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# Biodegradable, flexible silicon nanomembrane-based NO<sub>x</sub> gas sensor system with record-high performance for transient environmental monitors and medical implants

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## Abstract

A novel transient electronics technology that is capable of completely dissolving or decomposing in certain conditions after a period of operation offers unprecedented opportunities for medical implants, environmental sensors, and other applications. Here, we describe a biodegradable, flexible silicon-based electronic system that detects NO species with a record-breaking sensitivity of 136 Rs (5 ppm, NO<sub>2</sub>) and 100-fold selectivity for NO species over other substances with a fast response (~30 s) and recovery (~60 s). The exceptional features primarily depend on not only materials, dimensions, and design layouts but also temperatures and electrical operations. Large-scale sensor arrays in a mechanically pliable configuration exhibit negligible deterioration in performance under various modes of applied loads, consistent with mechanics modeling. In vitro evaluations demonstrate the capability and stability of integrated NO<sub>x</sub> devices in severe wet environments for biomedical applications.

## Introduction

Electronic materials and devices that are biologically benign or mechanically elastic have been utilized in versatile applications, including electronic skin (E-skin) and biomedicine, for the diagnosis and treatment of diseases due to their advantages of nontoxicity, bioresorption, and mechanical properties similar to those of human skin and organs. Published examples involve sweat-based analysis<sup>1–3</sup>, epidermal virtual reality<sup>4,5</sup>, self-healing electronics<sup>6,7</sup>, optoelectronic elements<sup>8,9</sup>, and neurorelated systems<sup>10–13</sup>. Although the technological advances in the corresponding areas delivered innovative output products,

a relatively unexplored research opportunity exists in detectable species indicative of occurrences in organisms that may affect the prevalence and severity of human health, such as infections, injuries, diseases, or environmental exposures. One of the key signaling elements is nitric oxide (NO), which is produced naturally in the human body and plays an essential role in many aspects of maintaining health by relaxing or widening blood vessels to enhance blood flow, allowing nutrients and oxygen to travel to every part of the human body<sup>14,15</sup>. In non-biological cases, highly reactive nitric oxide can be transformed into nitrogen dioxide (NO<sub>2</sub>) with other participants, contributing to acid rain deposition and harmful ozone generation<sup>16,17</sup>. Research approaches for NO species in previous works have explored high-performance channel materials such as graphene, carbon nanotubes, polymers, and metal oxides<sup>18–20</sup> or modifications to these structures for a high surface-to-volume ratio<sup>21–25</sup>, while several issues, including rigidity, low sensitivity, poor

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selectivity, high power consumption, and nonbiodegradability, still need to be considered for wearable and implantable electronic systems. Here, we report a flexible and bioresorbable single-crystal silicon nanomembrane (SC-Si NM)-based  $\text{NO}_x$  sensing system operating at room temperature with exceptional sensitivity and selectivity. The results focus on detailed studies of electrical responses of Si NMs under various conditions, and of the mechanical properties of the electronic system with theoretical considerations. In vitro assessments that incorporate semipermeable membranes provide information on their potential applicability in disposable environmental monitors and bioresorbable medical implants.

## Experimental procedure

### Fabrication of a transient $\text{NO}_x$ ( $\text{NO}/\text{NO}_2$ )-sensing system on various substrates

Device fabrication began with thinning down SC-Si NMs on silicon-on-insulator (SOI, SOITEC, France) wafers (top silicon  $\sim 300$  nm in thickness). Repeated oxidation of SC-Si NMs at an elevated temperature ( $1100^\circ\text{C}$ ) and removal of the thermally grown  $\text{SiO}_2$  by wet etching with hydrofluoric acid (HF, 49%, J. T Baker, USA), allowed to reach a desired target thickness of Si NMs ( $\sim 100$  nm). Phosphorus doping at  $950^\circ\text{C}$  with spin-on dopant (SOD, Filmtronics, USA) formed highly doped regions for electrical contacts of a  $\text{NO}_x$  sensing system. Undercut wet etching of box oxide released the thinned SC-Si NMs from the carrier wafers, enabling transfer printing onto a PMMA/diluted PI-coated temporary substrate. Active areas of the SC-Si NMs for  $\text{NO}_x$  gas, humidity, and temperature sensors were defined by sulfur hexafluoride ( $\text{SF}_6$ )-based reactive ion etching, followed by deposition of interdigitated electrodes (IDEs) with a layer of magnesium (Mg,  $\sim 300$  nm) using an electron-beam (e-beam) evaporator (VER5004, SNTEK, Korea). Formations of patterned dielectric layers and contact pads were generated by wet etching using buffered oxide etchant (BOE, 6:1, J. T Baker, USA) and e-beam evaporation. A geometry of mesh-typed bridges was employed to facilitate lifting off the device, enabling the release of the system from the temporary substrate. After removal of the bottom PI layer, the device was transfer printed onto degradable substrates for a complete transient electronic system.

