

Spatial distribution, habitat preference and colonization status of two alien terrestrial invertebrate species in Antarctica

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Abstract: The introduction of invasive species is one of the greatest threats to Earth's biodiversity, as they can reduce native biodiversity and alter ecosystem structure and function. Currently, the only two known non-native terrestrial invertebrates in Antarctica are the chironomid midge *Eretmoptera murphyi* and the enchytraeid worm *Christensenidrilus blocki*. These invertebrates were probably introduced to ground near Signy Research Station, South Orkney Islands, during transplantation experiments in the late 1960s. Between 2007 and 2009, this study surveyed the area around the introduction site for midge larvae and worms to assess any change over the last four decades in their spatial distribution, habitat preference and colonization status. *Eretmoptera murphyi* was found in concentrations up to 4.1×10^5 larvae m^{-2} (mean 2.1×10^4 larvae m^{-2}) at distances of up to 220 m from the probable introduction site ($c. 35\,000\,m^2$), while *C. blocki* was only found close to the introduction site in low numbers. Significantly more *E. murphyi* larvae were found in peat and dead organic material ($3.34 \times 10^4\,m^{-2}$) than in stony soil and gravel ($1.52 \times 10^4\,m^{-2}$) or living moss and other plant material ($1.16 \times 10^4\,m^{-2}$). *Eretmoptera murphyi* can no longer be considered a persistent alien as it clearly expanding its distribution, while *C. blocki* remains a persistent alien species.

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Introduction

Invasions by non-native species are one of the greatest threats to global biodiversity (McKinney & Lockwood 1999). Invasive species can alter existing species assemblages and ecosystem function (Mack *et al.* 2000) to the degree that entire landscapes can be transformed (Scott & Kirkpatrick 2008). The majority of terrestrial ecosystems on Earth have already been impacted by invasive species, including many of the sub-Antarctic islands but, although at risk, to date Antarctica has remained so far largely protected from impact by invasive species (Frenot *et al.* 2005, 2008, Chown *et al.* 2008, Convey 2008).

The Antarctic terrestrial biota includes both relict species that have been present in Antarctic refugia on multimillion year timescales throughout glacial cycles, some since the breakup of Gondwana (Convey *et al.* 2008), and species that have colonized since then (Barnes *et al.* 2006). Potential colonizing species have to overcome two major barriers, termed the transport barrier and the establishment barrier. The colonization of Antarctica by non-native species has been considered to be an unlikely event as Antarctica's extreme isolation and the circumpolar oceanic and atmospheric currents create a high barrier to natural transport and the extreme climatic conditions (low temperatures and water availability) and paucity of suitable habitat make establishment difficult. However, increased human activity in Antarctic, in particular tourism

and national governmental operator activities, has increased the opportunity for non-native species to be transported to Antarctica (Frenot *et al.* 2005, Whinam *et al.* 2005, COMNAP 2009, IAATO 2009, Hughes *et al.* 2009, Lee & Chown 2009). In addition, regional climate change may increase temperatures and water availability in some areas, making establishment more likely.

Since the arrival of humans on the sub-Antarctic islands for commercial activities in the late eighteenth and early nineteenth centuries, the number of non-native species has steadily increased, with *c.* 200 vertebrates, plants and invertebrates currently being known to be established (Frenot *et al.* 2005). This has resulted in major changes to the terrestrial environment (see reviews by Frenot *et al.* 2005, 2008). In contrast, only five non-native species are confirmed to have become established in the Antarctic terrestrial environment, including three grass and two invertebrate species. The two invertebrates, *Eretmoptera murphyi* Schaeffer and *Christensenidrilus blocki* Dozsa-Farkas & Convey, are found close to the United Kingdom's Signy Research Station (South Orkney Islands; 60°43'S 45°38'W), and may result from the importation of non-sterile soil and plants during transplant experiments from South Georgia (Edwards 1980, Block *et al.* 1984, Convey & Block 1996).

Introduction of non-Antarctic soil to Signy in the 1960s

Non-Antarctic soil was first intentionally introduced to Signy Island in 1963/64 when pieces of *Polytrichum*

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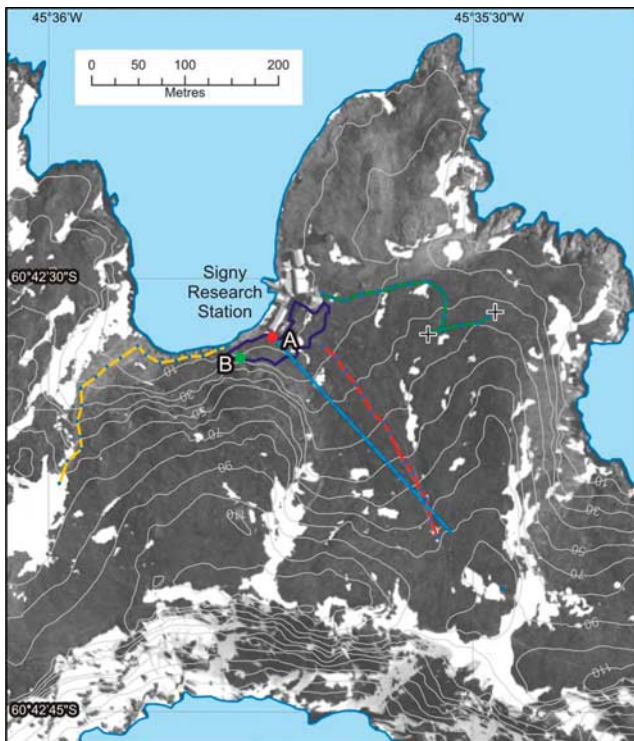


