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Extensive cytonuclear discordance in a crested newt from the Balkan Peninsula glacial refugium

BEN WIELSTRA^{1,2,*,•} and JAN W. ARNTZEN²

¹Institute of Biology Leiden, Leiden University, 2300 RA, Leiden, The Netherlands ²Naturalis Biodiversity Center, 2300 RA Leiden, The Netherlands

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Integration of multilocus data and species distribution modelling into phylogeography allows mitochondrial DNA (mtDNA)-based scenarios to be fine-tuned. We address the question of whether extensive mtDNA substructuring in the crested newt *Triturus macedonicus* from the Balkan Peninsula is matched in the nuclear genome. We determine the intraspecific population structure based on 52 nuclear DNA markers and project a species distribution model on climate layers for the Last Glacial Maximum. We show that *T. macedonicus* accumulated nuclear DNA population structure in an area predicted to have been climatically stable during the Pleistocene, with four nuclear DNA groups in the western part of the species range. The distribution of these nuclear DNA groups shows little agreement with that of mtDNA structuring, which shows three highly distinct species-specific clades and a fourth one introgressed from another crested newt species. This cytonuclear discordance conveys that historical biogeographical scenarios based on mtDNA exclusively should be interpreted with caution. Our findings further highlight the important role the Balkan Peninsula has played in the evolution and preservation of European biodiversity.

ADDITIONAL KEYWORDS: glacial refugium – historical biogeography – Ion Torrent – nuclear DNA – postglacial expansion – *Triturus macedonicus*.

INTRODUCTION

The aim of phylogeography is to reconstruct the biogeographical history of species from geographical patterns in genetic structure (Avise, 2000). Historically, phylogeographical surveys have relied mainly on mitochondrial DNA (mtDNA; Beheregaray, 2008), which is more susceptible to geographical structuring than the rest of the genome (Petit & Excoffier, 2009). This raises the question: how representative is the mtDNA geographical structure of the overall history of the taxon under study? With 'next-generation phylogeography' (Puritz *et al.*, 2012), in which many unlinked nuclear markers are consulted, this question can now be addressed (Ekblom & Galindo, 2011; McCormack *et al.*, 2013; Garrick *et al.*, 2015).

The field of phylogeography has been particularly prolific in unravelling the influence of the climate cycles of the Pleistocene Ice Age on biodiversity (Hewitt, 2004). During glacial cycles, many temperate

*Corresponding author. E-mail: b.m.wielstra@biology. leidenuniv.nl species had their ranges reduced to glacial refugia, whereas during interglacials, they could expand their ranges (Bennett & Provan, 2008). This cyclical contraction-and-expansion pattern has left its mark on genetic variation within species, with populations that persisted in glacial refugia showing relatively high genetic diversity and populations established during recent colonization being genetically depleted (Wallis & Arntzen, 1989; Taberlet et al., 1998; Hewitt, 2000). Furthermore, interglacial range expansion allowed secondary contact among populations and species, facilitating genetic admixture and introgression (Hewitt, 2011a; Garrick et al., 2019). The Last Glacial Maximum (LGM) is considered reasonably representative of previous glaciations (Hewitt, 2011b). Hence, species distribution models for the LGM can provide coarse estimates of those areas that were able to sustain populations during the glacial periods of the Pleistocene and those that became suitable during interglacials (Svenning et al., 2011; Alvarado-Serrano & Knowles, 2014).

In Europe, the Mediterranean peninsulas (the Iberian, Italian and Balkan Peninsulas) are

© 2020 The Linnean Society of London, *Biological Journal of the Linnean Society*, 2020, **130**, 578–585 578 This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com considered the dominant refugial areas for European temperate species (Taberlet et al., 1998; Provan & Bennett, 2008; Hewitt, 2011b). Phylogeographical studies have also uncovered genetically distinct, geographically coherent populations within the Mediterranean peninsulas, suggesting intraspecific fragmentation and differentiation throughout the Pleistocene (Hewitt, 2011b). The presence of multiple, discrete glacial refugia existing in an area formerly recognized as a single continuous glacial refugium has been coined 'refugia-within-refugia' (Gómez & Lunt, 2007; Abellán & Svenning, 2014). Many species in the Balkan Peninsula show extensive mtDNA structuring (Poulakakis et al., 2015), which raises the question: to what extent is the geographical structure observed in mtDNA paralleled in the nuclear genome?