### Characterization of soft, biodegradable $\text{NO}_x$ sensors

The electrochemical properties of a silicon-based gas detector were measured using an electrometer (Keithley 2636B) under a DC bias voltage of 0.01 V. Experiments were conducted in a temperature-adjustable chamber (RT  $\sim 50^\circ\text{C}$ , Lindberg, USA) with a constant flow rate of 1000 sccm controlled by mass flow controllers (MFCs). Electrical responses of sensors were determined by measurements of baseline resistances in dry  $\text{N}_2/\text{air}$  and

saturated resistances after exposure to target gases for 500 s. The responses to reducing and oxidizing gases were defined as  $R_0/R_{\text{gas}}$  and  $R_{\text{gas}}/R_0$ , where  $R_{\text{gas}}$  and  $R_0$  denote the resistances of the target gases and dry  $\text{N}_2/\text{air}$ , respectively.

### Cell culture and M1 activation

RAW 264.7 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 100 U/ml penicillin, and 100  $\mu\text{g}/\text{ml}$  streptomycin. Cells were incubated at  $37^\circ\text{C}$  with 5%  $\text{CO}_2$  in a humidified incubator. To induce M1 activation, 100 ng/ml LPS (lipopolysaccharide, Sigma) and 20 ng/ml IFN- $\gamma$  (interferon-gamma, Peprotech) were added to the culture medium, and it was incubated for 24 h.

### DAF-FM assay for NO measurement

After M1 activation, cells were incubated in L-arginine-free DMEM (Thermo, 88364) for 30 min, collected using a cell scraper and replated in 48-well plates. To measure NO production in cells, 10  $\mu\text{M}$  DAF-FM DA (Sigma, D1946) was added to medium with or without 150  $\mu\text{M}$  L-arginine, and the system was incubated for 30 min. The fluorescence intensity of DAF-FM-stained cells was measured using a Cytation 3 Cell Imaging Multi-Mode Reader (Bio-Tek).

### Immunofluorescence microscopy

Cells were fixed using 4% paraformaldehyde for 10 min at  $4^\circ\text{C}$  and permeabilized with 0.05% Triton X-100 for 5 min. After blocking with FBS (10%, v/v)-supplemented PBS for 30 min, cells were incubated with anti-iNOS antibody (1:500, Abcam, ab178945) for 60 min. Cells were then incubated with secondary antibodies (1:500, Bethyl, A120-100D4) for 60 min in the dark. The nucleus and F-actin were stained with DAPI (Invitrogen) and Alexa Fluor phalloidin (1:50, Invitrogen, A12379), respectively. The fluorescence confocal images were captured using confocal laser microscopy (A1R, Nikon) through a  $\times 60$  oil immersion objective (N.A. 1.4).

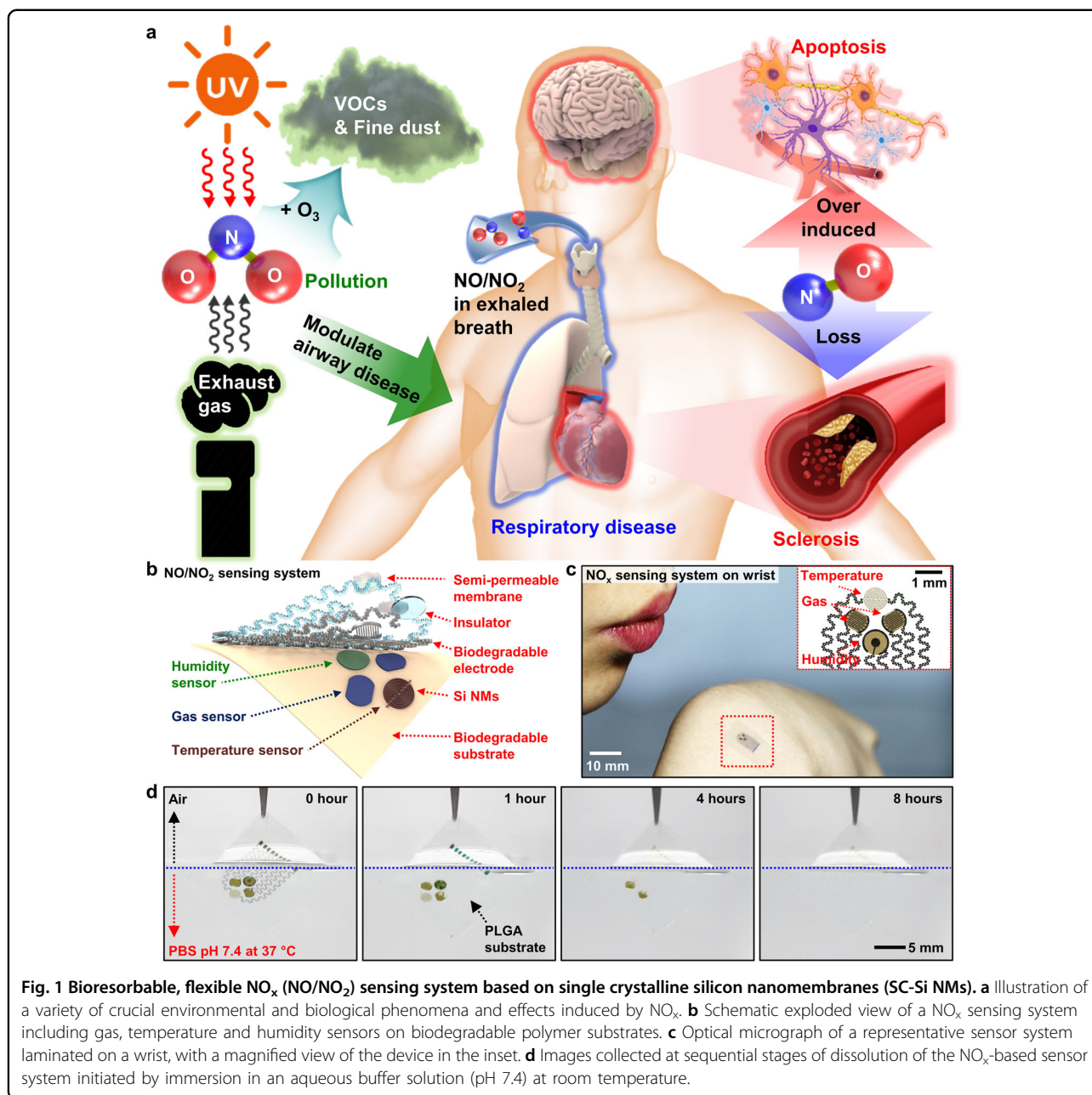
### Detection of $\text{NO}_x$ produced in M1-activated cells

