



Geographic variation in a South American clade of mormoopid bats, *Pteronotus (Phyllodia)*, with description of a new species

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The subgenus *Phyllodia* (genus *Pteronotus*) comprises 9 species ranging from the western coast of Mexico to central Brazil, including Greater and Lesser Antilles. Two of them, *Pteronotus rubiginosus* and *Pteronotus* sp. 1, form an endemic South American clade within *Phyllodia* and are reported in sympatry for several localities in Guyana, Suriname, French Guiana, and northern Brazil. We herein performed a comprehensive investigation to fully characterize the cranial variation and genetic intraspecific structuring within this clade. We also integrated genetic, morphological, and acoustic evidence to formally describe the species previously reported as *Pteronotus* sp. 1. Specimens of *P. rubiginosus* occurring in sympatry with the new species have a more distinctive cranial phenotype than those from allopatric areas, suggesting character displacement as a potential force promoting divergence by decreasing resource competition or reproductive interactions between them. Although the 2 species are sympatric in several localities, the divergence in their echolocation calls also may be promoting resource partitioning at the microhabitat level, with *P. rubiginosus* foraging in less cluttered areas and the new species restricted to more cluttered areas.

<i>Phyllodia</i>	<i>Pteronotus</i>		
		<i>Pteronotus rubiginosus</i> e	
<i>Pteronotus</i> sp.		<i>Phyllodia</i>	
		<i>Pteronotus</i> sp.	<i>P. rubiginosus</i>
			<i>P. rubiginosus</i>

Key words: Amazon, character displacement, echolocation calls, integrative taxonomy, morphometrics, new species

The genus *Pteronotus* inhabits the Neotropical region, from western Mexico to central and northeastern Brazil and the Caribbean (Patton and Gardner 2008; Rocha et al. 2011; Pavan and Marroig 2017). Species of *Pteronotus* are small- to medium-sized bats, ranging from 5 to 25 g (Mancina et al. 2012; Emrich et al. 2014; al. 2017). They

feed primarily on insects, including a high proportion of beetles (Coleoptera) in their diet, but also on other insects in the orders Lepidoptera, Diptera, Orthoptera, Hymenoptera, Hemiptera, and Odonata (Mancina 2005; Rolfe and Kurta 2012). These bats occupy several distinct habitats along their geographic range, from open areas and dry deciduous forests to evergreen

humid forests, from sea level up to 3,000 m (Herd 1983; Patton and Gardner 2008). Some of their external morphological characters include large and plate-like lower lips, short bristle-like hairs surrounding the mouth, and funnel-shaped ears with distal pinna lanceolate; the fur is short, fine, and dense, with hairs highly varying in color from reddish or orange to dark brown, depending on the individual's molt progress (Smith 1972; Simmons and Conway 2001).

The genus diversity traditionally included 6 extant species (Smith 1972; Simmons and Conway 2001), assigned to 3 subgenera: *Pteronotus*, including *P. davyi* Gray, 1838 and *P. gymnonotus* (Wagner, 1843); *Chilonycteris*, including *P. macleayi* (Gray, 1839), *P. personatus* (Wagner, 1843), and *P. quadridens* (Gundlach, 1840); and *Phyllodia*, with the sole species *P. parnellii* (Gray, 1843). Several investigations in the last years, however, suggested cryptic diversity in the genus (Lewis-Oritt et al. 2001; ; Borisenko et al. 2008; Molinari 2008), particularly in the *P. parnellii* species complex (subgenus *Phyllodia*), which is represented by multiple evolutionary lineages (Clare et al. 2011, 2013; Thoisy et al. 2014). In a recent study, Pavan and Marroig (2016) employed molecular and morphometric evidence to address a new phylogenetic hypothesis for the genus (Supplementary Data SD1). According to this study, *Pteronotus* comprises 16 extant species. Many taxa previously presumed to be species widely distributed across the continent actually represent species complexes replaced parapatrically or allopatrically by each other (see table S11 of Pavan and Marroig 2016; fig. S1 of Pavan and Marroig 2017). In this context, the subgenus *Phyllodia*, traditionally including only *Pteronotus parnellii* as its single living species (with a variable number of subspecies, according to different authors—see Smith 1972 and Patton and Gardner 2008), is, therefore, composed of 9 distinct species; 8 of them have valid and available names, namely *P. parnellii* (sensu stricto), *P. pusillus* (G. M. Allen, 1917), *P. portoricensis* (Miller, 1902), *P. mexicanus* (Miller, 1902), *P. mesoamericanus* (Smith, 1972), *P. fuscus* (J. A. Allen, 1911), *P. paraguayensis* (Linares and Ojasti, 1974), and *P. rubiginosus* (Wagner, 1843). One of the lineages in this complex of cryptic species does not have an available name and was referred as *Pteronotus* sp. 1 sensu Pavan and Marroig (2016), and *Pteronotus* sp. 3 sensu Clare et al. (2013).

South America is, therefore, inhabited by 4 species of the subgenus *Phyllodia*: *P. paraguayensis*, *P. fuscus*, *P. rubiginosus*, and *Pteronotus* sp. 1 (see Pavan and Marroig 2016). *P. paraguayensis* on the western coast of Venezuela; this taxon was recognized as a valid species based on morphometric data (Molinari 2008) and its phylogenetic position is still unknown. *P. fuscus* is known to occur in some islands in the Lesser Antilles, Colombia, Venezuela, and 3 localities in the highlands of northwestern Guyana; this species is more closely related to the *Phyllodia* species distributed in Central America (*P. mesoamericanus* and *P. mexicanus*) than to the South American species (Supplementary Data SD1; clade 4-B of Pavan and Marroig 2016). The other 2 species from South America are

P. rubiginosus, widely distributed in the Amazon and the Brazilian Cerrado biomes, and *Pteronotus* sp. 1, which has been found in Guyana, Suriname, French Guiana (hereafter referred to as the Guianas), and the Brazilian Amazon. These 2 species comprise an exclusively South American clade within *Phyllodia* (Supplementary Data SD1; clade 4-C of Pavan and Marroig 2016) and are reported in sympatry for several localities, being molecularly and acoustically discernible (Clare et al. 2013; Thoisy et al. 2014; Pavan and Marroig 2016; et al. 2017). Previous studies also suggested that these species might exhibit differences in cranial and dental measurements (Thoisy et al. 2014; al. 2017), although the morphometric dataset analyzed in the mentioned studies was small, both in sample size and geographic coverage.

Therefore, a more comprehensive investigation to characterize the phenotypic variation of the 2 species of the subgenus *Phyllodia* that occur in sympatry in Guianas and Brazil is still pending, as there is consistent molecular and acoustic evidence in the literature suggesting they represent independent lineages. This is an important topic to be addressed because the lack of knowledge on the morphological variation, i.e., how to differentiate these 2 species, is precluding researchers from acknowledging the real diversity of the genus *Pteronotus* in this geographic area and correctly identifying museum vouchers. Herein, we use highly complementary molecular and morphological datasets through most of the species range to investigate more thoroughly the populational structuring and the phenotypic divergence patterns between the 2 species of the South American clade of the subgenus *Phyllodia*, *Pteronotus* sp. 1, and *P. rubiginosus*. As a result, we formally describe the unnamed species of *Pteronotus* (*Pteronotus* sp. 1 sensu Pavan and Marroig 2016; *Pteronotus* sp. 3 sensu Clare et al. 2013) and redefine the taxonomic limits of *P. rubiginosus*, integrating the mitochondrial DNA (mtDNA), morphological, and bioacoustic markers here presented as well as those previously reported in the literature (Thoisy et al. 2014; Pavan and Marroig 2016; al. 2017).

MATERIALS AND METHODS

Sampling.—Our molecular dataset corresponded to a fragment of 651 bp of the cytochrome oxidase I (*COI*) mitochondrial gene. It included sequences of 157 individuals belonging to the lineages of *P. rubiginosus* and *Pteronotus* sp. 1 previously published by Clare et al. (2013), Thoisy et al. (2014), Pavan and Marroig (2016), and al. (2017) and available at GenBank and the Barcode of Life Data System (BOLD) (Appendix I). In addition, 11 specimens from 2 localities in Amazonas, Brazil, and 1 locality from Potaro-Siparuni, Guyana, were sequenced and added to this dataset, totaling 168 individuals. These new sequences are available at GenBank under the accession numbers MH017827–MH017837. The molecular dataset included 102 individuals also included in the morphological investigation (Appendix I).

For the morphological investigation, we examined 184 vouchers of common mustached bats sampled across the geographic range of this clade in South America, including 36

localities in the Guianas and Brazilian Amazon, where the 2 lineages were reported to occur in sympatry, and 12 localities in the Cerrado and southern Amazon of Brazil, where only populations of *P. rubiginosus* have been recorded (Fig. 1; see also Pavan and Marroig 2016). We also examined 15 specimens of *P. fuscus*, including the holotype and 3 individuals previously sequenced, to provide some morphological comparisons with *Pteronotus* sp. 1 and *P. rubiginosus* (see below, in the “Systematics” section). The specimens included in the present study are housed in the following institutions: Museu de

Natural History (AMNH), Royal Ontario Museum (ROM), and

list of examined specimens is described in Appendix II.

To gather additional evidence for delimiting species more properly, we incorporated the bioacoustic information provided by Thoisy et al. (2014) and al. (2017) for 40 specimens that we analyzed in our molecular and morphological investigation. Therefore, although we have not collected new acoustic data for the present study, some important findings of these 2 previous studies are reported in the results. Appendix I presents the correspondence between the molecular,

morphometric, and bioacoustics datasets and the GenBank accession numbers of all sequences included in the molecular analysis.

Molecular data.—Total genomic DNA was extracted from ethanol-preserved tissues using the Qiagen DNeasy Blood and Tissue Kit (Qiagen, Inc., Germantown, Maryland). The amplification and sequencing of the *COI* region was performed adopting the same primers and conditions described previously (Clare et al. 2007; Borisenko et al. 2008). Sequences were assembled and checked for quality using Geneious v.9.1 (Biomatters, Ltd., Auckland, New Zealand) and aligned with the sequences already available from GenBank using MEGA7 (Tamura et al. 2013). Molecular diversity indices for both species and estimates of genetic differentiation between them were calculated by MEGA7 and DnaSP v5 (Librado and Rozas 2009). We estimated the relationships among observed haplotypes using the median-joining network algorithm (Bandelt et al. 1999) by Network 5.0 (fluxus-engineering.com). We used haplotype network results to identify fixed polymorphic sites between the species and to show intraspecific population structuring.

