



Title	HIV-1 envelope glycoprotein variable loops are indispensable for envelope structural integrity and virus entry
Author(s)	Yuan, T; Li, J; Zhang, M
Citation	PLoS One, 2013, v. 8 n. 8, p. Article number 69789
Issued Date	2013
URL	http://hdl.handle.net/10722/185534
Rights	Creative Commons: Attribution 3.0 Hong Kong License

HIV-1 Envelope Glycoprotein Variable Loops Are Indispensable for Envelope Structural Integrity and Virus Entry

Tingting Yuan, Jingjing Li, Mei-Yun Zhang*

AIDS Institute, Department of Microbiology, Li Ka Shing Faculty of Medicine, The University of Hong Kong, Hong Kong, China

Abstract

HIV-1 envelope (Env) glycoprotein is a trimer of heterodimer of gp120 and gp41, and derives from a trimeric glycoprotein precursor, gp160. Gp120 contains five conserved regions that are interspersed with 5 variable loop regions (V1–V5). Env variations in variable loop length and amino acid composition may associate with virus pathogenesis, virus sensitivity to neutralizing antibodies (nAbs) and disease progression. To investigate the role of each variable loop in Env function, we generated a panel of JRFL gp160 loop deletion mutants and examined the effects of each loop deletion on Env expression, Env cell surface display and Env-mediated virus entry into permissive cells. We found that deletion of V1 and V2 ($\Delta V1V2$), or loop D (ΔlpD) abolished virus entry, the same effect as deletion of V3 ($\Delta V3$), while deletion of V3 crown ($\Delta V3C$) significantly enhanced virus assembly and entry. We further found that deletion of V4 ($\Delta V4$) or V5 ($\Delta V5$), or replacement of V4 or V5 with flexible linkers of the same lengths knocked out the receptor and coreceptor binding sites in gp120, but significantly enhanced the exposure of the N-trimer structure and the membrane proximal external region (MPER) in gp41. Although deletion of V4 or V5 did not affect Env expression, they negatively affected Env cell surface display, leading to the failure in virus assembly and subsequent entry. Taken together, we found that Env variable loops were indispensable for Env structural integrity and virus entry. Our findings may have implications for development of HIV-1 vaccine immunogens and therapeutics.

Citation: Yuan T, Li J, Zhang M-Y (2013) HIV-1 Envelope Glycoprotein Variable Loops Are Indispensable for Envelope Structural Integrity and Virus Entry. *PLoS ONE* 8(8): e69789. doi:10.1371/journal.pone.0069789

Editor: Jean-Pierre Vartanian, Institut Pasteur, France

Received: April 16, 2013; **Accepted:** June 17, 2013; **Published:** August 1, 2013

Copyright: © 2013 Yuan et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by Hong Kong Research Grants Councils (RGC) General Research Fund (GRF) (#785210 and #785112) and China 12th 5-year Mega project for HIV/AIDS (#2012ZX10001006) to MYZ. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: zhangmy@hku.hk

Introduction

HIV-1 envelope glycoprotein (Env) is a trimer of heterodimer of the mature surface glycoprotein gp120 and the transmembrane glycoprotein gp41. Env trimer mediates virus entry into permissive cells upon binding to the receptor (CD4) and coreceptor (CCR5 or CXCR4). Env gp120 contains five variable loops (V1–V5) that are interspersed with five sequence conserved regions (C1–5). Despite sequence and length variations, Env variable loops are important determinants or indicators for coreceptor tropism, virus sensitivity to neutralization by antibodies, virus pathogenesis, and disease progression [1,2,3,4,5,6,7,8,9]. It is well studied that V3 is the primary determinant of viral coreceptor usage although mutations outside the V3 have impact on coreceptor tropism of subtype B viruses [3]. V1V2 region may influence coreceptor binding and participate in shielding of neutralization-sensitive regions of the Env. V1V2 length and potential N-linked glycosylation sites (PNGS) were found to increase significantly through chronic infection before declining in late-stage infection [1], and an increase in the V1V2 length and/or the number of PNGS in the V1V2 region directly contributed to viral resistance to HIV-specific neutralizing antibodies (nAbs) [4,10]. Important structural motifs formed by the C3 and V4 regions and the epitopes within the motifs were also found to be major targets of the early

autologous neutralizing response in HIV-1 subtype C infection [7]. Extensive intra-patient V4 variability in length and number of PNGS has also been observed in clade B, G, and CRF02 isolates during early infection [2]. In addition, some variable loops constitute neutralizing determinants recognized by several known broadly neutralizing HIV-1 human monoclonal antibodies (bnmAbs). For example, bnmAb VRC01 binds to an epitope formed by the CD4 binding site (CD4bs), V5 and loop D (lpD) [11], and bnmAbs PG9/16, CH04 and PGT145 are V1/V2-directed antibodies [12], while a short β -strand segment of the V3 loop is involved in binding of PGT127 and 128 to Env trimer [13]. These observations suggest potentially important roles of variable loops in Env-mediated virus entry and virus pathogenesis. However, how changes in these loops affect Env function are not well studied.

In this study, we generated a series of loop deletion or replacement mutants of JRFL gp160, and investigated the effects of each loop deletion or replacement on Env expression, Env cell surface display, and virus assembly and subsequent virus entry into permissive cells. In addition to the five variable loops, we also investigated the importance of two other loops, the CD4 binding loop (CD4bl) and lpD, for Env-mediated viral function. We further investigated if deletion of Env cytoplasmic tail (CT) can

rescue the defects in Env expression and function caused by variable loop deletions.

Materials and Methods

Cells, Plasmids, Medium, Antibiotics, and Antibodies

93 T cells were purchased from ATCC. TZM-bl cell line was obtained from the NIH AIDS Research and Reference Reagent Program (ARRRP). The pSVIII-JRFL gp160 wild type (WT) and pcTAT plasmids were kindly provided by Yuxing Li, Richard Wyatt and Joseph Sodroski [14]. DMEM medium, fetal bovine serum (FBS) and penicillin-streptomycin (pen-strep) were purchased from Gibco. Purified gp120-specific polyclonal Abs D7324 were purchased from Aalto BioReagents. IgG1s b12 and 2G12 were obtained from ARRRP. IgG1 VRC01 was kindly provided by Xueling Wu and John Mascola. The rest of HIV-1 mAbs were produced in our laboratory by transient transfection of 293F cells (Invitrogen) followed by Protein A (GE Healthcare) affinity purification.