M1-activated cells were transferred into a sterile cell-culture flask containing 2 ml L-arginine-free DMEM. NO sensors were placed into the flask, and L-arginine was provided through needles embedded in stoppers. The NO sensors were connected with ACF cables to an electrometer, and all measurements were recorded through LabVIEW with the GPIB interface.

## Results and discussion

### Integrated bioresorbable, flexible $\text{NO}_x$ ( $\text{NO}/\text{NO}_2$ ) sensing system

Figure 1a illustrates examples of various adverse environmental and biological effects that may be caused by



**Fig. 1** Biosorbable, flexible  $\text{NO}_x$  ( $\text{NO}/\text{NO}_2$ ) sensing system based on single crystalline silicon nanomembranes (SC-Si NMs). **a** Illustration of a variety of crucial environmental and biological phenomena and effects induced by  $\text{NO}_x$ . **b** Schematic exploded view of a  $\text{NO}_x$  sensing system including gas, temperature and humidity sensors on biodegradable polymer substrates. **c** Optical micrograph of a representative sensor system laminated on a wrist, with a magnified view of the device in the inset. **d** Images collected at sequential stages of dissolution of the  $\text{NO}_x$ -based sensor system initiated by immersion in an aqueous buffer solution (pH 7.4) at room temperature.

nitrogen oxides ( $\text{NO}_x$ ). NO has been well recognized as a biomarker for nervous, vascular, and respiratory systems<sup>26–28</sup>, and a major air pollutant giving rise to acid rain and ozone generation after transformation into  $\text{NO}_2$  as a reaction intermediate. Figure 1b provides an exploded view drawing of a soft, biosorbable  $\text{NO}_x$  sensing system that includes highly sensitive and selective silicon nanomembrane (Si NM)-based gas sensors at room temperature, and a temperature and hydration monitor to adjust the properties of the gas sensors affected by those parameters. The electronic system consists of patterned single-crystal silicon nanomembranes (SC-Si NMs,

thickness  $\sim 100$  nm) doped with phosphorous using spin-on dopant (SOD, Filmtronics) for reactive materials, magnesium (Mg, thickness  $\sim 300$  nm) as electrodes, silicon dioxide ( $\text{SiO}_2$ , thickness  $\sim 100$  nm) as interlayer dielectric, and an elastomeric polymer (PDMS, thickness  $\sim 20$   $\mu\text{m}$ ) as a semipermeable membrane to maintain a stable gas sensing performance in humid and wet conditions. Sample preparation involves thinning of SC-Si NMs to a targeted value ( $\sim 100$  nm), doping with phosphorous at selected regions on silicon-on-insulator (SOI) wafers and transfer printing doped SC-Si NMs to a temporary foreign substrate. Electrodes and insulators were evaporated and

patterned through deposition, photolithography and etching, and transfer printing of the resulting components to a biodegradable substrate completed the whole process. In the case of measurements in humid or wet conditions, facile spin casting of PDMS was applied to the obtained sample to form a gas-permeable membrane. The bioresorbable, flexible NO<sub>x</sub> sensing system laminated on a wrist appears in Fig. 1c, with a magnified image of the individual gas, temperature, and humidity sensors in the inset (more images in Fig. S1). Figure 1d shows a set of images of the temporal degradation behavior of a representative NO<sub>x</sub> sensing system assembled on a poly(lactic-co-glycolic) acid (PLGA) substrate during half-immersion in a PBS (pH 7.4) at 37 °C for 0, 1, 4, and 8 h. All constituent materials dissolved via hydrolysis to form non-toxic end products. Mg electrodes, with a relatively rapid dissolution rate, disappeared faster than other components via the reaction  $\text{Mg} + 2\text{H}_2\text{O} \rightarrow \text{Mg}(\text{OH})_2 + \text{H}_2$ , and SC-Si NMs, SiO<sub>2</sub>, and PLGA gradually dissolved at rates similar to previously reported values while undergoing the reactions  $\text{Si} + 4\text{H}_2\text{O} \leftrightarrow \text{Si}(\text{OH})_4 + 2\text{H}_2$ ,  $\text{SiO}_2 + 2\text{H}_2\text{O} \rightarrow \text{Si}(\text{OH})_4$ , and poly(lactic-co-glycolic acid)  $\rightarrow$  lactic acid + glycolic acid + H<sub>2</sub><sup>29–31</sup>. Additional dissolution images at different stages appear in Fig. S2.

