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Research Article – Phytochemistry

Isolation, characterization and biological properties of betulin from *Entada africana* Guill. and Perr. (Mimosaceae)

A. Kwaji^{1*}, H. M. Adamu², I.Y. Chindo², Atiko, R¹

¹Department of Chemistry, PMB 127 Gombe State University, Gombe, Nigeria ²Department of Chemistry, PMB 0248 Abubakar Tafawa Balewa University, Bauchi, Nigeria

(Received: 01-03-2018; Accepted 24-03-2018; Published Online 26-03-2018) *Corresponding author E-mail: andrewkwaji@yahoo.com Tel: +2348036433710

Abstract

The present study is aimed at the isolation, characterization and evaluation of some biological properties of betulin from *Entada africana* stem bark extract. A dichloromethane soluble portion of the stem bark methanol/acetone (1:1 v/v) extract was subjected to gradient elution using ethyl acetate in hexane (5 – 30 %) on an open column. A pure compound was obtained with $R_f = 0.61$ in hexane/ethyl acetate (8:2 v/v) after repeated washing and recrystallization from methanol and coded Enac1. The pure compound was analyzed using IR, ¹H & ¹³C NMR and GC-MS. Clinical isolates of *Escherichia coli*, *Klebsiella pneumoniae*, *Salmonella typhi* and *Staphylococcus aureus* were used to assess the antibacterial activity of the pure compound while its preliminary Cytotoxicity was evaluated using brine shrimp nauplii. Based on the spectroscopic data obtained and in comparison with literature, the isolated compound was identified as betulin. The minimum inhibitory concentration (MIC) of betulin ranged between 62.50 - 250.00 µg/mL for all the four bacterial isolates in the study while its fifty percent lethal concentration (LC₅₀) was 10.00 µg/mL. Significant Cytotoxicity with moderate antibacterial activity was observed. The study therefore justifies the existence of bioactive compounds in the stem bark of *Entada africana* and its use in traditional medicine.

Keywords: Entada africana, isolation, betulin, characterization, cytotoxicity

Introduction

Entada africana is a West African medicinal plant that is quite useful in the treatment of many ailments and diseases such as fever, diabetes, diarrhea, hypertension, arrow poison and as tonic (Bako et al., 2005; Tibiri et al., 2010). The leaves, stem bark, roots, seeds and its gum resins are used in the management of several ailments in traditional medicine in different regions of West Africa such as Mali, Niger, Cameroon and Nigeria. Due to widespread use, the plant is now considered to be an endangered species (Banzouzi et al., 2007). Despite the existence of several reports on biological studies on the crude extracts of E. africana such as antileishmanial, antibacterial, antiinflammatory, antioxidant and wound healing properties (Tibiri et al., 2010; Mbatchou et al., 2011; Njayou et al., 2013; Marthe et al., 2014; Kwaji et al., 2017), only few reports exists on isolation of bioactive compounds such as the acidic wound healing polysaccharides (Diallo et al., 2001) and the antiproliferative triterpene saponins (Cioffi et al., 2006) from the root extracts of Entada africana. Therefore, this calls for further investigation of the bioactive constituents of Entada africana in order to understand more about its immense healing potential. Consequently the study is aimed at the isolation, characterization and evaluation of some biological properties of betulin from E. africana stem bark extract.

Materials and Methods

Isolation of Betulin from Entada africana Guill. and Perrott.

The isolation of compound was carried out in accordance with the method described by Teke *et al.* (2011)

with slight modification. Briefly, a dichloromethane soluble fraction (9.35 gram) of acetone/methanol (1:1 v/v) extract was loaded onto a silica gel 60 (60-200) column. Gradient elution was carried out on an open column with solvent combination of hexane/ethyl acetate (5-30%) to yield several fractions of 100 mL each. The different fractions were combined on their thin layer chromatography (TLC) profiles after concentration on a rotary evaporator at 45 °C. Fractions 27-32 was rechromatographed and eluted isocratically using silica gel 60 (70-230 mesh) with a solvent combination of petroleum ether/diethyl ether (7:3) and recrystallized from methanol to yield a pure compound, Enac1 (200 mg). The retention factor, Rf, value was found to be 0.61 in hexane/ethyl acetate (8:2). The pure compound was kept for spectroscopic analysis.

