- 1 Running Head: Increasing Nutrient Enrichment Effects
- 2 INCREASING EFFECTS OF CHRONIC NUTRIENT ENRICHMENT ON PLANT DIVERSITY
- 3 Loss and Ecosystem Productivity
- 4 Eric W. Seabloom^{1*}, Peter B. Adler², Juan Alberti³, Lori Biederman⁴, Yvonne M. Buckley⁵,
- 5 Marc W. Cadotte⁶, Scott L. Collins⁷, Laura Dee⁸, Philip A. Fay⁹, Jennifer Firn¹⁰, Nicole
- 6 Hagenah¹¹, W. Stanley Harpole^{12,13,14}, Yann Hautier¹⁵, Andy Hector¹⁶, Sarah E. Hobbie¹, Forest
- 7 Isbell¹, Johannes M. H. Knops¹⁷, Kimberly J. Komatsu¹⁸, Ramesh Laungani¹⁹, Andrew
- 8 MacDougall²⁰, Rebecca L. McCulley²¹, Joslin L. Moore²², John W. Morgan²³, Timothy Ohlert⁷,
- 9 Suzanne M. Prober²⁴, Anita C. Risch²⁵, Martin Schuetz²⁵, Carly J. Stevens²⁶, Elizabeth T. Borer¹
- ¹Department of Ecology, Evolution, and Behavior, University of MN, St. Paul, MN, 55108 USA. ²Dept. of Wildland
- Resources and the Ecology Center, 5230 Old Main, Utah State University, Logan, UT 84322 USA. ³Instituto de
- 12 Investigaciones Marinas y Costeras (IIMyC), FCEyN, UNMdP-CONICET. CC1260 (7600) Mar del Plata,
- 13 Argentina. ⁴Iowa State University, Ecology, Evolution, & Organismal Biology, 2200 Osborn Dr, Ames IA 50011
- 14 USA. ⁵Zoology, School of Natural Sciences, Trinity College Dublin, Dublin 2, Ireland. ⁶Department of Biological
- 15 Sciences, University of Toronto-Scarborough, 1265 Military Trail, Toronto, ON M1C 1A4, Canada. ⁷Department of
- 16 Biology, University of New Mexico, Albuquerque, NM 87131 USA. 8 University of Colorado Boulder, Department of
- 17 Ecology and Evolutionary Biology, Boulder CO USA. 9USDA-ARS Grassland, Soil, and Water Laboratory, 808 E.
- 18 Blackland Road, Temple, TX, 76502 USA. ¹⁰Queensland University of Technology (QUT), School of Earth,
- 19 Environmental and Biological Sciences, Science and Engineering Faculty, Brisbane, QLD, 4001, Australia.
- 20 II Mammal Research Institute, Department of Zoology and Entomology, University of Pretoria, Pretoria, South
- 21 Africa. ¹²Helmholtz Center for Environmental Research UFZ, Department of Physiological Diversity,
- 22 Permoserstrasse 15, 04318 Leipzig, Germany. ¹³German Centre for Integrative Biodiversity Research (iDiv),
- Deutscher Platz 5e, 04103 Leipzig, Germany. ¹⁴Martin Luther University Halle-Wittenberg, am Kirchtor 1, 06108
- Halle (Saale), Germany. ¹⁵Ecology and Biodiversity Group, Department of Biology, Utrecht University, Padualaan

25	8, 3584 CH Utrecht, The Netherlands. ¹⁶ Dept. of Plant Sciences, University of Oxford, OX1 3RB, UK. ¹⁷ School of
26	Biological Sciences, University of Nebraska, Lincoln, NE 68502, USA. ¹⁸ Smithsonian Environmental Research
27	Center, 647 Contees Wharf Road, Edgewater, MD 21037 USA. ¹⁹ Doane University, Department of Biology 1014
28	Boswell Ave Crete, NE 68333 USA. ²⁰ Department of Integrative Biology, University of Guelph, Guelph, Ontario,
29	Canada N1G 2W1. ²¹ Department of Plant & Soil Sciences, University of Kentucky, Lexington, KY 40536-0312.
30	²² School of Biological Sciences, Monash University, Clayton Campus VIC 3800, Australia. ²³ Department of
31	Ecology, Environment and Evolution, La Trobe University, Bundoora, Victoria 3086, Australia. ²⁴ CSIRO Land and
32	Water, Private Bag 5, Wembley, WA, Australia 6913. ²⁵ Swiss Federal Institute for Forest, Snow and Landscape
33	Research WSL, Zuercherstrasse 111, 8903 Birmensdorf, Switzerland. ²⁶ Lancaster Environment Centre, Lancaster
34	University, Lancaster, LA1 4YQ, UK.

* Email: seabloom@umn.edu

Abstract. Human activities are enriching many of Earth's ecosystems with biologically limiting mineral nutrients such as nitrogen (N), phosphorus (P), and potassium (K). In grasslands, this enrichment generally reduces plant diversity and increases productivity. The widely demonstrated positive effect of diversity on productivity suggests a potential negative feedback, whereby nutrient-induced declines in diversity reduce the initial gains in productivity arising from nutrient enrichment. In addition, plant productivity and diversity can be inhibited by accumulations of dead biomass, which may be altered by nutrient enrichment. Over longer timeframes, nutrient addition can increase soil fertility by increasing soil organic matter and nutrient pools. We examined the effects of 5-11 years of nutrient addition at 47 grasslands in twelve countries. Nutrient enrichment increased aboveground live biomass and reduced plant diversity at nearly all sites, and these effects became stronger through time. We did not find evidence that nutrient-induced losses of diversity reduced the positive effects of nutrients on biomass, however nutrient effects on live biomass increased more slowly at sites where litter was also increasing, regardless of plant diversity. This work suggests that short-term experiments underestimate the long-term effects of human-caused nutrient enrichment on global, grassland ecosystems.

Keywords: Nutrient Network, NutNet, Community Ecology, Biodiversity, Ecosystem Ecology

Grasslands

55

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

56 Introduction

57	Human activities have increased the input of limiting nutrients into many ecosystems through
58	burning of fossil fuels, use of agricultural fertilizers, and other activities that create and distribute
59	biologically available nutrients (Vitousek et al. 1997a, Vitousek et al. 1997b, Steffen et al. 2015)
60	This enrichment can alter ecosystem productivity, biogeochemistry, species richness, and species
61	composition (Lawes and Gilbert 1880, Vitousek et al. 1997b, Elser et al. 2007, Hillebrand et al.
62	2007, Lewandowska et al. 2016, Simkin et al. 2016, Midolo et al. 2019). Because of the tight
63	coupling between fluxes of inorganic and organic material and the effects of biodiversity on
64	ecosystem processes (Hobbie 2008, Tilman et al. 2014, Hobbie 2015, Riggs et al. 2015), there is
65	the potential for indirect effects or feedbacks that may increase or dampen the effects of global
66	nutrient enrichment over time (Smith et al. 2009, Isbell et al. 2013a, Avolio et al. 2014, Smith et
67	al. 2015).
68	Experimental work in grassland ecosystems illustrates the potential for indirect effects and
69	feedbacks to alter the impacts of nutrient enrichment over time. In many types of grasslands
70	(e.g., alpine tundra, Africa grassveld, montane meadows, mesic grasslands, prairies, desert
71	grasslands, and old fields), addition of limiting mineral nutrients, such as nitrogen and
72	phosphorus, often rapidly leads to an increase in biomass production and a loss of plant diversity
73	(Lawes and Gilbert 1880, Elser et al. 2007, Borer et al. 2014b, Fay et al. 2015, Gasarch and
74	Seastedt 2015, Ward et al. 2017). Because of the negative effect of plant diversity loss on
75	
-	productivity (Reich et al. 2001, Tilman et al. 2014), nutrient-induced losses of biodiversity may
76	productivity (Reich et al. 2001, Tilman et al. 2014), nutrient-induced losses of biodiversity may diminish the effect of nutrient addition on productivity over time (Isbell et al. 2013a). Nutrient

