Research Article

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Burial depths favor Italian ryegrass persistence in the soil seed bank

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Received March 26, 2019 Accepted October 23, 2019 ABSTRACT: Italian ryegrass (Lolium multiflorum Lam.) is a weed broadly found in fields cultivated with wheat and barley crops. Seed inputs into the soil before harvesting winter crops increase seed bank, ensuring survival. This study evaluated the persistence of Italian ryegrass seeds subjected to burial depths over time. Experiments were carried out in the field for two years in randomized block experimental design with four repetitions. Dormant seeds harvested from mother-plants were put in nylon bags containing soil. Previously, seed viability was evaluated in a 1 % tetrazolium solution, and 50 viable seeds by repetition were buried at 0.5, 5.0, 10, and 20 cm depths. Seed persistence was evaluated by the percentage of deteriorated and remaining seeds, non-dormant seeds, abnormal seedlings, and viable and non-viable dormant seeds at 0, 30, 60, 90, 180, 360, 540, and 720 days after burial (DAB). Seed persistence increased at 10 and 20 cm of burial depth compared to seeds in the soil layers up to 5 cm. Moreover, burial depth at 10 and 20 cm showed lower percentage of deteriorated seeds (10 % lower) compared to 0.5 cm at 360 DAB. For non-dormant seeds, a higher percentage was found at 90 DAB, regardless of seed burial depth. Dormancy breakage occurred until 180 DAB, and more rapidly at 10 and 20 cm depths. At 540 DAB, more than 95 % of seeds were unviable, demonstrating short persistence of Italian ryegrass in the soil seed bank.

Keywords: Lolium multiflorum Lam., seed bank weed management, seed longevity, seed dormancy, soil seed bank inputs

Introduction

Weeds are a major problem for agricultural production, causing significant yield reduction and economic losses worldwide. Italian ryegrass (*Lolium multiflorum* Lam.) is a winter annual grass species widely found in cropping systems, and considered the most important weed in wheat and barley fields (Vargas and Roman, 2005). Populational infestations have been increasing over the last years due to herbicide resistance, high seed production, premature dehiscence, seed dormancy, and absence of soil disturbance that contributes to seed bank feedback in the soil, providing survival in growing areas and enabling future infestations (Norris, 2007; Fernández-Moreno et al., 2017).

In addition, weed presence and interference capacity in growing areas depends on seed bank and dormancy degree, important factors capable of modifying the emergence moment and crop-weed competition (Ghersa et al., 1997). Seed dormancy in weeds is an evolutionary feature that causes germination delay over time and prevents the establishment under inadequate conditions, allowing population fluctuations (Graeber et al., 2012; Née et al., 2017).

In cropping systems, soil seed bank represents a potential target for management where temporary changes in environmental conditions and soil practices could affect weed population dynamic and exhausting the soil seed bank (Shaner and Beckie, 2014; Armengot et al., 2017). For example, mechanical control during the harvest of grain crops with the Harrington Seed Destructor Machine (HSD) could destroy more than 95 % of the annual ryegrass (*Lolium rigidum* Gaudin.) seeds, reducing seed inputs into the soil, essential to reduce weed population sustainably in the long-term in Australian fields (Walsh et al., 2012).

Persistence of viable seeds in soil seed bank without seed destruction depends on biotic and abiotic factors, such as germination cues, seed size and dormancy, predation, and microbial decay, which could be affected by environmental conditions as burial depth and persistence time (Korres et al., 2018). Some management practices, such as plowing and harrowing, cause temporary changes in temperature and water contents in the soil profile, altering the seed dormancy level and viability (Harker and O'Donovan, 2013; He et al., 2014). Conversely, viability of weed seed was higher after burial than in seeds on the soil surface (Sosnoskie et al., 2013; Jha et al., 2014).

Studies conducted for long periods to assess the effects of burial depth and time on viability of Italian ryegrass seeds are important to estimate seed bank persistence and develop weed management strategies. Therefore, this study evaluated persistence of Italian ryegrass seeds at burial depths over time.

Material and Methods

We evaluated Italian ryegrass seed persistence in a field experiment ($31^{\circ}80'08''$ S, $52^{\circ}50'13''$ W, altitude of 14 m) carried out from Oct 2016 to Oct 2018, using a randomized block experimental design with four repetitions. Previously, \pm 20.000 dormant seeds with viability of 96 % were sampled directly from mother-



plants and sorted using a diaphanoscope of light. Seed viability was evaluated on 400 seeds, using a 1 % tetrazolium solution kept in the dark at 30 °C for 6 h, to develop the embryo red color in viable seeds. (MAPA, 2009). Fifty viable seeds were put in naylon bags of 10 \times 10 cm with 500-micron pore openings, containing 50 g of soil, and buried in the field at different depths. The soil was classified as Endoaquult (USDA Soil Taxonomy), with 5.6 of pH, 1.5 % organic matter and 16 % clay. The meteorological conditions during the experiment are shown in Figure 1.

