

The HKU Scholars Hub



Title	Emergence of macrolide-resistant Mycoplasma pneumoniae in Hong Kong is linked to increasing macrolide resistance in the multilocus variable-number tandem-repeat analysis type 4-5-7-2
Author(s)	Ho, PL; Law, PYT; Chan, BWK; Wong, CW; To, KKW; Chiu, SS; Cheng, VCC; Yam, WC
Citation	Journal of Clinical Microbiology, 2015, v. 53 n. 11, p. 3560-3564
Issued Date	2015
URL	http://hdl.handle.net/10722/220171
Rights	Journal of Clinical Microbiology. Copyright © American Society for Microbiology.

1 JCM01983-15 revised

2	Emergence of macrolide-resistant Mycoplasma pneumoniae in Hong Kong is linked to						
3	increasing macrolide resistance in the multilocus variable-number tandem-repeat						
4	analysis type 4-5-7-2						
5							
6	Pak-Leung Ho, ^{a,b} Pierra Y. Law, ^a Betsy W. K. Chan, ^a Chun-Wai Wong, ^a Kelvin K. W. To, ^a						
7	Susan S. Chiu, ^c Vincent C. C. Cheng, ^{a,b} Wing-Cheong Yam ^{a,b}						
8							
9	Department of Microbiology, ^a Carol Yu Centre for Infection, ^b and Department of Pediatrics						
10	and Adolescent Medicine, ^c Queen Mary Hospital, The University of Hong Kong, Pokfulam,						
11	Hong Kong Special Administrative Region, China.						
12							
13	Running title: MLVA type associated with macrolide-R M. pneumoniae						
14	Keywords: molecular typing, Mycoplasma pneumoniae, antimicrobial resistance, pneumonia						
15							
16	*Corresponding author. P. L. Ho						
17	Department of Microbiology, The University of Hong Kong, Queen Mary hospital, Pokfulam						
18	Road, Pokfulam, Hong Kong SAR, CHINA. Tel: +852-2855 4897; Fax: +852-2855 1241; E-						
19	mail: <u>plho@hkucc.hku.hk</u>						
20							
21	Word counts: Abstract 207						
22							

24 ABSTRACT

Macrolide-resistant Mycoplasma pneumoniae (MRMP) is rapidly emerging in Asia but 25 26 information on the temporal relationship between the increase in macrolide resistance and changes in strain types is scarce. Between 2011 and 2014, M. pneumoniae infection was 27 28 diagnosed by PCR as part of routine care in a healthcare region in Hong Kong. Testing was 29 initiated by clinicians, mainly in patients with suspected M. pneumoniae pneumonia. 30 Specimens positive for *M. pneumoniae* were retrospectively investigated by macrolide 31 resistance genotyping and a four loci (Mpn13-16) multilocus variable-number tandem-repeat 32 analysis (MLVA) scheme. The overall percentage of *M. pneumoniae*-positive specimens was 17.9% with annual rates ranging from 9.8%-27.2%. Prevalence of MRMP had rapidly 33 increased from 13.6% in 2011, 30.7% in 2012, 36.6% in 2013 to 47.1% in 2014 (P = 0.038). 34 35 Two major MLVA types 4-5-7-2 and 3-5-6-2 accounted for 75%-85% of the infections each 36 year. MLVA types 4-5-7-2 and 3-5-6-2 predominated among macrolide-resistant and macrolide-sensitive groups, respectively. Increase in MRMP was mainly caused by 37 38 increasing macrolide resistance in the prevalent MLVA type 4-5-7-2, changing from 25.0% 39 in 2011, 59.1% in 2012, 89.7% in 2013 to 100% in 2014 (P < 0.001). In conclusion, 40 increasing MRMP in Hong Kong was linked to a single MLVA type which was both 41 prevalent and increasingly resistant to macrolides.