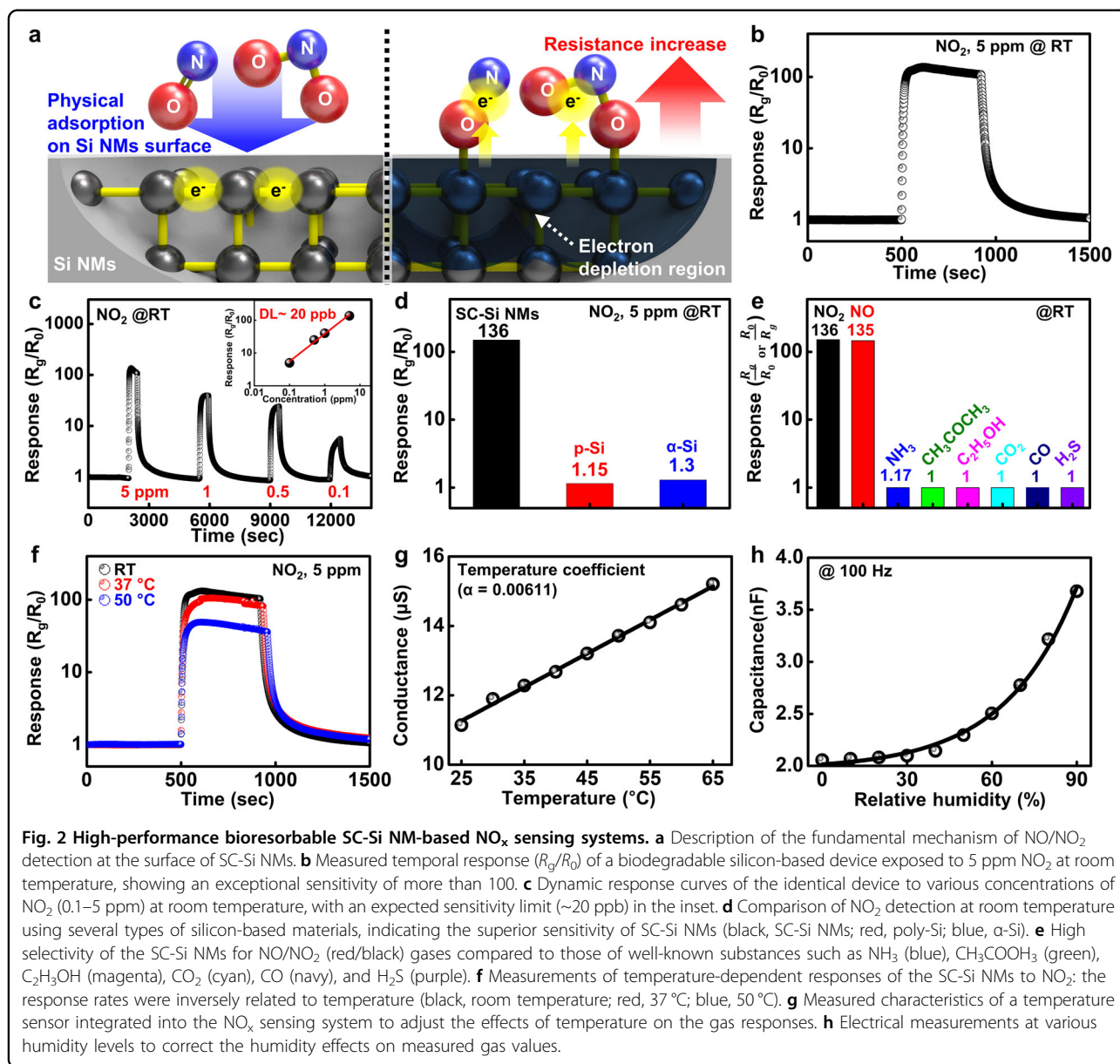
#### High-performance biodegradable NO<sub>x</sub> sensing system based on silicon nanomembranes

