

THE UNIVERSITY of EDINBURGH

Edinburgh Research Explorer

Soil carbon loss by experimental warming in a tropical forest

Citation for published version:

Nottingham, A, Meir, P, Velasquez, E & Turner, BL 2020, 'Soil carbon loss by experimental warming in a tropical forest', *Nature*. https://doi.org/10.1038/s41586-020-2566-4

Digital Object Identifier (DOI):

10.1038/s41586-020-2566-4

Link: Link to publication record in Edinburgh Research Explorer

Document Version: Peer reviewed version

Published In: Nature

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Article to Nature; accepted 6 March with only editorial corrections needed.
 2

Soil carbon loss from a tropical forest in response to *in situ* experimental warming

7 Andrew T. Nottingham^{1,2}, Patrick Meir^{1,3}, Esther Velasquez², Benjamin L. Turner²

¹School of Geosciences, University of Edinburgh, Crew Building, Kings Buildings, Edinburgh EH9
3FF, UK

10 ²Smithsonian Tropical Research Institute, 0843-03092, Balboa, Ancon, Republic of Panama

³*Research School of Biology, Australian National University, Canberra, ACT 2601, Australia*

12

3

4 5 6

Tropical soils contain a third of global soil carbon¹, so destabilization of soil organic matter 13 14 caused by the approximate 4°C warming predicted for tropical regions this century could 15 accelerate climate change by releasing additional carbon dioxide (CO₂) to the atmosphere²⁻⁵. 16 Theory predicts that warming should cause only modest carbon loss in tropical soils relative to 17 those at higher latitudes^{4,6}, but there have been no warming experiments in tropical forests to 18 test this prediction⁷. Here we show that *in situ* experimental warming of a lowland tropical forest 19 soil on Barro Colorado Island, Panama, caused an unexpectedly large increase in soil CO2 20 emissions. Two years of warming of the whole soil profile by 4°C increased CO₂ emission by 55% 21 compared to soils at ambient temperature. The additional CO₂ originated from heterotrophic rather than autotrophic sources and equated to a loss of 8.2 \pm 4.2 (\pm 1 SE) Mg C ha⁻¹ vr⁻¹ from 22 23 the breakdown of soil organic matter. During this time, we detected no acclimation of respiration 24 rates, no thermal compensation or change in temperature sensitivity of enzyme activities, and 25 no change in microbial carbon-use efficiency. These results demonstrate a high sensitivity of soil 26 carbon in tropical forests to warming, which represents a potentially substantial positive 27 feedback to climate change.

Tropical forests play a large role in the global carbon (C) cycle, because they exchange more CO₂ with the atmosphere than any other ecosystem, contain over two-thirds of terrestrial plant biomass⁸ and harbour over a quarter of global soil C (ref. 1). Between 30 and 50% of the C respired from tropical forests originates from soil, most of which is derived from the decomposition of organic matter⁹⁻¹¹. Thus, even a small increase in respiration from tropical forest soils could have a large effect on atmospheric CO₂ concentrations, with consequences for global climate.

34 There is considerable concern that increased global temperatures will destabilize soil C and increase the flux of CO_2 from soil to the atmosphere²⁻⁵. Experiments in temperate and arctic regions 35 have consistently found that short-term (< 2 years) warming increases the soil CO₂ efflux by an average 36 of $46 \pm 8\%$ compared to soil at ambient temperature^{2,3,5,12}. For the tropics, it is expected that the 37 38 response of soil C to warming will be smaller than at higher latitudes, because kinetic theory predicts 39 that the intrinsic temperature sensitivity of reaction rates is reduced at higher temperatures^{4,13}, and 40 meta-analyses of warming experiments have shown that the temperature sensitivity of soil C loss increases with latitude⁶. However, the extent to which intrinsic temperature sensitivity translates into 41 42 actual ('apparent') temperature sensitivity depends on co-variation of other environmental factors that influence respiration, such as soil moisture and substrate availability^{4,14}. As there have been no *in situ* 43 44 warming experiments conducted in tropical forests, the apparent temperature sensitivity of soil organic 45 matter in this biome remains unknown. As a result, earth-system models continue to use kinetic theory to define the temperature sensitivity of soil C^{15} , limiting how they predict the response of tropical 46 forests to global environmental change^{16,17}. 47