Fig. 1. Map of the Signy Research Station area showing the main introduction site (A), the vermiculite site (B). Blue line = back slope transect, yellow line = Stone Chute path, red line = Back Slope path, and green line = Study Site path (+ indicates position of cloches).

strictum Menzies ex Bridel turf were transplanted from South Georgia (R.I.L. Smith, personal communication 2008). In 1965, transplantation experiments continued using plants from the Falkland Islands and the Cairngorm Mountains, Scotland, all of which died in the first year and were incinerated in 1967 (Smith 1996).

In 1967 further transplantation experiments were performed, which probably resulted in the introduction of the two non-native invertebrate species to Signy from South Georgia (Edwards 1980). The main introduction site was on steeply sloping ground ~25 m inland from Factory Cove, close to Signy Research Station, in an eroded *Polytrichum alpestre* Hoppe moss bank which was dug over and substituted with additional soil from sites in the local area (labelled A on Fig. 1). This site faced NNE on a 20° slope around 13 m above sea level (Edwards & Greene 1973). A second experimental site was set up ~40 m WSW of the main site, where plants were grown in vermiculite in pots supported by stones on an unstable NNW-facing slope 10 m above sea level and c. 25 m from the coast (Edwards 1968, Edwards & Greene 1973) (labelled B on Fig. 1).

Vascular plants were collected from King Edward Point, Bore Valley and Husvik on South Georgia in early December 1967 and planted on Signy Island on 23 and 24 December 1967. A second collection of plants was made



Fig. 2. a. Photograph of the *Eretmoptera murphyi* adult. The adult midge is approximately 4.5 mm long. b. Photograph of *Eretmoptera murphyi* larvae. Each larva is approximately 3.5 mm long.

from East Cumberland Bay on 18 November 1968 and planted on Signy between 2–4 December 1968 (Edwards 1980). All South Georgian plant material was removed and destroyed in February 1973 at the end of the research project. Translocated plant species included *Acaena magellanica* (Lam.) Vahl., *A. tenera* Alboff., *Achillea millefolium* L., *Blechnum penna-marma* (Poir.) Kuhn., *Callitriche antarctica* Engelm., *Colobanthus subulatus* (D'Urv.) Hook. f., *C. quitensis* (Kunth) Bartl., *Cystopteris fragilis* (L.) Bernh., *Deschampsia antarctica* Desv., *Festuca contracta* T. Kirk, *Hymenophyllum falklandicum* Baker, *Juncus scheuchzerioides* Gaud., *Montia fontana* L., *Paradiochloa (Poa) flabellata* (Lam.) Hook. f., *Phleum alpinum* L., *Poa annua* L., *Poa pratensis* L., *Polystichum mohrioides* (Bory) Pr., *Ranunculus biternatus* Smith, *R. literatus* Smith, *R. repens* L., *Rostkovia magellanica* (Lam.) Hook. f., *Trifolium repens* L. and *Uncinia meridensis* Steyermark. Of these, *P. annua*, *P. pratensis*, *A. millefolium*, *T. repens* and *R. repens* were originally non-native to South Georgia (Edwards 1980, Smith 1996).

Burn (1982) first reported the discovery of a non-native enchytraeid worm and a flightless chironomid midge on Signy Island at the main experimental site (labelled 'A' in Fig. 1) on 27 September 1980, almost 13 years after the original probable introduction (Block *et al.* 1984). The two non-native species were later identified as the chironomid midge *Eretmoptera murphyi* (Fig. 2a & b) and the enchytraeid worm *Christensenidrilus blocki*. (Cranston 1985, Dózsa-Farkas & Convey 1997, 1998). Although the transplant experiments were the most probable source of the introductions, arrival via this route does not explain either the apparent 13 year delay in the discovery of the two non-native species in an area frequently visited by terrestrial biologists, nor why *E. murphyi* has yet to be found at the South Georgia plant collection sites. A plausible alternative explanation is that the species introduction was associated with people and cargo that was routinely moved between South Georgia and Signy Research Station over many years.

Eretmoptera murphyi

Eretmoptera murphyi is a terrestrial brachypterous chironomid midge originally endemic to South Georgia, from where it has been reported on just a few occasions (Brundin 1970, Ring *et al.* 1990, Allegrucci *et al.* 2006). Prior to the introduction of *E. murphyi*, no native dipterans had been recorded from the South Orkney Islands (Convey 1992).

