

Molecular studies of *DO* alleles reveal that *JO* is more prevalent than *HY* in Brazil, whereas *HY* is more prevalent in New York

L. CASTILHO, W. BALEOTTI, JR., E. TOSSAS, K. HUE-ROYE, K.R. RIBEIRO, C. LOMAS-FRANCIS, D. CHARLES-PIERRE, AND M.E. REID

Because of the scarcity of anti-Hy and anti-Jo^a, hemagglutination typing for the Dombrock blood group system antigens, Hy and Jo^a, is not feasible. The molecular bases associated with these antigens have been determined, making it possible to distinguish *HY* and *JO* from wild-type *DO*. This provides a tool to predict the probable phenotype of patients and to screen for antigen-negative donors. PCR-RFLP assays and a microchip assay were used to determine the frequency of *HY* and *JO* alleles in donors from Brazil and New York. DNA from random Brazilian donors, 288 by PCR-RFLP and 599 by the bead array method (BeadChip, BioArray Solutions, Warren, NJ), was tested to determine 323G/T (*HY*+/*HY*-) and 350C>T (*JO*+/*JO*-) single-nucleotide polymorphisms. In New York, 27,226 donors who self-identified as being African American were tested by hemagglutination with anti-Gy^a. Nonreactive and weakly reactive samples were tested by PCR-RFLP for the same alleles as listed above. In Brazil, 30 (3.4%) of the samples were *JO/DO* and 13 (1.4%) were *HY/DO*. In New York, of the samples that had *HY* or *JO* alleles, 14 were homozygous *HY/HY*, 132 were heterozygous *HY/DO*, 13 were heterozygous *HY/JO*, 14 were heterozygous *JO/DO*, and 3 were homozygous *JO/JO*. These results show that in donors from Brazil, *JO* (30 alleles) is more than twice as prevalent as *HY* (13 alleles), whereas in donors from New York, *HY* (173 alleles) was more than five times more common than *JO* (33 alleles). *Immunohematology* 2008;24:135-137.

Key Words: blood groups, DNA testing, Dombrock, molecular basis

The Dombrock (Do) blood group system consists of five antigens: a pair of antithetical antigens (Do^a and Do^b) and three high-prevalence antigens (Gy^a, Hy, and Jo^a).¹ The Gy(a-) phenotype is the null of the Dombrock system.² The Hy-negative phenotype is associated with weak expression of Do^b and Gy^a, and absent or very weak expression of Jo^a, and the Jo(a-) phenotype is associated with weak expression of Do^a and Hy.^{1,3}

In the transfusion setting, antibodies to antigens in the Dombrock blood group system have caused delayed, and rarely acute, transfusion reactions. In the prenatal setting, they have caused a positive direct antiglobulin test but not hemolytic disease of the newborn and fetus.⁴ Antigens in the Dombrock blood group system are carried on the Dombrock glycoprotein, which is encoded by the *DO* gene, also known as *ART4*.⁵ The molecular bases associated with the various Do phenotypes have been determined to be caused by single-nucleotide polymorphisms.⁵⁻⁷ Our ability to type RBCs for Hy and Jo^a by hemagglutination has been severely limited because of the scarcity of suitable antibodies. As the molecular bases of these Dombrock blood group system antigens have been determined, the ability to distinguish *HY* and *JO* makes it feasible to predict the probable phenotype of patients and to screen for antigen-negative donors. Based on this knowledge, we used PCR-RFLP assays and a bead microchip assay to determine the relative frequency of *HY* and *JO* alleles in donors from Brazil and New York.