We applied next-generation phylogeography to the crested newt *Triturus macedonicus* (Karaman, 1922), a Balkan endemic (Fig. 1). This species shows a degree of intraspecific mtDNA differentiation unparalleled in the genus *Triturus*, in line with a long-term presence in the Balkan Peninsula and a complex history of genetic differentiation *in situ* (Wielstra *et al.*, 2013; Fig. 1). We here compare and evaluate the spatial correspondence of the geographical substructuring in the nuclear and mitochondrial genomes of *T. macedonicus*.

MATERIAL AND METHODS

SAMPLING AND SEQUENCE DATA

We included three individuals from each of 39 populations, i.e. 117 individuals in total (Fig. 1; Supporting Information, Table S1). We re-analysed data previously used in studies that focused on interspecific hybridization, rather than intraspecific phylogeography. Sequence data for an mtDNA marker (ND4, 658 bp), including representatives of the eight other Triturus species and the related newt genus Calotriton, were taken from Wielstra et al. (2013). Sequence data for 52 nuclear DNA markers were taken from Wielstra et al. (2017). To obtain these nuclear markers, we used the Ion Torrent nextgeneration sequencing protocol described by Wielstra et al. (2014a). In brief, we amplified markers of ~140 bp in length (excluding primers), positioned in 3' untranslated regions, in five multiplex polymerase chain reactions. We pooled the multiplexes for each individual and ligated unique tags to be able to recognize the product belonging to each individual. We sequenced the amplicons on the Ion Torrent nextgeneration sequencing platform and processed the output with a bioinformatics pipeline that filters out poor-quality reads, identifies alleles and converts data

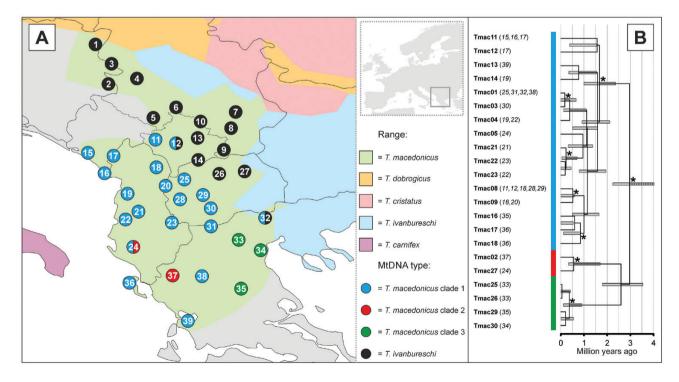


Figure 1. Distribution and sampling of *Triturus macedonicus*. In A, background colours reflect the range of *T. macedonicus* and neighbouring crested newt species. Populations are coloured according to the mitochondrial DNA (mtDNA) clade; those containing introgressed *Triturus ivanbureschi* mtDNA are shown in black. B, the calibrated mtDNA phylogeography. Asterisks denote nodes with a posterior probability ≥ 0.95 . Population numbers and mtDNA haplotype codes correspond to those in the Supporting Information (Tables S1, S2).

to a genotypic format that can easily be converted to other formats with the program CREATE (Coombs *et al.*, 2008). The total number of aligned Ion Torrent reads after data filtering was 6 293 299. The mean coverage was 791 reads (range 0–24 444) per marker– individual combination, and 98.9% of marker– individual combinations were considered successful (meaning that they had \geq 20 reads available).

MITOCHONDRIAL DNA RE-ANALYSIS

We obtained a dated phylogeny with BEAST v.2.1 (Bouckaert et al., 2014). A fossil dated at 24 Mya was used as a minimum estimate for the most recent common ancestor of the genus Triturus (Steinfartz et al., 2007) and given a lognormally distributed prior with an offset of 24 and a mean and standard deviation of 1.0. The origin of the Adriatic Sea at 5.33 Mya, at the end of the Messinian Salinity Crisis, was assumed to be the vicariant event causing the split between T. macedonicus and its sister species Triturus carnifex (Arntzen et al., 2007; Wielstra et al., 2019) and given a normally distributed prior with a small standard deviation (0.001). The most appropriate model of sequence evolution (GTR+G) was identified with jModelTest v.2.1.7 (Darriba et al., 2012), based on the Akaike information criterion. We applied the relaxed lognormal clock model and a Yule speciation model. We conducted two independent runs, each of 100 million generations, with a sampling frequency of 0.0001. The first half of the sampled trees was discarded as burn-in after analysis in TRACER v.1.6 (http://tree.bio.ed.ac. uk/software/tracer/); runs converged, and effective sample sizes were ≥ 200 .

