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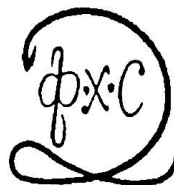
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*14th International Conference on
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INFLUENCE OF THE LOW FREQUENCY RANGES OF MAGNETIC FIELD ON *SACCHAROMYCES CEREVISIAE* RESPIRATION

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ABSTRACT

In this paper influence of the low frequency magnetic field on *Saccharomyces cerevisiae* respiration was examined. Influence of four low frequency magnetic field ranges was examined: 10-200 Hz, 200-300 Hz, 300-650 Hz and 650-1000 Hz. All analyzed frequency ranges gave the same influence on the yeast cells respiration.

INTRODUCTION

Over the years, due to the fast technological development, our environment has become more and more influenced by man-made low frequency electromagnetic fields (EMFs). Therefore it is not surprising that this increasing exposure of the population in everyday life to EMFs has earned such a wide public interest. Recently, effects of electric fields (EFs), magnetic fields (MFs) or EMFs on different microbes have become a very popular topic since the mentioned physical fields could potentially act as a stress factors and thus affect the survival of the microbial cells as well as their metabolism and behavior [1]. The influence of static (0 Hz) and 50 Hz magnetic fields on growth of the *Saccharomyces cerevisiae* (by the measurements of solution optical density at 600 nm) was already examined by many authors [2,3]. In this study the influence of different low frequency ranges of the magnetic fields (up to 1 kHz) were examined on the yeast cells respiration. To the best of our knowledge, this is the first time that the influence of magnetic fields is examined on yeast cells by using Micro-Oxymax® respirometer and constant frequency scan regime in the given ranges.

EXPERIMENTAL

All experiments were performed, in two specially designed glass bottles. Both bottles contained 3 mL of yeast suspension. One bottle was a control sample while the other one was set up inside of the spiral Cu-coil where the low frequency magnetic fields were generated. In order to avoid undesirable temperature increase of the yeast sample because of the heating effect of the coil, sample bottle was set up in a glass water recirculation jacket and all together wrapped in Cu-coil. Control vessel was connected in line with working vessel to minimize possible temperature differences between samples. Arbitrary function generator (Gw Instek AFG-2105, Good Will Instrument Co., Ltd, Taiwan) was used to set up a desired frequency range and a scan interval during which frequencies from set up range are continuously changed from lowest to highest. In this paper we examined influence of four different low frequency ranges: 100-200 Hz, 200-300 Hz, 300-650 Hz and 650-1000 Hz. In all experiments the scan interval was 100 s and total time of exposure was 24 h. In order to achieve the maximal effective current of 2,00 A trough the coil a signal amplifier was used. Maximal effective current of 2,00 A corresponds to magnetic induction of 33 mT. Before every experiment, oscilloscope was used to adjust maximal current. Respiration activity of *Saccharomyces cerevisiae* was measured by a twelve-channel Micro-Oxymax® respirometer (Columbus Instruments, USA). Cell respiration was measured every 20 min during 24 h. Cumulative O₂ consumption and cumulative CO₂ production (in mL) were determined. Experimental setup is shown in Figure 1.

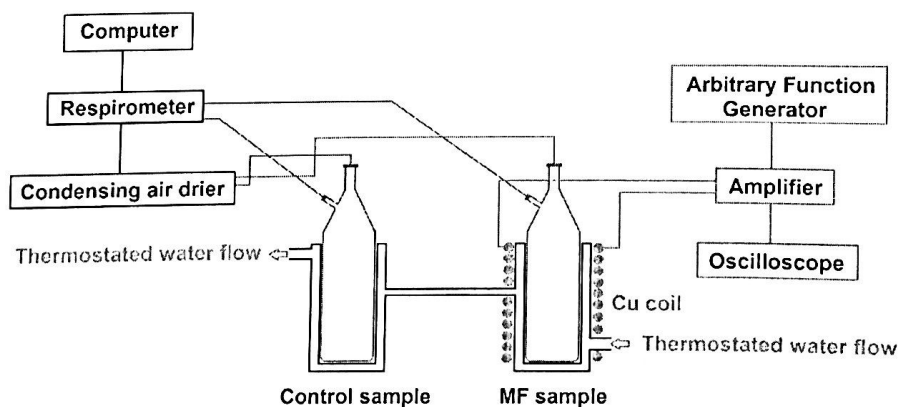


Figure 1. Experimental setup.

RESULTS AND DISCUSSION

Like it was mentioned in Experimental part of this work influence of four low frequency ranges (100-200 Hz, 200-300 Hz, 300-650 Hz and 650-1000 Hz) on yeast respiration were examined.