Morphological data.—Morphological data were collected from adults only, i.e., those having fused epiphyses of phalanges and metacarpals (Smith 1972). External and osteological characters were analyzed inter- and intraspecifically. Qualitatively, we studied facial structures, fur color, differences in foramina sizes, and variation in teeth morphology. We follow Simmons and Conway (2001) in assigning homology

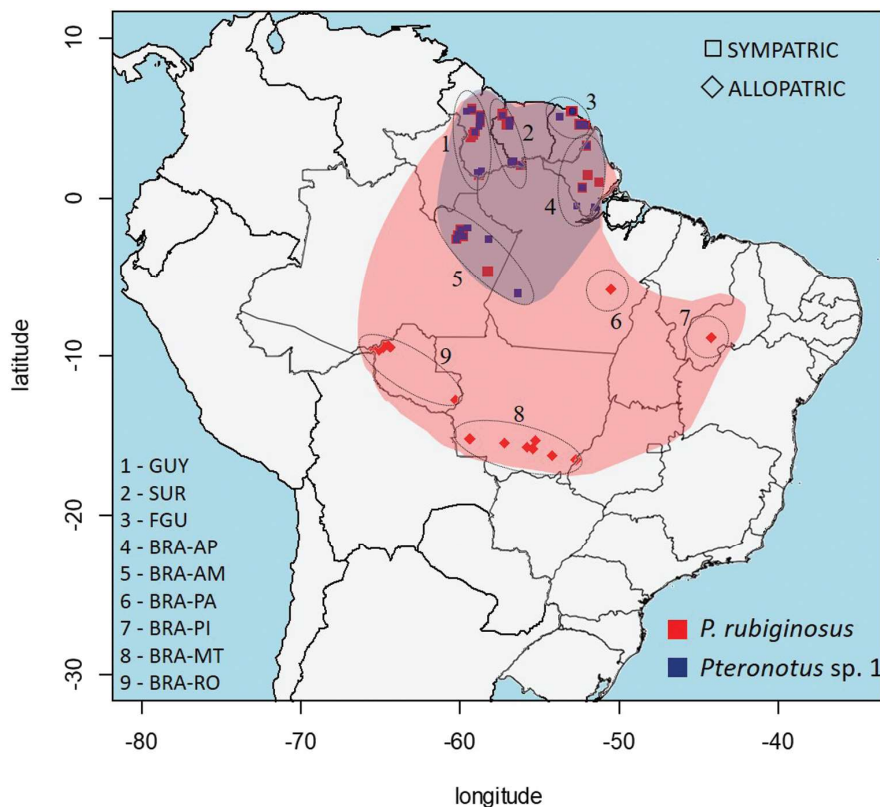


Fig. 1.—Sampling localities of the common mustached bat (*Pteronotus* cf. *rubiginosus*) included in the present study.

for the premolars and [al. \(2014\)](#) in the terminology of the dental structures evaluated in this study. Quantitatively, craniodental and mandibular measurements were recorded to the nearest 0.01 mm using digital calipers. Standard external measurements (forearm length [FL] and tibia length [TL]) are data collected in the field and were not used in the morphometric analysis. Nine linear measurements were taken from skulls and mandibles, 7 of them follow [Velazco and Patterson \(2014\)](#) and [Nogueira et al. \(2012\)](#), whereas the other 2 express the particular variation observed between these species of *Pteronotus* ([Supplementary Data SD2](#)). The measurements are:

Palatal width.—Distance across the palate, taken between the lingual margins of the alveoli of P3 at their contact with the canines.

Palatal length.—Measured from the anteriormost edge of palate bone, between the inner incisors, to its posteriormost margin, at the mesopterygoid fossa.

Greatest length of skull.—From the posteriormost point on the supraoccipital bone to the anteriormost point on the premaxilla (excludes incisors).

Interorbital breadth.—The least breadth across the postorbital constriction.

Braincase breadth.—The greatest breadth of the globular part of the braincase, excluding mastoid and paroccipital processes.

Rostral width at M2 (M2–M2).—The greatest width of the rostrum measured across the labial margins of the alveoli of M2.

Maxillary tooththrow length.—The distance from the anteriormost surface of the upper canine to the posteriormost surface of the crown of M3.

Mandibular length.—Greatest length of the dentary, from its anteriormost point (excluding the incisors) to its posteriormost point at the angular process.

Mandibular tooththrow length.—Distance from the anteriormost surface of the lower canine to the posteriormost surface of m3.

To better understand the morphological variation within this clade, we initially included in these comparisons only those specimens with molecular or bioacoustic information available to support their corresponding species assignment. Localities from Guyana (GUY), Suriname (SUR), French Guiana (FGU), and from the Brazilian states of Amazonas (BRA-AM), and

species ([Fig. 1](#)) because these areas have reports of sympatric occurrence ([Clare et al. 2013](#); [Thoisy et al. 2014](#); [Pavan and Marroig 2016](#); [Lopez-Baucells et al. 2017](#)). On the other hand, localities from the Brazilian states of Mato Grosso (BRA-MT),

were considered of exclusive occurrence of *P. rubiginosus* ([Pavan and Marroig 2016](#); [Fig. 1](#)).

Statistical analysis.—We firstly performed a principal component analysis (PCA) to evaluate which morphological variables most contributed to the variation of each axis and, consequently, which were more informative to discriminate populations and species. This information allowed us to perform a comprehensive investigation in the complete morphological dataset (i.e., those specimens without molecular or

acoustic information). To better understand how the cranial variables responded to geographic variation and differences between sexes as well as to the interaction between these 2 factors, we performed multivariate analyses of variance (MANOVA) in the complete distribution of each species. We additionally implemented MANOVA in different subsets of each species dataset to further investigate the sexual variation observed. For *P. rubiginosus*, we tested differences between sexes within one of the allopatric and sympatric populations with the largest samplings (BRA-MT and FGU, respectively) as well as for the total sampling of allopatric and sympatric regions. For *Pteronotus* sp. 1, tests were made for the largest population (FGU) and the Guianas as a whole. Divergence in variable means between sympatric and allopatric populations of *P. rubiginosus* was also tested using univariate analysis of variance (ANOVA). Discriminant function analysis (DFA) was then used to compare species across their ranges and to explore potential factors affecting the group morphometric variation, such as geographic overlap and sexual dimorphism. The PCA and MANOVA were conducted in the R software environment ([R Development Core Team 2013](#); functions *prcomp* and *lm*, respectively). The library *ggbiplot* was used for the PCA graph. The ANOVA and DFA were run with SYSTAT 11.

RESULTS

Molecular variation and population structuring.—The fragment of *COI* gene analyzed showed 80 variable positions, 29 singletons and 51 potentially parsimony informative sites, with 44 distinct haplotypes within the data. In accordance with phylogenetic data presented by previous studies ([Clare et al. 2013](#); [Thoisy et al. 2014](#); [Pavan and Marroig 2016](#)), the haplotype network inferred for the *COI* dataset presents 2 haplotype groups (clusters), corresponding to *P. rubiginosus* ($n = 114$) and *Pteronotus* sp. 1 ($n = 54$; [Fig. 2](#)). The K2P nucleotide divergence between them is 5.3%. As mentioned previously and highlighted in [Figure 2](#), these 2 clusters are not entirely distinguishable by geography: there are individuals inhabiting the same or nearby localities in Guyana, Suriname, French Guiana, and northern Brazil harboring highly divergent haplotypes, although Brazilian samples from MT, RO, PA, and PI are only recovered clustered in one of the haplogroups. A minimum of 20 fixed nucleotide mutations was found setting apart these 2 clusters of haplotypes. In addition, 41 sites are polymorphic across sequences of *P. rubiginosus* but are monomorphic within *Pteronotus* sp. 1 while 22 sites show the opposite variation between the 2 species.

It is also possible to notice intraspecific structuring for both species (1.9% for *P. rubiginosus* and 0.9% for *Pteronotus* sp. 1). Most of the *P. rubiginosus* specimens ($n = 72$; 63%) harbor the same haplotype (HAP1) while 1–4 individuals compose the other 31 haplotypes found for this species. For *Pteronotus* sp. 1, we could find 2 haplotypes at higher frequencies (HAP2 = 27 [54%] and HAP3 = 10 [18%]) and 10 haplotypes at lower frequencies within the sampling. Molecular diversity indices and neutrality tests estimated for both species

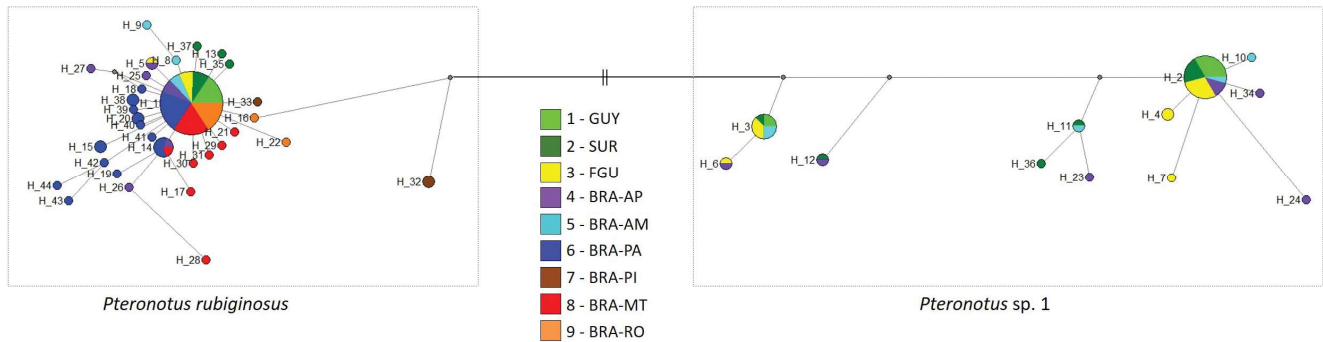


Fig. 2.—Haplotype network showing the geographic variation and intraspecific diversity in the *COI* fragment for the 2 South American sympatric species of common mustached bats (subgenus *Phyllodia*). Each haplotype found in our molecular dataset is labeled with a number. The haplotypes belonging to each cluster (representing the 2 species) differ in at least 20 mutational steps among them. Geographic samples within both species are identified by distinct colors according to the labels.

Table 1.—Molecular diversity indices and neutrality tests estimated for the *COI* region in both species. We excluded from the estimates 3 specimens with incomplete sequences and 2 specimens with uncertain assignment to species. Diversity indices presented below (Nei 1987): h = number of haplotypes; S = number of segregating sites; Hd = haplotype (gene) diversity; Pi = nucleotide diversity. Neutrality tests (significant values highlighted in bold): Tajima's D (Tajima 1989) and Fu's F_s (Fu 1997).

Species	n	h	Diversity indexes			Neutrality tests	
			S	Hd	Pi	D	F_s
<i>Pteronotus rubiginosus</i>	112	32	39	0.587	0.0018	-2.623	-43.280
<i>Pteronotus</i> sp. 1	51	12	22	0.686	0.0067	-0.355	-0.023

are presented in Table 1. We found a higher genetic diversity for the *COI* region in *Pteronotus* sp. 1, whereas *P. rubiginosus* seems to have gone through demographic expansion recently.

Morphometrics.—The morphometric dataset included 8 cranial measurements from 146 specimens. Table 2 describes sample sizes and mean values for selected external and cranial variables. Mean values of all cranial measurements differed among the 3 groups defined for morphometric analysis: *Pteronotus* sp. 1, *P. rubiginosus* – sympatric, and *P. rubiginosus* – allopatric; the range of values for most of these measurements, however, overlap. The result of the PCA allowed a general overview of the specimens belonging to the 2 distinct lineages in the morphospace (Fig. 3). The 1st principal component (PC1) explains ca. 86% of the total variation observed in the dataset and corresponds to size, being positively related to all 8 cranial variables; the 2nd principal component (PC2) is basically a contrast between the 2 width measurements of the cranial vault (interorbital breadth [IB] and braincase breadth [BB]) and the remaining variables, representing 4.7% of the total variation.

The PCA plot shows specimens from the same collecting localities but assigned to the distinct species in different areas of the morphospace, highlighting the dissimilarity between them. There is a general trend of size increase for both species in the northern part of their ranges (Guyana and Suriname). Specimens of *P. rubiginosus* from the allopatric (southernmost) region of the distribution (Fig. 3, populations numbered 6–9)

exhibit the smallest sizes for the species and are recovered in the overlapping area of the morphometric space.

The MANOVA results suggest the presence of geographic and sexual variation for both species, but no interaction between these 2 factors (Table 3). As a general pattern, we could not observe significant differences between sexes within any of the tested populations, but we did find differences at wider geographic scales. Thus, DFA was performed for the complete dataset but also for females and males separately. Geographic variation within *P. rubiginosus* was also explored through ANOVA, which compared the populations in sympatry with *Pteronotus* sp. 1 with those in allopatry, revealing that the 8 variables are statistically different, with individuals in sympatric populations being larger on average (Supplementary Data SD3).