Several factors could influence the apparent temperature sensitivity of soil organic matter. For example, soil warming is typically accompanied by soil drying, which can either reduce respiration in aerobic soils by reducing water availability, or increase respiration in waterlogged soils by increasing oxygen availability^{4,7,14}. Warming can also affect respiration rates by inducing changes in biotic processes, such as the physiological response or community composition of microbes, or changes in substrate availability to decomposers¹³. In experiments performed at higher latitudes, temperatureadaptive or compensatory responses of microbial communities and enzyme activities have been shown to modulate the effect of warming on the soil C cycle^{2,13}. In the tropics, future novel warm temperature maxima could exceed critical biochemical thresholds^{7,18}, with added complexity emerging from altered interactions among species-rich plant and microbial communities¹⁹, and from covarying changes in hydrological and nutrient cycles⁷.

59 Here, we present results from the first soil warming experiment in a lowland tropical forest (SWELTR: Soil Warming Experiment in Lowland Tropical Rainforest). The experiment tests the 60 response of the whole soil profile to the 4°C warming predicted for tropical latitudes by the end of this 61 century¹⁶ (Fig. 1A; Extended Data Figs. 1-3). SWELTR consists of five pairs of circular control and 62 63 warmed plots, evenly distributed within approximately 1 ha of seasonally-moist lowland tropical forest 64 on Barro Colorado Island, Panama. The soils are moderately weathered Dystric Eutrudepts (Inceptisols) that have developed on the volcanic facies of the Bohio Formation (Extended Data Table 65 1, see methods). Each warmed plot has a ground surface area of $\sim 20 \text{ m}^2$ heated to 1.2 m depth, resulting 66 in a total of 120 m³ of warmed soil across the experiment. 67

Two years of experimental warming increased soil CO₂ emissions by 55%, from 18.8 ± 1.9 Mg 68 C ha⁻¹ yr⁻¹ in control plots to 29.2 ± 5.0 Mg C ha⁻¹ yr⁻¹ in warmed plots (treatment effect, p < 0.05; Fig. 69 70 2; Extended Data Table 2). The soil CO₂ emission rate from unheated (i.e., control) plots is representative of tropical forests worldwide (8-40 Mg C ha⁻¹ yr⁻¹), including in the Amazon basin (12-71 24 Mg C ha⁻¹ yr⁻¹)²⁰. Using exclusion and ingrowth cores to partition respiration from heterotrophic 72 73 (soil-derived) and autotrophic (root-derived) sources, we find that the increase in CO₂ efflux was 74 derived predominantly from heterotrophic sources, whether from the decomposition of fresh-litter 75 inputs or pre-existing soil organic matter (Figs. 2-3, Extended Data Fig. 5). Soil-derived respiration increased from 12.0 ± 2.1 Mg C ha⁻¹ yr⁻¹ in control plots to 20.1 ± 4.2 Mg C ha⁻¹ yr⁻¹ in warmed plots 76 77 (a 68% increase of 8.2 Mg C ha⁻¹ yr⁻¹; treatment effect, p < 0.05), while root-derived respiration was not altered significantly (p = 0.21; 6.8 ± 1.2 and 9.0 ± 3.4 Mg C ha⁻¹ yr⁻¹ in control and warmed plots, respectively; Fig. 3, Extended Data Table 3).

80 It is possible that the large warming-induced increase in soil CO₂ efflux was due in part to soil 81 drying, because the warmed plots were slightly drier than the controls, particularly in the early wet 82 season (Extended Data Fig. 3). In wet soils, soil drying can increase respiration by increasing the 83 supply of oxygen to heterotrophic microbes. In contrast, soil drying under aerobic conditions can reduce heterotrophic respiration by promoting water limitation²⁰. Here, we find a marginally non-84 significant effect of soil moisture on CO_2 efflux across seasons (Extended Data Table 2; annual p =85 86 0.69, wet-season p = 0.07 and dry-season p = 0.06), consistent with the parabolic relationship of CO₂ flux with soil moisture for this site²⁰. However, there was no direct effect of warming on soil moisture 87 88 (for surface soils p = 0.19, whole-profile p = 0.24; Extended Data Tables 4-5), and the interaction 89 between soil moisture and warming in the CO₂ efflux model was not significant (annually and for 90 individual seasons; p > 0.2; Extended Data Table 2), indicating that the warming effect on CO₂ efflux 91 was not influenced by soil moisture. Furthermore, soil moisture was not correlated with soil CO₂ efflux 92 in the warmed plots (Extended Data Fig. 3); and drying during the early wet season in warmed soil 93 (Extended Data Fig. 3) should decrease rather than increase CO₂ efflux, because the soil was aerobic during this period and below the moisture content of $0.45 \text{ m}^3 \text{m}^{-3}$ at which soil CO₂ efflux peaks in this 94 95 forest²⁰. Our data thus show that although soil moisture influenced soil CO₂ efflux and that warmed 96 plots were slightly drier than control plots, particularly during the early wet season, this did not 97 contribute significantly to the increased CO₂ efflux from warmed soil.

