## Identification of Germline Variants in Tumor Genomic Sequencing Analysis

## To the Editor-in-Chief:

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Recently, the Association for Molecular Pathology (AMP), the American Society of Clinical Oncology (ASCO), and the College of American Pathologists (CAP) published consensus guidelines in *The Journal of Molecular Diagnostics* to standardize variant interpretation and reporting in oncology specimens.<sup>1</sup> This group's recommendations included guidance related to a dilemma commonly encountered in clinical practice—accurately distinguishing somatic mutations that arose in a patient's tumor from inherited germline variants.

Distinguishing somatic from germline variants is important for several reasons. First, most germline sequence variants will be clinically benign, and these should not be included in clinical reports of actionable somatic driver mutations. Second, however, some rare germline variants are pathogenic and may be associated with an inherited disorder, including familial cancer predisposition. It is important to recognize these variants as inherited to ensure appropriate management not only of the patient but also of family members. Moreover, even when germline variants are not associated with a high-penetrance cancer susceptibility syndrome, they may be functionally and even clinically significant. Finally, accurately recognizing that a sequence variant is of somatic origin may provide evidence of a clonal process, which can be diagnostically important in some neoplastic conditions, such as myeloid neoplasia.<sup>2</sup>

The task of distinguishing somatic and germline variants is made more difficult in tumor-only sequencing protocols. These workflows involve identification of mutations in neoplastic tissue without comparison to a normal sample from the patient. Many clinical laboratories have adopted such protocols for practical reasons and/or to avoid the substantial cost of sequencing paired normal specimens. However, tumor-only workflows carry some risk of misclassifying somatic and germline variants.

The ASCO/AMP/CAP guidelines make several recommendations to assist recognition of germline variants in tumor-only protocols.<sup>1</sup> Chief among these, the guidelines indicate that the main criterion for germline designation is variant allele fraction (VAF), which the authors note is expected to be near 50% or 100% for heterozygous and homozygous germline alleles, respectively, that are present at diploid copy number in all cells in a specimen. However, performance metrics of this recommendation have not been established. Moreover, as the authors of the guidelines acknowledge, there are certain limitations to using VAF as a screen for germline variants. For instance, somatic mutations may also be present near 50% or 100% VAF in tissue with high tumor burden, and germline variants may deviate from 50% or 100% VAF because of chromosomal aneuploidy or other changes in gene copy number, both of which are common in tumors.

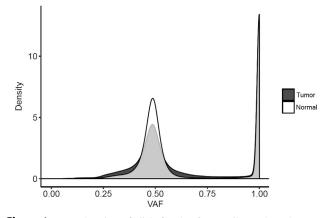
We examined the usefulness of the AMP/ASCO/CAP guideline for identification of germline variants in a cohort of 1310 cancer patients with paired sequencing of >200 genes in both a tumor and a normal sample (peripheral blood or tissue) as part of a large next-generation sequencing research program at our institution (LCCC1108/UNCSeq, NCT01457196).<sup>3</sup> A diverse group of neoplasms was represented in this cohort. Common sites of tumor origin included gynecologic tract (n = 402), breast (n = 212), head and neck (n = 203), gastrointestinal tract organs (n = 115), nervous system/brain (n = 95),

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**Figure 1** Density plots of allele fraction for germline variants in tumor and paired normal samples. Density plots are shown for the observed allele fraction of germline variants measured in tumor and paired normal samples from 1310 subjects. **Light gray regions** represent the area encompassed under both curves. For this analysis, variants detected in the paired normal sample were defined as germline variants. However, to be included in the study, variants were required to have a read depth of  $\geq 100 \times$  in both tumor and normal samples. Approximately 6.8 million variants met these inclusion criteria. Somatic variants present only in the tumor sample are not included in this plot. VAF, variant allele fraction.

genitourinary system (n = 94), hematologic/lymphoid (n = 57), soft tissue (n = 46), skin (n = 30), and lung (n = 28).

