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An innovative membrane bioreactor (MBR) system for simultaneous nitrogen and phosphorus removal

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Abstract

1
2 Membrane filtration was integrated with a post-denitrification process to form an innovative
3 membrane bioreactor (MBR) system for effective organic degradation and nutrient (N and P)
4 removal. The system comprised of an aerobic tank, an anoxic tank, an intermediate
5 sedimentation tank, and a membrane filtration tank. The sedimentation tank functioned not
6 only as a rough settler for sludge-water separation before membrane filtration, but also as an
7 anaerobic chamber for P release. While half of the influent flowed into the aerobic tank, the
8 other half was fed into the anoxic tank to favor the proliferation of phosphorus accumulating
9 organisms (PAOs). The experiment was conducted continuously for about 430 days. With a
10 short overall treatment time of less than 10 hrs for municipal wastewater, the MBR-based

11 process could achieve the total organic carbon, total nitrogen, and total phosphorus removals
12 of around 94%, 85%, and 87%, respectively. The growth and activity of PAOs in the MBR
13 system were evidenced by the significant P release in the anaerobic chamber followed by the
14 luxury P uptake in the membrane tank. With the DAPI and PAO_{mix} probe staining, the
15 increases of PAOs and polyhydroxybutyrate (PHB) in sludge during the experiment were
16 well observed under the fluorescent microscope.

17

18 **Keywords:** Biological nutrient removal; Enhanced biological phosphorus removal (EBPR);
19 Fluorescence *in situ* hybridization (FISH); Membrane bioreactor (MBR); Wastewater
20 treatment.

21

22 **1. Introduction**

23 Eutrophication has been recognized as one of the most serious water pollution problems.
24 Wastewater discharge brings nutrients, both nitrogen (N) and phosphorus (P), into the
25 receiving water, causing nutrient enrichment and algal blooms. For effective water
26 environment protection, a number of nutrient removal processes have been developed for N
27 and/or P removal from wastewaters [1]. Biological nitrogen removal involves the combined
28 nitrification by autotrophic bacteria under aerobic conditions and denitrification by
29 heterotrophic bacteria under anoxic conditions, while enhanced biological phosphorus
30 removal (EBPR) is achieved through P luxury uptake by phosphorus accumulating organisms
31 (PAOs) together with sludge discharge [2]. Biological N removal can be achieved by either
32 pre-denitrification or post-denitrification [3]. Compared to the pre-denitrification process,
33 post-denitrification would allow complete N removal [3], although the external carbon source
34 usually needs to be added into the anoxic reactor. More recently, it was reported that
35 simultaneous N and P removal could be achieved by placing an anaerobic tank before the

36 aerobic tank (i.e. post-anoxic type), for which external carbon sources might not be required
37 [4]. Although P removal was primarily driven by stored polyhydroxybutyrate (PHB) and
38 other organic substances, it was apparent that a portion of PAOs was able to utilize nitrate as
39 the electron acceptor [4].

40 Membrane bioreactor (MBR) is an attractive process that has been increasingly used for
41 biological wastewater treatment. With membrane filtration replacing the conventional
42 clarifier, MBR possesses a number of merits such as biomass enrichment, small footprint,
43 ensured sludge-effluent separation, easy manipulation of the hydraulic and sludge retention
44 times (HRT and SRT) and, most importantly, excellent effluent quality with little organic and
45 solid contents [5, 6]. However, an MBR tank without anaerobic and anoxic variations cannot
46 be simply used for nutrient removal in wastewater treatment. There is a need to develop
47 innovative MBR systems that incorporate membrane separation into the biological treatment
48 process for N and P removals.

49 In comparison to organic degradation, biological nutrient removal is more difficult to
50 achieve with the MBR, and P is the most difficult one to remove [1, 7, 8]. Incorporation of
51 membrane filtration into various processes for simultaneous N and P removal has been
52 attempted in the past decade [7, 8]. The system variations reported include sequencing batch
53 reactors [10, 11], the continually-operated pre-oxic system [8, 9, 12, 13], and post-oxic
54 denitrification type [4, 14]. For large-scale applications, the MBR has been used
55 conventionally to replace the final clarifiers for solids-effluent separation in common nutrient
56 removal activated sludge processes (e.g., the anaerobic-anoxic-oxic (A/A/O) process) (Table
57 1). These large-scale MBR systems often adopt a long treatment time or a low filtration rate
58 for wastewater treatment and nutrient removal. Besides, although nutrient removal can be
59 achieved in these systems, it has been found that denitrification sometimes may complicate

60 the EBPR as denitrifiers compete with PAOs for the limited carbon source and hence affect
61 the stability of the system performance [14].

62 Similar to any membrane separation process, membrane fouling in MBR is inevitable. A
63 number of studies has suggested that the membrane fouling rate in MBR correlates well with
64 the sludge concentration [5, 6]. Thus, lowering the sludge concentration in the membrane
65 chamber would help mitigate the fouling problem to some extent. However, the settleability
66 of sludge has not been fully utilized for regulating the sludge concentration before membrane
67 filtration in the MBR-based nutrient removal systems [10, 18]. Moreover, when membrane is
68 used to retrofit or upgrade existing biological wastewater treatment systems, some of the
69 secondary clarifiers would often become obsolete.

70 In the present study, we developed a new system to integrate membrane filtration with
71 the post-denitrification process for simultaneous N and P removal. The membrane chamber
72 was connected to an intermediate sludge settler. As such, the sedimentation tank not only was
73 used as a rough settler for sludge-water separation before membrane filtration but also
74 provided an anaerobic zone to allow the proliferation and function of PAOs for P removal
75 purpose. In addition to the development of an innovative and effective MBR wastewater
76 treatment system for simultaneous N and P removal, PAO abundance and the P concentration
77 profile throughout the system were characterized, and the long-term performance of the
78 system was evaluated.

79

80 **2. Materials and Methods**

81 ***2.1 Experimental set-up***

82 An MBR wastewater treatment system (Fig. 1) was developed and operated for the
83 experimental study on the simultaneous removals of organic pollutants, nitrogen, and
84 phosphorous. The system was a modification of the post-denitrification process that

85 comprised of an aerobic tank, an anoxic tank, an intermediate sludge settler, and a membrane
86 filtration tank. In addition to biomass concentration, the rough sludge settler also provided an
87 anaerobic zone for P release. In the MBR tank, a polyethylene hollow-fiber membrane
88 module (0.4 μm pore size, 0.075 m^2 effective area, Mitsubishi Rayon) was immersed. The
89 working volumes of the aerobic, anoxic, settling, and membrane tanks were 2, 1.4, 1.5, and
90 1.6 L, respectively. Aeration was provided through fine air diffusers from the bottom in the
91 aerobic tank and the membrane filtration tank, while sludge in the anoxic tank was suspended
92 by a paddle mixer at 30 rpm. Half of the feed wastewater was pumped into the aerobic tank
93 and the other half into the anoxic tank. The sludge recirculation ratios from the settling tank
94 to the aerobic tank and from the membrane tank to the anoxic tank ranged from 300 to 400%
95 (Table 2).