Analysis of Sample (Enac1)

Infrared spectrum of the sample was obtained and recorded on Perkin Elmer Universal ATR spectrum 100 FT-IR spectrometer, while the ¹H NMR and ¹³C NMR spectra were recorded on 400 MHz Bruker Avance II Ultrashield with the sample initially dissolved in deuterated chloroform (CDCl₃). The GC-MS analysis for molecular mass determination was carried out on Shimadzu GC-MS-QP2010SE. The complete analysis was done at the Department of Chemistry, University of Kwazulu-Natal, Durban South Africa.

Minimum Inhibitory Concentration

Modified macroboth resazurin assay was used to determine the minimum inhibitory concentration (MIC) as described by Satyajit *et al.* (2007) and Karuppusamy &

Rajasekaran, (2009). Briefly, two fold serial dilutions of plant extract (1.80 ml, 100 mg/mL) and gentamicin (1.80 ml, 1 mg/mL) in a set of 9 test tubes was carried out with Mueller Hinton broth. To each test tube was added 0.20 mL of standardized bacterial inoculums (10^6 cfu/mL). This was incubated for 12 hrs at 37°C for bacteria. Then 50 µL of 0.1% resazurin solution was added to each test tube and incubated for further 5 hrs. The MIC corresponds to the lowest concentration which prevented colour change from purple to pink in the test tubes.

Cytotoxicity Test

The brine shrimp lethality test was used to evaluate the preliminary cytotoxicity of Betulin. Brine shrimps nauplii were hatched from brine shrimp eggs in sterile artificial sea water (38 g/L NaCl and adjusted to pH 8.5 with 1M NaOH) in a plastic container with a porous middle partition through which hatched nauplii can move to an electrically illuminated zone of the container. A pinch of the brine shrimp eggs was added to one of the less illuminated compartment of the container and allowed to hatch overnight in the brine solution. Ten hatched nauplii per test tube were taken in 4.5 mL of brine solution. To each of the test tubes, 0.5 mL of either extract or standard at different concentrations (100 µg/mL, 1,000 µg/mL, 10,000 µg/mL) was added. Each test tube will now have a final concentration of 10 µg/mL, 100 µg/mL and 1000 µg/mL respectively. These were then incubated for 24 hrs under light. For each concentration, tests were carried out in triplicate. Percentage mortality rate for each concentration was calculated. The 50% lethality concentration was obtained using Probit analysis on software SPSS 16.0. Potassium dichromate was used as standard (Krishnaraju et al., 2005: Chaitali et al., 2010).

Results and Discussions

From 9.35 gram dichloromethane soluble fraction, the percentage recovery of Enac1 was found to be 2.10% as presented in Table 1 below. Enac1 was a powdery white substance that is soluble in hexane and chloroform.

 Table 1 Entada africana Isolated Compound

Sl. No.	Compound	Rf value	Mass of isolate	Percentage recovery
1	Enac1	0.61	200 mg	2.10 %

Enacl

The IR spectrum of Enac1(Table 2) showed a broad absorption band at 3331 cm⁻¹ and a sharp peak at 1042 corresponding to O-H and C-O stretching bands respectively. Other absorptions bands observed at 2925.43-2872.11 cm⁻¹ are typical of C-H bond stretching bands. A weak band at 1638.41 cm⁻¹ indicates the presence of a C=C absorption band (Elvira *et al.*, 2009; Correa *et al.*, 2014; Barakat & Saleh., 2016).

In the proton NMR (Table 3), a doublet of doublets was seen at δ 3.19 ppm which shows the presence of α -oriented hydrogen at C-3. Furthermore, doublets can also be seen at δ 4.67, 4.55 and 3.79 ppm together with a methyl group at δ 1.66 ppm clearly suggests the presence of a lupane-triterpene nucleus. This is further substantiated by the C-13 NMR spectrum (Table 3). The characteristic C=C double bond of a lupane type nucleus was observed as shifts at δ 150.98 and 109.32 ppm. Also observed are the typical chemical shifts at

δc 79.0 and 60.5 ppm of C-3 and C-28 to which hydroxyl (OH) groups are attached.