Larvae range from *c.* 1.5–5.0 mm in body length, pupae, *c.* 2.5–5.0 mm, and adults, *c.* 3.4–4.9 mm (Block *et al.* 1984, Cranston 1985), while live weights of freshly collected specimens range from 0.46–4.76 mg (larvae), 1.01–2.04 (pupae) and 0.18–1.16 mg (adults) (Block *et al.* 1984). On Signy, adult midges are generally observed emerging from pupal cases, with some degree of synchronicity, during calm sunny weather in early January. The adults crawl over the surface and are able to hold firm against wind currents (Block *et al.* 1984). Individuals produce single spherical egg batches containing *c.* 85 eggs, which are positioned beneath the substratum surface (Convey 1992, Convey & Block 1996). Larvae hatch in late February about 30–45 days after oviposition and spend the winter in the surface layers of the soil and vegetation. Convey (1992) suggested that *E. murphyi* at Signy Island may follow a two year life-cycle, including two winters. This is supported by the large variation in larval fresh weight (0.46–4.76 mg) which may represent cohorts hatched during different years (Convey & Block 1996). At Signy Island, the larvae may require a second year (or possibly more) to complete development, due to the shorter period during which habitat temperatures are suitable for activities including feeding (low positive temperatures). Typically, annual mean air temperatures are *c.* -3°C (range +11 to -30°C; data for 2006), while summer mean air temperatures are *c.* +1.0 (range +11 to -3°C; data for December 2006 to February 2007). Snow cover on Signy

Island is unpredictable and in years with little snow cover the winter soil temperatures can fall to below -10°C (Bokhorst *et al.* 2008). It is possible that *E. murphyi* on South Georgia are able to complete their life cycle in one year as a result of the less harsh climatic conditions compared to Signy Island.

Christensenidrilus blocki

The distribution, taxonomy, physiology and ecology of terrestrial annelid worms in Antarctica are poorly known (Block & Christensen 1985). The Acanthodrilidae and Lumbricidae extend only as far as the sub-Antarctic islands, but the Enchytraeidae penetrate further south to include the maritime Antarctic in their distribution (Dózsa-Farkas & Convey 1997, Frenot *et al.* 2005). Block & Christensen (1985) list 23 Enchytraeidae from the Antarctic and sub-Antarctic, including records of five new species from South Georgia that are common in northern Europe and which may have been introduced through human activities. Marine Antarctic enchytraeid species obtained from littoral collections at Signy Island include *Lumbricillus lineatus* Müller and *L. maximus* Michaelsen though no native terrestrial enchytraeids have been identified (Stephenson 1932). The enchytraeid introduced to Signy Island was identified as a new genus and named *Christensenidrilus blocki* (Block & Christensen 1985, Dózsa-Farkas & Convey 1997, 1998). Specimens, previously identified as *Marionina georgiana* Michaelsen and reclassified as *Christensenidrilus blocki* based on identical morphologies, were previously found on South Georgia within moss at East Cumberland Bay (Stephenson 1932). This is one of the locations from where plants were collected in 1968 for transplantation experiments on Signy (Edwards 1980).

Research and monitoring is essential if introduced species are to be managed and, if possible, eradicated. Since the introduction of *E. murphyi* and *C. blocki* to Signy Island, no systematic survey of their distribution has been undertaken. This study aims to assess the current spatial distribution, substratum preference and colonization status of terrestrial Antarctica's only two known non-native invertebrate species.

Methods

Quantification of non-native species in different substrata around the main introduction site

In January 2007, soil samples (four replicates) were taken at random (but within 50 cm) of points on a 5 × 5 m grid, centred on the main introduction site (labelled 'A' on Fig. 1), using a 25 mm diameter corer to a depth of about 2.5 cm. In addition, nine cores were collected from the main introduction site, which was *c.* 3 m by 3 m (using a 1.5 m matrix). Samples were immediately placed in sterile plastic bags and stored at +4°C until returned for analysis in the UK.

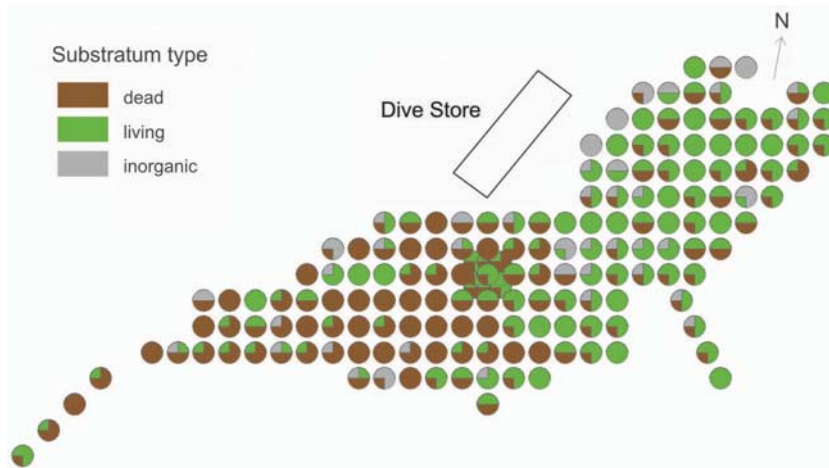


Fig. 3. Distribution of different substrates around the main introduction site. Each circle represents the proportion of each substratum-type found in the four replicates from each sample site. The samples locations are on a $5\text{ m} \times 5\text{ m}$ grid, with samples sites around the likely introduction site on a $1.5\text{ m} \times 1.5\text{ m}$ grid.

Quantification of non-native species on a transect away from the main introduction site

In January 2008 sampling points were identified at 20 m intervals for 280 m along a direct line between the introduction site and the 93 m spot height point on Observation Bluff (bearing of 135° ; see Fig. 1, blue line). Triplicate soil cores were collected within a 5 m radius of each sample point. Samples were not collected randomly, but cores were collected from dead moss or peat where available. Individual samples were placed in sample bags and returned to the UK at 4°C for analysis.