79 biomass. This accumulated dead biomass may limit plant productivity over time by creating a 80 physical barrier or reducing light at ground level (Seastedt et al. 1991, Foster and Gross 1998, 81 Coleman and Levine 2007, Clark and Tilman 2010, Seabloom 2010, Hobbie 2015). 82 Alternatively, positive feedbacks may increase the strength of nutrient effects over time. For 83 example, increased productivity can increase soil organic matter (Conant et al. 2001, Fornara and 84 Tilman 2012), which in turn may increase water holding and cation exchange capacity, thereby 85 reducing leaching and promoting nutrient retention (Hobbie 2008, Fornara and Tilman 2012, 86 Isbell et al. 2013b, Hobbie 2015), potentially leading to further increases in productivity. If 87 nutrient effects on ecosystems develop slowly over time due to long term feedbacks (Smith et al. 88 2009, Isbell et al. 2013a, Avolio et al. 2014, Smith et al. 2015), our understanding of nutrient 89 effects on ecosystems may be biased, because most experiments in ecology are relatively short-90 term (< 5 years) (Elser et al. 2007, Silvertown et al. 2010, Pierik et al. 2011, Smith et al. 2015, 91 Hughes et al. 2017). 92 While fertilization experiments in grasslands have found that nutrient addition often initially 93 increases aboveground plant biomass and decreases plant diversity (Elser et al. 2007, Borer et al. 94 2014b, Fay et al. 2015), a few longer term (> 5 yr) experiments suggest that the positive effects 95 of nutrients on aboveground live biomass will either remain constant or decline with time 96 (Jenkinson et al. 1994, Isbell et al. 2013a, Avolio et al. 2014), while the effects on diversity or 97 richness may continue to increase (Isbell et al. 2013a, Harpole et al. 2016) (but see, Pierik et al. 98 2011). However, it is difficult to draw general inference about how nutrient effects might unfold 99 through time, because the conflicting evidence comes from experiments conducted at different 100 sites and using different treatments and sampling methodologies.

Here we address this knowledge gap by examining how increased nutrient supply (nitrogen, phosphorus, potassium, and micronutrients) affects aboveground live biomass, dead biomass (plant litter), and diversity over time using a 5-11 year nutrient-addition experiment replicated at 47 grassland sites in twelve countries on six continents that are part of the Nutrient Network (NutNet) distributed experiment (Borer et al. 2014a, Borer et al. 2017). This experimental network allows novel insights, because each site uses identical treatments and sampling methodologies, in contrast to meta-analyses in which methodological and biological signals are often confounded (Elser et al. 2007, Gruner et al. 2008, Midolo et al. 2019). Furthermore, these treatments are replicated across a wide range of grassland ecosystems including alpine tundra, montane meadows, mesic grasslands, prairies, desert grasslands, and old fields, thereby capturing important environmental gradients of elevation (0-4241 m), latitude (38° S to 69° N), mean annual precipitation (250 – 1900 mm yr⁻¹), soil nutrient levels (e.g., 270-1200 ppm N, 10—230 ppm P), species richness (3-26 g m⁻²), and aboveground live biomass (28 – 870 g m⁻²), our measure of net primary production. Analyses of short-term NutNet data have shown that nutrient addition generally decreases diversity and increases aboveground biomass (Elser et al. 2007, Borer et al. 2014b, Fay et al. 2015). We focus on grasslands, because these systems have been used extensively to test the interactive effects of nutrient enrichment and biodiversity on ecosystem processes (Elser et al. 2007, Tilman et al. 2014), and because grasslands account for about a third of terrestrial productivity (Chapin et al. 2002), making them important regulators of the global carbon cycle and climate. From a conservation perspective, grasslands are among the most endangered of the terrestrial ecosystems due to extensive conversion to human-dominated land uses, biological invasions, and welldocumented loss of diversity in response to nutrient enrichment (Hoekstra et al. 2005,

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

Ramankutty et al. 2008, Seabloom et al. 2013). Grasslands are also logistically tractable, as the small stature of the plants allows for smaller plots sizes. In addition, a few plant families of similar structure dominate grasslands worldwide, which makes it easier to compare results across worldwide.

In our analyses, we explicitly test (1) if the strength of nutrient effects on plant biomass and diversity changes over time, (2) if nutrient-induced diversity loss reduces the positive effects of nutrient addition on plant biomass, and (3) whether the nutrient-induced accumulation of dead biomass reduces in the positive effects of nutrient addition on plant biomass?

132 Methods

Study System and experimental design.

This work was conducted within the Nutrient Network Distributed Experiment, a globally replicated nutrient and herbivore manipulation experiment (NutNet; www.nutnet.org) (Borer et al. 2014a, Borer et al. 2017). For this study, we used data from 47 sites that had been receiving nutrient addition for 5-11 years (Appendix S1: Appendix S1: Table S7). At each of these sites, we replicated an identical experiment that factorially combined three nutrient-addition treatments each at two levels (Control or Fertilized): Nitrogen Addition (+N; 10 g N m⁻² yr⁻¹ as timed-release urea), Phosphorus Addition (+P; 10 g P m⁻² yr⁻¹ as triple-super phosphate), and Potassium and Micronutrient Addition (+K; 10 g K m⁻² yr⁻¹ as potassium sulfate and 100 g m⁻² yr⁻¹ of a micronutrient mix (6% Ca, 3% Mg, 12% S, 0.1% B, 1% Cu, 17% Fe, 2.5% Mn, 0.05% Mo, and 1% Zn). N, P, and K were applied annually, and the micronutrient mix was applied once at the start of the study to avoid toxicity of largely immobile micronutrients.

The N addition rates (10 g N m⁻² yr⁻¹) were chosen to overcome N limitation, and are higher than would be used if the goal were to mimic anthropogenic N deposition (Clark and Tilman 2008, Stevens et al. 2015). Ammonium nitrate was used in 2007 instead of urea at some sites, but there were no detectable differences between these N sources on plant biomass or diversity (Seabloom et al. 2015).

Each site was set up as a completely randomized blocked design typically with 3 replicates (ranged from 1-6). The experimental unit was a 5 x 5 m plot.

Starting in the year prior to fertilization, we annually sampled aboveground plant biomass and plant community composition. Biomass was sampled by clipping all aboveground biomass (live and dead) in two 0.1 m x 1 m strips, sorting current year's biomass (live biomass) from previous year's biomass (dead biomass), drying the biomass to a constant mass at 60 °C, and weighing it to the nearest 0.01 g. The locations of the biomass strips were moved each year to avoid effect of the harvest on production estimates in subsequent years. We sampled plant community composition in a permanent 1 m x 1 m quadrat by visually estimating the areal cover of each species independently, such that the total summed cover may exceed 100% in multi-layer communities.

We collected surface soil samples in the pre-experimental duration and every 3-5 years thereafter. Several 10 cm deep cores were combined for each plot and air dried to a constant mass. All soils were then shipped to a lab at University of Minnesota (USA) for storage and processing prior to analyses. A subsample (~80-100g) from each plot was homogenized by grinding the soil with two steel beads (Daisy Premium 3/8" steel slingshot ammo) with 90 minutes of vigorous shaking using a paint shaker. Approximately 20 (18-25) mg of ground,

homogenized soils were then packed into 5 x 9 mm tin capsules for carbon and nitrogen analysis using dry combustion gas chromatography on an Elemental Analyzer (Costech ECS 4010 CHNSO Analyzer, Valencia, California, USA) calibrated with the analytical standard, atropine (C₁₇H₂₃NO₃). 20g of the ground, homogenized soils were sent to Waypoint Analytical (Memphis, TN, USA) to measure major nutrients, micronutrients, soil pH, organic matter, cation exchange capacity, and texture of the soil (percent sand, silt and clay; only measured in the Control Plots). Phosphorus, potassium, calcium, magnesium, sulfur, boron, copper, iron, manganese, zinc, and sodium (in parts per million) were measured using the Mehlich-3 method. Soil pH was measured with a water pH meter on a 1:1 soil:water suspension. Cation exchange capacity (CEC), reported here as meg/100 g (milliequivalents of charge per 100 g of dry soil), is a measure of the capacity of soil surfaces to retain cations and is used as an indicator of quality and productivity of the soil. CEC was calculated using the ppm of Ca, Mg, and K reported from the Mehlich-3 method using the following relationship: CEC = (ppm Ca / 200) + (ppm Mg / 120)+ (ppm K / 390)). Percent organic matter content in soil was measured using the Loss on Ignition (LOI) method (combustion for two hours at 400 °C). Values of organic matter reported here were not treated with acid prior to combustion. Texture was measured using the hydrometer method. In brief, the soil sample was shaken with Sodium Hexametaphosphate (HMP) solution, and then transferred to a settling cylinder and mixed. The percent sand, silt, and clay particles were calculated from hydrometer density readings taken at 40 seconds and two hours.

