





Brain & Development 43 (2021) 144–151

www.elsevier.com/locate/braindev

of Child Neurology

Case Report

Tremor as an early sign of hereditary spastic paraplegia due to mutations in ALDH18A1

Tibor Kalmár, Zoltán Maróti, Alíz Zimmermann, László Sztriha*,1

Department of Pediatrics, University of Szeged, Szeged, Hungary

Received 6 May 2020; received in revised form 23 July 2020; accepted 23 July 2020

Abstract

Background: The *ALDH18A1* gene is located at 10q24.1 and encodes delta-1-pyrroline-5-carboxylate synthetase (P5CS), a mitochondrial bifunctional enzyme that catalyzes the first two steps in *de novo* biosynthesis of proline, ornithine, citrulline, and arginine. *ALDH18A1*-related disorders have been classified into four groups, such as autosomal dominant and recessive hereditary spastic paraplegia (SPG9A and SPG9B, respectively), as well as autosomal dominant and recessive cutis laxa (ADCL3 and ARCL3A, respectively). Neurodegeneration is a characteristic feature of all groups.

Case report: Here, we report a girl with compound heterozygous disease-causing variants (c.-28-2A>G and c.383G>A, p. Arg128His) in the *ALDH18A1* gene, revealed by whole exome sequencing. The c.-28-2A>G variant in intron 1, inherited from the mother, is a novel mutation, while the c.383G>A variant in exon 4, inherited from the father, has already been reported. The patient presented with vigorous infantile tremor preceding progressive spastic paraplegia. Dysmorphic features included elongated face, deep-set ears, upturned nose, long philtrum and pointed chin. Intrauterine and postnatal growth retardation, microcephaly, global developmental delay and profound intellectual disability were also noticed. Blood fasting ammonia level, plasma proline, ornithine and arginine levels were normal, while citrulline level was slightly decreased. Brain MRI revealed moderate hypoplasia of the corpus callosum and reduction of white matter volume.

Conclusions: The patient represents SPG9B, a rare form of autosomal recessive hereditary spastic paraplegias. The early onset tremor, preceding lower limb spasticity appears to be a unique early manifestation of neurodegeneration in this case. © 2020 The Japanese Society of Child Neurology. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license

(http://creativecommons.org/licenses/by-nc-nd/4.0/).

Keywords: ALDH18A1-related disorders; Neurodegeneration; Hereditary spastic paraplegia type 9B; Growth retardation; Intellectual disability

* Corresponding author at: Department of Pediatrics, Division B, University of Szeged, Temesvári krt. 35-37, 6726 Szeged, Hungary.

E-mail address: sztriha.laszlo@med.u-szeged.hu (L. Sztriha).

¹ ORCID: http://orcid.org/0000-0002-8698-6514.

1. Introduction

Monoallelic and biallelic mutations in *ALDH18A1* (OMIM 138250), located at 10q24.1, can cause neurodegeneration in association with various nonneurological features [1–3]. Based on genotypic and phenotypic features the *ALDH18A1*-related disorders have been classified into four groups, such as autosomal dominant and recessive hereditary spastic paraplegia (SPG9A, OMIM 601162 and SPG9B, OMIM 616586,

https://doi.org/10.1016/j.braindev.2020.07.015

0387-7604/© 2020 The Japanese Society of Child Neurology. Published by Elsevier B.V.

This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Abbreviations: P5CS, delta-1-pyrroline-5-carboxylate synthetase; G5K, glutamate 5-kinase; G5PR, glutamate 5-phosphate reductase; SPG9A, autosomal dominant hereditary spastic paraplegia; SPG9B, autosomal recessive hereditary spastic paraplegia; ADCL3, autosomal dominant cutis laxa; ARCL3A, autosomal recessive cutis laxa; Gno-mAD, The Genome Aggregation Database