Construction of Loop Deletion Mutants

Full-length JRFL gp160 gene in pSVIII was used as a template for generation of various loop deletion or replacement mutants, including single loop deletion of V2 ($\Delta V2$), V3 ($\Delta V3$), V4 ($\Delta V4$), V5 ($\Delta V5$), loop D (ΔlpD), CD4 binding loop ($\Delta CD4bl$), and deletion of V2 crown ($\Delta V2C$) and V3 crown ($\Delta V3C$), and double loop deletion of V1 and V2 ($\Delta V1V2$), and loop D and V5 ($\Delta lpD\Delta V5$), and replacement of V4 ($\Delta V4fl$) and V5 ($\Delta V5fl$) with flexible linkers of the same lengths (Table 1). A panel of primers were designed and synthesized for amplification of different fragments of JRFL gp160 gene and for replacement of each loop with either a short flexible linker or a flexible linker of the same length as the original loop (in the case of V4 and V5) (Table 1 and 2). Primer pSV3for was paired with each anti-sense primer to amplify the upstream fragments of the Env, and primer pSV3rev was paired with each sense primer for amplification of the corresponding downstream fragments using the following PCR program: an initial denaturation at 94°C for 5 min, followed by 10 cycles of 95°C for 20 s, 50°C for 30 s and 72°C for 90 s, and 20 cycles of 95°C for 20 s, 55°C for 30 s and 72°C for 90 s, and a final extension at 72°C for 10 min. The upstream and the corresponding downstream fragments for each mutant were assembled by splice overlap extension (SOE) as follow: an initial denaturation at 94°C for 5 min, followed by 5 cycles of 95°C for 20 s, 55°C for 30 s and 72°C for 90 s. The assembled full-length gp160 loop deletion mutant genes were then amplified by PCR using pSV3for and pSV3rev as a pair of primers and the following PCR program: 20 cycles of 95°C for 20 s, 55°C for 30 s and 72°C for 3 min, and a final extension at 72°C for 10 min. All PCR products were gel-purified using QIAquick gel extraction kit (Qiagen), digested with KpnI and BamHI, and ligated to pSV3 plasmid digested with the same restriction enzymes. Ligation products were used to transform TG1 electroporation competent cells. Each loop deletion or replacement mutant was confirmed by DNA sequencing.

Transient Transfection of 293 T Cells and Flow Cytometry

The day before transfection, 293 T cells were seeded in DMEM growth medium (DMEM containing 10% FBS and 1% pen-strep). 70–80% confluent 293 T cells were co-transfected with pSVIII expression plasmid containing JRFL wild type (WT) gene or its loop deletion or replacement mutants, and pcTAT using PEI as a transfection reagent in DMEM medium containing 10% FBS. 4–6 h post transfection, the medium was changed to DMEM growth

medium. 48 h post transfection, 293 T cells were harvested, washed with FACS buffer (PBS+5% FBS) and stained with IgG1 2G12, or VRC01, or b12 as a primary antibody at 4°C for 1 h. Following washing three times with FACS buffer, cells were incubated with phycoerythrin (PE) conjugated to anti-human Fc (Sigma) (1:200) at 4°C for 45 min. Cells were then washed again three times with FACS buffer and analyzed on a BD flow cytometer. The results were analyzed by FlowJo software.

Generation of Env-pseudotyped Viruses

Env-pseudotyped viruses were assembled by co-transfection of 293 T cells with Env plasmid containing JRFL Env WT gene or its loop deletion or replacement mutants with or without the CT, and HIV-1 backbone plasmid pNL4-3 containing a luciferase reporter gene and HIV-1 structural genes. 48 h post transfection, the culture supernatants containing assembled pseudovirus were harvested and stored at -80°C , or directly used in virus titration by capture ELISA and subsequent virus entry assay.

Pseudovirus Entry Assay

Crude preparation of pseudovirus in culture supernatant was 3-fold serially diluted in DMEM growth medium and 100 μl of each dilution in triplicate were placed in 96-well flat-bottom cell culture plates. 100 μl of TZM-bl cell suspension in DMEM growth medium containing DEAE-Dextran (25 $\mu\text{g}/\text{ml}$) were dispensed to the wells to a final cell density of 10,000 cells per well and a final concentration of DEAE-Dextran of 10 $\mu\text{g}/\text{ml}$. The plates were then incubated at 37°C with 5% CO₂ for 48 h. Cells were washed with PBS once and lysed with 60 μl per well lysis buffer by incubating at RT for 30 min followed by addition of 25 μl of luciferin (Promega). Luminescence readings (RU) were determined by PE Victor3 luminometer. Relative pseudovirus entered into the cells was calculated as follow: $(\text{RU}_{\text{sample}} - \text{RU}_{\text{blank}}) / (\text{RU}_{\text{WT}} - \text{RU}_{\text{blank}}) \times 100$.

Capture ELISA

250 ng per well of D7324 in PBS were coated on high-binding 96-well half-well microplates by incubating the plates at 4°C overnight. Following blocking the wells with 2.5% skim milk in PBS (MPBS), 3-fold serially diluted whole cell lysate or pseudovirus-containing culture supernatant in lysis buffer (2% Triton X-100 from Sigma in PBS) were added to the ELISA plates. Captured JRFL gp160s were detected by using IgG1 2G12 as a primary antibody and HRP conjugated to anti-human F(ab')₂ (1: 2,500) as a secondary antibody, and ABTS (Roche) as substrate. The optical density at 405 nm (OD_{405nm}) was determined after color development at RT for 20 min. A similar capture ELISA set-up was used to determine structural integrity of loop deleted or replaced JRFL gp160 by measuring the binding of a panel of HIV-1-specific mAbs to each loop deleted or replaced Env mutant and WT captured on microplates. HIV-1-specific mAbs include CD4bs mAbs VRC01 and b12, CD4-induced (CD4i) mAbs X5 and 17b, and gp41-specific mAbs m47 (N-trimer-specific, unpublished), 2F5 and 4E10. Glycan-specific mAb 2G12 was included in the assay to measure each loop deleted or replaced Env mutant expressed in 293 T cells. A standard curve used for data calibration was generated using recombinant JRFLgp120 as antigen and 2G12 as a primary antibody in the same capture ELISA.

Immunohistochemistry (IHC) Assay

48 h post transfection, 293 T cells were fixed using 4% paraformaldehyde fixative solution by incubation at RT for

Table 1. JRFL gp160 loop deletion or replacement mutants.

JRFL Env loops	Original loop sequence	Replacing linker sequence and designated name of the construct
V1V2	VNATNTTNDSEGMTMERGEIKNCSFNITTSIRDEVQKEYALFYKLDVVPIDNNNTSYRL	GSGSG (Δ V1V2)
V2	NITTSIRDEVQKEYALFYKLDVVPIDNNNTSYRL	GSGSG (Δ V2)
V2 crown	<u>FYKLD</u>	AAAAA (Δ V2C)
loop D	NFTNNAKT	AAAA (Δ lpD)
V3	NTRKSIHIGPGRAFYTTGEIIG	GSGSG (Δ V3)
V3 crown	<u>GPGR</u>	AAAA (Δ V3C)
CD4bl	SSGGDPEIVMH	GSGSG (Δ CD4bl)
V4	NSTQLFNSTWNNTEGSNNTEGNTITLP	GSGSG (Δ V4); or GGGGSGGGGSGGGSGSGSG (Δ V4fl)
V5	INENGTEIFR	GSGSG (Δ V5); or GGGGSGSGSG (Δ V5fl)

The amino acid (AA) sequences of the original loops and flexible linkers for replacement are shown. V2 and V3 crown sequences are underlined. CD4bl: CD4 binding loop. Δ V4fl and Δ V5fl: V4 and V5 loops replaced with flexible linkers of the same lengths. Designated names of resultant constructs are indicated in parentheses. doi:10.1371/journal.pone.0069789.t001

Table 2. Primers used to construct JRFL gp160 loop deletion mutants.