Figure 2a illustrates the fundamental mechanism of SC-Si NMs reacting with nitrogen oxides. Changes in the electrical properties of *n*-type semiconductor sensors resulted from variations in a depletion layer on the surface through adsorption and reaction with the oxidizing gas molecules, i.e., electrons were extracted from the SC-Si NMs as the reaction  $\text{NO}_x(\text{gas}) + e^- \rightarrow \text{NO}_x^-(\text{ads})$  occurred. Figure 2b represents a measured response to 5 ppm of NO<sub>2</sub> at room temperature (RT) with a superb sensitivity of 136 Rs (13,600%) as well as fast response (30 s) and recovery (60 s) rates, which surpass those of previously reported room-temperature sensors based on various materials, including carbon nanotubes (1.14–1.2 Rs), graphene (1.12–1.6 Rs), polymers (5 Rs) and metal oxides (2.2 Rs), and a commercial gas sensor (7 Rs, MICS-2714, Amphenol SGX Sensortech, Switzerland) (Fig. S3). Investigations at a wide range of gas concentrations indicated that the detection had linear electrical responses, and the inset (Fig. 2c) provides a detection limit (DL) of ~20 ppb, which was estimated by a linear least squares fit for data collected at room temperature<sup>32</sup>. The excellent sensitivity sufficiently detects concentrations required in potential applications, such as concentrations in exhalation by asthma and halitosis patients (>100 ppb) and the atmosphere (0.6–5 ppm)<sup>33–35</sup>. The rationale that crystallinity, thickness, device layout, and electrical operation would affect the exceptional performance of the

bioresorbable silicon devices motivated various types of examinations. Figure 2d shows the measured responses using different kinds of silicon, such as SC-Si NMs (black), polycrystalline silicon (red, p-Si), and amorphous silicon (blue, α-Si), with identical dimensions (1.45 mm × 1.2 mm) and thicknesses (~100 nm), providing evidence of a dependence on Si crystallinity (Fig. S4). Analyses of other factors, e.g., film thickness, electrode layout, and applied voltage, are shown in Fig. S5, and dissolvable metals for coplanar interdigitated electrodes (IDEs) did not affect the functional properties (Fig. S6). The selective response to specific target gases is also an important criterion. We performed selectivity assays for well-known gases: oxidizing gases such as NO<sub>2</sub> (black) and NO (red) and reducing gases such as NH<sub>3</sub> (blue), CH<sub>3</sub>COOH<sub>3</sub> (green, acetone), CH<sub>2</sub>H<sub>3</sub>OH (magenta, ethanol), CO<sub>2</sub> (cyan), CO (navy), and H<sub>2</sub>S (purple). The resulting properties in Fig. 2e indicate that the responsiveness to NO and NO<sub>2</sub> was at least 100-fold greater than that to other substances (detailed data in Fig. S7), which was achieved without additional physical and chemical treatments. We note that the variation in the responses between NO and NO<sub>2</sub> gases was negligible (see Fig. S8). The temperature dependence of the sensitivity shown in Fig. 2f was evaluated to consider potential utility as medical implants and environmental monitors (black, RT; red, 37 °C; blue, 50 °C). The results were opposite to the typical behavior of conventional gas sensors, assuming that the electrical resistance/conductance of the silicon membranes would somewhat deviate from the optimum value at increased temperatures. Integration with temperature and humidity sensors can adjust the output values of gas detectors susceptible to ambient conditions for system-level functionality. The electrical characteristics of the resistive temperature device showed a sensitivity of ~9.6 nS/°C and a temperature coefficient of ~0.006 /°C (Fig. 2g), and changes in the capacitance of the humidity component were well resolved over a wide range of relative humidities (Fig. 2h).

