.82). A lower cut-off value for basal cortisol less than 140 nmol/L (likelihood ratio > 9) was used to diagnose hypoadrenalism and an upper cut-off value of greater than 370 nmol/L (likelihood ratio < 0.15) was used to exclude hypoadrenalism. To be eligible for inclusion in this study, adult and pediatric subjects had to be suspected of adrenal insufficiency from pituitary disease longer than four wk from prolonged exogenous glucocorticoid administration. Only studies with ITT or metyrapone test as a reference test were included (33). It seems reasonable to use the guidelines as proposed by these authors. Thus, in patients with a basal serum cortisol greater than 370 nmol/L without complaints, the likelihood to have adrenal insufficiency is very low, and screening using basal cortisol may suffice in asymptomatic patients.

We had the opportunity to review long-term follow-up data in a carefully followed cohort and to have the availability of multiple tests in the vast majority of patients in the presence of few missing data. Nonetheless, limitations of our study are the retrospective nature of the study and the fact that patients had been tested using different cortisol stimulation tests and assays. However, this does not affect our conclusions because the ITT, CRH, and metyrapone tests are all accepted tests for the evaluation of HPA function, and we have used unchanged cut-off values of cortisol throughout the years. Nevertheless, several reports suggest that the sensitivity of the CRH test is less than that of the ITT (34–38). This conclusion is partly related to the cut-off value of CRH-stimulated cortisol responses. Unfortunately, there are no large studies of the CRH test in healthy subjects across ages, body mass index, and gender.

Therefore, we used a restrictive approach and retested subjects with an insufficient response to CRH by ITT. The CRH test may not detect hypothalamic insufficiency, whereas the ITT is a test for the hypothalamus-corticotrope- adrenal cortex ensemble. However, we have no a priori reason to assume hypothalamic damage in our patients because they had no previous pituitary irradiation or very large tumors impinging on the hypothalamus. Furthermore, our findings are strengthened by the consecutive nature of the patient series and the yearly assessment of the pituitary function. For the evaluation we used preferably results of ITT. However, ongoing follow-up after the last ITT did not raise the suspicion of new adrenal deficiencies.

The recurrence of GH overproduction after initial cure by surgery may be due to either regrowth of residual tumor tissue or true recurrence (38-42). In our series we retested all patients cured by surgery at regular intervals. In our experience, recurrence of GH excess may occur, even after many years of postoperative cure documented by repeatedly normal IGF-I levels and normal GH nadir responses during the glucose tolerance test. This was also true after 1993, when a more sensitive GH assay was introduced. Even though this sequence of events may not exclude recurrence from persistent, but apparently longtime subclinical, postoperative adenomatous tissue, this observation indicates that the recurrence rate in our series is not merely the consequence of persistent postoperative disease. Moreover, in patients retested after surgical cure with regular intervals, we have previously documented that biochemical recurrence of GH excess after initial surgical cure clearly precedes radiological recurrence (42). Therefore, even in cases with biochemical recurrence of GH excess, it is highly unlikely that mass effects of adenomatous tissue were present. Finally, in the present study, we included the patients until the start of additional treatment of GH excess in the case of recurrent disease. Therefore, it is unlikely that recurrences of GH excess after years of biochemical remission affect our conclusions.

In conclusion, the incidence rate of late-onset adrenocortical insufficiency after successful surgery for acromegaly is very low (2/1000 person-years). We propose to repeat the dynamic test of HPA function 1 yr after surgery in patients with postoperative HPA insufficiency. Further research is required to assess whether yearly basal cortisol values may suffice to monitor adrenal function in asymptomatic patients. However, in case of low basal cortisol levels or symptoms suggestive of corticotrope insufficiency, additional dynamic testing should be performed.

References

- Melmed S, Colao A, Barkan A, Molitch M, Grossman AB, Kleinberg D, Clemmons D, Chanson P, Laws E, Schlegte J, Vance ML, Ho K, Giustina A; Acromegaly Consensus Group 2009 Guidelines for acromegaly management: an update
- 2. Melmed S 2006 Medical progress: Acromegaly. N Engl J Med 355:2558–2573
- Ronchi CL, Ferrante E, Rizzo E, Giavoli C, Verrua E, Bergamaschi S, Lania AG, Beck-Peccoz P, Spada A 2008 Long-term basal and dynamic evaluation of hypothalamic-pituitary-adrenal (HPA) axis in acromegalic patients. Clin Endocrinol (Oxf) 69:608–612
- Biermasz NR, Dekker FW, Pereira AM, van Thiel SW, Schutte PJ, van Dulken H, Romijn JA, Roelfsema F 2004 Determinants of survival in treated acromegaly in a single center: predictive value of serial insulin-like growth factor I measurements. J Clin Endocrinol Metab 89:2789–2796
- 5. Erturk E, Jaffe CA, Barkan AL 1998 Evaluation of the integrity of the hypothalamic-pituitaryadrenal axis by insulin hypoglycemia test. J Clin Endocrinol Metab 83:2350–2354
- 6. Jacobs HS, Nabarro JD 1969 Tests of hypothalamic-pituitary-adrenal function in man. Q J Med 38:475–491
- 7. Fish HR, Chernow B, O'Brian JT 1986 Endocrine and neurophysiologic responses of the pituitary to insulin-induced hypoglycemia: a review. Metabolism 35:763–780
- 8. Arlt W 2008 Adrenal insufficiency. Clin Med 8:211–215
- 9. Dorin RI, Qualls CR, Crapo LM 2003 Diagnosis of adrenal insufficiency. Ann Intern Med 139:194–204
- 10. Abdu TA, Elhadd TA, Neary R, Clayton RN 1999 Comparison of the low dose short synacthen test (1 microg), the conventional dose short synacthen test (250 microg), and the insulin tolerance test for assessment of the hypothalamo-pituitary-adrenal axis in patients with pituitary disease. J Clin Endocrinol Metab 84:838–843
- Mayenknecht J, Diederich S, Bahr V, Plockinger U, Oelkers W 1998 Comparison of low and high dose corticotropin stimulation tests in patients with pituitary disease. J Clin Endocrinol Metab 83:1558–1562
- Nye EJ, Grice JE, Hockings GI, Strakosch CR, Crosbie GV, Walters MM, Jackson RV 1999 Comparison of adrenocorticotropin (ACTH) stimulation tests and insulin hypoglycemia in normal humans: low dose, standard high dose, and 8-hour ACTH-(1–24) infusion tests. J Clin Endocrinol Metab 84:3648–3655

- Lytras N, Grossman A, Perry L, Tomlin S, Wass JA, Coy DH, Schally AV, Rees LH, Besser GM 1984 Corticotrophin releasing factor: responses in normal subjects and patients with disorders of the hypothalamus and pituitary. Clin Endocrinol (Oxf) 20:71–84
- Trainer PJ, Faria M, Newell-Price J, Browne P, Kopelman P, Coy DH, Besser GM, Grossman AB 1995 A comparison of the effects of human and ovine corticotropin-releasing hormone on the pituitary-adrenal axis. J Clin Endocrinol Metab 80:412–417
- Courtney CH, McAllister AS, McCance DR, Hadden DR, Leslie H, Sheridan B, Atkinson AB 2000 The insulin hypoglycaemia and overnight metyrapone tests in the assessment of the hypothalamic-pituitary-adrenal axis following pituitary surgery. Clin Endocrinol (Oxf) 53:309–312
- Fiad TM, Kirby JM, Cunningham SK, McKenna TJ 1994 The overnight single-dose metyrapone test is a simple and reliable index of the hypothalamic-pituitary-adrenal axis. Clin Endocrinol (Oxf) 40:603–609
- 17. Staub JJ, Noelpp B, Girard J, Baumann JB, Graf S, Ratcliffe JG 1979 The short metyrapone test: comparison of the plasma ACTH response to metyrapone and insulin-induced hypoglycaemia. Clin Endocrinol (Oxf) 10:595–601
- 18. Janssen YJ, Frölich M, Roelfsema F 1997 A low starting dose of genotropin in growth hormonedeficient adults. J Clin Endocrinol Metab 82:129–135
- 19. Rikken B, van Doorn J, Ringeling A, Van den Brande JL, Massa G, Wit JM 1998 Plasma levels of insulin-like growth factor (IGF)-I, IGF-II and IGF-binding protein-3 in the evaluation of childhood growth hormone deficiency. Horm Res 50:166–176
- 20. Wade M, Bald S, Calis K, Raff H, Sinaii N, Nieman L 2010 Technical details influence the diagnostic accuracy of the 1 μg ACTH stimulation test. Eur J Endocrinol; 162:109-113
- 21. Grinspoon SK, Biller BM 1994 Clinical review 62: Laboratory assessment of adrenal insufficiency. J Clin Endocrinol Metab 79:923–931
- 22. Weaver JU, Thaventhiran L, Noonan K, Burrin JM, Taylor NF, Norman MR, Monson JP 1994 The effect of growth hormone replacement on cortisol metabolism and glucorticoid sensitiviyu in hypopituitary adults. Clin Endocrinol (Oxf) 41:639-648
- Rodriguez-Arnao J, Perry L, Besser GM,Ross RJ 1996 Growth hormone treatment in hypopituitary GH deficient adults reduces circulating cortisol levels during hydrocortisone replacement therapy. Clin Endocrinol (Oxf) 45:33–37
- Tschöp M, Lahner H, Feldmeier H, Grasberger H, Morrison KM, Janssen OE, Attanasio AF, Strasburger CJ 2000 Effects of growth hormone replacement therapy on levels of cortisol and cortisol-binding globulin in hypopituitary adults. Eur J Endocrinol 145:769–773
- Isidori AM, Kaltsas GA, Perry L, Burrin JM, Besser GM, Monson JP 2003 The effect of growth hormone replacement therapy on adrenal androgen secretion in adult onset hypopituitarism. Clin Endocrinol (Oxf) 58:601–611

- 26. Giavoli C, Libé R, Corbetta S, Ferrante E, Lania A, Arosio M, Spada A, Beck-Peccoz P 2004 Effect of recombinant growth hormone (GH) replacement on the hypothalamic-pituitary-adrenal axis in adult GH-deficient patients. J Clin Endocrinol Metab 89:5397–5401
- 27. Trainer PJ, Drake WM, Perry LA, Taylor NF, Besser GM, Monson JP 2001 Modulation of cortisol metabolism by the growth hormone receptor antagonist pegvisomant in patients with acromegaly. J Clin Endocrinol Metab 86:2989–2992
- Moore JS, Monson JP, Kaltsas G, Putignano P, Wood PJ, Sheppard MC, Besser GM, Taylor NF, Stewart PM 1999 Modulation of 11beta-hydroxysteroid dehydrogenase isozymes by growth hormone and insulin-like growth factor:invivo and in vitro studies. J Clin Endocrinol Metab 84:4172–4177
- 29. Berg C, Meinel T, Lahner H, Mann K, Petersenn S 2010 Recovery of pituitary function in the late-postoperative phase after pituitary surgery: results of dynamic testing in patients with pituitary disease by insulin tolerance test 3 and 12 months after surgery. Eur J Endocrinol 162:853–859
- Arafah BM 1986 Reversible hypopituitarism in patients with large nonfunctioning pituitary adenomas. J Clin Endocrinol Metab 62:1173–1179
- 31. Webb SM, Rigla M, Wagner A, Oliver B, Bartumeus F 1999 Recovery of hypopituitarism after neurosurgical treatment of pituitary adenomas. J Clin Endocrinol Metab 84:3696–3700
- 32. Filipsson H, Johansson G 2009 GH replacement in adults: interactions with other pituitary hormone deficiencies and replacement therapies. Eur J Endocrinol 161 Suppl 1:S85–95
- Kazlauskaite R, Evans AT, Villabona CV, Abdu TA, Ambrosi B, Atkinson AB, Choi CH, Clayton RN, Courtney CH, Gonc EN, Maghnie M, Rose SR, Soule SG, Tordjman K 2008 Corticotropin tests for hypothalamic-pituitary-adrenal insufficiency: a metaanalysis. J Clin Endocrinol Metab 93:4245–4253
- 34. Dullaart RPF, Pasterkamp SH, Beentjes JAM, Sluiter WJ 1999 Evaluation of adrenal function in patients with hypothalamic and pituiatry disorders: comparison of serum cortisol, urinary free cortisol and the human-corticotrophin releasing hormone test with the insulin tolerance test. Clin Endocrinol (Oxf) 50:465–471
- 35. Lopez Schmidt L, Lahner H, Mann K, Petersenn S 2003 Diagnosis of adrenal insufficiency: evaluation of the corticotropin-releasing hormone test and basal serum cortisol in comparison to the insulin tolerance test in patients with hypothalamic-pituitary-adrenal disease. J Clin Endocrinol Metab 88:4193–4198
- 36. Dickstein G 2003 The assessment of the hypothalamo-pituitary-adrenal axis in pituitary disease: are there short cuts? J Endocrinol Invest 26:25–30
- 37. Grossman AB 2010 Clinical Review: The diagnosis and management of central hypoadrenalism. J Clin Endocrinol Metab 95:4855–4863
- Biermasz NR, van Dulken H, Roelfsema F 2000 Ten-year follow-up results of transsphenoidal microsurgery in acromegaly. J Clin Endocrinol Metab 85:4596–4602

- Giustina A, Barkan A, Casanueva FF, Cavagnini F, Frohman L, Ho K, Veldhuis J, Wass J, Von WK, Melmed S 2000 Criteria for cure of acromegaly: a consensus statement. J Clin Endocrinol Metab 85:526–529
- 40. Kristof RA, Grote A, Redel L, Neuloh G, Klingmuller D, Schramm J 2011 The common consensus criteria have high predictive values for long-term postoperative acromegaly remission. Acta Neurochir (Wien) 153:19–25
- 41. Nomikos P, Buchfelder M, Fahlbusch R 2005 The outcome of surgery in 668 patients with acromegaly using current criteria of biochemical 'cure'. Eur J Endocrinol 152:379–387
- 42. Zirkzee EJ, Corssmit EP, Biermasz NR, Brouwer PA, Wiggers-De Bruine FT, Kroft LJ, Van Buchem MA, Roelfsema F, Pereira AM, Smit JW, Romijn JA 2004 Pituitary magnetic resonance imaging is not required in the postoperative follow-up of acromegalic patients with long-term biochemical cure after transsphenoidal surgery. J Clin Endocrinol Metab 89:4320–4324

Chapter 6

Pituitary dysfunction in adult patients after cranial radiotherapy: a systematic review and meta-analysis

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Abstract

Context: Cranial radiotherapy is an important cause of hypopituitarism. The prevalence of hypopituitarism varies considerably between studies.

Objective: We conducted a systematic review and meta-analysis of reported prevalences of hypopituitarism in adults radiated for nonpituitary tumors.

Data sources: We searched PubMed, EMBASE, Web of Science and the Cochrane Library to identify potentially relevant studies.

Study selection: Studies were eligible for inclusion with following criteria: 1) cranial radiotherapy for nonpituitary tumors and/or total body irradiation for haematological malignancies 2) adult population (>18 yr old) 3) report on endocrine evaluation.

Data extraction: Data review was done by two independent reviewers. Besides extraction of baseline and treatment characteristics, also endocrine tests; definitions and cut-off values used to define pituitary insufficiency, were extracted.

Results: Eighteen studies with a total of 813 patients were included. These included 608 patients treated for nasopharyngeal cancer (75%) and 205 for intracerebral tumors. The total radiation dose ranged from 14 to 83 and 40 to 97 Gy for nasopharyngeal and intracerebral tumors, respectively. The point prevalence of any degree of hypopituitarism was 0.66 [98% confidence interval (CI), 0.55–0.76]. The prevalence of GH deficiency was 0.45 (95% CI, 0.33–0.57), of LH and FSH 0.3 (95% CI, 0.23–0.37), of TSH 0.25 (95% CI, 0.16–0.37), and of ACTH 0.22 (95% CI, 0.15–0.3), respectively. The prevalence of hyperprolactinemia was 0.34 (CI 0.15–0.6) There were no differences between the effects of radiotherapy for nasopharyngeal *vs.* intracerebral tumors.

Conclusion: Hypopituitarism is prevalent in adult patients after cranial radiotherapy for nonpituitary tumors. Therefore, all patients treated by cranial radiotherapy should have structured periodical assessment of pituitary functions.

Introduction

Pituitary insufficiency is a late onset sequel of cranial irradiation for intracerebral and nasopharyngeal tumors or total body radiotherapy for hematological malignancies in children (1-9). In the Childhood Cancer Survivor Study (CCSS) 43% of children treated for cerebral tumors had one or more endocrinopathies (10). Consequently, structured follow-up programs for childhood cancer survivors include endocrine assessments. In the last decades, survival rates of patients treated with cranial radiotherapy for various malignancies as well as for benign tumors have improved substantially by introduction of new surgical, radiotherapeutical, and chemotherapeutical options. In contrast to the long-term survivors of cranial radiotherapy in childhood, endocrine surveillance programs have not been routinely incorporated in adults treated with cranial radiotherapy. The prevalence of hypopituitarism after cranial radiotherapy is affected by several factors. First, the time interval between radiotherapy and the assessment of pituitary function is important because the development of pituitary failure is likely to increase in time after radiotherapy (11–13). Second, hypothalamic and pituitary insufficiencies are more likely to develop with increasing radiation exposure (10;14). Finally, methodological differences between the studies with respect to endocrine evaluation, like the use of different endocrine tests with different criteria for pituitary insufficiency, will also affect the prevalence of hypopituitarism.

The aim of this study was to systematically assess the reported prevalence of pituitary insufficiency after cranial or total body radiotherapy for intracerebral tumors, nasopharyngeal tumors or haematological malignancies at the adult age.

Design

Search strategy and eligibility criteria

We searched the following databases for studies on cranial radiotherapy and pituitary failure: PubMed, Cochrane Library, Web of Science, EMBASE, CINAHL database, Academic Search Premier, and Science Direct. The search was performed on August 14, 2010.

In collaboration with a trained clinical librarian, we composed a search strategy for the above mentioned databases, focusing on radiotherapy, pituitary function, cerebral tumors and nasopharyngeal tumors. We used all relevant keyword variations, including free text words. The complete strategy is provided in the Appendix 1. Furthermore, the references of relevant articles were checked for additional articles.

Only original articles in English were included. Studies were eligible for inclusion in this review if they fulfilled the following criteria: 1) cranial radiotherapy for nonpituitary tumors and/or total body irradiation for hematological malignancies 2) >18 yr old at the time of radiotherapy 3) report on endocrine evaluation.

In case of mixed cohorts (*i.e.* including both paediatric and adult patients), patients younger than 18 yr were filtered from the results. In case of duplication of reports involving the same patient cohort the results on the different axes were combined in the analyses and tables, and only the paper with the longest duration of follow-up was included.

Data review and analysis

Initial selection of studies by title and abstract was performed by two reviewers (N.M.A-D en N.E.K.). These studies were retrieved for full assessment. All studies were evaluated by the two reviewers independently. Disagreements were resolved by consensus. Data extraction was based on data from each study provided at the population level. The definition of hypopituitarism had to be stated in the paper, including the endocrine tests used for the evaluation, definitions and cut-off values used to define pituitary insufficiency for each axis, hormone assays and reference values provided by the authors.

Statistical analysis

The main outcome of the present meta-analysis was the pooled proportion of patients with pituitary insufficiency after cranial radiotherapy. For all studies, the proportion of patients with hypopituitarism was calculated as the number of patients with the pituitary insufficiency divided by the total number of tested patients.

Meta-analysis was performed using an exact likelihood approach. The method used was a logistic regression with a random effect at the study level (15). Given the expected clinical heterogeneity, a random effects model was performed by default, and no fixed effects analyses were performed. For meta-analysis of proportions, the exact likelihood approach based on a binomial distribution has advantages compared to a standard (DerSimonian and Laird) random effects model that is based on a normal distribution. First, estimates from a binomial model are less biased than estimates from models based on a normal approximation (15;16). This is especially the case for proportions that are close to 0 or 1. Secondly, no assumptions are needed for the exact approximation when dealing with zero-cells, whereas the standard approach needs to add an arbitrary value (often 0.5) when dealing with zero-cells. Adding values to zero-cells is known to contribute to the biased estimate of the model (17;18).

Meta-regression analyses were also performed with an exact likelihood approach. A random effects meta-regression was performed to address the question whether the tumor site (nasopharyngeal *vs* intracerebral) influences the prevalence of pituitary insufficiency. All analyses were performed with STATA 10.0 (Stata Corp, College Station, TX, USA).

Risk of bias assessment

An additional evaluation of the risk of bias was performed to identify components that could potentially bias an association between cranial radiotherapy and hypopituitarism. The following study characteristics were evaluated: 1) adequacy of exposure determination, 2) adequacy of inclusion and follow-up, and 3) adequacy of outcome determination. For exposure determination, one point was given if it was stated clearly that the pituitary was involved in the radiation field, and one point was given if the estimated dose at the pituitary was reported. For the evaluation of inclusion of patients, one point was given for each study that included (consecutive) non-selected patients. For outcome determination, one point was given when the hormonal evaluation also included dynamic tests, and one point was given if all patients in the study were tested. Consequently, each study could attain a maximum score of 5 points. Studies that scored 0–2 points were considered to have a high risk of bias, studies with 2–3 points as intermediate risk, and studies with 4–5 points as studies with a low risk of bias.

Results

Systematic literature search

The initial search resulted in a total of 849 articles (301 in PubMed, four in Cochrane Library, 131 in Web of Science, 353 in EMBASE, 16 in CINAHL database, 27 in Academic Search Premier, and 17 in Science Direct). Of these articles, 616 were unique without duplications (Figure 1). We excluded 378 papers based on title and abstract or language, 157 studies that evaluated patients younger than 16 yr, and seven papers which were not available for evaluation. Consequently, a total of 74 potentially relevant papers were retrieved for full assessment. Of these 74 publications, 51 papers were excluded from further analysis because the studies did not fulfill one or more of the eligibility criteria.

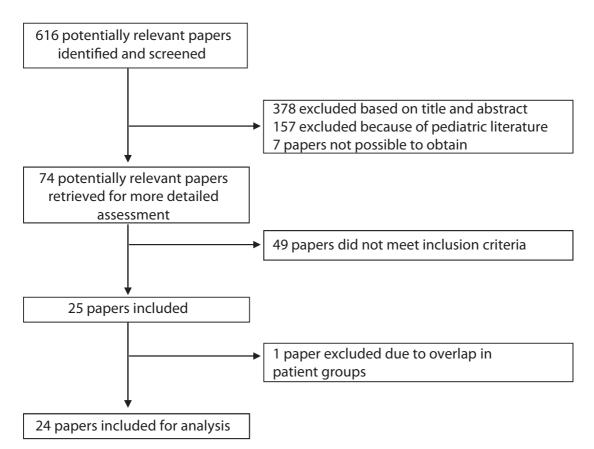


Figure 1. Flowchart of study assessment and exclusion stages

Therefore, ultimately, our search strategy resulted in 23 manuscripts meeting the inclusion criteria. However, some reports described data from the same patient cohort (19–26). In these cases, the results were combined as one study in the analysis, or the study with the longest follow-up was included. Consequently, a total of 18 studies were included in the present review, comprising 813 patients. The number of included patients per study ranged from only six (27) to 312 (20). Five studies included patients younger than 18 yr (26–30). However, in only three of these five studies (27;28;30) it was possible to obtain the numbers of patients treated after the age of 18 yr. The remaining two were included because of the low number of patients younger than 18 yr, or sub-analysis on the patient group younger than 18 yr was performed with no different outcome then the older group (Table 1).