98 There was no moderation of the warming-induced increase in soil CO₂ efflux over the two 99 years of the experiment. Such a moderation might be expected in the long-term, whether through 100 substrate limitation, adaptation of microbial communities (through changes in microbial carbon-use-101 efficiency; CUE) or thermal compensation of enzyme activities (reduced V_{max} at higher 102 temperatures)^{2,13,21}. We found no reduction in extractable or mineralized nitrogen or phosphorus with 103 warming, as would be expected under nutrient limitation (Extended Data Figs. 6-7). Almost all 104 hydrolytic enzymes were unaffected by warming (Extended Data Figs. 6-7), except for β-xylanase – 105 an enzyme involved in hemicellulose degradation – for which activity increased with warming during the wet season, an opposite response to that predicted by thermal compensation¹⁹. The temperature 106 107 sensitivity of enzyme activity (Q_{10} of V_{max}) was unaffected by warming (Extended Data Fig. 8), 108 indicating no dampening effect on soil C breakdown as a result of decreased enzyme flexibility (which 109 would cause decreased affinity of the enzyme for its substrate), as expected at warmer temperatures²². 110 Microbial CUE, which broadly represents C stabilised in biomass relative to C lost in respiration and can promote long-term (decadal) soil C loss^{2,21}, was unaffected by warming (Treatment effect, p =111 112 0.37; Extended Data Figs. 6-7). Indeed, microbial C increased with warming at the annual scale 113 (treatment effect, annual scale, p = 0.02; with a marginally non-significant increase at the seasonal scale, p < 0.1 Extended Data Fig. 7), suggesting slightly increased growth in response to greater 114 organic matter turnover in the absence of nutrient constraints to C degradation, which did not translate 115 116 into changes in CUE.

117 Our finding that tropical forest soil C has a high apparent temperature sensitivity challenges 118 the prevailing expectation that the temperature sensitivity of soil C is lower in the tropics compared to cooler ecosystems at higher latitudes^{4,6}. The 55% increase in total soil CO₂ emissions we report here 119 120 following two years of 4°C whole-profile warming (18.8 to 29.2 Mg C ha⁻¹ yr⁻¹) is larger than that 121 found in a temperate forest using a similar whole-soil-profile experimental design (34-37% increase over two years of 4°C whole-profile warming; 13 to 17.5 Mg C ha⁻¹ yr⁻¹)⁵. In addition, the rate of 122 123 additional soil C loss (8.2 Mg C ha⁻¹ yr⁻¹) is greater than for all the studies in a recent meta-analysis of surface-only soil warming experiments at higher latitude sites (all loss rates $< 5 \text{ Mg C} \text{ ha}^{-1} \text{ yr}^{-1}$)³. The 124 125 expectation that the temperature sensitivity of soil C breakdown is lower in the tropics compared to 126 higher latitudes, based in part on kinetic theory and commonly used to describe soil C responses in earth-system models¹⁵, is therefore not consistent with the ('apparent')^{4,14} temperature sensitivity of 127

the breakdown of tropical forest soil C reported here. Our results are, however, consistent with recent atmospheric and satellite measurements, which have shown a high sensitivity of ecosystem-scale C cycling in tropical regions in response to interannual temperature variation^{23,24}. Our findings suggest that soils contribute a major component of these ecosystem-scale responses to warming.

132 This high apparent temperature sensitivity of tropical forest soil C under in situ experimental 133 warming must arise through the temperature response of covarying ecosystem properties rather than 134 as the sole consequence of intrinsic kinetic processes. Although our data do not provide conclusive 135 mechanistic evidence for the marked increase in soil C loss from warmed soil, several findings point 136 to a possible explanation: i) the general absence of thermal compensation in enzyme activities (no 137 decrease in V_{max} ; ii) no change in the temperature sensitivity of enzymes under warming (no decrease in Q_{10} of V_{max}); and iii) no moderating thermal response of microbial CUE (Extended Data Figs. 6-138 7)^{21,25,26}. Together, our results indicate that organic matter degradation increased under warming with 139 140 no moderating responses or acclimation during our study period among microbial communities, or the 141 enzymes they synthesise.