145

respectively), as well as autosomal dominant and recessive cutis laxa (ADCL3, OMIM 616603 and ARCL3A, OMIM 219150, respectively) [1–3]. ALDH18A1 encodes delta-1-pyrroline-5-carboxylate synthetase (P5CS, EC 1.2.1.41 and 2.7.2.11), a mitochondrial bifunctional enzyme that catalyzes the first two steps in the biosynthesis of proline, ornithine, citrulline, and arginine from glutamate. It comprises two domains, with different enzymatic activities: an N-terminal glutamate 5-kinase (G5K) domain, responsible for the glutamate phosphorylation to gamma-glutamyl phosphate, and a Cterminal glutamate 5-phosphate reductase (G5PR) domain, which catalyzes the reduction and conversion to gamma-glutamyl semialdehyde, which is further metabolized to proline and ornithine [1-3]. Two isoforms of P5CS are generated, differing only by 2 amino acid insert in the G5K domain. The short P5CS isoform has high activity in gut, where it catalyzes an essential step in the arginine biosynthetic pathway. The long isoform of P5CS is expressed in multiple tissues and is necessary for the synthesis of proline from glutamate [4]. Although P5CS expression in the brain is not strong, it has a measurable activity [4].

Each of the *ALDH18A1*-related disorders are rare [1]. We extend the genotypic and phenotypic spectrum of SPG9B by reporting a girl with compound heterozygous *ALDH18A1* mutations who had intense tremor in infancy, preceding the development of spastic paraplegia.

2. Case report

The proband, a girl was born from the first uneventful pregnancy to heathy, non-consanguineous Caucasian parents on the 37th gestational week. Her birth weight was 1950 g (-2.2 SD), head circumference 30 cm (-2.0 SD) and length 44 cm (-1.4 SD). Dysmorphic features included elongated face, deep-set ears, upturned nose, long philtrum and pointed chin (Fig. 1A, B). She did not have any cutaneous involvement. At about 2 months of age fast head and hand tremor appeared in the form of rhythmic back-and-forth involuntary movements with low amplitudes both at rest and during action. Fasting did not worsen these movements. The tremor became quite vigorous with waxing amplitudes during infancy and gradually waned later. There was no tremor at rest by the age of 5 years; however, emotional distress, particularly fear still provoked it. She had hypotonia and was unable to sit at age of 10 months, or stand at 12 months of age. She had very short attention span and limited interest in her surroundings. Developmental Quotient (DQ) of 45 was found by Brunet-Lézine test at the age of 2 years.

Fasting ammonia level was normal. Plasma proline, ornithine and arginine levels were also within the normal range, while citrulline level was slightly decreased



Fig. 1. Photos and MR images of the patient. The patient at the age of 2.5 years (A and B). Dysmorphic features can be seen: elongated face, deep-set ears, upturned nose, long philtrum and pointed chin. Parental written permission has been gained to publish the patient's photos. T2-weigthed axial (C) MR image of the patient shows the paucity of white matter and thinning of the genu of the corpus callosum compared to age-matched control (E). T1 weighted sagittal MR image of the patient (D) demonstrates the hypoplasia of the corpus callosum (D) compared to control (F).

(9 μ mol/L, normal: 10–50 μ mol/L, borderline). The results of other blood tests were normal.

Brain MRI at the age of 2 years revealed reduced volume of white matter and moderate hypoplasia of the corpus callosum (Fig. 1C–F).

At the age of 5.5 years her head circumference was 46 cm (-3.5 SD), weight 15 kg, (-1.9 SD), and height 98 cm (-2.8 SD). By this age, marked spasticity through most of the range of motion (Modified Ashworth scale

2) developed in her lower limbs with brisk deep tendon reflexes. Wide based spastic gait was also observed. There was no speech and intellectual disability was evident. She had disruptive behavior hindering us from taking formal Intelligence Quotient (IQ) test.

2.1. Genetic analysis

Routine chromosomal analysis by G-banding showed normal 46,XX karyotype. Genomic DNA was extracted from peripheral blood samples with the Puregene kit (Gentra). Array comparative genomic hybridization (aCGH) showed normal genomic copy number (Quantitative Genomic Medicine Laboratories, S.L., Barcelona, Spain).

Trio analysis by whole exome sequencing (WES) was performed with CentoXome® Gold at Centogene AG (Rostock, Germany) as described earlier [5]. A heterozygous variant in intron 1 (NM 002860.4:c.-28-2A>G) another heterozygous variant in exon 4 and (NM_002860.4:c.383G>A, NP_002851.2:p.Arg128His) of the ALDH18A1 gene were detected (Fig. 2A, B). The c.-28-2A>G variant is likely pathogenic because it changes the acceptor splice site of intron 1 in the 5'-UTR, causing skipping of exon 2 (the start codon is in exon 2). The possible molecular effect of this variant was tested in silico using MutationTaster. It predicted that c.-28-2A>G variant is disease causing (prob: 0.969300883264394); the wild type splice site (tgcal GATA c.-28) has been lost. This variant is absent in the databases [GnomAD (The Genome Aggregation Database), dbSNP, Exome Variant Server, ClinVar]. However, the p.Arg128His variant has been previously reported in compound heterozygous state as diseasecausing for autosomal recessive spastic paraplegia [3]. The c.-28-2A>G variant was also detected in the mother (Fig. 2A) in a heterozygous state, whereas the c.383G>Awas detected in the father (Fig. 2B) also in a heterozygous state.