Quantification of non-native species along paths

In February 2009 a series of cores were collected along the three main paths leading away from the research station (see Fig. 1). One path followed the coast for about 100 m and then climbed steeply to about 75 m above sea level ('Stone Chute'; yellow line). This is the main path from the research station providing access to the main body of the island, and is used frequently by station staff. A second path ('Back Slope'; red line) from the station climbed steadily to a study site about

60 m a.s.l. This path is used regularly, but less than the Stone Chute path. The third path (Study Site; green line) leads to a small study site (*c.* 40 m a.s.l.) established in 1991 and has had much less use. For the Stone Chute and Back Slope paths, three sets of three samples were collected at *c.* 20 m intervals along the paths. One set was taken from the centre of the path, one each from about 10 m to either side. The samples were collected and analysed in a similar manner to those collected previously. For the Study Site path, triplicate samples were taken only from the path itself, and no on-path/off-path comparison of larvae concentrations was made.

Enumeration of non-native species in soil cores

Cores were removed from their sample bags, weighed and placed on trays for examination. Cores containing moss were teased apart using a pair of fine forceps while more friable material (peat and soil) was spread thinly over a white tray. Any midge larvae and enchytraeid worms were carefully removed and counted during a 15 min examination period using a low power stereo microscope. Cores were normally examined within four months of collection. Dead or

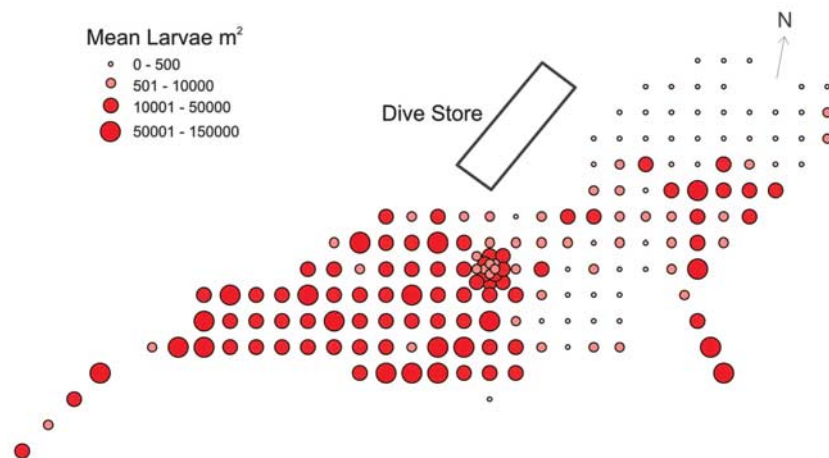


Fig. 4. Distribution of *E. murphyi* larvae around the main introduction site. Circles represent the mean number of *E. murphyi* larvae m^{-2} at each sample site.

decomposing midge larvae and worms were rarely found, suggesting that transport back to the UK had little impact on invertebrate survival; nevertheless a small number of dead organisms may have decayed beyond recognition. A note was made of the sample mass, location and predominant substratum type (three categories: i) dead organic material (dead moss and peat), ii) living organic material (mostly moss), and iii) inorganic material (stony soil and gravel)). The concentrations of larvae in each substratum were compared by one-way analysis of variance with Tukey's test using the MINITAB 15.1 package.

Wind speed and direction

Wind speed and direction was recorded near Signy Research Station using a series of anemometers between 1957 and 1995 to show the influence of wind on the distribution of the midge.

Estimation of larval mean fresh and dry mass

Records were made of the individual masses of 25 randomly selected fresh larvae, after which dry mass was measured by drying the larvae to constant mass at 45°C.

Results

Quantification of non-native species in different substrata around the main introduction site

Figure 3 shows the substrata sampled in the 5 m × 5 m grid around the main introduction site. Generally, dead moss and peat was the most common substrate in cores taken to the west while live moss was most common to the east.

Figure 4 shows the mean number of *E. murphyi* at each sample position. Larvae numbers were generally greater to the west of the main introduction site, although high larval densities were located at some other sites. The maximum number of larvae found in any single core within the grid was 150 which equates to over 3×10^5 larvae m⁻². The mean concentration of larvae found in the surveyed area around the introduction site was 2.11×10^4 larvae m⁻² (716 cores). However, on closer analysis, mean larval concentration varied to some degree depending upon the substratum type: i) dead organic material, 3.34×10^4 (SD ± 4.2×10^4) (305 cores), ii) living organic material, 1.16×10^4 (SD ± 4.1×10^4) (344 cores), and iii) inorganic material, 1.52×10^4 (SD ± 2.5×10^4) (67 cores). Larval concentrations in dead organic material were significantly different to those in living organic material and inorganic material ($P < 0.05$). Larvae concentrations in living organic material and inorganic material were not significantly different ($P > 0.05$).