96 The feed consisted of 90% synthetic wastewater prepared according to a classic recipe
97 [19] for typical municipal wastewater and 10% actual domestic sewage collected from a local
98 municipal wastewater treatment plant (Stanley Sewage Treatment Plant, Hong Kong). The
99 raw sewage was expected to supply trace elements for the biomass growth. The carbon source
100 in the synthetic wastewater was a mixture of 90% NaAc and 10% glucose, and the N and P
101 sources in the feed were supplied with NH_4Cl and a mixture of KH_2PO_4 and NaH_2PO_4 ,
102 respectively. The variations of the wastewater influent in chemical oxygen demand (COD)
103 and COD:N:P ratio are summarized in Table 2. NaHCO_3 was added to the feed at 50 mg/L or
104 higher to keep the pH between 6.5 and 7.5. Effluent was withdrawn through the membrane by
105 a suction pump (MasterFLEX, Cole-Parmer) that was set off for 2 min (for membrane
106 relaxation) every 10 min. Membrane fouling was indicated by the trans-membrane pressure
107 (TMP) increase, which was monitored with a manometer in mm Hg. The TMP increased
108 gradually with time from an initial value of about 5 mm Hg (0.67 kPa) to around 600 mm Hg
109 (80 kPa) and then, the fouled membrane was washed thoroughly with running tap water to

110 restore its permeability [20].

111

112 **2.2 MBR wastewater treatment experiment**

113 The wastewater treatment experiment was conducted in four phases in the laboratory at
114 room temperature (~25 °C) (Table 2). A short initial phase, termed as Phase 0, was recorded
115 for the original performance of the system in terms of organic and nutrient removals.
116 Afterwards, the system was operated and optimized for more than 400 days for enhanced
117 biological P removal together with N removal. The operation could be divided into three
118 phases according to the P removal performance in relation to the different COD:N:P ratios
119 and/or recirculation ratios (Table 2). From Phase I to Phase II, the P removal efficiency
120 improved significantly. Phase III was operated for a long period to demonstrate the stable
121 operation of the system and its nutrient removal performance. In addition, the organic content
122 or the COD:TN:TP ratio decreased in the influent in Phase III compared to Phase II to
123 increase the difficulty of biological nutrient removals. During this phase, the recirculation
124 ratio was increased as an adjustment to maintain a high nutrient removal efficiency (Table 2).
125 The treatment performance was evaluated in terms of the removal efficiencies of the total
126 organic carbon (TOC), total N (TN), and total P (TP) as well as the P concentration profile in
127 the liquid phase of the sludge suspensions through different tanks.

128

129 **2.3 Microscopic examination of PAOs**

130 The composition and spatial distribution of the microbial community of the sludge in
131 relation to PAO-based P removal were examined under a fluorescent microscope after
132 staining. DAPI (4',6'-diamidino-2-phenylindole) staining on all cells was performed
133 following the method described by Kawaharasaki et al [21] using a filtered DAPI solution (10
134 mg/mL in 25 mM Tris-HCl buffered saline, pH 7.0). For a sludge sample taken from the

135 aerobic tank, it was homogenized briefly using a vortex mixer (Maxi Mix II, Thermolyne) to
136 break up large flocs. The dispersed biomass was then air-dried on a slide and stained with the
137 DAPI solution. After 10 min of staining, the slide was washed using a phosphate buffer saline
138 (PBS) solution and then air-dried at room temperature. The sample was examined under a
139 fluorescent microscope (Eclipse, Nikon) with a 100 W high-pressure mercury lamp and a
140 filter set MWU (Olympus Optical, excitation 330-385 nm). The DAPI-DNA fluorescence
141 appeared to be blue white, while the fluorescence of either DAPI-poly-P or DAPI-lipid was
142 bright yellow.

143 Fluorescence *in situ* hybridization (FISH) technique was also employed to characterize
144 PAO in the sludge following the method described by Fu et al [22]. A sludge sample was first
145 homogenized briefly using the vortex mixer and then placed in a hybridization well on a
146 gelatin-coated microscopic slide plate. The PAO_{mix} probes (comprising the equal amount of
147 probes PAO₄₆₂, PAO₆₅₁, and PAO₈₄₆ (TechDragon, Hong Kong)) were used to target PAO
148 species. Meanwhile, EUB338 was used to target all eubacteria in the sludge sample. The
149 sample after staining was examined under an epifluorescent confocal laser scanning
150 microscope (CLSM) (LSM Pascal, Zeiss, Thornwood). The combined use of DAPI staining
151 and FISH also would display the abundance of PHB in the sludge [23].

152

153 **2.4 Analytical methods**

154 The influent and effluent of the MBR system were sampled twice a week for
155 determination of the overall treatment performance in terms of the removals of the organic
156 (TOC), total nitrogen (NH₄⁺-N and NO₃⁻-N), and total phosphorus (PO₄³⁻-P). In addition, the
157 sludge suspension was also sampled twice a week from each of the tanks and chambers for
158 detail analysis. For the sludge samples, the suspensions were filtered through a 0.4- μ m
159 polycarbonate membrane (25 mm, Osmonics) and the filtrates were analyzed. The TOC

160 concentration was determined by a TOC analyzer (IL550 TOC-TN Analyzers, Lachat) using
161 the combustion-infrared method. Ammonia nitrogen ($\text{NH}_4^+\text{-N}$) was analyzed using the
162 electrochemical method with an ammonia electrode and a potentiometer (920A, ORION).
163 Nitrate ($\text{NO}_3^-\text{-N}$) was analyzed by an UV/VIS spectrophotometer (Lambda 25, Perkin Elmer)
164 according to the Standard Methods [24]. The liquid-phase P concentration in the sludge
165 suspension was analyzed using a spectrophotometer (UV/VIS Lambda 12, Perkin Elmer) in
166 accordance to the Standard Methods [24]. The dissolved oxygen (DO) concentration in each
167 chamber was determined by a DO probe (97-08-99, Orion) with an electrometer (920A,
168 Orion). Mixed liquor suspended solids (MLSS) for the sludge concentration and COD for the
169 organic concentration were measured following the Standard Methods [24].

170

171 **3. Results and Discussion**

172 **3.1 *Organic and nitrogen removal***

173 The MBR wastewater treatment system was operated for over 430 days under various
174 conditions, including variations in food-to-microorganism (F/M) ratio, COD:TN:TP, and
175 internal recirculation ratio, as summarized in Table 2. The MBR system performed
176 consistently well in organic degradation and nitrification. In detail, the TOC removal
177 efficiency was more than 94% throughout the experiment (Fig. 2), even that a volumetric
178 organic loading rate up to 720 mg COD/L-d. The effluent contained a low organic
179 concentration with a TOC below 5 mg/L, which was not affected significantly by the
180 variations in operation. The organic content in the supernatant of the anoxic tank was
181 constantly below 25 mg TOC/L. Thus, it is deduced that the major portion of the influent
182 organic was degraded in the aerobic and anoxic tanks, with an average removal efficiency of
183 around 85%. The additional organic removal was attributable to the step of membrane
184 filtration, due to its effective retention of organic solutes [25, 26]. Moreover, the membrane

185 interception helped keep a relatively high biomass concentration in the MBR system, which
186 made the system less sensitive to the changes in operation [27, 28]. Meanwhile, the effluent
187 after membrane filtration was of high quality with a SS concentration of less than 1 mg/L.