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

Diversity Metrics

We measured plant diversity as the Effective Number of Species based on the Probability of Interspecific Encounter (*ENS*_{PIE}), as this measure provides a more scale-independent measure of potential biodiversity effects than richness and is much more robust to the effect of rare species

than species richness (Chase and Knight 2013). ENS_{PIE} estimates the number of equally abundant species and is equivalent to the Inverse Simpson's index of diversity. We calculated ENS_{PIE} as $1/\sum_{i=1}^{S} p_i^2$ where S is the total number of species and p_i is the proportion of the community cover represented by species i (Chase and Knight 2013). Simpson's evenness (E) is directly related to ENS_{PIE} through the relationship $E = ENS_{PIE}/S$ (Smith and Wilson 1996), thus we can factor diversity directly into its richness and evenness components through the relationship:

 $ENS_{PIE} = S * E$

 ENS_{PIE} was positively correlated with richness (r=0.74) but only weakly correlated with evenness (r=0.16) across all samples in our data. Richness and evenness were negatively correlated (r=-0.41).

200 Data Analyses

Ecosystem responses to nutrient enrichment could follow a variety of different shapes through time (e.g., no change or accelerating, decelerating, or linear change through time)(Smith et al. 2009, Smith et al. 2015), so we fit a flexible model that allowed for nutrient effects to increase or decrease in strength through time and for the shape of these curves to vary. Considering the case where nutrient effects are getting stronger through time, we might see a number of different curve shapes:

Decelerating effects through time: this would be the case where effects initially increase in strength, but this rate of change slows as the system starts equilibrating to a new level (e.g., Isbell et al. 2013a). In this case, a plot of effect size through time will be concave down.

- 210 Accelerating effects through time: in this case, we would see little response for a period of time
- and then a rapidly increasing effect size. This could occur in a buffered system, where a
- treatment needs to exceed a threshold before the system changes. In this case, a plot of effect
- size through time will be concave up.
- 214 Linear effects through time: While unrealistic in the long-run, it is possible we could observe a
- linear change in effect size over the duration of the experiment.
- We used a statistical model that allows all of these shapes. Specifically, our statistical model of
- biomass or diversity change through time was as follows:
- 218 $\log(y) = \beta_0 + \beta_1 N + \beta_2 \log(t) + \beta_3 \log(t) N$
- Where β_i is the *i*th regression coefficient, N is a dummy variable that indicates whether a sample
- is from a control plot (N=0) or a nutrient addition plot (N=1), and t is the duration of the
- experiment (years of treatment). For control plots (N=0), our predictive equation in log space is:
- $222 \quad \log(y) = \beta_0 + \beta_2 \log(t)$
- and for nutrient addition plots, our predictive equation was:
- 224 $\log(y) = (\beta_0 + \beta_1) + (\beta_2 + \beta_3)\log(t)$
- In the case where we were modeling plant biomass ($y=\log(\text{biomass})$, β_0 is the mean biomass
- (log g m⁻²) in a control plot at time = 1, β_0 is the marginal increase in biomass (log g m⁻²) due to
- adding nutrients in year 1, β_2 is the change in mean biomass (log g per log yr), and β_3 is the
- 228 marginal effect of adding nutrients on the change in mean biomass (log g per log yr).

229 In back-transformed space, we get following relationship:

230
$$y = e^{\beta_0 + \beta_1 N} t^{\beta_2 + \beta_3 N}$$

- For control plots (N=0), our predictive equation was:
- 232 $y = e^{\beta_0} t^{\beta_2}$
- In this case, if $\beta_2 < 1$ the rate of biomass change through time is slowing (Case 1), if $\beta_2 > 1$ the
- rate of biomass change through time is increasing (Case 2), and if $\beta_2 = 1$ there is a linear change
- 235 in biomass over time (Case 3). The mean biomass (g) in a control plot in year 1 (t=1) is e^{β_0} .
- For nutrient addition plots, our predictive equation is:
- 237 $y = e^{\beta_0 + \beta_1} t^{\beta_2 + \beta_3}$
- In this case, if $(\beta_2 + \beta_3) < 1$ the rate of biomass change through time is slowing (Case 1), if
- 239 $(\beta_2 + \beta_3) > 1$ the rate of biomass change through time is increasing (Case 2), and if $(\beta_2 + \beta_3) = 1$
- there is a linear change in biomass over time (Case 3). The mean biomass (g) in a fertilized plot
- 241 in year 1 (t=1) is $e^{(\beta_0 + \beta_1)}$.
- To test whether the effects of nutrient addition increased over time, we used a mixed-effects
- model with +N, +P, and +Kµ, and experimental duration. Experimental duration is the number of
- years the nutrient treatments had been applied; experimental duration was log₁₀ transformed. We
- included experimental duration and site as random effects. Because of the large number of
- parameters, the model with effects of all 8 nutrient treatments (+N \times +P \times +K μ) and their
- 247 interactions with experimental duration with random slope and intercepts for each site did not

converge. For this reason, we estimated treatment differences among sites (random slopes and intercept) over time using only the Control and All Nutrient (+N, +P, and +K μ). Model specifications in R are included with each table of regression results.

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

We tested for interactions between nutrient addition effects on live biomass, dead biomass, and diversity over time, to determine if nutrient effects became weaker over time at sites where nutrients lead to rapid declines in diversity or increases in dead biomass. To do this, we tested for correlations between the slopes of the nutrient effect on each of these metrics and experimental duration, as estimated by the nutrient effect by experimental duration interaction. For example, we tested if sites with increasing losses of diversity through time would also eventually have weakening nutrient effects on live biomass through time. Similarly, if nutrient enrichment induced increases in dead biomass that reduced productivity, we would expect a negative correlation between the nutrient by experimental duration interaction for live and dead biomass, indicating that nutrient effects on dead biomass were increasing and nutrient effects on live biomass were decreasing over time. We tested this relationship using standardized major axis (SMA) estimation (Warton et al. 2006). SMA summarizes the relationship between two variables, as opposed to predicting the value of one variable (i.e., the dependent variable in regression) using a second variable (the independent variable in regression). We used the sma function in the smatr R package.

We also used regressions to examine whether nutrient effects on live biomass, dead biomass, and plant diversity increased or decreased through time. At the site level, we included a water availability index (Mean Annual Precipitation/Potential Evapotranspiration), mean annual precipitation, mean annual temperature, mean diurnal temperature range, mean annual

temperature range, and temperature in the wettest quarter. The plot-level soil predictors were pre-treatment soil pH, total base cations (sum of Ca, Mg, and K), % soil N, and the soil C:N ratio. Note that % soil N, % soil organic matter, and % soil organic C are all highly correlated (r > 0.92 for all comparisons), so we only included % soil N in the regression models. Similarly, cation exchange capacity and base cations are highly correlated (r = 0.93), so we only include base cations in the regression models. The plant community measures included were pretreatment live biomass, dead biomass, and plant diversity. Finally, we included the change in nutrient effect over time as described in the correlation analysis above. For example, we included the change in nutrient effect over time on dead biomass and plant diversity as explanatory variables of the change in nutrient effect over time on live biomass. In our regression of site level characteristics that could predict the change in nutrient effects over time, many of our predictor variables co-varied (e.g., pre-treatment live biomass, soil N, soil C:N, and water availability index). To account for the potential for multiple models that had similar explanatory power due to this correlation, we used a multi-model approach, as described in Grueber et al. (2011) using the dredge and model.avg functions in the MuMIn library. We standardized the input variables using the arm library. In the model averaging, we included all models within 4 AICc units of the best model. We ran these models with and without soils data, as we were missing soils data at some sites. All analyses were conducted in R (v. 3.4.1; R Foundation for Statistical Computing, Vienna,

290 RESULTS

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

Austria). Mixed effects models were fit using the Imer function in the Ime4 R library.