Study characteristics

Details of the 18 included studies are summarized in Table 1 and 2. The studies were published between 1975 and 2009. Seventy-five percent of the patients (608 of 813) were treated for nasopharyngeal cancer (Table 1). The remaining 25% were treated for intracerebral tumors (Table 2). The majority of studies were cross-sectional studies with time after irradiation ranging from 4 months to 30 yr. However, the time after radiation varied considerably between individuals, even within one single study. Two studies did not report the time interval between irradiation and endocrine evaluation (28;31). None of the included papers evaluated patients treated with prophylactic body irradiation in the course of stem cell transplantation for haematological malignancies. In three reports, patient selection criteria were not stated (20;28;31), whereas four studies selected symptomatic patients by recruiting from a radiotherapy complication outpatient clinic or by inclusion of only patients suspected for any degree of hypopituitarism (21;30;32;33). For example, two studies used questionnaires on fatigue or diminished libido in combination with basal hormone samples to select patients for further endocrinological evaluation (22;28).

tumors	
l and neck	
Head	
ble	

	Number of							Pituitaı	Pituitary insufficiency per axis (%)	ency per	(%) sixe	
	patients,	Tumor	cilocuti Iciacu)	Time cinco Ptv		Bias						
Author	age anu gender	istics	tion (Gy)	(months)	Endocrine evaluation	ment	Any	Н	HPA	TSH	LH/FSH	PRL
Snyers, 2009(37)	Total n=32 ^a	Sinonasal cancer	44–66	107 (11–253)	Basal morning serum samples, if abnormal :	4	62% (13/21)	24% (5/21)	1 <i>9</i> % (4/21)	14% (3/21)	19% (4/21)	10% (2/21)
	n=21 endo- crine evalu- ation 56 (28-74) yr		Mean dose pituitary = 51–56 Mean dose hy- pothalamus = 44–52 Chemo n= 23		GHRH-arg							
Bhandare, 2008(19) ^b	Bhandare, Total n=312 2008(19) ^b 200M/112F n= 112 endocrine evaluation No age men- tioned		Nasal cavity Broad range: n=56 < 40 - >70 Nasopha- rynx n=119 Chemo n= 40 Frontal sinus n=2 Etmoid sinus n=5 Sphenoid sinus n=34 Maxillary Other n=37 Other n=37	63 (6–365)	Basal serum samples: TSH, fT ₄ , PRL, GH, Cor- tisol, LH, FSH, T If abnormal: ACTH-test, metyrapone, CRH test, ITT + vasopressin, GST, GHRH-test, Arginine- test, TRH test	7	60% (67/112)	36% (16/44)	32% (14/44)	70% (31/44)	27% (12/44)	15% (10/68)

	Number of							Dituita	Dituitary incutt ciency net avic (06)		vie (0/)	
		I		i		i						
	patients,	Tumor		Time		Bias						
	age and	character-	Cranial irradia-	since Rtx		assess-						
Author	gender	istics	tion (Gy)	(months)	Endocrine evaluation	ment	Any	НЭ	HPA	TSH	LH/FSH	PRL
Lam,	n=20	Nasopha-	Total dose: 60	60	Basal serum samples:	5	75%	55%	25%	15%	35%	30%
1991(22) ^c	1991(22) ^c 22M/9F	ryngeal			LH, FSH, PRL, TSH, T_4'		(15/20)	(11/20)	(5/20)	(3/20)	(7/20)	(6/20)
			Estimated dose		T, E ₂							
	M: 43.7±8.4		pituitary: 62									
	yr				ITT, LHRH 100 μg, TRH							
	F: 36.8±9.0 yr	F			200 µg							
Woo,	n=11	Nasopha-	Estimated dose	72–240	Basal serum samples:	4	82%	%06	18%	45%	55%	27%
1988(31)	8M/3F	ryngeal	pituitary: 62–67		LH, FSH, THS, T ₄ , cor- tisol. PRL_T_F		(11/6)	(9/10)	(2/11)	(5/11)	(6/11)	(3 /1 1)
	Ane.											
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	16 (40-cc)04				и т, спъл тоо ду, тъп 200 дg							
Samaan,	Total n=166	Nasopha-	Estimated dose	12–312	ITT, TRH 500 μg, LHRH	5	75%	75%	18%	20%	35%/20%	36%
1987(25) ^d	98M/68F	ryngeal	anterior pituitary:		100 µg;		(124/166)	(124/166) (124/166) (30/166) (33/166)	(30/166)	(33/166)	(58/166	(60/166)
		n=114	57 (4–75)		Total T ₄ , T ₃ resin uptake						FSH)	
	Age:	Paranasal									(33/166	
	47(6–80) yr	sinus tu-	Estimated dose								(H)	
		mors n=29	hypothalamus:									
		Optic	50 (11–75)									
		nerve/ eye										
		tumors										
		n=23										

Table 1.	Continued											
	Number of							Pituitaı	Pituitary insufficiency per axis (%)	iency per	axis (%)	
	patients,	Tumor		Time		Bias						
	age and	character-	Cranial irradia-	since Rtx		assess-						
Author	gender	istics	tion (Gy)	(months)	Endocrine evaluation	ment	Any	ВH	HPA	TSH	LH/FSH	PRL
Lam,	n=32	Nasopha-	46–60	60–204	n=32	m	25%	19%	6%	13%	16%	19%
1987(21) ^e	1987(21) ^e 21M/11F	ryngeal			Questionnaires on sexual impairement,		(8/32)	(6/32)	(2/32)	(4/32)	(5/32)	(6/32F)
	n=14 dyna-				galctorroea							
	mic testing				Basal serum samples: PRL, T _a , FTI, TSH, T							
	Age: 27–50 yr	,			lf abnormal: E ₂ , T, ITT. LHRH-test. TRH test							
Lam,		Nasopha-	46–61	>60	Basal serum samples:	5	100%	50%	50%	50%	25%	88%
1986(20)		ryngeal	Estimated dose		LH, FSH, ISH, PKL, TI4, Е ₂ , Т		(8/8)	(4/8)	(4/8)	(4/8)	(2/8)	(7/8)
	Age: 27–52 yr	L	to pituitary: 55–67		ITT (n=2) I HRH test							
			5		TRH test, GHRH-test (n=6), CRH test (n=4)							
Huang,	n=11	Nasopha-	Nasopharyngeal	12–156	LHRH 25 µg	Ś	82%	NR	NR	NR	82%/64%	100%
1979(33)	All females	ryngeal	area: 70				(6/11)				(9/11 LH)	(8/8)
											(7/11	
	Age: 19–40 yr	~	Estimated dose HP area: 60–65								FSH)	

CHAPTER 6

152

ents, andTumorTimeBiasand bercharactercranial irradiasince Rtxssess-derlotscharactercranial irradiasince Rtxssess-derlotston (Gy)(months)Endocrine evaluationment Any GH dersitcston (Gy)(months)Endocrine evaluationment Any GH HPa datestospha-55-6512-96Basal serum samples: T_{a} 3 67% 100%50%dateryngealTSH, FTI, T_{a} cortisol, LH,TSH, FTI, T_{a} cortisol, LH, Any GH HPa 35-66 yrS5-65TSH, GHTRH, FST, IT (n=2)3 67% 100% 50% 35-66 yrNasopha-Estimated dose $60-240$ IT, Chlorpromazin: 25 4 100% 60% 50% 0Nasopha-Estimated dose $60-240$ IT, Chlorpromazin: 25 4 100% 60% 50% 35ryngealHP area: $50-83$ mg, TRH test $500 \mug, 4, 7$ sresin $6/10$ $6/10$ $5/10$ $5/10$ 26-55S0-655 to thyroidNasopha-Estimated dose $100\mu, T_a/1$ sresin $6/10$ $5/10$ $5/10$ 26-55S0-655 to thyroidNasopha-Estimated dose $100\mu, T_a/1$ sresin 100% $5/10$ $5/10$ 26-55S0-655 to thyroidNasopha-Estimated dose $10\mu, Nasopha-10\mu, Nasopha-10\mu, Nasopha-10\mu, Nasopha-26-55$									Pituita	Pituitary insumciency per axis (%)	iency per	axis (%)	
andcharacterchanaltirradiasince Rtxassess-deristicstion (Gy)(months)Brdocrine evaluationment Any GH HPA derstoopha-55-6512-96Basal serum samples: $T_{a'}$ 3 67% 100%50%haleryngealTSH, FTI, $T_{a'}$ cortisol, LH,7 $(4/6)$ $(2/2)$ $(1/2)$ 35-66 yrS5-65TSH, FTI, $T_{a'}$ cortisol, LH, $(4/6)$ $(2/2)$ $(1/2)$ 35-66 yrNasopha-Estimated dose $60-240$ TT, Chlorpromazin: 25 4 100% 60% 0Nasopha-Estimated dose $60-240$ TT, Chlorpromazin: 25 4 100% 60% 50% 0Nasopha-Estimated dose $60-240$ TT, Chlorpromazin: 25 4 100% 60% 50% 0Nasopha-Estimated dose $60-240$ TT, Chlorpromazin: 25 4 100% 60% 50% 0Nasopha-Estimated dose $60-240$ TT, Chlorpromazin: 25 4 100% $(6/10)$ $(5/10)$ 26-5550-65 to thyroidNasob 100% 100% 100% 100% 100% 100% 100% 26-5550-65 to thyroidNasob 100% 100% 100% 100% 100% 100% 100% 26-5550-65 to thyroidNasob 100% 100% 100% 100% 100% 100% 26-5510006NasobNasobNasob		patients,	Tumor		Time		Bias						
deristicstion (Gy)(months)Endocrine evaluationment A_{IY} GH HPA Nasopha-55-6512-96Basal serum samples: $T_{4'}$ 3 67% 100%50%IaleryngealTSH, FTI, $T_{3'}$ cortisol, LH,3 67% 100%50%S5-66 yrTSH, GHTSH, GHRH est, ITT (n=2)(4/6)(2/2)(1/2)35-66 yrNasopha-Estimated dose $60-240$ IT, Chlorpromazin: 254100% 60% 50%0Nasopha-Estimated dose $60-240$ IT, Chlorpromazin: 254100% 60% 50%3FryngealHP area: 50-83mg, TRH test 500 µg, $T_{4'}$ sresin100% 60% 50%26-5550-65 to thyroiduptake, 24 hour 131-1Ktr:131-1KtriKtriA-58 wrIobes (n=12)thyroid uptakethyroid uptake $4/6$ $6/10$ $5/10$		age and	character-		since Rtx		assess-						
II, n=6Nasopha-55-6512-96Basal serum samples: T_4 367%100%50%All MaleryngealTSH, FTI, T_3 , cortisol, LH, $(4/6)$ $(2/2)$ $(1/2)$ Age: 35-66 yrAll MaleryngealFSH, GH $(4/6)$ $(2/2)$ $(1/2)$ Age: 35-66 yrNasopha-Estimated dose $60-240$ IT, Chlorpromazin: 25 4 100% 60% 50% $n=10$ Nasopha-Estimated dose $60-240$ IT, Chlorpromazin: 25 4 100% 60% 50% $7M/3F$ ryngealHP area: $50-83$ mg, TRH test 500 µg, 4 100% 60% 50% Age: $26-55$ 50-65 to thyroiduptake, 24 hour 131-1 $(6/10)$ $(5/10)$ $(5/10)$ $(5/10)$ Age Rtx:Iobes (n=12)thyroid uptakethyroid uptake	Author	gender	istics	tion (Gy)	(months)	Endocrine evaluation	ment	Any	НЭ	HPA	TSH	LH/FSH	PRL
Age: 35-66 yrTRH test, ITT (n=2)n=10Nasopha-Estimated dose $60-240$ ITT, Chlorpromazin: 254 100% 60% 50% 7M/3FryngealHP area: $50-83$ mg, TRH test 500 µg, LHRH 100 µg, T_4 , T_3 resin $(6/10)$ $(5/10)$ $(5/10)$ Age: $26-55$ $50-65$ to thyroid ubtake, 24 hour 131-1uptake, 24 hour 131-1thyroid uptakeAge Rtx:lobes (n=12)thyroid uptake	Rosenthal, 1976(34)	l, n=6 All Male	Nasopha- ryngeal	55-65	12–96	Basal serum samples: T ₄ , TSH, FTI, T ₃ , cortisol, LH, FSH, GH		67% (4/6)	100% (2/2)	50% (1/2)	67% (4/6)	NR	NR
n=10 Nasopha- Estimated dose 60–240 ITT, Chlorpromazin: 25 4 100% 60% 50% 7M/3F ryngeal HP area: 50–83 mg, TRH test 500 µg, (6/10) (5/10) Age: 26–55 50–65 to thyroid uptake, 24 hour 131-1 (6/10) (5/10) Age Rtx: lobes (n=12) thyroid uptake thyroid uptake 131-1		Age: 35–66	yr			TRH test, ITT (n=2)							
7M/3F ryngeal HP area: 50–83 mg, TRH test 500 μg, LHRH 100 μg, T ₄ , T ₃ resin (6/10) (5/10) Age: 26–55 50–65 to thyroid uptake, 24 hour 131-1 41 (8–58 vr) ⁹ 41 (8–58 vr) ⁹	Samaan,	n=10	Nasopha-	Estimated dose	60–240	ITT, Chlorpromazin: 25	4	100%	60%	50%	40%	30%	50%
50–65 to thyroid lobes (n=12)	1975(36)	7M/3F	ryngeal	HP area: 50–83		mg, TRH test 500 µg, LHRH 100 µg, T ₄ , T ₃ resin			(6/10)	(5/10)	(4/10)	(3/10)	(5/10)
lobes (n=12)		Age: 26–55		50–65 to thyroid		uptake, 24 hour 131-l							
41 (8–58 vr) ^g		Age Rtx:		lobes (n=12)		thyroid uptake							
		41 (8–58 yr) ^g	D										

PITUITARY DYSFUNCTION IN ADULT PATIENTS AFTER CRANIAL RADIOTHERAPY

	Number of							Pituita	Pituitary insufficiency per axis (%)	ency per a	(%) sixe	
	patients,		Cranial ir-			Bias						
	age and	Tumor	raidiation	Time since Endocrine	Endocrine	asses-						
Author	gender	characteristics	(Gy)	RTx (years)	Fx (years) Evaluation	ment	Any	НЭ	HPA	TSH	LH/FSH	PRL
Schneider, n= 44	n= 44	Glioma n= 40	NR	NR	Basal serum samples:	2	37%	27%	18%	16%	29.5%	7%
2006(30)	2006(30) 28 M/40 F	Neuroblastoma n= 1	_		cortisol, 24-h urinary		(17/44)	(12/44)	(8/44)	(7/44)	(13/44)	(3/44)
		Menigeoma n= 18	Chemo n=27		cortisol, T ₃ , T ₄ , TSH,							
	Age: 45	Schwanoma n= 5			PRL, FSH, LH, test, E ₂							
	(20–79) yr	Dysgerminoma n= 2	.									
		Neuroblastoma n= 1	_		GHRH-arg							
		Chondrosarcoma										
		n= 1										
		Hemangiopericy-										
		toma n= 1										
		Oesteosarcoma n= 1	_									
Agha,	n= 56	Glioma n= 43	Estima-	9	Basal serum samples:	5	41%	32%	21%	%6	27%	32%
2005(35)	28 M/28 F	Meningeoma n= 5	ted dose		FSH, LH, TSH, T ₄ , PRL,		(23/56)	(18/56)	(12/56)	(2/20)	(15/56)	(18/56)
		Pinealoma n= 3	pituitary:		IGF-I, T, E ₂							
	Age:	Medulloblastoma	54(4–97)									
	39.3±11.9	n= 2	Chemo n=7		ITT (n=25), Arginine							
	yr	Other n= 3			stim test, ACTH stim							
					test							

 Table 2.
 Primary intracerebral tumors

Cranial icCranial icTumorraidiationTime sinceLumorGy) \mathbf{FTX} (years)Low grade glioma 54 (45 - 59) 13.1 Low grade glioma 54 (45 - 59) 13.1 $n=27$ $6-25.6$) 13.1 $n=27$ $6-25.6$) 13.1 $n=27$ $6-25.6$) 13.1 $n=1$ 56 7.6 ± 0.7 $n=1$ 56 7.6 ± 0.7 $n=4$ 12.1 $(2-13)$ Pinealoma $n=2$ Estima- $n=4$ $25-30$ Low grade astrocy- $45-61$ Low grade oligo-Mean calcu-dendroma $n=6$ Mean calcu-low grade oligo-Mean calcu-dendroma $n=6$ Pituitary $36(0-50)$ Pituitary		Number of							Dituitar	v incuffici	ancy nor	ovic (0/)	
age and enderTunorraidiationTime sinceasses- towasses- tow Any GH HM $n = 33$ Low grade glioma $54(45-59)$ 13.1 Basal serum levels 3 64% NR 4% $n = 25$ Low grade glioma $54(45-59)$ 13.1 Basal serum levels 3 64% NR 4% $18M/15F$ $n = 27$ Low grade glioma $54(45-59)$ 13.1 Basal serum levels 3 64% NR 4% $18M/15F$ $n = 27$ High grade glioma $54(45-59)$ 13.1 Basal serum levels 3 64% NR 4% $n = 1$ $n = 7$ $(6-25.6)$ If abnormal: $(16/25)$ $(17/25)$ $(17/25)$ $(17/25)$ $n = 1$ $n = 1$ $(1-68)Y$ $(1-68)Y$ $(16/25)$ $(16/25)$ $(17/25)$ $(17/25)$ $Age Rix:n = 1(1-68)Y(17,6)(17,6)(17/25)(17/25)Age Rix:n = 1(1-68)Y(17,6)(17/25)(17/25)(17/25)Age Rix:n = 1(1-68)Y(17,6)(17/2)(17/25)(17/25)Age Rintn = 1(1-68)Y(17,6)(17/2)(17/2)(17/2)Age Rintn = 1(1-68)Y(17,6)(17/2)(17/2)(17/2)(14-68)Y(1-68)Y(17,6)(17/2)(17/2)(17/2)(17/2)(14-68)Y(1-68)Y(116)Y(160)Y(1$		patients,		Cranial ir-			Bias						
gendercharacteristics(Gy)RTx (years)Evaluationment Any GH HM $n=33$ low grade glioma $54(45-59)$ 13.1Basal senum levels3 64% NR 4% $18M/15F$ $n=27$ High grade glioma $54(45-59)$ 13.1Basal senum levels3 64% NR 4% $18M/15F$ $n=27$ High grade glioma $54(5-59)$ 13.1Basal senum levels3 64% NR 4% $n=1$ $n=2$ High grade glioma $56-25.6$ TRH test, CRH test $(16/25)$ (1725) (1725) $n=1$ $n=1$ $n=1$ $(-2)2.6$ TRH test, CRH test $(16/25)$ (1725) (1725) $n=1$ $n=1$ $n=1$ $(-2)2.6$ TRH test, CRH test $(16/25)$ (1725) (1725) $Age Rix:n=1(-2)2.6TRH test, CRH test(-2)2.6(-2)2.6(-2)2.6(-2)2.6Age Rix:n=1(-2)2.6TRH test, CRH test(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-2)2.6Age Rix:Age Rix:(-2)2.6TRH test, CRH test(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-2)2.6Age Rix:Age Rix:(-2)2.6(-2)2.6TRH test, CRH test(-2)2.6(-2)2.6(-2)2.6Age Rix:(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-2)2.6(-1-6.8)(-2)2.6(-2)2.6(-2)2.6<$		age and	Tumor	raidiation	Time since	Endocrine	asses-						
n=33Low grade glioma $54(45-59)$ 13.1 Basal serum levels 3 $64%$ NR $4%$ $18M/15F$ $n=27$ $(16/25)$ $(16/25)$ $(16/25)$ $(11/25)$ $(11/25)$ $(11/25)$ $8M/15F$ $n=27$ $(6-25.6)$ (17) $(16/25)$ $(16/25)$ $(11/25)$ $(11/25)$ $8notocrinen=5(6-25.6)(17)(16/25)(16/25)(11/25)(11/25)evaluationn=1(12)(12)(16/25)(11/25)(11/25)(11/25)Age Rtx:n=1(12)(12)(12)(12)(12)(12)Age Rtx:n=1(12)(12)(12)(12)(12)(12)Age Rtx:Age(14)(12)(12)(12)(12)(12)Age(14)(12)(12)(12)(12)(12)(12)Age(12)(12)(12)(12)(12)(12)(12)Age(12)(12)(12)(12)(12)(12)(12)Age(12)(12)(12)(12)(12)(12)(12)(12)Age(12)(12)(12)(12)(12)(12)(12)(12)Age(12)(12)(12)(12)(12)(12)(12)(12)(14)(12)(12)(12)(12)$	Author	gender	characteristics	(Gy)	RTx (years)	Evaluation	ment	Any	НD	HPA	TSH	LH/FSH	PRL
5F $n=27$ ($n=27$ ($16/25$) ($16/25$) ($1/25$) 119h grade glioma $n=1$ TRH test, CRH test ($16/25$) ($1/25$) 110n Anaplastic glioma $n=1$ $(1/25)$ ($1/25$) ($1/25$) 110n Anaplastic glioma $n=1$ $(1/25)$ $(1/25)$ $(1/25)$ 122 Medulloblastoma 56 7.6 ± 0.7 $(17, GHRH-GHRP6)$ 3 67% $(17, 67\%)$ NR 3) Yr $n=1$ $(2-13)$ $(17, GHRH-GHRP6)$ 3 67% $(1/6)$ $(4/6)$ $(4/6)$ $(1/6)$ 13 Yr $n=4$ $(2-13)$ $(2-13)$ $(17, GHRH-GHRP6)$ 3 67% $(1/6)$	Johan-	n= 33	Low grade glioma	54 (45–59)	13.1	Basal serum levels	m	64%	NR	4%	56%	16%	0
High grade gliomaTRH test, CRH testrinen=5n=1 $n=1$ tx: $(2-13)$ triandma n=2 $(1-11.5)$ toma n=7 $(1-11.5)$ toma n=6 $(1-11.5)$ troma n=6 $(1-11.5)$ trand n=7 $(1-11.5)$ troma n=6 $(1-11.5)$ trand n=7 $($	nesen,	18M/15F	n= 27		(6–25.6)	lf abnormal:		(16/25)		(1/25)	(14/25)	(4/25)	
tine n=5 tion Anaplastic glioma n=1 tx: tx: h=2 bx: bx: h=1 tx: h=2 h=1 tx: h=2 h=1 tx: h=2 h=1 tx: h=2 h=1 h=1 h=1 h=1 h=1 h=1 h=1 h=1	2003(28)	n= 25	High grade glioma			TRH test, CRH test							
tion Anaplastic glioma $ n=1 $ http://dotsing.org/linear controls/linear cont		endocrine	n= 5										
$n=1$ $n=1$ Low 56 7.6 ± 0.7 $TT, GHRH-GHRP6$ 3 67% $TT: 67\%$ NR -122 Medulloblastoma 56 7.6 ± 0.7 $TT, GHRH-GHRP6$ 3 67% $TT: 67\%$ NR -18 yr -13 $(2-13)$ $(2-13)$ $(4/6)$ $(4/6)$ $(4/6)$ $(4/6)$ -18 yr 22 $Estima (2-13)$ $(4/6)$ $(4/6)$ $(4/6)$ $(4/6)$ -18 yr 25 27% $(4/6)$ $(4/6)$ $(4/6)$ $(4/6)$ -18 yr $25-30$ $(2-6)$ 33% $(2-6)$ 33% -100 yr 100 yr 100 yr 100 yr $(10/13)$ $(4/13)$ $(4/13)$ -100 yr 100 yr 100 yr $(10/13)$ $(4/13)$ $(4/13)$ $(8/13)$ -100 yr 100 yr 100 yr 100 yr 100 yr 100 yr 100 yr		evaluation	Anaplastic glioma										
B)Yr 56 7.6 ± 0.7 ITT, GHRH-GHRP6 3 67% ITT: 67% NR = 22 Medulloblastoma 56 7.6 ± 0.7 ITT, GHRH-GHRP6 3 67% ITT: 67% NR -18yr n=4 (2-13) (2-13) (4/6) (4/6) (4/6) 33% -18yr n=4 (2-13) (2-13) (4/6) (4/6) (4/6) 33% Pinealoma n= 2 Estima- (2-13) (2-13) (4/16) (4/6) (4/6) (4/6) Pinealoma n= 2 Estima- (2-13) (10/12) (4/13) (4/13) (31% Pinealoma n= 7 1 (1-11.5) 33% (10/13) (4/13) (4/13) (4/13) I cow grade oligo- Mean calcu- (10-11.5) TRH test 200 µg 5 77% 31% 62% I cow grade oligo- Mean calcu- (10-11.5) TRH test 100 µg 10/13) (4/13) (4/13) (4/13) (4/13) (4/13) (4/13) (4/13) I cow grade oligo- I ated dose Gradh dose Grade oligo- I ated dose <td></td> <td></td> <td>n= 1</td> <td></td>			n= 1										
B) Yr 1= 22Medulloblastoma 18 yr 1= 4567.6 ± 0.7 ITT, GHRH-GHRP636.7%ITT. 67%NR·18 yrn=4(2-13)(2-13)(4/6)(4/6)(4/6)(4/6)(4/6)·18 yrn=4(2-13)(2-13)(4/6)(4/6)(4/6)(4/6)·18 yrted doseted doseted dose(2-13)(4/6)(4/6)(4/6)·18 yrted doseted doseted dose(2-13)(4/6)(4/6)(4/6)·10 w grade astrocy45-6138asal serum levels577%31%(62%·10 w grade astrocyMean calcu-(1-11.5)(1-11.5)(10/13)(4/13)(8/13)(8/13)·10 w grade oligo-Mean calcu-Ith test 200 µgfarH test 100 µg(10/13)(4/13)(8/13)·11 wrpituitaryintert 100 µgfarH test 100 µgintert 100 µgintert 100 µg·11 wrpituitarybituitaryfarH test 100 µgintert 100 µg·11 wrpituitarybituitarybituitaryintert 100 µg·11 wrpituitarybituitary		Age Rtx:											
B) yr = 22 Medulloblastoma 56 7.6 \pm 0.7 ITT, GHRH-GHRP6 3 67% ITT: 67% NR -18 yr = 22 medulloblastoma 55 7.6 \pm 0.7 iTT, GHRH-GHRP6 3 (4/6) (4/6) (4/6) (4/6) (4/6) (2		38											
1= 22 Medulloblastoma 56 7.6 ± 0.7 ITT, GHRH-GHRP6 3 67% ITT: 67% NR 18 yr n=4 (2-13) (2-13) (4/6) (4/1)<		(14–68) yr											
-18 yr n=4 (4/6) (4/6) (4/6) Pinealoma n= 2 Estima- ted dose ted dose pituitary: 25-30 Low grade astrocy- toma n=7 Low grade oligo- toma n=6 Mean calcu- Low grade oligo- Mean calcu- Low grade oligo- Mean calcu- Low grade oligo- Mean calcu- Low grade oligo- Mean calcu- Mean calcu- Low grade oligo- Mean calcu- Mean calcu- Mean calcu- Colop grade oligo- Mean calcu- Mean	Popovic,	Total n= 22		56	7.6 ± 0.7	ITT, GHRH-GHRP6	m	67%	ITT: 67%	NR	NR	NR	NR
Pinealoma n= 2 Estima- ted dose ted dose pituitary: GHRP6: 25-30 33% 25-30 33% Low grade astrocy- 45-61 3 low grade oligo- Mean calcu- TRH test 200 µg low grade oligo- Mean calcu- TRH test 100 µg low grade oligo- Mean calcu- TRH test 100 µg low grade oligo- Mean calcu- GnRH test 100 µg low grade oligo- Mean calcu- GnRH test 100 µg low grade oligo- Mean calcu- GnRH test 100 µg low grade oligo- Mean calcu- Mean calcu- low grade oligo- Mean calcu- Mean calcu- low g	2002(29)	n= 6 >18 yr	n=4		(2–13)			(4/6)	(4/6)				
ted dose ted dose GHRH- pituitary: 25-30 GHRP6: 25-30 25-30 33% 25-30 25-30 33% 25-30 25-30 33% 25-30 27% 31% 62% 10w grade astrocy- 45-61 3 833 10w grade astrocy- 45-61 3 83% 10w grade oligo- Mean calcu- (1-11.5) (10/13) (4/13) (8/13) 10w grade oligo- Mean calcu- TRH test 200 µg (10/13) (4/13) (8/13) 10w grade oligo- Mean calcu- TRH test 100 µg (10/13) (4/13) (8/13) 10w grade oligo- Mean calcu- GnRH test 100 µg (10/13) (4/13) (8/13) 10w grade oligo- Mean calcu- Gneb comp Gneb comp (10/13) (4/13) (8/13) 10w grade oligo- Mean calcu- Gneb comp Gneb comp (10/13) (4/13) (8/13) 10w grade oligo- Mean calcu- Gneb comp Gneb comp (10/13) (10/13) (10/13) (13/13)			Pinealoma n= 2	Estima-									
pituitary: GHRP6: 25–30 33% 25–30 33% 25–30 33% 100 grade astrocy- 45–61 3 100 grade astrocy- 45–61 3 833 100 grade astrocy- 45–61 3 833 100 grade astrocy- 45–61 3 833 100 grade oligo- Mean calcu- (1–11.5) (10/13) (4/13) (8/13) 100 grade oligo- Mean calcu- TRH test 200 µg (10/13) (4/13) (8/13) 101 yr Jated dose GnRH test 100 µg FCF test 100 µg 10/13) (4/13) (8/13) 101 yr Jated dose GnRH test 100 µg GnRH test 100 µg 10/13) (4/13) (8/13) 101 yr Jated dose GnRH test 100 µg GnRH test 100 µg 10/13) 10/13) 10/13) 10/13) 101 yr Jated dose GnBH test 100 µg Jated dose 10/13) 10/13) 10/13) 10/13) 101 yr Jated dose GnBH test 100 µg Jated dose 10/13) 10/13) 10/13) 10/13) </td <td></td> <td></td> <td></td> <td>ted dose</td> <td></td> <td></td> <td></td> <td></td> <td>GHRH-</td> <td></td> <td></td> <td></td> <td></td>				ted dose					GHRH-				
25-30 33% Low grade astrocy 45-61 3 8asal serum levels 5 77% 31% 62% I com a n= 7 (1-11.5) (10/13) (4/13) (4/13) (8/13) I com a n= 7 (1-11.5) TRH test 200 µg (10/13) (4/13) (8/13) Vr pituitary GnRH test 100 µg net test 100 µg 100 100 13 Vr pituitary GHRH 50 µg GHRH 50 µg 100 100 100				pituitary:					GHRP6:				
Low grade astrocy- 45-61 3 Basal serum levels 5 77% 31% 62% i toma n= 7 (1-11.5) (10/13) (4/13) (8/13) Low grade oligo- Mean calcu- TRH test 200 μg (10/13) (4/13) (8/13) Vr pituitary GnRH test 100 μg Frest 100 μg 5 77% 36(0-50) Yr 36(0-50) GHRH 50 μg GHRH 50 μg 5 77% 100 μg				25–30					33%				
Low grade astrocy- 45-61 3 Basal serum levels 5 7.1% 31% 62% : toma n= 7 (1-11.5) (1-11.5) (10/13) (4/13) (8/13) Low grade oligo- Mean calcu- TRH test 200 µg (10/13) (4/13) (8/13) Low grade oligo- Mean calcu- TRH test 200 µg (10/13) (4/13) (8/13) v dendroma n= 6 lated dose GnRH test 100 µg (10/13) (4/13) (8/13) .vr pituitary hCRF test 100 µg (10/13) (4/13) (8/13) .vr 36(0-50) GHRH 50 µg (10/13) (4/13) (8/13)	-				ſ	-	I		(2/6)				
9M/4Ftoma n= 7(1-11.5)(4/13)Low grade oligo-Mean calcu-TRH test 200 μgAge:dendroma n= 6lated doseGnRH test 100 μg24-66 yrpituitaryhCRF test 100 μg36(0-50)GHRH 50 μg	Taphoorn	, n= 13		45–61	m	Basal serum levels	S	77%	31%	62%	%0	15%	23%
Low grade oligo- Mean calcu- dendroma n= 6 lated dose 5 yr 36(0–50) 36(0–50)	1995(38)	9M/4F	toma n= 7		(1-11.5)			(10/13)	(4/13)	(8/13)		(2/13)	(3/13)
dendroma n= 6 lated dose 6 yr pituitary 36(0–50)			Low grade oligo-	Mean calcu-		TRH test 200 µg							
pituitary 36(0–50)		Age:	dendroma n= 6	lated dose		GnRH test 100 μg							
		24–66 yr		pituitary		hCRF test 100 µg							
				36(0–50)		GHRH 50 µg							