188 The average NH_4^+ -N removal efficiency of the MBR system was about 92%, indicating
189 sufficient biological nitrification (Fig. 2). The aerobic zone followed by the anoxic zone
190 formed a post-denitrification process for TN removal, and feeding the substrate into the
191 anoxic tank was proven to effectively facilitate denitrification. An increase of the nitrogen
192 loading rate or change of the COD/TN ratio did not significantly affect the NH_4^+ -N removal
193 efficiency. At a high volumetric nitrogen loading rate from 70 to 80 mg N/L-d, the average
194 TN removal efficiencies were 79.2 ± 4.3 , 78.8 ± 3.4 , 83.7 ± 3.0 , and $73.2\pm 1.3\%$ for Phases 0, I, II,
195 and III, respectively. However, the C:N ratio in the wastewater influent was important to the
196 denitrification efficiency. As the C:N ratio decreased from 54:5 to 46.5:5 in Phase III, the
197 effluent TN content increased considerably, due likely to the insufficient carbon source in the
198 influent for denitrification. The recirculation ratio appeared to be another important factor to
199 the TN removal result [7, 14, 29]. As NO_3^- -N counted for around 90% of the TN residue in
200 the MBR effluent, the TN removal efficiency by denitrification could be further improved if a
201 higher recirculation ratio was adopted.

202

203 **3.2 *Biologically-enhanced phosphorus removal***

204 The TP removal was no more than 40% during the start-up phase, or Phase 0 (Fig. 3),
205 which also suggested that The P removal via assimilation was clearly below 40%. The P
206 concentration in the supernatant of sludge at the outlet of the anaerobic tank (i.e. the rough
207 settler, $\text{DO} < 0.05$ mg/L) was not significantly higher than that in the aerobic MBR suspension.
208 This implies that the PAO community was not well developed in the system. For the EBPR
209 process, P release by PAOs under anaerobic condition is crucial to the luxury P uptake in the

210 subsequent aerobic stage [7, 30, 31]. Thus, conditions that favor PAO growth and anaerobic
211 phosphorus release should be provided. As such, a longer anaerobic period (to ~200 min) and
212 a shorter SRT (to ~10 d) were adopted from day 31 (i.e. Phase I). The anaerobic P release was
213 evidently improved gradually in the intermediate sedimentation chamber, and the TP removal
214 efficiency increased to nearly 50% with an average effluent TP concentration of about 5.2
215 mg/L. In order to enhance PAO accumulation further, the COD/TP ratio was increased to
216 more than 50 in Phase II. During this period, the TP removal efficiency increased gradually to
217 over 75% and the effluent TP concentration decreased eventually to 2.2 mg/L. The
218 phenomena of P release in the anaerobic phase (intermediate sedimentation chamber) and P
219 uptake in the aerobic phase (MBR tank) could be well observed. By the end of Phase II at a
220 volumetric phosphorus loading rate of 15 mg P/L-d, a fairly low effluent TP concentration of
221 less than 2.0 mg/L was attained and the average P removal efficiency reached to 87.1% (Fig.
222 3).

223 In Phase III, the organic content (COD concentration) in the wastewater influent was
224 reduced by more than 10%, resulting in lower COD/TN and COD/TP ratios (Table 2).
225 However, with a higher recirculation ratio applied ($4Q_{in}+4Q_{in}$), the TP removal efficiency
226 increased to over 88% with an average effluent TP concentration of around 0.8 mg/L.
227 Although decreased slightly, the TN removal rate was still around 80% (Fig. 2). The
228 experimental results showed that the MBR system developed in this study is highly effective
229 for sufficient organic degradation and simultaneous nutrient (N and P) removal. In this regard,
230 this MBR system with a short treatment time ($HRT < 10$ hr) and a high pollutant loading rate
231 (720 mg COD/L-d, 77 mg N/L-d and 15.4 mg P/L-d) is comparable to or more effective than
232 the MBR systems reported by others [11, 32] for simultaneous N and P removal. Moreover,
233 the long-term experimental operation also evidenced the stable performance of the
234 MBR-based treatment process. During the stable operation in Phase III, the specific nutrient

235 removal rates could be estimated for the sludge in different reactors or tanks, and the results
236 summarized in [Table 3](#) include the specific nitrification, denitrification, P release and P
237 uptake rates.

238 Phosphate release in the anaerobic zone is crucial to the PAO function for P-removal,
239 and its performance is affected considerably by the form of the carbon source [7, 31]. In the
240 anaerobic chamber (intermediate settler), acetate in the influent could be readily uptaken by
241 PAO cells, leading to the activation of acetyl-CoA. Two molecules of acetyl-CoA could
242 condense to form acetoacetyl-CoA, which would be transformed eventually to
243 poly- β -hydroxybutyrate (PHB) [7, 33]. Subsequently, in the aerobic stage with a rather low
244 organic concentration, PAOs could use PHB as the carbon and energy sources to grow and to
245 assimilate P forming poly-P [31, 32]. The configuration and operation of the wastewater
246 treatment process shown in [Fig. 1](#) was apparently favorable to the growth of PAOs in the
247 MBR system. In addition, a sufficient residence time of the sludge in the anaerobic chamber
248 would ensure the conversion of readily degradable substrates to PHB, which is essential to
249 the luxury P uptake in the aerobic stage [34, 35].

250 Phosphorus was then removed in the MBR via luxury P uptake and sludge discharge. The
251 intermediate settler functioned not only for sludge-water separation, but also as an anaerobic
252 chamber for P release. As half of the influent was fed into the anoxic tank, the organic carbon
253 would induce P release in the intermediate sludge settler. Subsequently, luxury P uptake took
254 place in the MBR to achieve P removal from the liquid phase. The P contents in the liquid
255 phases of the sludge mixture in different tanks were analyzed when a high TP removal
256 efficiency was maintained in Phase III ([Fig. 4](#)). With an influent P concentration of 8 mg/L,
257 the liquid-phase P concentrations in the aerobic tank, rough settler, and membrane chamber
258 were 6.7, 21.5, and 1.5 mg/L, respectively. In view of the sludge flow from the rough settler
259 to the aerobic tank and membrane chamber, it becomes apparent that anaerobic P release and

260 aerobic luxury P uptake occurred well in the system. The liquid-phase P contents showed the
261 characteristic P distribution profile of the EBPR process. The membrane filtration also played
262 a role in polishing the effluent for P removal, as indicated by a lower TP concentration in the
263 effluent (0.8 mg/L) than that in the MBR sludge supernatant (1.5 mg/L). Comparing Phase III
264 with Phase 0 for TP removal, it can be estimated that more than 56% of phosphorus (112 mg
265 P/d) was removed by the luxury P uptake of the sludge, whilst the other less than 40 mg P/d
266 was probably utilized for biomass growth.