The DFA showed very high classification rates of specimens into the categories (species) regardless which dataset was analyzed: only females, only males, or females and males combined (Supplementary Data SD4). Therefore, we herein explore the results of the DFA performed for the complete data. More than 90% of specimens were assigned to the correct category for both species, and values described for original and jack-knifed matrices are very close, highlighting the robustness of the discriminant function (DF) in differentiating the species (Klekca 1980; Pavan and Marroig 2016). Every misclassified specimen of *P. rubiginosus*, i.e., specimens belonging to *P. rubiginosus* but assigned to *Pteronotus* sp. 1 by their canonical scores, are from localities not overlapping with the range of the new species (allopatric populations of PI, MT, and RO in Brazil). When the range of DF values for each species is sorted by the different populations and modes of distribution (sympatric or allopatric), for localities of sympatry, species exhibit clear distinctive sizes, with *P. rubiginosus* exhibiting most of the size variation throughout its distribution (Fig. 4). Finally, males exhibited slightly larger values in the variable means than did females in both species, but there is no evident tendency in the DF values when both sexes are compared (data not shown).

Morphology.—We performed a careful and detailed comparative study on the skull morphology of specimens belonging

Table 2.—Measurements (mm) of the 2 external and 8 cranial variables collected from specimens of *Pteronotus* cf. *rubiginosus* according to the classification and mode of distribution established for morphometric analyses. For external measurements, mean values and sample sizes are presented. For cranial variables, samples sizes by sex (F = females; M = males; U = undetermined) and mean values (minimum–maximum range). BB = braincase breadth; GLS = greatest length of skull; IB = interorbital breadth; MdL = mandibular length; MdTL = mandibular tooththrow length; MxTL = maxillary tooththrow length; PL = palatal length; PW = palatal width.

Measurement	<i>Pteronotus rubiginosus</i>		<i>Pteronotus</i> sp. 1
	Allopatric	Sympatric	
Forearm length, mean (<i>n</i>)	62.8 (24)	64.9 (18)	61.6 (28)
Tibial length, mean (<i>n</i>)	25.5 (24)	26.2 (08)	24.4 (19)
<i>n</i> (cranial data)	38	49	60
	18 F, 17 M, 03 U	24 F, 24 M, 01 U	35 F, 23 M, 02 U
PW	4.28 (3.87–4.58)	4.49 (4.12–4.77)	4.20 (3.83–4.57)
PL	10.16 (9.76–10.51)	10.59 (10.06–11.20)	9.90 (9.53–10.42)
GLS	22.24 (21.4–23.11)	23.00 (22.24–23.64)	21.79 (21.11–22.59)
IB	4.39 (3.92–4.60)	4.51 (4.19–4.82)	4.60 (4.04–5.15)
BB	10.82 (10.32–11.40)	11.07 (10.13–11.45)	10.78 (10.08–11.30)
MxTL	9.67 (9.34–10)	10.09 (9.72–10.43)	9.43 (9.10–9.71)
MdL	17.04 (16.34–17.76)	17.73 (17.00–18.40)	16.73 (15.95–17.37)
MdTL	10.33 (9.96–10.70)	10.73 (10.35–11.14)	10.05 (9.55–10.40)

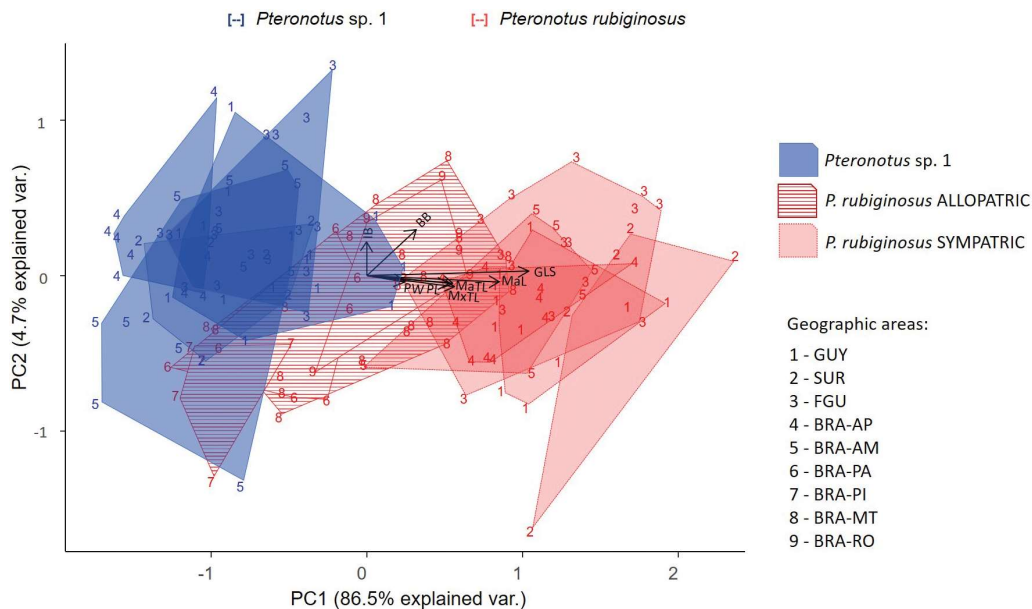


Fig. 3.—Principal component analysis displaying the cranial morphometric variation in the 2 sympatric species of the South American clade of *Phyllodia* (Clade 4C, [Supplementary Data SD1](#)). The correlation of the cranial variables with the 2 first principal components (PCs) is displayed in the center of the plot. The PC1 and PC2 correspond to 86.5% and 4.7% of the total variation, respectively. Higher values in PC1 represent larger cranial sizes while values in PC2 show variation in cranial breadth contrasted to the overall size. Numbers indicate the geographic location of individuals, whereas polygons delimit the area of each of these groups in the morphometric space.

Table 3.—Results of multivariate analysis of variance (MANOVA) performed in the complete dataset of each species (all) as well as in more restricted geographic regions within them. The *P*-values are provided in the rows, with significant values highlighted in bold. BRA-MT = Brazilian state of Mato Grosso; FGU = French Guiana.

Factor tested	<i>Pteronotus rubiginosus</i>					<i>Pteronotus</i> sp. 1	
	All	Allopatric	Sympatric	BRA-MT	FGU	All	Guianas
Sex	2.96×10^{-4}	0.01	0.05	0.40	0.39	0.04	0.02
Geography	3.71×10^{-9}	0.01	0.18			3.65×10^{-4}	0.39

to the 2 species. At first, evaluation was conducted among specimens sampled in the same or nearby localities to avoid a possible geographic bias. Further, groups of specimens were compared along the species ranges, allowing the definition of

some potential informative characters to be compared between the species. We contrasted the information we had about individual identification based on previous molecular results (*COI* and cytochrome b [*Cytb*] mitochondrial genes) with

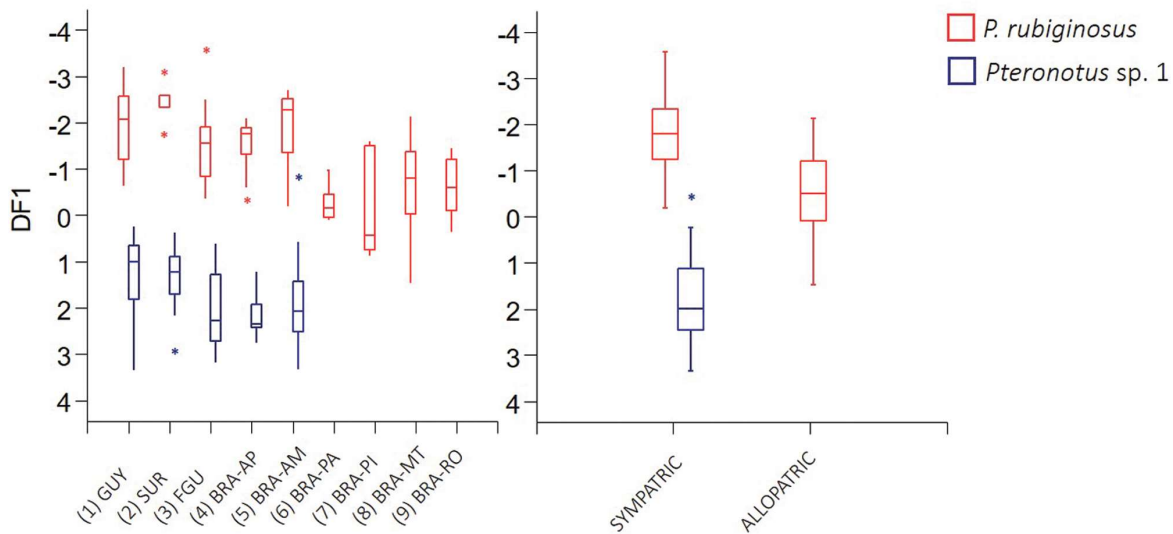


Fig. 4.—Range of the discriminant function (DF) values for the 2 species according to their geographic position or populations (left) and their mode of distribution (right). Notice the difference between the mean DF values in the populations of *Pteronotus rubiginosus* from the sympatric area of occurrence with *Pteronotus* sp. 1 (samples 1–5) and the populations from the allopatric area (samples 6–9).

the morphological data we obtained studying the specimens. We found a high correspondence between the mitochondrial clades and distinct patterns of skull morphology. The clade of *P. rubiginosus*, in general, is composed of specimens with larger skulls, with longer rostrum and palate, and proportionally smaller IB when compared to the skull of *Pteronotus* sp. 1. In addition, skulls of *P. rubiginosus* are characterized by a large pair of foramina in the pterygoid canal, which are noticeably smaller in the new species. Some morphometric traits are also useful for differentiating between the 2 species, especially the length of the maxillary tooththrow (see “Systematics” section below). This set of morphological characters varies geographically, being more variable in *P. rubiginosus* than in *Pteronotus* sp. 1. For example, both species tend to have larger skulls in the northern part of their distributions (Guyana and Suriname). Furthermore, individuals of *P. rubiginosus* vary not only in skull size, but in the robustness of the rostrum across the geographic range. Populations from the southern part of the distribution (Fig. 1: allopatric area) exhibit smaller skulls with more delicate rostra. Some specimens of *P. rubiginosus* from the Guiana Shield, on the other hand, have rostra with relatively wider IB, resembling the profile of *Pteronotus* sp. 1; these specimens are noticeably large, however, and can be easily distinguished from *Pteronotus* sp. 1 occurring across this range based on skull length (Supplementary Data SD5).

The only exceptions to the pattern mentioned above were 2 specimens (MPEG 41678 and AMNH 269115) whose mitochondrial haplotypes assign them to one of the lineages while the morphological characters strongly suggest that they may belong to the other. Because a 3rd source of evidence (acoustics) was not available for these individuals, we opted for not assigning them to any of the species when investigations requiring a priori classification of the data were performed; further discussion of these specimens will be done in the following sections. We also evaluated the variation of facial structures, fur length, and color, but

we found them to be uninformative for species differentiation.

Acoustic data.—Previous studies demonstrate that the 2 lineages included in this South American clade of *Phyllodia* represent 2 distinct phonic groups, which are easily discernible with no intermediate frequency values between them (Thoisy et al. 2014; al. 2017). Individuals of *P. rubiginosus* emit constant frequency (CF) calls around 53 kHz in French Guiana and 55 kHz in the central Amazon, whereas *Pteronotus* sp. 1 uses CF calls between 59 and 60 kHz in the same localities. Therefore, acoustic evidence suggests that there is a consistent difference of 5–7 kHz in the echolocation calls emitted by individuals belonging to these 2 species, even at different sites, with no overlap in the peak frequency range between them (see fig. 1 of al. 2017). These data also point to some level of geographical variation within each species although such variation does not compromise the ability of identifying acoustically between these 2 species in the field (al. 2017). The range of values of the frequency of maximum energy (FME) in *P. rubiginosus* seems larger than that observed for *Pteronotus* sp. 1.