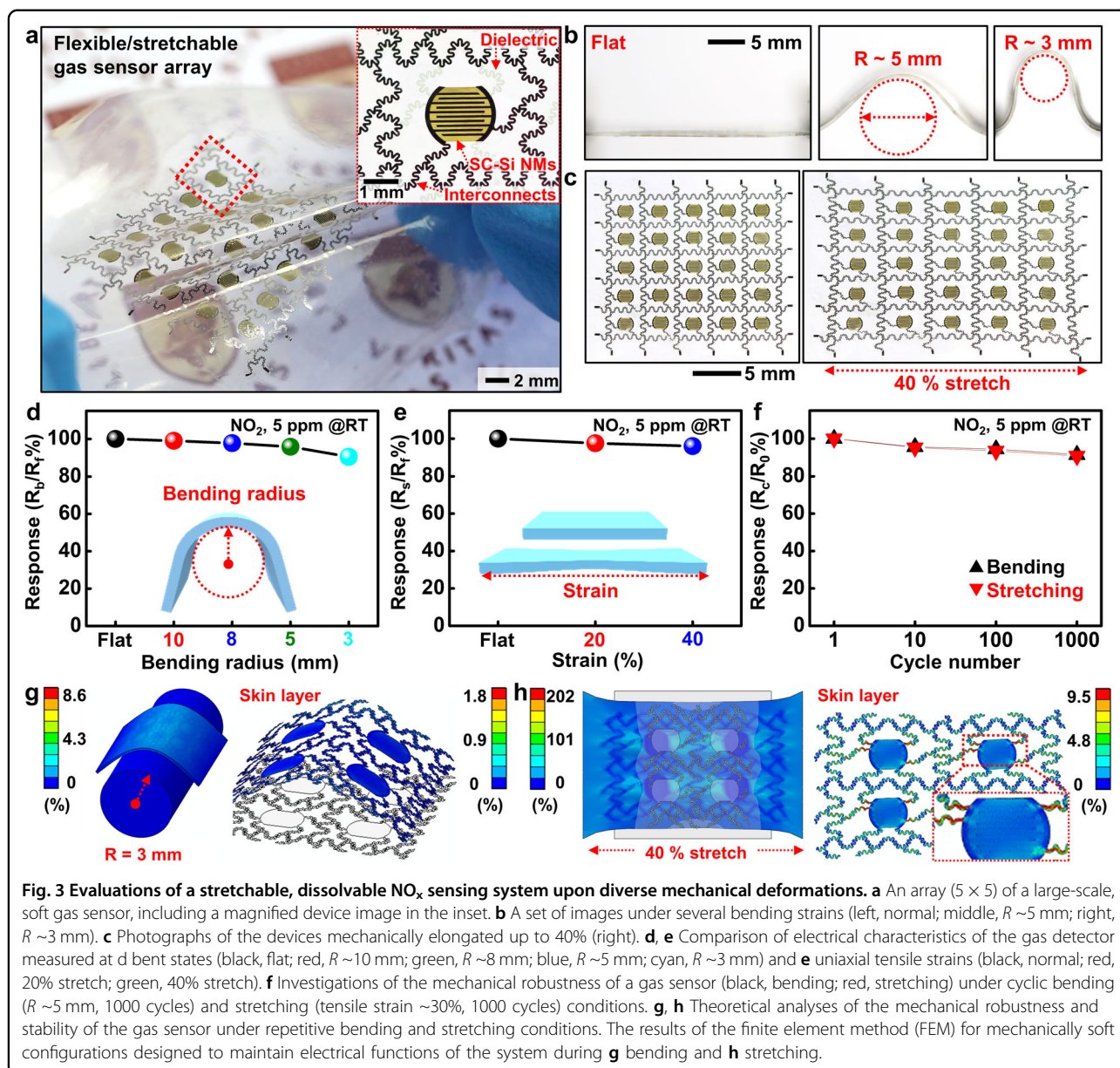
#### Mechanical stability of large-scale, flexible, and stretchable gas sensor arrays

As introduced in previous reports on deformable electronics, structures that involved device islands connected by serpentine bridges allowed devices to accommodate mechanical loads without deterioration in electrical performance<sup>36–38</sup>. Figure 3a represents a large-scale, biodegradable sensor array (5 × 5, 1.7 cm × 1.9 cm) deployed in a mechanically soft geometry, consisting of patterned SC-Si NMs as active components, Mg conductors for contact pads and interconnects, patterned SiO<sub>2</sub> layers for inter-layer dielectrics, and polycaprolactone (PCL) as substrate/encapsulant. The inset shows a magnified optical micrograph of an individual unit. Figure 3b, c show a set of



images under mechanical tests using several bending radii (left, flat; middle, 5 mm; right, 3 mm) and uniaxial tensile strains up to 40%, respectively. An experimental setup combined with a stretching system is shown in Fig. S9. The resulting values under those deformed conditions remained within effective ranges of 90% (Fig. 3d, bending) and 95% (Fig. 3e, stretching). In addition, the mechanical robustness and durability of the electronic system were investigated under cyclic bending with a radius of 5 mm (black) and stretching with 35% (red), as shown in Fig. 3f, and the decreases in the relative response rate in both cases after 1000 cycles were negligible. Three-dimensional finite element analysis was implemented in the commercial ABAQUS package to study the mechanical

deformation of the biodegradable Si-based gas sensor upon bending and stretching. Without losing generality, a  $2 \times 2$  array of gas sensors was considered in the simulation, and the substrate (100  $\mu\text{m}$ ) and encapsulation (50  $\mu\text{m}$ ) layers were modeled by an 8-node 3D element (C3D8R). PCL (Young's modulus, 0.25 GPa; Poisson ratio, 0.44) and semipermeable membranes (PDMS, 2.61 MPa; 0.49) were explored for the substrate and encapsulation in the bending test (Fig. 3d), whereas PDMS was used for the substrate in the tensile test (Fig. 3e). Because of their thinness, gas-sensitive Si nanomembranes (188 GPa; 0.28), Mg electrodes and interconnects (45 GPa; 0.29), and SiO<sub>2</sub> interlayer dielectrics (66.3 GPa; 0.15) could be modeled as skin layers, as shown in Fig. S10. For bending



with a radius of curvature of 3 mm in Fig. 3g, the maximum principal strains in the layers of Mg, SiO<sub>2</sub>, and Si were obtained at 1.66, 1.81, and 0.24% (Fig. S11), far below their corresponding fracture strains, which explains the mechanical stability. Upon stretching, the maximum principal strain in the skin layer occurred in the serpentine interconnect near the Si nanomembrane (Fig. 3h), and detailed strain distributions for each material are shown in Fig. S11. When the total stretchability of the system is defined at a level, where the maximum principal strain of half the width of one section exceeded the fracture strain of the metal interconnect (9–10% for Mg thin film<sup>39,40</sup>), as in a previous report<sup>37</sup>, the obtained total stretchability of  $\sim 40\%$  corresponded well to the

experimental value of Fig. 3e. The strain distribution in the SiO<sub>2</sub> layer was similar to that in the Mg layer, although the SiO<sub>2</sub> layer would fracture before the Mg layer due to its slightly smaller fracture strain of 8%<sup>41</sup>. In addition, the peak strain of 4.1% in the Si layer was immediately below its fracture strain of 4.25%<sup>42</sup>, which explained the fracture of the Si layer upon further stretching.