267

268 **3.3 PAOs and their role in phosphorus removal**

269 PAO accumulation and function are crucial to the P removal capability of the
270 MBR-based EBPR system. The PAO growth in sludge during the experiment was examined
271 by microscopic observations. The aerobic sludge flocs in different test phases (0, I, II, and III)
272 were stained by DAPI and the specific PAO probes (PAO_{mix}) [11, 31, 34]. After DAPI
273 staining, PAOs appeared as bright blue spots and polyhydroxybutyrate (PHB) appeared to be
274 yellow dots. As for the FISH images of the sludge labeled with the PAO probe, PAOs
275 appeared to be yellow clusters, as the probe would bind coccobacilli [35]. In the start-up
276 phase, the activated sludge (Fig. 5a₁ and a₂) showed only a little PAO and PHB signals. With
277 the development of the system for P removal, the PAO abundance in the sludge flocs
278 increased gradually. For the sludge in Phase I during which the MBR exhibited a moderate P
279 removal, the PAO and PHB contents increased considerably (Fig. 5b₁ and b₂).

280 In relation to the improvement of P removal in Phases II and III, the clusters of PAO
281 probes binding with the distinct and uniform yellow cells can be well observed (Fig. 5c₂ and
282 d₂). Moreover, large PHB granules were displayed by the bright yellow spots (Fig. 5c₁ and d₁),
283 which confirmed further the PAO activities. The apparent PAO content and PHB density
284 correlated well with the P removal performance of the EBPR system. For example, compared

285 to the early sludge samples, the sludge flocs in Phase III had more compact and larger PAO
286 clusters. It is believed that the massive PAO clustering signified the great P release and
287 uptake activities of the sludge under the anaerobic and aerobic conditions, respectively [2,
288 34].

289

290 **3.4 Sludge concentration profile**

291 The membrane fouling in MBR is caused mainly by sludge deposition on the membrane
292 surface [6, 20, 26]. Previous studies have shown that the membrane fouling rate in MBR is
293 affected to a certain extent by the bulk sludge concentration [20, 36, 37]. It would be
294 beneficial to the membrane fouling reduction if the sludge concentration in the membrane
295 chamber could be maintained at a low level (e.g. < 6 g MLSS/L) [28]. The present MBR
296 system employed a rough settler for sludge-water separation before the membrane chamber.
297 This would therefore help lower the sludge concentration in the membrane chamber while
298 increase the MLSS contents in other tanks to favor biological treatment activities. During the
299 operation of the MBR system for over 400 days, the average sludge concentration in the
300 membrane chamber (3 g MLSS/L) was found to be even lower than that in the aerobic tank
301 (3.6 g/L) (Fig. 6). If the rough settler was not employed, the membrane chamber would have
302 the highest sludge concentration in comparison to other tanks, which would likely lead to
303 more rapid membrane fouling [6, 36, 37]. During the stable operation in Phase III, the
304 overflow from the rough settler into the MBR had a sludge concentration (MLSS) of around
305 1.9 mg/L. This was considerably lower than the sludge concentration (~3.0 g/L) in the
306 upstream bioreactors. Because of the membrane separation, the effluent was free of solids and
307 sludge in the MBR tank was concentrated to about 2.4 g/L. The sludge mixture was
308 recirculated at a rate of $4Q_{in}$ ($Q_{in}=16.6$ L/d). Meanwhile, for a stable operation and effective
309 P removal, the sludge mixture was discharged from the MBR at a rate of about 0.7 L/d.

310

311 **4. Conclusions**

- 312 • An innovative MBR system was developed for effective organic degradation and nutrient
313 removal in wastewater treatment. The system integrated membrane filtration with the
314 post-denitrification activated sludge process, and the wastewater influent was split into
315 the aerobic tank and the anoxic tank. A rough settler was used not only for sludge-water
316 separation before membrane filtration but also as an anaerobic chamber for P release.
- 317 • The MBR-based system performed remarkably well for simultaneous organic, N and P
318 removals. With a short treatment time of less than 10 hrs, the system could achieve stable
319 TOC, TN, and TP removal efficiencies of around 94%, 85%, and 87%, with effluent
320 concentrations of less than 5, 6, and 1 mg/L, respectively.
- 321 • The proliferation and activity of PAOs in the MBR system were indicated by the
322 significant P release in the anaerobic chamber and the luxury P uptake in the aerobic
323 membrane chamber. PAOs and their P accumulating function were evidenced by the large
324 PAO clusters and PHB granules under the fluorescent microscope after DAPI and PAOM
325 probe staining.

326

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333

334 **References**

- 335 [1] Mino T, van Loosdrecht MCM, Heijnen JJ. Microbiology and biochemistry of the
336 enhanced biological phosphate removal process. *Water Res* 1998; 32: 3193–207.
- 337 [2] Oehmen A, Zeng RJ, Yuan ZG, Keller J. Anaerobic metabolism of propionate by
338 polyphosphate-accumulating organisms in enhanced biological phosphorus removal
339 systems. *Biotechnol Bioeng* 2005; 91: 43–53.
- 340 [3] Metcalf & Eddy. *Wastewater Engineering: Treatment and Reuse*, 4th ed. McGraw-Hill,
341 Boston, Massachusetts, USA; 2003.
- 342 [4] Coats ER, Mockos A, Loge FJ. Post-anoxic denitrification driven by PHA and glycogen
343 within enhanced biological phosphorus removal. *Biores Technol* 2011; 102: 1019–27.
- 344 [5] Judd S. *The MBR Book: Principles and Applications of Membrane Bioreactors in Water
345 and Wastewater Treatment*. Amsterdam, The Netherlands: Elsevier; 2006.
- 346 [6] Meng FG, Chae SR, Drew A, Kraume M, Shin HS, Yang FL. Recent advances in
347 membrane bioreactors (MBRs): Membrane fouling and membrane material. *Water Res*
348 2009; 43: 1489–512.
- 349 [7] Oehmen A, Lemos PC, Carvalho G, Yuan Z, Keller J, Blackall LL, Reis MAM. Advances
350 in enhanced biological phosphorus removal: From micro to macro scale. *Water Res* 2007;
351 41: 2271–300.
- 352 [8] Monclús H, Sipma J, Ferrero G, Comas J, Rodriguez-Rod I. Optimization of biological
353 nutrient removal in a pilot plant UCT-MBR treating municipal wastewater during
354 start-up. *Desalination* 2010; 250: 592–7.
- 355 [9] Adam C, Gnirss R, Lesjean B, Buisson H, Kraume M. Enhanced biological phosphorus
356 removal in membrane bioreactors. *Water Sci Technol* 2002 ; 46: 281–6.
- 357 [10] Zhang HM, Xiao JN, Cheng YJ, Liu LF, Zhang XW, Yang FL. Comparison between a
358 sequencing batch membrane bioreactor and a conventional membrane bioreactor.
359 *Process Biochem* 2006; 41: 87–95.