SYSTEMATICS

Family Mormoopidae Saussure, 1860

Genus *Pteronotus* Gray, 1838

Pteronotus alitonus sp. nov.

Chilonycteris rubiginosa [*rubiginosa*]: Rehn, 1904:200; part.

Chilonycteris rubiginosa rubiginosa: Husson, 1962:74; part.

Pteronotus [*Phyllodia*] *parnellii rubiginosus*: Smith, 1972:75; part.

Pteronotus parnellii: Honacki, Kinman and Koepl, 1982:150; part

Pteronotus sp. 3: Clare et al., 2013:14.

Pteronotus sp. 1: Pavan and Marrojo, 2016:190.

Holotype.—An adult male (INPA 6942; Fig. 5), preserved in alcohol with the skull removed and cleaned, deposited at the collection of the INPA, Amazonas, Brazil. It was collected on 6 September 2014 by Ricardo Rocha (field number PP02) at the Biological Dynamics of Forest Fragments Project (BDFFP) area, 80 km north of Manaus, Brazil (2°20'S, 60°6'W, elevation of 30–125 m). Body, skull, and mandible are in good condition. Tissue is preserved in ethanol and frozen at INPA under the same ID. Sequence of the mitochondrial gene *COI* is available under the GenBank accession number MH017835.

Paratypes.—MZUSP 35505, 35523; IEPA 417, 1847; INPA 6947; ROM 98128, 106659, 117545; MHNG 1978.077, 1978.082; AMNH 267851.

Other material.—The complete list of 82 specimens of *P. alitonus* analyzed in this study is described in Appendix II.

Distribution.—The new species of *Pteronotus* is known from several localities in the pristine forests of Guyana, Suriname, French Guiana, and the Brazilian Amazon (Fig. 1; Appendix II).

Etymology.—The specific epithet, *alitonus*, is composed of the Latin words *alius* (= different, changed) and *tonus* (sound, tone), in reference to the distinct echolocation call emitted by this species in comparison to *P. rubiginosus*.

Nomenclatural statement.—A Life Science Identifier (LSID) number was obtained for the new species *Pteronotus alitonus*: urn:lsid:zoobank.org:act:4B22D88F-77BA-4021-B031-B54B946DC52D.

Historical background.—In the synonymy presented above (only with 1st use of the names), we tentatively listed authors that implicitly included in their concepts of *Chilonycteris rubiginosa* or *P. parnellii*, populations from northern Brazil and the Guianas, which could be assigned to the new species here described. The 2 latter synonyms represent not binomial (and more formal) entries, but refer to specimens whose association to this new species is clear and explicit. Several studies have been published in recent years evidencing the existence of 2 sympatric lineages of common mustached bats in the Amazonian region. Each of these studies has focused on specific questions about the group, but also provided additional

sources of information to guide the present study. Based on information from 3 loci, Clare et al. (2013) described the existence of 4 distinct lineages in the continental range of the species formerly known as *P. parnellii*: 1 in Central America and 3 in South America. Two of these South American lineages, called *Pteronotus* sp. 3 and *Pteronotus* sp. 4, were found in sympatry in Guyana and Suriname. Clare et al. (2013) also described information from acoustics and morphometrics related to the geographic areas of occurrence of these lineages. Thoisy et al. (2014) showed that the 2 sympatric lineages of the *P. parnellii* complex found by Clare et al. (2013) in Guyana and Suriname were acoustically discernible and increased their ranges to French Guiana and northern Brazil. Thoisy et al. (2014) provided molecular and morphometric evidence indicating that one of these lineages (*Pteronotus* sp. 4) corresponds to *P. parnellii rubiginosus*, the taxon already known for that area. Later, Pavan and Marroig (2016) provided a new phylogenetic hypothesis and an updated taxonomic arrangement for the genus *Pteronotus*. Pavan and Marroig (2016) corroborated the existence of 8 distinct lineages within the *P. parnellii* complex (subgenus *Phyllodia*), 1 in Mexico, 1 in Central America, 3 in South America, and 3 in the Caribbean, linking them to the available names in the group (as subspecies and species names) and showing that one of the sympatric lineages from South America (*Pteronotus* sp. 1—sensu Pavan and Marroig 2016; *Pteronotus* sp. 3—sensu Clare et al. 2013) has no name available. Pavan and Marroig (2017) published a dated phylogeny for the genus *Pteronotus* based on the same molecular data published by Pavan and Marroig (2016). They discussed the historical processes related to the origin and diversification events within the group.

al. (2017) reported the existence of geographic variation in the echolocation calls of these 2 sympatric species of *Pteronotus* from South America. The study by al. (2017) is complementary to Thoisy et al. (2014) because it showed that, despite the existence of intraspecific variation, both species are acoustically discernible, i.e., the differences are consistent across geography. They also provided additional information on the distribution of the unnamed lineage of *Pteronotus* in Central Amazon. We



Fig. 5.—Dorsal, ventral, and lateral views of the skull and ventral and lateral views of the mandible of the holotype of *Pteronotus alitonus* sp. nov. (INPA 6942). Scale bar = 5 mm. INPA =

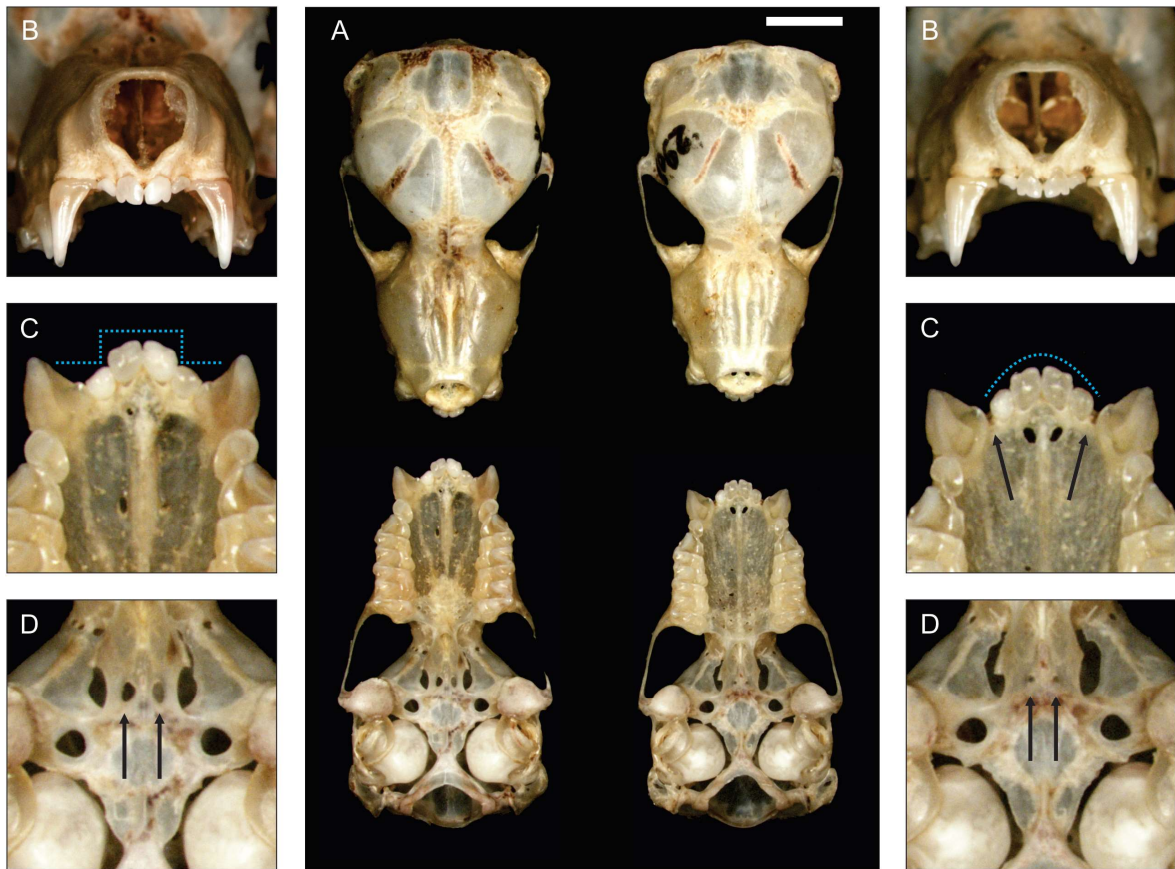


Fig. 7.—Comparison of the cranial morphology between *Pteronotus rubiginosus* (MHNG 1978.083) and *P. alitonus* sp. nov. (MHNG 1978.082) from Grotte Mathilde, French Guiana. A) Dorsal and ventral views of the skull of *P. rubiginosus* (left) and *P. alitonus* (right); B) frontal view of the skull, contrasting the sizes of the internal (I1) and external (I2) incisors in the 2 species; C) ventral view of 1st half of the palate, highlighting the form outlined by the incisors and the gap between I2 and canines in *P. alitonus*; D) ventral view of the basicranial region, with arrows indicating the foramina in the pterygoid canal with different sizes in the 2 species. Scale bar in top right of A = 5 mm. MHNG =

canal has a pair of foramina varying from almost indistinct perforations to small pits (less than 1/3 of the foramen ovale) in the new species; alternatively, *P. rubiginosus* specimens exhibit a large pair of foramina, as large as half of the diameter of the foramen ovale (Fig. 7D).

The dental formula is the same as for all other mormoopids, $i\ 2/2\ c\ 1/1\ p\ 2/3\ m\ 3/3 = 34$. The inner (or central) upper incisors (I1) are bilobed and usually have less than twice the height of the outer (or lateral) incisors (I2); the I1 are proportionally larger in *P. rubiginosus* than in *P. alitonus*, with more than twice the height of I2 (Fig. 7B). Ventrally, this character is also noticeable, with the margins of the incisors usually forming a continuous arc in *P. alitonus*, while the margins of the incisors are steeply uneven in *P. rubiginosus* (Fig. 7C), with the central ones forwardly projected. A small gap separating the outer upper incisors from the canines is sometimes present in *P. alitonus* (e.g., INPA 6942, MHNG 1978.082, AMNH 267851, ROM 106659) and is absent in all specimens of *P. rubiginosus*. The labial cingulum and the entire labial margin of the 1st upper premolar (P3 in homology—sensu Simmons and

Conway 2001) in *P. alitonus* exhibits a concave and rounded profile (C-shaped), with deeper notches on the molar tooththrow between the canine and the P3 and between the P4 and P3; by contrast, the labial cingulum of the P3 of *P. rubiginosus* exhibits a less concave, more open profile, with much less noticeable notches between P3 and the adjacent teeth. Moreover, the general shape of the P3 is usually different between species, being more rounded and narrow buccolabially in *P. alitonus* and more elongated and long buccolabially in *P. rubiginosus*. The inner lower incisors are trilobed and larger than the outer bilobed incisors in both species but for some individuals of *P. alitonus*, i1 and i2 crowns are not in contact. The 1st and 3rd lower premolars (p2 and p4) are large and have well-developed labial cingulids; in both species, the p3 is peg-like and compressed between the lingual edges of p2 and p4, although in *P. rubiginosus* it is usually larger than in *P. alitonus*.