142 This surprisingly large loss of soil C from warmed soil represents a substantial positive climate feedback over the period of this study. The additional C loss from warming observed here is of similar 143 magnitude to annual litterfall-C inputs at this site (5-7 Mg C ha⁻¹ yr⁻¹)²⁷ and is equivalent to 144 approximately 13% of the total soil C stock, or 30% of gross primary productivity (27.5 Mg C ha⁻¹ yr⁻ 145 1)²⁸. Extrapolation of the first two years of C loss in our experiment across the entire tropical forest 146 soil C stock (502 Pg C)¹ indicates a global loss of >65 Pg C with 4°C warming this century, which is 147 148 consistent with estimated C loss based on a five-year soil translocation experiment in tropical forests elsewhere²⁹. In the light of these findings, earlier estimates of global soil C loss under 4°C warming, 149 which were based on experiments performed at higher latitudes $(120-190 \text{ Pg C})^{2,30}$, underestimate the 150 151 magnitude of this global earth-atmosphere feedback.

We expect that the rate of soil C loss will eventually decline in warmed soils as substrate limitation increases, but we do not know how long this will take, nor whether the long-term soil C balance will be affected by plant-soil interactions or changes in soil microbial communities as they adapt to warmer temperatures^{2,12,13,21}. The nature of these longer-term responses will determine the strength of this positive earth-atmosphere feedback, already significant in the short-term, in contributing to further climate warming.

159 **References**

160

- Jackson, R. B. *et al.* The ecology of soil carbon: pools, vulnerabilities, and biotic and
 abiotic controls. *Annual Review of Ecology, Evolution, and Systematics* 48, 419-445,
 doi:10.1146/annurev-ecolsys-112414-054234 (2017).
- Melillo, J. M. *et al.* Long-term pattern and magnitude of soil carbon feedback to the
 climate system in a warming world. *Science* 358, 101-104,
 doi:10.1126/science.aan2874 (2017).
- van Gestel, N. *et al.* Predicting soil carbon loss with warming. *Nature* 554, E4-E5,
 doi:10.1038/nature20150 (2018).
- 169 4 Davidson, E. A. & Janssens, I. A. Temperature sensitivity of soil carbon decomposition
 170 and feedbacks to climate change. *Nature* 440, 165-173, doi:10.1038/nature04514
 171 (2006).
- 1725Hicks Pries, C. E., Castanha, C., Porras, R. C. & Torn, M. S. The whole-soil carbon flux in173response to warming. *Science* **355**, 1420-1422, doi: 10.1126/science.aal1319 (2017).
- 174 6 Carey, J. C. *et al.* Temperature response of soil respiration largely unaltered with
 175 experimental warming. *P Natl Acad Sci USA* **113**, 13797-13802,
 176 doi:10.1073/pnas.160536511 (2016).
- Wood, T. E. et al. in Ecosystem Consequences of Soil Warming: Microbes, Vegetation,
 Fauna and Soil Biogeochemistry (ed J. Mohan) Ch. 14, 385-439 (Academic Press, 2)
- *Fauna and Soil Biogeochemistry* (ed J. Mohan) Ch. 14, 385-439 (Academic Press, 2019).
 Pan, Y. *et al.* A large and persistent carbon sink in the world's forests. *Science* 333, 988-
- 180 993, doi:10.1126/science.1201609 (2011).
- Anderson-Teixeira, K. J., Wang, M. M. H., McGarvey, J. C. & LeBauer, D. S. Carbon
 dynamics of mature and regrowth tropical forests derived from a pantropical database
 (TropForC-db). *Global Change Biol* 22, 1690-1709, doi:10.1111/gcb.13226 (2016).
- 18410Malhi, Y. The productivity, metabolism and carbon cycle of tropical forest vegetation. J185Ecol 100, 65-75, doi:10.1111/j.1365-2745.2011.01916.x (2012).
- 186 11 Chambers, J. Q. *et al.* Respiration from a tropical forest ecosystem: Partitioning of
 187 sources and low carbon use efficiency. *Ecol Appl* 14, S72-S88 (2004). doi: 10.1890/016012.
- 189 12 Romero-Olivares, A. L., Allison, S. D. & Treseder, K. K. Soil microbes and their response
 190 to experimental warming over time: A meta-analysis of field studies. *Soil Biol Biochem*191 107, 32-40, doi:10.1016/j.soilbio.2016.12.026 (2017).