BAYESIAN CLUSTERING ANALYSIS

Interspecific

Triturus macedonicus is parapatric with three additional *Triturus* species (Fig. 1). Considering that Triturus species hybridize at their contact zones (Arntzen et al., 2014), we took the following approach to counter potential adverse effects of interspecific gene flow on the recovery of population structure within T. macedonicus. We conducted Bayesian species assignment using STRUCTURE v.2.3.4 (Pritchard et al., 2000) and compared the T. macedonicus dataset with the three crested newt species with which it shares a contact zone: Triturus cristatus, Triturus dobrogicus and Triturus ivanbureschi (reference data; three individuals for four populations per species, taken from the study by Wielstra et al., 2014a). We fixed the number of distinct gene pools (k) at four, because we were dealing with four parental species. We used the admixture model in combination with the correlated allele frequency model with 100 000 iterations, after 50 000 iterations of burn-in, and ran ten replicates, which were summarized with CLUMPAK (Kopelman *et al.*, 2015). Individuals allocated to *T. macedonicus* with a support < 0.95 were excluded.

Intraspecific

To explore population structure within *T. macedonicus*, we conducted non-spatial Bayesian clustering analysis with STRUCTURE v.2.3.4 and spatially explicit Bayesian clustering analysis using BAPS v.6 (Cheng et al., 2013). The upper limit of the range over which the number of gene pools (k) was tested was defined by the total number of populations included in these datasets (35, because four populations were excluded based on interspecific gene flow). To select the optimal value of k for STRUCTURE, we used the Δk criterion (Evanno et al., 2005) as implemented in CLUMPAK (Kopelman et al., 2015). BAPS determines the optimal value of k internally. For each program, we performed five independent runs. In STRUCTURE, we used the admixture model in combination with the correlated allele frequency model with 100 000 iterations, after 50 000 iterations of burn-in. CLUMPAK was used to average the outputs of multiple replicates. In BAPS, we conducted a spatial genetic mixture analysis. Given that individuals within populations share the same coordinates, we allowed BAPS to make input coordinates unique by introducing minor random variation.

SPECIES DISTRIBUTION MODELLING

We used MAXENT v.3.3.3k (Phillips et al., 2006) to create a species distribution model, because this program was developed for presence-only data, outperforms other algorithms and can be used reliably to hindcast to past environments (Elith et al., 2006; Hijmans & Graham, 2006). The feature type was restricted to hinge features because this produces a smoother model fit, emphasizing trends rather than data idiosyncrasies (Elith et al., 2010). We used a database of 131 T. macedonicus localities presented by Wielstra et al. (2014b). For climate layers, we used bioclimatic variables available from the WorldClim database v.1.4 (Hijmans et al., 2005; http://www.worldclim.org) at 2.5 arc-min resolution $(\sim 5 \text{ km} \times 5 \text{ km})$. Following Peterson (2011) and Guisan & Thuiller (2005), we selected a subset that showed low multicollinearity (a Pearson's correlation of r < 0.7) and was considered to reflect the physiological limitations of the study species (in this case, seasonality: bio10 = mean temperature of warmest quarter; bio11 = mean temperature of coldest quarter; bio15 = precipitation seasonality; bio16 = precipitation of wettest quarter; and bio17 = precipitation of driest quarter). To prevent the environmental range covered by pseudo-absence data, used by MAXENT to discriminate presence data from the environmental background, being either too narrow or too broad (Elith et al., 2011), we drew a buffer of radius 200 km around known Triturus localities (following VanDerWal et al., 2009). We tested that our species distribution model performed better than random expectation by testing its area under the curve of the receiver operating characteristic value against a null model (Raes & ter Steege, 2007) based on 99 models for random point data created in ENMTools v.1.3 (Warren et al., 2010). For hindcasting to the LGM (~21 000 years ago), we used bioclimatic variables, based on downscaled climate data from simulations of two global climate models: CCSM4 (Brady et al., 2013) and MIROC-ESM (Suevoshi et al., 2013), available from WorldClim.