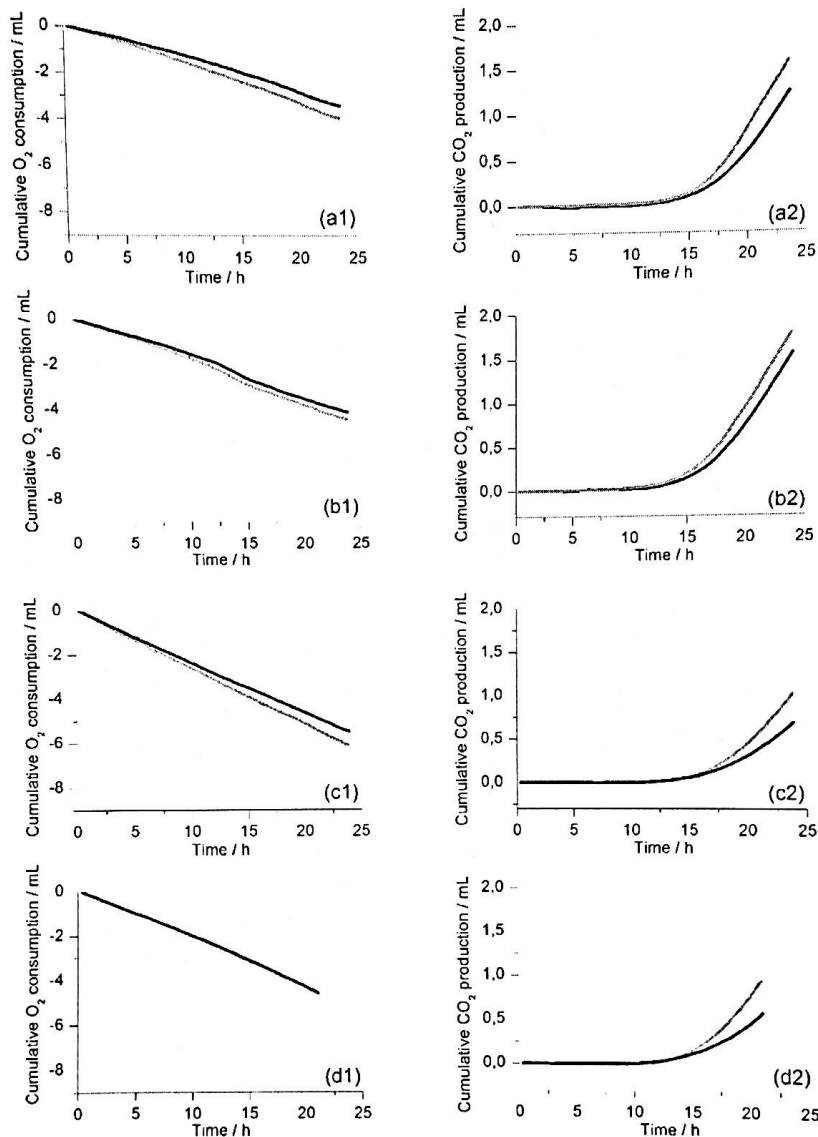


Figure 2. Experimentally obtained cumulative O₂ consumption in mL (a1, b1, c1, d1) and cumulative CO₂ production in mL (a2, b2, c2, d2) over 24 h of exposure, for different frequency ranges (100-200 Hz (a1, a2), 200-300 Hz (b1, b2), 300-650 Hz (c1, c2) and 650-1000 Hz (d1, d2)). Gray curve represents changes in control sample and black curve changes of the O₂ and CO₂ in magnetic field sample.

Cumulative O₂ consumption and cumulative CO₂ production were followed in control and magnetic field samples during 24h. Figure 2. shows cumulative O₂ consumption and cumulative CO₂ production in control and magnetic field samples over 24 h obtained for analyzed frequency ranges. Cumulative O₂ consumption is negative because oxygen at the beginning of the experiment (of 21% in air) is taken as reference (0 mL). As it can be seen from Figure 2, up to 15th hour the differences in cumulative CO₂ production between control and magnetic field sample at all investigated frequency ranges are negligible. After 15th hour those differences begin to grow and are the biggest at the end of exposure. Cumulative CO₂ production in magnetic field sample was lower in comparison to control sample for all frequency ranges. Cumulative O₂ consumption, for frequency ranges from 100-200 Hz, 200-300 Hz, 300-650 Hz, in magnetic field sample was lower in comparison to control sample but for 650-1000 Hz range difference is negligible.

CONCLUSION

All experiments showed lower cumulative O₂ consumption and lower cumulative CO₂ production in magnetic field which confirms magnetic effects. Considering mentioned lower consumption/production of gases, this influence is probably due to deceleration of yeast metabolism or cell division, which should be further investigated and confirmed by statistical tests. It is challenging task to design magnetic treatment to produce desired metabolic changes without using chemical reagents. Obtained results are preliminary and further investigations are necessary to verify proposed hypothesis.

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