- 360 [11] Yang S, Yang F, Fu Z, Wang T, Lei R. Simultaneous nitrogen and phosphorus removal
361 by a novel sequencing batch moving bed membrane bioreactor for wastewater treatment.
362 J Hazard Mater 2010; 175: 551–7.
- 363 [12] Wang Y, Huang X, Yuan Q. Nitrogen and carbon removals from food processing
364 wastewater by an anoxic/aerobic membrane bioreactor. Process Biochem 2005; 40:
365 1733–39.
- 366 [13] Shen J, He R, Han W, Sun X, Li J, Wang L. Biological denitrification of high-nitrate
367 wastewater in a modified anoxic/oxic-membrane bioreactor (A/O-MBR). J Hazard
368 Mater 2009; 172: 595–600.
- 369 [14] Vocks M, Adama C, Lesjean B, Gnirss R, Kraume M. Enhanced post-denitrification
370 without addition of an external carbon source in membrane bioreactors. Water Res 2005;
371 39: 3360–68.
- 372 [15] Hu Y, Wang XC, Zhang Y, Li Y, Chen H, Jin P. Characteristics of an A2O–MBR system
373 for reclaimed water production under constant flux at low TMP. J Membr Sci 2013; 431:
374 156–162.
- 375 [16] Gómez-Silván C, Arévalo J, Pérez J, González-López J, Rodelas B. Linking hydrolytic
376 activities to variables influencing a submerged membrane bioreactor (MBR) treating
377 urban wastewater under real operating conditions. Water Res 2013; 47: 66–78.
- 378 [17] Shen YX, Xiao K, Liang P, Sun JY, Sai SJ, Huang X. Characterization of soluble
379 microbial products in 10 large-scale membrane bioreactors for municipal wastewater
380 treatment in China. J Membr Sci 2012; 415–416: 336–345.
- 381 [18] Ivanovic I, Leiknes T. Impact of aeration rates on particle colloidal fraction in the
382 biofilm membrane bioreactor (BF-MBR). Desalination 2008; 231: 182–90.
- 383 [19] AEESP. Environmental Engineering Process Laboratory Manual. Champaign, IL, USA:
384 Association of Environmental Engineering and Science Professors; 2001.

- 385 [20] Chu HP, Li XY. Membrane fouling in a membrane bioreactor (SMBR): Sludge cake
386 formation and fouling characteristics. *Biotechnol Bioeng* 2005; 90: 323–31.
- 387 [21] Kawaharasaki M, Tanaka H, Kanagawa T, Nakamura K. In situ identification of
388 polyphosphate-accumulating bacteria in activated sludge by dual staining with
389 rRNA-targeted oligonucleotide probes and 4',6-diamidino-2-phenylindol (DAPI) at a
390 polyphosphate-probing concentration. *Water Res.* 1999; 33: 257–65.
- 391 [22] Fu Z, Yang F, An Y, Xue Y. Simultaneous nitrification and denitrification coupled with
392 phosphorus removal in a modified anoxic/oxic-membrane bioreactor (A/O–MBR).
393 *Biochem Eng J* 2009; 43: 191–6.
- 394 [23] Tsai CS, Liu WT. Phylogenetic and physiological diversity of tetrad-forming organisms
395 in deteriorated biological phosphorus removal systems. *Water Sci Technol* 2002; 46(1–2):
396 179–84.
- 397 [24] APHA–AWWA–WEF. *Standard Methods for the Examination of Water and Wastewater*,
398 20th ed. Washington, DC, USA: American Public Health Association/American Water
399 Works Association/Water Environment Federation; 1998.
- 400 [25] Liang S, Liu C, Song L. Soluble microbial products in membrane bioreactor operation:
401 behaviors, characteristics, and fouling potential. *Water Res* 2007; 41: 95–101.
- 402 [26] Wang XM, Li XY. Accumulation of biopolymer clusters in a submerged membrane
403 bioreactor and its effect on membrane fouling. *Water Res* 2008; 42: 855–62.
- 404 [27] Chae SR, Shin HS. Characteristics of simultaneous organic and nutrient removal in a
405 pilot-scale vertical submerged membrane bioreactor (VSMBR) treating municipal
406 wastewater at various temperatures. *Process Biochem* 2007; 42: 193–8.
- 407 [28] Monti A, Hall ER, Dawson RN, Husain H, Kelly H. Comparative study of biological
408 nutrient removal (BNR) processes with sedimentation and membrane-based separation.
409 *Biotechnol Bioeng* 2006; 96: 740–52.

- 410 [29] Rosenberger S, Evenblij H, Poele ST, Wintgens T, Laabs C. The importance of liquid
411 phase analyses to understand fouling in membrane assisted activated sludge
412 processes-six case studies of different European research groups. *J Membr Sci* 2005;
413 263: 113–26.
- 414 [30] Wachtmeister A, Kuba T, van Loosdrecht MCM, Heijnen JJ. A sludge characterization
415 assay for aerobic and denitrifying phosphorus removing sludge. *Wat Res* 1997; 31:
416 471–478.
- 417 [31] Liu Y, Zhang T, Fang HHP. Microbial community analysis and performance of a
418 phosphate-removing activated sludge. *Biores Technol* 2005; 96: 1205–14.
- 419 [32] Shin JH, Lee SM, Jung J.Y. Enhanced COD and nitrogen removals for the treatment of
420 swine wastewater by combining submerged membrane bioreactor (MBR) and anaerobic
421 upflow bed filter (AUBF) reactor. *Process Biochem* 2005; 40: 3769–76.
- 422 [33] Ersu CB, Ong SK, Arslankaya E, Lee YW. Impact of solids residence time on biological
423 nutrient removal performance of membrane bioreactor. *Water Res* 2010; 44: 3192–202.
- 424 [34] Seviour RJ, Mino T, Onuki M. The microbiology of biological phosphorus removal in
425 activated sludge systems. *FEMS Microbiol Rev* 2003; 27: 99–127.
- 426 [35] Lee N, Janssen JL, Aspegren H, Henze M, Nielsen PH, Wagner M. Population dynamics
427 in wastewater treatment plants with enhanced biological phosphorus removal operated
428 with and without nitrogen removal. *Water Sci Technol* 2002; 46: 163–70.
- 429 [36] Li XY, Wang XM. Modelling of membrane fouling in a submerged membrane bioreactor.
430 *J Membr Sci* 2006; 278: 151–61.
- 431 [37] Meng F, Zhang H, Yang F, Zhang S, Li Y, Zhang X. Identification of activated sludge
432 properties affecting membrane fouling in submerged membrane bioreactors. *Sep Purif*
433 *Technol* 2006; 51: 95–103.

