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REVIEW ARTICLE
Human enterovirus 71 and hand, foot and mouth disease

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SUMMARY

Hand, foot and mouth disease (HFMD) is generally a benign febrile exanthematous childhood disease caused by human enteroviruses. The route of transmission is postulated to be faeco-oral in developing areas but attributed more to respiratory droplet in developed areas. Transmission is facilitated by the prolonged environmental survival of these viruses and their greater resistance to biocides. Serious outbreaks with neurological and cardiopulmonary complications caused by human enterovirus 71 (HEV-71) seem to be commoner in the Asian Pacific region than elsewhere in the world. This geographical predilection is unexplained but could be related to the frequency of intra- and inter-typic genetic recombinations of the virus, the host populations' genetic predisposition, environmental hygiene, and standard of healthcare. Vaccine development could be hampered by the general mildness of the illness and rapid genetic evolution of the virus. Antivirals are not readily available; the role of intravenous immunoglobulin in the treatment of serious complications should be investigated. Monitoring of this disease and its epidemiology in the densely populated Asia Pacific epicentre is important for the detection of emerging epidemics due to enteroviruses.

Key words: Hand, foot and mouth disease, enterovirus, enterovirus 71.

VIROLOGY

The family Picornaviridae contains 12 genera of non-enveloped, linear positive-sense, single-stranded RNA viruses which include *Aphthovirus*, *Cardiovirus*, *Enterovirus*, *Erbovirus*, *Hepatovirus*, *Kobuvirus*, *Parechovirus*, *Teschovirus*, *Tremovirus*, *Avihepatovirus*, *Senecavirus*, and *Sapelovirus* [1]. The last five genera have not so far been associated with human infections to date. The currently recognized species and types of *Enterovirus* are listed in Table 1 [1, 2].

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In addition to the classical communicable disease syndromes such as poliomyelitis, herpangina, hepatitis A, and common cold, several new picornaviruses are found in humans. These newly described viruses include human parechoviruses, Saffold Cardiovirus, Klassevirus, Salivirus and other yet unclassified viruses, but their association with clinical disease is still unclear [3–8].

The genome of human enterovirus 71 (HEV-71) is about 7·4 kb in size, which is flanked by 5′ and 3′ untranslated regions. The protein-coding region can be divided into three regions: P1 encodes for the structural proteins VP4, VP2, VP3, and VP1; P2 and P3 for the non-structural proteins 2A, 2B, 2C and 3A, 3B, 3C, 3D, respectively [9, 10]. A single, long

Table 1. *Enteroviral species and types currently recognized*

Enterovirus species	Types
Human enterovirus A	Human Coxsackievirus A2–8, 10, 12, 14, 16. Human enterovirus 71, 76, 89–92.
Human enterovirus B	Human Coxsackievirus A9, B1–6. Human echovirus 1–9, 11–21, 24–27, 29–33. Human enterovirus 69, 73–75, 77–88, 93, 97, 98, 101, 106, 107.
Human enterovirus C	Human poliovirus 1–3. Human Coxsackievirus A1, 11, 13, 15, 17–22, 24. Human enterovirus 95, 96, 99, 102, 104, 105, 109.
Human enterovirus D	Human enterovirus 68, 70, 94.
Human rhinovirus A, B, C	
Porcine enterovirus B	
Bovine enterovirus	
Simian enterovirus A	

polyprotein is translated from the viral RNA which is then cleaved to form the individual proteins. VP1 carries most of the type-specific neutralizing antibody epitopes. Mutations in VP1 have been associated with increased virulence in animal models [11].

HEV-71 has three genotypes (A, B, and C) based on *VP1* and *VP4* gene sequences. Genotypes B and C are each further divided into five subtypes B1–B5 and C1–C5, respectively. More recently, analysis of complete genome sequences of HEV-71 (including non-structural protein genes) suggested that sub-genotype C4 should be classified as a new genotype D (Fig. 1) [12]. Retrospective analysis of strains of HEV-71 isolated in The Netherlands from 1963 to 1967 suggested that there is a new subgenotype B0 [13].

Genetic recombination among RNA viruses was first noted in poliovirus [14]. Recombination occurs through template switch and other mechanisms mainly in the non-structural protein region of the genome [14, 15]. Intra-typic recombinations are commoner than inter-typic ones owing to the higher degree of sequence homology between the gene segments. Recombination is also frequent and a major source of genetic variation in other non-polio enteroviruses [16–20]. Most recombination occurs within the same species of virus; natural inter-species recombination is uncommon. Intra-typic and inter-typic HEV-71 recombinants have been detected in outbreaks in the Asia Pacific region. In Malaysia, inter-typic recombinants of HEV-71 have been described with the P3 region being derived from CV-A16 [21]. In China, intra-typic recombinants were found to be circulating in the 2008 outbreak of HEV-71 [22]. Although recombinants can be a driving force in the genesis of new epidemics, other factors such as cross-protection with other genotypes, virulence, and transmissibility may also play a role in determining the size and outcome of epidemics. Examples of both intra-typic and inter-typic recombinations are shown in Figure 2. Bootscan analysis of a recent EV-71 strain identified in Guangzhou suggested that intra-typic recombination occurs between EV-71 genotypes B and C at junctions 2A–2B and 3C; however, the analysis of a recent HEV-71 strain identified in Shanghai suggested inter-typic recombination between HEV-71 genotype C and CA16 G-10.

PATHOGENESIS AND PATHOLOGY

At least three human cellular receptors of HEV-71 have recently been identified [23–25]. The relative importance of these receptors in different tissues or in different phases of the infection awaits clarification. Human dendritic cells are susceptible to infection by HEV-71 through DC-SIGN [dendritic cell-specific intracellular adhesion molecules (ICAM)-3 grabbing non-integrin, also known as CD209] [23]. Infected dendritic cells serve as antigen-presenting cells with the ability to prime T cells to generate protective immune responses. DC-SIGN is unlikely to be the sole receptor for HEV-71 since cell types not expressing DC-SIGN can also be infected by the virus.

Another HEV-71 cellular receptor is the human P-selectin glycoprotein ligand-1 (CD162) [24]. CD162 is a cell surface glycoprotein which plays an important role in the binding of leukocytes to endothelial cells and platelets. It is expressed on the surface of cells of haematopoietic origin but not on parenchymal cells of most tissues. Cells expressing the ligand include circulating leukocytes, dendritic cells, tissue macrophages (such as those in liver, lung, bowel, and Langerhans cells in the skin) and myeloid progenitor cells [26]. Its presence on the macrophages in the
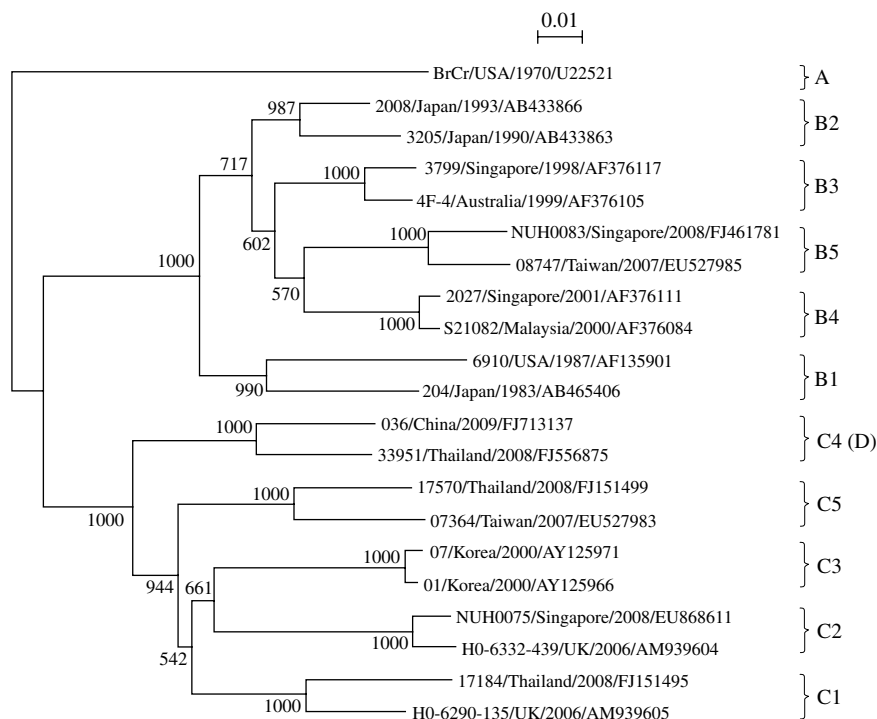
0.01