42

44 INTRODUCITON

45 Mycoplasma pneumoniae (MP) is a common cause of community-acquired pneumonia and 46 other respiratory tract infections (19). Community epidemics occur at intervals of 3 to 7 years. 47 Infections develop in persons of all ages but it is primarily a disease of children and teenagers (2). When treatment is indicated, a macrolide is usually the drug of choice (2,19). However, 48 49 macrolide-resistant *M. pneumoniae* (MRMP) have become increasingly prevalent worldwide and high rates (>80%) have been found in certain parts of the world (14,16,18,23). MRMP 50 51 infections have been associated with persistence of symptoms, slower reduction in bacterial 52 load, longer hospital stays, requirement for alternative therapy and higher frequency of 53 complications (6,19,26). Strain typing is important for understanding changes in disease 54 epidemiology and for investigation of outbreaks. In 2009, a multilocus variable-number 55 tandem-repeat analysis (MLVA) scheme based upon five loci (Mpn1, Mpn13-16) was 56 developed for the molecular typing of MP (8). It was initially used for investigation of 57 isolates, but was later modified for directly typing MP in respiratory specimens (4,10,24). An 58 amended 4 loci MLVA scheme was later proposed after studies had raised concerns on the 59 instability of the Mpn1 locus (1,21). In clinical laboratories, culture and characterization of MP is seldom performed. Therefore, MP typing was usually carried out on isolates collected 60 61 from sporadic cases and outbreaks (1,8,9), limiting the inferences that can be made about 62 trends in MP infections. In addition, information on the temporal relationship between the 63 increase in macrolide resistance and changes in strain types is scarce (9). Here, MLVA 64 analysis was used to investigate the MP strain type and macrolide resistance genotype in respiratory specimens collected consecutively from patients in a healthcare region in Hong 65 66 Kong over a 4-year period.

67

69 MATERIALS AND METHODS

70 Study design. This retrospective study was conducted in a healthcare region in Hong Kong 71 comprising one university-affiliated hospital with 1600 beds, three extended-care hospitals 72 with a total of 1600 beds, and one paediatric hospital with 160 beds. Diagnostic PCR assay 73 for MP was provided as a routine service for inpatients by a clinical microbiology laboratory 74 (5,6). Testing was initiated by clinicians, mainly in patients with features suspicious of MP pneumonia (2,15). Nasopharyngeal aspirate was collected in viral transport medium (7). 75 76 Sputum and other respiratory specimens were collected using standard techniques (5). 77 Patients were included if their respiratory specimens were obtained for MP testing by PCR 78 between January 2011 and December 2014. During the study period, a total of 1657 79 respiratory specimens from 1433 patients were investigated by real-time PCR test for the 80 presence of MP. Overall, 257 (17.9%) patients, including 274 (16.5%) specimens were 81 positive for MP. The 274 MP-positive specimens comprised 264 nasopharyngeal 82 aspirates/swabs, five pleural specimens and five other respiratory specimens (sputum, 83 bronchial aspirate). The data was analysed by five age groups: 0-1 year (infants, n = 11), 2-84 11 years (children, n = 195), 12-17 years (teenagers, n = 33), 18-64 years (adults, n = 16), ≥ 65 years (seniors, n = 2). The patients were diagnosed with pneumonia (n = 231), upper 85 86 respiratory tract infection (n = 7), non-specific respiratory illness (n = 9) and acute 87 bronchiolitis (n = 1). In nine patients, no information on the syndromic diagnosis was 88 available. Clinical features and macrolide resistance genotyping results for 101 of the patients 89 have been reported previously (5,6). Nucleic acid extracts from the 257 patients with positive 90 MP results were retrospectively retrieved for further testing. Only one specimen from each 91 patient was included.

93 Nucleic acid extraction. Nucleic acid extraction was performed by using the NucliSENS 94 easyMAG extraction system (bioMérieux, France) and stored at -80 °C, as described 95 previously (5). All testing was performed on nucleic acid extracted from the clinical 96 specimens. Culture for MP was not performed.