- 19213Tang, J. et al. in Ecosystem Consequences of Soil Warming: Microbes, Vegetation, Fauna193and Soil Biogeochemistry (ed J. Mohan) Ch. 8, 175-201 (Academic Press, 2019).
- 194 14 Nottingham, A. T. *et al.* Climate warming and soil carbon in tropical forests: insights
 195 from an elevation gradient in the Peruvian Andes. *Bioscience* 65, 906-921,
 196 doi:10.1093/biosci/biv109 (2015).
- 197 15 Todd-Brown, K. E. O. *et al.* Causes of variation in soil carbon predictions from CMIP5
 198 Earth system models and comparison with observations. *Biogeosciences* 10, 1717–1736,
 199 doi:10.5194/bgd-9-14437-2012 (2013).
- 16 Hoegh-Guldberg, O. et al. in Global Warming of 1.5°C. An IPCC Special Report on the
 impacts of global warming of 1.5°C above pre-industrial levels and related global
 greenhouse gas emission pathways, in the context of strengthening the global response to
 the threat of climate change, sustainable development, and efforts to eradicate poverty
 (Cambridge University Press, 2018).
- 20517Cox, P. M. *et al.* Sensitivity of tropical carbon to climate change constrained by carbon206dioxide variability. *Nature* **494**, 341-344, doi:10.1038/Nature11882 (2013).
- 207 18 Mora, C. *et al.* The projected timing of climate departure from recent variability. *Nature*208 502, 183, doi: 10.1038/Nature12540 (2013).
- 20919Steidinger, B. S. *et al.* Climatic controls of decomposition drive the global biogeography210of forest-tree symbioses. *Nature* **569**, 404, doi:10.1038/s41586-019-1128-0 (2019).
- 211 20 Rubio, V. E. & Detto, M. Spatiotemporal variability of soil respiration in a seasonal
 212 tropical forest. *Ecol Evol* 7, 7104-7116, doi:10.1002/ece3.3267 (2017).
- 213 21 Frey, S. D., Lee, J., Melillo, J. M. & Six, J. The temperature response of soil microbial
 214 efficiency and its feedback to climate. *Nat Clim Change* 3, 395-398,
 215 doi:10.1038/Nclimate1796 (2013).
- 21622Bradford, M. A. Thermal adaptation of decomposer communities in warming soils. Front217Microbiol 4, doi:Artn 333, doi 10.3389/Fmicb.2013.00333 (2013).
- 218 23 Wang, X. H. *et al.* A two-fold increase of carbon cycle sensitivity to tropical temperature
 219 variations. *Nature* 506, 212–215, doi:10.1038/nature12915 (2014).
- 22024Liu, J. J. *et al.* Contrasting carbon cycle responses of the tropical continents to the 2015-2212016 El Nino. Science **358**, doi:10.1126/science.aam5690 (2017).
- 22225Karhu, K. *et al.* Temperature sensitivity of soil respiration rates enhanced by microbial223community response. *Nature* **513**, 81-84, doi:10.1038/nature13604 (2014).
- 224 26 Dacal, M., Bradford, M. A., Plaza, C., Maestre, F. T. & Garcia-Palacios, P. Soil microbial
 225 respiration adapts to ambient temperature in global drylands. *Nat Ecol Evol* 3, 232-+,
 226 doi:10.1038/s41559-018-0770-5 (2019).
- 227 27 Wieder, R. K. & Wright, S. J. Tropical forest litter dynamics and dry season irrigation on
 228 Barro Colorado Island, Panama. *Ecology* 76, 1971-1979, doi: 10.2307/1940727 (1995).
- 229 28 Chave, J. *et al.* Spatial and temporal variation of biomass in a tropical forest: results from
 230 a large census plot in Panama. *J Ecol* **91**, 240-252, doi: 10.1046/j.1365231 2745.2003.00757.x (2003).
- Nottingham, A. T. *et al.* Microbial responses to warming enhance soil carbon loss
 following translocation across a tropical forest elevation gradient. *Ecol Lett* 22, 18891899, doi:10.1111/ele.13379 (2019).
- 235 30 Crowther, T. W. *et al.* Quantifying global soil carbon losses in response to warming.
 236 *Nature* 540, 104-108, doi:10.1038/nature20150 (2016).
 237
- 238 Supplementary Information is linked to the online version of the paper
- 239