3. Discussion

The patient in this report is compound heterozygous for two *ALDH18A1* disease-causing mutations. Both mutations affect the G5K domain of the P5CS enzyme. The phenotypic characteristics, such as intrauterine growth retardation, dysmorphic features, short statue, microcephaly, global developmental delay, cognitive impairment, progressively developing spastic paraplegia and lack of cutaneous manifestations in association with biallelic *ALDH18A1* mutations meet the criteria of SPG9B [1–3,6–8].Fifteen patients in 8 families have been described so far with this disorder, including the patient in this report (Table 1). The compound heterozygous mutations in 5 families and homozygous mutations in 3 families distributed randomly in the *ALDH18A1* gene, affecting both the G5K and G5PR domains (Table 1). Corpus callosum hypoplasia and thin white matter, found in our patient, have been described in *ALDH18A1*-related disorders, however they are rare in SPG9B (Table 1). Autopsy findings or neuropathology have never been reported.

The vigorous infantile tremor, the presenting sign in our patient, was unusual and has never been reported so early in SPG9B. Tremor, reported in other cases of SPG9B, started later, at around the 7th and 15th years of age [2,3] (Table 1). It can be regarded as a manifestation of neurodegeneration [9]. Tremor has also been seen in patients with ARCL3 due to biallelic *ALDH18A1* mutations [10,11]. Tremor can occur in other types of hereditary spastic paraplegias as well, like in SPG4 due to *SPAST* mutation [12] and SPG11 due to mutations in the *spatacsin* gene [13,14].

Neurodegeneration seems to be a common feature of autosomal recessive and autosomal dominant ALDH18A1-related disorders [1]. A unifying view for these disorders has been hypothesized, claiming that the different presentations conform to a disease continuum of decreasing severity from the cutis laxa forms ARCL3A and ADCL3 to the motor syndromes SPG9B and SPG9A [1]. Global developmental delay and intellectual disability are major manifestations of the central nervous system involvement with equal frequency in both autosomal recessive forms, i.e. in ARCL3A and SPG9B [1]. While hypotonia is a consistent feature in ARCL3A, occasionally followed by pyramidal signs [1,10,15], hypotonia seems to be rare in SPG9B; progressive hypertonia, spasticity and pyramidal signs prevail instead [1,3], as in our patient. A transition between these autosomal recessive conditions might be represented by patients reported with biallelic mutations without both cutis laxa and spastic paraplegia [[16], Patient 2 in [17]].

Data has been collected and reviewed by Marco-Marin and coworkers in favor of the view that the severity of the various syndromes in ALDH18A-related disorders would correspond to higher or lower degrees of loss of P5CS function [1]. Decreased serum P5CS activity was found in a patient with SPG9B due to homozygous p.Ser242Asn mutation in the G5K domain; however, the P5CS protein level and its mitochondrial localization in HeLa cells transfected with the mutant ALDH18A1 plasmids remained unchanged [7]. In another patient with SPG9B due to compound heterozygous mutations in the G5PR domain of ALDH18A1 (p.Arg371Gln and p.Ser497Asn) residual activity of the P5CS was also observed [8]. Indeed, these findings may suggest that the SPG9B phenotype could be associated with residual P5CS activity, while some patients with biallelic null mutations in ALDH18A1 and cutis laxa phenotype exhibited pronounced reduction or total absence of P5CS protein [1,11,18,19].