Fig. 1. Phylogenetic analysis of all genotypes of EV-71 represented by their most recent isolates based on alignment of *VP1* gene sequence available in GenBank (891 nucleotide positions in each *VP1* region were included in the analysis). The scale bar indicates the estimated number of substitutions per 100 bases. Phylogenetic tree construction was performed using neighbour-joining method with GrowTree using Kimura's two-parameter correction, with bootstrap values calculated from 1000 trees (Genetics Computer Group Inc., USA)

mucosa-associated lymphoid tissues of the alimentary tract has been postulated to represent the primary site of viral multiplication after infection and the infection and activation of Langerhans cells in the skin reflects the genesis of the skin lesions typical of hand, foot and mouth disease (HFMD) [24].

A third receptor is the scavenger receptor BII (SR-BII) which has the physiological function of mediating high-density lipoprotein uptake into and cholesterol efflux from cells [25, 27, 28]. It is expressed in significant amount in various organs and cells, including the liver, spleen, testes, retinal pigment epithelial cells, osteoblasts, macrophages, and importantly, the brain [29–31]. The scavenger receptor class B is also involved in the uptake of the hepatitis C viruses [32, 33].

The ubiquity of HEV-71 receptors in different organs may account for the systemic nature of HEV-71 infection in severe cases and the predilection for involvement of the central nervous system (CNS). In poliovirus infection, the poliovirus receptor CD155 is present in a large number of organs and tissues, yet not all these organs are sites of viral replication

or exhibit the pathology of infection [34, 35]. Hence, the presence of receptors *per se* may be a necessary but not sufficient prerequisite for the pathogenesis of the infection. Both polioviruses and HEV-71 are neurotropic, thus explaining their propensity to cause neurological complications such as acute flaccid paralysis. Neuropathological examination of fatal human encephalomyelitis patients showed that inflammation involved the whole grey matter of the spinal cord, tegmentum of the midbrain, hypothalamus, and subthalamic and dentate nuclei, and more focally and less intensely in the cerebral cortex, especially the motor cortex. The virus may enter the CNS via the motor pathway of the peripheral nervous system, possibly through retrograde axonal transport [36, 37]. This is supported by the *in vitro* finding that infection of the human neuronal cell line (SK-N-SH) by HEV-71 resulted in the highest level of viral replication when compared to human laryngeal (HEp-2), human glial (U373MG), and African green monkey kidney (Vero) cell lines [38]. Experimental evidence suggested that HEV-71-infected cells (including neurons and Vero cells) undergo apoptosis
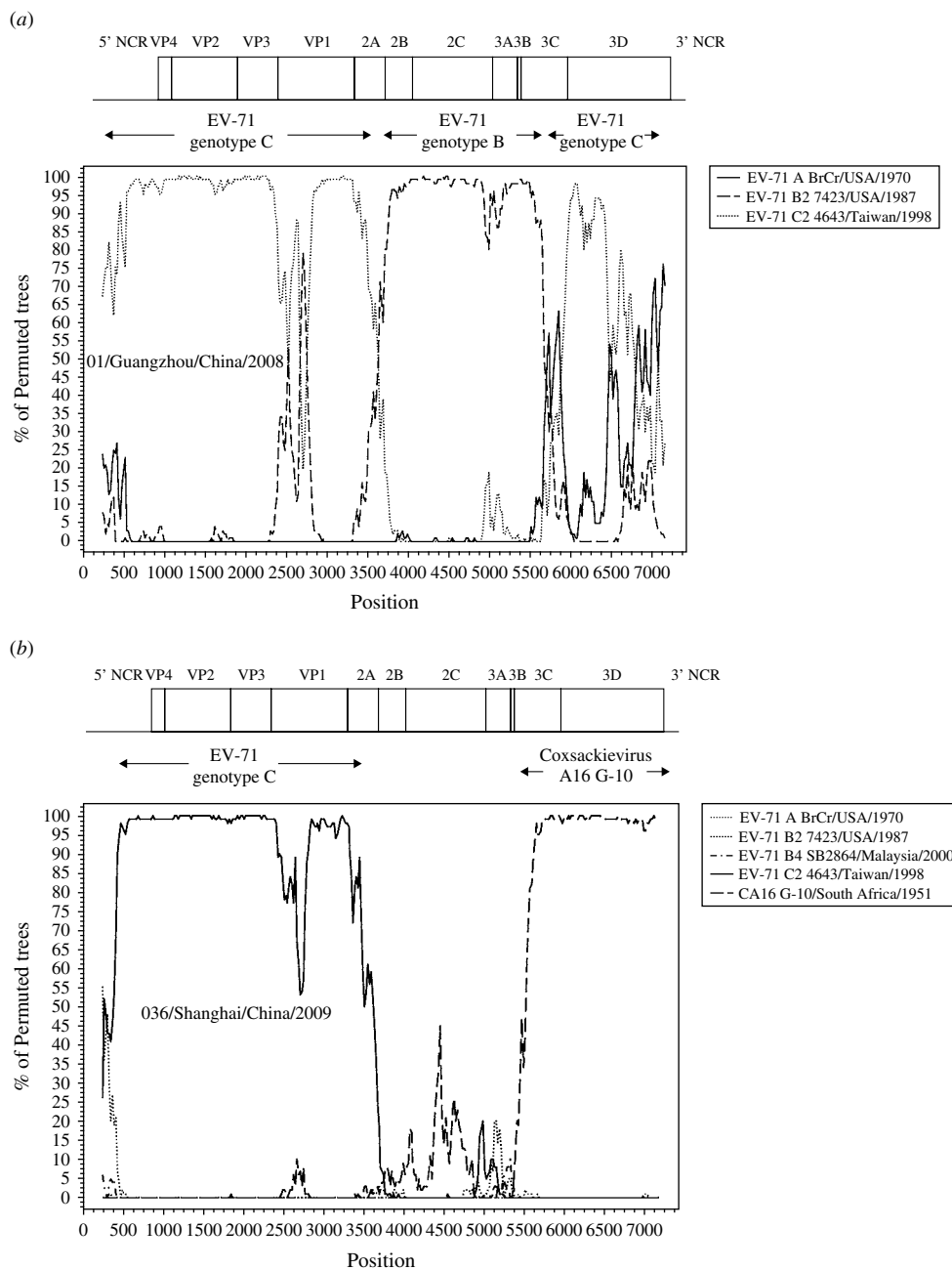
(a)

(b)

Fig. 2. (*a*) A recent example of intra-typic recombination reviewed by bootscan analysis [bootscanning was conducted with Simplot version 3.5.1 (Kimura distance model: window size 500 bp, step 20 bp) on a gapless nucleotide alignment, generated with ClustalX, with the genome sequence of the EV-71 strain (01/Guangzhou/China/2008) as the query sequence]. (*b*) A recent example of inter-typic recombination reviewed by bootscan analysis [bootscanning was conducted with Simplot version 3.5.1 (Kimura distance model: window size 500 bp, step 20 bp) on a gapless nucleotide alignment, generated with ClustalX, with the genome sequence of the EV-71 strain (036/Shanghai/China/2009) as the query sequence].

through a variety of pathways [39–43]. In addition, neuronal damage can also be caused by HEV-71-induced cellular autophagy [44]. Finally, HEV-71 infection of endothelial cells can lead to activation and apoptosis; this may serve as an alternative mechanism of end organ damage in systemic infections [43].