240	Acknowledgements This study was supported by a European Union Marie-Curie Fellowship FP7-2012-		
241	329360 and a Smithsonian Tropical Research Institute Tupper Fellowship to ATN, a Smithsonian Institution		
242	Scholarly Studies Grant to BT and Klaus Winter, and a NERC grant NE/K01627X/1 (NERC HTMF-BALI) to		
243	PM, who was also supported by an Australian Research Council (ARC) grant DP170104091. We thank Oris		
244	Acevado, Dayana Agudo, Aleksandra Bielnicka, Melissa Cano, David Dominguez, Milton Garcia, Matthew		
245	Larsen, Maria Jose Montero, Julio Rodriguez, Irene Torres, William Wcislo, Klaus Winter and Joe Wright for		
246	their contributions to SWELTR.		
247			
248	Author contributions ATN conceived the study with BLT and PM. ATN, EV and BLT performed the		
249	study. ATN analysed the data and wrote the paper with BLT and PM.		
250			
251	Author Information The authors declare no competing financial interests. Reprints and permissions		
252	information is available at www.nature.com/reprints. Correspondence and requests for materials should be		
253	addressed to A.T.N. (anotting@ed.ac.uk).		
254			
255	Data availability statement The data supporting the findings of this study are available within the paper and		
256	its supplementary information files, with further information available from the corresponding author upon		
257	reasonable request.		
258			
259	Supplementary Information and Extended Data for this manuscript include the following:		
260	Supplementary Materials and Methods		
261	Extended Data Table 1 to 5		
262	Extended Data Figs. 1 to 8		
263			
264			
265			





Figure 1 Mean differences in (a) soil temperature and (b) soil moisture content between control and warmed plots by depth. Data are the means of continuous measurements from integrated soil temperature and moisture probes (Campbell CS655) for the two-year period after the warming treatment began (Dec 2016 – Dec 2018). The error bars represent one standard error of the temporal variation across five plots (Treatment effect p < 0.001 for all comparisons, based on the temporal variation across n = 5 plots). The vertical lines show the mean soil temperature across the soil profile, which is 26.18°C for control plots and 30.14°C for warmed plots (3.97°C difference).

- 274
- 275
- 276
- 277
- 278
- 270
- 279
- 280
- 281











Figure 3 | The annual soil CO₂ efflux (total), partitioned into soil-derived and root-derived components. a. The pre-treatment period (Jan-May 2016; predominantly dry-season). b. The two years with warming. The figures show total CO₂ (Total) and the root-derived and soil-derived components calculated using CO₂ efflux from partition cores (equations 1-2). Differences between control and warmed plots are shown by asterisks where $p \le 0.05$ or as non-significant (ns); determined using mixed effect models (Extended Data Tables 2-3). The error bars represent one standard error of the variation across the five plots over the study period.

318 **METHODS**

319

320 Site and experiment. The experiment is situated in approximately 1 ha area of seasonally moist lowland tropical forest on Barro Colorado Island, Panama³¹. Within the plot area the dominant tree 321 species include Anarcardium excelsum and Poulsenia armata. The soils are Inceptisols (Fine, 322 323 isohyperthermic, Dystric Eutrudepts) that are rich in clay (~54% profile-weighted clay concentration) 324 and secondary metal oxides and are developed on the volcanic facies of the Bohio Formation, a basaltic conglomerate of Oligocene age³². Although these moderately weathered Inceptisols are less infertile 325 326 than soils under large areas of lowland tropical forests, Inceptisols still account for 14% of total land area in the tropics (Ultisols and Oxisols account for 20% and 23%, respectively)³³, and soil respiration 327 328 in the control plots in our experiment is comparable to that in lowland tropical forests in general, 329 including those on Ultisols and Oxisols (see main text, ref. 20).

330 The SWELTR experiment consists of 10 circular plots (five paired plots 'warm' and 'control'). 331 Each plot measures 5 m diameter, with approximately 10 m between each plot-pair and a minimum of 332 20 m between different plot-pairs. The warmed plots contain two heating structures, each consisting 333 of eight 1-m long stainless-steel rods, connected by approximately 50 cm of flexible stainless-steel 334 conduit. We used stainless-steel T-junctions at the top of each rod (adjoining the flexible conduit) and 335 conical caps at the bottom of each rod. The final structure was 1.2 m tall. Inside each of the structures, 336 we threaded 25 m of heating cable (SLMCAB10120BF, Briskheat, Columbus, USA) and filled the 337 remaining space in the rods and conduit with quartz sand, selected for its high thermal conductivity. 338 The complete structure was welded to seal the heating cable and sand inside. Two of these structures 339 were buried around a 3.5 diameter circumference, with the top of the flexible conduit 5 cm 340 belowground. Thus, each plot contained 50 m of heating cable inserted to 1.2 m depth, encircling a 3.5 m diameter area; with an effective heated plot area of 5 m diameter. The experiment heats 341