97

98 **Real-time qPCR for the detection of** *M. pneumoniae*. Real-time quantitative (qPCR) was 99 conducted for the detection of MP using TaqMan universal PCR master mix (Applied 100 Biosystems) in a StepOnePlus instrument (Applied Biosystems, Foster City, CA), as previously described (5). A series of 6 log₁₀ dilutions equivalent to ten to 1×10^6 copies per 101 102 reaction mixture were prepared from a plasmid (pC-RII-TOPO vector; Invitrogen, CA) 103 containing the corresponding target bacterial sequence to generate calibration curves; these 104 were ran in parallel with the test specimens. The detection limit of the qPCR assay was 105 approximately ten copies per reaction mixture (5).

106

107 **MRMP genotype detection.** SimpleProbe real-time PCR coupled to melting curve analysis 108 (SimpleProbe PCR) was performed on the extracted nucleic acid from specimens to identify 109 MRMP. The MRMP assay was done by using the LightCycler FastStart DNA master 110 HybProbe kit (Roche Diagnostics, Germany) according to a published protocol (5). The 111 detection limit of SimpleProbe PCR for both wild-type and mutant was 10³ copies per 112 reaction. A randomly chosen subset of the specimens was subjected to Sanger sequencing for 113 confirmation.

114

MLVA typing. Previously published primers were used to amplify four variable-number tandem-repeat (VNTR) loci (Mpn13-16) (24). Our initial testing showed that nonspecific bands were commonly observed if all loci were amplified together in one multiplex reaction.

118 After optimization, good results were obtained through amplification of the loci in two duplex reactions, one for Mpn13 and Mpn15, and one for Mpn14 and Mpn16. The PCR was 119 performed using 2 μ l nucleic acid, 15 μ l 2 \times QIAGEN multiplex master mixes (QIAGEN), 120 121 concentrations for each primers were 0.2 µM for Mpn13 and Mpn15, 0.4 µM for Mpn14 and 122 0.08 µM for Mpn16. The total reaction mixture volume was made up to 30 µl with nuclease-123 free water. A Veriti 96-well thermal cycler (Applied Biosystems) was used for amplification. 124 Cycling conditions were as follows: denaturation step of 15 min at 95 °C, amplification step 125 of 40 cycles of 30 s at 95 °C, 30 s at 62 °C, and 45 s at 72 °C. The products were then pooled 126 into one tube for product size determination in one lane by capillary electrophoresis using an 127 ABI 3130 genetic analyser (Applied Biosystem), and the data was analysed using 128 GeneMapper software (version 4.0, Applied Biosystems). The primers were fluorescently 129 labelled at the 5' end with VIC (green, Mpn13 and Mpn15), NED (yellow, Mpn14) or 6-130 FAM (blue, Mpn16) (Applied Biosystem). The fluorescent labels for the targets together with 131 the expected product sizes of each locus allowed sizing of the amplicons for all four loci in 132 one reaction mixture. The number of repeats for each locus was calculated according to the 133 PCR fragment size. The MLVA type was designated by the numeric combination of the number of tandem repeats at four loci (Mpn13-16), as suggested previously (1,9). The 134 135 number of repeats was rounded up to an integer value (8,24). The number of repeats in each 136 locus (1 to 2 specimens for each product size) was confirmed by Sanger sequencing.

137

138 **Statistical analysis.** Statistical analysis was performed using SPSS Statistics version 23 for 139 Windows. Chi-square tests were used to compare categorical variables. A P value of < 0.05 140 was considered statistically significant.

141

142

143 **RESULTS**

144**Prevalence of** *M. pneumoniae.* The percentage of test-positive patients per month from 2011145to 2014 is shown in Fig 1. Higher positive rates (more than 1 standard deviation above146average for the entire period) were observed in 2012 (April, June, July, September) and 2013147(May to July). The percentage of test-positive patients by year was 9.8% in 2011, 27.2% in1482012, 24.3% in 2013 and 11.4% in 2014 (P < 0.001).

The MP-positive rate was highest among children aged 2-11 years (33.2%) and teenagers aged 12-17 years (30.6%), then in infants aged 0 to 1 year (7.6%); it was lowest in adults aged 18 to 64 years (4.0%) and seniors aged 65 years (1.0%) (P < 0.001). The positive rate was higher in females than in males (21.8% versus 14.5%, P < 0.001).