Analysis was restricted to regions with  $100\times$  or deeper coverage in both the tumor and normal samples. Within these regions, a total of 6.8 million variants were identified in the normal samples as candidate germline variants. Density plots of observed VAFs for these variants are shown in Figure 1. As expected, measured VAF of germline variants in normal tissue showed a bimodal distribution, with peaks centered at 50% and 100%. Although a bimodal pattern was also observed for germline variants in tumor samples, the Gaussian distribution was broader around the 50% VAF peak, indicating a wider range of VAFs for these variants within the tumor samples (Figure 1).

To further explore this observation, VAFs of variants detected in normal samples were plotted against VAFs of the same variants in tumor samples (tumor VAF) (Figure 2). As expected, germline variants were densest in both normal and tumor VAFs at around 50% and 100%. In the tumor sample, 46% of all germline variants were present at VAFs between 40% and 60%, as expected for a heterozygous allele, and 35% of all germline variants were present at a VAF of >95%. However, 19% of candidate germline variants exhibited a VAF in the tumor sample outside of the ranges suggested by the AMP/ASCO/CAP guidelines. This deviation was particularly striking for heterozygous germline variants, many of which showed considerable scatter away from 50% VAF in the tumor sample (Figure 2). Specifically, 22% of germline variants that were detected at a VAF between 40% and 60% in the normal sample (representing heterozygous alleles) had a

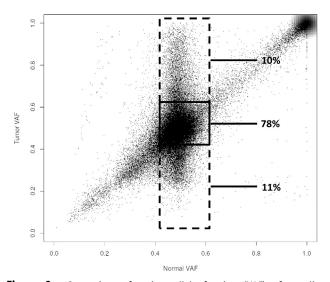
VAF outside of this range in the paired tumor samples (10% with a VAF of >60% and 11% with a VAF of <40%) (Figure 2).

Next, the percentage of all variants with a tumor VAF between 40% and 60% that were germline versus somatic in origin was determined. Of a total of 3.1 million variants with a tumor VAF between 40% and 60%, most (96%) were confirmed as germline by presence in the normal sample (data not shown).

Our observations indicate that the VAF of a substantial percentage of germline variants will deviate markedly from 50% or 100% in tumor samples. Although performance metrics are likely to be affected by several variables, including tumor type and tumor percentage, these data provide an informative first estimate of the sensitivity and specificity of the AMP/ASCO/CAP guideline for identification of germline variants in a cohort of patients with diverse tumors. As such, our observations have important implications for variant interpretation in tumor-only platforms.

In our cohort, VAF discordance between normal and tumor samples was most prominent for heterozygous alleles. In a subset of these cases, the finding of a VAF of <40% or >60% in tumor may indicate loss of heterozygosity of a pathogenic germline variant, a possibility worthy of future study. However, given that such discordance was common in our cohort, it may be more often the case that changes in VAF in tumor are passenger effects of an unstable genome.

Ultimately, there is no true substitute to tumor-normal pairs in the accurate distinction of germline from somatic variants. However, given persistent reimbursement issues in this space,<sup>4</sup> the additional expense of paired normal



**Figure 2** Comparison of variant allele fraction (VAF) of germline variants in tumor and paired normal samples. Observed VAF for germline variants in normal samples (*x* axis) and tumor samples (*y* axis). Results from a random sample of 100,000 variants (of 6.8 million total variants) are plotted for visualization. Among variants at 40% to 60% VAF in the normal sample, 78% are also detected at 40% to 60% VAF in tumor (**boxed area**), 10% were detected at >60% VAF in tumor (**top dashed boxed area**), and 11% were detected at <40% VAF in tumor (**bottom dashed boxed area**).

sequencing may prove cost prohibitive for many laboratories. Although the AMP/ASCO/CAP recommendations provide helpful guidance to such laboratories, reporting should emphasize the inherent inability of definitively distinguishing germline and somatic variants in tumor-only sequencing assays. If suspicion for a germline-inherited variant is present, follow-up testing of a normal sample should be recommended.

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