RESULTS

MITOCHONDRIAL DNA RE-ANALYSIS

Triturus macedonicus possesses three main mtDNA clades that split > 2.5 Mya (Fig. 1). Along the entire north-eastern fringe of its range, *T. macedonicus* has captured mtDNA derived from another crested newt species, *T. ivanbureschi*.

BAYESIAN CLUSTERING ANALYSIS

Interspecific

The comparison in STRUCTURE with the three parapatric *Triturus* species revealed signs of nuclear genetic admixture with *T. ivanbureschi* only, in 15 individuals from seven *T. macedonicus* populations

from near the contact zone (Fig. 2; Supporting Information, Table S1). In four populations, all individuals showed introgression, whereas in each of the other three only a single individual was affected.

Intraspecific

Results of the non-spatial and spatially explicit Bayesian population assignment analyses at the level of the population are presented in Figure 2 (details on the assignment of individuals are in Supporting Information, Table S1). The STRUCTURE analysis suggested k = 2 as the most likely number of nuclear DNA gene pools based on Evanno's Δk criterion, with a second smaller peak found for k = 4. A spatially explicit Bayesian clustering analysis in BAPS also considered k = 4 as the most likely number of gene pools. Given the tendency of STRUCTURE to converge on k = 2 under the Δk criterion (Janes *et al.*, 2017), we also explored population subdivision under k = 4. In all analyses, one 'north-eastern group' was found in the northeastern part of the range, and three additional groups found in BAPS and STRUCTURE under k = 4 fitted in a 'south-western group' recovered by STRUCTURE under k = 2. Results for BAPS were similar to those of STRUCTURE under k = 4, although less mingling between gene pools was suggested. There was little apparent agreement between the distribution of these nuclear DNA groups and the mtDNA clades (Figs 1, 2).

SPECIES DISTRIBUTION MODELLING

The species distribution model predicted that the western part of the current range of T. macedonicus was also suitable during the LGM (Fig. 3). Two

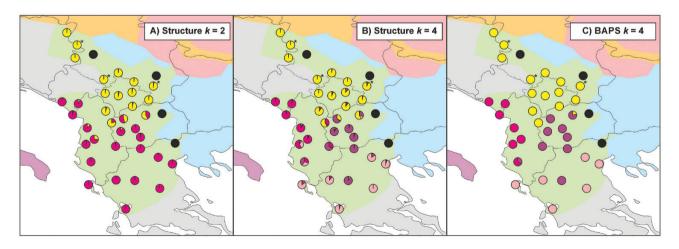


Figure 2. Bayesian structuring results for *Triturus macedonicus*. Pies reflect admixture proportions to genetic clusters at the level of the population for STRUCTURE under k=2 (A) and k=4 (B) and BAPS under k=4 (C). Populations affected by interspecific gene flow are marked with an asterisk if one individual was affected (and only the remaining two individuals were included) and coloured black if all individuals were affected (and the entire population was excluded).

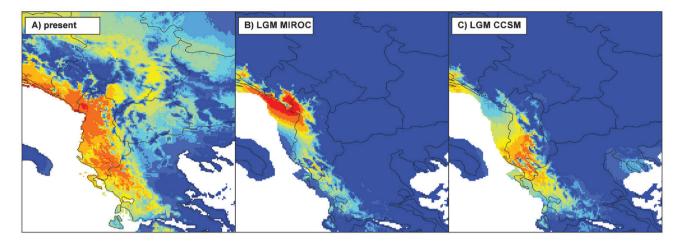


Figure 3. Species distribution model for *Triturus macedonicus*. The species distribution model is projected on climate layers for the present (A) and the Last Glacial Maximum based on the MIROC (B) and CCSM (C) climate models. Warmer colours indicate higher predicted suitability.

clades of species-specific mtDNA and all four nuclear DNA groups occur in this area today. There was no suggestion that the distribution range was fragmented at the LGM. The eastern part of the current range became suitable only after the LGM. This includes the area where mtDNA clade 3 occurs, where introgressed mtDNA is found and where the 'north-eastern nuclear DNA group' has its main distribution.

DISCUSSION

Although mtDNA has driven the field of phylogeography for more than two decades (Avise, 2000; Beheregaray, 2008), there is currently a shift taking place towards the use of large panels of unlinked nuclear markers (Garrick *et al.*, 2015). Next-generation phylogeography allows us to test, refine and adjust the biogeographical hypotheses derived from mtDNA (Spinks *et al.*, 2014; Dufresnes *et al.*, 2020). Here, we have explored intraspecific genetic structuring of mitochondrial and nuclear genomes within the Balkan Peninsula glacial refugium, using the crested newt *T. macedonicus* as a model (Wallis & Arntzen, 1989).