Christensenidrilus blocki was absent from the cores taken directly within the main introduction site, and was found in only three dead moss cores (containing 1, 5 and 12

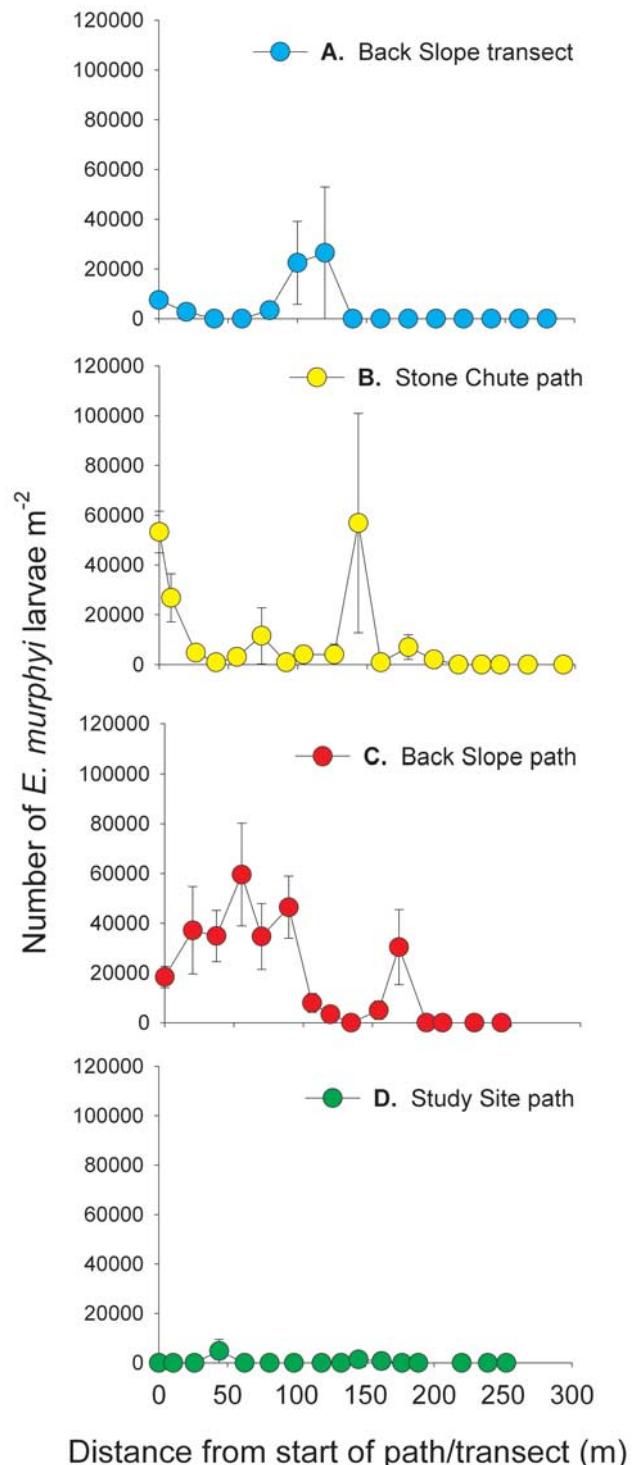


Fig. 5a–d. Plot of *E. murphyi* larval numbers along a transect and paths leading away from the research station. Distances between the likely introduction site (labelled A in Fig. 1) and the start of the Stone Chute, Back Slope and Study Site paths are 51, 55 and 71 m, respectively.

worms) at two sample sites situated 5 m apart and ~7 m south of the main introduction site. The mean number of

worms at each site was *c.* 6600 m⁻² and 2000 m⁻², although there was a high degree of aggregation. It is possible that *C. blocki* was more widely distributed, but either at a greater depth or in numbers sufficiently low to be missed by our sampling method. No other enchytraeid worms were found during the study.

Quantification of larvae on a transect away from the main introduction site

Figure 5a shows the mean number of midge larvae per core sampled at 20 m intervals along a line from the probable introduction site towards Observation Bluffs. Comparatively low numbers of larvae were found close to the introduction site ($< 1.0 \times 10^4$ m⁻²) with none found at the 40 and 60 m sample points. However, the highest numbers were found between 80 and 120 m from the introduction site, with one replicate core (at 120 m) containing 8.0×10^4 larvae m⁻². No larvae were detected in core samples beyond the 120 m point, which may indicate either a maximum extent of larval colonization along that line, or that larval concentrations were below the sensitivity of the sampling methodology. This transect line crossed undisturbed areas and did not follow any particular paths. However, it did cross the Back Slope path at *c.* 75 m a.s.l.

Quantification of larvae on paths leading away from the research station

There was no significant difference between the numbers of larvae in cores collected on the Stone Chute or Back Slope paths compared with the numbers of larvae in cores collected either side of the path (*c.* 10 m to either side of the path), and thus these data were pooled. Figure 5b–d shows the concentrations of larvae along the Stone Chute, Back Slope and Study Site paths, respectively. Therefore, for Fig. 5b & c, each data point represents the counts from nine cores per data point). Figure 5d shows the mean values of three on-path replicates for the Study Site path.

Overall, larvae were not recorded on or around any of the paths beyond 198 m along a path, which is around 220 m from the main introduction site. Larval concentration on the Stone Chute path declined initially with distance from the station; however, a high mean concentration of almost 5.7×10^4 larvae m⁻² was encountered 144 m from the start of the path (including a single core with a concentration of 4.1×10^5 larvae m⁻²). The Back Slope path showed high levels of colonization by the midge for the first 90 m, with mean larval concentrations of between 1.8×10^4 and 6.0×10^4 larvae m⁻². However, at points beyond 90 m, numbers were less than 1×10^4 larvae m⁻², with the exception of a point 169 m along the path where over 3.0×10^4 larvae m⁻² were found. On the Study Site path, larvae were largely absent, but where found, were in concentrations of less than 5.0×10^3 larvae m⁻².