Additional comparisons.—We examined the cranial morphology of specimens of the geographically contiguous species *P. fuscus* and compared them with specimens of *P. rubiginosus* and *P. alitonus* from Guyana to provide a few insights on

their differentiation. *P. fuscus* has its easternmost distribution in the highlands of northwestern Guyana (Clare et al. 2013), near the border with Venezuela, whereas *P. rubiginosus* and *P. alitonus* extend northward to central and southeastern Guyana (Fig. 1). So far, these 3 species have not been recorded at the same locality and, therefore, *P. fuscus* is considered to have a parapatric distribution relative to *P. rubiginosus* and *P. alitonus*. In general, specimens of *P. fuscus* have a skull shape more similar to *P. rubiginosus*, but size more similar to *P. alitonus* (Supplementary Data SD8). Comparatively, the skull of *P. fuscus* has a narrow and delicate rostrum; the nasal bones are parallel and flat. *P. fuscus* can be easily differentiated from *P. rubiginosus* based on the smaller size (greatest length of skull [GLS] = 21.4 mm \pm 0.36; MxTL = 9.33 mm \pm 0.2). When compared to *P. alitonus*, specimens of *P. fuscus* have noticeably narrower rostra, which are not slightly upturned in their anterior most part as for *P. alitonus*. The maxillary bones are less inflated in their suture with nasals; morphometrically, this feature is noticed by the greatest width across the molars (M2–M2), generally < 8.0 mm in *P. fuscus*, and > 8.18 mm in *P. alitonus* from Guyana.

Natural history.—Little is known about the biology or behavior of this new species, but our data suggest that it forages preferentially in highly cluttered forested areas as there are no reports on the species occurrence in more open areas such as savannas or karstic regions in the Amazon.

Remarks.—As a consequence of the description of *P. alitonus*, *P. rubiginosus* needs to be redefined. In the present study, we described several morphological features that allow the identification of *P. rubiginosus*, including quantitative and qualitative characters (see previous section; see also Table 2 and Fig. 7). *P. rubiginosus* is found in the Amazon and Cerrado biomes of South America. The occurrence of this species has been confirmed by molecular data for Guyana, Suriname, French Guiana, northern and central Brazil (and states of Pavan and Marroig 2016; A. C. Pavan, pers. obs.). There are also records of *Pteronotus* cf. *rubiginosus* in the Amazonian regions of Peru, Colombia, and Venezuela but the status of these populations need to be reviewed.

DISCUSSION

Our comparative study evaluated the existence of an unnamed evolutionary lineage of *Pteronotus* (*Pteronotus* sp. 1—sensu Pavan and Marroig 2016; *Pteronotus* sp. 3—sensu Clare et al. 2013) occurring in sympatry with *P. rubiginosus* in several localities in the Guianas and Brazilian Amazon (Clare et al. 2013; Thoisy et al. 2014; Pavan and Marroig 2016; Baucells et al. 2017). Using cranial, genetic, and acoustic data, we corroborate the hypothesis of a new species of *Pteronotus* and formally describe *P. alitonus*.

These 2 species, *P. rubiginosus* and *P. alitonus*, diverge in genetic and acoustic traits throughout their geographic ranges. Despite their external similarity, we herein provided skull diagnostic traits that will be useful for future identification of

material in collections. We also provided some insights on their evolution and ecology, which are essential for the adequate diagnosis of the new species.

Genetic divergence and population structuring.—The COI haplotype network reconstruction recovers both species, *P. rubiginosus* and *P. alitonus*, as cohesive clusters, but with intraspecific structuring. Molecular diversity indexes (Table 1) point to a higher genetic diversity in *P. alitonus* compared to *P. rubiginosus*. In *P. alitonus*, the 2 most frequent haplotypes (HAP2 and HAP3) differ in 9 mutated positions from each other, with 3 intermediary haplotypes (median vectors) inferred by the analysis (Fig. 2). No spatial correlation seems to exist, however, for the distribution of these haplotypes; both are widespread across the species geographic range. *P. rubiginosus* exhibits 1 central haplotype (HAP1), surrounded by several related haplotypes at low frequency and showing little difference among them. This pattern suggests these low-frequency haplotypes originated recently and agrees with a scenario of recent demographic expansion (Wakeley 2004; Ferreri et al. 2011). This premise is confirmed by the significant values of neutrality tests found for *P. rubiginosus* (Table 1). A more divergent haplotype occurs in *P. rubiginosus* (HAP32), represented

northeastern Brazil, but unfortunately these specimens were not available for the morphological analyses. Alternatively, we (Appendix II), and these specimens are unequivocally assigned to *P. rubiginosus*, although exhibiting the smallest average sizes of our *P. rubiginosus* sampling.

Morphological variation and character displacement.—We corroborate previous studies (Thoisy et al. 2014; Baucells et al. 2017), showing that specimens of *P. rubiginosus* have larger skulls than specimens of *P. alitonus* where these species occur in sympatry. In addition, we present some cranial characters useful to distinguish between these 2 lineages, the most consistent being the foramina in the pterygoid canal. This structure exhibits distinct states (Fig. 7D): in *P. rubiginosus*, 81 out of the 87 specimens (93%) have a relatively large pair of foramina, averaging from 1/3 to 1/2 of the diameter of the foramen ovale; whereas in *P. alitonus*, approximately one-half of specimens (31 of 60) exhibit a small-sized pair of foramina (less than 1/3 of the diameter of foramen ovale) and the other half (29 of 60) exhibit foramina that are barely perceptible as perforations in the pterygoid region. We also found that the 2 species are more easily distinguishable by the ratio of PL to IB: individuals of *P. rubiginosus* usually have longer palates but proportionally narrower constrictions than *P. alitonus*.

Quantitatively, the 2 species also markedly differ in 2 measurements related to the rostrum, i.e., the length of palate and maxillary tooththrow. The difference in these characters, however, is more conspicuous in sympatry. Individuals of *P. rubiginosus* are larger in areas where the species range overlaps with *P. alitonus*. Therefore, the morphological variation of specimens of *P. rubiginosus* from the allopatric area (Brazilian samples of PA, PI, MT, and RO) seems to slightly blur this clear separation, since they exhibit smaller sizes and more polymorphic

cranial characters. The DF1 values in the different populations (Fig. 4) clearly outline such a distinctive pattern. Also, most of the individuals displaying small pairs of foramina or higher IB/PL ratios (0.45) are found in the allopatric distribution of *P. rubiginosus*. Our results suggest that specimens of *P. rubiginosus* occurring in sympatry with *P. alitonus* are more dissimilar than those occurring in allopatry, at least concerning the cranial phenotype.

The difference in morphology of *P. rubiginosus* in sympatry and allopatry with *P. alitonus* agrees with character displacement theory, which hypothesizes that phenotypic differences between species are enhanced where they occur together, to minimize or avoid resource competition or reproductive interactions between them (Pfennig and Pfennig 2009). Character displacement is believed to be a ubiquitous phenomenon in nature, although difficult to test in the face of alternative hypothesis explaining divergence on closely related sympatric species (sensu Losos 2000; Pfennig and Pfennig 2009). This competitively mediated divergence can finalize a process of allopatric speciation (Pfennig and Pfennig 2010). Several criteria have been proposed to test the strength of a particular adaptive hypothesis, including that phenotypic divergence: 1) must be nonrandom, 2) has to show a genetic basis, and 3) should reflect differences in resource use (Schluter and McPhail 1992; Losos 2000). Testing for all premises is beyond the scope of the present study. Our ANOVA results, however, do reveal a significant morphological divergence between sympatric and allopatric populations of *P. rubiginosus* for all cranial characters (Supplementary Data SD3). Alternatively, half of the characters are different when allopatric populations are tested among themselves, and no significant differences are found in the variable means among the sympatric populations (data not shown). This finding seems to meet the 1st criteria, that differences are nonrandom. It also suggests a higher phenotypic variation within the more widespread species *P. rubiginosus*, especially in its allopatric range with *P. alitonus*, a factor that is expected to be present in species undergoing character displacement (Pfennig and Pfennig 2009). *P. alitonus*, on the other hand, is more conservative in its size throughout its known (smaller) range. In addition, geographic variation in the echolocation calls has also been reported for these 2 species (Baucells et al. 2017). It is possible that the variation in the calls of *P. rubiginosus* is even greater in its allopatric area of occurrence. However, more evidence is needed in the southernmost region of the *P. rubiginosus* distribution to test this hypothesis.

Echolocation calls and foraging behavior.—Both *P. rubiginosus* and *P. alitonus* find prey using high-duty cycle signals that consist of a long CF component followed by a short downward FM terminal component. Long CF-FM signals are associated with Doppler shift compensation and are used by bats that search for prey in narrow spaces (Schnitzler and Denzinger 2011; Denzinger and Schnitzler 2013). Pulse duration is similar in the 2 species but the FME always differ by 5–7 kHz between them (al. 2017). There is a clear relationship between call frequency and the size of prey that insectivorous bats can detect. While bats calling at higher frequencies are

expected to catch smaller insects with more efficiency, those emitting at lower frequencies will target larger insects more successfully (Jones 1997; Kingston et al. 2001). However, it has been discussed that small differences in the call frequencies such as the one described between *P. rubiginosus* and *P. alitonus* are not sufficient to significantly influence target specificity, and thus cannot promote resource partitioning associated with prey size (Jones and Barlow 2004; Kingston et al. 2001; Clare et al. 2013).

Alternatively, the hypothesis of competitive exclusion between these 2 species, as suggested by the cranial phenotypic traits, may be taking place spatially instead of acting on prey-size selection. Accordingly, the 2 species may search similarly for their targets, but exploit distinct microhabitats in their sympatric area of occurrence. Because *P. rubiginosus* emits calls at lower frequencies, its foraging strategy might be more associated with less cluttered environments, while the higher frequencies of *P. alitonus* increase the target detection in more cluttered areas. For example, *P. rubiginosus* crosses open areas between islands created by the Balbina hydroelectric dam, while *P. alitonus* is restricted to the continuous forest (Ponzio 2017). Also, some Amazonian localities characterized

as well as the Cerrado of Brazil, a savanna-like environment, are areas where only *P. rubiginosus* occurs. This pattern seems to be more related to some specific requirement of *P. alitonus* than an actual preference of *P. rubiginosus* since the latter also forages in cluttered areas when not in sympatry with *P. alitonus* (De Oliveira et al. 2015; Ponzio 2017). Based on this observation, we hypothesize that *P. rubiginosus* may be exploiting more open habitats within the Amazon Region, such as the Amazonian savannas, whereas *P. alitonus* is mostly restricted to more cluttered microhabitats while searching for food.

Finally, distinctive patterns of microspatial segregation of these 2 species might be related to the molecular results that we observed. The lower nucleotide diversity and the large number of individuals sharing 1 single haplotype in *P. rubiginosus* may be a consequence of its high dispersal capability and, consequently, the higher gene flow among the populations. *P. alitonus* seems to exhibit a smaller geographic range and shows a greater intraspecific structuring that may reflect its foraging behavior associated with highly cluttered areas and limitations to cross open landscapes.

Inconsistencies between morphology and molecular data.—Two specimens included in our analysis presented disparities regarding molecular and morphological data. MPEG 41678 (Itaituba, Brazil) has a mitochondrial haplotype of *P. rubiginosus*; its phenotype, however, resembles *P. alitonus*, being small (FL = 58.9 mm; GLS = 21.2 mm) and exhibiting diagnostic cranial characters such as small pterygoid foramina, PL = 9.71 mm and high IB/PL ratio (0.47). AMNH 269115 (Cayenne, French Guiana), on the other hand, had a haplotype of *P. alitonus* while its cranial features mostly agree with *P. rubiginosus* (GLS = 23.5 mm; PL = 10.71 mm; IB/PL ratio = 0.44; large pterygoid foramina). Three distinct interpretations of these results are possible: 1) the defined

cranial morphological characters are not completely consistent for species differentiation, exhibiting polymorphic states for some individuals of the distinct species; 2) the mtDNA exhibits some level of incomplete lineage sorting between the species; and 3) there is ongoing gene flow between the species due to secondary contact, leading to introgression of mtDNA from one species to the other. We cannot assert which of these scenarios is the most likely without a deeper investigation including more molecular markers, preferably from independent systems.