Mitochondrial DNA is a poor predictor for the contemporary population structure of *T. macedonicus* because its geographical structure (Fig. 1) is highly discordant to that of the nuclear DNA groups (Fig. 2). In the west of its range, *T. macedonicus* possesses three highly distinct mtDNA clades that appear to have radiated before the Pleistocene (< 2.58 Mya; green, red and blue in Fig. 1). In fact, *T. macedonicus* shows the deepest divergence in mtDNA of any *Triturus* species (Wielstra *et al.*, 2013). Mitochondrial DNA clade 1 (blue in Fig. 1) covers a large area and includes many distinct

haplotypes with unclear relationships, indicative of longterm demographic stability (Hewitt, 1999). In contrast, clades 2 (red) and 3 (green) contain few haplotypes and are localized. Clades 1 and 2 occur in areas consistently predicted to have been suitable during the LGM based on species distribution modelling (Fig. 3). Clade 3 occurs slightly further eastwards of this suitable area, suggesting that the LGM distribution by the model is at least somewhat underestimated, because clade 3 must have survived here locally.

The nuclear DNA in T. macedonicus is structured into four groups that show extensive genetic admixture in the STRUCTURE analysis (but not in BAPS), all occurring in the west of the T. macedonicus range (Fig. 2). The distribution of nuclear DNA groups has little bearing on the distribution of mtDNA clades; one nuclear DNA group contains all three mtDNA clades, and one mtDNA clade is found in three of the nuclear DNA groups (Figs 1, 2). In general, mtDNA appears to be relatively stationary in comparison to the average nuclear marker, resulting in stronger and more permanent geographical structuring (Petit & Excoffier, 2009; Toews & Brelsford, 2012). Hence, this mtDNA structuring could reflect ancient population subdivision, whereas the signal in the nuclear DNA was lost at a later point owing to genetic swamping upon secondary contact, resulting in the maintenance of mtDNA clades that are no longer reflected by nuclear DNA clusters. Several other Balkan newt species are also known to possess such mtDNA 'ghost lineages' (Recuero et al., 2014; Pabijan et al., 2015).

In most of its postglacially colonized range, as predicted by the species distribution model (Fig. 3), *T. macedonicus* has captured mtDNA derived from another crested newt species, *T. ivanbureschi* (Wielstra *et al.*, 2017; Fig. 1). Although all the populations that carry introgressed mtDNA also belong to the same nuclear DNA group, this nuclear DNA group additionally encompasses individuals from further west that are known not to possess any diagnostic *T. ivanbureschi* nuclear alleles (Wielstra *et al.*, 2017). Hence, rather than introgression of *T. ivanbureschi* nuclear alleles defining this nuclear DNA group, we would argue that strong genetic drift attributable to postglacial expansion from a small source unites these populations in the Bayesian clustering analyses. The source of this expansion would be northern Albania, right at the eastern edge of the area predicted to have harboured suitable conditions during the LGM (Figs 2, 3).

CONCLUSION

Based on several dozen nuclear DNA markers, we find that T. macedonicus comprises multiple geographical genetic groups. Although species distribution modelling suggests a reduced glacial range, it does not support range fragmentation. We find the same number of partitions based on mtDNA and nuclear DNA, but the two datasets show geographical discordance. This 'evolutionary melting pot' pattern (Dufresnes et al., 2016) suggests recurrent interglacial mingling of populations that have been diverging gradually during the glacial periods (Garrick et al., 2019). For T. macedonicus, hybridization upon secondary contact even resulted in the capture of mtDNA from a different crested newt species (Wielstra et al., 2017). Our genetic data are in line with a refugia-within-refugia scenario (Abellán & Svenning, 2014). Considering a large number of taxa with deep mtDNA structuring previously identified in the Balkan Peninsula (Poulakakis et al., 2015), the generality of this scenario can be tested in future next-generation phylogeographical studies (Arbogast & Kenagy, 2001).

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REFERENCES

- Abellán P, Svenning J-C. 2014. Refugia within refugia patterns in endemism and genetic divergence are linked to Late Quaternary climate stability in the Iberian Peninsula. *Biological Journal of the Linnean Society* **113**: 13–28.
- Alvarado-Serrano DF, Knowles LL. 2014. Ecological niche models in phylogeographic studies: applications, advances, and precautions. *Molecular Ecology Resources* 14: 233–248.