	f						Pituita	Pituitary insufficiency per axis (%)	iency per	axis (%)	
	Tumor	Cranial ir- raidiation	Time since	Time since Endocrine	Bias asses-						
	characteristics	(Gy)	RTx (years)	RTx (years) Evaluation	ment	Any	НŊ	HPA	TSH	LH/FSH	PRL
	Total n=65 Glioma n= 43 n=30 Medulloblas n= 7 (>18 yr) Meningioma n= 5 Ependymoma n= 4 Age: Others n= 5 36.5(19–65) No biopsy n= 2 yr	Various amounts n = 47 cra- nial Rtx n= 18 crani- ospinal Rtx Chemo n=8	NR	T ₄ and TSH n= 65 PRL n= 47 No dynamic tests n= 35 cranial Rtx	~	ж Z	ж Z	N N N N N N N N N N N N N N N N N N N	25% (7/28)	Only measu- red in 9 people	29% (7/24)
Mecha- n= 15 nick, 5M/10F 1986(32) Age: 22–59 yr	Astrocytomas n= 14 40–50 to Medulloblast n= 1 whole bra 1–22 loca boost Chemo n	40–50 to whole brain 1–22 local boost Chemo n=13	2-9	PRL n=9,T ₄ , T ₃ , T ₃ -resin uptake TRH 200 μg	7	60% (9/15)	Х Х	N	14.3% (1/7)	N N	100% (9/9)

CHAPTER 6

156

	Number of							Pituitar	Pituitary insufficiency per axis (%)	ency per a	(%) sixe	
	patients,		Cranial ir-			Bias						
	age and	Tumor	raidiation	Time since	Time since Endocrine	asses-						
Author	gender	characteristics	(Gy)	RTx (years)	.) Evaluation	ment	Any	НD	HPA	TSH	LH/FSH	PRL
Harrop,	Total n=17	Astrocytoma: n= 2	40–52	9	ITT (n= 6), GST (n= 6),	4	62.5%	50%	12.5%	12.5%	37.5%	NR
1976(26)	n= 8 > 18 yr	n= 8 > 18 yr Meningeoma: n= 3		(1-15)	TRH (n= 8) 200 µg		(5/8)	(4/8)	(1/8)	(1/8)	(3/8)	
		Angioma: n= 1			250 µg Synacthen,							
	Group 1 :	No biopsy: n= 2			Clomiphene stim test							
	tumors re-				(n= 5), LHRH 100 μg							
	mote from											
	the HP-area											
	n= 5											
	3M/2F											
	Group 2:											
	tumors in											
	hypothala-											
	mic area											
	n= 3											
	1M/2F											
ACTH adren Gy, Gray; HP Rtx, radioth a Evaluatior	iocorticotrope h Ά, hypothalamu erapy; Τ, testost η galactorrhea, l	ACTH adrenocorticotrope hormone; E ₂ , estrogens; F, female; FSH, follicle st Gy, Gray; HPA, hypothalamus pituitary axis; ITT, insulin tolerance test; LH, lu Rtx, radiotherapy; T, testosterone; T _a , thyroxine; T _a , triiodothyronine ;TRH, t a Evaluation galactorrhea, libido, menstrual function, texture of skin and	, female; FSH, foll in tolerance test; 'iiodothyronine ; 'n, texture of skii	icle stimulatin LH, luteinizing TRH, thyreotro n and hair, wei	ACTH adrenocorticotrope hormone; E ₂ , estrogens; F, female; FSH, follicle stimulating hormone; GH, growth hormone; GHRH, growth hormone releasing hormone; GST, glucagon stimulation test; Gy, Gray; HPA, hypothalamus pituitary axis; ITT, insulin tolerance test; LH, luteinizing hormone; LHRH, luteinizing hormone releasing hormone; N, number; NR, not reported; M, male; PRL, prolactin; Rtx, radiotherapy; T, testosterone; T ₄ , thyroxine; T ₃ , triiodothyronine ;TRH, thyreotropin releasing hormone, TSH, thyroid stimulating hormone; N, number; NR, not reported; M, male; PRL, prolactin; a Evaluation galactorrhea, libido, menstrual function, texture of skin and hair, weight change, constipation, leg cramping, heat/cold intolerance. When abnormal quationaire: basal hormone	none; GHR hormone r , thyroid st. eg crampir	tH, growth h eleasing ho imulating hc ig, heat/colo	ormone relé rmone; N, nı ormone; d intoleranc	easing horm umber; NR, r :e. When abı	one; GST, glı 10t reported 10tmal quat	ucagon stimu t; M, male; PRL tionaire: basa	lation test; , prolactin; hormone

 $\ensuremath{\mathsf{P}}\xspace{\mathsf{ITUITARY}}$ dysfunction in adult patients after cranial radiotherapy

levels, 2 patients did fill out questionnaire but did not proceed to testing.

Risk of bias assessment

One study was classified as high risk (received 1 point) (28), eight studies as intermediate risk (20;22;29–31;33–35), and nine studies as low risk for selection bias (21;23;26;27;32;36–39).

Radiotherapy

The radiation dose was reported in all, but one study (31) and ranged from 14 to 83 and 40 to 97 Gy in patients treated for nasopharyngeal carcinomas and intracerebral tumors, respectively. A total of nine studies (six involving patients treated for nasopharyngeal tumors and three for intracerebral tumors) also calculated the estimated dose delivered to the pituitary, ranging from 46 to 83 Gy (in patients treated for nasopharyngeal tumors) and 25 to 97 Gy (in patients with intracerebral tumors) (21;23;26;29;30;36–39).

Endocrine assessment

The overall prevalence of any degree of hypopituitarism differed considerably between the studies, ranging from 25% (22) to 100% (21;37) in studies involving patients treated for nasopharyngeal tumors, and from 37% (31) to 77% (39) in patients treated for intracerebral tumors (Figure 2, Table 1 and 2).

GH-IGF-I axis (n=724)

Fourteen studies evaluated the GH-IGF-I axis. However, in only 61% (440 of 724) of the patients the axis was evaluated using basal serum IGF-I and/or GH levels and/or a stimulation test [glucagon stimulation test, insulin tolerance test (ITT), arginine test, and combined GHRH plus arginine tes] (20–23;26;27;30–32;35–40). The prevalence of GH deficiency varied between 24 and 100%. The prevalence of 100% was assessed in one study of six patients that evaluated only two patients using ITT (35).

Hypothalamic-pituitary-adrenal (HPA) axis (n=751)

The HPA axis was evaluated in 14 studies. In only 61% (460/751) of the patients the axis was tested by basal serum cortisol levels and/or a stimulation test (CRH test, ITT, glucagon or ACTH test). Adrenal insufficiency was diagnosed in 0-50% of patients with nasopharyngeal tumors and in 3-62% of the patients with intracerebral tumors.

Hypothalamic-pituitary-thyroidal-axis (n=488)

Sixteen studies evaluated thyroid function using either basal serum hormone levels or a TRH-stimulation test. TSH deficiency was diagnosed in 26% of the patients (126/488). The prevalence rates ranged from 0 to 67% and 13 to 25% in patients with nasopharyngeal cancers and intracerebral tumors, respectively.

Hypothalamic-pituitary-gonadal axis (n=469)

The pituitary-gonadal axis was assessed in 14 studies. Hypogonadotropic hypogonadism was present in 30% (143/469): in 30-82% of cases treated for nasopharyngeal cancer and in 38-61% of cases treated for intracerebral tumors.

Hyperprolactenemia (n=502)

Prolactin levels were measured in 15 studies, documenting hyperprolactinemia in 144 of 502 patients (29%, 2–100% in patients treated for nasopharyngeal cancers, and 7–100% in patients treated for intracerebral tumors). One study did not use basal prolactin levels for definition of a prolactin secretion disturbance but defined abnormal prolactin secretion as a failure to rise more than three-fold in response to a TRH test (39). Therefore, this study was not included in the analysis on this axis.

Deficiency	Studies		Prevalence of pituitary deficiendies (95% CI)
ACTH deficiency	14		0.22 (0.15, 0.30)
TSH deficiency	16	_ —	0.25 (0.16, 0.37)
LH/FSH deficiency	14		0.30 (0.23, 0.37)
Prolactin deficiency	14		0.34 (0.15, 0.60)
GH deficiency	14	_ 	0.45 (0.33, 0.57)
Any deficiency	17		0.66 (0.55, 0.76)
		0.5	1

Figure 2A. Random effects meta-analysis of prevalence of pituitary insufficiency after cranial radiotherapy.

Localization	Deficiency S	Studies		nce of pituitary cies (95% Cl)
Intracerebral tumors	TSH deficiency	7	•	0.16 (0.08, 0.32)
Intracerebral tumors	ACTH deficiency	5	_ •	0.19 (0.07, 0.40)
Intracerebral tumors	Prolactin deficienc	y 5	•	0.24 (0.02, 0.83)
Intracerebral tumors	LH/FSH deficiency	5	-•	0.25 (0.19, 0.33)
Intracerebral tumors	GH deficiency	5	_	0.33 (0.25, 0.42)
Intracerebral tumors	Any deficiency	7	•	0.54 (0.42, 0.66)
Nasopharyngeal cancer	ACTH deficiency	9	_ —	0.25 (0.16, 0.36)
Nasopharyngeal cancer	TSH deficiency	9	•	0.33 (0.19, 0.50)
Nasopharyngeal cancer	LH/FSH deficiency	9	_ 	0.33 (0.23, 0.45)
Nasopharyngeal cancer	Prolactin deficienc	y 9	-	0.38 (0.18, 0.62)
Nasopharyngeal cancer	GH deficiency	9	- _	0.49 (0.32, 0.65)
Nasopharyngeal cancer	Any deficiency	10	-	0.74 (0.57, 0.86)
	. ,			
		(D .5	1

Figure 2B. Random effects meta-analysis of prevalence of pituitary insufficiency according to tumor site

Meta-analysis (Figure 2A and B)

The pooled prevalence of any degree of hypopituitarism was 0.66 [95% confidence interval (CI), 0.55–0.76). GH deficiency was the most prevalent pituitary deficiency, with a prevalence of 0.45 (95% CI, 0.33–0.57), followed by LH/FSH deficiency 0.3 (95% CI, 0.23–0.37) and TSH deficiency 0.25 (95% CI, 0.16–0.37), respectively. The prevalence of hyperprolactinemia was 0.34 (95% CI, 0.15–0.6) and ACTH deficiency had the lowest prevalence 0.22 (95% CI, 0.15–0.3).

In a random effects meta-regression, the effect of tumor localization (nasopharyngeal *vs* cerebral) on the prevalence of deficiencies was assessed. There was no statistically significant association between the probability of any pituitary deficiency (P = 0.14), GH deficiency (P = 0.36), ACTH deficiency (P = 0.75), TSH deficiency (P = 0.11) LH/FSH deficiency (P = 0.21) as well as hyperprolactinemia (P = 0.44) and the indication for radiotherapy (nasopharyngeal cancer *vs*. intracerebral tumors).

A sensitivity analysis with four studies explicitly mentioning the inclusion of consecutive unselected patients was performed (23;29;36). The pooled prevalence of any pituitary deficiency was 0.62 (95% CI 0.45–0.77), which is similar to the pooled prevalence of 0.66 when combining all the studies.

Pituitary insufficiency related to duration of follow-up after radiotherapy

Two studies reported on the occurrence in time of hypopituitarism (23;26). The prevalence of pituitary failure in patients treated for nasopharyngeal tumors was 6% after 1 yr, 35% after 2 yr, 56% after 3 yr and 62% after 4 and 5 yr (23). Samaan *et al.* (26) reported on the classical sequential order of failure of individual pituitary functions in time. GH deficiency occurred after a mean of 2.6 yr, followed by failure of the pituitary-gonadal axis and hyperprolactinemia after approximately 3.8 yr, ACTH insufficiency after 6 yr, and finally TSH insufficiency after a mean of 11 yr.

Discussion

This systematic review and meta-analysis demonstrated that pituitary insufficiency is a highly prevalent condition in adult patients after cranial radiotherapy for nasopharyngeal and intracerebral tumors. The prevalence of any form of hypopituitarism was 0.66 (95% CI, 0.55–0.76). There were considerable variations in the reported prevalence rates of hypopituitarism after cranial radiotherapy, ranging from hardly any effect on pituitary function to almost 100% of the patients being affected. These variations were associated with differences in the number of patients included in the study and the manner of endocrine evaluation.

The risk of development of hypopituitarism after cranial radiotherapy is a well recognized phenomenon in children. The likelihood to develop hypothalamic-pituitary insufficiency increases with increasing radiation exposure and with prolonged duration of follow-up up after radiotherapy (11;12). In the CCSS 43% of pediatric patients treated for cerebral tumors had one or more endocrinopathies (10). Our meta-analysis showed that hypopituitarism is present in approximately two thirds of all adult patients previously treated with cranial irradiation. The prevalence of hypopituitarism after cranial radiotherapy is affected by several factors including radiation dose and techniques. Furthermore, the time interval between radiotherapy and the assessment of pituitary functions is important because the development of pituitary failure is likely to increase during prolongation of follow-up after radiotherapy (11;12).

A random effects meta-regression revealed no significantly different effects of underlying disease on pituitary function between the two groups of adult patients reported in literature; *i.e.* those treated for nasopharyngeal cancer *vs.* intracerebral tumors. In addition, the overall difference in radiation dosage did not differ between groups: 40–83 Gy for the patients treated for nasopharyngeal carcinoma and 40–97 Gy for the patients with intracerebral tumors. The use of different radiotherapeutical techniques, however, will most likely affect the rate of subsequent hypopituitarism because higher cumulative radiation doses are associated with increasing incidence rates of pituitary failure (11;12).

Patients treated for nasopharyngeal tumors are usually treated with a higher average dosage and additive high dose single tumor boost. Therefore, a separate analysis of the studies concerning nasopharyngeal tumors vs. the other studies was performed (Figure 2A). However, there were no significant differences in the prevalence of any pituitary insufficiency between both groups. There are various possible explanations for this lack of significant differences between the two patient groups despite differences in irradiation dose. In nine studies from which the estimated doses delivered to the pituitary could be extracted (six involving patients treated for nasopharyngeal tumors and three for intracerebral tumors), the dose ranges were wide in both patient groups and showed a considerable overlap. In patients treated for nasopharyngeal tumors, the dose ranged from 46-83 Gy, and in patients with intracerebral tumors the dose ranged from 25-97 Gy. Duration of follow-up since radiotherapy varied from 11-253 months in the patient group treated for nasopharyngeal tumors and from 12 - 156 months in the group treated for intracerebral tumors. Finally, the overall prevalence of pituitary insufficiency is already high in both groups with 0.54 (95% CI, 0.42-0.66) for intracerebral tumors vs. 0.74 (95% CI, 0.57-0.86) for nasopharyngeal tumors. This 0.2 difference in prevalence, however, did not result in significant differences between groups (P = 0.14).

According to our risk of bias stratification, nine studies were considered as being at low risk of bias. The majority of studies used cross-sectional study designs with large differences in the time of evaluation in relation to previous radiotherapy. The selection of patient groups differed largely between the studies. Patient selection criteria were not stated in some of the reports (20;28;31), whereas the description of the selection procedures in other studies suggests preselection of patients, like recruitment from a radiotherapy complication outpatient clinic or by including only symptomatic patients suspected for any degree of hypopituitarism (21;30;32;33). Two studies, (n=186), used a prospective design; the reasons for loss to follow-up were not mentioned in any of the studies, precluding definite answers after 5 yr of follow-up (23;26). We additionally performed a sensitivity analysis of studies that qualified as a potential low – intermediate risk of bias (23;29;36;39). This analysis revealed a pooled proportion of any pituitary deficiency of 0.62 (95% CI,

0.45-0.77), which is remarkably close to the overall found prevalence of 0.66 (95% CI, 0.55-0.76) calculated for all studies. This outcome illustrates that signs and symptoms of hypopituitarism after cranial irradiation apparently are not predictive of hypopituitarism in individual patients with their condition.

Differences in endocrine evaluation may also affect the reported prevalence. The majority of patients were not evaluated by proper stimulation tests. Therefore, it is likely that the estimates of hypopituitarism after cranial radiotherapy represent a rather conservative estimation of the true prevalence of hypopituitarism. In addition, studies that did perform stimulation tests used different tests and cut-off values, and only 10 of 18 studies assessed all pituitary axes (but even then, not all patients were tested for each axis). Moreover, the hypothalamicpituitary-thyroidal axis and the hypothalamic-pituitary-gonadal axis can also be influenced by the use of alkylating chemotherapeutics and exposure of the thyroid gland and gonads to irradiation (19; 41–43). Primary hypothyroidism in patients treated for nasopharyngeal carcinoma occurs up to 20-30% within the first year after treatment (44). In the random effects regression model the prevalence of TSH deficiency was 0.33 (95% CI, 0.19–0.5), and 0.16 (95% CI, 0.08–0.32) for the patients treated for intracerebral tumors. These differences were not significant, with P=0.11. Hypogonadism after cranial radiotherapy was difficult to quantify because some studies reported testosterone or estrogen levels, whereas others used delayed or insufficient LH/FSH responses to GnRH as a criterion for hypogonadism. In addition, primary gonadal failure is highly prevalent in patients treated with chemotherapy, especially when treated with alkalyting agents which could overestimate the results (41). Patients treated for nasopharyngeal tumors were more likely to receive chemotherapy but, again, no significant differences between groups were found.

Prolactin might be another possible tool to estimate the likelihood of radiation induced damage of the hypothalamic area. Because hyperprolactinemia might be a consequence of decreased hypothalamic dopamine secretion, hyperprolactinemia is variable in severity and often subclinical; it diminishes and might even normalize in time due to slowly evolving radiationinduced damage of lactotrophs. If hyperprolactinemia is to be considered as an indicator of disturbed hypothalamic dopamine secretion, one could expect the prolactin level to be high in the first years after radiation and normal after several years when lactrotroph function has declined. Unfortunately, none of the studies provided information on prolactin levels in relation to the time interval since radiotherapy.

Considering the high prevalence of hypopituitarism found, 0.66 (95% CI, 0.55–0.76), and that there was no significant difference between groups, assessment of pituitary function should be included in the long-term follow-up of all cranially irradiated patients. Current literature does not provide a timeline or a sequence of axis failure. Hypopituitarism can occur as soon as the first year after treatment, but can also occur 11 yr after treatment (23;26). Taken into consideration the improved survival of patients, duration of follow-up for at least 15 yr should be advisable. This follow-up period should include a basal morning hormone sample and dynamic testing of the HPA axis in every patient not on corticosteroids. Dynamic testing of the somatotrope axis should be defined per patient, since GH failure will not have therapeutical consequences, although it might be an indicator for radiation induced pituitary damage.

This systematic review underscores the need for structured, periodical, endocrine assessments of all patients who survive after cranial radiotherapy for all kinds of diagnosis. Therefore, an increasing number of patients will require a structured tailored periodical evaluation of pituitary functions after cranial radiotherapy.