Primer Name	Primer Sequence (5' to 3')	Use
pSV3for	ACCATGCTCCTGGGATGTTGATG	annealing to plasmid pSVIII
pSV3rev	TCTCAAGCGGTGGTAGCTGAAGAG	annealing to plasmid pSVIII
delLpDF	GACGCTGCAGCAGCTATAATAGTACAGCTGAAAGAATC	generating Δ lpD
delLpDR	AGCTGCTGCAGCGTCAGATCTAATTACTACCTC	
Delv1v2F	GGTAGCGGATCAGGTATAAGTTGTGACACCTCAGTC	generating Δ V1V2
Delv1v2R	ACCTGATCCGCTACCCTCTGCAATTTAAAGTAAC	
V2crownF	GCTGCTGCAGCTGCTGTAGTACCAATAGATAATAATAATACC	generating Δ V2C
V2crownR	AGCAGCTGCAGCAGCAAGAGCATATTCTTTCTGCAC	
delfullV2F	GGTTCAGGATCTGGCATAAGTTGTGACACCTCAGTC	generating Δ V2
delfullV2R	GCCAGATCCTGAACCGAAAGAGCAGTTTTTTATTCTC	
V3crownF	TAGCAGCAGCTGCAGCATTATACTACAGGAG	generating Δ V3C
V3crownR	CTGCAGCTGCTGTATATGTATACTTTTTCTTG	
Delfullv3F	GGTTCTGGATCAGGTGATATAAGACAAGCACATTGTAAC	generating Δ V3
Delfullv3R	ACCTGATCCAGAACCCTGTTGGGTCTGTACAATTAATTC	
V4delFnew	GGCTCTGGTCTGGGTGCGAATAAAACAATATAAAACATG	generating Δ V4 with a short linker
V4delRnew	CCCAGAACCAGAGCCACAGTAGAAAAATTCTCCTCCAC	
V5delFnew	GGTAGCGGATCAGGCCCTGGAGGAGGAGATATGAG	generating Δ V5 with a short linker
V5delRnew	GCCTGATCCGCTACCACCACCATCTCTGTTAATAG	
DelCD4bIF	GGTCTGGATCAGGTAGTTTTAATGTGGAGGAGAATTTTTTC	generating Δ CD4bl
DelCD4bIR	ACCTGATCCAGAACCCTGATTAAGACTATTGTTTTATTCTC	
V4FFL	GGTGGAGGCGTTTCAGGCGGAGGTGGCTCTGGCGGTGGCGGATCA GGCTCTGGTCTGGG	generating Δ V4 with a linker of the same length
V4RFL	CCGCTGAACCGCTCCACCACAGTAGAAAAATTCTCCTCCAC	
V5FFL	GGTGGAGGCGTTTCAGGTAGCGGATCAGGCCCTGGAGG	generating Δ V5 with a linker of the same length
V5RFL	CTACCTGAACCGCTCCACCACCACCATCTCTGTTAATAG	
JRFLCTfor	CCTGGATGGAGTGGGAAAGAG	generating Δ CT
JRFLCTrev	CTCTTCCCACCTCCATCCAGG	

doi:10.1371/journal.pone.0069789.t002

20 min. If necessary, cells were permeabilized using 0.2% Triton X-100 by incubation at RT for 30 min. Cells were then incubated with IgG1 2G12 in 2% MPBS at 37°C for 1 h, washed two times with PBS, and incubated with FITC conjugated to anti-human Fc (Jackson ImmunoResearch) at 37°C for 1 h. Following washing two times with PBS, cells were permeabilized using 0.2% Triton X-100 by incubation at RT for 30 min, and incubated with Hoechst 33258 (Invitrogen) at RT for 2 min to stain the nucleus.

Results

Effects of Variable Loop Deletions on Env Cell Surface Display

We co-transfected 293T cells with pSVIII encoding JRFL gp160 wild type (WT) or loop deletion mutants, and pCtAT encoding HIV-1 Tat. 48 h post transfection, we did flow cytometry to analyze cell surface display of Env gp160 WT and the mutants using mAb 2G12 as a primary antibody and PE-anti-human IgG, F(ab)² as a secondary antibody. We found that $\Delta V4$, $\Delta V5$, and $\Delta lpD\Delta V5$ almost abolished Env cell surface display (Fig. 1A, Table 3). Other loop deletion mutants also decreased their cell surface display compared to the WT, including $\Delta V2$, $\Delta V2C$, $\Delta V3$, $\Delta V3C$, ΔlpD , and $\Delta V1V2$ (Fig. 1A, Table 3).

To determine whether CT deletion can rescue the defects in cell surface display of Env loop deletion mutants, we deleted the CT in JRFL gp160 WT and each loop deletion mutant, and generated another panel of loop deletion mutants in combination with CT deletion (ΔCT) (Table 3). A CD4 binding loop ($\Delta CD4bl$) deletion mutant in combination with ΔCT ($\Delta CD4bl\Delta CT$) was included in this panel of mutants and used as a control in the subsequent pseudovirus assembly and entry assay. We found that ΔCT alone significantly enhanced Env expression and cell surface display (Fig. 1B), which is consistent with the result previously reported by others. But deletion of CT did not rescue the defects of $\Delta V4$ and $\Delta V5$ Envs in cell surface display (Fig. 1B, Table 3). The other loop deletion mutants in combination with ΔCT had increased levels of cell surface display compared to the corresponding loop deletion alone mutants. The rescue was significant for ΔlpD mutant.

To determine whether decreased cell surface display of $\Delta V4$ and $\Delta V5$ Envs was caused by decreased transfection efficiency or decreased Env expression, we measured the transfection efficiency by IHC assay and determined the total $\Delta V4$ and $\Delta V5$ Env expression in the whole cell lysates of transfected 293T cells by capture ELISA using 2G12 as primary antibody. The percentages of positive cells transfected with $\Delta V4$ and $\Delta V5$ Env plasmids, as well as the rest of loop deletion mutant plasmids showed no significant difference compared to the WT (Fig. 2A, and data not