In contrast to our expectations, we found increasing effects of nutrient addition on live biomass through time at most sites despite increasing losses of diversity (Figs. 1 & 2), and there was an overall significant interaction between treatment duration and fertilization (Table S2). While the nutrient effects grew stronger, the rate of change was decelerating in most cases. Nutrient enrichment, especially the addition of N, increased live biomass but did not have a consistent effect on dead biomass and there was not an interaction between fertilization and treatment duration (Figs. 1 & 3; Appendix S1: Tables S1 & S2). The effects of nutrient enrichment on live biomass increased at most sites for up to 11 years (Fig. 1; Appendix S1: Tables S1 & S2). In contrast to the consistent effects of nutrient addition on live biomass, nutrient effects on dead biomass became stronger at some sites and weaker at others (Fig. 1; Appendix S1: Tables S1 & S2). Nutrient addition caused increasingly strong reductions in plant diversity (ENS_{PIE}) at nearly all sites (Figs. 2 & 3; Appendix S1: Table S1 & S2), primarily due to the effects of N addition (Fig. 3, Appendix S1: Table S1). This loss in diversity was primarily caused by increasing losses of richness at all sites (Figs. 2 & 3). In contrast to richness, nutrient effects on evenness did not show a consistent increase over time (Fig. 2). The change in nutrient effects on diversity over time were not correlated with change in nutrient effects on live biomass over time (Fig. 4), however there was a negative correlation between the trajectory of nutrient effects on live and dead biomass over time (Fig. 4). In addition to the negative effects of dead biomass on live biomass, we found evidence that nutrient effects on live biomass were stronger at more productive sites with higher pre-treatment biomass (Appendix S1: Fig. S6; Appendix S1: Tables S4 & S5). Similarly, we found that

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

nutrients effects on diversity loss increased in strength faster at sites that had high diversity at the

start of the experiment (Appendix S1: Fig. S6; Appendix S1: Tables S4 & S5). Nutrient effects on dead biomass were weaker at sites with high levels of pre-treatment dead biomass when we included soil chemistry in the models which include 37 of 47 sites, but this effect was not significant for the full set of sites (Appendix S1: Fig. S6; Appendix S1: Tables S4 & S5). A potential determinant of dead biomass is domestic grazing, as the size sites with domestic grazers present had lower levels of dead biomass in control plots (p =0.011). There was no evidence that estimates of the change in nutrient effects over time were affected by among-site differences in experiment duration, which ranged from 5-11 years (Appendix S1: Tables S4 & S5).

Furthermore, the change in nutrient effects over time was not dependent on a site's climate or soil chemistry (Appendix S1: Tables S4 & S5).

Nutrient addition increased soil nutrient pools, carbon, and organic matter. Specifically, N addition led to increased soil C and N (Appendix S1: Fig. S7; Appendix S1: Table S6), and P addition increased soil P, cation exchange capacity, and base cations, possibly due to the calcium (Ca) in the P source (triple-super-phosphate; Appendix S1: Fig. S7; Appendix S1: Table S6).

328 DISCUSSION

Short- and long-term studies have identified differing responses to chronic nutrient enrichment. Although an analysis of nutrient responses at 42 grassland sites in the NutNet experiment found increasing effects of fertilization on live biomass over the short-term (3 years) (Fay et al. 2015), we expected, based on previous long-term fertilization experiments, that the positive effects of nutrients on aboveground live biomass would either plateau and remain constant or decline with time (Jenkinson et al. 1994, Isbell et al. 2013a, Avolio et al. 2014). In contrast, we found that the effect of nutrient enrichment on live biomass continued to increase at nearly all sites for 11 years

(Fig. 1; Appendix S1: Table S1 & S2), despite increasing losses of plant diversity and increasing dead biomass accumulation at some sites. Accumulation of dead biomass was the strongest predictor of the trajectory of nutrient effects over time, with increased biomass accumulation reducing the effects of nutrient on live biomass. We did not find evidence that increasing losses of diversity could cause nutrient effects on biomass to decline over time. A possible mechanism explaining the increasingly strong effects of fertilization on biomass is a concomitant increase in soil nutrient pools and buildup of soil C and organic matter (Fornara and Tilman 2012, Crowther et al. 2019), which could lead to increased water holding and cation exchange capacity and greater nutrient retention (Fornara and Tilman 2012, Isbell et al. 2013b, Hobbie 2015). We found that N addition led to increased soil C and N (Appendix S1: Fig. S7; Appendix S1: Table S4), and P addition increased soil P, cation exchange capacity, and base cations, possibly due to the calcium (Ca) in the P source (triple-super-phosphate; Appendix S1: Fig. S7; Appendix S1: Table S6). These nutrient-induced alterations to soil chemistry could act to further increase productivity. While many studies have examined nutrient effects on plant productivity and biomass (Elser et al. 2007), there is little basis for *a priori* expectations of how nutrient effects on dead biomass accumulation would change through time, although we did expect a general coupling between live and dead biomass (Hobbie 2015, Grace et al. 2016). Dead biomass accumulation can directly reduce plant growth by acting as a physical barrier and reducing light to young plants (Foster and Gross 1998, Coleman and Levine 2007, Clark and Tilman 2010, Seabloom 2010). In the longer term, dead biomass can affect productivity through the influence of senesced plant material on nutrient cycling, either facilitating or delaying nutrient release to plants (Hobbie 2015). In contrast to the consistently positive effects of nutrients on live biomass, nutrients

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

effects on the accumulation of dead biomass increased at some sites and decreased at others (Fig. 1; Appendix S1: Table S1 & S2). In meta-analysis spanning grasslands, forests, and tundra ecosystems, N effects on decomposition has been shown to vary depending on N addition rates, ambient N deposition rates, an litter quality (e.g., lignin content) (Knorr et al. 2005). Based on this work, the N addition rate we used (10 g N m⁻² yr⁻¹) would be expected to increase decomposition rates.

While we did not measure decomposition rates directly, our results suggest that nutrient addition may be increasing the rate of turnover of dead biomass, because fertilization increased biomass production and soil carbon, but it did not consistently increase total standing dead biomass. The effects of nutrients on decomposition are governed by complex feedbacks between primary productivity, plant tissue chemistry, and soil nutrient cycling (Hobbie 2008, Hobbie 2015), and the effects can vary among different locations (Hobbie 2008). Given the potential for increased nutrient supply rates to alter decomposition and ultimately carbon storage, developing a general understanding of nutrient supply on decomposition remains an important unresolved challenge in ecosystem ecology (Hobbie 2008).

Based on past work, we expected nutrients to continuously reduce diversity over time (Isbell et al. 2013a, Harpole et al. 2016), and we found this to be the case (Fig. 2; Appendix S1: Table S1 & S2), primarily due to the increasingly strong effects of N addition (Appendix S1: Table S1). Declines in diversity could arise either through reduced richness, due to increased extinction or reduced colonization rates, or reduced evenness, reflecting an increasingly skewed abundance distribution of species. We found that loss in diversity in response to nutrients was driven primarily by continuing richness loss at all sites (Fig. 2). In contrast to richness, nutrient addition did not have a consistent effect on evenness (Tables S1 & S2), and nutrient effects on evenness

did not show a consistent increase over time (Fig. 2), although evenness varied widely among sites and across years and nutrient addition did change evenness at individual sites (Figure S5). Past work suggested that the positive effects of nutrient enrichment on live biomass would decline over time at sites where there was a concurrent loss in plant diversity (Isbell et al. 2013a) or increase in dead biomass. We tested for this relationship by examining the correlations between the change over time in the nutrient effects on live biomass, dead biomass, and plant diversity. We did not find evidence of a negative feedback between fertilization induced diversity loss and live biomass; live biomass continued to increase in response to nutrients for up to a decade at nearly all sites. Furthermore, sites experiencing ongoing diversity loss did not have decelerating nutrient effects on live biomass through time (Fig. 4), as would be expected if the nutrient induced diversity loss were counteracting the effects of nutrients on productivity (Isbell et al. 2013a). It is possible that diversity loss could eventually start to counteract the positive effects of nutrient on plant biomass; the experiment that found this effect had been adding nutrients for more than 25 years (Isbell et al. 2013a). However, the longest running nutrient addition experiment has not shown large shifts after more than a century (Jenkinson et al. 1994). Ultimately, standardized, long-term experiments replicated at sites representing a range of conditions are necessary to distinguish biological from methodological effects of nutrient addition among sites and studies (Borer et al. 2017). For example, Avolio et al. (2014) found that a decade of N and P addition did not alter the richness in a tallgrass prairie community, while Isbell et al. (2013a) found that the effects of nutrient addition on richness strengthened through time, but in a different tallgrass prairie using different methods. In contrast, our standardized experiment collocated at with these previous studies showed a consistent response of increasingly strong declines in richness over time (konz.us & cdcr.us; Figures S1, S3, & S4),