approximately 120 m³ soil in total (5 plots x 5 m diameter by 1.2 m depth). The plot design and heating
methodology follows that of Hanson et al. (ref. 34) and Hicks Pries et al. (ref. 5).

344

345 Temperature control. Each warming and control plot was connected to a thermostat system, which 346 maintained soil temperature in the warmed plots at 4°C above ambient temperature. The thermostat 347 system consisted of three integrated temperature and moisture sensors per plot (CS655 Reflectometer, 348 Campbell Scientific) inserted to 0-20, 50-70 and 100-120 cm depth at the mid-radius point in each 349 plot, which were connected to a control unit (one control unit for each plot pair; five in total). The 350 control units consisted of waterproof (IP68) enclosures containing a solid-state power controller 351 (DA10-24C0-0000, Watlow), relay (12V single channel), datalogger (CR1000, Campbell Scientific) 352 and 12V and 120V power supply. Temperature in each warmed plot was therefore maintained at 4°C 353 above the temperature in each corresponding paired control plot, based on the average temperature 354 from 0-120 cm depth at the mid-radius point in each plot. The average temperature differential over 355 two years was 3.97°C, which was the average of 2.7°C at 0-20 cm depth, 4.0°C at 50-70 cm depth and 356 5.2°C at 100-120 cm depth.

357 This experimental design has been shown to warm the soil approximately uniformly across the 358 soil volume, with minor anomalies of warmer soil very close to the heating rods (< 10 cm) and slightly cooler surface soils due to heat-transfer to the air^{5, 34}. Therefore, surface soils were slightly cooler 359 360 compared to subsoils, although the response of surface rather than subsoils will likely dominate the 361 warming response across the soil profile because they contain greater organic matter (two thirds of the 362 C stock occurs in the upper 50 cm of the soil profile; Extended data Table 1). The heating structures 363 were installed during May-July 2016, and plots were tested during June-October 2016. The testing 364 phase consisted of heating each plot by 4°C for approximately 2-week periods. The experiment was 365 switched on in full on 1 November 2016.

366

367 Soil gas-exchange and partitioning. Soil CO₂ efflux was measured every two weeks at four 368 systematically distributed locations within each plot from 2016 until 2019 using an infra-red gas 369 analyser (IRGA Li-8100; LI-COR Biosciences, Nebraska, USA). The soil collars for soil CO₂ efflux 370 measurements were assigned to zones within each plot ('centre 1', centre 2', 'side 1' and 'side 2') and 371 were relocated randomly within each zone every three months, for long-term within-plot spatial 372 independence. Soil CO₂ efflux was also measured every two weeks for four root-partition cores per plot ($2 \times$ root-exclusion and $2 \times$ root-ingrowth) to determine soil- and root-derived components of the 373 374 CO₂ efflux. At the same time as soil CO₂ efflux measurements, we measured soil temperature (using 375 a HI98509 thermometer probe; Hanna Instruments, USA) and soil moisture (using a Thetaprobe; 376 Delta-T, Cambridge, UK) at 0-20 cm soil depth for a random location within 1 m radius of each soil 377 collar, or within the root-partition cores.

Root-exclusion cores were made from PVC tubing (30 cm height, 10 cm diameter) with a 1 μ m nylon mesh base for drainage. Root-ingrowth cores (i.e. disturbance controls) had additional windows (~340 cm²) covered with 2 mm mesh around the sides³⁵. In each plot, two root-exclusion cores and two root-ingrowth cores were buried within each plot, approximately 30 cm from the heating cable (where the soil profile is warmed on average by 4°C).