- Arbogast BS, Kenagy GJ. 2001. Comparative phylogeography as an integrative approach to historical biogeography. *Journal of Biogeography* 28: 819–825.
- Arntzen JW, Espregueira Themudo G, Wielstra B. 2007. The phylogeny of crested newts (*Triturus cristatus* superspecies): nuclear and mitochondrial genetic characters suggest a hard polytomy, in line with the paleogeography of the centre of origin. *Contributions to Zoology* 76: 261–278.
- Arntzen JW, Wielstra B, Wallis GP. 2014. The modality of nine *Triturus* newt hybrid zones, assessed with nuclear, mitochondrial and morphological data. *Biological Journal of the Linnean Society* 113: 604–622.
- Avise JC. 2000. *Phylogeography: the history and formation of species*. Cambridge: Harvard University Press.
- **Beheregaray LB. 2008.** Twenty years of phylogeography: the state of the field and the challenges for the Southern Hemisphere. *Molecular Ecology* **17:** 3754–3774.
- Bennett KD, Provan J. 2008. What do we mean by 'refugia'? Quaternary Science Reviews 27: 2449–2455.
- Bouckaert R, Heled J, Kühnert D, Vaughan T, Wu C-H, Xie D, Suchard MA, Rambaut A, Drummond AJ. 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. *PLoS Computational Biology* **10**: e1003537.
- Brady EC, Otto-Bliesner BL, Kay JE, Rosenbloom N. 2013. Sensitivity to glacial forcing in the CCSM4. *Journal of Climate* 26: 1901–1925.
- Cheng L, Connor TR, Sirén J, Aanensen DM, Corander J. 2013. Hierarchical and spatially explicit clustering of DNA sequences with BAPS software. *Molecular Biology and Evolution* 30: 1224–1228.
- Coombs JA, Letcher BH, Nislow KH. 2008. CREATE: a software to create input files from diploid genotypic data for 52 genetic software programs. *Molecular Ecology Resources* 8: 578–580.
- **Darriba D, Taboada GL, Doallo R, Posada D. 2012.** jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods* **9:** 772.
- Dufresnes C, Litvinchuk SN, Leuenberger J, Ghali K, Zinenko O, Stöck M, Perrin N. 2016. Evolutionary melting pots: a biodiversity hotspot shaped by ring diversifications around the Black Sea in the Eastern tree frog (*Hyla orientalis*). *Molecular Ecology* **25**: 4285–4300.
- Dufresnes C, Nicieza AG, Litvinchuk SN, Rodrigues N, Jeffries DL, Vences M, Perrin N, Martínez-Solano Í. 2020. Are glacial refugia hotspots of speciation and cytonuclear discordances? Answers from the genomic phylogeography of Spanish common frogs. *Molecular Ecology* 29: 986–1000.
- **Ekblom R**, **Galindo J. 2011.** Applications of next generation sequencing in molecular ecology of non-model organisms. *Heredity* **107:** 1–15.
- Elith J, Graham CH, Anderson RP, Dudík M, Ferrier S, Guisan A, Hijmans RJ, Huettmann F, Leathwick JR, Lehmann A, Li J, Lohmann LG, Loiselle BA, Manion G, Moritz C, Nakamura M, Nakazawa Y, Overton JMcCM, Townsend Peterson A, Phillips SJ, Richardson K, Scachetti-Pereira R, Schapire RE, Soberón J, Williams S, Wisz MS, Zimmermann NE. 2006. Novel

methods improve prediction of species' distributions from occurrence data. *Ecography* **29:** 129–151.