References

- 1. Alvarez JA, Scully RE, Miller TL, Armstrong FD, Constine LS, Friedman DL, Lipshultz SE 2007 Long-term effects of treatments for childhood cancers. Curr Opin Pediatr 19:23–31
- 2. Constine LS, Woolf PD, Cann D, Mick G, McCormick K, Raubertas RF, Rubin P 1993 Hypothalamicpituitary dysfunction after radiation for brain tumors. N Engl J Med 328:87–94
- Oeffinger KC, Mertens AC, Sklar CA, Kawashima T, Hudson MM, Meadows AT, Friedman DL, Marina N, Hobbie W, Kadan-Lottick NS, Schwartz CL, Leisenring W, Robison LL 2006 Chronic health conditions in adult survivors of childhood cancer. N Engl J Med 355:1572–1582
- Schmiegelow M, Lassen S, Poulsen HS, Feldt-Rasmussen U, Schmiegelow K, Hertz H, Muller J 2000 Cranial radiotherapy of childhood brain tumours: growth hormone deficiency and its relation to the biological effective dose of irradiation in a large population based study. Clin Endocrinol (Oxf) 53:191–197
- Schmiegelow M, Feldt-Rasmussen U, Rasmussen AK, Poulsen HS, Muller J 2003 A populationbased study of thyroid function after radiotherapy and chemotherapy for a childhood brain tumor. J Clin Endocrinol Metab 88:136–140
- 6. Shalet SM, Beardwell CG, Jones PH, Pearson D 1976 Growth hormone deficiency after treatment of acute leukaemia in children. Arch Dis Child 51:489–493
- 7. Shalet SM, Beardwell CG, Morris-Jones P, Bamford FN, Ribeiro GG, Pearson D 1976 Growth hormone deficiency in children with brain tumors. Cancer 37:1144–1148
- Shalet SM, Beardwell CG, MacFarlane IA, Jones PH, Pearson D 1977 Endocrine morbidity in adults treated with cerebral irradiation for brain tumours during childhood. Acta Endocrinol (Copenh) 84:673–680
- 9. Constine LS, Woolf PD, Cann D, Mick G, McCormick K, Raubertas RF, Rubin P 1993 Hypothalamicpituitary dysfunction after radiation for brain tumors. N Engl J Med 328:87–94
- Gurney JG, Kadan-Lottick NS, Packer RJ, Neglia JP, Sklar CA, Punyko JA, Stovall M, Yasui Y, Nicholson HS, Wolden S, McNeil DE, Mertens AC, Robison LL 2003 Endocrine and cardiovascular late effects among adult survivors of childhood brain tumors: Childhood Cancer Survivor Study. Cancer 97:663–673
- 11. Biermasz NR, van DH, Roelfsema F 2000 Long-term follow-up results of postoperative radiotherapy in 36 patients with acromegaly. J Clin Endocrinol Metab 85:2476–2482
- 12. Toogood AA 2004 Endocrine consequences of brain irradiation. Growth Horm IGF Res 14 Suppl A: S118–S124

- 13. Littley MD, Shalet SM, Beardwell CG, Ahmed SR, Applegate G, Sutton ML 1989 Hypopituitarism following external radiotherapy for pituitary tumours in adults. Q J Med 70:145–160
- 14. Darzy KH, Shalet SM 2009 Hypopituitarism following radiotherapy. Pituitary 12:40–50
- 15. Hamza TH, van Houwelingen HC, Stijnen T 2008 The binomial distribution of meta-analysis was preferred to model within-study variability. J Clin Epidemiol 61:41–51
- 16. Chang BH, Waternaux C, Lipsitz S 2001 Meta-analysis of binary data: which within study variance estimate to use? Stat Med 20:1947–1956
- 17. Platt RW, Leroux BG, Breslow N 1999 Generalized linear mixed models for meta-analysis. Stat Med 18:643–654
- Sweeting MJ, Sutton AJ, Lambert PC 2004 What to add to nothing? Use and avoidance of continuity corrections in meta-analysis of sparse data. Stat Med 23:1351–1375
- Bhandare N, Kennedy L, Malyapa RS, Morris CG, Mendenhall WM 2007 Primary and central hypothyroidism after radiotherapy for head-and-neck tumors. Int J Radiat Oncol Biol Phys 68:1131–1139
- 20. Bhandare N, Kennedy L, Malyapa RS, Morris CG, Mendenhall WM 2008 Hypopituitarism after radiotherapy for extracranial head and neck cancers. Head Neck 30:1182–1192
- 21. Lam KS, Wang C, Yeung RT, Ma JT, Ho JH, Tse VK, Ling N 1986 Hypothalamic hypopituitarism following cranial irradiation for nasopharyngeal carcinoma. Clin Endocrinol (Oxf) 24:643–651
- 22. Lam KS, Ho JH, Lee AW, Tse VK, Chan PK, Wang C, Ma JT, Yeung RT 1987 Symptomatic hypothalamic-pituitary dysfunction in nasopharyngeal carcinoma patients following radiation therapy: a retrospective study. Int J Radiat Oncol Biol Phys 13:1343–1350
- Lam KS, Tse VK, Wang C, Yeung RT, Ho JH 1991 Effects of cranial irradiation on hypothalamicpituitary function--a 5-year longitudinal study in patients with nasopharyngeal carcinoma. Q J Med 78:165–176
- 24. Samaan NA, Bakdash MM, Caderao JB, Cangir A, Jesse J, Ballantyne AJ 1975 Hypopituitarism After External Irradiation. Annals of Internal Medicine 83:771
- Samaan NA, Vieto R, Schultz PN, Maor M, Meoz RT, Sampiere VA, Cangir A, Ried HL, Jesse RH, Jr. 1982 Hypothalamic, pituitary and thyroid dysfunction after radiotherapy to the head and neck. Int J Radiat Oncol Biol Phys 8:1857–1867
- Samaan NA, Schultz PN, Yang KP, Vassilopoulou-Sellin R, Maor MH, Cangir A, Goepfert H 1987 Endocrine complications after radiotherapy for tumors of the head and neck. J Lab Clin Med 109:364–372
- 27. Harrop JS, Davies TJ, Capra LG, Marks V 1976 Hypothalamic-pituitary function following successful treatment of intracranial tumours. Clin Endocrinol (Oxf) 5:313–321
- Constine LS, Rubin P, Woolf PD, Doane K, Lush CM 1987 Hyperprolactinemia and hypothyroidism following cytotoxic therapy for central nervous system malignancies. J Clin Oncol 5:1841–1851

- 29. Johannesen TB, Lien HH, Hole KH, Lote K 2003 Radiological and clinical assessment of longterm brain tumour survivors after radiotherapy. Radiotherapy and Oncology 69:169–176
- Popovic V, Pekic S, Golubicic I, Doknic M, Dieguez C, Casanueva FF 2002 The impact of cranial irradiation on GH responsiveness to GHRH plus GH-releasing peptide-6. J Clin Endocrinol Metab 87:2095–2099
- Schneider HJ, Rovere S, Corneli G, Croce CG, Gasco V, Ruda R, Grottoli S, Stalla GK, Soffietti R, Ghigo E, Aimaretti G 2006 Endocrine dysfunction in patients operated on for nonpituitary intracranial tumors. Eur J Endocrinol 155:559–566
- Woo E, Lam K, Yu YL, Ma J, Wang C, Yeung RT 1988 Temporal lobe and hypothalamic-pituitary dysfunctions after radiotherapy for nasopharyngeal carcinoma: a distinct clinical syndrome. J Neurol Neurosurg Psychiatry 51:1302–1307
- 33. Mechanick JI, Hochberg FH, LaRocque A 1986 Hypothalamic dysfunction following wholebrain irradiation. J Neurosurg 65:490–494
- 34. Huang KE 1979 Assessment of hypothalamic-pituitary function in women after external head irradiation. J Clin Endocrinol Metab 49:623–627
- 35. Rosenthal MB, Goldfine ID 1976 Primary and secondary hypothyroidism in nasopharyngeal carcinoma. JAMA 236:1591–1593
- Agha A, Sherlock M, Brennan S, O'Connor SA, O'Sullivan E, Rogers B, Faul C, Rawluk D, Tormey W, Thompson CJ 2005 Hypothalamic-pituitary dysfunction after irradiation of nonpituitary brain tumors in adults. J Clin Endocrinol Metab 90:6355–6360
- 37. Samaan NA, Bakdash MM, Caderao JB, Cangir A, Jesse RH, Jr., Ballantyne AJ 1975 Hypopituitarism after external irradiation. Evidence for both hypothalamic and pituitary origin. Ann Intern Med 83:771–777
- 38. Snyers A, Janssens GORJ, Twickler MB, Hermus AR, Takes RP, Kappelle AC, Merkx MAW, Dirix P, Kaanders JHAM 2009 Malignant Tumors of the Nasal Cavity and Paranasal Sinuses: Long-Term Outcome and Morbidity With Emphasis on Hypothalamic-Pituitary Deficiency. International Journal of Radiation Oncology Biology Physics 73:1343–1351
- Taphoorn MJ, Heimans JJ, van der Veen EA, Karim AB 1995 Endocrine functions in long-term survivors of low-grade supratentorial glioma treated with radiation therapy. J Neurooncol 25:97–102
- 40. Chen MS, Lin FJ, Huang MJ, Wang PW, Tang S, Leung WM, Leung W 1989 Prospective hormone study of hypothalamic-pituitary function in patients with nasopharyngeal carcinoma after high dose irradiation. Jpn J Clin Oncol 19:265–270
- 41. Levine J, Canada A, Stern CJ 2010 Fertility preservation in adolescents and young adults with cancer. J Clin Oncol 28:4831–4841
- 42. Jereczek-Fossa BA, Alterio D, Jassem J, Gibelli B, Tradati N, Orecchia R 2004 Radiotherapyinduced thyroid disorders. Cancer Treat Rev 30:369–384

- 43. Mercado G, Adelstein DJ, Saxton JP, Secic M, Larto MA, Lavertu P 2001 Hypothyroidism: a frequent event after radiotherapy and after radiotherapy with chemotherapy for patients with head and neck carcinoma. Cancer 92:2892–2897
- 44. Lin Z 2010 A Longitudinal Study on the Radiation-Induced Thyroid Gland Changes After External Beam Radiotherapy of Nasopharyngeal Carcinoma.
- 45. Lam KS, Tse VK, Wang C, Yeung RT, Ma JT, Ho JH 1987 Early effects of cranial irradiation on hypothalamic-pituitary function. J Clin Endocrinol Metab 64:418–424

Chapter 7

Growth hormone replacement therapy in elderly growth hormone deficient patients: a systematic review

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Abstract

Context: Recombinant human GH (rhGH) is prescribed for the treatment of adultsz with GH deficiency (GHD). However, conflicting data are available on the efficacy of rhGH treatment in elderly GHD patients.

Objective: To assess the efficacy of rhGH treatment in elderly GHD subjects.

Methods: We searched PubMed, Cochrane Library, Web of Science and EMBASE.

Study selection: Eligible studies included GHD patients, aged > 60 years, treated with rhGH. Data extraction was performed by two reviewers independently.

Results: We found 11 eligible studies with a total of 534 patients. Only two studies had prospective, randomized, placebo-controlled study designs of rhGH treatment with a duration of 6 (n=15) and 12 months (n=62), respectively. Treatment with rhGH decreased total and low density lipoprotein (LDL) cholesterol levels by 4–8 and 11–16%, respectively, but did not alter high density lipoprotein or triglyceride levels. RhGH did not affect body mass index, but decreased waist circumference (by ~3 cm) and waist/hip ratio. RhGh did not consistently affect blood pressure or bone mineral density. RhGH increased lean body mass by 2–5% and decreased total fat mass by 7–10% in four studies, but did not affect body composition in two other studies. RhGH consistently improved quality of life (QoL) parameters reflected in AGHDA-scores. There are no explicit data on elderly GHD patients aged > 80 years.

Conclusion: RhGH replacement in elderly subjects with GHD decreases LDL cholesterol levels and improves QoL, but the effects on other parameters are not unequivocal. There are no data on the efficacy and safety of rhGH treatment in octogenarians with GHD.

Introduction

In healthy adults, GH secretion declines with increasing age (1;2). Some of the clinical features of normal aging resemble the manifestations of pathological GH deficiency (GHD). These features include changes in body composition (BC), such as increased total fat mass, decreased lean body mass (LBM) and decreased bone mass, as well as a higher prevalence of cardiovascular risk factors and diminished cardiac function (3).

Consequently, a number of studies have examined the effect of recombinant human GH (rhGH) on various clinical parameters in otherwise healthy elderly subjects (4) as well as in elderly patients with GHD. Rudman et al. (1990) (5) were the first to report that 6 months of rhGH treatment in healthy elderly men reduced adiposity and increased muscle mass and bone mineral density (BMD). Other studies observed similar beneficial effects, suggesting a potential role for rhGH as anti-aging therapy (6-8). A recent systematic review and metaanalysis of randomized controlled trials observed that rhGH treatment decreased overall fat mass decreased by ~2.1 kg and increased overall LBM increased by ~2.1 kg (CI, 1.3-2.9) (P<0.001), without any effect on weight. However, rhGH was associated with increased rates of adverse effects (4). Moreover, in healthy elderly subjects, higher physiological insulin like growth factor-I (IGF-I) concentrations are associated with increased mortality (9). Several studies assessed the clinical benefits of rhGH therapy in elderly GHD patients. Some studies in elderly patients with GHD documented that, rhGH improved QoL (10;11), BC (12–14) and lipoprotein profiles (11, 15, 16), although another study showed no effects (17). A consensus statement on the treatment of GHD adults states that 'the age-related decline in the GH-IGF-I status does not warrant rhGH supplementation, but patients with proved GHD should be treated'. These guidelines indicate that the dose of rhGH should be adjusted with advancing age, because of the normal age-related decline in GH secretion (18). Apparently, there is no clear age limitation in treating elderly GHD adults with rhGH.

The aim of the present study was to critically assess the available literature in order to evaluate the available evidence for treatments of elderly patients with GHD. Therefore, we performed a structured review of the available literature on this subject.

Subjects and methods

Search strategy

We performed a systematic search in the following database: PubMed, EMBASE, Web of Science, Cochrane Library, CINAHL database and Academic Search Premier. The search strategy included three main issues: growth hormone deficiency, age > 60 years and growth hormone replacement therapy. We used all relevant keyword variations, including free text words. This resulted in the following search string: ((("Growth *deficiency*"[*ti*] OR "Growth Hormone/deficiency" [Majr] hormone OR "GH deficiency"[ti] OR "Growth hormone deficient"[ti] OR "GH deficient"[ti] OR GHD[ti]) AND ("aged"[mesh] OR elderly[tw] OR oldest old OR Nonagenarians OR Nonagenarian OR septuagenarian OR septuagenarians OR Octogenarians OR Octogenarian OR Centenarians OR Centenarian)) OR (("Growth hormone deficiency"[ti] OR "Growth Hormone/deficiency"[Majr] OR "GH deficiency"[ti] OR "Growth hormone deficient"[ti] OR "GH deficient"[ti] OR GHD[ti] OR "growth hormone deficiency" OR "growth hormone deficient" OR "gh deficiency" OR "gh deficient") AND ("Growth Hormone/administration and dosage" [Mesh] OR "Growth Hormone/therapeutic use" [Mesh] OR "Growth Hormone/ therapy"[Mesh] OR ((growth hormone OR growth hormones OR Somatotropin OR Somatotropins) AND (therapy OR therapeutic OR replacement))) AND ("aged" [mesh] OR elderly [tw] OR oldest old OR Nonagenarians OR Nonagenarian OR septuagenarian OR septuagenarians OR Octogenarians OR Octogenarian OR Centenarians OR Centenarian))) AND (English [lang]). Furthermore, the references of relevant articles were checked for additional articles. The following exclusion criteria were used: age < 60 year, non-GHD subjects, no rhGH therapy.

Data review

The following data were extracted from each study: 1) age, gender and number of patients, 2) the endocrine tests used to diagnose GHD, 3) criteria used to define GHD, 4) duration of treatment and treatment dose, and 6) the effect of GH on individual outcome parameters.

Results

The initial search resulted in a total of 577 articles (527 in PubMed, 3 in Cochrane Library, 15 in Web of Science, and 26 in EMBASE, 0 CINAHL database and 6 Academic Search Premier). Of these 577 articles, 534 were unique without duplications. We excluded 403 papers based on title and abstract (studies on GHD without specific focus on rhGH replacement therapy and/or age < 60 yr (n=326), reviews (n=77)). In 89 additional papers, which included patients with an age> 60 yrs, the individual data of the patients could not be extracted. Two additional papers were not available for evaluation.

Therefore, a total of 40 potentially relevant manuscripts were retrieved for full assessment, of which 26 studies were excluded from further analysis because those studies did not meet one or more of the eligibility criteria (age < 60 yrs, no rhGH therapy, healthy elderly).

Ultimately, the search strategy resulted in a total of 14 manuscripts meeting our inclusion criteria. However, only eight different cohorts of patients were described in these 14 studies, because several studies described data from the same patient cohort. The studies by Götherström *et al.* 2010 (12) and Götherström *et al.* 2005 (19) described the same patient cohort (n=24) after 5 and 10 years of rhGH treatment. The studies by Elzgyri *et al.* (16) and Fernholm *et al.*(13) (n=31) as well as Gill *et al.* (14) and Toogood *et al.*(20) (n=12) also described the same patient cohort. Therefore, the data of these studies are described in combination (12–14;16;19;20). The studies reported from the KIMS database included different numbers of patients in each publication (n=64, n=125, n=135, n=64). Although it is likely that similar patients have been included, the different numbers of subjects preclude combination of the data of these studies are studies and these studies are studies.

Consequently, a total of 11 studies were included in the present review, comprising 534 patients (Figure 1).

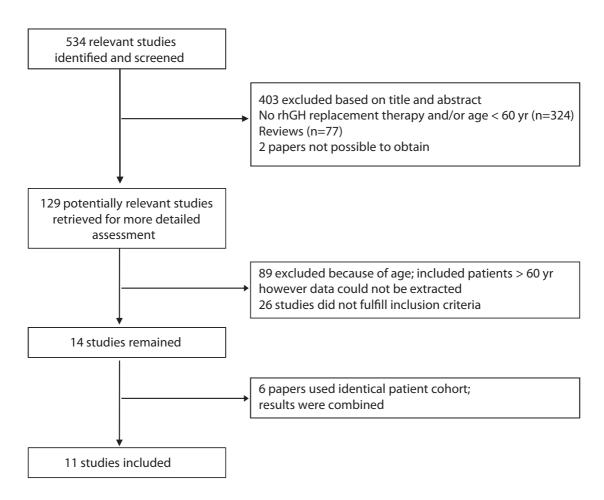


Figure 1. Summary of study assessment and exclusion stages

Study designs

Two studies (n=65) had a prospective placebo-controlled, randomized design assessing the effects of rhGH treatment during 6 (n=15) and 12 n=62) months, respectively, in elderly GHD patients (16;17). The study by Elzgyri *et al.* (2004) (16), that evaluated the effects of rhGH versus placebo for six months, was continued for another 12 months using a nonrandomized prospective study design. The other nine studies had a non-randomized prospective study design, in which the basal data prior to rhGH treatment were used to assess the effects of rhGH (n=469). Four of these studies (n=388) used patients derived from the KIMS database (10;11;15;21).

In this review, we only included studies with patients above the age of 60 years. Two studies did not specify the age of the patient, but only indicate that all patients are >60 years (10;11). One study assessed

patients between the age 60 and 70 years (22), and in three studies, the age ranged between 60 and 75 years (12;21;23). Five studies included patients >75 years and only two of these studies included patients with age >80 years (14–17;24).

Endocrine evaluation

All 11 studies used stimulation tests for the evaluation of GH reserve (Table 1). Different tests, however, were used, including insulin tolerance test (ITT), combined GHRH plus arginine (GHRH-arginine) test, glucagon stimulation test or stimulation with arginine alone (Table 1). The ITT was used in the total population or in the majority of patients in seven studies (11;12;15;17;21–23). The study by Koltowska *et al.* (2009) (10) did not mention which tests were used to diagnose GHD. However, that study also described patients from the KIMS database, and, therefore, it can be assumed that the ITT was also used in the majority of these patients. The remaining three studies either used GHRH tests (16), arginine tests (14) or combined GHRH-arginine tests (24). All but one studies applied the generally used cut-off value for severe GHD of a peak GH < 3 μ g/L.

In the study by De Marinis *et al.* (24), which used the GHRH-arginine test, severe GHD was defined as a GH peak < $15 \mu g/L$, without corrections for the effects of body mass index (BMI).

Duration and dose of rhGH treatment

The duration of treatment with rhGH ranged from 6 months (n=15) to 10 years (n=24).

Six studies (n=390) calculated the GH dose based on bodyweight (11;12;15;16;21;24). One of these studies used a predefined dose per kg bodyweight per day (0.0119 mg/kg/day) (12). The other five studies titrated the rhGH dose per kg per week (0.017 – 0.042 mg/kg/wk) (11;15; 16;21;24).

Of the remaining five studies, one study gave three fixed doses of rhGH (0.17, 0.33, and 0.5 mg/day) for 12 weeks. These patients received the highest dose of rhGH, *i.e.* 0.5 mg/day (14). In the remaining four studies, rhGH was titrated on an individual basis with the aim to reach IGF-I levels within the normal age- and sex-related range or clinical improvements, taking QoL and BC into account. The mean dose of rhGH ranged from 0.11 to 0.37 mg/day (11;15;21;24).

IGF-I levels and SD scores

Seven of the 11 studies used age-adjusted IGF-I levels measured in a control population to titrate rhGH dose (11;12;14;16;21–23). Götherström *et al.* and Franco *et al.* 2005 both used the same references values measured in a group of 392 patients aged between 25 and 64 years, as described by Landin-Wilhelmsen *et al.* 1994 (25). Feldt-Rasmussen and Monson and colleagues both refer to a study by Drake *et al.* 1998 (26). However, that study does not mention the IGF-I reference values, but refers to a dose finding study by Janssen and colleagues 1997 (27), in which normative data were based on 54 healthy control subjects aged 20–70 years. The study by Elzgyri *et al.* 2004 use reference values derived from the study by Hilding *et al.* 1999 (28), in which IGF-I values are measured in a population of 448 healthy controls aged 20–96 years. The remaining two studies measure reference values in an own reference population of 450 (18–80 years) (22) and 124 (60–84 years) (14) healthy controls, respectively.

All studies titrated the rhGH dose with the aim of normalizing IGF-I S.D. scores, *i.e.* aiming at IGF-I SDS in physiological levels for age and sex (between -2 and +2). However, four studies also took the clinical response and BC into account for titration of the rhGH dose. Feldt-Rasmussen *et al.* 2004 and Monson *et al.* 2000 both took the clinical response into account when titrating rhGH dose, referring to a study by Drake *et al.* 1998 (26), in which waist/hip (W/H) measurements and improvement of QoL measured by AGHDA were taken into account for the titration of the dose. The remaining two studies (12;23) both state that, when adjusting the rhGH dose, the aim is to 'normalizing IGF-I and BC.', both referring to a study by Johannssen *et al.* 1997 (7), in which individualized doses of rhGH are Q2 compared with doses based on body weight. In their study, normalization of BC was of great importance and normal values derived from a study by Bruce *et al.* 1980 (29), comprising 376 patients in the age 20–70 years, were used to evaluate patients and adjust rhGH levels.

Effects of rhGH on cardiovascular and metabolic parameters

Five studies (n=424) assessed the effects on plasma lipid profiles (11;15;16;21;23). In general, rhGH treatment decreased total and low density lipoprotein (LDL) cholesterol levels by 4-8% (15;16;23) and by 11–16%, respectively, whereas rhGH increased high density lipoprotein

(HDL) only by 17% in one other study (16). Treatment with rhGH did not affect triglyceride (TG) levels at all.

Six studies assessed the effects of rhGH on body weight, height, BMI and/or W/H ratios (11;12;15;21;23;24). In two studies, rhGH did not affect BMI (12;24). In three of five studies which reported W/H ratios, there was a significant decrease in waist circumference (3 cm in de study by Franco *et al.* 2006 (23)) and W/H ratios (11;15;21;23). However, the two other studies did not find any effect of rhGH on W/H ratios (15;24).

Five studies assessed the effects of rhGH on blood pressure (BP) (n=379) (11;15;16;21;23). There were no clear consistent effects of rhGH treatment on BP. Treatment with rhGH did not affect (16), only transiently decreased BP (23), or decreased diastolic BP only (11;15;21).

One study used an exercise test to evaluate cardiac function (n=31). Treatment with rhGH induced a transient increase in heart rate at rest and exercise. However, rhGH treatment did not affect cardiac structural and functional parameters (16).

Effects of rhGH on bone parameters

The effects of rhGH therapy on bone metabolism were evaluated in three studies (n=65). Treatment with rhGH did not affect BMD. One study found that treatment with rhGH increased osteocalcin and calcium levels without any change in PTH levels (23). Another study found that rhGH treatment lowered PTH and urinary cAMP levels, associated with higher adjusted calcium and bone turnover markers, indicating a higher PTH target organ sensitivity (22). The third study observed that treatment with rhGH induced higher markers for bone formation (bone-specific alkaline phosphatase activity, osteocalcin and procollagen I carboxyl-terminal peptide in serum) (13). The effects of rhGH treatment on fracture incidence were not described.

Effects of rhGH on body composition

Six studies assessed the effects of rhGH on body composition using DEXA scan (n=138) (11;12;14;16;23;24). Two studies (n=35) found no effect of rhGH on body composition (23;24). In contrast, the other 4 studies (n=103) found that 6 months of rhGH treatment induced a significant increase in lean body mass (LBM) by 2–5% and a significant decrease in total body fat by 7–10% (12;14;16). Moreover, these effects of rhGH on body composition were reversed, when rhGH therapy was subsequently

stopped (n=12) (14). One study used a four compartment model to assess body fat, body cell mass and extra cellular weight, but these parameters were not affected by rhGH therapy (12;19).

Effects of rhGH on QoL and cognitive functioning

The effects of rhGH treatment on quality of life (QoL) parameters was assessed using only the AGHDA questionnaire in five studies (n=400) (10;11;14;15;21). The majority of these patients were from the KIMS database (n=388). Treatment with rhGH induced significant improvements of AGHDA scores in all studies.

Only one study assessed cognitive functioning (n=34) using computerized psychometric test package (Neurobehavorial Examination-System 2). However, compared with placebo rhGH therapy was not associated with improvement in cognition after 12 months.