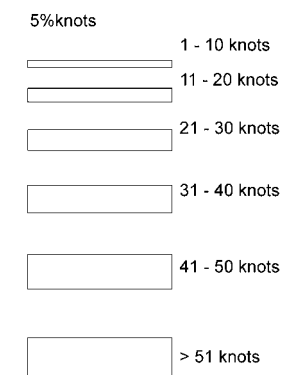
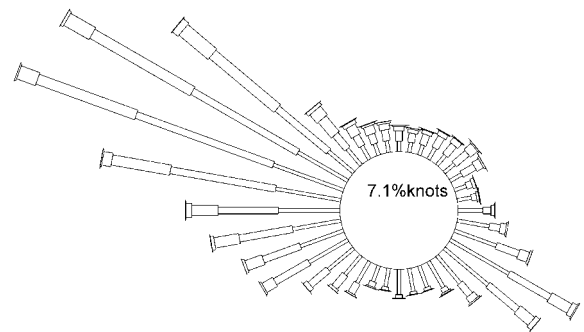


Fig. 6. Wind speed and direction recorded at the meteorological station at Signy Research Station (summary of data collected between 1957 and 1995).

Wind speed and direction

Figure 6 shows the wind speed and direction recorded at the meteorological station at Signy Research Station. Strong south-easterly and north-westerly winds dominate.

Estimation of larval mean fresh and dry mass

The water content of the midge larvae varied considerably from 3.00 to 1.71 g g⁻¹ with a mean of 2.65 ± 0.46 . The mean fresh mass was 1.36 mg (SD ± 0.45) and the mean dry mass 0.36 mg (SD ± 0.13).

Discussion

Distribution and colonization success

This study is the first detailed survey of the distribution of Antarctica's only two known terrestrial non-native invertebrates, *E. murphyi* and *C. blocki*, since their probable introduction to Signy Island in the late 1960s. Table 1

Table I. Spatial distribution and concentration of *E. murphyi* larvae and *C. blocki* at Signy Island, South Orkney Islands.

Reference	Study date(s)	<i>E. murphyi</i> colonized area (concentration)	<i>C. blocki</i> colonized area (concentration)	Comments
Burns 1982	1980	~1 m ² (13 000 larvae m ⁻²)	~1 m ²	'Larvae have been found in highest densities (1.3 cm ⁻²) in the top few millimetres of almost pure stands of <i>Polytrichum alpestre</i> that have colonized the site.'
Block <i>et al.</i> 1984	1981/82	> 1 m ² (25 718 larvae m ⁻² highly aggregated)	~1 m ² (3243 worms m ⁻² highly aggregated)	'Extensive examination of the ground immediately adjacent to the site has produced little evidence of spread by either taxon.' 'A single larva of <i>E. murphyi</i> was found 40 cm from the edge of the site in 1982, whilst six adults were seen on moss <i>c.</i> 25 m from the site the following year.'
Cranston 1985	1981/82	(7500 larvae m ⁻²)		
Smith unpublished	1985	~15 m ²		'2 January 1985. J. Edwards' main experimental site. During period of brilliant sun, numerous adult midges seen crawling over surface of peat. Larvae abundant in soil. Eggs under stones and pieces of loose peat lying as lumps on surface. Eggs and many larval skins + occasional adults seen beneath stones and pieces of peat up to 30 cm outside site and 3 m downhill from site (downhill and to left).'
Smith 1996	1989	~2000 m ²	~1 m ²	'No enchytraeids were found beyond the [introduction] site but, by 1989, chironomid larvae had spread through out a radius of about 25 m from the original experimental plot.'
Convey 1992	1989/90 1990/91	> 600 m ²		'The species [<i>E. murphyi</i>] now occurs frequently within an area of at least 600 m ² centred on the point of origin.'
Smith, unpublished data	1991		(?)	Enchytraeid worm found <i>c.</i> 10 m from introduction site.
Dózsa-Farkas & Convey 1997	1995	~ 16 000 m ² (?)		'Both introduced species appear to be well pre-adapted to the more rigorous conditions of the maritime Antarctic, the midge having spread approximately 100 m from the introduction site...'
This study	2007–09	~35 000 m ² (<i>c.</i> 21 000 larvae m ⁻²)	<i>C. blocki</i> only found at two sample sites, 5 m apart, 7 m south of the main introduction site (2000 and 6600 worms m ⁻²)	<i>E. murphyi</i> found up to 220 m from main introduction site. <i>C. blocki</i> highly aggregated in an area close to the introduction site.

summarizes the limited available data on the extent and concentration of the two species since 1982, largely in the form of *ad hoc* observations. Most previous reports detail the extent of the midge and worm centred on the main introduction site (Fig. 1, site A). However, it is possible that the non-native species were introduced to more than one site in the immediate vicinity of the station, e.g. Fig. 1, site B (R.I.L. Smith, personal communication 2008).

Frenot *et al.* (2005) defined a persistent alien as having 'survived, established and reproduced for many years in a restricted locality, but has not expanded range from that location'. In this context, after around 15 years with apparently little expansion from the original establishment site, *E. murphyi* has extended its distribution range since at least the mid-1980s to an area of *c.* 35 000 m² and as a result its colonization status may no longer be considered 'persistent'. Further research is required to show whether

or not the midge should be considered an invasive alien, i.e. has spread into native communities and displaced native species (Frenot *et al.* 2005).