Materials and Methods

Genomic DNA was extracted from the buffy coat fraction from blood samples using a DNA extraction kit (QIAamp DNA Blood Mini Kit, QIAGEN, Inc., Valencia, CA). PCR was performed using the following conditions: 100 ng of each primer (synthesized by Life Technologies, Inc., Gaithersburg, MD), 200 μM of each dNTP, 2.5 mM (for nt 323 and nt 350

of *DO*) or 3.0 mM MgCl₂ (for nt 793 of *DO*), 1.0 U DNA polymerase (HotStar Taq, QIAGEN), and buffer in a total volume of 50 µL. Amplification was performed in a thermal cycler (GeneAmp PCR System 2400, Perkin Elmer, Norwalk, CT) with the following profile: 95°C for 15 minutes; followed by 35 cycles of 94°C for 20 seconds; 58°C (for nt 323 and nt 350 of *DO*) or 62°C (for nt 793 of *DO*) for 20 seconds and 72°C for 20 seconds; then 72°C for 7 minutes.^{7,8} PCR products were analyzed by electrophoresis in 1% agarose gel. PCR-RFLP assays were performed as previously described.^{7,8} The sequence of primers, PCR product size, restriction enzyme used to digest each PCR-amplified product, and expected restriction fragment sizes are given in Table 1. Digested products were analyzed by electrophoresis in 8% polyacrylamide gel.

In Brazil, DNA samples from random Brazilian donors were tested by PCR-RFLP (n = 288) and by HEA bead microchip (n = 599; BeadChip, BioArray Solutions, Warren, NJ) to determine 793A>G (*DO**A/*DO**B), 323G>T (*HY*+/-), and 350C>T (*JO*+/-) single-nucleotide polymorphisms. The donors were predominantly of European and African descent, and a small number were of Asian descent. In New York, 27,226 (from September 1999 to April 2007) donors who self-identified as being African American were tested by hemagglutination with a polyclonal anti-Gy^a by the IAT. Nonreactive and weakly reactive samples were tested by PCR-RFLP for the same alleles as listed above.

Results

In Brazil, 21 (2.4%) typed as *JO/DO**B, 9 (1.0%) as *JO/DO**A, 5 (0.5%) as *HY/DO**A, and 8 (0.9%) as *HY/DO**B. Thus, of donors who had *HY* or *JO* alleles, 30 were heterozygous *JO* in *trans* to a *DO**B or *DO**A, and 13 were heterozygous *HY* in *trans* to *DO**A or *DO**B (Table 2). All donors with *HY* or *JO* alleles were Afro-Brazilians.

Table 1. Primers used for PCR-RFLP analyses

Primer Name	Primer Sequence (5' to 3')	Uncut Size (bp)	Restriction Enzyme (nt)	Restriction Fragment Size (allele)	
DoF	TACCTCACCTCAGCAATCCAGCTGCTGAGGAGAGAC	368	<i>Bse</i> RI	326, 42	268, 58, 42
DoR	TTTAGCAGCTGACAGTTATATGTGCTCAGGTTC		(nt 793)	(<i>DOA</i>)	(<i>DOB</i>)
DoX2F	TCAGTACCAAGGCTGTAGCA	220	<i>Bsa</i> JI (nt 323)	120, 92, 8 (<i>DO</i>)	212, 8 (<i>HY</i>)
Do378R	AGTAAAGTCAGAATGAACATTGCTGCACAAT		<i>Xcm</i> I (nt 350)	167, 53 (<i>DO</i>)	220 (<i>JO</i>)

nt = nucleotide; bp = base pairs; *DO* = *DOA* or *DOB*.

Table 2. Distribution of *HY* and *JO* alleles in donors of African descent in Brazil and New York

Allele Combinations	Brazilian Donors (n = 43)	New York Donors (n = 176)
<i>HY/DO</i>	13 (30%)	132 (75%)
<i>HY/HY</i>	0	14 (8%)
<i>HY/JO</i>	0	13 (7%)
<i>JO/DO</i>	30 (70%)	14 (8%)
<i>JO/JO</i>	0	3 (2%)

In New York, of the samples that had *HY* or *JO* alleles, 14 were homozygous *HY/HY*, 132 were heterozygous *HY* in *trans* to a *DO**A or *DO**B, 13 were heterozygous *HY/JO*, 3 were homozygous *JO/JO*, and 14 were heterozygous *JO* in *trans* to a *DO**A or *DO**B (Table 2).