Effects of rhGH on muscle strength

One study assessed the effects of 5 and 10 years of rhGH treatment on muscle strength (n=24) (12). Treatment with rhGH induced a transient improvement only in knee flexor strength. However, rhGH treatment protected from most of the normal age-related decline in muscle performance and neuromuscular function.

Adverse effects

Six of the 11 studies mention possible adverse effects of rhGH treatment. In two studies the number of adverse events (AEs) was similar for younger and older patients with GHD (11;15). However, younger patients appeared to have more AEs related to fluid retention (*i.e.* headaches, oedema and arthralgias), whereas patients > 65 yrs had more AEs related to glucose metabolism, cerebrovascular events and neoplasms (11). One of the two placebo-controlled studies mentioned AEs and found no differences between the placebo and rhGH groups (17).

In the study by Fernholm *et al.* 25% of the patients (8/31) developed side effects probably due to fluid retention (peripheral edema, joint stiffness and muscle pain).

However, these side effects subsided spontaneously or after minor dose reduction (13). The study using the highest dose, found AEs in 3 of the 12 patients. The AEs subsided when the dose was down titrated (20). One study needed to reduce the dose of rhGH because of symptoms of the carpal tunnel syndrome (22).

	Treat-						
	ment						
Author Patients	Definition GHD durat	duration Dose	IGF-I SDS	Evaluation	Effect of treatment		
					Metabolic syndrome		
							Body com- QoL/
					Anthropometrics Lipids	Bone	position Cognition
Götherström n=24	ITT(n=22) 10 yr	Mean initial	After 10 yrs:	Physical exam.	BW: ↓ -	1	BF:↓ -
<i>et al.</i> ; 2010 ¹² M: 11	GHRH-pyr (n=1)	dose 0.72 mg/	\uparrow -1.10(1.08) to	BC (DEXA)	Height: ↓		LBM:↑
65 (61–74) yr	Low IGF-I (n=2)	day in 5	1.17(1.52)		BMI: =		
		years	/	strength*			
Götherström et al < 2005 ¹⁹	Peak GH < 3 μ g/L 5 yr	day					4-comp model
		:					
		Aim normalizing	D				
		IGF-I SDS (-2 to					and
		+2) and BC					ECW: =
Koltowska KIMS	ITT 6 yr	Mean after 1 yr: Baseline:	: Baseline:	QoL (AGHDA)		ı	- Both
<i>et al.</i> ; 2009 ¹⁰ n=64 > 60 yr	Arginine	0.26-0.40 mg/	-1.3(1.19) to				groups:
	GHRH	day	-2.1(1.75)				AGDHA: +
n=286 < 60 yr	Glucagon						
			After 1 year:				
	Peak GH < 3 μg/L		0.8(1.4) to				
			0.5(1.31)				

 Table 1.
 Studies on rhGH therapy in GHD elderly patients

Chapter 7

			Treat-							
			ment							
Author	Patients	Definition GHD	duration Dose	Dose	IGF-I SDS	Evaluation	Effect of treatment	Ŀ		
							Metabolic Syndrome	•		
							Automotion Linit	G	Body com- QoL/	QoL/ Cognition
										COGINICOL
Feldt-Ras-	KIMS		12	Mean 0.2 mg/	Baseline:	Physical exam.		n=107): -	Both	Both
mussen <i>et a</i> i	mussen <i>et al.</i> ; n=125 > 65 yr	Arginine	months	day	Between 0	Lipids	Waist (n=114): 🔱		groups: groups:	groups:
2004 ¹¹		GHRH			and -4	BC (DEXA)	↑ FDL:↓	→ 	LBM	AGHDA
	n=2469 < 65 yr	Glucagon		Titration based		QoL (AGHDA)	W/H-ratio: \ HDL	HDL; TG: =	(n=47):↑	(n=47):↑ (n=98): +
	M: 1249			on clinical res- After 12 months:	After 12 month:	5:				
		Peak GH < 3 µg/L		ponse and IGF-I Between -2	Between -2		BP:			
				SDS	and +2		< 65 yr: DBP			
							± →			
							> 65 yr: DBP			
							×			
Monson	KIMS	ITT 55%	12	Start dose max Baseline:	Baseline:	Physical exam.	Both groups: TC: (1	ı	Both
<i>et al.</i> ; 2003 ²¹	n=135 > 65 yr	Arginine 14%	months	0.042-0.083	M: median -1.60 Lipids) Lipids	Waist (n=93): \ LDL (n=78):	(n=78):		groups:
	(65–74)	Glucagon 5%		mg/ kg/week	(-3.19 to +0.28) QoL (AGHDA)	QoL (AGHDA)	W/H-ratio: 🔶 🗼			AGHDA
	M: 83	Other 26 %			F: median -1.78		HDL	HDL; TG: =		(n=78): +
					(-3.81 to 0.06)		BP(n=123):			
	n= 1395 < 65 yr	Peak GH < 3 µg/L					< 65 yr: DBP			
					After 1 yr:		± →			
					Between -2		> 65 yr: DBP			
					and +3		₩↑			

Table 1.	Table 1. Continued										
			Treat-								
			ment								
Author	Patients	Definition GHD duration Dos	duration	Dose	IGF-I SDS	Evaluation	Effect of treatment	tment			
							Metabolic Syndrome	drome			1
									I	Body com- QoL/	QoL/
							Anthropometrics Lipids	s Lipids	Bone	position	Cognition
Monson	KIMS	ITT 44%	> 6	Mean:	Between -2	6 months	Waist:	TC:↓	ı	I	Both
<i>et al.</i> ; 2000 ¹⁵ n=64	⁵ n=64	Arginine 19%	months	0.37 mg /day	and+2	(n=64):	< 65 yr \	LDL: U			groups:
	68 (65–82) yr	GHRH 13%				ВР	Only F > 65	HDL; TG: =			AGHDA: +
		Glucagon 11%		Titration based		QoL (AGHDA)	yr ↓				
	n=863 < 65 yr			on clinical res-							
		Peak GH < 3 μg/L		ponse and IGF-I	_	12 months	BP:				
				SDS		(n=22):	> 65 yr: DBP↓				
						Lipids	W				
						Bone					
Sathiava-	n=34	ITT	12	Mean:	Target:	Fasting glucose, HbA1c, in-	, HbA1c, in-		I	I	Cognition:
geeswaran	M: 22	Arginine	months	0.16 mg/day	Between +1	insulin, HbA1c sulin, fasting	sulin, fasting				No bene-
<i>et al.</i> ; 2007 ¹	<i>et al</i> .; 2007 ¹⁷ 66 (60–77) yr			(0.10-0.30)	and +2	Cognitive func- glucose: no	glucose: no				fits
		Peak GH < 3 µg/L				tion	differences				
	n=16 rhGH			Titration based		Mood					
	n=18 placebo			on IGF-I SDS							

CHAPTER 7

184

Author Pati											
			ment								
	Patients	Definition GHD	duration Dose	Dose	IGF-I SDS	Evaluation	Effect of treatment	ment			
							Metabolic Syndrome	rome			
										Body com- QoL/	QoL/
							Anthropometrics Lipids	Lipids	Bone	position	Cognition
Franco <i>et al.</i> ; n=24	4	ITT (n=41)	2 yr	Mean:	Target:	Physical exam.	Both groups:	> 65 yr:	Both	Both	
2006 ²² M: 15	5	GHRH (n=3),		0.31(0.03) mg/	0.31(0.03) mg/ Between 0 and	BC (DEXA)	BW:=	LDL: U	groups:	groups:	
68 ((68 (65–75) yr	Glucagon (n=1)		day	+2	Bone (DEXA)		TC:↓		No diffe-	
						Lipids	W/H-ratio: ↓		cin:↑	rences	
n=24	4	Peak GH < 3 μg/L		Normalizing age	0	Glucose metab			Calcium: ↑	←	
M: 15	5			adjusted IGF-I			Transient BP 🔱		PTH: unaf-		
37 (2	37 (27–46) yr			levels and BC					fected		
									BMD: no		
									differen-		
									ces		
White et al.; n=10	0	TTI	12	Mean:	Baseline:	Bone	ı		Both	ı	ı
2005 ²³ M: 5			months	0.29(0.03) mg/	Mean -2.72 ±				groups:		
90-6	60–68 yr	Peak GH < 3 μg/L		day	1.17				PTH tar-		
									get-organ	-	
n=22	2			Maintaining	After 1 month:				sensiti-		
M: 6				IGF-I within 2 SDMean -0.38 ±) Mean -0.38 ±				vity:⊠**		
26-5	26–57 yr			of age related	0.96						
				ref range							

Continued	
Table 1.	

					ion													
				1- QoL/	position Cognition													
				Body com- QoL/	position													
					Bone													
		itment	drome		s Lipids	6 months	Both			LDL:	A LDL/HDL	ratio: ↓		12 months	†:C:↓	↑:TDL:↓	HDL:↑	
		Effect of treatment	Metabolic Syndrome		Anthropometrics Lipids	6 and 12	months	No changes	on cardiac	noninvasive	structural and LDL/HDL	functional	parameters.					
		Evaluation				BP, heart rate	Lipids	ECG	Exercise tests			i				10		
		IGF-I SDS				Baseline:	3 All patients	below normal	mean for age		336 months:	6.9–18.5 nmol/L	(10.4–32.8)		12 months:	Mean 18.8+/-1.6		
Treat-	ment	duration Dose				GHRH test n=29 6 months 1 month: 0.017 Baseline:	GH: n=15 5 months: 0.033 All patients	Placebo: mg/kg/week	n=16	1 month 0.017	12 11 months 0.0336 months:	months mg/kg/week	GH: n=28	Normal range	IGF-I assessed	by n=448 he-	althy subjects	(20–96 yr)
		Definition GHD duration Dos				GHRH test n=29	ITT n=2		Peak GH < 3 μg/L n=16									
		Patients				<i>l</i> .; n=31	M: 25	68 (60–79) yr										
		Author				Elzgyri et al.; n=31	2004 ¹⁶											

		Treat- ment					
Author Patients	s Definition GHD	duration Dose	ise IGF-I SDS	Evaluation	Effect of treatment		
					Metabolic Syndrome		
						Body com- QoL/	/70
					Anthropometrics Lipids	Bone position C	Cognition
Fernholm				BC (DEXA)	BP:=	BMD: no 6 and 12	
<i>et al</i> .; 2000 ¹³				Bone		changes months:	
						Markers Only	
						bone for- in M	
						mation: ↑ Placebo:	
						no res-	
						ponse	
						GH	
						groups:	
						LBM:↑	
						TBF:	
De Marinis n=11	GHRH-arg	12 0.0	0.06 – 0.12 IU/ NR	BMI	BMI: =	- No chan	
<i>et al.</i> ; 2002 ²⁴ M: 6		months kg	kg/wk	Waist	W/H ratio: =	ges	
60–78 yr	r Peak GH < 15			BC			
	hg/L	Ke	Keep IGF-I levels				
n=39		ini	in psychological				
M: 19		rar	range for age				
18–57 yr	L	ano	and sex				

			Treat-						
			ment						
Author	Patients	Definition GHD	duration Dose	IGF-I SDS	Evaluation	Effect of treatment			
						Metabolic Syndrome			I
								Body com- QoL/	· QoL/
						Anthropometrics Lipids	Bone	position	Cognition
Gill <i>et al.</i> ;	n=12	Arginine	9 months Acute study:	Baseline:	BC (DEXA)			FM: (AGHDA: +
1999 ¹⁴	M: 9		single bolus 0.1 Between	Between	QoL (AHGDA)			LBM:↑	
	68 (62–85) yr	Peak GH < 3 μg/L		-2 and 0					
			Chronic study.	0 17 mg/dav					
			0.17 mg/day	normal limits					
			0.33 mg/day	0.33 mg/day:					
			0.5 mg/day,	n=2 > +2					
			Each dose 3	0.50 mg/day:					
			months	n=6 > +2					
			per eidzeniteled	C					
				U					
			and IGF-I asses-						
			sed in group						
			n=124 (60-87						
			yr) with specific						
			equations***						

188

Chapter 7

Muthor Patients Definition GHD duration IGF-1 SDS Evaluation Effect of treatment nonconstructure Anthropometrics Lippin Anthropometrics Body constructure Ool Toopood Lippin Leptin Actute study: Bondy constructure Cognition Cognitio				Treat- ment						
Metabolic Syndrome Anthropometrics Lipids Bone Anthropometrics Lipids Bone Insulin Leptin Acute study: P Insulin Leptin: P BC (DEXA) Insulin: Chronic study: Eptin: BC (DEXA) Insulin: Chronic study: Eptin: Eptin: BC (DEXA) Insulin: Chronic study: Eptin:	Author	Patients	Definition GHD	duration Dose	IGF-I SDS	Evaluation	Effect of treatment			
Body.com- Ool/ Anthropometrics Lipids Body.com- Ool/ Body.com- Ool/ Anthropometrics Lipids Bone Bosition Cognition Toogood Leptin Acute study: Insulin Leptin: 1 Borie position Cognition Foldy com- Col/ et di, 1999 Insulin Leptin: 1 Acute study: Insulin: 1 Connic study: Insulin: 1 Connic study: Insulin: 1 Chronic study: Insulin: 1 Chronic study: Insulin: 1 Chronic study: Insulin: 1 Chronic study: Insulin: 1 Insulin: 1							Metabolic Syndrome			
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Toogood Leptin Acute study: insulin Insulin Leptin: ↑ <i>et al.</i> ; 1999 Leptin: ↑ Ecptin: ↑ BC (DEXA) Insulin: ↑ Ecptin: ↑ AGHD, Assessment of Growth Hormone Deficiency in Adults; BC, body composition; BCM, body real mass; BF, body fat; BMD, hormone; GHRH, growth hormone; GL, quality of life; SBP, systolic blood pressure; IC, traisly regiveredes; WH, waist/hip; =, no changes							Anthropometrics Lipids	Bone	position	Cognition
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** PTH↓ nephro cAMP↑ adjusted calcium ↑, bone turnover markers ↑ *** Relationship between age and serum IGF-I is expressed by the following equation: serum IGF-I (μg/L)= (-1.9347 3 x age) + 286.14; sd = 52.67 μg/L.

The age-specific sd score was calculated using the following formula: sd score = [serum IGF-I – (1.9347 3 age)/52.665]

Discussion

This systematic literature review assessed the effects of rhGH treatment in elderly GHD patients. The data indicate that rhGH treatment positively affects total and LDL cholesterol levels, and quality of life parameters. There is controversy about the influence of rhGH therapy on other cardiovascular risk factors, including insulin, HDL-cholesterol and triglyceride levels blood pressure or body composition. Some studies show improvement after treatment with rhGH, whereas others find no changes. Moreover, treatment with rhGH did not improve BMD in elderly subjects with GHD. Finally, studies on the effects of rhGH treatment in elderly GHD patients on clinically relevant endpoints, e.g. cardiovascular morbidity, fractures and mortality, have not been reported.

There is hardly any information on the treatment of very old GHD patients with rhGH. Although 2 studies included patients > 80 years, but data of these patients could not be extracted. Therefore, at present, there is no information with respect to the efficacy and safety of rhGH treatment in GHD octogenarians.

All patients described in the studies included in this systematic review were diagnosed with severe GHD based on different endocrine stimulation tests. However, because of the decline in GH secretion during ageing, aging may have affected the cut-off values of the GH stimulation tests. Studies using the insulin tolerance test (ITT) have been performed in these patients with various results. A study by Finucane *et al.* show that the ITT is a safe test even in elderly patients (30). However, other studies do show a lower GH response to ITT in the elderly (31). Nonetheless, the ITT is contra-indicated in patients with cardiac ischemia or arrhythmias. There are discrepancies between the studies on the cut-off values of the combined GHRH-arginine test and of the arginine test. In previous studies age seemed to be of no influence when using these tests (32;33). However, there is a significantly lower peak GH response in elderly compared to younger patients in the GHRH-arginine test (34). Therefore, the omission to reduce the cut-off values of GH stimulation tests in aging subjects may result in an erroneous diagnosis of GHD in some of these subjects. It is at present uncertain to which extent this may have affected the conclusions of the studies.

During ageing GH secretion decreases, associated with a decline of IGF-I levels. Therefore, age-adjusted IGF-I SD scores are necessary to be able to assess the treatment response to rhGH. All studies titrated the rhGH dose with the aim of normalizing IGF-I SD scores, *i.e.* aiming at IGF-I SDS in physiological levels for age and sex (between -2 and +2). However, from the analysis described in the results section, it becomes evident that in some studies SD scores were higher. In addition, some studies included the response of body composition to titrate the rhGh dose. Finally, some studies used IGF-I scores from reference populations with a different age distribution. Therefore, there are methological differences between the included studies that may have affected the relation between physiological rhGH replacement and responses in elderly subjects.

In GHD elderly subjects, treatment with rhGH had undisputed positive effects on total and LDL cholesterol levels, and on QoL (10;11;14;15;21). Treatment with rhGH decreased total and LDL-cholesterol levels by 4–8 % and 11–18 %, respectively. RhGH decreased W/H ratio in 3 of the 5 studies that report this parameter, but this was not confirmed in 2 other studies. RhGH increased lean body mass (LBM) by 2–5% and decreased fat mass by 7–10% (12;14;16) in 4 studies (n=192) (11;12;14;16), but this was not confirmed in 2 other studies (n=35) (23;24). One study documented that these positive effects of rhGH on body composition were reversed when rhGH therapy was subsequently stopped, even after only 3 months (n=12) (14). Therefore, there are undeniable effects of rhGH substitution in elderly subjects with GHD for some, but not all, parameters.

Several animal models of GH deficiency show prolonged, rather than decreased, longevity. Mice with mutations that cause GHD or GH resistance, live longer than their genetically normal siblings (9;35–38). In addition, adult body size, which can be considered a biological outcome marker of GH actions, was negatively correlated with longevity in other species, including rats (39), horses (40) and domestic dogs (41;42). Therefore, from an evolutionary perspective the natural decrease of GH and IGF-I levels during normal aging may even be beneficial. Epidemiological studies in humans, however, documented an association between both decreased and increased IGF-I levels and increased mortality, indicating that the optimal relation between IGF-I and rhGH dose may not be simple (9;43). Accordingly, it is presently not straightforward that all elderly subjects with GHD should be treated unconditionally.

In conclusion, only a small number of randomized placebo controlled trials have assessed the beneficial effects of rhGH therapy in the elderly. These studies show relatively limited effects. Therefore, the question remains whether the treatment with rhGH is clinically relevant in elderly, and especially very old, patients with GHD.

References

- 1. Corpas E, Harman SM & Blackman MR. Human growth hormone and human aging. Endocr Rev 1993 14 20–39
- Arvat E, Giordano R, Gianotti L, Broglio F, Camanni F & Ghigo E. Neuroendocrinology of the human growth hormone-insulin-like growth factor I axis during ageing. Growth Horm IGF Res 1999 9 Suppl A 111–115
- de Boer H, Blok GJ & Van dVAE. Clinical aspects of growth hormone deficiency in adults. Endocr Rev 1995 16 63–86
- 4. Liu H, Bravata DM, Olkin I, Nayak S, Roberts B, Garber AM *et al.* Systematic review: the safety and efficacy of growth hormone in the healthy elderly. Ann Intern Med 2007 146 104–115
- 5. Rudman D, Feller AG, Nagraj HS, Gergans GA, Lalitha PY, Goldberg AF *et al*. Effects of human growth hormone in men over 60 years old. N Engl J Med 1990 323 1–6
- Papadakis MA, Grady D, Black D, Tierney MJ, Gooding GA, Schambelan M *et al.* Growth hormone replacement in healthy older men improves body composition but not functional ability. Ann Intern Med 1996 124 708–716
- Johannsson G, Marin P, Lonn L, Ottosson M, Stenlof K, Bjorntorp P *et al.* Growth hormone treatment of abdominally obese men reduces abdominal fat mass, improves glucose and lipoprotein metabolism, and reduces diastolic blood pressure. J Clin Endocrinol Metab 1997 82 727–734
- 8. Franco C, Brandberg J, Lonn L, Andersson B, Bengtsson BA & Johannsson G. Growth hormone treatment reduces abdominal visceral fat in postmenopausal women with abdominal obesity: a 12-month placebo-controlled trial. J Clin Endocrinol Metab 2005 90 1466–1474
- van Bunderen CC, van N, I, van Schoor NM, Deeg DJ, Lips P & Drent ML. The association of serum insulin-like growth factor-I with mortality, cardiovascular disease, and cancer in the elderly: a population-based study. J Clin Endocrinol Metab 2010 95 4616–4624
- Koltowska-Haggstrom M, Mattsson AF & Shalet SM. Assessment of quality of life in adult patients with GH deficiency: KIMS contribution to clinical practice and pharmacoeconomic evaluations. Eur J Endocrinol 2009 161 Suppl 1 S51–S64
- 11. Feldt-Rasmussen U, Wilton P & Jonsson P. Aspects of growth hormone deficiency and replacement in elderly hypopituitary adults. Growth Horm IGF Res 2004 14 Suppl A S51–S58

- Gotherstrom G, Elbornsson M, Stibrant-Sunnerhagen K, Bengtsson BA, Johannsson G & Svensson J. Muscle strength in elderly adults with GH deficiency after 10 years of GH replacement. Eur J Endocrinol 2010 163 207–215
- Fernholm R, Bramnert M, Hagg E, Hilding A, Baylink DJ, Mohan S *et al.* Growth hormone replacement therapy improves body composition and increases bone metabolism in elderly patients with pituitary disease. J Clin Endocrinol Metab 2000 85 4104–4112
- 14. Gill MS, Toogood AA, Jones J, Clayton PE & Shalet SM. Serum leptin response to the acute and chronic administration of growth hormone (GH) to elderly subjects with GH deficiency. J Clin Endocrinol Metab 1999 84 1288–1295
- Monson JP, Abs R, Bengtsson BA, Bennmarker H, Feldt-Rasmussen U, Hernberg-Stahl E *et al.* Growth hormone deficiency and replacement in elderly hypopituitary adults. KIMS Study Group and the KIMS International Board. Pharmacia and Upjohn International Metabolic Database. Clin Endocrinol (Oxf) 2000 53 281–289
- 16. Elgzyri T, Castenfors J, Hagg E, Backman C, Thoren M & Bramnert M. The effects of GH replacement therapy on cardiac morphology and function, exercise capacity and serum lipids in elderly patients with GH deficiency. Clin Endocrinol (Oxf) 2004 61 113–122
- 17. Sathiavageeswaran M, Burman P, Lawrence D, Harris AG, Falleti MG, Maruff P *et al.* Effects of GH on cognitive function in elderly patients with adult-onset GH deficiency: a placebocontrolled 12-month study. Eur J Endocrinol 2007 156 439–447
- 18. Ho KK. Consensus guidelines for the diagnosis and treatment of adults with GH deficiency II: a statement of the GH Research Society in association with the European Society for Pediatric Endocrinology, Lawson Wilkins Society, European Society of Endocrinology, Japan Endocrine Society, and Endocrine Society of Australia. Eur J Endocrinol 2007 157 695–700
- 19. Gotherstrom G, Bengtsson BA, Sunnerhagen KS, Johannsson G & Svensson J. The effects of five-year growth hormone replacement therapy on muscle strength in elderly hypopituitary patients. Clin Endocrinol (Oxf) 2005 62 105–113
- 20. Toogood AA & Shalet SM. Growth hormone replacement therapy in the elderly with hypothalamic-pituitary disease: a dose-finding study. J Clin Endocrinol Metab 1999 84 131-136
- 21. Monson JP & Jonsson P. Aspects of growth hormone (GH) replacement in elderly patients with GH deficiency: data from KIMS. Horm Res 2003 60 112–120
- 22. White HD, Ahmad AM, Durham BH, Patwala A, Whittingham P, Fraser WD *et al.* Growth hormone replacement is important for the restoration of parathyroid hormone sensitivity and improvement in bone metabolism in older adult growth hormone-deficient patients. J Clin Endocrinol Metab 2005 90 3371–3380
- Franco C, Johannsson G, Bengtsson BA & Svensson J. Baseline characteristics and effects of growth hormone therapy over two years in younger and elderly adults with adult onset GH deficiency. J Clin Endocrinol Metab 2006 91 4408–4414