Regarding the 1st assumption, our data show that there is variation for some of the cranial traits, including overlapping in some quantitative markers: these 2 specimens could represent outliers of our dataset, exhibiting the extreme of the variation expressed by the species molecularly delimited. On the other side, incomplete lineage sorting and interspecific gene flow (with introgression) are frequently described phenomena for recently diverged species (Degnan and Rosenberg 2009; Petit and Excoffier 2009). An event of introgression has been already reported in the genus *Pteronotus* (Pavan and Marroig 2016). One specimen of *P. gymnonotus* was found to have the mtDNA of *P. fulvus*, and these species diverged earlier than *P. alitonus* and *P. rubiginosus* (see Pavan and Marroig 2017). Therefore, introgression could be the cause of the inconsistency described above. These 2 specimens were collected in the sympatric area of occurrence of *P. rubiginosus* and *P. alitonus* but in opposite sides of this range (Cayenne = north of the sympatry zone; Itaituba = south of the sympatry zone), which means that, if introgression has caused this pattern, 2 independent events are necessary to explain the finding. In addition, the direction of the introgression would have to be different in the 2 events. Still, these phenomena do not preclude divergence in the rest of the genome, particularly if we assume that mtDNA is evolving under neutrality and, therefore, not related to the speciation process (Feder et al. 2013). Alternatively, specific cranial traits might be susceptible to evolutionary constraints due to highly specialized functions of the skull (Cheverud 1982; Santana and Lofgren 2013), such as the echolocation in high-duty cycle echolocating bats, and may be more tightly related to factors leading to reproductive isolation (Kingston et al. 2001; Clare et al. 2013). Based on this, we tentatively suggest that the disparity is being caused by conflicting information from the mtDNA, and we opted for identifying these specimens according to their phenotypes (Appendix I).

The case described above is an excellent example of how multiple sources of evidence are important for assigning individuals to species with confidence. Several studies have shown that bioacoustic data is a useful tool for the identification of bat species (Barataud et al. 2013; Thoisy et al. 2014; Baucells et al. 2016), and in the case of the subgenus *Phyllodia* it may be essential to understand the contact areas between species and aspects of their biology. In some cases, only morphological information is available for taxonomic studies, such as for fossils and highly endangered or extremely rare species. Nevertheless, in all other cases, an integrative approach should be attempted.

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SUPPLEMENTARY DATA

Supplementary data are available at *Journal of Mammalogy* online.

Supplementary Data SD1.—Phylogenetic hypothesis proposed for the genus *Pteronotus* according to Pavan and Marroig (2016). The phylogeny represents a species tree estimated from 6 molecular markers, depicting the relationships among major clades according to the multispecies coalescent approach (see Pavan and Marroig 2016 for more details).

Supplementary Data SD2.—The 9 cranial and mandibular measurements taken in the present study: palatal width (PW), palatal length (PL), greatest length of skull (GLS), interorbital breadth (IB), braincase breadth (BB), greatest width of rostrum measured across the labial margins of the alveoli of M2 (M2–M2), maxillary toothrow length (MxTL), mandibular length (MaL), and mandibular toothrow length (MaTL). The M2–M2 measurement was taken from a few specimens for the comparative morphology description (systematic account) and, therefore, was not included in the morphometric analysis. Scale bar = 0.5 cm. Drawings of skull by Ivan Akirov and taken from

Supplementary Data SD3.—Average measurements in allopatric (ALLO, $n = 38$) and sympatric (SYM, $n = 49$) populations of *Pteronotus rubiginosus* and result of analysis of variance (ANOVA) between them.

Supplementary Data SD4.—Classification matrix of the discriminant function analysis (DFA) performed with all specimens of both species compared to the classification rates of the DFA performed with only females (F) or males (M).

Supplementary Data SD5.—Comparison between the new species *Pteronotus alitonus* sp. nov. (ROM117576, left) and *P. rubiginosus* (ROM 117608, right) from Suriname (Blanche Marie Vallen, Sipaliwini), showing the difference in the size and shape of the nasal bones.

Supplementary Data SD6.—Comparison between the echolocation signals emitted by *Pteronotus rubiginosus* and *P. alitonus* sp. nov. The specimens were recorded on the islands of the Balbina Hydroelectric Reservoir (BHR), a man-made reservoir

(1°01′–1°55′S; 60°29′–59°28′W). These recordings were only used to illustrate the sonograms and were not part of the analyses since no bat was captured.

Supplementary Data SD7.—Comparison between the rostral pad (arrows) of *Pteronotus rubiginosus* (MZUSP 35519, left) and *P. alitonus* sp. nov. (MZUSP 35523, right). Both specimens are from Amaila Falls, Potaro-Siparuni, Guyana.

Supplementary Data SD8.—Comparison of the cranial morphology among the 3 species of *Phyllodia* distributed in the Guiana Shield. *Pteronotus rubiginosus* (ROM 98127, left) and *P. alitonus* sp. nov. (ROM 98128, middle) occur sympatrically in the Guianas and Brazilian Amazon, whereas *Pteronotus fuscus* (ROM 100871, right) is found in the highlands of north-western Guyana and in Venezuela.

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APPENDIX I

Specimens included in the molecular, morphometric, and acoustic datasets.

A = acoustic dataset; COI = sequence of the cytochrome oxidase I gene; FN = field number; GN = GenBank number; M = morphometric dataset; S = sex; VN = voucher number.

AMNH = American Museum of Natural History; IEPA =

= Instituto Nacional de Pesquisas
=

ESALQ; MHNG =

MPEG = = Museu de
= Royal Ontario

Museum; TTU = Texas Tech University; UEMA = Universidade
= Universidade Federal de

Minas Gerais.

BRA-AM = Brazilian state of Amazonas; BRA-AP = Brazilian
= Brazilian state of Mato Grosso;

BRA-PA = = Brazilian state of
= French

Guiana; GUY = Guyana; SUR = Suriname.

X = type of data included for the specimen.

VN	FN	S	Species	Locality	Country	Population	M	A	COI	GN
MZUSP	RB06	M	<i>Pteronotus rubiginosus</i>		Brazil	BRA-MT	X		X	KX590144
MZUSP	RB09	F	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590145
MZUSP	AC1334	F	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KF636799
MZUSP	AC1362	M	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590131
MZUSP	AC1396	F	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590153
MZUSP	FM43	F	<i>P. rubiginosus</i>	Jangada, Mato Grosso	Brazil	BRA-MT	X		X	KX590154
MZUSP	FM45	F	<i>P. rubiginosus</i>	Jangada, Mato Grosso	Brazil	BRA-MT	X		X	KX590155
MZUSP	FM47	F	<i>P. rubiginosus</i>	Jangada, Mato Grosso	Brazil	BRA-MT	X		X	KX590157
MZUSP	FM52	M	<i>P. rubiginosus</i>	Jangada, Mato Grosso	Brazil	BRA-MT	X		X	KX590159
MZUSP	MN70_14	M	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590160
MZUSP	MN70_30	F	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590162
MZUSP	MN70_42	F	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590146
MPEG	ACP32	F	<i>P. alitonus</i>		Brazil	BRA-AM	X		X	KX590147
MZUSP	ACPI35	F	<i>P. alitonus</i>		Brazil	BRA-AM	X		X	KX590148
MZUSP	ACPI58	F	<i>P. alitonus</i>		Brazil	BRA-AM	X		X	KX590149
MZUSP	ACPI73	F	<i>P. alitonus</i>		Brazil	BRA-AM	X		X	KX590129
MZUSP	AC1118	F	<i>P. rubiginosus</i>		Brazil	BRA-PA	X		X	KX590134
MZUSP	AC1524	M	<i>P. rubiginosus</i>		Brazil	BRA-PA	X			
MZUSP	AC1023	F	<i>P. rubiginosus</i>		Brazil	BRA-PA	X			
MPEG	AC254	F	<i>P. rubiginosus</i>		Brazil	BRA-PA	X			
MPEG	AC377	F	<i>P. rubiginosus</i>		Brazil	BRA-PA	X			
MZUSP	AC670	M	<i>P. rubiginosus</i>		Brazil	BRA-PA	X			
MZUSP	AC1911	F	<i>P. rubiginosus</i>		Brazil	BRA-PA	X		X	KX590139
MZUSP	AC1046	F	<i>P. rubiginosus</i>		Brazil	BRA-PA	X		X	KX590127
MZUSP	AC939	M	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590143
MZUSP	MN70_07	F	<i>P. rubiginosus</i>	Tesouro, Mato Grosso	Brazil	BRA-MT	X		X	KX590158
MZUSP	MN70_37	F	<i>P. rubiginosus</i>		Brazil	BRA-MT	X		X	KX590161
MZUSP	EG153	M	<i>P. rubiginosus</i>	Serra das Araras, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	EG183	F	<i>P. rubiginosus</i>	Serra das Araras, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	EG191	U	<i>P. rubiginosus</i>	Serra das Araras, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	EG206	M	<i>P. rubiginosus</i>	Serra das Araras, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	EG235	M	<i>P. rubiginosus</i>	Serra das Araras, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	EG246	M	<i>P. rubiginosus</i>	Serra das Araras, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	29504	M	<i>P. rubiginosus</i>	Pontes Lacerda, Mato Grosso	Brazil	BRA-MT	X			
MZUSP	35701	M	<i>P. rubiginosus</i>		Brazil	BRA-RO	X		X	KX590163
MZUSP	35699	F	<i>P. rubiginosus</i>		Brazil	BRA-RO	X		X	KX590135
MZUSP	AC1820	M	<i>P. rubiginosus</i>		Brazil	BRA-RO	X		X	KX590136
MZUSP	CCA1840	F	<i>P. rubiginosus</i>	Rio Madeira, UHE Jirau, Porto Velho,	Brazil	BRA-RO	X			
MZUSP	CCA1899	M	<i>P. rubiginosus</i>	Rio Madeira, UHE Jirau, Porto Velho,	Brazil	BRA-RO	X			
MZUSP	CCA919	M	<i>P. rubiginosus</i>	Rio Madeira, UHE Jirau, Porto Velho,	Brazil	BRA-RO	X			
MZUSP	CCA1226	U	<i>P. rubiginosus</i>	Rio Madeira, UHE Jirau, Porto Velho,	Brazil	BRA-RO	X		X	KX590150
MZUSP	CCA1267	U	<i>P. rubiginosus</i>	Rio Madeira, UHE Jirau, Porto Velho,	Brazil	BRA-RO	X		X	KX590151
MZUSP	CCA1324	U	<i>P. rubiginosus</i>	Rio Madeira, UHE Jirau, Porto Velho,	Brazil	BRA-RO	X		X	KX590152