#### In vitro assessments of dissolvable gas sensors using a semipermeable membrane

Humidity-free gas meters enable the maintenance of robust performance even in harsh environments in envisioned areas of interest. Figure 4a illustrates the

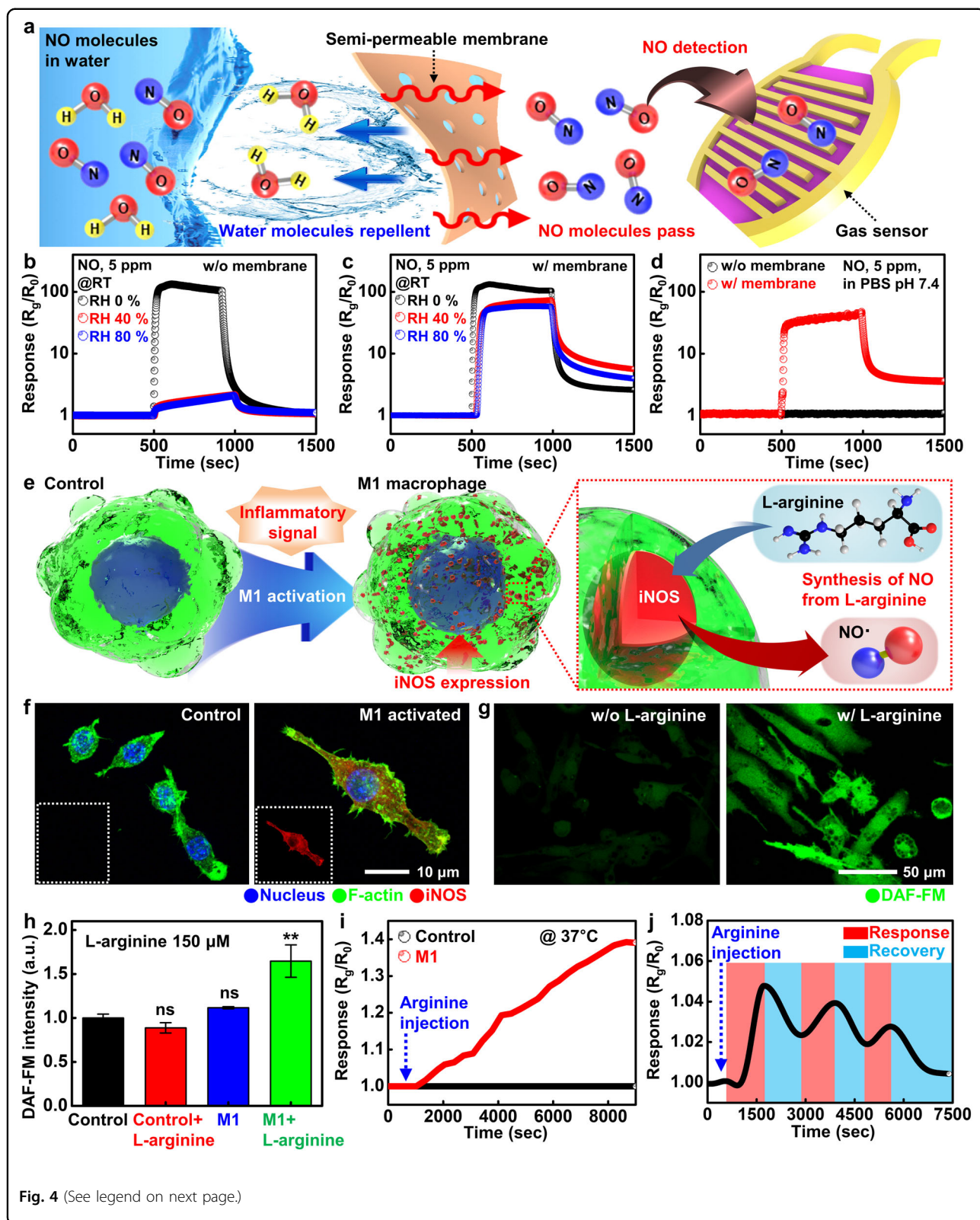


Fig. 4 (See legend on next page.)