Table 2 FT-IR Spectrum of Enac1

Sl. No.	Frequency (cm ⁻¹)	Type of Vibration	
1	3331.91	O-H stretching	
2	2925.43	C-H stretching due to CH ₃ ,	
3	2872.11	CH ₂ stretching of the first ring	
4	1638.41	C=C stretching	
5	1379.57	CH ₃ and CH ₂ bending vibrations	
6	1042.68	C-O stretching due to CH ₂ OH	
7	1014.60	C-O stretching of methine carbon	
8	879.37	Ω (H-C-CH) of an umbrella alkene	

The Mass spectrum data of Enac1 (Table 4) revealed the molecular ion mass to be 442 [M^+ C₃0H₅₀O₂]. Subsequent fragment peaks of m/z 411, 234, 203, 189 and 81 confirm the presence of a lupane-type nucleus (Maryam *et al.*, 2013). Consequently based on comparison of spectra data with literature, Enac1 was found to be lup-20(29)-ene-3,28-diol with the molecular formula C₃₀H₅₀O₂ commonly known as betulin. The structural formula is as shown in Fig. 1 below. This is the first time that betulin is isolated from the stem bark extract of *Entada africana*.

Despite the discovery of this molecule about 200 years ago from other sources, there is still a growing interest in betulin due to its wide spectrum of biological activities such as antiviral, anticancer, antibacterial, anti-inflammatory (Maryam et al., 2013) and hepatoprotective with potential for application in several therapeutic domains such as the anti-infectives, oncology and immunology (Sylwia et al., 2015). A comparative study of the hepatoprotective effects of betulin, betulinic acid and oleanolic acid against ethanolinduced toxicity indicates that betulin exhibited the best hepatoprotective activity and even stimulated cell growth at 10-5 mM alcohol concentration. Lupeol, a structurally related triterpene to betulin reportedly offered hepatoprotective activity in cadmium-treated rats by improving tissue redox reaction and by inhibition of lipid peroxidation. The hepatoprotective mechanism of betulin is assumed to follow the same pattern (Agnieszka & Martyna, 2005). It is considered as the drug of the future partly due to its potency, availability and high selectivity index which are critical properties of a molecule with drug properties (Boryczka et al., 2013).



Fig. 1 Betulin Structure

Antibacterial Activity of Betulin

The lowest MIC for compound Enac1 was 62.50 $\mu\text{g/mL}$

Carbon	Literature		Enac1	Literature	DEPT
C-atom	δC	δC	δΗ	δΗ	Enac1
1	38.8	38.3			CH ₂
2	27.4	27.5		1.52	CH_2
3	79.0	79.0	3.19(1H, dd)	3.15	CH
4	38.3	38.1			С
5	55.4	55.3			CH
6	18.3	18.3			CH ₂
7	34.3	34.3			CH_2
8	41.0	40.8			С
9	50.6	50.5			CH
10	37.4	37.2			С
11	20.9	20.9			CH_2
12	25.6	25.2			CH_2
13	37.0	37.2			CH
14	42.8	42.8			С
15	27.1	27.4			CH_2
16	29.3	29.7			CH_2
17	47.8	48.0			С
18	47.8	48.3			CH
19	48.8	48.3			CH
20	150.6	150.9			С
21	29.8	29.9			CH_2
22	34.0	34.3			CH_2
23	28.0	28.0	0.95(3H, s)	0.99	CH ₃
24	15.3	15.4	0.77(3H, s)	0.80	CH ₃
25	16.1	16.1	0.81(3H, s)	1.00	CH ₃
26	16.1	16.1	0.95(3H, s)	1.02	CH ₃
27	14.7	14.6	1.01(3H, s)	1.04	CH ₃
28	60.8	60.5	3.70(1H, d)	3.56	CH_2
29	109.6	109.9	4.67(1H, s)	5.11	CH_2
30	19.4	19.3	1.66(3H, s)	1.82	CH ₃

Table 3 δ ¹³C and ¹H NMR for Enac1 and Literature Values

¹H- and ¹³C-NMR Appendix I (a & b).