The brains of patients who developed HEV-71 encephalitis generally showed oedema, vascular congestion, and typical pathological features of

encephalitis [45–47]. There is a predominant involvement of the grey matter especially in the brainstem; inflammation in cerebellum, spinal cord, and the meninges is often present. Involvement of the cerebrum is generally less intense. Neuronal degeneration and necrosis are common. The inflammatory response is characterized by perivascular mononuclear cell infiltration. Micro-abscess formation and glial nodules may also be seen. The lung of a patient with cardiopulmonary failure is characterized pathologically by oedema, diffuse alveolar damage and the presence of inflammatory infiltrates [45]. Viral myocarditis is not a feature of HEV-71-associated cardiopulmonary failure in patients who developed brainstem encephalitis. The myocardium showed coagulative myocytolysis and myofibrillar degeneration, which suggested that the pathogenesis is one of neurogenic cardiac damage rather than direct involvement by infection [48, 49].

PROTECTIVE IMMUNITY

There is clinical and experimental evidence on the roles of different arms of immune responses in the protective immunity against HEV-71 infection. The cellular and humoral immune responses are both essential for decreasing the viral load and mortality in mice [50]. In humans, cellular immunity is important in preventing the development of serious complications after HEV-71 infection [51, 52]. On the other hand, the neutralizing antibodies from the humoral response appear to be crucial in the protective immunity against infection [53–55]. During the 1998 epidemic in Taiwan, attack and case-fatality rates were lowest in seropositive infants aged <6 months, suggesting a protective role of maternal antibodies [56, 57]. In humans, presence of maternal anti-HEV-71 antibodies has also been demonstrated in neonates, the prevalence and titre of which correlate with those levels in the mothers [58]. In mice, transplancental transfer of antibodies following maternal immunization against EV-71 protects against lethal infection of newborn mice [59]. Thus, it appears that the seroprevalence of neutralizing antibodies in women of childbearing age is important in protecting infants aged <6 months. The protective role of breastfeeding also needs to be studied: breast milk contains lactoferrin which inhibits binding of HEV-71 to host cells. Whether secretory IgA in maternal milk contributes to the mucosal immunity against HEV-71 (as has been proven in the case of poliovirus) is not known.

CLINICAL DISEASE

Many picornaviral infections occur mainly in childhood. The propensity to cause outbreaks is an important feature of some of these viruses, most notably poliomyelitis in the pre-vaccination era and now, enteroviruses. Enterovirus outbreaks range from small community clusters of acute haemorrhagic conjunctivitis due to Coxsackieviruses to large nationwide HEV-71 epidemics. Most enteroviral infections are asymptomatic which adds to the difficulty in controlling spread in the community.

Clinical syndromes typically associated with enteroviral infections include undifferentiated fever; neurological manifestations (acute flaccid paralysis, aseptic meningitis, meningoencephalitis); respiratory infections with exanthems and/or enathems (HFMD, herpangina); eye infections (acute haemorrhagic conjunctivitis); cardiovascular infections (pericarditis, myocarditis); muscle diseases (pleurodynia and Bornholm disease); and systemic infections.

HFMD is a common illness in children aged <10 years. The infection typically has an incubation period of 3–7 days. The main manifestations are fever, lymphadenopathy, followed in 1–2 days' time by the appearance of vesicles on the palmar and plantar skin, buccal mucosa, and tongue. Papular and vesicular lesions can also occur on other parts of the body. The oral enanthem helps to distinguish HFMD from other causes of childhood exanthems, although cases without oral lesions have been described. Uncomplicated HFMD usually resolves in 5–7 days. CV-A16 and HEV-71 are the commonest causes of HFMD, the latter is especially common in the Asia Pacific region. Other enteroviruses causing HFMD include CV-A4 to A7, A9, A10, A24, B2–B5, echoviruses 1, 4, 11, 18, and HEV-18. Clinical features of HFMD caused by these viruses are indistinguishable. In contrast to CV-A16, HFMD caused by HEV-71 is more likely to cause a high fever ($\geqslant 39\,^{\circ}$C) and fever for >3 days, more severe illness, and a higher risk of developing complications and fatalities [60, 61]. HFMD is rare in adults but cases due to HEV-71 have been reported. Adults can also develop severe HEV-71 infections such as encephalitis as a result of intra-familial transmission [62, 63].

HEV-71 infection commonly manifests as HFMD or herpangina. In a few patients, neurological and cardiopulmonary complications with substantial mortality may occur. No specific genotype is associated with more severe disease [64]. Children aged <5 years

have the highest incidence of severe complications [65–67]. Fatal cases of HFMD due to HEV-71 were more often associated with vomiting and a lower incidence of mouth ulcers [61]. The predominant forms of neurological syndrome include aseptic meningitis, acute flaccid paralysis, brainstem encephalitis, or cerebellitis and vary in different epidemics. These complications often appear early, at 2–5 days after the onset of illness [68]. Over an 8-year period from 1998 to 2005, the case-fatality rate of complicated enteroviral infections – most of which were caused by HEV-71 – ranged from 10·0% to 25·7% [65].

Long-term neurological sequelae are common in survivors who had more serious CNS disease and cardiopulmonary failure. Late complications include limb weakness, dysphagia requiring tube-feeding, cerebellar dysfunction, delayed neurodevelopment, and impaired cognitive functions [69, 70].

Acute cardiopulmonary failure as a complication of systemic HEV-71 infection has a high mortality. Pulmonary oedema is related to cerebral compression or increased intracranial pressure which leads to sympathetic hyperactivity [71]. Pulmonary oedema and the associated hypoxaemia and acute respiratory distress syndrome are the commonest causes of death in severe HEV-71 infections. Similarly, brainstem encephalitis leads to acute heart failure in 19% of patients and this complication has a high mortality rate of 77% [48]. Again, hyperactivity of sympathetic stimulation to the heart leading to a 'catecholamine storm' and neurogenic cardiac damage is believed to be the mechanism of cardiac damage in this infection.

Risk factors associated with the progression to CNS involvement without pulmonary oedema in HEV-71 infection included fever for $\geqslant 3$ days, peak body temperature of $\geqslant 39$ °C, the presence of headache, lethargy, vomiting, seizure, and hyperglycaemia. Hyperglycaemia, leucocytosis, and limb weakness were found to be risk factors for pulmonary oedema [72]. In some series, a higher level of leucocytosis was found in patients with CNS involvement [73]. In a series of 333 patients with CNS involvement due to non-poliovirus in Taiwan, severe CNS disease was associated with age <4 years, leucocytosis (over 13×10^9/l), seizure, myoclonic jerks, and a higher incidence of skin rash and oral ulcers [74].