- 24. De Marinis L, Mancini A, Giampietro A, Gentilella R, Bianchi A, Perrelli M *et al.* GH deficiency syndrome in elderly patients. J Endocrinol Invest 2002 25 40–41
- 25. Landin-Wilhelmsen K, Wilhelmsen L, Lappas G, Rosen T, Lindstedt G, Lundberg PA *et al.* Serum insulin-like growth factor I in a random population sample of men and women: relation to age, sex, smoking habits, coffee consumption and physical activity, blood pressure and concentrations of plasma lipids, fibrinogen, parathyroid hormone and osteocalcin. Clin Endocrinol (Oxf) 1994 41 351–357
- 26. Drake WM, Coyte D, Camacho-Hubner C, Jivanji NM, Kaltsas G, Wood DF *et al.* Optimizing growth hormone replacement therapy by dose titration in hypopituitary adults. J Clin Endocrinol Metab 1998 83 3913–3919
- 27. Janssen YJ, Frolich M & Roelfsema F. A low starting dose of genotropin in growth hormonedeficient adults. J Clin Endocrinol Metab 1997 82 129–135
- 28. Hilding A, Hall K, Wivall-Helleryd IL, Saaf M, Melin AL & Thoren M. Serum levels of insulin-like growth factor I in 152 patients with growth hormone deficiency, aged 19–82 years, in relation to those in healthy subjects. J Clin Endocrinol Metab 1999 84 2013–2019
- 29. Bruce A, Andersson M, Arvidsson B & Isaksson B. Body composition. Prediction of normal body potassium, body water and body fat in adults on the basis of body height, body weight and age. Scand J Clin Lab Invest 1980 40 461–473
- 30. Finucane FM, Liew A, Thornton E, Rogers B, Tormey W & Agha A. Clinical insights into the safety and utility of the insulin tolerance test (ITT) in the assessment of the hypothalamopituitary-adrenal axis. Clin Endocrinol (Oxf) 2008 69 603–607
- Eskes SA, Tomasoa NB, Endert E, Geskus RB, Fliers E & Wiersinga WM. Establishment of reference values for endocrine tests. Part VII: growth hormone deficiency. Neth J Med 2009 67 127–133
- Toogood AA, Jones J, O'Neill PA, Thorner MO & Shalet SM. The diagnosis of severe growth hormone deficiency in elderly patients with hypothalamic-pituitary disease. Clin Endocrinol (Oxf) 1998 48 569–576
- 33. Ghigo E, Goffi S, Nicolosi M, Arvat E, Valente F, Mazza E *et al.* Growth hormone (GH) responsiveness to combined administration of arginine and GH-releasing hormone does not vary with age in man. J Clin Endocrinol Metab 1990 71 1481–1485
- Colao A, Di SC, Savastano S, Rota F, Savanelli MC, Aimaretti G *et al*. A reappraisal of diagnosing GH deficiency in adults: role of gender, age, waist circumference, and body mass index. J Clin Endocrinol Metab 2009 94 4414–4422
- 35. Bartke A & Brown-Borg H. Life extension in the dwarf mouse. Curr Top Dev Biol 2004 63 189–225
- Bartke A. Growth hormone and aging: a challenging controversy. Clin Interv Aging 2008 3 659–665

- 37. Bonkowski MS, Pamenter RW, Rocha JS, Masternak MM, Panici JA & Bartke A. Long-lived growth hormone receptor knockout mice show a delay in age-related changes of body composition and bone characteristics. J Gerontol A Biol Sci Med Sci 2006 61 562–567
- 38. Coschigano KT, Holland AN, Riders ME, List EO, Flyvbjerg A & Kopchick JJ. Deletion, but not antagonism, of the mouse growth hormone receptor results in severely decreased body weights, insulin, and insulin-like growth factor I levels and increased life span. Endocrinology 2003 144 3799–3810
- 39. Rollo CD. Growth negatively impacts the life span of mammals. Evol Dev 2002 4 55–61
- 40. Brosnahan MM & Paradis MR. Assessment of clinical characteristics, management practices, and activities of geriatric horses. J Am Vet Med Assoc 2003 223 99–103
- 41. Greer KA, Canterberry SC & Murphy KE. Statistical analysis regarding the effects of height and weight on life span of the domestic dog. Res Vet Sci 2007 82 208–214
- 42. Patronek GJ, Waters DJ & Glickman LT. Comparative longevity of pet dogs and humans: implications for gerontology research. J Gerontol A Biol Sci Med Sci 1997 52 B171–B178
- 43. Dekkers OM, Biermasz NR, Pereira AM, Romijn JA & Vandenbroucke JP. Mortality in acromegaly: a metaanalysis. J Clin Endocrinol Metab 2008 93 61–67

Chapter 8

General discussion and summary

Contents

I.	Introduction	199
II.	Evaluation of pituitary function in patients after Traumatic	
	Brain Injury (TBI)	200
III.	Dynamic tests of pituitary function in other pituitary	
	diseases	203
IV.	Treatment of GHD	208
V.	Concluding remarks	210

I. Introduction

Many tests are available for the assessment of pituitary function. The thyroid, gonadotropic and prolactine axes can be appropriately assessed by the combination of clinical symptoms and unstimulated serum hormone levels. However, stimulation tests are mandatory for appropriate evaluation for the assessment of the HPA and GH-IGF-I axes (1).

Different stimulation tests are available, for which different cut-off values have been reported. When using a stimulation test, it is of great importance to take other confounding factors into account such as age, gender, BMI and medication. GH secretion decreases resulting in lower serum levels with increasing age and BMI (2–4). Medication can alter hormone levels and therefore influence test outcome. The pituitary gland does not necessarily be stable, but can change over time, e.g. after traumatic brain injury (TBI) or pituitary surgery (5;6). Consequently, pituitary stimulation tests performed immediately after surgery may give altered results. In this thesis several studies are reported aiming to provide better insight into the complexity of different endocrine tests used for the evaluation of possible pituitary insufficiency and in the treatment of patients with pituitary insufficiency.

II. Evaluation of pituitary function in patients after Traumatic Brain Injury (TBI)

After TBI patients experience persistent, invalidating complaints that resemble those observed in patients with hypopituitarism, such as impaired cognition, depression, fatigue and impaired quality of life (QoL) (7–9). Consequently, pituitary insufficiency following TBI may contribute to the problems reported by these patients. This condition is important to identify since it can be treated by hormone replacement therapy resulting in improved QoL (10).

In the past decade a high prevalence of pituitary insufficiency following TBI has been reported (11-24). However, there is great variation in the reported prevalence rates. Several factors influence the prevalence of hypopituitarism after TBI: the time interval between TBI and endocrine assessment, the type and severity of the trauma and also the methods (*i.e.* endocrine tests, assays and criteria) used for the diagnosis of hypopituitarism. Some reviews have addressed TBI-related hypopituitarism and concluded that hypopituitarism is a common complication of TBI and might contribute to morbidity and poor recovery after brain injury (25). However, these reviews did not take into account the variability in diagnostic strategies and definitions of pituitary insufficiency. We hypothesized that methodological differences may have contributed, at least in part, to the discrepancies in prevalence rates of hypopituitarism after TBI. Therefore, the aim of our systematic review in **chapter 2** was to critically compare the pituitary function tests, and definitions of hypopituitarism between studies that assessed the long-term outcome of TBI on pituitary function for each pituitary axis.

We found that the reported prevalence rates of pituitary insufficiency indeed vary considerably and that this is associated with major differences in endocrine and analytic methods of assessment en definitions used for the diagnosis hypopituitarism. The studies in the review used different endocrine tests, cut-off values and analytic methods. Moreover, several confounding factors (such as BMI) were not taken into account when assessing the pituitary axes. This is especially of importance for the assessment of the GH axis given the decrease of GH concentration with increasing BMI. This all may result in an overestimation of hypopituitarism following TBI in obese subjects. These discrepancies limit the possibility to compare the results of studies on TBI. Future studies should be designed to ensure a high diagnostic robustness for proper identification of reliable predictors, as the results may be highly dependent on diagnostic pitfalls (26).

Because of the large variations published on prevalence rates reported and the variations in endocrine and analytical methods to assess pituitary functions, we performed a cross-sectional study in the Netherlands in a large cohort of TBI patients evaluated after long-term follow-up (described in **chapter 3**). We included 112 patients with TBI, hospitalized for at least 3 days and duration of follow-up > 1 yr after TBI from 5 (neurosurgical) referral centers. Evaluation of pituitary function included fasting morning hormone measurements and insulin tolerance test (ITT n=90) or, when contraindicated, ACTH-stimulation and/or CRH-stimulation test and a GHRH-arginine test (n=22).

Our study demonstrates that prevalence of hypopituitarism after TBI after long-term follow-up is low. Using a standardized evaluation that included the golden standard tests for the evaluation of GH and cortisol secretory reserves in the majority of the patients, we found a prevalence of any pituitary insufficiency of only 5.4% (severe growth hormone deficiency (2.8%), hypogonadism (0.9%), adrenal insufficiency (1.8%)). This prevalence is much lower compared to the prevalence rates reported in the majority of the previous studies (15–90%)(5–17). This discrepancy might be explained by the use of different endocrine tests and different cut-off values in the previous studies. If possible we used the golden standard test: the insulin tolerance test. In accordance with the data by Klose *et al.* (21), this resulted in lower prevalence rates of GHD and adrenal insufficiency. If ITT was contraindicated, we used

the combined stimulation with GHRH and arginine to assess the GH axis which has been shown to be a good alternative test, provided higher cut-off GH levels are used. As mentioned above, GH secretion decreases with increasing BMI (2-4). We, therefore, used BMI-adjusted cutoff values for the combined stimulation with GHRH and arginine. In addition to differences in endocrine test, the time interval between TBI and endocrine evaluation as well as trauma severity may also affect the reported prevalence rates. Studies have reported that in the acute phase after TBI hormone alterations mimicking pituitary insufficiency can be present (21;27). To avoid this transient effect of TBI, we evaluated patients at least 1 year post TBI. Increased trauma severity increases the risk of pituitary insufficiency. Therefore, we included only patients with more severe trauma. Patients had to be hospitalized for at least 3 days and GCS was evaluated. In contrast to many previous studies, the prevalence of pituitary insufficiency appeared to be low in patients with more severe trauma at least one year after trauma, using these inclusion criteria and golden standard test for pituitary assessment whenever possible and BMI-adjusted cut-off values if necessary.

Our results indicate that consensus for a more uniform endocrine evaluation of pituitary function in general and after TBI in particular is needed. Nonetheless, pituitary failure, even if present after TBI in a very small proportion of patients, is potentially treatable, may be life-saving, and is likely to ameliorate quality of life (7;10).

III. Dynamic tests of pituitary function in other pituitary diseases

Pituitary adenomas can be treated by transsphenoidal surgery (TS), additional radiotherapy and/or medication. Pituitary insufficiency is a complication that can be attributed to the tumor itself (compression), surgery and/or radiotherapy. Therefore, accurate assessment of pituitary function is critical for appropriate management of patients with pituitary adenoma after surgery with or without irradiation.

Endocrine assessment after pituitary surgery

After TS the assessment of the hypothalamic-pituitary adrenal (HPA) axis is of clinical relevance to judge the need for hydrocortisone replacement therapy at discharge. The ITT is the golden standard to evaluate the HPA axis in patients suspected of secondary adrenal insufficiency. Because of contraindications for the induction of hypoglycemia different other dynamic tests of the HPA axis are available such as the metyrapone test, the ACTH stimulation test and the corticotrophin releasing hormone (CRH)-test. In our clinic (from 1990 onwards) adrenal function of patients directly after surgery has been evaluated by stimulation with CRH. Based on the test result it is decided whether the patients were discharged with our without hydrocortisone replacement therapy. Specific data on the clinical applicability of the CRH test directly after TS are hardly available. Therefore, in **chapter 4** we retrospectively evaluated the clinical relevanceof the CRH stimulation test in assessing pituitary adrenal function after TS. We performed a retrospective chart review of all patients who had been treated by TS in our center. We included a total of 144 non-Cushing patients of whom data were available on postsurgical CRH tests, of whom second (confirmation) tests were also available and who had not been subjected to confounding factors like use of exogenous glucocorticoids, re-operation or postsurgical radiotherapy. Forty-two patients were diagnosed with hypocortisolism of whom 13 (31%) had sufficient adrenal function during follow-up.

A possible explanation for these discrepant results is the use of different cut-off values. For the ITT (golden standard) regularly accepted cut-off values have been defined. However, for the CRH test different cut-off values for peak cortisol responses have been proposed. Because in our center the CRH test is used as a screening test to identify those patients that require hydrocortisone supplementation after TS, we applied a generally accepted stringent criterion of 550 nmol/L (28;29). Aiming for a higher sensitivity will be at the expense of a lower specificity, i.e a greater proportion of patients will be incorrectly diagnosed with adrenal insufficiency.

Another possible explanation is the recovery of preoperative adrenal insufficiency after TS within one year (5;6). This has been described in a study that compared the ITT response at 3 and 12 months after TS. In agreement, we found a normal adrenal function in 8 patients within the first year after surgery who were initially diagnosed as being adrenal insufficient. This indicates the necessity of an extensive follow-up in patiens after surgery within one year.

A normal function of the HPA axis was assessed in 102 of the 144 patients. However, fourteen of these patients (14%) appeared to have hypocortisolism based on a second test. These discrepant results can be potentially life-threatening because these patients are at risk for adrenal crises. It is possible that additional pituitary insufficiency influenced the test results of these patients. Growth hormone and thyroid hormone deficiency can influence the test results(5;30–32). Moreover, growth hormone replacement therapy in patients with GHD may also play an important role because of the influence of GH on the cortisol metabolism. Growth hormone stimulates $11-\beta$ hydroxysteroid dehydrogenase (11 β HSD-1), leading to increased cortisol-cortisone conversion (31). The use of GH replacement therapy in GH-deficient patients may therefore unmask cortisol deficiency (30;32).

Based on our results we conclude that the CRH test can be safely used to guide hydrocortisone substitution after TS. Nonetheless, the cortisol response to this test can not reliably predict adrenal function in all patients during longer follow-up after TS. We therefore recommend to perform a second test of pituitary adrenal function during longer follow-up, e.g. 3–6 months after surgery. This approach is not required in patients with an impaired postoperative cortisol response to CRH, who have multiple pituitary insufficiencies.

In chapter 4 we retrospectively assessed the HPA function in all patients who had been treated by TS in our center, whereas in chapter 5 we focused on postoperative assessment of HPA function in a specific postoperative group; patients after TS for GH secreting adenomas *i.e.* patients with acromegaly. A recent study by Ronchi and colleagues evaluated the HPA axis in acromegalic patients after TS. They found a remarkably high prevalence of adrenal insufficiency (32%) after TS in these patients. They concluded that the function of the HPA axis may worsen over time and should be carefully monitored by dynamic testing in all acromegalic patients, independently from the type of treatment. This recommendation has obvious implications for the longterm management of non-irradiated patients with acromegaly (33). Therefore, the aim of **chapter 5** was to evaluate the prevalence of adrenal insufficiency during long-term follow-up in our own unselected cohort of consecutive patients in remission of GH excess after transsphenoidal surgery.

We retrospectively reviewed the assessment of corticotrope function in 91 consecutive patients in remission after transsphenoidal surgery using ITT, CRH stimulation, metyrapone test and ACTH stimulation tests. We found insufficient adrenal function in 16 patients (18%) in the early postoperative period, which was transient in 8 but irreversible in 8 other patients within the first year of postoperative follow-up. Therefore, after the first year of follow-up after curative surgery for acromegaly, the prevalence of adrenal insufficiency was only 9%. Late, new-onset adrenal insufficiency developed in only 3 patients, 13, 18 and 24 years after surgery, resp. The incidence rate of late adrenal insufficiency after successful surgery was only 2/1000 person years. After long-term followup, with a median duration of 8.1 yr (range 1–31 yr), the prevalence of secondary adrenal insufficiency was 12% in patients in remission after surgery for acromegaly. Therefore, new-onset adrenal insufficiency after TS for acromegaly is not frequently present. The discrepancies in prevalence with the study by Ronchi *et al.* and our study may be explained by differences in study design and study population but also by patient selection and differences in surgical techniques. We used the golden standard test (ITT) and CRH test in a large whereas Ronchi and colleagues used a low-dose ACTH test in patients (33). Other potential mechanisms of influence may be changed cortisol binding globulin levels (CBG) in acromegaly (31;32;34;35), the presence of postoperative GH deficiency (30;35;36) and the possibility of recovery of preoperative adrenal insufficiency following transsphenoidal surgery (5;6;37), although this is most likely a rare event.

Limitations of our study are the retrospective nature of the study and the fact that patients had been tested by different cortisol stimulation tests and assays. However, this does not affect our conclusions, since the ITT, CRH and metyrapone tests are all accepted tests for the evaluation of HPA function and we have used unchanged cut-off values of cortisol throughout the years.

We propose to repeat dynamic test of HPA function 1 yr post surgery in patients with postoperative HPA insufficiency. Further research is required to assess whether yearly basal cortisol values may suffice to monitor adrenal function in asymptomatic patients. However, in case of low basal cortisol levels, symptoms suggestive of corticotrope insufficiency or progressive impairment of other pituitary functions, additional dynamic testing of the HPA axis should be performed.

Endocrine assessment following cranial radiotherapy

Patients with nonpituitary intracranial and/or nasopharyngeal tumors are frequently treated by radiotherapy, in which the pituitary gland is involved in the radiation field. These patients are at risk for pituitary insufficiency. This is well described in children treated with cranial radiotherapy (38–46), but the assessment of pituitary function during long term follow-up has not been implemented in the guidelines of patients treated by cranial radiotherapy for nonpituitary tumors. To assess the prevalence of pituitary insufficiencies after cranial radiotherapy in these patients, we performed a systemic literature search and meta-analysis focusing on the prevalence of pituitary dysfunction in adult patients treated with radiotherapy for nonpituitary tumors, which is described in **chapter 6**.

Our review ultimately included 18 studies (n=813) evaluating patients treated for nasopharyngeal cancer or intracerebral tumors (47-64). There were considerable variations in the reported prevalence rates of hypopituitarism after cranial radiotherapy, ranging from hardly any effect on pituitary function to almost 100% of the patients being affected. These variations may be associated with differences in radiotherapeutic techniques, study design, time of evaluation, patient selection and differences in endocrine evaluation. The majority of patients was not evaluated by pituirary stimulation tests. If stimulation tests had been used, different cut-off values and diagnostic criteria were used. Our meta-analysis showed that any hypopituitarism is present in approximately two thirds of all adult patients previously treated by cranial radiotherapy (0.66, CI 0.55-0.76). The prevalence of growth hormone deficiency was 0.45 (CI 0.33-0.57), of LH and FSH 0.3 (CI 0.23-0.37), of TSH 0.25 (CI 0.16-0.37), and of ACTH 0.22 (CI 0.15-0.3), respectively. The prevalence of hyperprolactinemia was 0.34 (CI 0.15–0.6) There were no differences between the effects of radiotherapy for nasopharyngeal versus for intracerebral tumors.

Based on these data we conclude that hypopituitarism is rather prevalent in adult patients after cranial radiotherapy for nonpituitary tumors. Considering this high prevalence of hypopituitarism, the evaluation of pituitary function should be included in the guidelines of long-term follow-up of all patients treated by cranial radiotherapy.

IV. Treatment of GHD

When growth hormone deficiency is considered, the therapeutical implications should be carefully evaluated, especially in elderly subjects in whom normal growth hormone secretion and IGF-I levels are low compared to young adults and GH and IGF-I levels overlap between normal and growth hormone deficient subjects.

In **chapter 7**, we performed a systematic review, to critically assess the available literature on the evidence of clinical efficacy of rhGH in elderly patients with GHD. We ultimately included only 11 eligible studies with a total of 534 patients (65–78). The studies show that there are undeniable effects of rhGH substitution in elderly subjects with GHD for some, but not all, parameters. RhGH treatment unequivocally positively affects total and LDL cholesterol levels and QoL parameters. However, there is controversy on the effects on other cardiovascular risk factors, including insulin, HDL cholesterol, BP and BC, whereas rhGH therapy does not improve plasma triglyceride levels. Moreover, treatment with rhGH did not improve BMD in elderly subjects with GHD. Studies in octogenarians have not been performed. Finally, there are no data on the effects of rhGH on clinically relevant end points, like cardiovascular disease or fractures.

Several factors should be taken into account in the assessment of the effects of rhGH therapy in elderly subjects. With increasing age GH secretion decreases. This decrease in GH levels may affect the response to stimulation tests and, therefore, affect the cut-off values of the GH stimulation tests. Studies using the ITT and GHRH-arginine stimulation test have been performed in these patients with various results. Some studies show a lower peak of GH response in elderly compared to younger patients (79;80), whereas other studies show no differences (81). Nonetheless, the omission to reduce the cut-off values of GH stimulation tests in aging subjects might result in an erroneous diagnosis of GHD in at least some of these subjects. The extent to which this may have affected the conclusions is uncertain at present. The decline of GH levels during aging is associated with a decline in IGF-I levels. Therefore, age-adjusted IGF-I SD scores are necessary to enable to assess the treatment response to rhGH. In this respect, there were methodological differences between the included studies, which may have affected the relation between physiological rhGH replacement and responses in elderly subjects. Moreover, from an evolutionary perspective, the natural decrease of GH and IGF-I levels during normal aging may even be beneficial. In animal models of decreased GH-IGF-I function longevity was increased (82–89). Accordingly, it is presently not straightforward that all elderly subjects with GHD should be treated unconditionally.

In conclusion, only a small number of randomized placebo-controlled trials have assessed the beneficial effects of rhGH therapy in elderly with GHD. These studies show relatively limited effects. Therefore, the question remains whether the treatment with rhGH is clinically relevant in elderly patients with GHD. There are no data whatsoever on the effects of rhGH in octogenarians with GHD.

V. Concluding remarks

The conclusions of the studies described in this thesis can be summarized as follows:

Pituitary function after TBI

There is a wide variation in the reported prevalence rates of hypopituitarism after TBI. This is at least in part caused by differences in definitions, endocrine assessments of hypopituitarism, and confounding factors. These methodological issues prohibit simple generalizations of results of original studies on TBI-associated hypopituitarism in the perspective of meta-analyses or reviews.

The prevalence of hypopituitarism during long-term follow-up after TBI is most likely very low, if stringent criteria and appropriate pituitary tests are used. The reported prevalence rates of pituitary insufficiency after TBI are most likely overestimated.

Pituitary function after transsphenoidal surgery

The CRH test is a valuable tool to define clinically relevant cortisol deficiency immediately after pituitary surgery. This test can be safely used to define hydrocortisone dependency at discharge until a second test is performed.

In patients with acromegaly cured by transsphenoidal surgery, the prevalence of adrenal insufficiency very low: 9% one year after surgery and only 2/1000 person-years in patients in long term remission after surgery. Therefore, development of late-onset adrenal insufficiency is a

very infrequent complication in patients with acromegaly in remission after transsphenoidal surgery only.

Pituitary function after cranial radiotherapy

Hypopituitarism is very prevalent in adult patients after cranial radiotherapy for nonpituitary tumors. Therefore, all patients treated by cranial radiotherapy should have structured periodical assessment of pituitary function during follow-up. This should be implemented in the guidelines of follow-up of these patients.

Pituitary function in elderly subjects

Recombinant GH replacement in elderly subjects with GHD decreases LDL cholesterol levels and improves QoL, but the effects on other parameters are not unequivocal. There are no data on the efficacy and safety of rhGH treatment in octogenarians with GHD. There are no data on clinically relevant endpoints like cardiovascular disease or fractures.