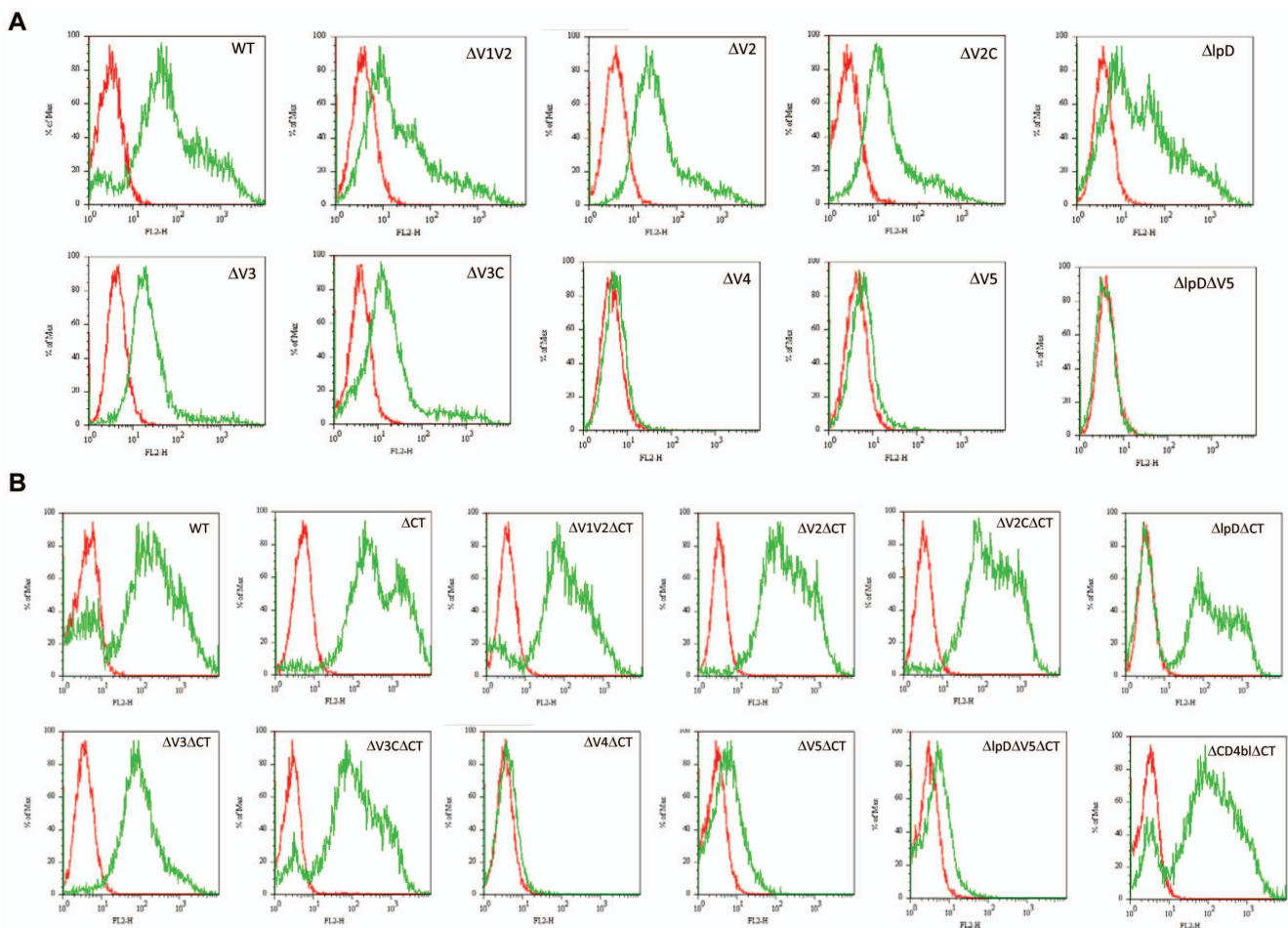


Figure 1. Effects of various loop deletions on Env cell surface display. A: Flow cytometry of 293T cells co-transfected with various loop deletion Env plasmids and pCtAT; B: Flow cytometry of 293 T cells co-transfected with various loop deletions and CT deletion plasmids and pCtAT. Cells were incubated with HIV-1 mAb 2G12 and bound 2G12 measured by FITC-anti-human IgG, F(ab)². doi:10.1371/journal.pone.0069789.g001

Table 3. Effects of loop deletions with or without the CT deletion on JRFL gp160 cell surface display, virus assembly, and subsequent virus entry.

ENV variants (JRFL)	Mean value in flow cytometry	Relative Env cell surface display (%)	Relative pseudovirus assembled (%)	Relative pseudovirus entry (%)
WT	312	100	100	100
JRFLΔV1V2	178	57	43	0
JRFLΔV2	148	48	442	31
JRFLΔV2C	159	51	258	35
JRFLΔlpD	87	28	137	0
JRFLΔV3	118	38	313	0
JRFLΔV3C	101	32	1,828	343
JRFLΔV4	9	3	1	NT
JRFLΔV5	9	3	6	NT
JRFLΔCT	457	147	2,259	6,375
JRFLΔV1V2ΔCT	255	82	55	0
JRFLΔV2ΔCT	217	70	1,733	91
JRFLΔV2CΔCT	437	140	1,095	253
JRFLΔlpDΔCT	428	137	612	0
JRFLΔV3ΔCT	236	76	1,822	0
JRFLΔV3CΔCT	320	103	5,669	1,195
JRFLΔV4ΔCT	6	2	1	NT
JRFLΔV5ΔCT	9	3	2	NT
JRFLΔlpDV5ΔCT	6	2	4	NT
JRFLΔCD4blΔCT	322	103	5,106	0

Cell surface displayed Env proteins were measured by flow cytometry. Mean values are shown and cell surface displayed Env proteins relative to the WT are calculated as a percentage. The amount of assembled pseudovirus in culture supernatant was measured by capture ELISA. Three-fold serially diluted culture supernatants with a starting volume of 50 μ l were tested in luciferase assay. Relative pseudovirus entry was defined as a percentage of luminescence reading (response units, RU) of each mutant virus relative to the WT when same amount of assembled pseudovirus was used. In this case, the same amount of pseudovirus gave OD405nm of 0.95 in the capture ELISA (Fig. 2C). NT: not tested.

doi:10.1371/journal.pone.0069789.t003

shown), suggesting that loop deletion mutant plasmids can transfect 293T cells as efficient as the WT plasmid. Capture ELISA result showed that $\Delta V4$ and $\Delta V5$ Envs were well expressed in the cells albeit to a decreased optical density (OD) at 405nm (Fig. 2B), which may attribute to the loss of PNGS in the V4 and V5, leading to decreased binding to 2G12. The rest of loop deletion mutants expressed well in 293T cells and bound well to 2G12 (Fig. 2B).

Effects of Variable Loop Deletions on Virus Assembly and Entry into Permissive Cells

The defects of $\Delta V4$ and $\Delta V5$ Envs in cell surface display led to the failure in pseudovirus assembly and subsequent virus entry (Fig. 2C, Table 3). Assembled pseudovirus was undetectable in the culture supernatant of 293T cells co-transfected with pSVIII- $\Delta V4$, or - $\Delta V5$, or - $\Delta lpDV5$, and pcTAT and pNL4-3 backbone plasmids. Interestingly, we found that $\Delta V1V2$ also affected virus assembly (Fig. 2C), and abolished virus entry (Table 3). More interestingly, ΔlpD did not affect virus assembly (Fig. 2C), but the assembled pseudovirus was not able to enter the permissive cells (Table 3). Both $\Delta V2$ and $\Delta V2C$ enhanced virus assembly, but negatively affected virus entry. As expected, $\Delta V3$ abolished virus entry although the amount of assembled pseudovirus increased. To our surprise, $\Delta V3C$ led to almost 20-fold increase in virus assembly and 3-fold increase in virus entry compared to the WT (Table 3). Deletion of CT in the WT Env significantly enhanced pseudovirus assembly and subsequent entry into the cells, which is

consistent with the observation previously reported by others. But deletion of CT did not rescue $\Delta V1V2$ and ΔlpD viruses to enter the cells although $\Delta lpD\Delta CT$ had 5-fold increase in pseudovirus assembly compared to ΔlpD alone (Table 3). Interestingly, we found that ΔCT significantly enhanced assembly of $\Delta V2$, $\Delta V3$, $\Delta V2C$, and $\Delta V3C$ pseudoviruses (3-6-fold increases), and enhanced the entry of $\Delta V2C$ and $\Delta V3C$ pseudoviruses into the cells (Table 3). Combination of $\Delta CD4bl$ with ΔCT ($\Delta CD4bl\Delta CT$) also significantly enhanced pseudovirus assembly compared to the WT (50-fold increase) although the assembled pseudoviruses could not enter the cells, as expected (Table 3).