In those patients who developed serious CNS disease due to non-poliovirus infection, poor prognostic factors included age <2 years, higher peak leucocyte counts (over 17×10^9/l), a higher incidence of skin rash, and a lower yield of virus from the cerebrospinal fluid [74]. For HEV-71 cardiopulmonary failure, poorer clinical outcomes were associated with higher troponin I level, lower initial systolic blood pressure, longer duration of hypotension, greater requirements for inotropic support, lower PaO_2:FiO_2 ratio, higher white blood cell counts in the cerebrospinal fluid, and the lowest Glasgow coma score. Fatality correlated best with the troponin I level [75].

LABORATORY DIAGNOSIS

Aetiological diagnosis of HFMD can be achieved by examining the vesicular fluid aspirated from the skin lesion and naso-/oro-pharyngeal swabs. In complicated cases with brainstem encephalitis and cardiopulmonary failure with sparse skin lesions HEV-71 may still be detectable in the cerebrospinal fluid, naso-/oro-pharyngeal secretions, or faeces. The commonest rapid diagnostic test is by reverse transcription–polymerase chain reaction (RT–PCR) of the RNA extracted from these specimens, targeting towards the 5′ untranslated or *VP1* region of the viral genome [76–78]. Isolation of virus from clinical specimens is possible using conventional cell culture or rapid shell viral culture with rhabdomyosarcoma (RD), HEp-2, colonic carcinoma (CaCo-2), or Vero cell lines and cytopathic effects can be seen in 3–7 days [79]. Cell lines infected by HEV-71 or CV-A16 can be differentiated by immunostaining with specific monoclonal antibodies against their VP1 proteins. The isolates can be genotyped by PCR sequencing of the *VP1* and/or *VP4* genes. IgM antibody to HEV-71 has been detected as early as 2 days after onset of illness but, as the test is not yet widely available, serological diagnosis generally requires demonstration of a fourfold rise in neutralizing antibody titre taken 10–14 days after the onset of illness [80].

EPIDEMIOLOGY

HEV-71 was first detected in 1969 in California in an infant suffering from encephalitis. Initial isolations of HEV-71 were made in the USA and in Australia in the early 1970s, and outbreaks of HFMD occurred in Sweden and Japan [81, 82]. In the late 1970s, Bulgaria (1975) and Hungary (1978) witnessed large epidemics of HEV-71 infection with prominent neurological manifestations (aseptic meningitis, encephalitis, acute flaccid paralysis) [83, 84]. Since the late 1990s the densely populated Asia Pacific region has

been the hotspot for epidemics: Taiwan, Singapore, Malaysia, China, Vietnam, and Australia have experienced recurrent epidemics of various sizes. The reason for this geographical distribution is uncertain, but the association between HLA-A33 (which is common in some Asian populations) and susceptibility to HEV-71 infection has been suggested as a possible explanation [85]. Other factors such as genetic predisposition (glucose-6-phosphate dehydrogenase deficiency, polymorphisms in cytotoxic T lymphocyte antigen-4 and scavenger receptor BII), food and water hygiene, and micronutrient deficiencies require further studies to confirm their significance [51, 86–91].

Recent epidemics of HFMD disease in the Asia Pacific region were mainly caused by HEV-71 (Table 2). However, more than one subgenotype of HEV-71 can be found co-circulating in the same epidemic, as well as other non-HEV-71 enteroviruses, such as CV-A16 (Table 3). Co-infection is possible in enteroviral HFMD [92, 93]. In Sarawak, Malaysia, co-infection of HEV-71 with other viruses occurred in 10% of patients [93]. Co-infection does not appear to result in more severe clinical disease [92].

The Taiwan epidemic of HEV-71 HFMD in 1998 was the largest outbreak in the Asia Pacific region until 2008, when 488955 cases of HFMD were reported in China (Table 2), certainly an underestimation of the actual burden of disease. More than 600 000 cases were reported from March to June 2009, but this could be because HFMD became a notifiable disease in China from May 2008. The epidemic centre in China in 2008 was the southeastern provinces of Guangdong, Zhejiang, and Anhui. The overall case-fatality rate reported from China in 2008 and 2009 was about 0·03%, but in certain local outbreaks, such as the 2008 outbreak in Fuyang City of Anhui Province, the case-fatality rate was as high as 0·3% [94].

Humans are the only known natural hosts of HEV-71. Intra-familial transmission of HEV-71 occurs commonly. In a prospective cohort in Taiwan, transmission rates from infected children to siblings were as high as 84% [66]. The rate of symptomatic infection after household transmission is higher than that in other community settings (94% vs. 29%) which is attributed to more prolonged contact with the cases and possibly a larger infective dose. Faeces and oropharyngeal secretions are likely to be important in the transmission of the virus. Shedding of non-polio enteroviruses in the stool can persist for 3–11 weeks after the onset of illness, while the duration of shedding from oral secretion is shorter [95, 96]. Enteroviruses are also detectable in throat swabs. During the 1998 Taiwan HEV-71 epidemic, detection of the virus from the throat swabs was more frequent than from stool specimens; the time to positivity by viral culture was also shorter [97]. The superior recovery rate of enteroviruses, mainly HEV-71 and CV-A16, from throat swabs over stool samples (with or without the testing of vesicle swabs) in HFMD was confirmed in another study in Sarawak, Malaysia [98]. A recent study from Mongolia showed that hand washing after defecation was associated with a lower risk of infection by non-polioviruses in households, whilst having a bathroom in the house was a risk factor for infection [99]. These findings suggested that the faeco-oral route is probably important. However, other hygiene measures did not affect the incidence of virus isolation. Hence, modes of transmission other than faeco-oral could also play an important role in the households. The relative importance or infectivity of oropharyngeal secretions vs. faeces in real-life situations have not been determined. We postulate that in developing countries, sanitation plays a more important role. In developed countries, although personal hygiene and sanitation facilities are much better, there is also a much larger number of facilities where great numbers of susceptible children congregate. Under such circumstances, respiratory transmission may become more significant. Faeco-oral transmission may contribute more to an endemic disease in the community, whereas the respiratory tract, whose secretions contain a higher viral load but have a shorter duration of shedding than faeces, may contribute more to the epidemic spread of the viruses during outbreak situations. In the 1998 Taiwan epidemic, in addition to intrafamilial contact with cases, attendance at a kindergarten or childcare centre and residence in a rural area were significant risk factors associated with HEV-71 infection. This suggests that close contacts at schools are important in the epidemiology of the disease by acting as sources of spread to the community [56]. Contact transmission from blister fluid may have a minor role.