References

- van Aken MO, Lamberts SW 2005 Diagnosis and treatment of hypopituitarism: an update. Pituitary 8:183–191
- 2. Iranmanesh A, Lizarralde G, Veldhuis JD 1991 Age and relative adiposity are specific negative determinants of the frequency and amplitude of growth hormone (GH) secretory bursts and the half-life of endogenous GH in healthy men. J Clin Endocrinol Metab 73:1081–1088
- 3. Veldhuis JD, Liem AY, South S, Weltman A, Weltman J, Clemmons DA, Abbott R, Mulligan T, Johnson ML, Pincus S, . 1995 Differential impact of age, sex steroid hormones, and obesity on basal versus pulsatile growth hormone secretion in men as assessed in an ultrasensitive chemiluminescence assay. J Clin Endocrinol Metab 80:3209–3222
- Veldhuis JD, Roelfsema F, Keenan DM, Pincus S 2011 Gender, age, body mass index, and IGF-I individually and jointly determine distinct GH dynamics: analyses in one hundred healthy adults. J Clin Endocrinol Metab 96:115–121
- 5. Arafah BM 1986 Reversible hypopituitarism in patients with large nonfunctioning pituitary adenomas. J Clin Endocrinol Metab 62:1173–1179
- 6. Webb SM, Rigla M, Wagner A, Oliver B, Bartumeus F 1999 Recovery of hypopituitarism after neurosurgical treatment of pituitary adenomas. J Clin Endocrinol Metab 84:3696–3700
- Blair JC 2010 Prevalence, natural history and consequences of posttraumatic hypopituitarism: a case for endocrine surveillance. Br J Neurosurg 24:10–17
- Fleminger S, Ponsford J 2005 Long term outcome after traumatic brain injury. BMJ 331:1419–1420
- van Aken MO, Lamberts SW 2005 Diagnosis and treatment of hypopituitarism: an update. Pituitary 8:183–191
- Maric NP, Doknic M, Pavlovic D, Pekic S, Stojanovic M, Jasovic-Gasic M, Popovic V 2010 Psychiatric and neuropsychological changes in growth hormone-deficient patients after traumatic brain injury in response to growth hormone therapy. J Endocrinol Invest 33:770–775
- Lieberman SA, Oberoi AL, Gilkison CR, Masel BE, Urban RJ 2001 Prevalence of neuroendocrine dysfunction in patients recovering from traumatic brain injury. J Clin Endocrinol Metab 86:2752–2756
- Kelly DF, Gonzalo IT, Cohan P, Berman N, Swerdloff R, Wang C 2000 Hypopituitarism following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a preliminary report. J Neurosurg 93:743–752

- Bondanelli M, De ML, Ambrosio MR, Monesi M, Valle D, Zatelli MC, Fusco A, Bianchi A, Farneti M, gli Uberti EC 2004 Occurrence of pituitary dysfunction following traumatic brain injury. J Neurotrauma 21:685–696
- 14. Agha A, Rogers B, Sherlock M, O'Kelly P, Tormey W, Phillips J, Thompson CJ 2004 Anterior pituitary dysfunction in survivors of traumatic brain injury. J Clin Endocrinol Metab 89:4929–4936
- 15. Aimaretti G, Ambrosio MR, Di SC, Gasperi M, Cannavo S, Scaroni C, Fusco A, Del MP, De ME, Faustini-Fustini M, Grimaldi F, Logoluso F, Razzore P, Rovere S, Benvenga S, gli Uberti EC, De ML, Lombardi G, Mantero F, Martino E, Giordano G, Ghigo E 2005 Residual pituitary function after brain injury-induced hypopituitarism: a prospective 12–month study. J Clin Endocrinol Metab 90:6085–6092
- 16. Popovic V, Pekic S, Pavlovic D, Maric N, Jasovic-Gasic M, Djurovic B, Medic SM, Zivkovic V, Stojanovic M, Doknic M, Milic N, Djurovic M, Dieguez C, Casanueva FF 2004 Hypopituitarism as a consequence of traumatic brain injury (TBI) and its possible relation with cognitive disabilities and mental distress. J Endocrinol Invest 27:1048–1054
- Leal-Cerro A, Flores JM, Rincon M, Murillo F, Pujol M, Garcia-Pesquera F, Dieguez C, Casanueva FF 2005 Prevalence of hypopituitarism and growth hormone deficiency in adults long-term after severe traumatic brain injury. Clin Endocrinol (Oxf) 62:525–532
- Schneider HJ, Schneider M, Saller B, Petersenn S, Uhr M, Husemann B, von RF, Stalla GK 2006 Prevalence of anterior pituitary insufficiency 3 and 12 months after traumatic brain injury. Eur J Endocrinol 154:259–265
- 19. Tanriverdi F, Senyurek H, Unluhizarci K, Selcuklu A, Casanueva FF, Kelestimur F 2006 High risk of hypopituitarism after traumatic brain injury: a prospective investigation of anterior pituitary function in the acute phase and 12 months after trauma. J Clin Endocrinol Metab 91:2105–2111
- Herrmann BL, Rehder J, Kahlke S, Wiedemayer H, Doerfler A, Ischebeck W, Laumer R, Forsting M, Stolke D, Mann K 2006 Hypopituitarism following severe traumatic brain injury. Exp Clin Endocrinol Diabetes 114:316–321
- 21. Klose M, Juul A, Poulsgaard L, Kosteljanetz M, Brennum J, Feldt-Rasmussen U 2007 Prevalence and predictive factors of post-traumatic hypopituitarism. Clin Endocrinol (Oxf) 67:193–201
- 22. Bushnik T, Englander J, Katznelson L 2007 Fatigue after TBI: association with neuroendocrine abnormalities. Brain Inj 21:559–566
- 23. Wachter D, Gundling K, Oertel MF, Stracke H, Boker DK 2009 Pituitary insufficiency after traumatic brain injury. J Clin Neurosci 16:202–208
- 24. Tanriverdi F, Ulutabanca H, Unluhizarci K, Selcuklu A, Casanueva FF, Kelestimur F 2008 Three years prospective investigation of anterior pituitary function after traumatic brain injury: a pilot study. Clin Endocrinol (Oxf) 68:573–579

- 25. Schneider HJ, Kreitschmann-Andermahr I, Ghigo E, Stalla GK, Agha A 2007 Hypothalamopituitary dysfunction following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a systematic review. JAMA 298:1429–1438
- 26. Klose M, Feldt-Rasmussen U 2008 Does the type and severity of brain injury predict hypothalamo-pituitary dysfunction? Does post-traumatic hypopituitarism predict worse outcome? Pituitary 11:255–261
- 27. Klose M, Juul A, Struck J, Morgenthaler NG, Kosteljanetz M, Feldt-Rasmussen U 2007 Acute and long-term pituitary insufficiency in traumatic brain injury: a prospective single-centre study. Clin Endocrinol (Oxf) 67:598–606
- 28. Trainer PJ, Faria M, Newell-Price J, Browne P, Kopelman P, Coy DH, Besser GM, Grossman AB 1995 A comparison of the effects of human and ovine corticotropin-releasing hormone on the pituitary-adrenal axis. J Clin Endocrinol Metab 80:412–417
- Lytras N, Grossman A, Perry L, Tomlin S, Wass JA, Coy DH, Schally AV, Rees LH, Besser GM 1984 Corticotrophin releasing factor: responses in normal subjects and patients with disorders of the hypothalamus and pituitary. Clin Endocrinol (Oxf) 20:71–84
- 30. Giavoli C, Libe R, Corbetta S, Ferrante E, Lania A, Arosio M, Spada A, Beck-Peccoz P 2004 Effect of recombinant human growth hormone (GH) replacement on the hypothalamic-pituitaryadrenal axis in adult GH-deficient patients. J Clin Endocrinol Metab 89:5397–5401
- 31. Moore JS, Monson JP, Kaltsas G, Putignano P, Wood PJ, Sheppard MC, Besser GM, Taylor NF, Stewart PM 1999 Modulation of 11beta-hydroxysteroid dehydrogenase isozymes by growth hormone and insulin-like growth factor: in vivo and in vitro studies. J Clin Endocrinol Metab 84:4172–4177
- Rodriguez-Arnao J, Perry L, Besser GM, Ross RJ 1996 Growth hormone treatment in hypopituitary GH deficient adults reduces circulating cortisol levels during hydrocortisone replacement therapy. Clin Endocrinol (Oxf) 45:33–37
- Ronchi CL, Ferrante E, Rizzo E, Giavoli C, Verrua E, Bergamaschi S, Lania AG, Beck-Peccoz P, Spada A 2008 Long-term basal and dynamic evaluation of hypothalamic-pituitary-adrenal (HPA) axis in acromegalic patients. Clin Endocrinol (Oxf) 69:608–612
- 34. Weaver JU, Thaventhiran L, Noonan K, Burrin JM, Taylor NF, Norman MR, Monson JP 1994 The effect of growth hormone replacement on cortisol metabolism and glucocorticoid sensitivity in hypopituitary adults. Clin Endocrinol (Oxf) 41:639–648
- 35. Tschop M, Lahner H, Feldmeier H, Grasberger H, Morrison KM, Janssen OE, Attanasio AF, Strasburger CJ 2000 Effects of growth hormone replacement therapy on levels of cortisol and cortisol-binding globulin in hypopituitary adults. Eur J Endocrinol 143:769–773
- Isidori AM, Kaltsas GA, Perry L, Burrin JM, Besser GM, Monson JP 2003 The effect of growth hormone replacement therapy on adrenal androgen secretion in adult onset hypopituitarism. Clin Endocrinol (Oxf) 58:601–611

- 37. Berg C, Meinel T, Lahner H, Mann K, Petersenn S 2010 Recovery of pituitary function in the late-postoperative phase after pituitary surgery: results of dynamic testing in patients with pituitary disease by insulin tolerance test 3 and 12 months after surgery. Eur J Endocrinol 162:853–859
- Alvarez JA, Scully RE, Miller TL, Armstrong FD, Constine LS, Friedman DL, Lipshultz SE 2007 Long-term effects of treatments for childhood cancers. Curr Opin Pediatr 19:23–31
- 39. Constine LS, Woolf PD, Cann D, Mick G, McCormick K, Raubertas RF, Rubin P 1993 Hypothalamicpituitary dysfunction after radiation for brain tumors. N Engl J Med 328:87–94
- 40. Oeffinger KC, Mertens AC, Sklar CA, Kawashima T, Hudson MM, Meadows AT, Friedman DL, Marina N, Hobbie W, Kadan-Lottick NS, Schwartz CL, Leisenring W, Robison LL 2006 Chronic health conditions in adult survivors of childhood cancer. N Engl J Med 355:1572–1582
- Schmiegelow M, Lassen S, Poulsen HS, Feldt-Rasmussen U, Schmiegelow K, Hertz H, Muller J 2000 Cranial radiotherapy of childhood brain tumours: growth hormone deficiency and its relation to the biological effective dose of irradiation in a large population based study. Clin Endocrinol (Oxf) 53:191–197
- 42. Shalet SM, Beardwell CG, Jones PH, Pearson D 1976 Growth hormone deficiency after treatment of acute leukaemia in children. Arch Dis Child 51:489–493
- 43. Schmiegelow M, Feldt-Rasmussen U, Rasmussen AK, Poulsen HS, Muller J 2003 A populationbased study of thyroid function after radiotherapy and chemotherapy for a childhood brain tumor. J Clin Endocrinol Metab 88:136–140
- 44. Shalet SM, Beardwell CG, Morris-Jones P, Bamford FN, Ribeiro GG, Pearson D 1976 Growth hormone deficiency in children with brain tumors. Cancer 37:1144–1148
- 45. Shalet SM, Beardwell CG, MacFarlane IA, Jones PH, Pearson D 1977 Endocrine morbidity in adults treated with cerebral irradiation for brain tumours during childhood. Acta Endocrinol (Copenh) 84:673–680
- 46. Gurney JG, Kadan-Lottick NS, Packer RJ, Neglia JP, Sklar CA, Punyko JA, Stovall M, Yasui Y, Nicholson HS, Wolden S, McNeil DE, Mertens AC, Robison LL 2003 Endocrine and cardiovascular late effects among adult survivors of childhood brain tumors: Childhood Cancer Survivor Study. Cancer 97:663–673
- 47. Snyers A, Janssens GO, Twickler MB, Hermus AR, Takes RP, Kappelle AC, Merkx MA, Dirix P, Kaanders JH 2009 Malignant tumors of the nasal cavity and paranasal sinuses: long-term outcome and morbidity with emphasis on hypothalamic-pituitary deficiency. Int J Radiat Oncol Biol Phys 73:1343–1351
- Bhandare N, Kennedy L, Malyapa RS, Morris CG, Mendenhall WM 2008 Hypopituitarism after radiotherapy for extracranial head and neck cancers. Head Neck 30:1182–1192
- Lam KS, Tse VK, Wang C, Yeung RT, Ho JH 1991 Effects of cranial irradiation on hypothalamicpituitary function--a 5-year longitudinal study in patients with nasopharyngeal carcinoma. Q J Med 78:165–176

- 50. Woo E, Lam K, Yu YL, Ma J, Wang C, Yeung RT 1988 Temporal lobe and hypothalamic-pituitary dysfunctions after radiotherapy for nasopharyngeal carcinoma: a distinct clinical syndrome. J Neurol Neurosurg Psychiatry 51:1302–1307
- Samaan NA, Schultz PN, Yang KP, Vassilopoulou-Sellin R, Maor MH, Cangir A, Goepfert H 1987 Endocrine complications after radiotherapy for tumors of the head and neck. J Lab Clin Med 109:364–372
- 52. Lam KS, Ho JH, Lee AW, Tse VK, Chan PK, Wang C, Ma JT, Yeung RT 1987 Symptomatic hypothalamic-pituitary dysfunction in nasopharyngeal carcinoma patients following radiation therapy: a retrospective study. Int J Radiat Oncol Biol Phys 13:1343–1350
- 53. Lam KS, Wang C, Yeung RT, Ma JT, Ho JH, Tse VK, Ling N 1986 Hypothalamic hypopituitarism following cranial irradiation for nasopharyngeal carcinoma. Clin Endocrinol (Oxf) 24:643–651
- 54. Huang KE 1979 Assessment of hypothalamic-pituitary function in women after external head irradiation. J Clin Endocrinol Metab 49:623–627
- 55. Rosenthal MB, Goldfine ID 1976 Primary and secondary hypothyroidism in nasopharyngeal carcinoma. JAMA 236:1591–1593
- Samaan NA, Bakdash MM, Caderao JB, Cangir A, Jesse RH, Jr., Ballantyne AJ 1975 Hypopituitarism after external irradiation. Evidence for both hypothalamic and pituitary origin. Ann Intern Med 83:771–777
- 57. Schneider HJ, Rovere S, Corneli G, Croce CG, Gasco V, Ruda R, Grottoli S, Stalla GK, Soffietti R, Ghigo E, Aimaretti G 2006 Endocrine dysfunction in patients operated on for nonpituitary intracranial tumors. Eur J Endocrinol 155:559–566
- Agha A, Sherlock M, Brennan S, O'Connor SA, O'Sullivan E, Rogers B, Faul C, Rawluk D, Tormey W, Thompson CJ 2005 Hypothalamic-pituitary dysfunction after irradiation of nonpituitary brain tumors in adults. J Clin Endocrinol Metab 90:6355–6360
- 59. Johannesen TB, Lien HH, Hole KH, Lote K 2003 Radiological and clinical assessment of longterm brain tumour survivors after radiotherapy. Radiother Oncol 69:169–176
- 60. Popovic V, Pekic S, Golubicic I, Doknic M, Dieguez C, Casanueva FF 2002 The impact of cranial irradiation on GH responsiveness to GHRH plus GH-releasing peptide-6. J Clin Endocrinol Metab 87:2095–2099
- 61. Taphoorn MJ, Heimans JJ, van d, V, Karim AB 1995 Endocrine functions in long-term survivors of low-grade supratentorial glioma treated with radiation therapy. J Neurooncol 25:97–102
- 62. Constine LS, Rubin P, Woolf PD, Doane K, Lush CM 1987 Hyperprolactinemia and hypothyroidism following cytotoxic therapy for central nervous system malignancies. J Clin Oncol 5:1841–1851
- 63. Mechanick JI, Hochberg FH, LaRocque A 1986 Hypothalamic dysfunction following wholebrain irradiation. J Neurosurg 65:490–494
- 64. Harrop JS, Davies TJ, Capra LG, Marks V 1976 Hypothalamic-pituitary function following successful treatment of intracranial tumours. Clin Endocrinol (Oxf) 5:313–321

- 65. De ML, Mancini A, Giampietro A, Gentilella R, Bianchi A, Perrelli M, Vezzosi C, Milardi D, Fusco A, Valle D, Bernabei R 2002 GH deficiency syndrome in elderly patients. J Endocrinol Invest 25:40–41
- 66. Elgzyri T, Castenfors J, Hagg E, Backman C, Thoren M, Bramnert M 2004 The effects of GH replacement therapy on cardiac morphology and function, exercise capacity and serum lipids in elderly patients with GH deficiency. Clin Endocrinol (Oxf) 61:113–122
- 67. Feldt-Rasmussen U, Wilton P, Jonsson P 2004 Aspects of growth hormone deficiency and replacement in elderly hypopituitary adults. Growth Horm IGF Res 14 Suppl A:S51–S58
- 68. Fernholm R, Bramnert M, Hagg E, Hilding A, Baylink DJ, Mohan S, Thoren M 2000 Growth hormone replacement therapy improves body composition and increases bone metabolism in elderly patients with pituitary disease. J Clin Endocrinol Metab 85:4104–4112
- 69. Franco C, Johannsson G, Bengtsson BA, Svensson J 2006 Baseline characteristics and effects of growth hormone therapy over two years in younger and elderly adults with adult onset GH deficiency. J Clin Endocrinol Metab 91:4408–4414
- 70. Gill MS, Toogood AA, Jones J, Clayton PE, Shalet SM 1999 Serum leptin response to the acute and chronic administration of growth hormone (GH) to elderly subjects with GH deficiency. J Clin Endocrinol Metab 84:1288–1295
- 71. Gotherstrom G, Bengtsson BA, Sunnerhagen KS, Johannsson G, Svensson J 2005 The effects of five-year growth hormone replacement therapy on muscle strength in elderly hypopituitary patients. Clin Endocrinol (Oxf) 62:105–113
- Gotherstrom G, Elbornsson M, Stibrant-Sunnerhagen K, Bengtsson BA, Johannsson G, Svensson J 2010 Muscle strength in elderly adults with GH deficiency after 10 years of GH replacement. Eur J Endocrinol 163:207–215
- 73. Koltowska-Haggstrom M, Mattsson AF, Shalet SM 2009 Assessment of quality of life in adult patients with GH deficiency: KIMS contribution to clinical practice and pharmacoeconomic evaluations. Eur J Endocrinol 161 Suppl 1:S51–S64
- 74. Monson JP, Abs R, Bengtsson BA, Bennmarker H, Feldt-Rasmussen U, Hernberg-Stahl E, Thoren M, Westberg B, Wilton P, Wuster C 2000 Growth hormone deficiency and replacement in elderly hypopituitary adults. KIMS Study Group and the KIMS International Board. Pharmacia and Upjohn International Metabolic Database. Clin Endocrinol (Oxf) 53:281–289
- 75. Monson JP, Jonsson P 2003 Aspects of growth hormone (GH) replacement in elderly patients with GH deficiency: data from KIMS. Horm Res 60:112–120
- 76. Sathiavageeswaran M, Burman P, Lawrence D, Harris AG, Falleti MG, Maruff P, Wass J 2007 Effects of GH on cognitive function in elderly patients with adult-onset GH deficiency: a placebo-controlled 12-month study. Eur J Endocrinol 156:439–447
- 77. Toogood AA, Shalet SM 1999 Growth hormone replacement therapy in the elderly with hypothalamic-pituitary disease: a dose-finding study. J Clin Endocrinol Metab 84:131–136

- 78. White HD, Ahmad AM, Durham BH, Patwala A, Whittingham P, Fraser WD, Vora JP 2005 Growth hormone replacement is important for the restoration of parathyroid hormone sensitivity and improvement in bone metabolism in older adult growth hormone-deficient patients. J Clin Endocrinol Metab 90:3371–3380
- 79. Colao A, Di SC, Savastano S, Rota F, Savanelli MC, Aimaretti G, Lombardi G 2009 A reappraisal of diagnosing GH deficiency in adults: role of gender, age, waist circumference, and body mass index. J Clin Endocrinol Metab 94:4414–4422
- Eskes SA, Tomasoa NB, Endert E, Geskus RB, Fliers E, Wiersinga WM 2009 Establishment of reference values for endocrine tests. Part VII: growth hormone deficiency. Neth J Med 67:127–133
- 81. Finucane FM, Liew A, Thornton E, Rogers B, Tormey W, Agha A 2008 Clinical insights into the safety and utility of the insulin tolerance test (ITT) in the assessment of the hypothalamo-pituitary-adrenal axis. Clin Endocrinol (Oxf) 69:603–607
- 82. Bartke A, Brown-Borg H 2004 Life extension in the dwarf mouse. Curr Top Dev Biol 63:189–225
- Bartke A 2008 Growth hormone and aging: a challenging controversy. Clin Interv Aging 3:659–665
- 84. Bonkowski MS, Pamenter RW, Rocha JS, Masternak MM, Panici JA, Bartke A 2006 Long-lived growth hormone receptor knockout mice show a delay in age-related changes of body composition and bone characteristics. J Gerontol A Biol Sci Med Sci 61:562–567
- 85. Brosnahan MM, Paradis MR 2003 Assessment of clinical characteristics, management practices, and activities of geriatric horses. J Am Vet Med Assoc 223:99–103
- 86. Coschigano KT, Holland AN, Riders ME, List EO, Flyvbjerg A, Kopchick JJ 2003 Deletion, but not antagonism, of the mouse growth hormone receptor results in severely decreased body weights, insulin, and insulin-like growth factor I levels and increased life span. Endocrinology 144:3799–3810
- 87. Greer KA, Canterberry SC, Murphy KE 2007 Statistical analysis regarding the effects of height and weight on life span of the domestic dog. Res Vet Sci 82:208–214
- 88. Patronek GJ, Waters DJ, Glickman LT 1997 Comparative longevity of pet dogs and humans: implications for gerontology research. J Gerontol A Biol Sci Med Sci 52:B171–B178
- 89. Rollo CD 2002 Growth negatively impacts the life span of mammals. Evol Dev 4:55-61

Chapter 9

Nederlandse samenvatting

Inhoud

I.	Inleiding	221
II.	Evaluatie van de functie van de hypofyse na traumatisch	
	hersenletsel	223
III.	Dynamisch testen van de hypofysefunctie bij andere	
	hypofysaire aandoeningen	226
	Endocriene evaluatie na hypofyse operatie	226
	Endocriene evaluatie na craniële radiotherapie	230
IV.	Behandeling van GHD	231
V.	Slotopmerkingen	233

I. Inleiding

Voor de beoordeling van de functie van de hypofyse zijn diverse diagnostische testen beschikbaar. Er kan gebruik gemaakt worden van basale hormoonspiegels, maar ook van hormoon stimulatietesten. Voor de beoordeling van de schildklier-as, de gonadotrope-as en het prolactine is het voldoende om basale (niet-gestimuleerde) hormoonspiegels te bepalen vroeg in de ochtend (8–9 uur) en daarbij rekening te houden met de klinische symptomen van de patiënt. Echter, de afgifte van cortisol kenmerkt zich door een sterk dag-nacht ritme terwijl de groeihormoon (GH) secretie wordt gekenmerkt door afgifte in pulsen. Voor de beoordeling van de hypofyse bijnier- en GH-IGF-I as is het dus niet mogelijk alleen basale serum hormoonspiegels te bepalen. Voor de juiste evaluatie van deze assen is het gebruik van een stimulatie test noodzakelijk.

Er zijn verschillende stimulatietesten beschikbaar, waarvoor verschillende afkapwaarden worden gebruikt. Wanneer er gebruik wordt gemaakt van een stimulatietest is het van belang rekening te houden met verschillende factoren die van invloed kunnen zijn, zoals leeftijd, geslacht, body mass index (BMI) en medicatiegebruik. Uit onderzoek is gebleken dat met vorderende leeftijd en hogere BMI de secretie van GH afneemt, resulterend in lagere GH spiegels.

Na traumatisch hersenletsel, een operatie aan de hypofyse, of bestraling van de hypofyse kunnen zowel de hypofyse als de hypothalamus beschadigd raken, met uitval van de hypofysefuncties (hypopituïtarisme) als gevolg. Uit onderzoek is echter gebleken dat hypofyseweefsel in staat is te herstellen. Dit gebeurt voornamelijk binnen het eerste jaar na trauma en/of operatie. Wanneer in deze periode een stimulatie test wordt gebruikt om de functie van de hypofyse te bepalen, kan dit dus tot een verkeerde interpretatie van de uiteindelijke restfunctie van de hypofyse leiden.

Het doel van dit proefschrift is om beter inzicht te krijgen in de complexiteit van de verschillende testen die beschikbaar zijn voor de evaluatie van de hypofysefuncties bij verschillende aandoeningen die gepaard kunnen gaan met passagière of persisterende schade aan de hypofyse en/of de hypothalamus. Het effect van het gebruik van verschillende endocriene testen op de gevonden mate van uitval wordt hierbij kritisch tegen het licht gehouden. Ook is dit proefschrift bedoeld om beter inzicht te krijgen in de behandeling van patiënten met hypofyse uitval.

II. Evaluatie van de functie van de hypofyse na traumatisch hersenletsel

Patiënten die een traumatisch hersenletsel hebben ondergaan kunnen persisterende, invaliderende klachten hebben die lijken op de klachten van patiënten met hypopituïtarisme (zoals verminderde cognitie, depressie, vermoeidheid en een verminderde kwaliteit van leven (QoL)). Het kan dan ook zo zijn dat aanwezige, maar niet gediagnostiseerde uitval van de hypofysefuncties na trauma bijdraagt aan de klachten van deze patiënten. De behandeling van hypopituïtarisme is relatief makkelijk (toedienen van de ontbrekende hormonen) en uit onderzoek is gebleken dat de behandeling de kwaliteit van leven significant kan verbeteren. Het is dus van belang hypopituïtarisme bij patiënten na een hersentrauma vast te stellen, dan wel uit te sluiten.

In de afgelopen jaren is er veelvuldig onderzoek gedaan naar de relatie tussen traumatisch hersenletsel en hormoonuitval. Hierbij hebben verschillende onderzoeken een hoog percentage hypofyse-uitval na TBI beschreven. Er is echter wel een grote variatie in deze beschreven prevalentie. Tijdens de beoordeling van de functie van de hypofyse na hersentrauma is het van belang rekening te houden met de volgende factoren die van invloed kunnen zijn: het tijdsinterval tussen trauma en de endocriene beoordeling, het type en de ernst van het trauma, maar ook de methoden (d.w.z. endocriene testen en criteria) die gebruikt worden om de diagnose hypopituïtarisme te stellen. Sommige studies hebben geconcludeerd dat hypopituïtarisme een veel voorkomende complicatie is na TBI, maar zij hebben geen rekening gehouden met de grote variabiliteit in diagnostische testen en definities van hypofyse uitval. Het doel van onze systematische review in **hoofdstuk 2** was dan ook om kritisch te kijken naar de gebruikte stimulatie testen en de definities van hypopituïtarisme van studies die hebben gekeken naar de lange termijn uitkomsten na hersentrauma.

Uit onze review blijkt dat er inderdaad een grote variatie bestaat tussen de gerapporteerde prevalentie cijfers, waarschijnlijk gebaseerd op de grote verschillen in endocriene en analytische methoden. De studies in de review gebruiken verschillende stimulatie testen, afkapwaarden en analytische methoden. Bovendien is geen rekening gehouden met factoren die mogelijk van invloed kunnen zijn, zoals BMI. Dit is vooral van belang voor de beoordeling van de GH-IGF-I-as, aangezien de concentratie GH daalt met een hogere BMI. Dit kan resulteren in een overschatting van het percentage GH uitval na TBI bij patiënten met overgewicht. Al deze verschillen maken het moeilijk de resultaten van studies over TBI te vergelijken. Er is meer onderzoek nodig naar voorspellende factoren voor het optreden van hypopituïtarisme na TBI.