Seed viability of Italian ryegrass was evaluated at four burial depths (0.5, 5.0, 10, and 20 cm), and seeds were recovered at 0, 30, 60, 90, 120, 360, 540, and 720 DAB (Figure 2). Field samples collected in each time were washed in running water using a set of 16, 32, and 60 mesh sieves. After washing, the samples were placed on filter paper for 24 h, and remaining seeds were



Figure 1 – Meteorological conditions (rainfall, maximum, and minimum temperature) during the experiment on persistence of Italian ryegrass in soil seed bank.

collected manually and separated for the germination test. This study was performed at the seed laboratory using germination boxes ($11.5 \times 11.5 \times 3.5$ cm), where seeds were distributed over two sheets of blotter paper, previously moistened with distilled water at an amount equivalent to 2.5-fold the paper weight. The germination test was carried out in a chamber of biochemical oxygen demand (BOD) for 14 d under alternating temperature ($20 \ ^{\circ}$ C for 16 h in dark and $30 \ ^{\circ}$ C for 8 h in light), according to the Brazilian Rules for Seed Testing (MAPA, 2009).

The percentage of deterioration (seed losses in the field + dead seeds found in the germination test), remaining seeds, non-dormant seeds, abnormal seedlings, viable and non-viable dormant seeds were measured in each time, using as parameter the initial number of seeds buried. After the germination test, non-germinated seeds were longitudinally cut through the embryo and kept in a 1 % tetrazolium solution for imbibition in the dark for 6 h at 30 °C, according Brazilian Rules for Seed Testing (MAPA, 2009).

Data were subjected to the analysis of variance ($p \le 0.05$) and the means were compared by the Tukey test ($p \le 0.05$) using the Statistical Analysis System software (SAS, version 9.0). Bar charts were created using the Sigmaplot software (SIGMAPLOT, version 12.3), considering interactions between time and burial depths for each variable.

Results and Discussion

For all variables measured to study persistence of Italian ryegrass in soil seed bank, significant differences were found between time and burial depths (Figure 3, 4, 5, and 6). The percentage of deteriorated seeds after burial increased over time at all depths, and higher values were observed for the seeds buried at 0.5 cm



Figure 2 – Experimental layout of Italian ryegrass seed persistence, demonstrating the design plot, burial depths, seed bags, and remaining seeds over time.

Seed persistence is affected by burial

than for burial depths of 10 and 20 cm (Figure 3A). Similar results were reported for seed viability in *Conyza bonariensis* (L.) Cronq. with losses up to 59 % for periods shorter than 360 DAB (Vargas et al., 2018). At 180 DAB, burial depth of 0.5 cm exhibited approximately 45 % of deteriorated seeds compared to 5-20 cm burial depths with values 35 and 28 %, respectively. However, more than 95 % of seeds were lost up to 540 DAB, regardless of the depth evaluated (Figure 3A). Similar results were found for *L. rigidum* Gaudin. in which about 99 % of seeds were lost after 16 months of burial, demonstrating low persistence in soil seed bank (Narwal et al., 2008).

A significant decrease of remaining seeds was found at all burial depths, especially in evaluations performed between 90 and 360 DAB, where more than 85 and 65 % of seeds were lost at 0.5 and 20 cm depths, respectively (Figure 3B). Changes in weed dynamic in soil seed bank over time may occur due to losses by germination, migration, and degradation/predation of microorganisms and invertebrates (Chee-Sanford et al., 2006). The presence of weed seeds on surface soil layer



Figure 3 – Percentage of deteriorated (A) and remaining seeds (B) evaluating at different times and burial depths. Error bars represent the minimum significant difference in each time by the Tukey test (p < 0.05).

favors mortality due to frequent changes in temperature and soil moisture, factors that contribute to break seed dormancy, promoting seed germination and increasing predation and microbial activity (Main et al., 2006).

Italian ryegrass seeds buried at 20 cm depth showed faster dormancy breakage compared to 0.5 and 5.0 cm, where germination of non-dormant seeds was approximately 10 % higher at 30 DAB (Figure 4A). However, the maximum germination rate was obtained for evaluations performed between 90 and 180 DAB with more than 55 % of seeds germinated due to dormancy breakage, regardless of burial depths (Figure 4A). Moreover, a decrease in the non-dormant Italian ryegrass seed number after 180 DAB was observed as a result of increased mortality, which was demonstrated until 540 DAB when more than 95 % of seeds were lost in soil bank for all depths.