Enteroviraemia has been demonstrated in healthy Scottish blood donors at a predicted prevalence of 0·023% [100]. The viruses being detected were primarily Coxsackieviruses and human echoviruses. Similarly, enterovirus, as well as cytomegalovirus, parvovirus B9, and adenovirus genomes have been detected in explanted heart myocardial tissues from

Table 2. *Recent outbreaks of HFMD due to HEV-71 in the Asia Pacific region*

Year	Location	Reported/ confirmed cases	Deaths	Major complications	Genotype of HEV-71 and other co-circulating viruses	References
1997	Peninsular and Sarawak Malaysia	n.a.	⩾35	Encephalomyelitis, cardiopulmonary failure	EV-71 B3, B4 (C1, C2) (CV-A16, A2, A4, A6, A9; CV-B5; EV-1, EV-4, EV-5, EV-7)	[43, 170–172]
1998	Taiwan	129 106*	78	Aseptic meningitis. encephalitis, meningoencephalitis, acute flaccid paralysis, acute pulmonary oedema/ haemorrhage (⩾405 cases)	EV-71 C2 (CV-A16; CV-B1, B2, B3, B5; EV-6, 7, 11, 22, 27)	[57, 173, 174]
1999	Australia	6000	n.a.	Aseptic meningitis, Guillain–Barré syndrome, acute transverse myelitis, acute cerebellar ataxia, opso-myoclonus syndrome, benign intracranial hypertension, febrile convulsion	EV-71 C2	[175, 176]
2000	Australia	200	n.a.	Acute pulmonary oedema	EV-71 B4	[175]
2000	Singapore	3790	4 (+1 possible case)	Acute pulmonary oedema and haemorrhage, encephalitis, aseptic meningitis	EV-71 B4 (CV-A16, A3, A4, A5, A6, A10, A23; HEV-18)	[82]
2000	Taiwan	80 677	41	291 cases	EV-71 B4 (CV-A16, A9, A24; CV-B1, B3, B4; EV-4, EV-9)	[123, 174]
2001	Taiwan	n.a.	58	389 cases	EV-71 B4 (CV-A16, A6, A9, A24; CV-B4, B5; EV-4, EV-6)	[65, 174]
2000	Sarawak, Malaysia	n.a.	n.a.		EV-71 B4 (CV-A16)	[124]
2000	Peninsular Malaysia	n.a.	n.a.		EV-71 C1, B4	[170, 171]
2003	Sarawak, Malaysia	n.a.	n.a.		EV-71 B4, B5, C1 (CV-A16)	[170, 171]
2005	Peninsular Malaysia	n.a.	n.a.		EV-71 B5, C1	[171]
2006	Singapore	15 282	n.a.	1·8 % cases hospitalized†	EV-71 B5, CV-A16	[120]
2006	Brunei	1681	3	Neurological	EV-71 B5	[177]
2007	China	83 344	17			[178]
2008	China	488 955	126		EV-71 C4	[178, 179]
2009	China	614 901 (March to June 2009)	200		EV-71 C4	[22, 180]

n.a., Not available; CV-A, Coxsackievirus A; HEV, human enterovirus; EV, echovirus. Viruses or genotypes in parentheses are not the predominant strains.

* Actual number of cases estimated to be ten times higher.

† Compared to 0·7–0·8 % in epidemics due to CV-A16.

Table 3. *Summary of predominant EV-71 genotypes in the Asia Pacific region*

Year	B1	B2	B3	B4	B5	C1	C2	C3	C4	C5
				Genotypes						
1986						Sydney				
1987						Sydney				
1989						Sydney				
1990						Sydney				
1991						Sydney				
1993							**Japan**			
1994						Sydney				
1995						Sydney	Australia			
1996						Sydney				
1997			**Sarawak Malaysia, peninsular Malaysia**, Singapore, Japan	**Peninsular Malaysia**, Singapore, **Japan**		**Peninsular Malaysia**	**Peninsular Malaysia**, Japan	China		
1998			Sarawak Malaysia, Singapore	Peninsular Malaysia, Taiwan		Sarawak Malaysia, peninsular Malaysia, Singapore	Peninsular Malaysia, **Taiwan**		China	
1999			Sarawak Malaysia, Singapore, **Perth**	Peninsular Malaysia, Taiwan		Peninsular Malaysia	Peninsular Malaysia, **Perth**		China	
2000				**Sydney**, Sarawak Malaysia, **peninsular Malaysia**, **Taiwan**, **Singapore**	**Peninsular Malaysia, Japan**	Sarawak Malaysia, peninsular Malaysia, Perth	Peninsular Malaysia	Korea	China	
2001				Sarawak Malaysia, Sydney, **Taiwan**, Singapore					China	
2002				Sarawak Malaysia, Taiwan, Singapore		Sarawak Malaysia, Singapore, Thailand			China	
2003				Taiwan	**Sarawak** Malaysia		Sarawak Malaysia		China, Japan	
2004									China, Taiwan	
2005					**Peninsular Malaysia**	**Peninsular Malaysia**, Vietnam			Taiwan, Vietnam	**Vietnam**
2006					**Singapore, Brunei**, Taiwan					Taiwan
2007					Taiwan				**China**	Taiwan
2008									**China**	
2009									**China**	

Boldface indicates major outbreaks as indicated by the reports showing a significant increase in incidence above their local baseline.

Data from references [22, 64, 124, 171, 175, 179, 181–186].

heart transplant donors and recipients in Germany. The prevalence of enterovirus as detected by RT–PCR reaction ranged from 21% to 60% of the heart samples [101]. During the HEV-71 epidemic in Taiwan in 1998, 20% of patients in one study had viraemia as detected by RT–PCR [73]. Although these findings suggested that transfusion of blood products and transplantation could be potential routes of transmission, such cases have not yet been reported. One case of intrauterine infection by HEV-71 was reported and the pregnancy was complicated by still-birth with virological evidence of brain and liver involvement [102]. Another baby who developed perinatal HEV-71 infection with HFMD and aseptic meningitis had a benign course and recovered without long-term neurological sequelae [103].

The contribution of environmental factors in the ecology and transmission of HEV-71 is not well understood. Few environmental studies specifically addressed HEV-71, although one may draw inferences from studies involving other better studied picornaviruses such as poliovirus and Coxsackie-viruses. Enteroviruses have a higher level of resistance to biocides compared to other enveloped viruses. They are fairly tolerant to temperature, salinity, pH, and sewage treatment procedures. Different types of enteroviruses – including HEV-71 – can readily be detected in sewage and other environmental waters [104, 105]. The finding of enteroviruses in surface waters has no correlation with the level of other indicator organisms of water pollution. Outbreaks of picornavirus meningitis have occurred with the sources traced to potable or recreational water sources. Examples include swimming pools (echovirus 30 aseptic meningitis), campsite swimming pool (echovirus 9 aseptic meningitis), nature-like swimming pond (echoviruses 30 and 13), tap and bottled drinking water (echoviruses 30 and 6, CV-B5) [106–109]. Inadequate chlorination might have accounted for some outbreaks; however, the usual levels of chlorination in swimming pools and potable water (2–4 ppm and 1 ppm free chlorine, respectively) do not reliably inactivate picornaviruses. Viable enteroviruses can also be isolated from bottled and tap water (vaccine strain of poliovirus, Coxsackievirus B, echoviruses) [110–112]. The waterborne outbreak in Belarus in 2003, mainly caused by Echovirus 30 and to a lesser extent, Echovirus 6, was one of the largest outbreaks, affecting 1222 children; 57·5% of the patients developed meningitis [109]. Interestingly, during the 1998 Taiwan outbreak, usage of tap water was found to be a risk factor for acquisition of HFMD or herpangina in univariate analysis [56]. However, the study did not specify whether the contact involved drinking unboiled tap water, rinsing of mouth during tooth brushing or face washing. The role of cross-infection caused by backwashing of infected saliva onto drinking fountains used in schools and other public facilities has not been investigated.

In addition to drinking water, sewage contamination of coastal and marine waters can also lead to accumulation of enteroviruses in molluscs. Clams, mussels, oysters, and crabs have been found to have a high prevalence of enteroviruses (ranging from 8% to 40% in some studies), frequently in the presence of a normal coliform count in the samples [112–116]. Foodborne transmission of HEV-71 in humans has not been demonstrated. However, it would not be surprising if this turns out to be an important route of transmission in settings where raw shellfish is frequently consumed. Apart from hepatitis A virus, another picornavirus, Aichi virus, has been found to be involved in causing outbreaks of gastroenteritis related to consumption of oysters and other seafood in France, and the seroprevalence among adults in the general populations was up to 85% [117–119].