Deletion of V4 or V5 did not Cause gp120 Shedding

To investigate the possibility that $\Delta V4$ and $\Delta V5$ may enhance gp120 shedding, resulting in lack of Env cell surface display, we did a capture ELISA to detect soluble gp120 that may be present in the culture supernatant of 293T cells co-transfected with recombinant pSVIII plasmid encoding JRFL gp160 WT, or $\Delta V4$, or $\Delta V5$, and pcTAT plasmid. IgG1 2G12 was used as a primary antibody in the capture ELISA. The result showed that gp120 was absent in the culture supernatant of 293T cells co-transfected with the pSVIII gp160 $\Delta V4$ or $\Delta V5$ mutant plasmid, and pcTAT, while gp120 was present in the culture supernatant of 293T cells co-transfected with Env WT plasmid and pcTAT (data not shown). This result indicates that undetectable $\Delta V4$ and $\Delta V5$ Envs on cell surface may be attributed to the lack of Env cell surface display, not to gp120 shedding.

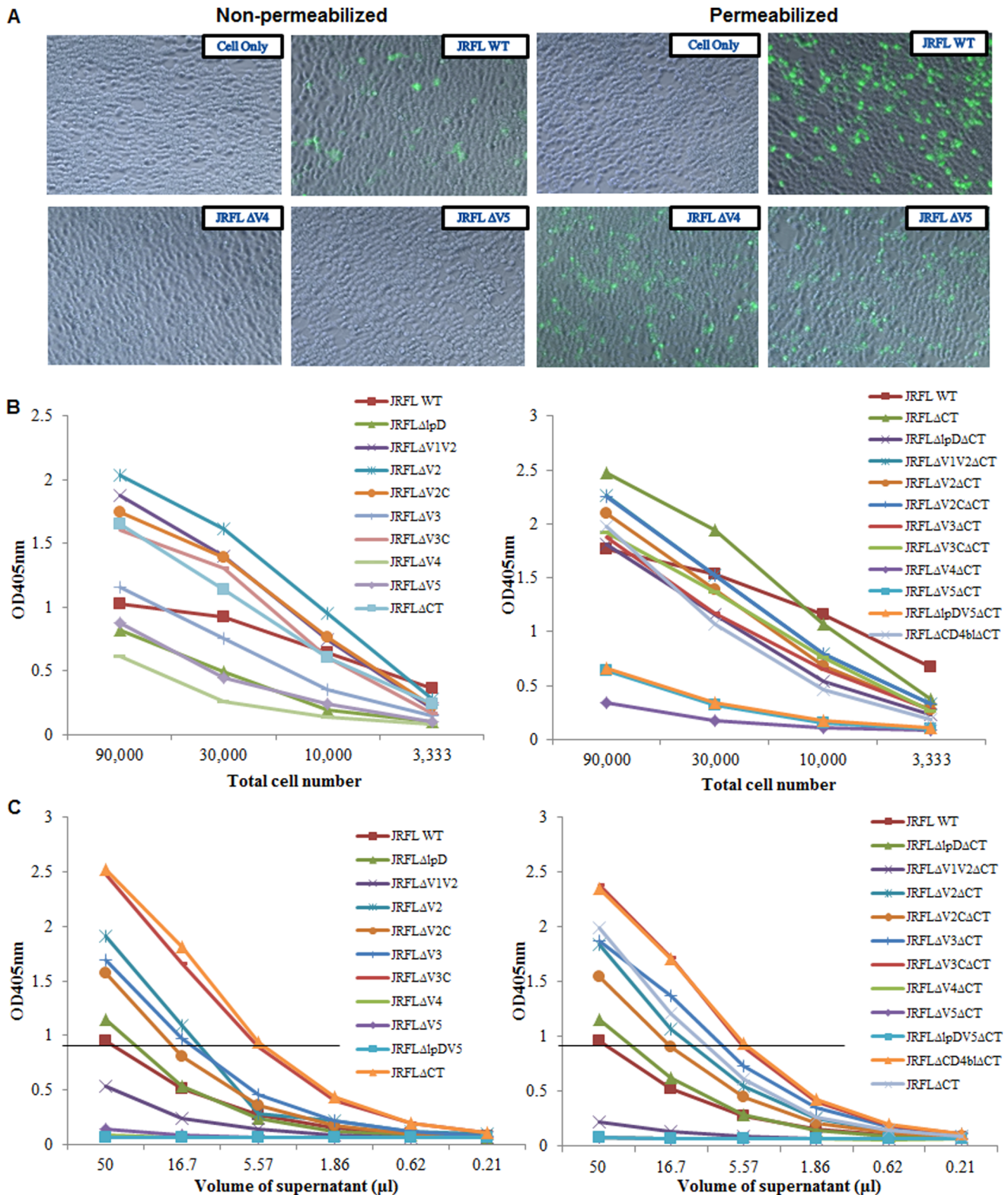


Figure 2. Effects of various loop deletions on total Env expression in 293T cells, Env binding to mAb 2G12, and pseudovirus assembly. A: Immunostaining of 293T cells co-transfected with ΔV4 or ΔV5 Env plasmid and pCTAT with or without permeabilization prior to staining with mAb 2G12; B: Binding of Env loop deletion mutant proteins in the whole cell lysates to 2G12 by capture ELISA; C: Titration of pseudovirus in the culture supernatants by capture ELISA. The volume of each supernatant that contains the same amount of pseudovirus is indicated with a flat line when OD405nm = 0.95.

doi:10.1371/journal.pone.0069789.g002

Effects of V4 and V5 Loop Deletions on Env Structural Integrity

Since $\Delta V4$ and $\Delta V5$ Env proteins can be expressed in cells, but cannot be displayed on cell surface, we examined the possible conformational changes occurred to the Env proteins by measuring their bindings to various mAbs by ELISA. We found that $\Delta V4$ and $\Delta V5$ Env proteins lost the binding to CD4bs mAbs b12 and VRC01, and CD4i mAbs X5 and 17b, but had increased binding to gp41-specific mAbs m47 (N-trimer-specific)(unpublished), 2F5 and 4E10 (MPER-specific) (Fig. 3A-G). The result indicates that deletion of V4 or V5 may destroy Env structural integrity, resulting in loss of the receptor and coreceptor binding sites, and enhanced exposure of the N-trimer structure and the MPER. Binding of glycan-specific mAb 2G12 to $\Delta V4$ and $\Delta V5$ Env proteins decreased, to some extent, which may be attributed to the decreased number of PNGS in the Envs due to the deletion of V4 and V5 (Fig. 3H).