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

405 suggesting that methodological differences, such nutrient addition rates or sampling protocols, 406 may explain differences in the inference between these two long-term experiments (Isbell et al. 407 2013a, Avolio et al. 2014). 408 Dead biomass accumulation has been shown to suppress plant recruitment in a variety of 409 herbaceous ecosystems (van der Valk 1986, Foster and Gross 1998, Coleman and Levine 2007, 410 Clark and Tilman 2010, Seabloom 2010), and our results supported the generality of this 411 relationship. The smallest change in nutrient effects on live biomass over time occurred at sites 412 where nutrient addition led to increased dead biomass accumulation (Fig. 4). 413 Our experiment was replicated across a wide range of conditions globally, underscoring the 414 generality of the strengthening of nutrient effects over time. At nearly all sites, nutrient effects on 415 live biomass and diversity became stronger over time. This suggests that short-term experiments 416 (< 5 years) will underestimate the effects of nutrient enrichment on ecosystems and communities. 417 Experiments spanning multiple decades have shown that short-term experiments may generate 418 biased estimates of long-term treatment effects (Reich et al. 2012, Isbell et al. 2013a), 419 highlighting the value of long-term ecological data (Hughes et al. 2017). 420 Over multiple decades, nutrient addition can lead to soil carbon accumulation (Fornara and 421 Tilman 2012), which can increase soil moisture and nutrient holding capacity (Hobbie 2015). 422 Via this pathway, plant growth may increase ecosystem productivity, rather than simply 423 depleting limiting nutrients. In our experiment, nitrogen addition consistently increased both soil 424 nitrogen and soil carbon across sites suggesting that nutrient enrichment could increase soil 425 fertility over time, and potentially inducing the continued increase in live biomass in response to 426 nutrients.

Many experiments and models have demonstrated that nutrient addition in grassland ecosystems can induce plant diversity declines (Lawes and Gilbert 1880, Miller et al. 2005, Clark et al. 2007, Harpole and Tilman 2007, Hautier et al. 2009, Borer et al. 2014c, Harpole et al. 2016, Ward et al. 2017), and the few long-term experiments have shown that these effects can continue for decades (Lawes and Gilbert 1880, Jenkinson et al. 1994, Isbell et al. 2013a, Ward et al. 2017). Here we have shown that an increasing effect of nutrients on diversity loss through time is a general phenomenon in many grassland ecosystems. This result suggests that current understanding, which is largely based on short-term experiments (< 5 years) (Elser et al. 2007), may be underestimating the severity of effect of nutrient enrichment on biodiversity. The mismatch between short experimental duration and the time for ecosystems to fully respond to nutrient enrichment is particularly problematic given that human activities are continuing to increase the supplies of limiting nutrients into many of Earth's ecosystems (Vitousek et al. 1997a, Galloway et al. 2008, Steffen et al. 2015). In interpreting our results, it is important to note that the N addition rates (10 g N m⁻² yr⁻¹) we use were chosen to overcome N limitation, and are higher than would be used if the goal were to mimic anthropogenic N deposition (Clark and Tilman 2008, Stevens et al. 2015). Lower rates of N addition would likely lead to weaker effects, though still positive, positive effects on live biomass and richness and could potentially inhibit decomposition rates (Knorr et al. 2005, Isbell et al. 2013a, Midolo et al. 2019). Human activities are concurrently altering a variety of interacting environmental factors that drive ecosystem processes and ultimately ecosystem services that are necessary for human wellbeing (Vitousek et al. 1997a, Vitousek et al. 1997b, Steffen et al. 2015), and the ecosystem-

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

level effects of many of these factors may not be fully evident for a decade or more (Reich et al.

2012, Isbell et al. 2013a). For this reason, long-term experiments are critical for predicting the effects of humans on ecological systems (Silvertown et al. 2010, Hughes et al. 2017).

Nevertheless, long-term experiments in ecology remain relatively rare, and funding such experiments is increasing difficult (Silvertown et al. 2010, Hughes et al. 2017). Human impacts on ecosystems also vary spatially, and ecologists have recently started replicating experiments at global scales (Borer et al. 2014a, Borer et al. 2017). Now that some distributed experiments have been in place for more than a decade, we can ask novel questions about the factors that determine the trajectory of ecosystem response to global change (Borer et al. 2017). Here we have shown a remarkably consistent increasing effect of nutrient addition on plant production and concomitant decline in diversity in wide array of grassland ecosystems including deserts, prairies, and alpine tundra. However, these responses were not fully coupled; nutrient-induced declines in diversity did not reduce the effects of nutrients on live biomass. These results suggest that, due to their short duration, many experiments to date have likely underestimated the effects of human driven eutrophication on biodiversity loss and ecosystem productivity.

ACKNOWLEDGMENTS

This work was generated using data from the Nutrient Network (http://www.nutnet.org) experiment, funded at the site scale by individual researchers. Coordination and data management have been supported by funding to E. Borer and E. Seabloom from the National Science Foundation Research Coordination Network (NSF-DEB-1042132) and Long Term Ecological Research (NSF-DEB-1234162 & DEB-1831944 to Cedar Creek LTER) programs, and the University of Minnesota's Institute on the Environment (DG-0001-13). We also thank the Minnesota Supercomputer Institute for hosting project data and the Institute on the Environment for hosting Network meetings.

473 REFERENCES

495

496

497

498

499

500

501

502

503

504

505

506

507

508

509

- Avolio, M. L., S. E. Koerner, K. J. La Pierre, K. R. Wilcox, G. W. T. Wilson, M. D. Smith, and S. L. Collins. 2014. Changes in plant community composition, not diversity, during a decade of nitrogen and phosphorus additions drive above-ground productivity in a tallgrass prairie. Journal of Ecology **102**:1649-1660.
- Borer, E. T., J. B. Grace, W. S. Harpole, A. S. MacDougall, and E. W. Seabloom. 2017. A decade of insights into grassland ecosystem responses to global environmental change. Nature Ecology & Evolution 1:0118.
- Borer, E. T., W. S. Harpole, P. B. Adler, E. M. Lind, J. L. Orrock, E. W. Seabloom, and M. D. Smith. 2014a. Finding generality in ecology: a model for globally distributed experiments. Methods in Ecology and Evolution 5:65-+.
- 484 Borer, E. T., E. W. Seabloom, D. S. Gruner, W. S. Harpole, H. Hillebrand, E. M. Lind, P. B. 485 Adler, J. Alberti, T. M. Anderson, J. D. Bakker, L. Biederman, D. Blumenthal, C. S. 486 Brown, L. A. Brudvig, Y. M. Buckley, M. Cadotte, C. J. Chu, E. E. Cleland, M. J. 487 Crawley, P. Daleo, E. I. Damschen, K. F. Davies, N. M. DeCrappeo, G. Z. Du, J. Firn, Y. 488 Hautier, R. W. Heckman, A. Hector, J. HilleRisLambers, O. Iribarne, J. A. Klein, J. M. 489 H. Knops, K. J. La Pierre, A. D. B. Leakey, W. Li, A. S. MacDougall, R. L. McCulley, B. 490 A. Melbourne, C. E. Mitchell, J. L. Moore, B. Mortensen, L. R. O'Halloran, J. L. Orrock, 491 J. Pascual, S. M. Prober, D. A. Pyke, A. C. Risch, M. Schuetz, M. D. Smith, C. J. 492 Stevens, L. L. Sullivan, R. J. Williams, P. D. Wragg, J. P. Wright, and L. H. Yang. 493 2014b. Herbivores and nutrients control grassland plant diversity via light limitation. 494 Nature **508**:517-+.
 - Borer, E. T., E. W. Seabloom, C. E. Mitchell, and J. P. Cronin. 2014c. Multiple nutrients and herbivores interact to govern diversity, productivity, composition, and infection in a successional grassland. Oikos **123**:214-224.
 - Chapin, F. S., M. P. A., and H. A. Mooney. 2002. Principles of terrestrial ecosystem ecology New York: Springer
 - Chase, J. M., and T. M. Knight. 2013. Scale-dependent effect sizes of ecological drivers on biodiversity: why standardised sampling is not enough. Ecology Letters **16**:17-26.
 - Clark, C. M., E. E. Cleland, S. L. Collins, J. E. Fargione, L. Gough, K. L. Gross, S. C. Pennings, K. N. Suding, and J. B. Grace. 2007. Environmental and plant community determinants of species loss following nitrogen enrichment. Ecology Letters 10:596-607.
 - Clark, C. M., and D. Tilman. 2008. Loss of plant species after chronic low-level nitrogen deposition to prairie grasslands. Nature **451**:712-715.
 - Clark, C. M., and D. Tilman. 2010. Recovery of plant diversity following N cessation: effects of recruitment, litter, and elevated N cycling. Ecology **91**:3620-3630.
 - Coleman, H. M., and J. M. Levine. 2007. Mechanisms underlying the impacts of exotic annual grasses in a coastal California meadow. Biological Invasions **9**:65-71.
- Conant, R. T., K. Paustian, and E. T. Elliott. 2001. Grassland management and conversion into grassland: Effects on soil carbon. Ecological Applications 11:343-355.
- Crowther, T. W., C. Riggs, E. M. Lind, E. T. Borer, E. W. Seabloom, S. E. Hobbie, J. Wubs, P. B. Adler, J. Firn, L. Gherardi, N. Hagenah, K. S. Hofmockel, J. M. H. Knops, R. L.
- McCulley, A. S. MacDougall, P. L. Peri, S. M. Prober, C. J. Stevens, and D. Routh. 2019.
- Sensitivity of global soil carbon stocks to combined nutrient enrichment. Ecology Letters **22**:936-945.