Although the role of environmental viruses in the transmission and maintenance of HEV-71 infection remains to be defined, their contribution needs to be further studied, especially in developing countries where sewage treatment, quality of potable water supply, and food hygiene could be suboptimal. Nevertheless, these environmental factors will not be the sole determinants of the epidemiology of HEV-71 infections. Some developing countries such as India has not reported major outbreaks, whereas high prevalence of enteroviruses has been found in potable water in developed countries like Korea, and epidemics of HEV-71 HFMD have occurred in developed countries such as Singapore and Taiwan [120]. The policy of surveillance for HFMD and HEV-71 and class suspension in cases of school outbreaks may have helped to limit the size of outbreaks in several of the endemic Southeast Asian countries [121, 122]. Various control measures at epidemic sites such as school closure or discontinuation of drinking fountains have not been adequately evaluated. Such measures may theoretically reduce interpersonal transmission at these sites and so prevent further community spread of the virus.

The peak season for HFMD and HEV-71 infection is generally the summer months. Epidemics of HEV-71 infection tend to occur once every 2–5 years. The cyclical appearance of epidemics could be due to the accumulation of susceptible individuals in the community. On the other hand, introduction of new genotypes or emergence of new genotypes or strains has led to outbreaks in the Asia Pacific region [64]. Studies in Taiwan showed that the level of seroprevalence against HEV-71 in different parts of the island and the age-specific seroprevalence are correlated with the incidence of severe disease and mortality rates [56]. Emergence of a new genotype or subgenotype of HEV-71 could contribute to the occurrence of outbreaks. The two changes in the prevailing genotype of HEV-71 in Taiwan can be seen in Table 3, with the epidemics in 1998 and 2000 being caused by C2 and B4 strains, respectively [123]. Similarly, surveillance studies in Sarawak also demonstrated the emergence of a new subgenotype C1 in the 2003 epidemic [124]. New variants of the virus can also be generated from intra-typic and inter-typic recombinations of the pre-existing viruses (Fig. 2) [22, 125, 126]. Nevertheless, epidemic genotypes or subgenotypes of the virus could circulate for a long time before causing outbreaks. Similarly, the introduction of new genotypes into a susceptible population does not always result in large epidemics. The lack of herd immunity towards the prevailing strains of HEV-71 is apparently not sufficient to generate an epidemic. Virulence of the viral strains could also be pivotal. However, this area has not been fully studied. A comparison of the HEV-71 strains from fatal and non-fatal cases in Taiwan showed that they have a very high degree of genomic identity, with only minor differences in the homology of the 3C protease [127]. The 3C protease induces apoptosis in human neural cells *in vitro*, but its role in determining the virulence of the virus has not been documented [128]. Neither has the role of temperature-sensitivity of the virus been shown to be of significance in determining virulence in an animal model [129]. The contribution of micronutrient deficiencies (such as selenium and vitamin E) in determining the outcome of HEV-71 infection is uncertain.

TREATMENT, PREVENTION, AND CONTROL

Most HFMD cases are self-limiting and only required supportive treatment. The case-fatality rate for HFMD of all causes ranges from 0·06% to 0·11% [61]. The rapid progression to neurological and cardiopulmonary complications after the onset of HEV-71-associated disease (usually within 3–5 days) suggests that viral replication and direct cytopathic effects of the virus on the host cells are important in the pathogenesis of severe manifestations [45, 67, 130]. Thus, early administration of an effective antiviral agent should theoretically be beneficial. There is currently no specific antiviral approved for HEV-71 infections. Of the available antivirals, pleconaril is the best studied agent against picornaviruses. It binds to the hydrophobic pocket in the viral capsid VP1, thereby inhibiting the attachment, entry, and uncoating of the virus [131]. Pleconaril is readily absorbed after oral administration and penetrates well into body fluids including the cerebrospinal fluid. However, the drug is not readily available in most countries and there are limited data showing that it lacks *in vitro* activity against HEV-71 [132, 133]. Pleconaril has been used in a small number of patients suffering form HEV-71 encephalitis and pulmonary oedema but its efficacy cannot be ascertained to date [133, 134]. In one study, ribavirin showed antiviral activity against HEV-71 *in vitro* and in animal models but there are no reports of its use for treating human EV-71 infections [135]. One animal experiment suggested that type I interferons may have a useful therapeutic role in EV-71 infection [136].

Agammaglobulinaemic individuals are prone to the development of chronic enteroviral meningoencephalitis. Prophylaxis and treatment with intravenous immunoglobulin (IVIG) has been successful in this group of patients and there are anecdotal reports of successful treatment of enteroviral meningoencephalitis in other immuncompromised patients [137–139]. Since maternal antibody appears to be protective against enteroviruses including HEV-71, transfusion of maternal plasma had been used in a case of neonatal disseminated echovirus infection [140]. IVIG has also been used in patients with complicated HEV-71 infections; in addition to the presence of neutralizing antibodies and hence the suppression of viral replication, immunoglobulin may also play a role in limiting organ damage through its anti-inflammatory activities [133, 141–143]. However, its benefits have not yet been demonstrated in randomized controlled trials. Different preparations of IVIG also vary in their titres of antibodies against enteroviruses [144]. Based on the deleterious effects

of dexamethasone in animals, control of the inflammatory response using systemic corticosteroids cannot be recommended at present [136]. The use of milrinone – a bipyridine inotropic agent used in the treatment of heart failure – has been recommended based on a historically controlled study in Taiwan which demonstrated clinical and survival benefits in milrinone-treated patients with pulmonary oedema due to severe HEV-71 infection [145, 146].

With the observation that neutralizing antibodies offer protection against infection and mortality in humans and animal models, vaccination is an obvious pathway towards prevention of HEV-71 infection and epidemics. Initial work on various models of HEV-71 vaccines (using VP1 subunits, inactivated viruses, DNA vaccines, virus-like particles, or oral vaccination with VP1 protein) in animals appears promising, although still in the early stages of development [147–151]. Animal studies with attenuated EV-71 vaccine and human seroepidemiological studies suggested there may be cross-protecting neutralizing antibodies between different EV-71 genotypes [152, 153]. However, the relevance of these findings to the development of vaccines (in terms of the choice of and the need periodically to change vaccine strains) is uncertain. Rapid changes in *VP1* as a result of recombination necessitate a robust surveillance of circulating viruses. The generally mild disease manifestations, prevalence and lack of monitoring in some developing countries means the incentive for vaccine development could be low. A novel potential means of preventing HEV-71 infection is the use of oral lactoferrin. *In vitro*, lactoferrin inhibits binding of EV-71 to cells and neonatal mice fed with recombinant porcine lactoferrin were protected against lethal infection by EV-71 [154–156].

In the absence of vaccine, prevention of human EV-71 infections still relies on basic infection control measures, especially in schools and childcare centres. Monitoring for the incidence of HEV-71-associated diseases such as HFMD and herpangina is essential, as well as virological studies to detect emergence of new genotypes of the virus.