153

154 Prevalence of macrolide-resistant genotype. The MP-positive specimens for 16 patients 155 were of an insufficient amount and not investigated further. Macrolide-resistant genotyping 156 could be successfully carried out on all specimens from the remaining 241 patients. 157 SimpleProbe real-time PCR coupled to melting curve analysis identified 34.9% (84/241) of the unique patient specimens as MRMP genotype. The A2063G transition was the only 158 159 mutation identified. A subset of 88 specimens, including 61 with MSMP genotype and 27 160 with MRMP genotype was further analysed by Sanger sequencing. The results were 100% 161 concordant with melting curve analysis. The annual prevalence of MRMP among all MP-162 positive patients had significantly increased from 13.6% (3/22) in 2011 to 30.7% (23/75) in 2012, 36.6% (34/93) in 2013 and 47.1% (24/51) in 2014 (P = 0.038). The prevalence of 163 MRMP genotype was higher among children (aged 0-1 years, 30.0%; aged 2-11 years, 36.1%; 164 165 aged 12-17 years, 39.7%) than in adults (aged 18-64 years, 20.0%, aged \geq 65 years, 0%) but

166 the difference was not statistically significant (P = 0.122). MRMP prevalence among males 167 (33.7%) and females (35.7%) were similar (P = 0.742).

168

Temporal changes in macrolide resistance rate and MLVA types. Specimens from the 241 patients with sufficient DNA extracts were further investigated by MLVA typing and successful results were obtained for 205 (85.1%) patients. The number of repeats in the four loci were 3 to 5 for Mpn13, 4 to 6 for Mpn14, 6 to 7 for Mpn15 and 2 to 3 for Mpn16, giving seven distinct MLVA types. The major types were 3-5-6-2 (44.4%), 4-5-7-2 (36.6%) and 4-5-7-3 (14.1%). Other rare types, including 3-6-6-2 (n = 4), 5-5-7-2 (n = 3), 4-6-7-3 (n = 2) and 4-4-7-3 (n = 1) only accounted for 4.9% of the total.

176 During the four year period, types 4-5-7-2 and 3-5-6-2 were predominant (Fig. 2). 177 Together the two types comprised 75% to 85% of all positive specimens in each year. The 178 proportion of type 4-5-7-2 had an increasing trend from 29% in 2011, 34% in 2012, 36% in 179 2013 and 43% in 2014 but the difference was not statistically significant (P = 0.686). Of the 180 seven MLVA types, five and four MLVA types were found among specimens with 181 macrolide-sensitive *M. pneumoniae* (MSMP) and MRMP genotypes, respectively (Fig. 3A). 182 The two major MLVA types (3-5-6-2 and 4-5-7-2) occurred in the MRMP and MSMP groups 183 at different frequencies. The prevalence of MLVA type 4-5-7-2 was substantially higher in 184 the MRMP group than in the MSMP group (89.6% versus 10.9%, P < 0.001). In contrast, MLVA type 3-5-6-2 was more prevalent in the MSMP group than in the MRMP group 185 186 (64.5% vs. 3.0%, P < 0.001). The other five MLVA types were found at low frequencies in either the MRMP group (5-5-7-2 and 4-6-7-3) or the MSMP group (4-5-7-3, 4-4-7-3 and M3-187 188 6-6-2) only. Stratification by year revealed that macrolide resistance rate of MLVA type 4-5-189 7-2 had significantly increased from 25.0% in 2011 to 100% in 2014 (Fig. 3B, P < 0.001).

191 **DISCUSSION**

192 In this study, an increase in the rate of MP infection was noted in 2012 and 2013, suggesting 193 that there was an epidemic outbreak during this period. During the entire period, changes in 194 the annual cycle of positive rates were irregular, with peaks in early summer (in 2012), mid-195 summer (in 2013) or early autumn (in 2014). This is in line with reports describing more MP 196 infections with increased relative humidity and ambient temperature (17,22). In our neighborhood areas, recent epidemic outbreaks of MP infections were also noted in South 197 198 Korea from 2010-2011, in Japan from 2011 to 2012 and in Beijing and Shanghai, China in 199 2012 (13,16,20). Notably, a substantial increase in the prevalence of MRMP was noted in the 200 areas during or shortly after those epidemics (12-14,16,20). Our data revealed that the MRMP 201 rate had increased by more than three folds from 13.6% in 2011 to 47.1% in 2014. Reported 202 rates of MRMP range from 62.9% in South Korea (12) and >80% to 90% in China and Japan 203 (14,26), compared to $\leq 10\%$ in Europe and the United States (9,19). The relationship between 204 MP epidemics and MRMP emergence is likely to be complex, involving selection pressure 205 from widespread administration of macrolides (12).