(see figure on previous page)

**Fig. 4 In vitro detection of NO produced in macrophages using transient silicon sensors with semipermeable membranes.** **a** Schematic illustration of the principle of enabling gas measurements in humid conditions and aqueous solutions. Semipermeable membranes (polydimethylsiloxane, PDMS, thickness  $\sim 50$   $\mu\text{m}$ ) selectively allow passage of gas molecules while acting as a barrier to water molecules. **b, c** Evaluations of the electrochemical responses of integrated gas sensors **b** without and **c** with a thin sheet of semipermeable membrane (thickness  $\sim 50$   $\mu\text{m}$ ) to 5 ppm NO gas at several levels of relative humidity (black, 0%; red, 40%; blue, 80%). **d** Measured electrical properties of integrated gas sensors without (black) and with (red) semipermeable membranes in phosphate-buffered saline (PBS, pH 7.4) solution. **e** Description of key procedures for nitric oxide (NO) production through the inflammatory response of macrophages (left). The production of NO from L-arginine through enzymatic reactions by inducible nitric oxide synthase (iNOS) is shown in the inset (right). **f** Representative images of control (left) and M1-activated (right) RAW 264.7 macrophages. Nuclei (blue), F-actin (green), and iNOS (red) were visualized by maximum intensity projection of immunofluorescence confocal images. Note that iNOS expressed only M1-activated macrophages. **g** Microscopic images of NO production in M1 macrophages by L-arginine treatments (left, 0  $\mu\text{M}$ ; right, 150  $\mu\text{M}$ ). **h** Quantification of DAF-FM intensity for NO measurements. Values were normalized to the control values, and significant differences were evaluated by one-way analysis of variance (ANOVA) for each group (\*\* $p < 0.01$ , NS;  $p > 0.05$ ). **i** In vitro experiments assessing electrical responses to NO generated from control and M1 cells after injection with 150  $\mu\text{M}$  L-arginine. **j** Measured dynamic NO responses over time as periodic excitations and inhibitions.

fundamental principle that a PDMS-based semipermeable membrane can effectively deliver desired gas components to active areas of an electronic system and repel water and/or aqueous components<sup>43,44</sup>. The PDMS membrane provides hydrophobicity and gas permeability since methyl groups (Si-CH<sub>3</sub>) on the surface of PDMS reduce the surface energy, and siloxane backbones (Si-O) inside the polymer network form a pathway for NO diffusion. In comparative experiments without (Fig. 4b) or with (Fig. 4c) membranes, absolute values of the responsiveness were slightly reduced, while the electrical characteristics still exhibited a high level of sensitivity compared to those in the tests where there were nearly no responses with even a small amount of moisture in the absence of the membrane. Similar behaviors were found during immersion in aqueous media, in which the membrane enabled stable detection in phosphate-buffered saline (PBS, pH 7.4) solution, whereas no signals were observed without the membrane (Fig. 4d and experimental setup appeared in Fig. S12). Detailed studies of the correlation between membrane thickness and system performance could enhance the sensitivity under optimized conditions (Fig. S13).

In vitro assessment of the NO detection capability indicated the feasibility of using silicon-based bioresorbable devices for applications in temporary implants. Figure 4e briefly describes the overall procedure of NO production that is recognized as a regulator of inflammatory responses in living macrophages<sup>26</sup>. Macrophages undergo M1 activation in response to pathogen-specific molecules, lipopolysaccharide (LPS) or proinflammatory cytokines such as TNF- $\alpha$  and interferon- $\gamma$  (INF- $\gamma$ )<sup>45</sup>. As a result of inflammatory activation, macrophages produce inducible nitric oxide synthase (iNOS), one of the key enzymes that acts as a catalyst to produce NO by consuming L-arginine<sup>46</sup>. A set of fluorescence confocal images verified the expression of iNOS, cellular morphological changes (Fig. 4f) and NO production in the presence of L-arginine (Fig. 4g). Quantitative analysis of immunofluorescence intensity is shown in Fig. S14. The produced NO reacted with DAF-FM to

increase the fluorescence intensity in Fig. 4h, suggesting that both iNOS expression and the presence of L-arginine were essential factors. Figure 4i shows the measured responses after an injection of L-arginine (150  $\mu\text{M}$ ) into control (black) and M1-activated macrophages (red) in medium (L-arginine-free DMEM (2 ml)). Details of the experimental setup are shown in Fig. S15. The results showed that there was no response in the control group without M1 activation, while a linear response was clearly seen in the activated group, as expected. Evaluation of the dynamic response over time in Fig. 4j was performed through periodic L-arginine injections and ventilations, verifying the capability of the system for reliable and repeatable measurements of produced NO concentrations.

## Conclusion

The concepts introduced here provide materials, a manufacturing strategy and device layouts for a soft, transient NO<sub>x</sub> gas detector integrated with temperature and hydration meters. Electrical measurements reveal that the thickness, crystallinity, and operation voltage have a strong influence on the record-high sensitivity and selectivity with rapid response and recovery rates. Upon mechanical deformations by diverse modalities, the performance of large-scale arrays change negligibly, consistent with those determined by theoretical calculations. In vitro studies provide the feasibility of stable operation in harsh conditions, suggesting practical uses in environmental monitoring and biomedical implants.

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#### Conflict of interest

The authors declare that they have no conflict of interest.

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