VN	FN	S	Species	Locality	Country	Population	M	A	COI	GN
MZUSP	QTPC13	U	<i>P. rubiginosus</i>		Brazil	BRA-MT			X	KX590164
MZUSP	QTPC18	U	<i>P. rubiginosus</i>		Brazil	BRA-MT			X	KX590165
MZUSP	30035	M	<i>P. rubiginosus</i>		Brazil	BRA-PI	X			
MZUSP	30120	U	<i>P. rubiginosus</i>		Brazil	BRA-PI	X			
MZUSP	30142	U	<i>P. rubiginosus</i>		Brazil	BRA-PI	X			
MZUSP	30155	F	<i>P. rubiginosus</i>		Brazil	BRA-PI	X			
MZUSP	30210	F	<i>P. rubiginosus</i>		Brazil	BRA-PI	X			
MZUSP	35518	F	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X			MH017827
MZUSP	35519	M	<i>P. rubiginosus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X		X	MH017828
MZUSP	35520	M	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X		X	MH017827
MZUSP	35521	M	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X			
MZUSP	35522	F	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X			
MZUSP	35523	F	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X		X	MH017829
MZUSP	35525	M	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X		X	MH017830
MZUSP	35527	F	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY				
MZUSP	35528	F	<i>P. rubiginosus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X		X	MH017831
MZUSP	35529	F	<i>P. altionus</i>	Amalla Falls, Potaro-Siparuni	Guyana	GUY	X		X	MH017832
LM-ESALQ	ABX58	F	<i>P. rubiginosus</i>	Paca-Mirim, margem direita Rio Abacaxis,	Brazil	BRA-AM	X		X	MH017833
LM-ESALQ	ABX60	F	<i>P. rubiginosus</i>	margem direita Rio Madeira, Amazonas	Brazil	BRA-AM			X	
UFMG	BOC541	M	<i>P. rubiginosus</i>	Paca-Mirim, margem direita Rio Abacaxis,	Brazil	BRA-PA			X	KX590311
UFMG	BOC548	M	<i>P. rubiginosus</i>	margem direita Rio Madeira, Amazonas	Brazil	BRA-PA			X	KX590314
UFMG	BOC575	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590317
UFMG	BOC602	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590318
UFMG	BOC652	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590319
UFMG	BOC655	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590320
UFMG	VCT1072	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590323
UFMG	VCT1103	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590324
UFMG	VCT1135	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590325
UFMG	VCT1741	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590328
UFMG	VCT1773	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590331
UFMG	VCT1775	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590332
UFMG	VCT1869	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590334
UFMG	VCT1871	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590335
UFMG	VCT1873	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590336
UFMG	VCT2824	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590342
UFMG	VCT3048	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590343
UFMG	VCT3055	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590344
UFMG	VCT3846	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590347
UFMG	VCT3880	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590348
UFMG	VCT4953	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590349
UFMG	VCT5139	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590351
UFMG	VCT6099	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590352
UFMG	VCT6224	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590353
UFMG	VCT6228	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590355
UFMG	VCT6255	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590361
UFMG	VCT6370	F	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590362
UFMG	VCT6376	M	<i>P. rubiginosus</i>		Brazil	BRA-PA			X	KX590363

VN	FN	S	Species	Locality	Country	Population	M	A	COI	GN
UFMG	ARI03	M	<i>P. rubiginosus</i>		Brazil	BRA-MT			X	KX590276
UFMG	ARI08	F	<i>P. rubiginosus</i>		Brazil	BRA-MT			X	KX590277
UFMG	ARI16	F	<i>P. rubiginosus</i>		Brazil	BRA-MT			X	KX590278
UFMG	ARI33	M	<i>P. rubiginosus</i>		Brazil	BRA-MT			X	KX590279
UFMG	TG22	M	<i>P. rubiginosus</i>		Brazil	BRA-RO			X	KX590126
UEMA	U.62	F	<i>P. rubiginosus</i>		Brazil	BRA-PI			X	KX590281
UEMA	91.3	M	<i>P. rubiginosus</i>		Brazil	BRA-PI			X	KX590282
UEMA	92.1	U	<i>P. rubiginosus</i>		Brazil	BRA-PI			X	KX590280
IEPA	MAP276	M	<i>P. alitonus</i>	Rio Cupixi, RDS Iratapuru, Pedra Branca	Brazil	BRA-AP	X			
IEPA	MAP311	F	<i>P. alitonus</i>	Rio Cupixi, RDS Iratapuru, Pedra Branca	Brazil	BRA-AP	X		X	KX590104
IEPA	MAP313	M	<i>P. alitonus</i>	Rio Cupixi, RDS Iratapuru, Pedra Branca	Brazil	BRA-AP	X		X	KX590105
IEPA	MAP314	M	<i>P. rubiginosus</i>	Rio Cupixi, RDS Iratapuru, Pedra Branca	Brazil	BRA-AP	X		X	KX590106
IEPA	MAP324	M	<i>P. rubiginosus</i>	Rio Cupixi, RDS Iratapuru, Pedra Branca	Brazil	BRA-AP	X		X	KX590107
IEPA	MAP336	F	<i>P. alitonus</i>	P.N.Tumucumaque, Rio Anoteite, Oiapoque,	Brazil	BRA-AP	X		X	KX590108
IEPA	MAP358	F	<i>P. rubiginosus</i>	P.N.Tumucumaque, Rio Anoteite, Oiapoque,	Brazil	BRA-AP	X		X	KX590109
IEPA	MAP377	F	<i>P. rubiginosus</i>		Brazil	BRA-AP	X		X	KX590110
IEPA	MAP385	M	<i>P. rubiginosus</i>		Brazil	BRA-AP	X		X	KX590111
IEPA	MAP406	F	<i>P. rubiginosus</i>		Brazil	BRA-AP	X		X	KF636800
IEPA	MAP437	M	<i>P. rubiginosus</i>	km	Brazil	BRA-AP	X		X	KX590112
IEPA	MAP463	F	<i>P. rubiginosus</i>	km	Brazil	BRA-AP	X		X	KX590113
IEPA	MAP473	F	<i>P. rubiginosus</i>	km	Brazil	BRA-AP	X		X	KX590114
IEPA	MAP475	F	<i>P. rubiginosus</i>	km	Brazil	BRA-AP	X		X	KX590115
IEPA	UHSA22	F	<i>P. alitonus</i>		Brazil	BRA-AP	X		X	KX590116
IEPA	UHSA32	M	<i>P. alitonus</i>		Brazil	BRA-AP	X		X	KF636815
IEPA	UHSA36	F	<i>P. alitonus</i>		Brazil	BRA-AP	X		X	KX590117
IEPA	UHSA42	F	<i>P. alitonus</i>		Brazil	BRA-AP	X		X	KX590118
IEPA	UHSA15	F	<i>P. alitonus</i>		Brazil	BRA-AP	X		X	KX590119
INPA	PP01	M	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MH017834
INPA	PP02	M	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MH017835
INPA	PP03	F	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547497
INPA	PP05	M	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	
INPA	PP11	F	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547493
INPA	PP14	F	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547498
INPA	PP16	M	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MH017836
INPA	PP21	F	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547495
INPA	PP22	F	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547496

VN	FN	S	Species	Locality	Country	Population	M	A	COI	GN
INPA	6950	M	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547494
INPA	6951	F	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547499
INPA	6952	F	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MH017837
INPA	6953	F	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547500
INPA	6954	F	<i>P. alitonus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547501
INPA	6955	F	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-AM	X	X	X	MF547502
INPA	6085	F	<i>P. rubiginosus</i>	PDBFF, Manaus, Amazonas	Brazil	BRA-RO	X	X	X	KX590120
INPA	6100	M	<i>P. rubiginosus</i>		Brazil	BRA-RO	X	X	X	KX590121
INPA	6120	M	<i>P. rubiginosus</i>		Brazil	BRA-RO	X	X	X	KX590122
INPA	6129	M	<i>P. rubiginosus</i>		Brazil	BRA-RO	X	X	X	KX590123
INPA	6162	F	<i>P. rubiginosus</i>		Brazil	BRA-RO	X	X	X	KX590124
INPA	6172	M	<i>P. rubiginosus</i>		Brazil	BRA-RO	X	X	X	KX590125
AMNH	267283	F	<i>P. rubiginosus</i>	Paracou	French Guiana	FGU	X			
AMNH	267286	M	<i>P. rubiginosus</i>	Paracou	French Guiana	FGU	X			KX590080
AMNH	269115	M	<i>P. rubiginosus</i>	Paracou	French Guiana	FGU	X		X	KX590081
AMNH	267851	F	<i>P. alitonus</i>	Paracou	French Guiana	FGU	X			
AMNH	267288	M	<i>P. rubiginosus</i>	Paracou	French Guiana	FGU	X			
AMNH	267284	M	<i>P. rubiginosus</i>	Paracou	French Guiana	FGU	X			
AMNH	267285	M	<i>P. rubiginosus</i>	Paracou	French Guiana	FGU	X			
AMNH	267406	F	<i>P. alitonus</i>	Paracou	French Guiana	FGU	X			
AMNH	267405	F	<i>P. alitonus</i>	Paracou	French Guiana	FGU	X		X	KX590086
ROM	100391	F	<i>P. rubiginosus</i>	Mango Landing, Corentyne River, East Berbice-Corentyne	Guyana	GUY	X		X	JF448242
ROM	100397	F	<i>P. rubiginosus</i>	Mango Landing, Corentyne River, East Berbice-Corentyne	Guyana	GUY	X		X	JF448220
ROM	100427	F	<i>P. alitonus</i>	Mango Landing, Corentyne River, East Berbice-Corentyne	Guyana	GUY	X		X	JF448247
ROM	97957	M	<i>P. rubiginosus</i>	Karanambo, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448258
ROM	98128	F	<i>P. alitonus</i>	Kurupukari, East Bank, Essequibo River, Upper Demerara-Berbice	Guyana	GUY	X		X	JF448217
ROM	97963	M	<i>P. rubiginosus</i>	Annai, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448259
ROM	97964	M	<i>P. rubiginosus</i>	Annai, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448260
ROM	98127	F	<i>P. rubiginosus</i>	Kurupukari, East Bank, Essequibo River, Upper Demerara-Berbice	Guyana	GUY	X		X	JF448216
ROM	102991	M	<i>P. rubiginosus</i>	5 km SE Surama, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448393
ROM	103375	F	<i>P. rubiginosus</i>	Tropenbos, 20 km SSE Mabura Hill, Upper Demerara-Berbice	Guyana	GUY	X		X	JF448377
ROM	104705	F	<i>P. alitonus</i>	Iwokrama Reserve, 25 km SSW Kurupukari, Potaro-Siparuni	Guyana	GUY	X		X	JF448408
ROM	106575	F	<i>P. rubiginosus</i>	Chodikar River, 55 km SW Gunn's Strip, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448290
ROM	106585	F	<i>P. alitonus</i>	Chodikar River, 55 km SW Gunn's Strip, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448291
ROM	106776	F	<i>P. alitonus</i>	Gunn's Strip, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448295
ROM	102973	M	<i>P. alitonus</i>	5 km SE Surama, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448391