206 Two MLVA types (3-5-6-2, 44.4% and 4-5-7-2, 36.6%) accounted for 81.0% of the infections during the study period. Both types occur worldwide and were among the 207 208 predominant types in many studies. Amongst international collections of MP isolates 209 collected over decades, 14.0%-20.8% and 50.6%-55.1% were of MLVA types 3-5-6-2 and 4-210 5-7-2, respectively (1,8). During the periods with a high proportion of MP-positive specimens, 211 the major genotypes did not change (Figure 2A). This suggests that increased detection of MP infections is likely a result of increased transmission of co-circulating MLVA types, 212 213 rather than introduction of new types to the community.

This study found that increase in MRMP was predominantly a result of increasing resistance in MLVA type 4-5-7-2. Macrolide resistance rate of this type had drastically

216 increased from 25% in 2011 to 100% in 2014. All the other MLVA types including the 217 prevalent 3-5-6-2 type remained largely macrolide-sensitive. Qu et al previously reported the 218 same significant correlation between macrolide resistance and susceptibility with type 4-5-7-219 2 and type 3-5-6-2, respectively (20). In the two international MP collections described by 220 Degrange et al and Benitez et al, four (33.3%) of 12 and nine (90%) of ten MRMP isolates 221 were of MLVA type 4-5-7-2, respectively (1,8). In the United States, 13 (68.4%) of 19 222 MRMP isolates identified through CDC-assisted investigations across the country between 223 2006 and 2013 belonged to MLVA type 4-5-7-2 (9). In China, this MLVA type accounted for 224 >90% of the MRMP isolates identified in Beijing from 2010 to 2013 (20,24,25). Of the 21 225 distinct types that could be distinguished by the amended four loci MLVA scheme (1,3,4,8-226 11,18,20,21,23-25) (Table S1, supplementary file), MRMP has been detected in 12 types. 227 Among the published reports, prevalence of the other 11 types among MRMP were low and 228 occurrence was sporadic (1,8,9,11,18,20,21,23-25).

229 As far as we know, this is the first report to demonstrate a link between changes in 230 MRMP prevalence and increasing resistance within a single MLVA type. Inclusion of 231 consecutive specimens from a 4-year period and a relatively large sample size are the 232 strengths of this study. Given that this retrospective analysis only examined specimens from 233 inpatients of whom the majority were diagnosed with pneumonia, the findings may not be representative of mild MP infections in the community. MP is a genetically conserved 234 235 organism. Pairwise comparison of the four published MP genomes (strains M29, M129, 309 236 and FH) revealed that the difference between strains of different MLVA types (4-5-7-2 237 versus 3-5-6-2) was 0.3%-0.5% while difference between strains of the same MLVA types 238 was 0.08% (Table S2, supplementary file). Therefore, whole genome sequencing may be the 239 ultimate approach for resolving whether there is any MRMP subclone within type 4-5-7-2.

240	In summary, we demonstrated a link between increasing macrolide resistance and the
241	expansion of MRMP strains of the MLVA type 4-5-7-2 in Hong Kong during 2011-2014. It
242	is worrying that type 4-5-7-2 may be associated with more severe disease (20). Increasing
243	public awareness, enhancing access to rapid diagnostics and improving surveillance for MP
244	and macrolide resistance is necessary to inform case and outbreak management and to
245	understand the burden of disease.
246	
247	
248	
249	ACKNOWLEDGEMENTS:
250	This work was supported by grants from the Health and Medical Research Fund of the Food
251	and Health Bureau of the Government of the Hong Kong Special Administrative Region.
252	
252	
255	
<i>437</i>	

FIG 1. *M. pneumoniae* positive rate in respiratory specimens by date of request for a healthcare region in Hong Kong, January 2011-December 2014. Histogram shows the monthly number of specimens tested. Dotted line shows the percentage of specimens positive for MP in each month. Horizontal line showed the average positive percentage for the entire period. Only one specimen per patient was included.