Table 1. Summary of a few large-scale MBR processes for biological nutrient removals in wastewater treatment

Process	Capacity (m ³ /d)	Membrane material	Operating conditions			Effluent quality			Membrane cleaning cycle	Features and limitations	Ref.
			HRT (h)	SRT (d)	MLSS in MBR (g/L)	COD (mg/L)	TN (mg/L)	TP (mg/L)			
A ₁ -A ₂ -O-MBR	2000	0.1 μm PVDF	13.8	50	5.4~8.2	14.2±5.8	8.6±3.4	0.3±0.2	1-2 weeks	High quality reclaimed water; but a low filtration flux.	[15]
Post-denitrification +MBR	200	PVDF	38	20	4	20.1±14.8	24.6±3.6	n.a.	weekly	Stable operation; but a long HRT.	[16]
A ₁ -A ₂ -O-MBR	60,000	0.04 μm, PVDF	17.3	20	7.00~8.50	15~20	15~25	0.30~0.50	n.a.	Stable pollutant removal; but a long HRT and low filtration flux.	
A ₁ -A ₂ -A ₂ -O-MBR	50,000	0.1 μm, PVDF	12~13	25~32	5.65~10.44	8~53	4.20~22.70	0.03~1.45	n.a.	Treatment of both municipal and industrial wastewater; but less stable pollutant removal.	[17]
A ₁ -A ₂ -O-A ₂ -MBR	20,000	0.4 μm PVDF	33~38	27~37	7.07~11.59	3~36	3.02~13.10	0.03~0.67	n.a.	Treatment of both municipal and industrial wastewater; but a long HRT.	
A ₁ -A ₂ -O-MBR	20,000	0.04 μm PVDF	10~14	14	3.24~12.00	8~91	3.42~18.66	0.05~0.89	n.a.	High resistance to the seasonal shocking; but low filtration flux.	

Note: A₁ - anaerobic; A₂ - anoxic; O - oxic (aerobic); n.a.- not available.

Table 2. The experimental condition in different phases for the MBR nutrient removal study.

Phase	Duration (d)	SRT (d)	HRT (hr)	COD (mg/L)	TOC (mg/L)	F/M (g COD/g MLSS-d)	TN (mg/L)	COD:TN:TP	Recirculation rate
0	30	> 12	11.6	320	120	~0.32	40	32:4:1	3Q _{in} + 3Q _{in}
I	83	~ 10	11.6	320	120	~0.22	40	32:4:1	3Q _{in} + 3Q _{in}
II	153	~ 10	9.3	432	160	~0.26	40	54:5:1	3Q _{in} + 3Q _{in}
III	165	~ 10	9.3	372	140	~0.28	40	46.5:5:1	4Q _{in} + 4Q _{in}

Table 3. Specific biological N and P removal rates estimated for the sludge in different reactors during the stable operation in Phase III.

Specific nutrient removal rate	Aerobic tank	Anoxic tank + sludge settler	MBR
Volume (L)	2.0	1.4	1.5
HRT (hr)	2.9	2.0	2.1
MLSS (g/L)	3.0	3.0	3.0
Nitrification (mg N/g MLSS-d)	45		70
Denitrification (mg N/g MLSS-d)		100	
P release (mg P/g MLSS-d)			180
P uptake (mg P/g MLSS-d)			230

Figure captions

- Fig. 1. Schematic diagram of the new MBR system for organic degradation and simultaneous N and P removal.
- Fig. 2. Organic and total nitrogen removal performance of the MBR system (OM: organic matter measured by mg TOC/L).
- Fig. 3. Total phosphorus removal in different phases (Phase 0, I, II, and III) of the MBR experiment.
- Fig. 4. Change of the phosphate concentration in the liquid phase of the sludge in different tanks of the MBR system during Phase III operation.
- Fig. 5. Microscopic images of the PAOs in the MBR sludge after (1) DAPI staining (bright blue spots - live bacteria stained by DAPI; yellow dots - PHB stained by DAPI) and (2) FISH-CLSM observation (green spots - bacteria stained by EUB338; yellow area - PAO stained by PAO_{mix}): (a) seed sludge in Phase 0, (b) sludge flocs in Phase I, (c) sludge flocs in Phase II, and (d) sludge flocs in Phase III.
- Fig. 6. The MLSS concentrations of the sludge in the aerobic tank and MBR chamber.

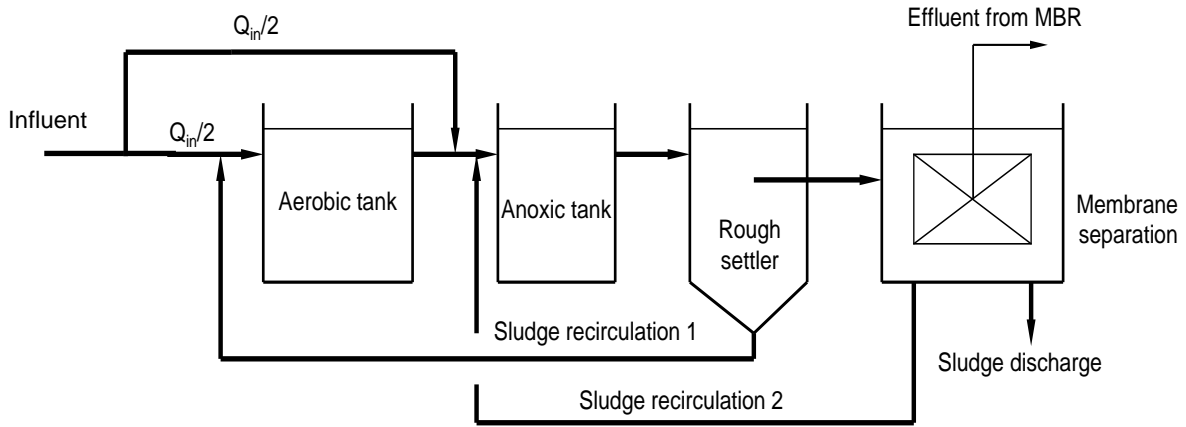


Fig. 1. Schematic diagram of the new MBR system for organic degradation and simultaneous N and P removal.