VN	FN	S	Species	Locality	Country	Population	M	A	COI	GN
ROM	103374	F	<i>P. alitonus</i>	Tropenbos, 20 km SSE Mabura Hill, Upper Demerara-Berbice	Guyana	GUY	X		X	JF448376
ROM	103420	M	<i>P. rubiginosus</i>	Tropenbos, 20 km SSE Mabura Hill, Upper Demerara-Berbice	Guyana	GUY	X		X	JF448379
ROM	106659	M	<i>P. alitonus</i>	Kamoa River, 50 km SWW Gunn's Strip, Upper Takutu-Upper Essequibo	Guyana	GUY	X		X	JF448292
ROM	120275	F	<i>P. alitonus</i>	Iconja Landing, Sipaliwini River, Sipaliwini	Suriname	SUR	X		X	HQ545477
ROM	120294	M	<i>P. alitonus</i>	Iconja Landing, Sipaliwini River, Sipaliwini	Suriname	SUR	X		X	HQ919657
ROM	120589	F	<i>P. alitonus</i>	Kutari River Camp, Sipaliwini	Suriname	SUR	X		X	JQ601193
ROM	120631	M	<i>P. alitonus</i>	Sipaliwini River Camp, Sipaliwini	Suriname	SUR	X		X	JQ601223
ROM	120645	F	<i>P. alitonus</i>	Sipaliwini River Camp, Sipaliwini	Suriname	SUR	X		X	JQ601237
ROM	120408	F	<i>P. rubiginosus</i>	Sipaliwini Village, Sipaliwini	Suriname	SUR	X		X	HQ919758
ROM	117576	F	<i>P. alitonus</i>	Blanche Marie Vallén, Sipaliwini	Suriname	SUR	X		X	EU096919
ROM	117608	M	<i>P. rubiginosus</i>	Blanche Marie Vallén, Sipaliwini	Suriname	SUR	X		X	EU096918
ROM	117654	F	<i>P. rubiginosus</i>	Blanche Marie Vallén, Sipaliwini	Suriname	SUR	X		X	EU096924
ROM	117282	M	<i>P. rubiginosus</i>	Bakhuis, Transect 13, Sipaliwini	Suriname	SUR	X		X	EU096906
ROM	117338	F	<i>P. rubiginosus</i>	Bakhuis, Transect 14, Sipaliwini	Suriname	SUR	X		X	EU096908
ROM	117545	F	<i>P. alitonus</i>	Bakhuis, Area 8 Recon Fly Camp, Sipaliwini	Suriname	SUR	X		X	EU096917
ROM	113978	M	<i>P. rubiginosus</i>	Brownsberg Nature Park, jeep trail, Brokopondo	Suriname	SUR			X	JF448284
ROM	115699	F	<i>P. alitonus</i>	Iwokrama field Station, Potaro-Siparuni	Guyana	GUY	X	X	X	JF448175
MHNG	1972_050	U	<i>P. rubiginosus</i>		French Guiana	FGU	X			
MHNG	1972_051	U	<i>P. alitonus</i>		French Guiana	FGU	X			
MHNG	1972_052	U	<i>P. alitonus</i>	Kaw: Grotte Mathilde	French Guiana	FGU	X			
MHNG	1978_076	M	<i>P. rubiginosus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636795
MHNG	1978_077	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636809
MHNG	1978_078	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636811
MHNG	1978_079	M	<i>P. rubiginosus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636797
MHNG	1978_080	F	<i>P. rubiginosus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636798
MHNG	1978_081	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636810
MHNG	1978_082	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636805
MHNG	1978_083	M	<i>P. rubiginosus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636796
MHNG	1978_084	F	<i>P. rubiginosus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636794
MHNG	1978_085	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636807
MHNG	1978_086	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636806
MHNG	1978_087	M	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636812
MHNG	1978_088	F	<i>P. rubiginosus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636793
MHNG	1978_089	F	<i>P. alitonus</i>	Regina, Grotte Mathilde	French Guiana	FGU	X		X	KF636808
MHNG	1979_073	M	<i>P. rubiginosus</i>	Cacao: Va-Joua	French Guiana	FGU	X		X	
MHNG	1980_093	M	<i>P. alitonus</i>		French Guiana	FGU	X		X	
MHNG	1980_094	F	<i>P. alitonus</i>	Kaw: Grotte Mathilde	French Guiana	FGU	X		X	
MHNG	1983_043	M	<i>P. alitonus</i>	Cacao	French Guiana	FGU	X		X	
MHNG	1983_058	M	<i>P. alitonus</i>	Cacao	French Guiana	FGU	X		X	
MHNG	1983_064	M	<i>P. rubiginosus</i>	Cacao	French Guiana	FGU	X		X	
MHNG	1983_065	M	<i>P. rubiginosus</i>	Cacao	French Guiana	FGU	X		X	
MHNG	1983_069	M	<i>P. alitonus</i>	Cacao	French Guiana	FGU	X		X	
MHNG	C1228	U	<i>P. alitonus</i>	Montagne de Gouffres, Kaw	French Guiana	FGU			X	KX590097
MHNG	C1229	U	<i>P. alitonus</i>	Montagne de Gouffres, Kaw	French Guiana	FGU			X	KX590098

VN	FN	S	Species	Locality	Country	Population	M	A	COI	GN
MHNG		U	<i>P. alitonus</i>	Montagne de Couffres, Kaw	French Guiana	FGU			X	KX590099
TTU	TK151456	M	<i>P. alitonus</i>	Sipaliwini	Suriname	SUR			X	KX590264
TTU	TK145282	M	<i>P. rubiginosus</i>	Sipaliwini	Suriname	SUR			X	KX590259
TTU	TK145283	F	<i>P. rubiginosus</i>	Sipaliwini	Suriname	SUR			X	KX590260
TTU	TK145284	M	<i>P. alitonus</i>	Sipaliwini	Suriname	SUR			X	KX590261
TTU	TK145285	F	<i>P. rubiginosus</i>	Sipaliwini	Suriname	SUR			X	KX590262

APPENDIX II

List of specimens examined. This list refers to all specimens included in the morphological study regardless if they were included or not in the morphometric analysis. AMNH = American Museum of Natural History; IEPA =

= Instituto Nacional de Pesquisas da

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MPEG =

= Museu de

= Royal Ontario

Museum.

Pteronotus alitonus sp. nov. ($n = 82$).—**BRAZIL**:—_____: Rio Cupixi, Reserva de Iratapuru, Pedra Branca do Amapari, 0.58N 52.32W (IEPA 380, 415, 417); Parque Nacional Montanhas do Tumucumaque, Rio Anoteie, Oiapoque, 3.22N 52.02W (IEPA 456); Rio Jari, Laranjal do Jari, 0.62S 51.52W (IEPA 1833, 1843, 1847, 1853, 1893). **Amazonas**: Caverna Maroaga

(INPA 332, 339, 343, 345, 351, 358, 359, 361); Estrada S-2,

do Cuieiros, Base de Apoio ZF-2, Manaus, 2.59S 60.21W (INPA 2ZFII03, 2ZFII10); Manaus, 80 km N, 2.41S 59.88W (INPA 6942, 6945, 6947, 6948, 6949, 6950, 6952, 6954). ____: Itaituba, 6.05S 56.30W (MPEG 41678, MZUSP 35503, 35504, 35505). **FRENCH GUIANA**:—**Cayenne**: Sinnamary, Paracou, 5.38N 52.95W (AMNH 267851, 267405, 267406). ____: Grotte Mathilde, 4.52N 52.12W (MHNG 1972.051, 1978.077, 1978.078, 1978.081, 1978.082, 1978.085, 1978.086, 1978.087, 1978.089, 1980.094). **Roura**: Cacao, 4.57N 52.45W (MHNG

4.62N 52.28W (MHNG 1972.052). **Saint Elie**

5.02N 53.66W (MHNG 1980.093). **GUYANA**:—**East Berbice-**

Corentyne: Mango Landing, Corentyne River, 5.17N 57.3W

(ROM 100427). **Potaro-Siparuni**: Amaila Falls, 5.52N 59.26W

(MZUSP 35518, 35520, 35521, 35522, 35523, 35524, 35525,

35526, 35527, 35529); Iwokrama Reserve, 25 km SSW of

Kurupukari, 4.47N 58.78W (ROM 104705). **Upper Demerara-**

Berbice: Kurupukari, East Bank, Essequibo River, 4.67N

58.68W (ROM 98128); Tropenbos, 20 km SSE of Mabura Hill,

5.15N 58.7W (ROM 103374). **Upper Takutu-Upper Essequibo**:

Chodikar River, 55 km SW of Gunn's Strip, 1.37N 58.77W

(ROM 106585); Gunn's Strip, 1.65N 58.63W (ROM 106776); 5

km SE of Surama, 4.1N 59.05W (ROM 102973); Kamao River,

50 km SWW of Gunn's Strip, 1.54N 58.85W (ROM 106659).

SURINAME:—**Sipaliwini**: Iconja Landing, Sipaliwini River,

1.99N 56.09W (ROM 120275, 120294); Kutari River Camp,

2.18N 56.79W (ROM 120589); Sipaliwini River Camp, 2.29N

56.61W (ROM 120631, 120645); Blanche Marie Vallen, 4.76N

56.88W (ROM 117576); Bakhuis, Area 8 Recon Fly Camp,

4.45N 56.86W (ROM 117545).

Pteronotus rubiginosus ($n = 102$).—**BRAZIL**:—_____: Rio

Cupixi, Reserva de Iratapuru, Pedra Branca do Amapari,

0.58N 52.32W (IEPA 418, 428); Parque Nacional Montanhas

do Tumucumaque, Rio Anoteie, Oiapoque, 3.22N 52.02W (IEPA 476); Parque Nacional Montanhas do Tumucumaque, 554); Fazenda Aricari/BR156, km 147, Tartarugalzinho, 0.95N 51.25W (IEPA 722, 748, 758, 760). Amazonas: Caverna 59.95W (INPA 333, 353); Manaus, 80 km N, 2.41S 59.88W (INPA 6941, 6943, 6944, 6946, 6951, 6955); Paca-Mirim, margem direita Rio Abacaxis, 4.59S 58.22W (LM-ABX58); 2.59S, 60.21W (INPA 2ZFIII15). Mato Grosso 55.77W (35151, 35152); Jangada, 15.27S 55.22W (MZUSP 35147, 35148, 35149, 35150); Pontes Lacerda, 15.2S 59.37W 35155); Serra das Araras, 15.48S 57.19W (MZUSP 35702, 35703, 35704, 35705, 35706, 35707). ____: Floresta Nacional 38828, MZUSP 35506, 35507, 35508, 35509, 35510, 35512). ____ 44.17W (MZUSP 30035, 30120, 30142, 30155, 30210, 30226). ____ Pedras, Porto Velho, 9.16S 64.63W (INPA 6085, 6093, 6100, 9.16S 64.71W (INPA JRDA27); Rio Jaci-Parana, Porto velho - margem direita do rio madeira, 9.45S 64.37W (INPA 6116, (MZUSP 35511, 35516, 35517); Vilhena, 12.72S 60.26W

(MZUSP 35699, 35700, 35701). **FRENCH GUIANA**:—Cayenne: Sinnamary, Paracou, 5.38N 52.95W (AMNH 267283, 267284, 267285, 267286, 267288, 269115). ____: Grotte Mathilde, 4.52N 52.12W (MHNG 1978.076, 1978.079, 1978.080, 1978.083, 1978.084, 1978.088). Roura: Cacao, Natural Preservation, 4.62N 52.28W (MHNG 1972.050). **GUYANA**:—East Berbice-Corentyne: Mango Landing, Corentyne River, 5.17N 57.3W (ROM 100391, 100397). Potaro-Siparuni: Amaila Falls, 5.52N 59.26W (MZUSP 35519, 35528). Upper Demerara-Berbice: Kurupukari, East Bank, Essequibo River, 4.67N 58.68W (ROM 98127); Tropenbos, 20 km SSE of Mabura Hill, 5.15N 58.7W (ROM 103375, 103420). Upper Takutu-Upper Essequibo: Annai, 3.95N 59.13W (ROM 97963, 97964); Chodikar River, 55 km SW of Gunn's Strip, 1.37N 58.77W (ROM 106575); 5 km SE of Surama, 4.1N 59.05W (ROM 102991); Karanambo, 3.75N 59.3W (ROM 97957). **SURINAME**:—Sipaliwini: Sipaliwini Village, 2.03N 56.12W (ROM 120408); Blanche Marie Vallen, 4.76N 56.88W (ROM 117608, 117654); Bakhuis, transect 13, 4.55N 57.06W (ROM 117282); Bakhuis, Transect 14, 4.58N 57.05W (ROM 117338). *Pteronotus fuscus* ($n = 15$).—**GUYANA**:—Barima-Waini: Baramita, Old World, 7.36N 60.48W (ROM 100871, 100948, 101046). **VENEZUELA**:—Aragua: Rancho Grande, 10.37N 67.68W (AMNH 144842, 144845); Carabobo: San Esteban, 10.43N 68.02W (AMNH 31565, 31566, 31568, 31569, 31570, 31571, 31576); Las Quigas, 10.40N 68.00W (AMNH 31561, 31563, 31564).