260 261

FIG 2. MLVA type in *M. pneumoniae* specimens in a healthcare region in Hong Kong, 2011-2014. The percentage of each MLVA type for each year is shown. The number of patients in each year is shown within parentheses.

265 266

FIG 3. Macrolide resistance in *M. pneumoniae* specimens in a healthcare region in Hong Kong, 2011-2014. (A) MLVA type according to macrolide resistance genotype. The proportions of MLVA types for MRMP (n = 67) and MSMP (n = 138) groups are shown in the outer and inner doughnuts, respectively. Others in the MRMP group included types 4-6-7-3 and 5-5-7-2. Others in the MSMP group included types 3-6-6-2 and 4-4-7-3. (B) Changes in macrolide resistance rate of MLVA type 4-5-7-2 during 2011-2014. The number of patients in each year is shown within parentheses.

274

275

277 **REFERENCES**

Benitez, A. J., M. H. Diaz, B. J. Wolff, G. Pimentel, M. K. Njenga, A. Estevez, and
 J. M. Winchell. 2012. Multilocus variable-number tandem-repeat analysis of
 Mycoplasma pneumoniae clinical isolates from 1962 to the present: a retrospective
 study. J Clin.Microbiol 50:3620-3626. doi:JCM.01755-12 [pii];10.1128/JCM.01755-12
 [doi].

283 2. Bradley, J. S., C. L. Byington, S. S. Shah, B. Alverson, E. R. Carter, C. Harrison, S. 284 L. Kaplan, S. E. Mace, G. H. McCracken, Jr., M. R. Moore, S. D. St Peter, J. A. 285 Stockwell, J. T. Swanson, and Pediatric Infectious Diseases Society and the 286 Infectious Diseases Society of. 2011. The management of community-acquired 287 pneumonia in infants and children older than 3 months of age: clinical practice 288 guidelines by the Pediatric Infectious Diseases Society and the Infectious Diseases 289 Society of America. Clin.Infect Dis. 53:e25-e76. doi:cir531 [pii];10.1093/cid/cir531 290 [doi].

- Chalker, V., T. Stocki, D. Litt, A. Bermingham, J. Watson, D. Fleming, and T.
 Harrison. 2012. Increased detection of Mycoplasma pneumoniae infection in children
 in England and Wales, October 2011 to January 2012. Euro.Surveill 17.
- Chalker, V., T. Stocki, M. Mentasti, D. Fleming, and T. Harrison. 2011. Increased
 incidence of *Mycoplasma pneumoniae* infection in England and Wales in 2010:
 multiocus variable number tandem repeat analysis typing and macrolide susceptibility.
 Euro.Surveill 16.
- 298 5. Chan, K. H., K. K. To, B. W. Chan, C. P. Li, S. S. Chiu, K. Y. Yuen, and P. L. Ho.
 2013. Comparison of pyrosequencing, Sanger sequencing, and melting curve analysis

for detection of low-frequency macrolide-resistant mycoplasma pneumoniae
quasispecies in respiratory specimens. J Clin.Microbiol 51:2592-2598. doi:JCM.0078513 [pii];10.1128/JCM.00785-13 [doi].