Effects of V4 or V5 Replacement with a Flexible Linker of the Same Length on Env Expression and Function

To investigate if V4 and V5 length, or amino acid composition, or both contribute to the importance of V4 and V5 for Env cell surface display and virus assembly, we generated two loop replacement mutants, designated $\Delta V4fl$ and $\Delta V5fl$, in which V4 or V5 was replaced with a flexible linker of the same length as the original loop (Table 1). Flow cytometry and ELISA results showed that there were no significant differences between $\Delta V4$ and $\Delta V4fl$, or between $\Delta V5$ and $\Delta V5fl$ in Env cell surface display (Fig. 4A and 4B), and in total Env protein expression in the cells (Fig. 4C and 4D), suggesting that amino acid composition of V4 and V5 may play a more important role than the loop length in preserving Env structural integrity and Env function.

Discussion

Engineering Env is one of approaches for HIV-1 vaccine development. Understanding the importance of variable loops for Env-mediated viral functions may help develop Env-based vaccine immunogens and therapeutics against HIV-1 infection. Env CD4 binding loop and V3 are known to be involved in binding to the receptor and coreceptor during virus entry. In this study, we found that V1V2 and loop D were also critical for Env-mediated virus entry into permissive cells. Therefore, V1V2 and loop D could be key components of vaccine immunogens. Indeed, follow-up studies on the recent RV144 trial demonstrate the importance of V1V2 loop for inducing protective antibodies against HIV-1 infection [15]. Furthermore, V1V2 and loop D constitute partially the neutralizing epitopes of bnmAbs PG9/16 and VRC01, respectively. The critical roles of V1V2 and loop D in virus entry may also be explored for developing entry inhibitors targeting these regions. We did sequence alignment and found that six out of eight amino acids in loop D were highly conserved in all HIV-1 subtype viruses and the rest two relatively conserved (data not shown). Targeting conserved loop D may be an alternative to current approaches that target the CD4bs and/or glycans on Env trimer. We further found that V4 and V5 were indispensable for Env structural integrity, but deletion of V4 or V5 enhanced the exposure of the N-trimer structure and the MPER of Env. Several MPER-specific bnmAbs have been identified, including 2F5, 4E10, Z13e1 and 10e8 [16,17], and many MPER-based peptides have been designed, but they failed to induce the same or similar bnAbs. Our $\Delta V4$ and $\Delta V5$ Env proteins may be tested in animals (rabbits or rhesus macaques) for inducing MPER-specific bnAbs. Engineered Env with enhanced exposure of the MPER may help induce MPER-specific nAbs in vivo.

Recently revealed cryo-EM structure of JRFL Env trimer suggested the involvement of V1V2 and V3 in gp120 trimer association [18]. Deletion of V1V2 may result in a decreased association of gp120 with Env trimer. Therefore, viral escape from nAbs by increased length and increased number of PNGS in

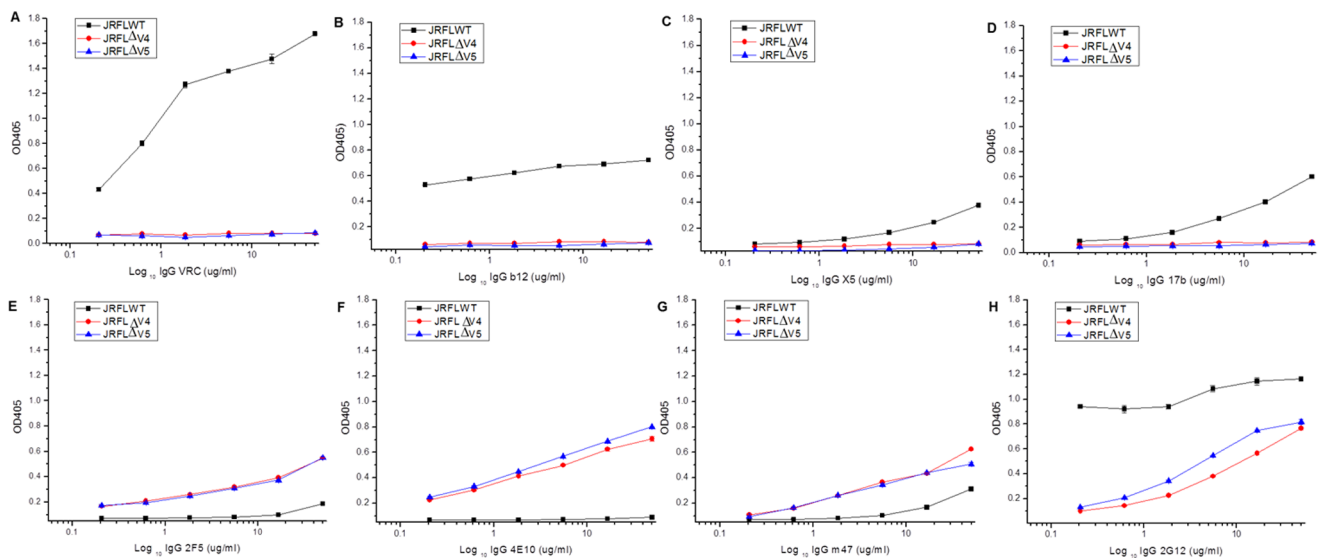


Figure 3. Effects of V4 and V5 loop deletions on Env structural integrity. 293T cells were co-transfected with recombinant pSVIII plasmid encoding JRFL gp160 WT, or $\Delta V4$, or $\Delta V5$ mutant and pCAT plasmid. 48h post transfection, the transfected cells were collected and lysed with cell lysis buffer, and added to the ELISA plates coated with anti-HIV antibody D7324. Captured Env proteins were detected using gp120-specific CD4bs mAbs VRC01, b12, or CD4i mAbs X5, 17b, or glycan-specific mAb 2G12, or gp41-specific mAbs 2F5, 4E10, m47 as primary antibody and HRP-anti human IgG, F(ab')₂ as secondary antibody. doi:10.1371/journal.pone.0069789.g003