In weeds, dormancy is an important evolutionary mechanism to prevent germination under unfavorable environmental conditions, contributing to the spatiotemporal adjustment, regulating the appropriate timing of seed germination under optimal conditions to ensure species development and survival (Gardarin and Colbach, 2014). Furthermore, seed dormancy breakage is regulated by several environmental and endogenous signals that may control seed physiological and biochemical processes (Née et al., 2017). Changes of temperature and soil moisture in surface layers provide faster dormancy breakage and promote mortality increase, allowing lower persistence in soil seed bank (Davis et al., 2008; Donohue et al., 2010). However, unfavorable conditions for seed germination can induce secondary dormancy to ensure germination capacity and persistence increase (Nambara et al., 2010; Graeber et al., 2012).

Percentage of abnormal seedlings at 0.5 cm burial depth was 2 % higher than at the other depths from 60 to 90 DAB, and similar after 180 days of the experiment start (Figure 4B). Abnormal seedlings indicate seed vigor loss and reduce plant development, growth, and capacity of occupying soil seed bank, even under highly favorable environments of temperature, soil moisture, and light (Barth Neto et al., 2014). Abnormal seedlings on upper soil layers can occur due to increase of predation and microbial deterioration as well as changes of temperature and soil moisture compared to burial depths at 10 and 20 cm depths, which are more homogeneous environments (Née et al., 2017; Korres et al., 2018).

Considering dormant viable seeds, the field experiment started with more than 92 % viable seeds evaluated by the tetrazolium test. Strong reduction in the percentage of viable dormant seeds was found until 90 DAB, with values from 5 to 17 % depending on the burial depth and seed dormancy breakage in the field (Figure 5A). Similarly, the maintenance of seeds near the surface layer favored dormancy breakage and increased the percentage of non-viable dormant seeds compared to seeds buried deeper than 5 cm (Figure 5B).



Figure 4 – Percentage of non-dormant seeds (A) and abnormal seedlings (B) evaluated at different times and burial depths. Error bars represent the minimum significant difference in each time by the Tukey test (p < 0.05).

germination In agricultural systems, and biological processes, such as deterioration and predation of weed seeds, are stimulated by the presence of seeds on the soil surface, providing better conditions for management of soil seed bank (Goggin et al., 2012; Scherner et al., 2016; Vargas et al., 2018). Other characteristics associated the soil seed bank dynamic involve complex interactions with each kind of weed species (dormancy), environmental conditions (light, temperature, and soil moisture), biological processes, such as predation and allelopathy, and soil management practices (harrowing and/or plowing) that can affect persistence of soil seed banks (Graeber et al., 2012; Scherner et al., 2016). Moreover, crop rotation during summer and winter periods and/or the use of cover plants could be adopted by farmers to reduce the input of seeds into the soil (Galvan et al., 2015). Crop rotation is important to change herbicides used at the pre- and post-emergence of crops, whereas cover plants could reduce weed growth due to competition, reducing weed seed production.



Figure 5 – Percentage of viable dormant (A) and non-viable dormant seeds (B) evaluated at different times and burial depths. Error bars represent the minimum significant difference in each time by the Tukey test (p < 0.05).

Our results for viable seeds, considering the sum of abnormal seedlings, non-dormant seeds, and viable dormant seeds, show that these seeds can occupy the environment and generate plants capable of completing the life cycle and producing seeds. Thus, burial at 10 and 20 cm depths reduced deterioration and increased persistence of Italian ryegrass seeds in soil bank (Figure 6). In addition, more than 94 % of the seeds were not recovered at 540 DAB, regardless of the burial depth, indicating short persistence of Italian ryegrass in soil seed bank (Figure 6). Low viability of weed seeds was also reported in Amaranthus palmeri S. Watson (Korres et al., 2018), Conyza bonariensis (L.) Crong. (Vargas et al., 2018) and Lolium rigidum Gaudin. (Narwal et al., 2008), which presented short persistence, depending on the burial depths.

The low viability of Italian ryegrass and maintenance of seeds on soil surface are key factors to establish more efficient management strategies and reduce soil seed bank, avoiding future infestations. Our suggestions to farmers include integrated management



Figure 6 – Deterioration and viability of seeds buried at 0.5 cm (A), 5.0 cm (B), 10 cm (C), and 20 cm (D) depths evaluated for two years in the field experiment.

practices, such as cover crops for weed growth suppression and crop rotation, along with the chemical control to reduce infestation and seed production of Italian ryegrass.

Conclusions

Greater burial depths of Italian ryegrass seeds increase persistence in soil bank due to lower deterioration compared to seeds in the soil surface layer.

Italian ryegrass persistence in soil seed bank is short and lower than 540 days, regardless of depth.

Farmers can adopt management practices to prevent new seed inputs into the soil due to short persistence of Italian ryegrass seeds.

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Authors's Contributions

Conceptualization: Cechin, J; Agostinetto, D.; Vargas, L. Data acquisition: Cechin, J.; Schmitz, M.F.; Henckes, J.R. Data analysis: Cechin, J.; Design of Methodology: Cechin, J.; Vargas, A.A.M.; Agostinetto, D.; Vargas, L. Writing and editing: Cechin, J.

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