- Elith J, Kearney M, Phillips S. 2010. The art of modelling range-shifting species. *Methods in Ecology and Evolution* 1: 330–342.
- Elith J, Phillips SJ, Hastie T, Dudík M, Chee YE, Yates CJ. 2011. A statistical explanation of MaxEnt for ecologists. *Diversity and Distributions* 17: 43–57.
- **Evanno G**, **Regnaut S**, **Goudet J. 2005.** Detecting the number of clusters of individuals using the software structure: a simulation study. *Molecular Ecology* **14**: 2611–2620.
- Garrick RC, Banusiewicz JD, Burgess S, Hyseni C, Symula RE. 2019. Extending phylogeography to account for lineage fusion. *Journal of Biogeography* 46: 268–278.
- Garrick RC, Bonatelli IAS, Hyseni C, Morales A, Pelletier TA, Perez MF, Rice E, Satler JD, Symula RE, Thomé MTC, Carstens BC. 2015. The evolution of phylogeographic data sets. *Molecular Ecology* 24: 1164–1171.
- Gómez A, Lunt DH. 2007. Refugia within refugia: patterns of phylogeographic concordance in the Iberian Peninsula. In: Weiss S, Ferrand N, eds. *Phylogeography of Southern European refugia*. Dordrecht: Springer, 155–188.
- Guisan A, Thuiller W. 2005. Predicting species distribution: offering more than simple habitat models. *Ecology Letters* 8: 993–1009.
- Hewitt G. 2000. The genetic legacy of the Quaternary ice ages. Nature 405: 907–913.
- Hewitt GM. 1999. Post-glacial re-colonization of European biota. *Biological Journal of the Linnean Society* 68: 87–112.
- **Hewitt GM. 2004.** The structure of biodiversity insights from molecular phylogeography. *Frontiers in Zoology* **1:** 4.
- Hewitt GM. 2011a. Quaternary phylogeography: the roots of hybrid zones. *Genetica* 139: 617–638.
- Hewitt GM. 2011b. Mediterranean peninsulas: the evolution of hotspots. In: Zachos FE, Habel JC, eds. *Biodiversity hotspots: distribution and protection of conservation priority areas*. Berlin: Springer, 123–147.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. 2005. Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology* 25: 1965–1978.
- Hijmans RJ, Graham CH. 2006. The ability of climate envelope models to predict the effect of climate change on species distributions. *Global Change Biology* 12: 2272–2281.
- Janes JK, Miller JM, Dupuis JR, Malenfant RM, Gorrell JC, Cullingham CI, Andrew RL. 2017. The K= 2 conundrum. *Molecular Ecology* 26: 3594–3602.
- Kopelman NM, Mayzel J, Jakobsson M, Rosenberg NA, Mayrose I. 2015. CLUMPAK: a program for identifying clustering modes and packaging population structure inferences across K. Molecular Ecology Resources 15: 1179–1191.
- McCormack JE, Hird SM, Zellmer AJ, Carstens BC, Brumfield RT. 2013. Applications of next-generation sequencing to phylogeography and phylogenetics. *Molecular Phylogenetics and Evolution* **66**: 526–538.
- Pabijan M, Zieliński P, Dudek K, Chloupek M, Sotiropoulos K, Liana M, Babik W. 2015. The dissection of

a Pleistocene refugium: phylogeography of the smooth newt, *Lissotriton vulgaris*, in the Balkans. *Journal of Biogeography* **42:** 671–683.

- Peterson AT. 2011. Ecological niche conservatism: a timestructured review of evidence. *Journal of Biogeography* 38: 817–827.
- Petit RJ, Excoffier L. 2009. Gene flow and species delimitation. *Trends in Ecology & Evolution* 24: 386–393.
- Phillips SJ, Anderson RP, Schapire RE. 2006. Maximum entropy modeling of species geographic distributions. *Ecological Modelling* 190: 231–259.
- Poulakakis N, Kapli P, Lymberakis P, Trichas A, Vardinoyiannis K, Sfenthourakis S, Mylonas M. 2015. A review of phylogeographic analyses of animal taxa from the Aegean and surrounding regions. Journal of Zoological Systematics and Evolutionary Research 53: 18–32.
- Pritchard JK, Stephens M, Donnelly P. 2000. Inference of population structure using multilocus genotype data. *Genetics* 155: 945–959.
- Provan J, Bennett KD. 2008. Phylogeographic insights into cryptic glacial refugia. *Trends in Ecology & Evolution* 23: 564–571.
- **Puritz JB**, Addison JA, Toonen RJ. 2012. Next-generation phylogeography: a targeted approach for multilocus sequencing of non-model organisms. *PLoS ONE* 7: e34241.
- Raes N, ter Steege H. 2007. A null-model for significance testing of presence-only species distribution models. *Ecography* 30: 727–736.
- Recuero E, Buckley D, García-París M, Arntzen JW, Cogălniceanu D, Martínez-Solano I. 2014. Evolutionary history of *Ichthyosaura alpestris* (Caudata, Salamandridae) inferred from the combined analysis of nuclear and mitochondrial markers. *Molecular Phylogenetics and Evolution* 81: 207–220.
- Spinks PQ, Thomson RC, Shaffer HB. 2014. The advantages of going large: genome-wide SNPs clarify the complex population history and systematics of the threatened western pond turtle. *Molecular Ecology* 23: 2228–2241.
- Steinfartz S, Vicario S, Arntzen JW, Caccone A. 2007. A Bayesian approach on molecules and behavior: reconsidering phylogenetic and evolutionary patterns of the Salamandridae with emphasis on *Triturus* newts. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution* **308B:** 139–162.
- Sueyoshi T, Ohgaito R, Yamamoto A, Chikamoto MO, Hajima T, Okajima H, Yoshimori M, Abe M, O'Ishi R, Saito F, Watanabe S, Kawamiya M, Abe-Ouchi A. 2013. Set-up of the PMIP3 paleoclimate experiments conducted using an Earth system model, MIROC-ESM. Geoscientific Model Development 6: 819–836.
- Svenning J-C, Fløjgaard C, Marske KA, Nógues-Bravo D, Normand S. 2011. Applications of species distribution modeling to paleobiology. *Quaternary Science Reviews* 30: 2930–2947.
- Taberlet P, Fumagalli L, Wust-Saucy A-G, Cosson J-F. 1998. Comparative phylogeography and postglacial colonization routes in Europe. *Molecular Ecology* 7: 453-464.