- Elser, J. J., M. E. S. Bracken, E. E. Cleland, D. S. Gruner, W. S. Harpole, H. Hillebrand, J. T. Ngai, E. W. Seabloom, J. B. Shurin, and J. E. Smith. 2007. Global analysis of nitrogen and phosphorus limitation of primary producers in freshwater, marine and terrestrial ecosystems. Ecology Letters **10**:1135-1142.
- Fay, P. A., S. M. Prober, W. S. Harpole, J. M. H. Knops, J. D. Bakker, E. T. Borer, E. M. Lind,
 A. S. MacDougall, E. W. Seabloom, P. D. Wragg, P. B. Adler, D. M. Blumenthal, Y.
 Buckley, C. J. Chu, E. E. Cleland, S. L. Collins, K. F. Davies, G. Z. Du, X. H. Feng, J.
 Firn, D. S. Gruner, N. Hagenah, Y. Hautier, R. W. Heckman, V. L. Jin, K. P. Kirkman, J.
 Klein, L. M. Ladwig, Q. Li, R. L. McCulley, B. A. Melbourne, C. E. Mitchell, J. L.
 Moore, J. W. Morgan, A. C. Risch, M. Schutz, C. J. Stevens, D. A. Wedin, and L. H.
 Yang. 2015. Grassland productivity limited by multiple nutrients. Nature Plants 1:5.
- Fornara, D. A., and D. Tilman. 2012. Soil carbon sequestration in prairie grasslands increased by chronic nitrogen addition. Ecology **93**:2030-2036.
 - Foster, B. L., and K. L. Gross. 1998. Species richness in a successional grassland: Effects of nitrogen enrichment and plant litter. Ecology **79**:2593-2602.

532

533

534

535

536

537

- Galloway, J. N., A. R. Townsend, J. W. Erisman, M. Bekunda, Z. C. Cai, J. R. Freney, L. A. Martinelli, S. P. Seitzinger, and M. A. Sutton. 2008. Transformation of the nitrogen cycle: Recent trends, questions, and potential solutions. Science **320**:889-892.
- Gasarch, E. I., and T. R. Seastedt. 2015. Plant community response to nitrogen and phosphorus enrichment varies across an alpine tundra moisture gradient. Plant Ecology & Diversity 8:739-749.
- Grace, J. B., T. M. Anderson, E. W. Seabloom, E. T. Borer, P. B. Adler, W. S. Harpole, Y.
 Hautier, H. Hillebrand, E. M. Lind, M. Pärtel, J. D. Bakker, Y. M. Buckley, M. J.
 Crawley, E. I. Damschen, K. F. Davies, P. A. Fay, J. Firn, D. S. Gruner, A. Hector, J. M.
 H. Knops, A. S. MacDougall, B. A. Melbourne, J. W. Morgan, J. L. Orrock, S. M.
 Prober, and M. D. Smith. 2016. Integrative modelling reveals mechanisms linking
 productivity and plant species richness. Nature 529:390-393.
- Gruner, D. S., J. E. Smith, E. W. Seabloom, S. A. Sandin, J. T. Ngai, H. Hillebrand, W. S.
 Harpole, J. J. Elser, E. E. Cleland, M. E. S. Bracken, E. T. Borer, and B. M. Bolker. 2008.
 A cross-system synthesis of consumer and nutrient resource control on producer biomass.
 Ecology Letters 11:740-755.
- Harpole, W. S., L. L. Sullivan, E. M. Lind, J. Firn, P. B. Adler, E. T. Borer, J. Chase, P. A. Fay,
 Y. Hautier, H. Hillebrand, A. S. MacDougallm, E. W. Seabloom, R. Williams, J. D.
 Bakker, M. W. Cadotte, E. J. Chaneton, C. J. Chu, E. E. Cleland, C. D'Antonio, K. F.
 Davies, D. S. Gruner, N. Hagenah, K. Kirkman, J. M. H. Knops, K. J. La Pierre, R. L.
 McCulley, J. L. Moore, J. W. Morgan, S. M. Prober, A. C. Risch, M. Schuetz, C. J.
 Stevens, and P. D. Wragg. 2016. Addition of multiple limiting resources reduces
 grassland diversity. Nature 537:93-96.
- Harpole, W. S., and D. Tilman. 2007. Grassland species loss resulting from reduced niche dimension. Nature **446**:791-793.
- Hautier, Y., P. A. Niklaus, and A. Hector. 2009. Competition for Light Causes Plant Biodiversity Loss After Eutrophication. Science **324**:636-638.
- Hillebrand, H., D. S. Gruner, E. T. Borer, M. E. S. Bracken, E. E. Cleland, J. J. Elser, W. S.
 Harpole, J. T. Ngai, E. W. Seabloom, J. B. Shurin, and J. E. Smith. 2007. Consumer
 versus resource control of producer diversity depends on ecosystem type and producer

- community structure. Proceedings of the National Academy of Sciences of the United States of America **104**:10904-10909.
- Hobbie, S. E. 2008. Nitrogen effects on litter decomposition: a five-year experiment in eight temperate grassland and forest sites. Ecology **89**:2633-2644.
- Hobbie, S. E. 2015. Plant species effects on nutrient cycling: revisiting litter feedbacks. Trends in Ecology & Evolution **30**:357-363.
- Hoekstra, J. M., T. M. Boucher, T. H. Ricketts, and C. Roberts. 2005. Confronting a biome crisis: global disparities of habitat loss and protection. Ecology Letters 8:23-29.
- 571 Hughes, B. B., R. Beas-Luna, A. K. Barner, K. Brewitt, D. R. Brumbaugh, E. B. Cerny-572 Chipman, S. L. Close, K. E. Coblentz, K. L. De Nesnera, S. T. Drobnitch, J. D. Figurski, B. Focht, M. Friedman, J. Freiwald, K. K. Heady, W. N. Heady, A. Hettinger, A. 573 574 Johnson, K. A. Karr, B. Mahoney, M. M. Moritsch, A. M. K. Osterback, J. Reimer, J. 575 Robinson, T. Rohrer, J. M. Rose, M. Sabal, L. M. Segui, C. C. Shen, J. Sullivan, R. 576 Zuercher, P. T. Raimondi, B. A. Menge, K. Grorud-Colvert, M. Novak, and M. H. Carr. 577 2017. Long-Term Studies Contribute Disproportionately to Ecology and Policy. 578 Bioscience 67:271-281.
 - Isbell, F., P. B. Reich, D. Tilman, S. E. Hobbie, S. Polasky, and S. Binder. 2013a. Nutrient enrichment, biodiversity loss, and consequent declines in ecosystem productivity. Proceedings of the National Academy of Sciences of the United States of America 110:11911-11916.