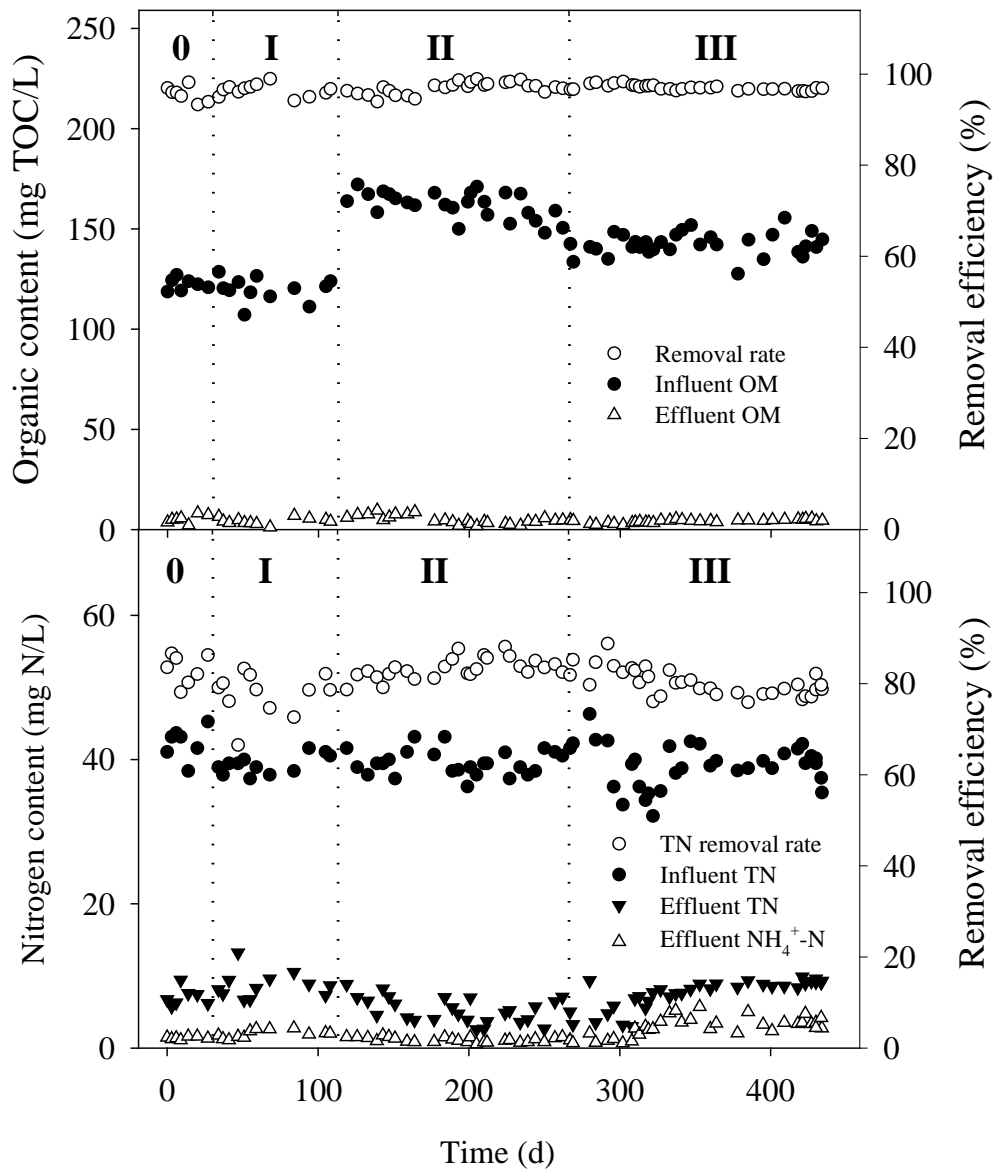


Fig. 2. Organic and total nitrogen removal performance of the MBR system (OM: organic matter measured by mg TOC/L).

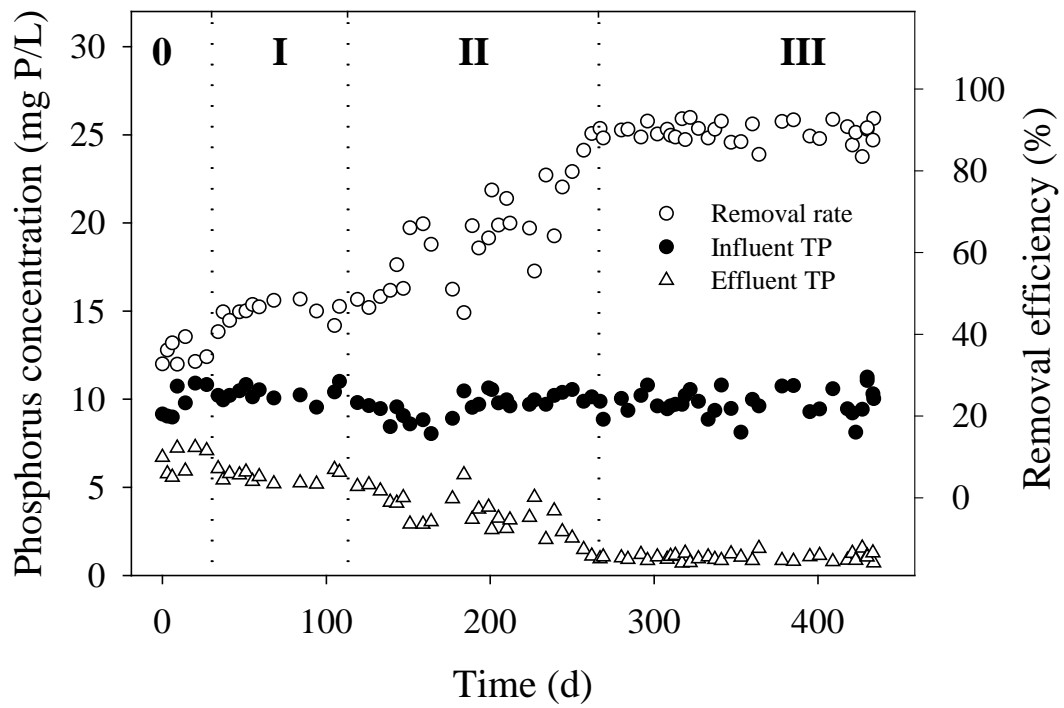


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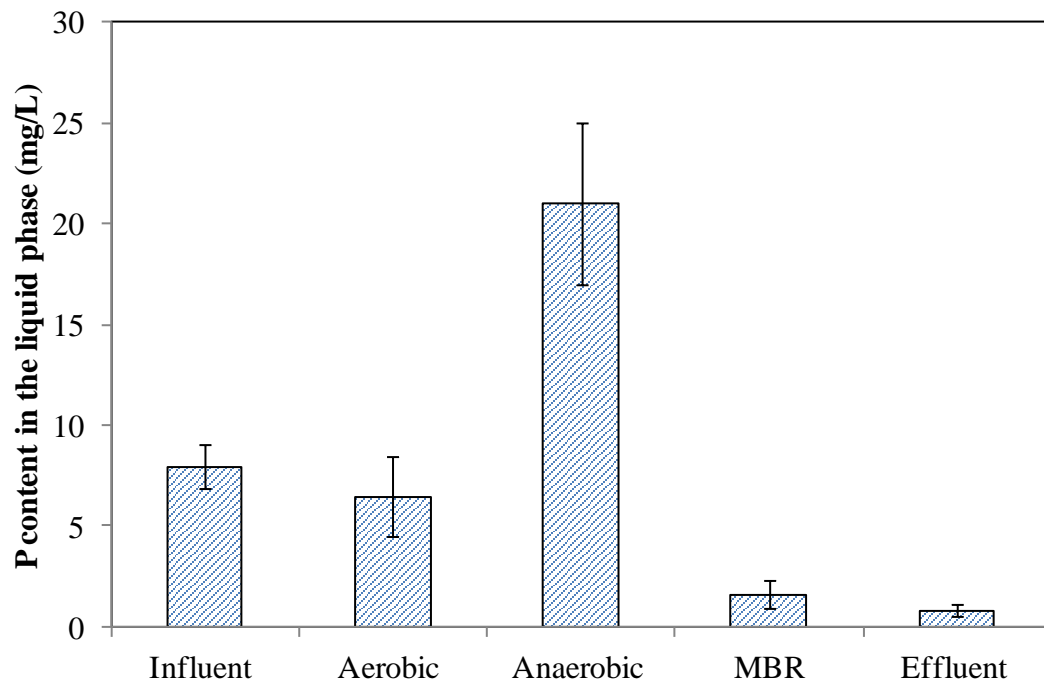