 Table 4 Mass spectral data of Enac1

Enac1	442[M ⁺ C ₃₀ H ₅₀ O ₂], 411(5.8), 313(00), 288(04), 271(00), 245(06), 234(9.6), 220(7.7), 203(32.6), 189(100) , 175(38),
GC-MS	161(26), 147(42), 135(36.5), 121(0), 107(48), 95(42.2), 81(36.5), 69(28.8), 55(34.6), 41(30.7), 21(9.6)
m/z	

on Salmonella typhi while the highest MIC value was observed for Klebsiella pneumoniae and Staphylococcus aureus at 250 µg/mL. This implies that Enac1 is very active on S. typhi and therefore could potentially be used in the management of infections or diseases associated with S. typhi. Additionally, it may also be used as a bacteriostatic agent for K. pneumoniae and Staph. aureus related infections. The gentamicin (control) lowest MIC value of 3.96 µg/mL was observed for Staphylococcus aureus, a gram positive bacterium with a less complicated cell wall structure compared to the gram negative bacteria (E. coli, K. pneumonia and S. typhi). However, the highest MIC value of gentamicin was seen for Salmonella typhi at 31.25 µg/mL as expected (Table 5). According to Teke et al. (2011) classification of antibacterial activity, MIC values of 24.4 to 78.2 µg/mL is considered significant (i.e. MIC < 100 $\mu g/mL$), while 100 < MIC = 625 $\mu g/mL$ is moderate or weak (MIC > 625 μ g/mL) against various resistant pathogens. On the basis of such a classification, betulin may be considered to possess a significant to moderate antibacterial activity. Several compounds isolated from Entada abyssinica, a member of the same genus as Entada africana exhibited moderate activity against several pathogenic organisms. The reported antibacterial activity is consistent with those obtained in this study. Two of the isolated compounds (methyl-3,4,5-trihydroxybenzoate, and

benzene-1,2,3-triol) with MIC values ranging between 250 - $500 \mu g/mL$ demonstrated moderate antibacterial activity.

Table 5 Antibacterial of Activity of Betulin

Bacteria	Betulin	Gentamicin
	MIC (µg/mL)	MIC(µg/mL)
Escherichia coli	125	7.81
Kleb. pneumoniae	250	15.63
Salmonella typhi	62.50	31.25
Staph. aureus	250	3.96

Cytoxicity of Betulin

Probit analysis of the brine shrimp test data (Appendix Ic) of the isolated compound betulin gave an LC₅₀ of 10.00 μ g/mL while that of the covariate standard substance potassium dichromate gave LC₅₀ of 8.33 μ g/mL. According to Meyer *et al.* (1982) and McLaughlin *et al.* (1998) the isolate is cytotoxic and indicates the presence of a bioactive compound or a potential anticancer agent. Adoum (2009) reported that the crude chloroform fraction of *Entada sudanica* (syn. *Entada africana*) stem bark extract had quite significant cytotoxic effect with LC₅₀ value of 10 μ g/mL while the ethanol and aqueous fractions had LC₅₀ values of 8 μ g/mL and 6 μ g/mL respectively. The observed higher Cytotoxicity values of the crude extracts could be indicative of the synergistic (additive) effects of the various phytocompounds. In herbal medicine the plant had been

used for the treatment of cancer, ulcers and malaria diseases. This high cytotoxic nature of the plant justifies its use in traditional medicine as an anticancer agent both within and outside Nigeria by some native tribes (Adoum, 2009).

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Supplementary data

Supplementary data related to this article can be found at <u>https://doi.org/10.21839/jaar.2018.v3i2.138</u>

References

- Adoum, O. A. (2009). Determination of Toxicity Levels of Some Savannah Medicinal Plants Using Brine Shrimp Test. Bayero Journal of Pure and Applied Sciences, 2(1), 135-138.
- Agnieszka, S. & Martyna, K. (2005). Protective Effects of Betulin and Betulinic acid against Ethanol-induced Cytotoxicity in HepG2 Cells. *Pharmacological Reports*, 57, 588-595.
- Bako, S. P., Bakfur, M. J., John, I.& Bala E. I. (2005). Ethnomedicinal and Phytochemical Profile of Some Savannah Plant Species In Nigeria. *International Journal of Botany*,1(2). 147-150.
- Banzouzi, J. T., Tibiri, A., Traore, A., Nacoulma, G. A., Guissou, I. P. & Mbatchi, B. (2007): Toxicological Assessment of Methanolic Stem bark and Leaf Extracts of *Entada africana* Guill. and Perr. (Mimosaceae). *International Journal of Pharmacology*, 3(5):393-399.
- Barakat, K. & Saleh, M. (2016). Bioactive Betulin Produced by Marine Paecilomyces WE3-F. *Journal of Applied Pharmaceutical Sciences*, 6(03), 034-040.
- Boryczka, S., Bebenek, E., Wietrzyk, J., Kempinska, K., Jastrzebska, M., Kusz, D. (2013). Synthesis, Structure and Cytotoxic Activity of New Acetylenic Derivatives of Betulin. *Molecules*, 18, 4526-4543.
- Chaitali, H. V., Nikhil, S. M., Sonali, S. B.& Sandip, B. B. (2010). Cytotoxicity Screening of Selected Indian Medicinal Plants Using Brine Shrimp Lethality Bioassay. Advances In Natural Applied Sciences, 4(3). 389-395.
- Cioffi, G., Piaz, F. D., Caprarris, P. D., Sanogo, R., Marzocco, S., Autore, G. & Tommasi, N. D. (2006). Antiproliferative Triterpene Saponins from *Entada* africana. Journal of Natural Products, 69 (9), 1323-1329.
- Correa, G. M., Abreu, V. G., Martins, D. A., Takahashi, J. A., Fontoura, H. D., Cara, D. C., *et al.* (2014). Antiinflammatory and Antimicrobial Activities of Steroids and Triterpenes Isolated from Aerial parts of *Justicia acuminatissima* (Acanthaceae). *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(6), 75-81.