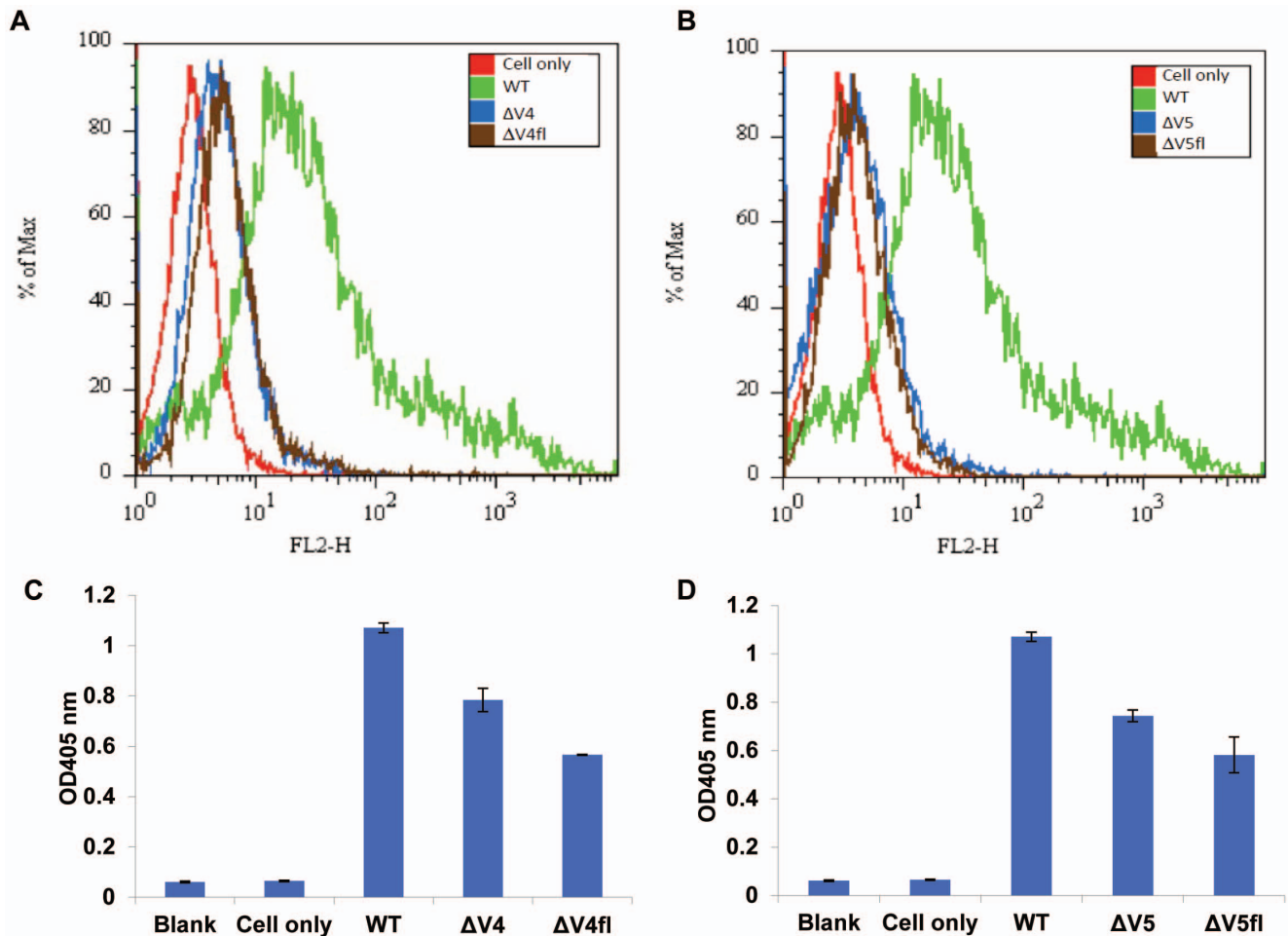


Figure 4. Expression of Env mutants with V4 and V5 replaced with flexible linkers of the same lengths as the original loops in 293T cells as measured by flow cytometry (A and B) and ELISA (C and D). 293T cells were co-transfected with recombinant pSVIII plasmid encoding JRFL gp160 WT, or Env mutants with V4 (A and C) or V5 (B and D) replaced with a flexible linker of the same length as the original loop, and pcTAT plasmid. 48h post transfection, the cells were detached and stained with 2G12 as primary antibody and PE-anti-human IgG, F(ab')₂ as secondary antibody (A and B), or cells were lysed with cell lysis buffer and the expressed Envs in the cell lysate captured by anti-HIV antibody D7324 (5 μg/ml) coated on high binding ELISA plates and bound Env proteins measured by using 2G12 as primary antibody and HRP-anti human IgG, F(ab')₂ as secondary antibody (C and D). doi:10.1371/journal.pone.0069789.g004

V1V2 region during chronic infection may come at the cost of viral fitness. Loop D may be directly or indirectly involved in Env interaction with the receptor CD4, so deletion of loop D abolished virus entry. This notion is supported by the observation that amino acid substitutions in loop D cause viral escape from CD4bs bnmAb VRC01 [19]. Interestingly, we found that deletion of V2, or V2 crown, or V3, or V3 crown significantly enhanced pseudovirus assembly, especially when the CT was also deleted (Table 3), but the effects of these deletions on pseudovirus entry were different. Deletion of V2 or V2 crown negatively affected pseudovirus entry, while deletion of V3 abolished pseudovirus entry. But to our surprise, we found that deletion of V3 crown significantly enhanced pseudovirus entry, and this enhancement was even more significant in the absence of CT. When the CT was deleted, deletion of V2 crown also enhanced pseudovirus entry. Both V2 crown and V3 crown sequences were conserved, and V3 crown was very immunogenic and served as a target for developing V3-specific nAbs and inhibitory peptides [20]. It was postulated that V3 crown was critical for coreceptor binding based on the observation that V3 crown structurally mimicked the β2-β3

loop in the CXC and CC chemokines [21]. But our study indicated that V3 crown may not be involved in binding to the coreceptor (either CCR5 or CXCR4). This is supported by two observations. First, V3 crown was not involved in coreceptor selectivity [21]. Second, antibodies targeting V3 stem showed broad neutralizing activity [22].

Deletion of V4 or V5 did not lead to the failure in Env protein expression, but significantly affect Env cell surface display, as well as pseudovirus assembly and subsequent entry into the cells. According to the crystal structure of gp120 core containing V4 and V5, deletion of V4 essentially removed a conserved glycan at position 386 and half of the beta sheet-19, which may destroy the CD4 binding site, while deletion of V5 removed half of the beta sheet-24, which may in turn destabilize the neighboring CD4 binding loop. Nevertheless, expression of ΔV4 and ΔV5 Envs in the cells was not significantly affected. Failure in exporting ΔV4 and ΔV5 Envs to cell surface suggested that V4 and V5 may be involved in Env binding to host factors and/or matrix, which is required for transportation machinery to display Envs on cell surface. Loss of Env structural integrity caused by deletion of V4

or V5 may be associated with the loss of binding of Env to the host factors and/or matrix [23]. We found that replacement of V4 or V5 with a flexible linker of the same length did not alleviate the defects in Env cell surface display compared to the loop deletion mutants (replaced with significantly shorter linkers), indicating that V4 and V5 lengths may not be determining factors for virus sensitivity to nAbs and disease progression. Instead, increased numbers of PNGS in V4 and V5 along with the increased loop lengths may play more important roles in viral escape from nAbs and disease progression.

We tested the effect of Δ CT in combination with each loop deletion on Env protein expression, Env cell surface display, and pseudovirus assembly and subsequent entry. HIV-1 Env CT is unusually long (150 amino acids) and highly conserved. It affects Env cell surface display and subsequent virus assembly and entry [23,24]. Interaction between matrix and CT is required for Env incorporation into membrane and virus infectivity [25]. In this study, we also observed that Δ CT alone led to increased Env expression and Env cell surface display, and significantly enhanced pseudovirus assembly and entry. But deletion of CT did not rescue the defects of Δ V4 and Δ V5 Envs in cell surface display, and in pseudovirus assembly and entry into the cells, suggesting that V4 and V5 deletion or replacement may be detrimental to Env structure and function.