The percentage contributions of fine roots and rhizo-microorganisms (root-derived) and freeliving heterotrophic microorganisms (soil-derived) to the total soil CO₂ efflux were calculated as follows:

```
386 Equation 1: Soil-derived (%) = (root-exclusion core CO<sub>2</sub> efflux / root-ingrowth core CO<sub>2</sub> efflux) x
387 100
```

388 Equation 2: Root-derived (%) = 100 - Soil-derived (%)

389

Total soil CO_2 efflux measured for soil collars was multiplied by the results from equations 1–2 to estimate the absolute contributions of root-derived (roots, rhizo-microbial and mycorrhizal) and soil392 derived (free-living microbial through the decomposition of litter and soil organic matter) 393 components³⁵. The partition cores were buried to 0-25 cm depth, where 95% fine roots occur³⁶ – 394 therefore the soil component consists of soil-derived CO₂ from the entire soil profile (with very 395 marginal contribution from fine roots at > 20 cm depth).

396

397 Soil properties. Soil was sampled before the experimental treatments began (to 100 cm depth; 398 Extended Data Table 1) and then every three months following the beginning of the experiment (0-10 399 cm; average responses in Extended Data Fig. 4) within the plots at a point where the surface soil is 400 evenly heated (at approximately 30 cm distance from the heating structure), and analysed for soil 401 properties: total elements, available nutrients, microbial CNP and enzyme activities using standard 402 procedures (see SI methods). We calculated microbial carbon-use-efficiency (CUE) using microbial 403 CNP and enzyme activity data using a stoichiometric method³⁷. Here we describe the responses 404 following two years of warming, by using the average change in soil properties over two years (average 405 of eight temporal measurements per plot, with n = 5 per plot). Full details on these analytical methods 406 are provided in SI.

407

408 Statistical analyses. Treatment (warming) effects on time-averaged total and partitioned CO₂ 409 emissions and other soil properties (nutrients, microbial properties), were tested using ANOVA. 410 Treatment effects on soil CO₂ emissions were further tested using mixed effects models with CO₂ 411 emission as the response variable and warming-treatment, soil moisture, season, warming \times soil moisture and season \times soil moisture as fixed effects and plot number as random effect³⁸. We tested 412 413 both with and without a repeated measures effect because within-plot soil CO₂ efflux measurements 414 were partially spatially independent (within-plot locations of soil collars were changed every three 415 months, see soil gas exchange measurements above). Treatment effects on partitioned root- and soil-416 derived CO₂ efflux components were tested using repeated measures mixed models with time and plot

417	number as random effects. Treatment effects on soil-surface moisture (0-20 cm depth) were tested		
418	using mixed effects models with soil moisture as the response variable and warming-treatment, season,		
419	warming \times season as fixed effects and plot number as random effect. For whole-profile soil moisture		
420	we included time as an additional random effect (soil-surface moisture and temperature measurements		
421	were fully independent, whereas whole-profile measurements were repeated measures of fixed points).		
422	Further details on these approaches are provided in SI. All statistical analyses were performed in R		
423	(version 3.5.2).		
424			
425 426	31	Leigh, E. G. J. <i>Tropical Forest Ecology: A View from Barro Colorado Island</i> . (Oxford University Press, 1999)	
427 428	32	Woodring, W. P. Geology of Barro Colorado Island. <i>Smithsonian Miscellaneous Collections</i> 135 1 – 39 (1958)	
429 430	33	Sanchez, P. A. & Logan, T. J. Myths and Science About the Chemistry and Fertility of Soils in the Tropics SSA Spec Publ 29 35-46 (1992)	
431 432	34	Hanson, P. J. <i>et al.</i> A method for experimental heating of intact soil profiles for application to climate change experiments. <i>Global Change Biol</i> 17 , 1083-1096,	
433		doi:10.1111/j.1365-2486.2010.02221.x (2011).	
434	35	Nottingham, A. T., Turner, B. L., Winter, K., van der Heijden, M. G. A. & Tanner, E. V. J.	
435		Arbuscular mycorrhizal mycelial respiration in a moist tropical forest. <i>New Phytol</i> 186 ,	
436	26	957-967, doi:10.1111/J.1469-8137.2010.03226.X (2010).	
437	30	montane rain-forest in Panama <i>Plant Soil</i> 142 , 187-201 (1992)	
439	37	Sinsabaugh, R. L. et al. Stoichiometry of microbial carbon use efficiency in soils.	
440	-	<i>Ecological Monographs</i> 86 , 172-189, doi:10.1890/15-2110.1 (2016).	
<i>1.1</i> .1	30	Zuur A E Jono E N & Smith C M Introduction to mixed modelling Stat Biol Health	

441 38 Zuur, A. F., Ieno, E. N. & Smith, G. M. Introduction to mixed modelling. *Stat Biol Health*,
442 125-142 (2007).

443