- Elvira, E. K., Kemal, D., Zdenka, K. & Emin, S. (2009). Identification and Isolation of Pharmacologically Active Triterpenes In Betuale cortex, *Betula pendula* Roth, Betulaceae. *Bosnian Journal of Basic Medical Sciences*,9(1), 31-38.
- Karuppusamy, S.& Rajasekaran, K. M. (2009). High throughput Antibacterial Screening of Plant Extracts by Resazurin Redoxwith Special Reference to Medicinal Plants of Western Ghats. *Global Journal of Pharmacology*,3(2). 63-68.
- Krishnaraju, A. V., Rao, T. V. N., Sundararaju, D., Vanisree, M., Tsay, H.& Subbaraju, G. V. (2005). Assessment of Bioactivity of Indian Medicinal Plants Using Brine Shrimp (Artemia salina) Lethality Assay. International Journal of Applied Science and Engineering,3(2).125-134.
- Kwaji, A., Adamu, H. M. & Chindo, I. Y. (2017). Phytochemical analysis, Antibacterial and Antioxidant activities of *Entada africana* Guill. and Perrott Stem bark extracts. *Research Journal of Chemical Sciences*, 7(10), 10-15.
- Marthe, E. S. T, Aime, G. F., Armelle, T. M., Ernestine, T. N., Jackson, A. S., Francesco, K. T.*et al.* (2014). Activities of Selected Medicinal Plants against Multidrug resistant Gram negative bacteria in Cameroon.*African Health Sciences*, 14(1).167-172.
- Maryam, A., Ghazala, H. R., Faryal, V. M., Iffat, M., Viqar, U. A. & Shaukat, M. (2013). A Triterpenoid Antioxidant Agent Found In *Holoptelea Integrifolia* (Roxb) Planch. *International Journal of Pharmaceutical, Chemical and Biological Sciences*, 3(1), 63-67.
- Mbatchou, V. C., Ayebila, A. J.& Apea, O. B. (2011). Antibacterial Activity of Phytochemicals FromAcacia nilotica, Entada africana and Mimosa nigra L. on Salmoella typhi. Journal of Animal and Plant Sciences, 10(1). 1248-1258.
- Njayou, F. N., Aboudi, E. C. E., Tandjang, M. K., Tchana, A. K., Ngadjui, B. T., Moundipa, P. F. (2013). Hepatoprotective and Antioxidant Activities of Stem bark extract of *Khaya grandifoliola* (Welw) CDC and *Entada africana* Guill. Et Perr. *Journal of Natural Products*, **6**.73-80.
- Satyajit, D. S., Lutfun, N.& Yashodharan, K. (2007). Microtitre plate based antibacterial assay incorporating resazurin as an indicator of cell growth and its application in *in-vitro* antibacterial screening of phytochemicals. *Sciencedirect, Methods*, **42**. 321-324.
- Tibiri, A., Sawadogo, R. W., Ouedraogo, Banzouzi, J. T., Guissou, I. P., Nacoulma, G. O. (2010). Evaluation of Antioxidant Activity, Total Phenolic and Flavonoid Content of *Entada africana* Guill. Et Perr. (Mimosaceae) Organ Extracts. *Research Journal of Medical Sciences*,4(2). 81-87.