- 303 6. Cheong, K. N., S. S. Chiu, B. W. Chan, K. K. To, E. L. Chan, and P. L. Ho. 2014.
 304 Severe macrolide-resistant *Mycoplasma pneumoniae* pneumonia associated with
 305 macrolide failure. J Microbiol Immunol.Infect. doi:S1684-1182(14)00230-8
 306 [pii];10.1016/j.jmii.2014.11.003 [doi].
- 307 7. Chiu, S. S., P. L. Ho, M. J. Peiris, K. H. Chan, and E. L. Chan. 2014. Population308 based hospitalization incidence of respiratory viruses in community-acquired
 309 pneumonia in children younger than 5 years of age. Influenza.Other Respir.Viruses.
 310 8:626-627. doi:10.1111/irv.12277 [doi].
- Bebear. 2009. Development of multiple-locus variable-number tandem-repeat analysis
 for molecular typing of *Mycoplasma pneumoniae*. J Clin.Microbiol 47:914-923.
 doi:JCM.01935-08 [pii];10.1128/JCM.01935-08 [doi].
- 9. Diaz, M. H., A. J. Benitez, and J. M. Winchell. 2015. Investigations of *Mycoplasma pneumoniae* Infections in the United States: Trends in Molecular Typing and Macrolide
 Resistance from 2006 to 2013. J Clin.Microbiol 53:124-130. doi:JCM.02597-14
 [pii];10.1128/JCM.02597-14 [doi].
- 319 10. Dumke, R. and E. Jacobs. 2011. Culture-independent multi-locus variable-number
 320 tandem-repeat analysis (MLVA) of *Mycoplasma pneumoniae*. J Microbiol Methods
 321 86:393-396. doi:S0167-7012(11)00222-3 [pii];10.1016/j.mimet.2011.06.008 [doi].

322	11. Dumke, R., C	. Schnee, M. V	W. Pletz, J. Rup	p, E. Jacobs, K.	Sachse, a	and G. Rohde.				
323	2015. Mycople	2015. Mycoplasma pneumoniae and Chlamydia spp. infection in community-acquired								
324	pneumonia,	Germany,	2011-2012.	Emerg.Infect	Dis.	21 :426-434.				
325	doi:10.3201/eid2103.140927 [doi].									

- Hong, K. B., E. H. Choi, H. J. Lee, S. Y. Lee, E. Y. Cho, J. H. Choi, H. M. Kang, J.
 Lee, Y. M. Ahn, Y. H. Kang, and J. H. Lee. 2013. Macrolide resistance of *Mycoplasma pneumoniae*, South Korea, 2000-2011. Emerg.Infect Dis. 19:1281-1284.
 doi:10.3201/eid1908.121455 [doi].
- 330 13. Kim, E. K., Y. S. Youn, J. W. Rhim, M. S. Shin, J. H. Kang, and K. Y. Lee. 2015. Epidemiological comparison of three Mycoplasma pneumoniae pneumonia epidemics 331 332 in а single hospital over 10 years. Korean J Pediatr. **58**:172-177. doi:10.3345/kjp.2015.58.5.172 [doi]. 333
- 14. Liu, Y., X. Ye, H. Zhang, X. Xu, and M. Wang. 2012. Multiclonal origin of
 macrolide-resistant *Mycoplasma pneumoniae* isolates as determined by multilocus
 variable-number tandem-repeat analysis. J Clin.Microbiol 50:2793-2795.
 doi:JCM.00678-12 [pii];10.1128/JCM.00678-12 [doi].
- Mandell, L. A., R. G. Wunderink, A. Anzueto, J. G. Bartlett, G. D. Campbell, N. C.
 Dean, S. F. Dowell, T. M. File, Jr., D. M. Musher, M. S. Niederman, A. Torres, and
- Society consensus guidelines on the management of community-acquired pneumonia in
 adults. Clin.Infect Dis. 44 Suppl 2:S27-S72. doi:CID41620 [pii];10.1086/511159 [doi].

C. G. Whitney. 2007. Infectious Diseases Society of America/American Thoracic

340

343 16. Okada, T., M. Morozumi, T. Tajima, M. Hasegawa, H. Sakata, S. Ohnari, N.
344 Chiba, S. Iwata, and K. Ubukata. 2012. Rapid effectiveness of minocycline or

345 doxycycline against macrolide-resistant Mycoplasma pneumoniae infection in a 2011
346 outbreak among Japanese children. Clin.Infect Dis. 55:1642-1649. doi:cis784
347 [pii];10.1093/cid/cis784 [doi].