Fig. 2. Mutations in the *ALDH18A1* gene. (A) A single nucleotide change (boxed) near the 5' UTR in the mutant compared to normal (wild type) sequence. The patient inherited this mutation from her mother (R = A/G). (B) A single nucleotide change (boxed) in the exon 4 in the mutant compared to normal (wild type) sequence. The patient inherited this mutation from her father (R = A/G). The mutated positions are highlighted blue in the DNA chromatograms. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

References	This study	[6]	[2]			[3]	
Number of affected siblings	1		2	1			4	
Gender	F	М	F	М	F	F	М	F
Ethnicity	Caucasian	Japanese	Japanese	Caucasian	Caucasian	Caucasian	Caucasian	Caucasian
Intron/exon location	Intron 1	Exon 2	Exon 2	Exon 3	Exon 4	Exon 4	Exon 4	Exon 4
	Exon 4	Exon 4	Exon 4	Exon 14	Exon 15	Exon 15	Exon 15	Exon 15
Nucleotide variation	c28-2A>G	c.30C>A	c.30C>A	c.251G>A	c.383G>A	c.383G>A	c.383G>A	c.383G>A
	c.383G>A	c.383G>A	c.383G>A	c.1741G>A	c.1910T>C	c.1910T>C	c.1910T>C	c.1910T>C
		(two homozygous mutations)	(two homozygous mutations)			1.0011	1. 10011	1 100X
Protein variation	?	p.Phe10Leu p.	p.Phe10Leu	p.Arg84Gln	p.Arg128His	p.Arg128His	p.Arg128His	p.Arg128His
	p.Arg128His	Arg128His	p.Arg128His	p.Glu581Lys	p.Leu63/Pro	p.Leu63/Pro	p.Leu63/Pro	p.Leu63/Pro
		(two homozygous mutations)	(two homozygous mutations)					
Protein domain	G5K/G5K	G5K/G5K	G5K/G5K	G5K/G5PR	G5K/G5PR	G5K/G5PR	G5K/G5PR	G5K/G5PR
Dysmorphic features	Yes	NA	NA	NA	Yes	No	Yes	Yes
Microcephaly	Yes	NA	NA	NA	Yes	Yes	No	No
Growth retardation	Yes	NA	NA	NA	Yes	Yes	No	Yes
Developmental delay	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Cognitive impairment								
Lower limb spasticity	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Ataxia	No	No	No	No	No	No	No	NA
Cerebellar signs								
Tremor (age at onset, years)	Yes (<1)	No	No	Yes (15)	No	No	No	NA
Epilepsy	No	No	No	Yes	No	No	No	No
Cutaneous findings	No	No	No	No	No	No	No	No
Ocular findings	No	No	No	No	NA	NA	Probable cataract	NA
Brain MRI	Corpus callosum hypoplasia Thin white matter	Normal	Normal	Normal	NA	NA	Thin corpus callosum Periventricular white matter anomalies Mild cortical atrophy	NA
Plasma amino acids (proline, citrulline, ornithine, arginine)	Normal	NA	NA	Normal	NA	NA	NA	NA

Table 1 Reported biallelic *ALDH18A1* variants associated with autosomal recessive spastic paraplegia (SPG9B).

(continued on next page)

References	[7]	[8]		[(5]	[3]	
Number of affected siblings	1	1 2			2	2	
Gender	F	М	Μ	М	М	М	Μ
Ethnicity	Chinese	Caucasian	Caucasian	Japanese	Japanese	Caucasian	Caucasian
Intron/exon location	Exon 7	Exon 10	Exon 10	Exon 12	Exon 12	Exon 17	Exon 17
		Exon 10	Exon 10	Exon 16	Exon 16		
Nucleotide variation	c.725G>A	c.1112G>A	c.1112G>A	c.1321C>T	c.1321C>T	c.2143G>C	c.2143G>C
	(homozygous)	c.1490G>A	c.1490G>A	c.1994G>A	c.1994G>A	(homozygous)	(homozygous)
Protein variation	p.Ser242Asn	p.Arg371Gln	p.	p.Arg441Ter	p.Arg441Ter	p.Asp715His	p.Asp715His
	(homozygous)	p.Ser497Asn	Arg371Gln	p.Arg665Gln	p.Arg665Gln	(homozygous)	(homozygous)
			p.Ser497Asn				
Protein domain	G5K	G5PR/G5PR	G5PR/	G5PR/GSPR	G5PR/GSPR	GSPR	GSPR
			G5PR				
Dysmorphic features	NA	Yes	Yes	NA	NA	Yes	Yes
Microcephaly	NA	Yes	Yes	NA	NA	Yes	Yes
Growth retardation	NA	Yes	Yes	NA	NA	NA	NA
Developmental delay	No	Yes	Yes	Yes	Yes	Yes	Yes
Cognitive impairment							
Lower limb spasticity	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Ataxia	No	No	No	Yes	Yes	NA	No
Cerebellar signs							
Tremor (age at onset, years)	No	No	No	No	No	Yes (7)	Yes (7)
Epilepsy	No	Yes	Yes	No	No	No	No
Cutaneous findings	No	No	No	No	No	No	No
Ocular findings	No	No	No	No	No	NA	NA
Brain MRI	Normal	Increase in the prominence of the	NA	Mild cerebellar	Mild cerebellar	Normal	NA
		cortical sulci		atrophy	atrophy		
Plasma amino acids (proline, citrulline,	NA	NA	NA	Normal	Normal	Normal	NA
ornithine, arginine)							