Discussion

In this study, in the Afro-Brazilians, *JO* (30 alleles) is about twice as frequent as *HY* (13 alleles). In contrast, in African American donors from New York, *HY* (173 alleles) is more than five times as common as *JO* (33 alleles). It is likely these findings reflect that Africans brought to South America were from a different region of Africa than those brought to the East Coast of North America.

Antibodies to antigens in the Dombrock blood group system are difficult to identify, and the paucity of reliable monospecific antisera hampers studies involving the Dombrock blood group system. At least one anti-Hy has caused biphasic destruction of Hy+ RBCs.⁹ Other examples of anti-Hy as well as anti-Gy^a and anti-Jo^a have caused moderate transfusion reactions (reviewed in Reid¹). PCR-based testing for *DOA*, *DOB*, *HY*, and *JO* alleles provides a tool to predict the probable phenotype of patients and blood donors. This is an advantage for screening a large number of donors to find those who are Do(a-), Do(b-), Hy-, or Jo(a-), a feat not possible by hemagglutination. Thus, for Dombrock typing, DNA-based assays are not only feasible, they are more reliable

than hemagglutination. Our findings emphasize the importance of testing populations with different ethnic backgrounds to define their *DO*, and other blood group, alleles.

Acknowledgments

We thank Robert Ratner for assistance in preparation of the manuscript.

References

1. Reid ME. The Dombrock blood group system: A review. *Transfusion* 2003;43:107–14.
2. Banks JA, Hemming N, Poole J. Evidence that the Gy^a, Hy and Jo^a antigens belong to the Dombrock blood group system. *Vox Sang* 1995; 68:177–82.
3. Scofield TL, Miller JP, Storry JR, et al. Evidence that Hy⁻ RBCs express weak Jo^a antigen. *Transfusion* 2004;44:170–2.
4. Reid ME, Lomas-Francis C. Blood Group Antigen FactsBook. 2nd ed. San Diego: Academic Press, 2004.
5. Gubin AN, Njoroge JM, Wojda U, et al. Identification of the Dombrock blood group glycoprotein as a polymorphic member of the ADP-ribosyltransferase gene family. *Blood* 2000;96:2621–7.
6. Rios M, Hue-Roye K, Lee AH, et al. DNA analysis for the Dombrock polymorphism. *Transfusion* 2001;41:1143–6.
7. Rios M, Hue-Roye K, Øyen R, et al. Insights into the Holley-negative and Joseph-negative phenotypes. *Transfusion* 2002;42:52–8.
8. Storry JR, Westhoff CM, Charles-Pierre D, et al. DNA analysis for donor screening of Dombrock blood group antigens. *Immunohematology* 2003;19:73–6.
9. Beattie KM, Castillo S. A case report of a hemolytic transfusion reaction caused by anti-Holley. *Transfusion* 1975;15:476–80.

Lilian Castilho, PhD, Hemocentro, Universidade de Campinas, Campinas, São Paulo, Brazil, Wilson Baleotti, Jr, PhD, Faculdade de Medicina de Marília, Marília, São Paulo, Brazil, Edith Tossas, New York Blood Center, New York, NY, Kim Hue-Roye, BA, New York Blood Center, New York, NY, Karina R. Ribeiro, MSc, Hemocentro, Universidade de Campinas, Campinas, São Paulo, Brazil, Christine Lomas-Francis, MSc, New York Blood Center, New York, NY, Daisy Charles-Pierre, BS, New York Blood Center, New York, NY, and Marion E. Reid, Ph.D. (corresponding author), Director, Immunohematology Laboratory, New York Blood Center, 310 East 67th Street, New York, NY 10021.

Free Classified Ads and Announcements

Immunohematology will publish classified ads and announcements (SBB schools, meetings, symposia, etc.) **without charge**. Deadlines for receipt of these items are as follows:

Deadlines

- 1st week in January for the March issue
- 1st week in April for the June issue
- 1st week in July for the September issue
- 1st week in October for the December issue

E-mail or fax these items to Cindy Flickinger, Managing Editor, at (215) 451-2538 or flickingerc@usa.redcross.org.