- 348 17. Onozuka, D., M. Hashizume, and A. Hagihara. 2009. Impact of weather factors on
 349 *Mycoplasma pneumoniae* pneumonia. Thorax 64:507-511. doi:thx.2008.111237
 350 [pii];10.1136/thx.2008.111237 [doi].
- 18. Pereyre, S., A. Touati, J. Petitjean-Lecherbonnier, A. Charron, A. Vabret, and C.
 Bebear. 2013. The increased incidence of *Mycoplasma pneumoniae* in France in 2011
 was polyclonal, mainly involving *M. pneumoniae* type 1 strains. Clin.Microbiol Infect
 19:E212-E217. doi:10.1111/1469-0691.12107 [doi].
- 355 19. Principi, N. and S. Esposito. 2013. Macrolide-resistant *Mycoplasma pneumoniae*: its
 356 role in respiratory infection. J Antimicrob.Chemother. 68:506-511. doi:dks457
 357 [pii];10.1093/jac/dks457 [doi].
- Qu, J., X. Yu, Y. Liu, Y. Yin, L. Gu, B. Cao, and C. Wang. 2013. Specific
 multilocus variable-number tandem-repeat analysis genotypes of *Mycoplasma pneumoniae* are associated with diseases severity and macrolide susceptibility. PLoS
 One. 8:e82174. doi:10.1371/journal.pone.0082174 [doi];PONE-D-13-32693 [pii].
- 362 21. Sun, H., G. Xue, C. Yan, S. Li, L. Cao, Y. Yuan, H. Zhao, Y. Feng, L. Wang, and Z.
 363 Fan. 2013. Multiple-locus variable-number tandem-repeat analysis of *mycoplasma*364 *pneumoniae* clinical specimens and proposal for amendment of MLVA nomenclature.
 365 PLoS One. 8:e64607. doi:10.1371/journal.pone.0064607 [doi];PONE-D-13-08530 [pii].
- 366 22. Xu, Y. C., L. J. Zhu, D. Xu, X. F. Tao, S. X. Li, L. F. Tang, and Z. M. Chen. 2011.
 367 Epidemiological characteristics and meteorological factors of childhood *Mycoplasma*

368

369

pneumoniae pneumonia in Hangzhou. World J Pediatr. 7:240-244. doi:10.1007/s12519-011-0318-0 [doi].

370 23. Xue, G., Q. Wang, C. Yan, N. Jeoffreys, L. Wang, S. Li, G. L. Gilbert, and H. Sun.

2014. Molecular characterizations of PCR-positive *Mycoplasma pneumoniae* specimens
 collected from Australia and China. J Clin.Microbiol **52**:1478-1482. doi:JCM.03366-13

373 [pii];10.1128/JCM.03366-13 [doi].

- Yan, C., H. Sun, G. Xue, H. Zhao, L. Wang, Y. Feng, and S. Li. 2014. A SingleTube Multiple-Locus Variable-Number Tandem-Repeat Analysis of *Mycoplasma pneumoniae* Clinical Specimens by Use of Multiplex PCR-Capillary Electrophoresis. J
 Clin.Microbiol 52:4168-4171. doi:JCM.02178-14 [pii];10.1128/JCM.02178-14 [doi].
- 378 25. Zhao, F., G. Liu, B. Cao, J. Wu, Y. Gu, L. He, F. Meng, L. Zhu, Y. Yin, M. Lv, and

J. Zhang. 2013. Multiple-locus variable-number tandem-repeat analysis of 201
 Mycoplasma pneumoniae isolates from Beijing, China, from 2008 to 2011. J
 Clin.Microbiol 51:636-639. doi:JCM.02567-12 [pii];10.1128/JCM.02567-12 [doi].

26. Zhou, Y., Y. Zhang, Y. Sheng, L. Zhang, Z. Shen, and Z. Chen. 2014. More
complications occur in macrolide-resistant than in macrolide-sensitive *Mycoplasma pneumoniae* pneumonia. Antimicrob.Agents Chemother. 58:1034-1038.
doi:AAC.01806-13 [pii];10.1128/AAC.01806-13 [doi].

- 386
- 387
- 388