 Table 1 (continued)

 Reported biallelic ALDH18A1 variants associated with autosomal recessive spastic paraplegia (SPG9B).

Abbreviations: M: male, F: female, G5K: glutamate 5-kinase domain, G5PR: gamma-glutamyl phosphate reductase domain, NA: not available.

Measurements of blood ammonia and amino acid levels led to inconstant results. Increased blood ammonia and low plasma proline, ornithine, citrulline and arginine were described in both SPG9A and ARCL3A [3,10]. In contrast, the ammonia and amino acid levels were in the normal or low normal range in SPG9B patients regardless of whether the mutation affected the G5K, as in our case, the G5PR, or both domains [2,3,6]. The patient population with ALDH18A1-related disorders is small, hampering the comparison of the sometimes contradictory results gained by various methods in singular cases, or families with different phenotypes.

The pathophysiology of the neurological impairment in *ALDH18A*-related disorders remains to be clarified. Reduced cerebral proline and/or creatine synthesis might have a role [1,19], however moonlighting of P5CS protein cannot be ruled out [19,20]. Further research warranted to elucidate the mechanism of neurodegeneration in these conditions.

4. Conclusion

We report a girl with a rare form of autosomal recessive hereditary spastic paraplegia (SPG9B) due to compound heterozygous mutations in the *ALDH18A1* gene. The c.-28-2A>G variant in intron 1 is a novel mutation; it was inherited from her mother. The other, c.383G>A variant in exon 4, inherited from her father, has already been published. Vigorous infantile tremor preceding progressive spastic paraplegia was a unique clinical manifestation of the disease. Intrauterine and postnatal growth retardation, dysmorphic features, microcephaly, delayed development and intellectual disability were the other characteristic features of the disorder.

Acknowledgments

The authors are grateful to the patient's parents for their collaboration.

The authors thank to Professor Eva Morava-Kozicz MD, PhD for the amino acid analysis.

Funding

This study was supported by the GINOP-2.3.2-15-2 grant (TK and ZM) provided by the National Research, Development and Innovation Office (Hungary).

Ethical approval

Written informed parental consent has been obtained.

Written permission has been gained from the parents to publish the patient's photos in a scientific journal.

The study was approved by the Human Investigation Review Board at Albert Szent-Györgyi Clinical Centre, University of Szeged, Hungary.

Conflict of Interest Disclosures

The authors declare no competing interests.