References

- Curlin ME, Zioni R, Hawes SE, Liu Y, Deng W, et al. (2010) HIV-1 envelope subregion length variation during disease progression. *PLoS Pathog* 6: e1001228.
- Castro E, Belair M, Rizzardi GP, Bart PA, Pantaleo G, et al. (2008) Independent evolution of hypervariable regions of HIV-1 gp120: V4 as a swarm of N-Linked glycosylation variants. *AIDS Res Hum Retroviruses* 24: 106–113.
- Monno L, Saracino A, Scudeller L, Punzi G, Brindicci G, et al. (2011) Impact of mutations outside the V3 region on coreceptor tropism phenotypically assessed in patients infected with HIV-1 subtype B. *Antimicrob Agents Chemother* 55: 5078–5084.
- van Gils MJ, Bunnik EM, Boeser-Nunnink BD, Burger JA, Terlouw-Klein M, et al. (2011) Longer V1V2 region with increased number of potential N-linked glycosylation sites in the HIV-1 envelope glycoprotein protects against HIV-specific neutralizing antibodies. *J Virol* 85: 6986–6995.
- Bunnik EM, Euler Z, Welkers MR, Boeser-Nunnink BD, Grijsen ML, et al. (2010) Adaptation of HIV-1 envelope gp120 to humoral immunity at a population level. *Nat Med* 16: 995–997.
- Wei X, Decker JM, Wang S, Hui H, Kappes JC, et al. (2003) Antibody neutralization and escape by HIV-1. *Nature* 422: 307–312.
- Moore PL, Gray ES, Choge IA, Ranchohe N, Mlisana K, et al. (2008) The c3-v4 region is a major target of autologous neutralizing antibodies in human immunodeficiency virus type 1 subtype C infection. *J Virol* 82: 1860–1869.
- Rong R, Gnanakaran S, Decker JM, Bibollet-Ruche F, Taylor J, et al. (2007) Unique mutational patterns in the envelope alpha 2 amphipathic helix and acquisition of length in gp120 hypervariable domains are associated with resistance to autologous neutralization of subtype C human immunodeficiency virus type 1. *J Virol* 81: 5658–5668.
- Rong R, Li B, Lynch RM, Haaland RE, Murphy MK, et al. (2009) Escape from autologous neutralizing antibodies in acute/early subtype C HIV-1 infection requires multiple pathways. *PLoS Pathog* 5: e1000594.
- Rong R, Bibollet-Ruche F, Mulenga J, Allen S, Blackwell JL, et al. (2007) Role of V1V2 and other human immunodeficiency virus type 1 envelope domains in resistance to autologous neutralization during clade C infection. *J Virol* 81: 1350–1359.
- Zhou T, Georgiev I, Wu X, Yang ZY, Dai K, et al. (2010) Structural basis for broad and potent neutralization of HIV-1 by antibody VRC01. *Science* 329: 811–817.
- McLellan JS, Pancera M, Carrico C, Gorman J, Julien JP, et al. (2011) Structure of HIV-1 gp120 V1/V2 domain with broadly neutralizing antibody PG9. *Nature* 480: 336–343.
- Pejchal R, Doores KJ, Walker LM, Khayat R, Huang PS, et al. (2011) A potent and broad neutralizing antibody recognizes and penetrates the HIV glycan shield. *Science* 334: 1097–1103.
- Pancera M, Wyatt R (2005) Selective recognition of oligomeric HIV-1 primary isolate envelope glycoproteins by potentially neutralizing ligands requires efficient precursor cleavage. *Virology* 332: 145–156.
- Haynes BF, Gilbert PB, McElrath MJ, Zolla-Pazner S, Tomaras GD, et al. (2012) Immune-correlates analysis of an HIV-1 vaccine efficacy trial. *N Engl J Med* 366: 1275–1286.
- Song L, Sun ZY, Coleman KE, Zwick MB, Gach JS, et al. (2009) Broadly neutralizing anti-HIV-1 antibodies disrupt a hinge-related function of gp41 at the membrane interface. *Proc Natl Acad Sci U S A* 106: 9057–9062.
- Huang J, Ofek G, Laub L, Louder MK, Doria-Rose NA, et al. (2012) Broad and potent neutralization of HIV-1 by a gp41-specific human antibody. *Nature* 491: 406–412.
- Mao Y, Wang L, Gu C, Herschhorn A, Xiang SH, et al. (2012) Subunit organization of the membrane-bound HIV-1 envelope glycoprotein trimer. *Nat Struct Mol Biol* 19: 893–899.
- Li Y, O'Dell S, Walker LM, Wu X, Guenaga J, et al. (2011) Mechanism of Neutralization by the Broadly Neutralizing HIV-1 Monoclonal Antibody VRC01. *J Virol* 85: 8954–8967.
- Gazarian KG, Palacios-Rodriguez Y, Gazarian TG, Huerta L (2013) HIV-1 V3 loop crown epitope-focused mimotope selection by patient serum from random phage display libraries: Implications for the epitope structural features. *Mol Immunol* 54: 148–156.
- Cardozo T, Kimura T, Philpott S, Weiser B, Burger H, et al. (2007) Structural basis for coreceptor selectivity by the HIV type 1 V3 loop. *AIDS Res Hum Retroviruses* 23: 415–426.
- Stanfield RL, Gorny MK, Zolla-Pazner S, Wilson IA (2006) Crystal structures of human immunodeficiency virus type 1 (HIV-1) neutralizing antibody 2219 in complex with three different V3 peptides reveal a new binding mode for HIV-1 cross-reactivity. *J Virol* 80: 6093–6105.
- Piller SC, Dubay JW, Derdeyn CA, Hunter E (2000) Mutational analysis of conserved domains within the cytoplasmic tail of gp41 from human immunodeficiency virus type 1: effects on glycoprotein incorporation and infectivity. *J Virol* 74: 11717–11723.
- Checkley MA, Lutge BG, Freed EO (2011) HIV-1 envelope glycoprotein biosynthesis, trafficking, and incorporation. *J Mol Biol* 410: 582–608.
- Freed EO, Martin MA (1996) Domains of the human immunodeficiency virus type 1 matrix and gp41 cytoplasmic tail required for envelope incorporation into virions. *J Virol* 70: 341–351.

Conclusions

HIV-1 Env variable loops are indispensable for Env structural integrity and Env-mediated virus entry. Deletion of V1V2, or V3, or loop D abolished virus entry. Deletion of V4 or V5, or replacement of V4 or V5 with flexible linkers of the same lengths knocked out receptor and coreceptor binding sites in gp120, but enhanced the exposure of the N-trimer structure and the MPER in gp41. Deletion of V4 or V5 significantly affected Env cell surface display, leading to the failure in virus assembly and subsequent entry. Our findings suggest that, in addition to the CD4b1 and V3, other Env variable loops, such as V1V2, loop D, V4 and V5, may be explored for development of vaccine immunogens and therapeutics against HIV-1 infection.

Acknowledgments

We wish to thank Yuxing Li, Richard Wyatt and Joseph Sodroski for providing pSVIII-JRFLgp160 and pcTAT plasmids, Tongqing Zhou and Dimitar S Dimitrov for helpful discussions.

Author Contributions

Conceived and designed the experiments: MYZ TY. Performed the experiments: TY JL. Analyzed the data: MYZ TY. Wrote the paper: MYZ.