580

581

582

583

584

585

586

587

588

589

590

591

592

- Isbell, F., D. Tilman, S. Polasky, S. Binder, and P. Hawthorne. 2013b. Low biodiversity state persists two decades after cessation of nutrient enrichment. Ecology Letters **16**:454-460.
- Jenkinson, D. S., J. M. Potts, J. N. Perry, V. Barnett, K. Coleman, and A. E. Johnston. 1994.

 Trends in herbage yields over the last century on the Rothamsted Long-term Continuous Hay Experiment. Journal of Agricultural Science 122:365-374.
- Knorr, M., S. D. Frey, and P. S. Curtis. 2005. Nitrogen additions and litter decomposition: A meta-analysis. Ecology **86**:3252-3257.
- Lawes, J. B., and J. H. Gilbert. 1880. Agricultural, botanical, and chemical results of experiments on the mixed herbage of permanent meadow, conducted for more than twenty years in succession on the same lands. Part 1. Philosophical Transactions of the Royal Society of London, Series B **171**:289-416.
- 594 Lewandowska, A. M., A. Biermann, E. T. Borer, M. A. Cebrian-Piqueras, S. A. J. Declerck, L. 595 De Meester, E. Van Donk, L. Gamfeldt, D. S. Gruner, N. Hagenah, W. S. Harpole, K. P. 596 Kirkman, C. A. Klausmeier, M. Klever, J. M. H. Knops, P. Lemmens, E. M. Lind, E. 597 Litchman, J. Mantilla-Contreras, K. Martens, S. Meier, V. Minden, J. L. Moore, H. O. 598 Venterink, E. W. Seabloom, U. Sommer, M. Striebel, A. Trenkamp, J. Trinogga, J. 599 Urabe, W. Vyverman, D. B. Van de Waal, C. E. Widdicombe, and H. Hillebrand. 2016. 600 The influence of balanced and imbalanced resource supply on biodiversity-functioning 601 relationship across ecosystems. Philosophical Transactions of the Royal Society B-602 Biological Sciences 371.
- Midolo, G., R. Alkemade, A. M. Schipper, A. Benitez-Lopez, M. P. Perring, and W. De Vries. 2019. Impacts of nitrogen addition on plant species richness and abundance: A global meta-analysis. Global Ecology and Biogeography **28**:398-413.
- Miller, T. E., J. H. Burns, P. Munguia, E. L. Walters, J. M. Kneitel, P. M. Richards, N. Mouquet,
 and H. L. Buckley. 2005. A critical review of twenty years' use of the resource-ratio
 theory. American Naturalist 165:439-448.

- 609 Pierik, M., J. van Ruijven, T. M. Bezemer, R. Geerts, and F. Berendse. 2011. Recovery of plant 610 species richness during long-term fertilization of a species-rich grassland. Ecology 611 **92**:1393-1398.
- Ramankutty, N., A. T. Evan, C. Monfreda, and J. A. Foley. 2008. Farming the planet: 1. Geographic distribution of global agricultural lands in the year 2000. Global Biogeochemical Cycles **22**:1-19.
- Reich, P. B., J. Knops, D. Tilman, J. Craine, D. Ellsworth, M. Tjoelker, T. Lee, D. Wedin, S.
 Naeem, D. Bahauddin, G. Hendrey, S. Jose, K. Wrage, J. Goth, and W. Bengston. 2001.
 Plant diversity enhances ecosystem responses to elevated CO2 and nitrogen deposition.
 Nature 410:809-812.
- Reich, P. B., D. Tilman, F. Isbell, K. Mueller, S. E. Hobbie, D. F. B. Flynn, and N. Eisenhauer.

 2012. Impacts of Biodiversity Loss Escalate Through Time as Redundancy Fades.

 Science **336**:589-592.
- Riggs, C. E., S. E. Hobbie, E. M. Bach, K. S. Hofmockel, and C. E. Kazanski. 2015. Nitrogen addition changes grassland soil organic matter decomposition. Biogeochemistry **125**:203-219.
- Seabloom, E. W. 2010. Spatial and Temporal Variability in Propagule limitation of California Native Grasses. Oikos **120**:291–301.
- 626 627 Seabloom, E. W., E. T. Borer, Y. Buckley, E. E. Cleland, K. Davies, J. Firn, W. S. Harpole, Y. Hautier, E. Lind, A. Macdougall, J. L. Orrock, S. M. Prober, P. Adler, J. Alberti, T. M. 628 629 Anderson, J. D. Bakker, L. A. Biederman, D. Blumenthal, C. S. Brown, L. A. Brudvig, 630 M. Caldeira, C. J. Chu, M. J. Crawley, P. Daleo, E. I. Damschen, C. M. D'Antonio, N. M. 631 Decrappeo, C. R. Dickman, G. Z. Du, P. A. Fay, P. Frater, D. S. Gruner, N. Hagenah, A. 632 Hector, A. Helm, H. Hillebrand, K. S. Hofmockel, H. C. Humphries, O. Iribarne, V. L. 633 Jin, A. Kay, K. P. Kirkman, J. A. Klein, J. M. H. Knops, K. J. La Pierre, L. M. Ladwig, J. 634 G. Lambrinos, A. D. B. Leakey, Q. Li, W. Li, R. McCulley, B. Melbourne, C. E. 635 Mitchell, J. L. Moore, J. Morgan, B. Mortensen, L. R. O'Halloran, M. Partel, J. Pascual, 636 D. A. Pyke, A. C. Risch, R. Salguero-Gomez, M. Sankaran, M. Schuetz, A. Simonsen, 637 M. Smith, C. Stevens, L. Sullivan, G. M. Wardle, E. M. Wolkovich, P. D. Wragg, J.
 - Wright, and L. Yang. 2013. Predicting invasion in grassland ecosystems: is exotic dominance the real embarrassment of richness? Global Change Biology **19**:3677-3687.
 - Seabloom, E. W., E. T. Borer, Y. M. Buckley, E. E. Cleland, K. F. Davies, J. Firn, W. S. Harpole, Y. Hautier, E. M. Lind, A. S. MacDougall, J. L. Orrock, S. M. Prober, P. B.
- Harpole, Y. Hautier, E. M. Lind, A. S. MacDougall, J. L. Orrock, S. M. Prober, P. B.
 Adler, T. M. Anderson, J. D. Bakker, L. A. Biederman, D. M. Blumenthal, C. S. Brown,
 L. A. Brudvig, M. Cadotte, C. J. Chu, K. L. Cottingham, M. J. Crawley, E. I. Damschen,
- C. M. Dantonio, N. M. DeCrappeo, G. Z. Du, P. A. Fay, P. Frater, D. S. Gruner, N.

639

- Hagenah, A. Hector, H. Hillebrand, K. S. Hofmockel, H. C. Humphries, V. L. Jin, A.
- Kay, K. P. Kirkman, J. A. Klein, J. M. H. Knops, K. J. La Pierre, L. Ladwig, J. G.
- Lambrinos, Q. Li, W. Li, R. Marushia, R. L. McCulley, B. A. Melbourne, C. E. Mitchell,
- J. L. Moore, J. Morgan, B. Mortensen, L. R. O'Halloran, D. A. Pyke, A. C. Risch, M.
- Sankaran, M. Schuetz, A. Simonsen, M. D. Smith, C. J. Stevens, L. Sullivan, E.
- Wolkovich, P. D. Wragg, J. Wright, and L. Yang. 2015. Plant species' origin predicts
- dominance and response to nutrient enrichment and herbivores in global grasslands.

 Nature Communications **6**:8.
- Seastedt, T. R., J. M. Briggs, and D. J. Gibson. 1991. CONTROLS OF NITROGEN
 LIMITATION IN TALLGRASS PRAIRIE. Oecologia 87:72-79.