If *E. murphyi* has, at a minimum, a two year life-cycle as suggested by Convey & Block 1996, then the midge has survived on Signy Island for around 20 generations. We do not know if the rate of colonization of new habitat on Signy by the midge has increased, though any change could be due to a reduction in the severity of environmental conditions on the island due to climate change, or a change in the physiology of the midge. Alternatively, earlier reports on the extent of the area colonized by the larvae may have been of limited accuracy and the midge population may be undergoing an unchanged rate of expansion made possible by existing behavioural and physiological adaptations.

Within the 2775 m² grid surveyed around the introduction site (equal in area to a 53 m × 53 m square)

there were estimated to be 5.9×10^7 larvae. Using our rough calculation of mean larval mass, this may equate to a larval fresh mass of up to 80 kg. With mean fresh and dry masses estimated to be *c.* 29 and 8 g m^{-2} , respectively, it is likely that the midge larvae are having an impact on nutrient turnover in the colonized area. In contrast with *E. murphyi*, since the 1960s, the *C. blocki* population has not expanded its spatial distribution greatly, being absent from the introduction site and only found in two locations $\sim 7 \text{ m}$ away. Within the sub-Antarctic, invasive status of the non-native earthworms *Dendrodriulus rubidus tenuis* (Eisen) has been reported to vary with location; on Possession Island it has not spread much beyond the research station and is persistent, while on Kerguelen it is invasive (Frenot *et al.* 2005). Little information exists on non-native invertebrates that have been transient over a timescale of decades within the Antarctic and sub-Antarctic literature although there are several reports of winged insects existing as transient aliens on sub-Antarctic islands (Hänel *et al.* 1998, Greenslade *et al.* 1999, Convey 2005). In the southern polar region, invertebrate invasive species have received relatively little attention by environmental managers compared to invasive vertebrates such as rats, cats and rabbits (Chapuis *et al.* 2001, Bester *et al.* 2002). Nevertheless, sub-Antarctic invasive invertebrates have been shown capable of altering ecosystem structure and function (Chown *et al.* 2008). A predatory beetle, *Trechisibus antarcticus* Dejean, introduced to South Georgia and invading the coastal lowlands around Husvik, has affected the endemic detritivorous/herbivorous beetle *Hydromedion sparsutum* Müll by decreasing its abundance and causing an increase in mean body size (Ernsting *et al.* 1995, 1999). On Kerguelen, the flightless cold-temperate predatory carabid beetle *Merizodus soledadinus* Guerin-Méneville threatens population of native invertebrates, particularly in the littoral zone, while the necrophagous blowfly *Calliphora vicina* Robineau-Desvoidy may lead to intra-specific resource competition between the larvae of the native wingless fly *Anatalanta aptera* Eaton (Chevrier *et al.* 1997, Frenot *et al.* 2008). In lowland plant communities of sub-Antarctic Marion Island, caterpillars of the indigenous flightless moth *Pringleophaga marioni* Viette were thought to dominate nutrient cycling. However, Hänel & Chown (1998) suggested that the introduced parthenogenetic chironomid midge, *Limnophyes minimus* Meigen, may be undertaking substantial nutrient turnover comparable with that of *P. marioni*.

Dispersal processes

The midge larvae have an aggregated and patchy distribution in the vicinity of the Signy Island Research Station but it is unclear if they are being dispersed by processes other than crawling. Wing reduction (brachyptery) is a characteristic exhibited by some insects found in low temperature or high wind environments such as exist in Antarctica (e.g. *Belgica*

antarctica Jacobs) and the sub-Antarctic islands (Convey & Block 1996). The wind direction around the research station is often from the WNW, which could blow the midge adults up the back slope (Fig. 6). However, wind patterns can differ greatly near the ground surface and if wind were a successful dispersal method, we might expect a greater distribution of the midge larvae in suitable substrates further from the introduction site, given that up to 40 years may have elapsed since the initial introduction of the midge. Temporary drainage streams are unlikely to assist in adult midge distribution as all streams in the area flow north towards the main introduction site and the sea. Birds may play a part in distribution to other areas of the South Orkney Islands and beyond, but at present we have no data to confirm this.

We found no evidence that midge larvae numbers were higher on the paths than on similar substrates off the paths. However, other workers have shown human movement along paths to be an effective means of aiding dispersal of non-native species (see Frenot *et al.* 2008). An island-wide survey was beyond the scope of this study; however, it is possible that the midge and/or worm have been distributed to other areas of Signy Island in peat or mud attached to clothing, footwear or equipment. Although no evidence supporting this has been reported by other researchers on the island, a generic challenge to any assessment of the status and distribution of small terrestrial invertebrates such as these is the ability to detect new founding populations at an early stage after establishment.

Habitat availability and colonization

Physical features around the main introduction sites (e.g. the research station to the north, ocean to the west and cliffs to the south-west) limit the area where suitable substrata are available for colonization by the midge larvae and worm. Our study has shown that the midge larvae have a habitat preference for dead moss and peat substrata over stony soil and gravel or live moss and other plants. *Eretmoptera murphyi* is likely to be a detritovore, and live mosses may not be a favourable substratum (Convey & Block 1996). Mosses with dense, closely packed stems may restrict access of the larvae to sub-surface layers, although the midge has also been reported in association with lichens and in particular *Cladonia* spp. (Block *et al.* 1984). Given that the midge larvae can colonize a wide range of substratum types, there is clearly a large area of suitable habitat on Signy Island as a whole. On South Georgia, *E. murphyi* is found in fellfield areas up to 300 m a.s.l. in the moss *Racomitrium austro-georgicum* (Ring *et al.* 1990), while on Signy Island it has been found up to 40 m a.s.l. mainly in peat (this study).