- Toews DPL, Brelsford A. 2012. The biogeography of mitochondrial and nuclear discordance in animals. *Molecular Ecology* 21: 3907–3930.
- VanDerWal J, Shoo LP, Graham C, Williams SE. 2009. Selecting pseudo-absence data for presence-only distribution modeling: how far should you stray from what you know? *Ecological Modelling* 220: 589–594.
- Wallis GP, Arntzen JW. 1989. Mitochondrial-DNA variation in the crested newt superspecies: limited cytoplasmic gene flow among species. *Evolution* 43: 88–104.
- Warren DL, Glor RE, Turelli M. 2010. ENMTools: a toolbox for comparative studies of environmental niche models. *Ecography* 33: 607–611.
- Wielstra B, Arntzen JW. 2020. Data from: Extensive cytonuclear discordance in a crested newt from the Balkan Peninsula glacial refugium. *Dryad Digital Repository*. doi:10.5061/dryad.xgxd254d0
- Wielstra B, Burke T, Butlin RK, Arntzen JW. 2017. A signature of dynamic biogeography: enclaves indicate past species replacement. *Proceedings of the Royal Society B: Biological Sciences* 284: 20172014.

- Wielstra B, Crnobrnja-Isailović J, Litvinchuk SN, Reijnen BT, Skidmore AK, Sotiropoulis K, Toxopeus AG, Tzankov N, Vukov T, Arntzen JW. 2013. Tracing glacial refugia of *Triturus* newts based on mitochondrial DNA phylogeography and species distribution modeling. *Frontiers in Zoology* **10**: 13.
- Wielstra B, Duijm E, Lagler P, Lammers Y, Meilink WRM, Ziermann JM, Arntzen JW. 2014a. Parallel tagged amplicon sequencing of transcriptome-based genetic markers for *Triturus* newts with the Ion Torrent next-generation sequencing platform. *Molecular Ecology Resources* 14: 1080–1089.
- Wielstra B, McCartney-Melstad E, Arntzen JW, Butlin RK, Shaffer HB. 2019. Phylogenomics of the adaptive radiation of *Triturus* newts supports gradual ecological niche expansion towards an incrementally aquatic lifestyle. *Molecular Phylogenetics and Evolution* 133: 120–127.
- Wielstra B, Sillero N, Vörös J, Arntzen JW. 2014b. The distribution of the crested and marbled newt species (Amphibia: Salamandridae: *Triturus*) an addition to the New Atlas of Amphibians and Reptiles of Europe. *Amphibia*-*Reptilia* 35: 376–381.

SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

Table S1. Sampling details and Bayesian population assignment for *Triturus macedonicus*.**Table S2.** GenBank accession numbers for the mitochondrial DNA haplotypes identified in *Triturus macedonicus*.

SHARED DATA

Sequence data and files associated with analyses are available from the Dryad Digital Repository: Wielstra & Arntzen (2020).