References

- Marco-Marin C, Escamilla-Honrubia JM, Llácer JL, Seri M, Panza E, Rubio V. Δ¹-Pyrroline-5-carboxylate synthetase deficiency: An emergent multifaceted urea cycle-related disorder. J Inherit Metab Dis 2020. <u>https://doi.org/10.1002/jimd 12220</u>.
- [2] Steenhof M, Kibaek M, Larsen MJ, Christensen M, Lund AM, Brusgard K, et al. Compound heterozygous mutations in two different domains of *ALDH18A1* do not affect the amino acid levels in a patient with hereditary spastic paraplegia. Neurogenet 2018;19:145–9.
- [3] Coutelier M, Goizet C, Durr A, Habarou F, Morais S, Dionne-Laporte A, et al. Alteration of ornithine metabolism leads to dominant and recessive hereditary spastic paraplegia. Brain 2015;138:2191–205.
- [4] Hu CA, Lin WW, Obie C, Valle D. Molecular enzymology of mammalian Δ¹-pyrroline-5-carboxylate synthase. Alternative splice donor utilization generates isoforms with different sensitivity to ornithine inhibition. J Biol Chem 1999;274:6754–62.
- [5] Zombor M, Kalmár T, Nagy N, Berényi M, Telcs B, Maróti Z, et al. A novel *WDR62* missense mutation in microcephaly with abnormal cortical architecture and review of the literature. J Appl Genet 2019;60:151–62.
- [6] Koh K, Ishiura H, Beppu M, Shimazaki H, Ichinose Y, Mitsui J, et al. Novel mutations in the ALDH18A1 gene in complicated hereditary spastic paraplegia with cerebellar ataxia and cognitive impairment. J Hum Genet 2018;63:1009–13.
- [7] Wei Q, Dong HL, Pan LY, Chen CX, Yan YT, Wang RM, et al. Clinical features and genetic spectrum in Chinese patients with recessive hereditary spastic paraplegia. Transl Neurodegener 2019;8:19.
- [8] Magini P, Marco-Marin C, Escamilla-Honrubia JM, Martinelli D, Dionisi-Vici C, Faravelli F, et al. P5CS expression study in a new family with *ALDH18A1*-associated hereditary spastic paraplegia SPG9. Ann Clin Transl Neurol 2019;6:1533–40.
- [9] Torres-Russotto D. Clinical approach to tremor in children. Parkinsonism Relat Disord 2019;59:111–6.
- [10] Baumgartner MR, Rabier D, Nassogne MC, Dufier JL, Padovani JP, Kamoun P, et al. Δ^1 -pyrroline-5-carboxylate synthase deficiency: neurodegeneration, cataracts and reduced ornithine, citrulline, arginine and proline. Eur J Pediatr 2005;164:31–6.
- [11] Fischer B, Callewaert B, Schröter P, Coucke PJ, Schlack C, Ott CE, et al. Severe congenital cutis laxa with cardiovascular manifestations due to homozygous deletions in *ALDH18A1*. Mol Genet Metab 2014;112:310–6.
- [12] de Bot ST, van den Elzen RTM, Mensenkamp AR, Schelhaas HJ, Willemsen MAAP, Knoers NVAM, et al. Hereditary spastic paraplegia due to SPASTmutations in 151 Dutch patients: new clinical aspects and 27 novel mutations. J Neurol Neurosurg Psychiatry 2010;81:1073–8.
- [13] Anheim M, Lagier-Tourenne C, Stevanin G, Fleury M, Durr A, Namer IJ, et al. SPG11 spastic paraplegia. A new cause of juvenile parkinsonism. J Neurol 2009;256:104–8.
- [14] Schneider SA, Mummery CJ, Mehrabian M, Houlden H, Bain PG. SPG11 presenting with tremor. Tremor Other Hyperkinet Mov (NY) 2012;2 tre-02-104-666-1.

- [15] Wolthuis DFGJ, van Asbeck E, Mohamed M, Gardeitchik T, Lim-Melia ER, Wevers RA, et al. Cutis laxa, fat pads and retinopathy due to *ALDH18A1* mutation and review of the literature. Eur J Paediatr Neurol 2014;18:511–5.
- [16] Kremer LS, Bader DM, Mertes C, Kopajtich R, Pichler G, Iuso A, et al. Genetic diagnosis of Mendelian disorders via RNA sequencing. Nat Commun 2017;8:15824.
- [17] Handley MT, Mégarbane A, Meynert AM, Brown S, Freyer E, Taxlor MS, et al. Loss of *ALDH18A1* function is associated with a cellular lipid droplet phenotype suggesting a link between autosomal recessive cutis laxa type 3A and Warburg Micro syndrome. Mol Genet Genomic Med 2014;2:319–25.
- [18] Skidmore DL, Chitayat D, Morgan T, Hinek A, Fischer B, Dimopoulou A, et al. Further expansion of the phenotypic spectrum associated with mutations in *ALDH18A1*, encoding Δ^1 -pyrroline-5-carboxylate synthase [P5CS]. Am J Med Genet Part A 2011;155:1848–56.
- [19] Martinelli D, Häberle J, Rubio V, Giunta C, Hausser I, Carrozzo R, et al. Understanding pyrroline-5-carboxylate synthase deficiency: clinical, molecular, functional, and expression studies, structure-based analysis, and novel therapy with arginine. J Inherit Metab Dis 2012;35:761–76.
- [20] Jeffery C. Protein moonlighting: what is it, and why is it important?. Philos Trans R Soc B 2017;373:20160523.