- 655 Silvertown, J., J. Tallowin, C. Stevens, S. A. Power, V. Morgan, B. Emmett, A. Hester, P. J. 656 Grime, M. Morecroft, R. Buxton, P. Poulton, R. Jinks, and R. Bardgett. 2010.
- Environmental myopia: a diagnosis and a remedy. Trends in Ecology & Evolution **25**:556-561.
- Simkin, S. M., E. B. Allen, W. D. Bowman, C. M. Clark, J. Belnap, M. L. Brooks, B. S. Cade, S.
 L. Collins, L. H. Geiser, F. S. Gilliam, S. E. Jovan, L. H. Pardo, B. K. Schulz, C. J.
 Stevens, K. N. Suding, H. L. Throop, and D. M. Waller. 2016. Conditional vulnerability
 of plant diversity to atmospheric nitrogen deposition across the United States.
- Proceedings of the National Academy of Sciences of the United States of America 113:4086-4091.
- Smith, B., and J. B. Wilson. 1996. A consumer's guide to evenness indices. Oikos **76**:70-82.
- 666 Smith, M. D., A. K. Knapp, and S. L. Collins. 2009. A framework for assessing ecosystem
 667 dynamics in response to chronic resource alterations induced by global change. Ecology
 668 **90**:3279-3289.
- Smith, M. D., K. J. La Pierre, S. L. Collins, A. K. Knapp, K. L. Gross, J. E. Barrett, S. D. Frey,
 L. Gough, R. J. Miller, J. T. Morris, L. E. Rustad, and J. Yarie. 2015. Global
 environmental change and the nature of aboveground net primary productivity responses:
 insights from long-term experiments. Oecologia 177:935-947.
- Steffen, W., K. Richardson, J. Rockström, S. E. Cornell, I. Fetzer, E. M. Bennett, R. Biggs, S. R.
 Carpenter, W. de Vries, C. A. de Wit, C. Folke, D. Gerten, J. Heinke, G. M. Mace, L. M.
 Persson, V. Ramanathan, B. Reyers, and S. Sörlin. 2015. Planetary boundaries: Guiding human development on a changing planet. Science.
- Stevens, C. J., E. M. Lind, Y. Hautier, W. S. Harpole, E. T. Borer, S. Hobbie, E. W. Seabloom,
 L. Ladwig, J. D. Bakker, C. J. Chu, S. Collins, K. F. Davies, J. Firn, H. Hillebrand, K. J.
 La Pierre, A. MacDougall, B. Melbourne, R. L. McCulley, J. Morgan, J. L. Orrock, S. M.
 Prober, A. C. Risch, M. Schuetz, and P. D. Wragg. 2015. Anthropogenic nitrogen
 deposition predicts local grassland primary production worldwide. Ecology 96:14591465.
 - Tilman, D., F. Isbell, and J. M. Cowles. 2014. Biodiversity and Ecosystem Functioning. Pages 471-493 *in* D. J. Futuyma, editor. Annual Review of Ecology, Evolution, and Systematics, Vol 45. Annual Reviews, Palo Alto.
 - van der Valk, A. G. 1986. The impact of litter and annual plants on recruitment from the seed bank of a lacustrine wetland. Aquatic Botany **24**:13-26.
- Vitousek, P. M., J. D. Aber, R. W. Howarth, G. E. Likens, P. A. Matson, D. W. Schindler, W. H.
 Schlesinger, and D. G. Tilman. 1997a. Human alteration of the global nitrogen cycle:
 Sources and consequences. Ecological Applications 7:737-750.
- Vitousek, P. M., H. A. Mooney, J. Lubchenco, and J. M. Melillo. 1997b. Human domination of Earth's ecosystems. Science **277**:494-499.
- Ward, D., K. Kirkman, and Z. Tsvuura. 2017. An African grassland responds similarly to longterm fertilization to the Park Grass experiment. Plos One 12.

Data accessibility statement:

683

684

685

686

687

Data supporting the results will be archived at the Environmental Data Initiative

(https://environmentaldatainitiative.org/). The data DOI will be included at the end of the article

upon acceptance.

_	\sim	\sim	
1	11	11	
- /	١,	,	

FIGURE LEGENDS

701	Figure 1. Effect of nutrient enrichment on live and dead aboveground plant biomass in grassland
702	ecosystems. Colored lines indicate individual sites, and the solid black line shows the mean
703	response across sites. Models were fit using log10 (Treatment/Control) versus log10 (Number of
704	Years of Treatment), and back-transformed for plotting as the difference between Treatment (i.e.
705	all nutrients added) and Control plots. Standard errors of all parameter estimates are presented in
706	Appendix S1: Table S2, and site species models with raw data are presented in Figures S1 and
707	S2.
708	Figure 2. Effects of nutrient enrichment on diversity (<i>ENS_{PIE}</i>), richness (<i>S</i> , species m ⁻²), and
709	evenness (ENS _{PIE} S ⁻¹) in grassland ecosystems. Colored lines indicate individual sites, and the
710	solid black line shows the mean response across sites. Models were fit using
711	log ₁₀ (Treatment/Control) versus log ₁₀ (Number of Years of Treatment), and back-transformed for
712	plotting as the difference between Treatment (i.e., all nutrients added) and Control plots.
713	Standard errors of all parameter estimates are presented in Appendix S1: Table S2, and site
714	species models with raw data are presented in Figures S3, S4, and S5.
715	Figure 3. Effects of nutrient addition treatments on live and dead aboveground plant biomass
716	and diversity (ENS_{PIE}), richness (S , species m ⁻²), and evenness ($ENS_{PIE}S^{-1}$) in grassland
717	ecosystems. Error bars indicate 1 standard error (SE) among site means for each treatment.
718	Parameter estimates and standard errors for all treatment effects are shown in Appendix S1:
719	Table S1.

Figure 4. Change over time of the effects of nutrient enrichment on live biomass, dead biomass, and diversity at individual sites (slopes in Figure 1). Values of zero (dashed lined) indicate effects that are constant over time, positive values indicate increasing effects through time, and negative values indicate decreasing effects through time. All slopes are from log₁₀ transformed variables versus log₁₀ transformed number of years of treatment. Diversity is measured as *ENS*_{PIE}. Correlations and significant tests are based on standardized major axis (SMA) estimation and are detailed in Appendix S1: Table S3. Large open circles represent the mean value across all sites with error bars representing two standard errors of the mean (SEM).

Figure 1.

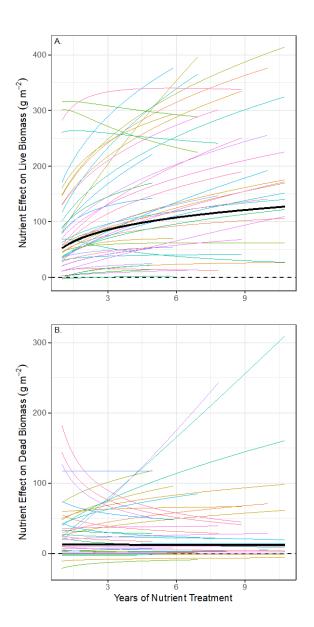


Figure 2.

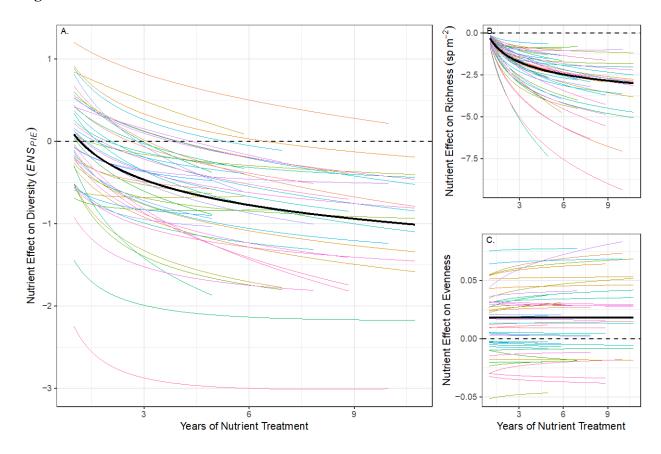


Figure 3.

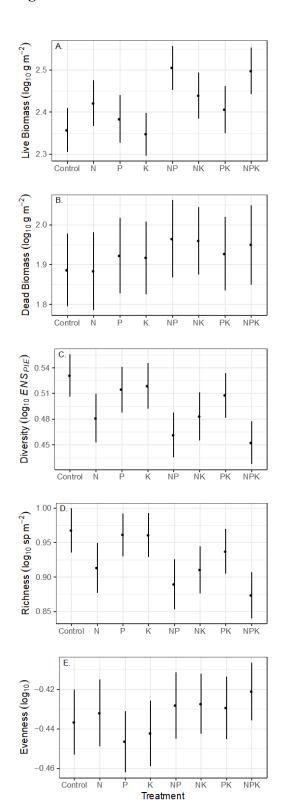


Figure 4.