Naar aanleiding van de grote variatie in prevalentie cijfers en de variatie in de endocriene en analytische methoden om de hypofyse functies te beoordelen, hebben we in **hoofdstuk 3** onderzoek gedaan naar de lange termijn consequenties bij een grote groep patiënten na hersentrauma. In dit onderzoek hebben we 112 patiënten geïncludeerd die in het verleden traumatisch hersenletsel hadden opgelopen. De patiënten moesten minstens drie dagen in het ziekenhuis hebben gelegen en het trauma moest meer dan een jaar geleden plaats hebben gevonden. Vijf verschillende ziekenhuizen in Nederland hebben geparticipeerd in dit onderzoek. De functie van de hypofyse werd geëvalueerd middels nuchtere basale hormoonspiegels in combinatie met een insuline tolerantie test (ITT n = 90) of, wanneer een ITT gecontraïndiceerd was, een ACTH stimulatie en/of CRH stimulatie test en een gecombineerde GHRH-arginine test (n = 22).

Uit dit onderzoek bleek dat de prevalentie van hypopituïtarisme na traumatisch hersenletsel na lange follow-up laag was. Wanneer gebruik wordt gemaakt van de gouden standaard test (de ITT) voor de evaluatie van GH- en cortisol secretie in de meerderheid van de patiënten, vonden wij een percentage hypofyse uitval van slechts 5.4% (een ernstige groeihormoondeficiëntie (2.8%), hypogonadisme (0.9%), bijnierinsufficiëntie (1.8%)). Dit percentage uitval is veel lager in vergelijking met de prevalentie cijfers gerapporteerd in eerdere soortgelijke studies (15–90%). Dit kan worden verklaard door het gebruik van verschillende endocriene testen en verschillende afkapwaarden om de diagnose hypopituïtarisme te stellen. Waar mogelijk hebben wij in onze studie gebruik gemaakt van de gouden standaard test: de ITT. In overeenstemming met onderzoek verricht door Klose en collegae, resulteerde dit in een lage prevalentie van GHD en bijnierinsufficiëntie. Als de ITT was gecontraïndiceerd, gebruikten we de gecombineerde stimulatie met GHRH en arginine voor de beoordeling van de GH-as. Het is aangetoond dat dit een goede alternatieve test is, mits de gebruikte afkapwaarden worden aangepast aan de BMI. In ons onderzoek hebben we wel gebruik gemaakt van aangepaste afkapwaarden.

Naast verschillen in endocriene testen, kan ook het tijdsinterval tussen trauma en de endocriene evaluatie, alsook de ernst van het trauma van invloed zijn op de gerapporteerde prevalentie cijfers. In de acute fase na een traumatisch hersenletsel zijn voorbijgaande hormoonveranderingen die lijken op hypopituïtarisme veelvuldig beschreven. Om te voorkomen dat dit van invloed zou kunnen zijn op de endocriene evaluatie in onze studie hebben wij alle patiënten minimaal een jaar na trauma geëvalueerd. Een ernstig trauma verhoogt het risico van het optreden van hypofyse uitval, zodat we tevens besloten hebben om in onze studie alleen patiënten met een ernstig trauma te evalueren. We hebben derhalve alleen patiënten geïncludeerd met een opname duur (ten gevolge van het trauma) van minstens drie dagen. Daarnaast is de Glasgow Coma Scale (GCS) ten tijde van opname beoordeeld om een indruk te krijgen van de ernst van het trauma. Ondanks het feit dat wij met al deze factoren rekening hebben gehouden, bleek het percentage hormoonuitval uiteindelijk toch laag.

Als er sprake is van hypofyse-uitval, kan behandeling hiervan van levensbelang zijn en een aanzienlijke verbetering van de kwaliteit van leven geven. Daarom is het van belang om vooralsnog bij elke patiënt rekening te houden met mogelijke uitval van hypofysefuncties na hersentrauma, ondanks het feit dat op basis van onze resultaten dit mogelijk niet-frequent voorkomt. Bovendien is het van belang dat er consensus komt voor een meer uniforme endocriene evaluatie van hypofysefuncties in het algemeen, en na traumatisch hersenletsel in het bijzonder.

III. Dynamisch testen van de hypofysefunctie bij andere hypofysaire aandoeningen

Hypofyse adenomen kunnen behandeld worden middels transsfenoidale chirurgie (TS), aanvullende radiotherapie en/of medicatie. Een complicatie die kan optreden ten gevolge van de tumor zelf (compressie), of als gevolg van de behandeling (chirurgie en/of radiotherapie) is uitval van de hypofysefuncties. Voor een passende behandeling van patiënten met hypofyse adenomen is het daarom van essentieel belang de functie van de hypofyse te beoordelen ook na operatie en na bestraling.

Endocriene evaluatie na hypofyse operatie

Na TS is het van groot klinisch belang om de hypothalamus-hypofysebijnieras (HPA as) te beoordelen, aangezien het belangrijk is te weten of de patiënt bijnierschorshormoon (hydrocortison) afhankelijk is. De gouden standaard test voor de evaluatie van de HPA as is de ITT. Als de ITT vanwege contra-indicaties (epilepsie en/of coronair lijden) niet kan worden uitgevoerd, zijn er verschillende andere dynamische testen beschikbaar, zoals de metyrapon test, de ACTH stimulatie test en de corticotrofine releasing hormoon (CRH) test. Sinds 1990 wordt in ons ziekenhuis de bijnierfunctie van patiënten direct na hypofyse operatie beoordeeld door stimulatie met CRH. Op basis van de testuitslag wordt vervolgens besloten of de patiënten met of zonder hydrocortison substitutietherapie naar huis wordt ontslagen. Gebaseerd op onze klinische ervaring is dit een goede test om de patiënten te screenen op uitval van HPA as. Er is echter weinig literatuur beschikbaar met betrekking tot de klinische toepasbaarheid van de CRH test direct na TS. Daarom hebben we in **hoofdstuk 4** gekeken naar de klinische toepasbaarheid van deze test bij patiënten behandeld voor niet-ACTH producerende hypofyse-adenomen in ons ziekenhuis.

We hebben retrospectief gekeken naar data van alle patiënten behandeld met TS in ons centrum. In totaal hebben we 144 niet-Cushing patiënten geïncludeerd van wie gegevens beschikbaar waren met betrekking tot de CRH test direct na operatie, van wie tijdens de follow-up periode een bevestigingstest beschikbaar was en waarbij geen sprake was van het gebruik van exogene glucocorticoïden, of een tweede behandeling (zoals heroperatie of aanvullende radiotherapie) die een vergelijk met de eerste test onmogelijk zou maken.

Op basis van de CRH test bleek er bij 42 patiënten sprake te zijn van hypocortisolisme. Echter, 13 van deze patiënten (31%) bleek uiteindelijk, op basis van de bevestigingstest, geen uitval van de HPA as te hebben. Hierbij kon de hydrocortsion suppletie worden gestopt. Een mogelijke verklaring voor deze discrepante resultaten is het gebruik van verschillende afkapwaarden. Voor de ITT zijn er alom geaccepteerde afkapwaarden beschikbaar. Daarentegen, voor de CRH stimulatie test zijn er verschillende afkapwaarden gedefinieerd die allemaal resulteren in een andere sensitiviteit en specificiteit van de test. In ons ziekenhuis wordt de CRH test als screeningstest gebruik om hypocortisolisme direct na TS op te sporen. Wij gebruiken daarom een stringente en hoge afkap waarde van 550 nmol/L. Echter, het streven naar een sensitieve test gaat ten koste van de specificiteit van een test. Hierdoor zal dus een grotere aantal patiënten onjuist als bijnierinsufficiënt worden gediagnosticeerd. Een andere verklaring zou het herstel van de HPA as kunnen zijn. Een recente studie heeft drie en twaalf maanden na TS de HPA as beoordeeld middels ITT. Hieruit bleek dat er binnen het jaar herstel was opgetreden van de HPA as. In overeenstemming met deze studie hebben wij ook 8 patiënten die in eerste instantie gediagnosticeerd waren met bijnieruitval, maar waarbij binnen een jaar na chirurgie herstel van de HPA as optrad. Dit toont aan dat een langdurige, uitgebreide follow-up van patiënten na TS van belang is.

Bij 102 patiënten bleek er direct na TS sprake te zijn van een normale functie van de HPA as. Echter, bij 14 van deze patiënten bleek dat er alsnog sprake was van hypocortisolisme op basis van een tweede test. Deze discrepante resultaten kunnen potentieel levensbedreigend zijn, aangezien deze patiënten at risk zijn voor het optreden van een adrenale crisis. Waarschijnlijk speelt uitval van overige hypofyse assen hierbij een rol, daar 13 van deze 14 patiënten namelijk ook uitval bleken te hebben van andere hypofyse hormonen. Uit onderzoek is gebleken dat GH en TSH deficiëntie van invloed kunnen zijn op deze testuitslagen. Bovendien kan rhGH therapie bij patiënten met GHD ook een belangrijke rol spelen, vanwege de invloed van GH op het cortisol metabolisme. Groeihormoon stimuleert een enzym dat cruciaal is voor de werking van cortisol op weefselniveau, het 11- β hydroxysteroid dehydrogenase (11 β HSD-1), dat de omzetting van cortisol-cortison reguleert. Het gebruik van rhGH bij GHD patiënten kan daarom een cortisol deficiëntie ontmaskeren.

Op basis van onze resultaten concluderen wij dat de CRH test veilig kan worden gebruikt om te evalueren of een patiënt na TS met of zonder HC naar huis kan worden ontslagen. Desalniettemin is de CRH test niet bij alle patiënten een betrouwbare voorspeller. Wij adviseren daarom om de HPA as tijdens follow-up middels een tweede test te evalueren, bijvoorbeeld 3–6 maanden na operatie. Bij patiënten met een verminderde reactie van cortisol op CRH direct postoperatief, die uitval van meerdere hypofyse hormonen blijken te hebben, is hertesten niet noodzakelijk.

In hoofdstuk 4 hebben we retrospectief gekeken naar alle patiënten die behandeld zijn met TS in ons centrum. In hoofdstuk 5 hebben we ons gericht op een specifieke groep patiënten; patiënten behandeld met TS voor een GH producerende tumor d.w.z. patiënten met acromegalie. De aanleiding voor ons onderzoek was een recente studie van Ronchi en collegae die de functie van de HPA as bij acromegalie patienten behandeld middels TS hebben onderzocht. Zij vonden een opmerkelijk hoge prevalentie bijnierinsufficiëntie (32%) bij deze patiënten. Zij concludeerden dat de functie van de HPA as kan afnemen na verloop van tijd en dat deze as dus zorgvuldig tijdens follow-up na behandeling gecontroleerd moet blijven worden in alle acromegalie patiënten, ongeacht de aard van de behandeling. Deze aanbeveling heeft duidelijk gevolgen voor de lange termijn behandeling van patiënten met acromegalie. Het doel van hoofdstuk 5 was dan ook om de prevalentie van bijnierinsufficiëntie na lange termijn follow-up te bepalen in ons eigen cohort van patiënten met acromegalie in remissie na behandeling met TS. We hebben retrospectief gekeken naar de gegevens van 91 patiënten in remissie na TS waarbij de HPA as was beoordeeld met behulp van de ITT, CRH stimulatie, metyrapon- of ACTH stimulatie test. In ons cohort bleek er bij 16 patiënten (18%) in de direct postoperatieve periode sprake te zijn van bijnierinsufficiëntie. Dit bleek van voorbijgaande aard te zijn bij 8 patiënten en blijvend bij de overige 8 gedurende het eerste jaar na operatie. Dus een jaar na operatie was de prevalentie bijnierinsufficiëntie uiteindelijk slechts 9%. Slechts bij drie patiënten ontstond alsnog een bijnierinsufficiëntie 13, 18 en 24 jaar na de operatie. De incidentie van late bijnierinsufficiëntie na een succesvolle operatie was dus slechts 2 / 1000 persoonsjaren. De prevalentie van bijnierinsufficiëntie bij patiënten in remissie na chirurgie voor acromegalie na langdurige follow-up (mediane follow-up 8.1 jaar (range 1–31 jaar)), was 12%. Op basis van onze gegevens kunnen wij zeggen dat het ontstaan van bijnierinsufficiëntie na TS niet vaak voorkomt.

Het verschil in prevalentie tussen onze studie en de studie van Ronchi en collegae kan worden verklaard door verschillen in onderzoeksopzet en patiëntenpopulatie, maar ook door verschillen in patiëntenselectie en chirurgische technieken. In ons cohort hebben we in de meerderheid van de patiënten gebruik gemaakt van de gouden standaard test (ITT) en een CRH test terwijl in de studie van Ronchi gebruik is gemaakt van een lage dosis ACTH test. Daarnaast zouden de volgende factoren ook mogelijk van invloed kunnen zijn: een veranderd niveau van cortisol bindend globuline (CBG) in acromegalie, de aanwezigheid van postoperatieve GH deficiëntie en de mogelijkheid van herstel van preoperatieve bijnierinsufficiëntie na TS.

Beperkingen van onze studie zijn de retrospectieve aard van de studie en het feit dat de HPA as is beoordeeld middels verschillende cortisol stimulatie testen. Dit doet echter geen afbreuk aan onze conclusies, omdat de ITT, CRH en metyrapon tests allen worden geaccepteerd als test voor de evaluatie van de HPA-as.

Op basis van onze gegevens stellen wij voor om de HPA as een jaar na operatie opnieuw dynamisch te testen bij alle patiënten met postoperatie HPA insufficiëntie. Verder onderzoek is nodig om te beoordelen of jaarlijks basale cortisol waarden kunnen volstaan om de bijnierfunctie te monitoren bij asymptomatische patiënten. Wanneer er echter sprake is van lage basale cortisol spiegels, symptomen van corticotrope insufficiëntie of overige hypofyse uitval, moeten aanvullende dynamisch testen van de HPA as worden uitgevoerd.

Endocriene evaluatie na craniële radiotherapie

Patiënten met niet-hypofysaire hersentumoren en/of tumoren in het neus- en keel gebied worden vaak behandeld met radiotherapie. Tijdens deze behandeling ligt de hypofyse ook in het bestralingsgebied wat kan resulteren in hypofyse insufficiëntie. Voor kinderen is dit een goed beschreven complicatie na radiotherapie. Echter voor volwassenen is de beoordeling van de hypofysefunctie nog niet opgenomen in de richtlijnen voor de follow-up van deze patiënten. In **hoofdstuk 6** rapporteren we een systematisch review waarbij we hebben gekeken naar de prevalentie van hypofyse insufficiëntie na craniële radiotherapie voor niet-hypofysaire tumoren bij volwassen patiënten.

We hebben uiteindelijk slechts 18 studies (n=813) kunnen includeren. Er zijn grote verschillen in de gerapporteerde prevalentie van hypopituïtarisme na craniale radiotherapie, variërend van nauwelijks effect op de hypofyse functie tot bijna 100% van de patiënten met uitval.

Het verschil in prevalentie kan worden toegekend aan verschillen in radiotherapeutische technieken, verschil in studie design, tijdstip van de evaluatie, selectie van patiënten en de verschillen in endocriene evaluatie. Bij de meerderheid van de patiënten werd geen gebruik gemaakt van hormoon stimulatietesten om de hypofysefunctie te beoordelen. Wanneer er wel gebruik werd gemaakt van stimulatietesten, werden er verschillende afkapwaarden en diagnostische criteria gebruikt. De uitgevoerde meta-analyse toonde aan dat er bij ongeveer tweederde van alle volwassenen sprake is van enige vorm van hypofyse-uitval (0.66, 95% CI 0.55–0.76). De prevalentie van groeihormoondeficiëntie was 0.45 (95% CI 0.33–0.57), van LH en FSH 0.3 (95% CI 0.23–0.37), van TSH 0.25 (95% CI 0.16–0.37), en van ACTH 0.22 (95% CI 0.15–0.3). De prevalentie van hyperprolactinemie was 0.34 (95% CI 0.15-0.6). Er waren geen verschillen tussen de effecten van radiotherapie voor nasofaryngeale versus intracerebrale tumoren. Op basis van deze gegevens kunnen we concluderen dat hypopituïtarisme een veelvoorkomende complicatie is ook bij volwassen patiënten die behandeld zijn met craniële radiotherapie. Gezien deze hoge prevalentie zou de evaluatie van hypofysefuncties moeten worden opgenomen in de richtlijnen voor de follow-up van alle patiënten behandeld met craniële radiotherapie.

IV. Behandeling van GHD

Groeihormoon deficiëntie (GHD) kan behandeld worden door toediening van recombinant GH (rhGH). Echter bij de behandeling van patiënten met GHD moet onder andere rekening worden gehouden met de leeftijd aangezien de normale afgifte van GH afneemt met de leeftijd.

In **hoofdstuk** 7 hebben we kritisch gekeken naar de beschikbare literatuur met betrekking tot de behandeling van de oudere GHD patiënt. We hebben gezocht naar artikelen die de effecten van rhGH bij patiënten boven de 60 jaar hebben geëvalueerd. Uiteindelijk konden we 11 bruikbare studies met een totaal van 543 patiënten meenemen in onze review. De studies laten zien dat er inderdaad een positief effect op totaal- en LDL-cholesterol is en dat ook de kwaliteit van leven bij deze patiënten verbeterd. Voor de overige uitkomstmaten wordt er echter geen positief effect beschreven. Ook zijn er geen gegevens over het effect van rhGH op klinisch relevante eindpunten zoals hart- en vaatziekten of het voorkomen van botbreuken.

Bij de beoordeling van het effect van rhGH therapie bij oudere patiënten moet rekening worden gehouden met verschillende factoren. Allereerst is het van belang rekening te houden met het feit dat met toenemende leeftijd, de GH productie en dus secretie afneemt. Deze afname kan invloed hebben op de uitkomsten van de stimulatietesten en indirect zal het dus ook van invloed zijn op de afkapwaarden die worden gebruikt. Studies die hebben gekeken naar het gebruik van de ITT en de gecombineerde GHRH met arginine test bij oudere patiënten laten verschillende uitkomsten zien. Sommige studies geven aan dat er een lagere GH piek is bij ouderen vergeleken met jongere patiënten, terwijl andere studies geen verschil laten zien. Wanneer er helemaal geen rekening wordt gehouden met leeftijd bij het bepalen van de afkapwaarden kan het zijn dat patiënten onterecht gediagnosticeerd worden met GHD. Ondanks deze controversiële uitkomsten moet er toch rekening gehouden worden met leeftijd. In hoeverre dit van invloed is voor het effect van rhGH therapie bij deze patiëntengroep is nog onduidelijk.

De afname van de GH concentratie gaat gepaard met een daling van de IGF-I concentraties. IGF-I SD scores worden gebruikt voor de diagnose van GHD, maar ook voor het goed instellen van de rhGH behandeling. Het is dus ook noodzakelijk voor leeftijd gecorrigeerde IGF-I SD scores te bepalen. Vanuit een evolutionair perspectief blijkt de natuurlijke daling van GH en IGF-I tijdens veroudering zelfs gunstig. In onderzoek met dieren blijkt dat een verminderde GH-IGF-I functie de levensduur van de dieren verlengd. In hoeverre dit van toepassing is bij de mens is nog niet duidelijk. Maar een klein aantal studies laat een gunstig effect zien van rhGH bij oudere patiënten. De vraag blijft dus of de behandeling van ouderen met rhGH klinisch relevant is. Er is meer onderzoek nodig in deze patiënten groep om dit goed te kunnen beoordelen.

V. Slotopmerkingen

Hypofyse functie na TBI

Er is een grote variatie in gerapporteerde prevalentie cijfers van hypopituïtarisme na traumatisch hersenletsel. Dit wordt voor een deel veroorzaakt door het gebruik van verschillende endocriene testen, verschillende afkapwaarden en het niet rekening houden met overige factoren die van invloed (kunnen) zijn (zoals BMI). Door deze methodologische verschillen is het niet mogelijk de uitkomsten van deze verschillende studies met elkaar te vergelijken.

Wanneer juiste endocriene testen en strenge criteria worden gebruikt is de prevalentie van hypopituïtarisme na een traumatisch hersenletsel waarschijnlijk zeer laag. De gerapporteerde prevalentie van hypofyse insufficiëntie na een traumatisch hersenletsel is zeer waarschijnlijk overschat.

Hypofyse functie na transsfenoïdale chirurgie

De CRH test lijkt een waardevolle aanvulling in het diagnostisch arsenaal van endocrinologische testen om klinisch relevant cortisol tekort te definiëren onmiddellijk na hypofyse chirurgie. Deze test is weinig belastend en kan veilig worden gebruikt om te beslissen of patiënten met of zonder hydrocortison uit het ziekenhuis worden ontslagen. Gedurende follow-up is het echter wel noodzakelijk de HPA as nogmaals te testen.

De prevalentie van bijnierinsufficiëntie bij patiënten in remissie van acromegalie na TS is zeer laag (9%) een jaar na operatie. Het optreden van 'late-onset' bijnierinsufficiëntie is een zeer zeldzame complicatie bij patiënten met acromegalie in remissie na transsfenoidale chirurgie.

Hypofyse functie na craniële radiotherapie

Hypopituïtarisme is een veelvoorkomende complicatie bij volwassenen na craniële radiotherapie voor niet-hypofysaire tumoren. Daarom moeten alle patiënten die worden behandeld met craniële radiotherapie tijdens follow-up een gestructureerde periodieke beoordeling van de hypofyse functies ondergaan. Dit moet worden opgenomen in richtlijnen voor de behandeling en follow-up van deze patiënten.

Hypofyse functie bij ouderen

RhGH therapie bij oudere patiënten met GHD verlaagt het LDLcholesterol gehalte en verbetert de kwaliteit van leven, maar de overige effecten zijn niet eenduidig. Er zijn geen gegevens over de werkzaamheid en veiligheid van rhGH behandeling in tachtigjarige patiënten met GHD. Er zijn tot slot ook geen gegevens bij oudere patiënten met GHD over klinisch relevante eindpunten zoals cardiovasculaire aandoeningen of breuken. Gezien de hoge kosten van de behandeling en de potentiële schadelijke effecten van suprafysiologische rhGH substitutie op het ontstaan van maligniteiten adviseren we gerandomiseerd onderzoek bij de oudere patiënt met GHD met een weloverwogen lange termijn kostenbaten analyse.

Curriculum Vitae

Nieke Eva Kokshoorn werd geboren op 4 december 1982 te Voorburg. Zij behaalde in 1999 eerst haar HAVO diploma gevolgd door haar VWO diploma in 2001. Beide aan het Veurs College te Leidschendam. Aangezien ze het vak Scheikunde miste om mee te mogen loten voor de opleiding Geneeskunde heeft ze deze in 2001 behaald bij het Kades College te Den Haag. Ze werd ingeloot voor de opleiding Geneeskunde aan de Universiteit van Maastricht. Na zes mooie jaren in Maastricht te hebben doorgebracht heeft zij in 2008 haar artsenbeul behaald. In september 2008 startte zij als promovenda bij de afdeling Endocrinologie van het Leids Universitair Medisch Centrum onder leiding van Prof. dr. J.A. Romijn, Prof. dr. J.W.A. Smit en Dr. A.M. Pereira. De resultaten van haar promotieonderzoek staan beschreven in dit proefschrift.

List of publications

- Kokshoorn NE, Wassenaar MJ, Biermasz NR, Roelfsema F, Smit JW, Romijn JA, Pereira AM. Hypopituitarism following traumatic brain injury: prevalence is affected by the use of different dynamic tests and different normal values. Eur J Endocrinol. 2010 Jan;162(1):11-8. Epub 2009 Sep 25. Review.
- Tiemensma J, Kokshoorn NE, Biermasz NR, Keijser BJ, Wassenaar MJ, Middelkoop HA, Pereira AM, Romijn JA. Subtle cognitive impairments in patients with long-term cure of Cushing's disease J Clin Endocrinol Metab. 2010 Jun;95(6):2699-714. Epub 2010 Apr 6.
- 3. **Kokshoorn NE**, Biermasz NR, Roelfsema F, Smit JW, Pereira AM, Romijn JA. GH replacement therapy in elderly GH-deficient patients: a systematic review. Eur J Endocrinol. 2011 May;164(5):657-65. Epub 2011 Feb 21. Review.
- **4. Kokshoorn NE**, Burgers AM, Pereira AM, Roelfsema F, Smit JW, Biermasz NR, Romijn JA. Low incidence of adrenal insufficiency after transsphenoidal surgery in patients with acromegaly: a long-term follow-up study. J Clin Endocrinol Metab. 2011 Jul;96(7):E1163-70. Epub 2011 Apr 6.
- Kokshoorn NE, Appelman-Dijkstra NM, Dekkers OM, Neelis KJ, Biermasz NR, Romijn JA, Smit JW, Pereira AM. Pituitary dysfunction in adult patients after cranial radiotherapy: systematic review and meta-analysis. J Clin Endocrinol Metab. 2011 Aug;96(8):2330-40. Epub 2011 May 25.
- Kokshoorn NE, Smit JW, Nieuwlaat WA, Tiemensma J, Bisschop PH, Groote Veldman R, Roelfsema F, Franken AA, Wassenaar MJ, Biermasz NR, Romijn JA, Pereira AM. Low prevalence of hypopituitarism after traumatic brain injury: a multicenter study. Eur J Endocrinol. 2011 Aug;165(2):225-31. Epub 2011 Jun 6.
- 7. **Kokshoorn NE**, Romijn JA, Roelfsema F, Rambach AHJH, Smit JWA, Biermasz NR, Pereira AM. The use of an early postoperative CRH test to assess adrenal function after transsphenoidal surgery for pituitary adenomas. Pituitary. 2011 Sep 9. [Epub ahead of print].

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