Signy Island has undergone substantial deglaciation over the last 50 years (Smith 1990), creating new areas of ice-free ground suitable for colonization by vegetation and

Table II. Colonization status of known non-native species in the Antarctic terrestrial environment.

Species	Location	Date introduced	Colonization status	Colonization area	Notes	References
<i>Eretmoptera murphyi</i>	Signy Research Station, South Orkney Islands, Scotia Arc.	1967, 1968(?)	Expanding	~35 000 m ²		this manuscript and references therein
<i>Christensenidrilus blocki</i>	Signy Research Station, South Orkney Islands, Scotia Arc.	1967, 1968(?)	Persistent	<150 m ²		this manuscript and references therein
<i>Poa annua</i>	Arctowski Station, King George Island, South Shetland Islands	1985/86	Expanding	Has spread over 500 m into the vicinity of the nearby Antarctic Specially Protected Area (ASPA) 128 Western Shore of Admiralty Bay. Expansion may be attributed to regional warming	1985/86: first found in metal grating at Arctowski Station main building. 1990: spread to greenhouse area and above subterranean hot water pipes within a single area of c. 0.4 km ² . 1991/92: found in a number of locations with disturbed ground. 2005/06: found growing amongst indigenous plant communities for the first time. <i>Poa annua</i> invasive on four sub-Antarctic islands	Olech 1996 Smith 1996 Olech 2003 Frenot <i>et al.</i> 2005 Chwedorzewska 2008
<i>Poa pratensis</i>	Primavera Station, Cierva Point, Palmer Archipelago, Antarctic Peninsula	1954/55	Persistent	Limited to an area c. 40 cm across, possibly because the microclimate is unsuitable for growth outside this restricted area.	Transplant experiments of <i>Nothofagus antarctica</i> and <i>N. pumilo</i> in soil imported from Ushuaia (Terra del Fuego). <i>P. pratensis</i> invasive on three sub-Antarctic islands.	Corte 1961 Smith 1996 (Last report 1995) Frenot <i>et al.</i> 2005
<i>Poa trivialis</i>	Syowa Station, Enderby Land, East Antarctica	1993(?)	Eradicated	Single plant	Removed in 2007 (S. Imura, personal communication 2007)	Japan 1996

associated fauna at both lower and higher altitudes. The midge may be restricted to lower altitudes on Signy Island as its physiological pre-adaptations may not be adequate to survive the more severe climatic conditions likely to be experienced at higher altitude. Some doubt remains as to whether or not *E. murphyi* larvae are freeze susceptible (Block *et al.* 1984) or freeze tolerant (Harrison & Block 1988, Convey & Block 1996), and further clarification is needed. Block *et al.* (1984) found trehalose, glucose, fructose and glycerol in low concentrations (< 1% of fresh body weight of larvae, pupae and adults), but these concentrations are unlikely to have a significant effect on the organism's supercooling capacity. Nevertheless, at both South Georgia and Signy Island, the soil and moss habitat chosen by the midge allows it to be buffered from climatic extremes (Davey *et al.* 1992). No males have been reported, and the species is considered to be an obligate parthenogenetic species, which may be an adaptation to allow rapid reproduction under harsh conditions (Cranston 1985, Convey 1992, Hullé *et al.* 2003). Further expansion of the midge may not be limited by direct competitors or predators, as these are largely absent on Signy Island. Of the five known alien species in terrestrial Antarctica, *E. murphyi* and *P. annua* (King George Island, South Shetland Islands) may currently present the highest risk to their local indigenous communities (see Table II). Their impact on the Antarctic terrestrial ecosystem may be increased greatly if they are distributed to other Antarctic locations, by either natural or anthropogenic processes.

Past and current biosecurity precautions

Those undertaking the transplantation experiments at Signy Research Station were aware of the recommendations for conservation of the Antarctic flora and fauna mentioned in the Antarctic Treaty, and a specific permit was obtained for their work (Edwards 1980). Measures were taken to ensure that the imported plants did not spread beyond the study sites. After completion of the transplantation experiments in February 1973, all plant remains were removed and burnt, and the imported soil disposed of at sea. However, these measures may have proven inadequate to prevent the establishment of the enchytraeid worm and chironomid midge.

There is no evidence to suggest that either *E. murphyi* or *C. blocki* have been translocated from Signy to other UK research stations served by the British Antarctic Survey ships that resupply Signy Research Station. However, both species were found recently in soil and vegetation moved inadvertently on industrial plant equipment between South Georgia and Rothera Research Station (Adelaide Island, Antarctic Peninsula) (Hughes *et al.* 2009). Nevertheless, the risk of expanding the area colonized by these non-native species through human activities should be reduced to a minimum. The British Antarctic Survey now requires

everyone leaving Signy Island to wash soil and mud from their footwear using disinfectant at the earliest opportunity and before landing at any other Antarctic location. Eradication of *C. blocki*, or possibly *E. murphyi*, may be feasible, but would result in the destruction of a large area of rich terrestrial habitat, of considerable scientific value, close to the research station. The implications of any eradication attempt would need to be considered carefully.

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