Proper environmental disinfection in school and childcare centres and avoidance of possible public sources of viral acquisition may be necessary [104, 106, 108, 157, 158]. Only one study specifically examined the efficacy of a peroxygen disinfectant on HEV-71 [159]. In general, effective environmental disinfectants against picornaviruses include sodium hypochlorite, glutaraldehyde, chlorine dioxide, and peroxygen compounds [159–162]. As an antiseptic, povidone iodine is highly effective against non-enveloped viruses [163]. Alcohol hand rubs commonly used in healthcare settings contain 60–70% v/v alcohols (such as ethanol, isopropanol, and *n*-propanol). These preparations are less effective in inactivating non-enveloped viruses than bacteria and enveloped viruses. Higher concentrations of alcohol (such as 85–95% v/v ethanol) are required for reliable virucidal activities against non-enveloped viruses [164]. Alternative hand disinfectants that can be considered include 0·2% peracetic acid with 80% (v/v) ethanol and other newer combination alcohol-based formulations [165–167]. These products should be used in situations where active outbreaks of enterovirus infection are ongoing.

Countries in the Asia Pacific region in particular should develop national plans for future epidemics of HEV-71 infection. In Hong Kong, for example, the following strategies have been developed in anticipation of possible community outbreaks: surveillance and laboratory support; clinical management and infection control in healthcare settings; emergency preparedness (including stepping up of surveillance and case investigation, mobilization of clinical and intensive-care facilities, enhanced laboratory support, data management, outbreak control measures, and risk communication); health education and capacity building; and applied research [168].

CONCLUSION

In the past decade, HFMD due to HEV-71 has become a major public health concern in the Asia Pacific region, which appeared to be the epicentre for the generation of epidemic genotypes (as with influenza). Control and prevention of the disease is a difficult task because of the stability of the virus in the environment and its high transmissibility, frequency of genetic recombinations, the lack of an effective antiviral and vaccine, and the relatively low priority for vaccine development. Development and studies in the only available anti-picornavirus agent pleconaril have been minimal towards HEV-71, and newer technologies for screening antiviral compounds, for example, using chemical genetics and screening of chemical libraries, may help in the discovery of a novel agent for treatment [169]. The contributions of genetic, environmental, and viral factors in the genesis of epidemics in different regions of the world need to be clarified in order to understand the varying

epidemiology of the disease in different countries. Similarly, the role of public health measures to reduce the impact of outbreaks, such as school closure, warrant further studies.

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REFERENCES

1. **Picornaviridae Study Group.** (http://www.picornastudygroup.com/types/index.html). Accessed 22 September 2009.
2. **Brown B, et al.** Complete genomic sequencing shows that polioviruses and members of human enterovirus species C are closely related in the noncapsid coding region. *Journal of Virology* 2003; **77**: 8973–8984.
3. **Wolthers KC, et al.** Human parechoviruses as an important viral cause of sepsis-like illness and meningitis in young children. *Clinical Infectious Diseases* 2008; **47**: 358–363.
4. **Zoll J, et al.** Saffold virus, a human Theiler's-like cardiovirus, is ubiquitous and causes infection early in life. *PLoS Pathogens* 2009; **5**: e1000416.
5. **Holtz LR, et al.** Klassevirus 1, a previously undescribed member of the family *Picornaviridae*, is globally widespread. *Virology Journal* 2009; **6**: 86.
6. **Kapoor A, et al.** A highly prevalent and genetically diversified *Picornaviridae* genus in South Asian children. *Proceedings of the National Academy of Sciences USA* 2008; **105**: 20482–20487.
7. **Holtz LR, et al.** Identification of a novel picornavirus related to cosaviruses in a child with acute diarrhea. *Virology Journal* 2008; **5**: 159.
8. **Li L, et al.** A novel picornavirus associated with gastroenteritis. *Journal of Virology* 2009; **83**: 12002–12006.
9. **Muir P, et al.** Molecular typing of enteroviruses: current status and future requirements. The European Union concerted action on virus meningitis and encephalitis. *Clinical Microbiology Reviews* 1998; **11**: 202–227.
10. **McMinn PC.** An overview of the evolution of enterovirus 71 and its clinical and public health significance. *FEMS Microbiology Reviews* 2002; **26**: 91–107.
11. **Chua BH, et al.** The molecular basis of mouse adaptation by human enterovirus 71. *Journal of General Virology* 2008; **89**: 1622–1632.
12. **Chan YF, Sam IC, AbuBakar S.** Phylogenetic designation of enterovirus 71 genotypes and subgenotypes using complete genome sequences. *Infection, Genetics and Evolution* 2009 (in press).
13. **Van der Sanden S, et al., on behalf of the Dutch Working Group for Clinical Virology.** Epidemiology of enterovirus 71 in the Netherlands, 1963–2008. *Journal of Clinical Microbiology* 2009; **47**: 2826–2833.
14. **Cuervo NS, et al.** Genomic features of intertypic recombinant Sabin poliovirus strains excreted by primary vaccinees. *Journal of Virology* 2001; **75**: 5740–5751.
15. **Lukashev AN.** Role of recombination in evolution of enteroviruses. *Reviews in Medical Virology* 2005; **15**: 157–167.
16. **Santti J, et al.** Evidence of recombination among enteroviruses. *Journal of Virology* 1999; **73**: 8741–8749.
17. **Oprisan G, et al.** Natural genetic recombination between co-circulating heterotypic enteroviruses. *Journal of General Virology* 2002; **83**: 2193–2200.
18. **Lukashev AN, et al.** Recombination in circulating enteroviruses. *Journal of Virology* 2003; **77**: 10423–10431.
19. **Oberste MS, Peñaranda S, Pallansch MA.** RNA recombination plays a major role in genomic change during circulation of coxsackie B viruses. *Journal of Virology* 2004; **78**: 2948–2955.
20. **Simmonds P, Welch J.** Frequency and dynamics of recombination within different species of human enteroviruses. *Journal of Virology* 2006; **80**: 483–493.
21. **Chan YF, AbuBaker S.** Recombinant human enterovirus 71 in hand, foot and mouth disease patients. *Emerging Infectious Diseases* 2004; **10**: 1468–1470.
22. **Ding NZ, et al.** Appearance of mosaic enterovirus 71 in the 2008 outbreak of China. *Virus Research* 2009; **145**: 157–161.
23. **Lin YW, et al.** Enterovirus 71 infection of human dendritic cells. *Experimental Biology and Medicine (Maywood)* 2009; **234**: 1166–1173.
24. **Nishimura Y, et al.** Human P-selectin glycoprotein ligand-1 is a functional receptor for enterovirus 71. *Nature Medicine* 2009; **15**: 794–797.
25. **Yamayoshi S, et al.** Scavenger receptor B2 is a cellular receptor for enterovirus 71. *Nature Medicine* 2009; **15**: 798–801.
26. **Moore KL.** Structure and function of P-selectin glycoprotein ligand-1. *Leukemia and Lymphoma* 1998; **29**: 1–15.
27. **Mulcahy JV, Riddell DR, Owen JS.** Human scavenger receptor class B type II (SR-BII) and cellular cholesterol efflux. *Biochemical Journal* 2004; **377**: 741–747.
28. **Webb NR, et al.** SR-BII, an isoform of the scavenger receptor BI containing an alternate cytoplasmic tail, mediates lipid transfer between high density lipoprotein

and cells. *Journal of Biological Chemistry* 1998; **273**: 15241–15248.

29. **Thilakawardhana S, et al.** Quantification of apolipo-protein E receptors in human brain-derived cell lines by real-time polymerase chain reaction. *Neurobiology of Aging* 2005; **26**: 813–823.

30. **Eckhardt ER, et al.** High density lipoprotein endo-cytosis by scavenger receptor SR-BII is clathrin-dependent and requires a carboxyl-terminal dileucine motif. *Journal of Biological Chemistry* 2006; **281**: 4348–4353.