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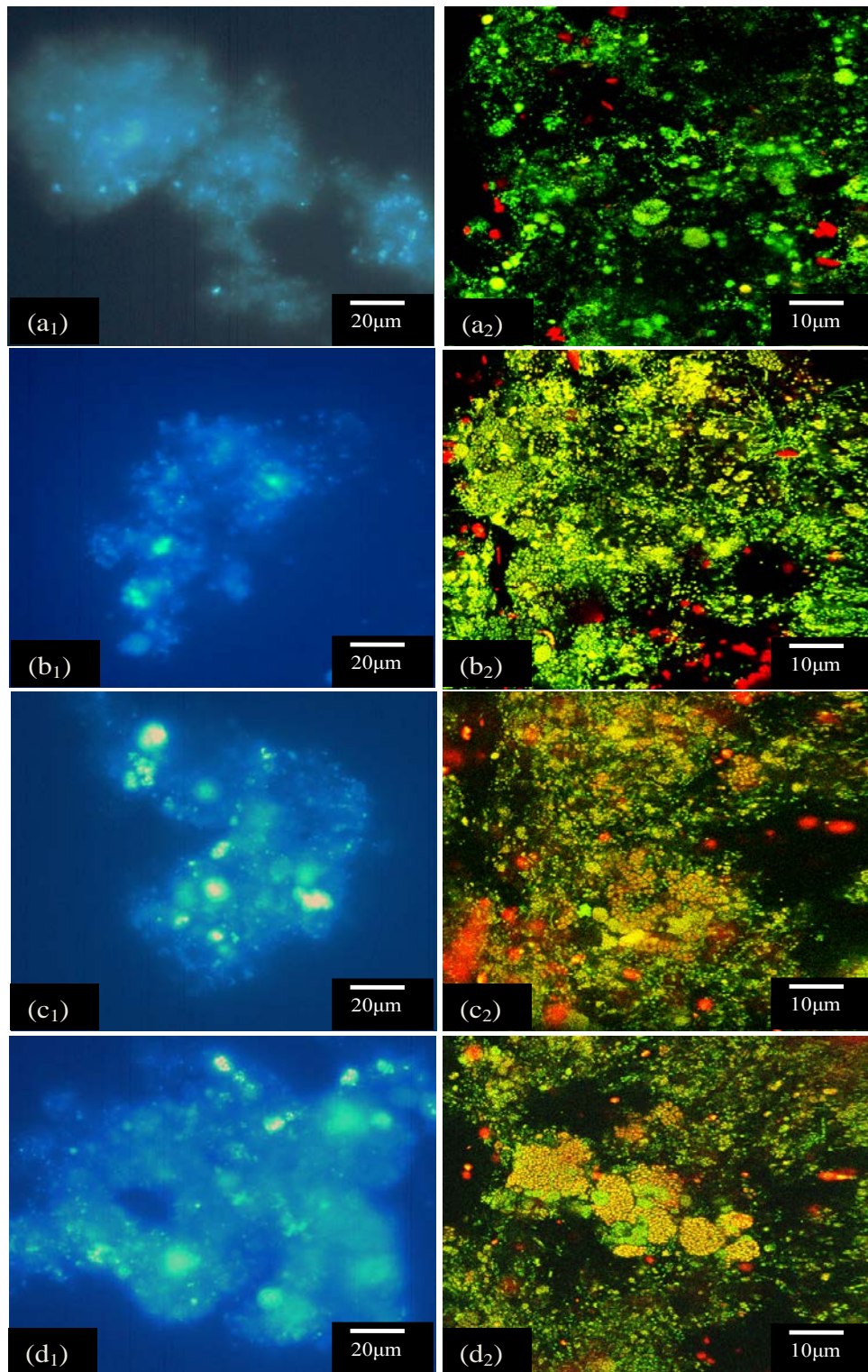


Fig. 5. Microscopic images of the PAOs in the MBR sludge after (1) DAPI staining (bright blue spots - live bacteria stained by DAPI; yellow dots - PHB stained by DAPI) and (2) FISH-CLSM observation (green spots - bacteria stained by EUB338; yellow area - PAO stained by PAO_{mix}): (a) seed sludge in Phase 0, (b) sludge flocs in Phase I, (c) sludge flocs in Phase II, and (d) sludge flocs in Phase III.

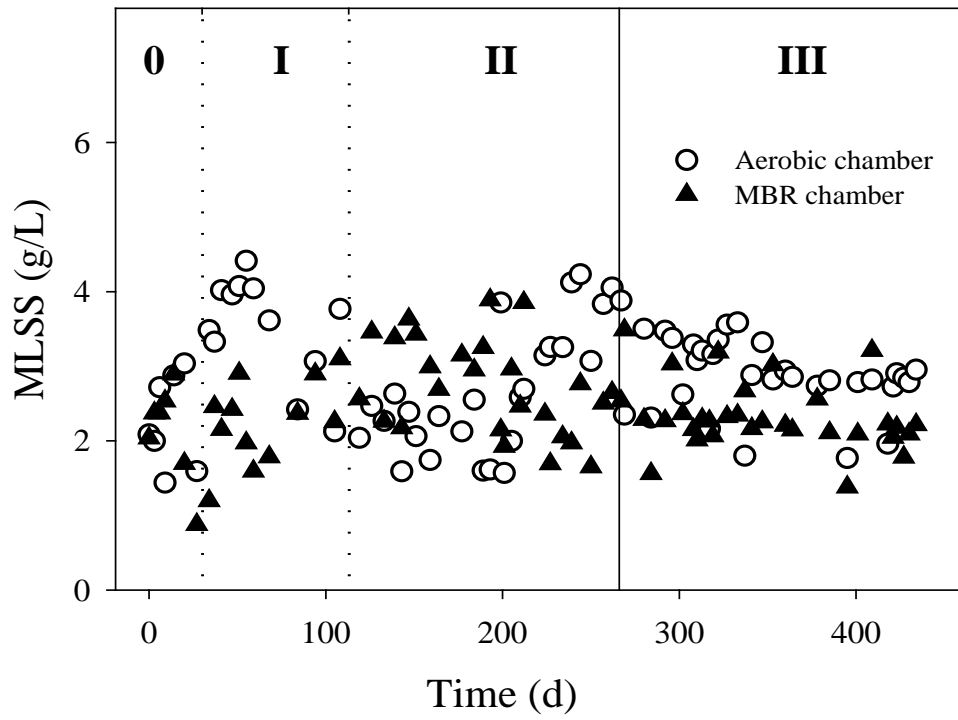


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