31. **Brodeur MR, et al.** Scavenger receptor of class B ex-pressed by osteoblastic cells are implicated in the uptake of cholesteryl ester and estradiol from LDL and HDL3. *Journal of Bone and Mineral Research* 2008; **23**: 326–337.

32. **Grove J, et al.** Scavenger receptor BI and BII expression levels modulate hepatitis C virus infectivity. *Journal of Virology* 2007; **81**: 3162–3169.

33. **Barth H, et al.** Scavenger receptor class B is required for hepatitis C virus uptake and cross-presentation by human dendritic cells. *Journal of Virology* 2008; **82**: 3466–3479.

34. **Mueller S, Wimmer E, Cello J.** Poliovirus and polio-myelitis: a tale of guts, brains, and an accidental event. *Virus Research* 2005; **111**: 175–193.

35. **Racaniello VR.** One hundred years of poliovirus pathogenesis. *Virology* 2006; **344**: 9–16.

36. **Wong KT, et al.** The distribution of inflammation and virus in human enterovirus 71 encephalomyelitis suggests possible viral spread by neural pathways. *Journal of Neuropathology and Experimental Neurology* 2008; **67**: 162–169.

37. **Chen CS, et al.** Retrograde axonal transport: a major transmission route of enterovirus 71 in mice. *Journal of Virology* 2007; **81**: 8996–9003.

38. **Wen YY, et al.** Comparative study of enterovirus 71 in-fection of human cell lines. *Journal of Medical Virology* 2003; **70**: 109–118.

39. **Kuo RL, et al.** Infection with enterovirus 71 or ex-pression of its 2A protease induces apoptotic cell death. *Journal of General Virology* 2002; **83**: 1367–1376.

40. **Chan YF, Abubakar S.** Enterovirus 71 infection induces apoptosis in Vero cells. *Malaysian Journal of Pathology* 2003; **25**: 29–35.

41. **Chang SC, et al.** Diverse apoptotic pathways in enterovirus 71-infected cells. *Journal of Neurovirology* 2004; **10**: 338–349.

42. **Liang CC, et al.** Human endothelial cell activation and apoptosis induced by enterovirus 71 infection. *Journal of Medical Virology* 2004; **74**: 597–603.

43. **Chen TC, et al.** Enterovirus 71 triggering of neuronal apoptosis through activation of Abl-Cdk5 signalling. *Cellular Microbiology* 2007; **9**: 2676–2688.

44. **Huang SC, et al.** Enterovirus 71-induced autophagy detected *in vitro* and *in vivo* promotes viral repli-cation. *Journal of Medical Virology* 2009; **81**: 1241–1252.

45. **Chan LG, et al.** Deaths of children during an out-break of hand, foot, and mouth disease in Sarawak, Malaysia: clinical and pathological characteristics of the disease. For the Outbreak Study Group. *Clinical Infectious Diseases* 2000; **31**: 678–683.

46. **Lum LC, et al.** Fatal enterovirus 71 encephalomyelitis. *Journal of Pediatrics* 1998; **133**: 795–798.

47. **Yang Y, et al.** Neuropathology in 2 cases of fatal en-terovirus type 71 infection from a recent epidemic in the People's Republic of China: a histopathologic, immunohistochemical, and reverse transcription poly-merase chain reaction study. *Human Pathology* 2009; **40**: 1288–1295.

48. **Fu YC, et al.** Cardiac complications of enterovirus rhombencephalitis. *Archives of Disease in Childhood* 2004; **89**: 368–373.

49. **Shieh WJ, et al.** Pathologic studies of fatal cases in outbreak of hand, foot, and mouth disease, Taiwan. *Emerging Infectious Diseases* 2001; **7**: 146–148.

50. **Lin YW, et al.** Lymphocyte and antibody responses reduce enterovirus 71 lethality in mice by decreasing tissue viral loads. *Journal of Virology* 2009; **83**: 6477–6483.

51. **Yang KD, et al.** Altered cellular but not humoral reac-tions in children with complicated enterovirus 71 infec-tions in Taiwan. *Journal of Infectious Diseases* 2001; **183**: 850–856.

52. **Chang LY, et al.** Status of cellular rather than humoral immunity is correlated with clinical outcome of entero-virus 71. *Pediatric Research* 2006; **60**: 466–471.

53. **Yu CK, et al.** Neutralizing antibody provided pro-tection against enterovirus type 71 lethal challenge in neonatal mice. *Journal of Biomedical Science* 2000; **7**: 523–528.

54. **Foo DG, et al.** Passive protection against lethal en-terovirus 71 infection in newborn mice by neutralizing antibodies elicited by a synthetic peptide. *Microbes and Infection* 2007; **9**: 1299–1306.

55. **Wu TC, et al.** Immunity to avirulent enterovirus 71 and coxsackie A16 virus protects against enterovirus 71 infection in mice. *Journal of Virology* 2007; **81**: 10310–10315.

56. **Chang LY, et al.** Risk factors of enterovirus 71 infec-tion and associated hand, foot, and mouth disease/ herpangina in children during an epidemic in Taiwan. *Pediatrics* 2002; **109**: e88.

57. **Ho M, et al.** An epidemic of enterovirus 71 infection in Taiwan. Taiwan Enterovirus Epidemic Working Group. *New England Journal of Medicine* 1999; **341**: 929–935.

58. **Luo ST, et al.** Enterovirus 71 maternal antibodies in infants, Taiwan. *Emerging Infectious Diseases* 2009; **15**: 581–584.

59. **Chiu CH, et al.** Protection of neonatal mice from lethal enterovirus 71 infection by maternal immunization with attenuated *Salmonella enterica* serovar Typhimurium expressing VP1 of enterovirus 71. *Microbes and Infec-tion* 2006; **8**: 1671–1678.

60. **Chang LY, et al.** Comparison of enterovirus 71 and coxsackie-virus A16 clinical illnesses during the Taiwan enterovirus epidemic, 1998. *Pediatric Infectious Disease Journal* 1999; **18**: 1092–1096.

61. **Chong CY, et al.** Hand, foot and mouth disease in Singapore: a comparison of fatal and non-fatal cases. *Acta Paediatrica* 2003; **92**: 1163–1169.

62. **Tai WC, Hsieh HJ, Wu MT.** Hand, foot and mouth disease in a healthy adult caused by intrafamilial transmission of enterovirus 71. *British Journal of Dermatolology* 2009; **160**: 890–892.

63. **Hamaguchi T, et al.** Acute encephalitis caused by intrafamilial transmission of enterovirus 71 in adult. *Emerging Infectious Diseases* 2008; **14**: 828–830.

64. **Cardosa MJ, et al.** Molecular epidemiology of human enterovirus 71 strains and recent outbreaks in the Asia-Pacific region: comparative analysis of the *VP1* and *VP4* genes. *Emerging Infectious Diseases* 2003; **9**: 461–468.

65. **Centers for Disease Control, Republic of China (Taiwan).** The activity of human enterovirus in Taiwan between 1998 and 2006 (http://www.cdc.gov.tw/public/Attachment/7121014355971.pdf). Accessed 22 July 2009.

66. **Chang LY, et al.** Transmission and clinical features of enterovirus 71 infections in household contacts in Taiwan. *Journal of the American Medical Association* 2004; **291**: 222–227.

67. **Yang TT, et al.** Clinical features and factors of unfavorable outcomes for non-polio enterovirus infection of the central nervous system in northern Taiwan, 1994–2003. *Journal of Microbiology, Immunology, and Infection* 2005; **38**: 417–424.

68. **Huang CC, et al.** Neurologic complications in children with enterovirus 71 infection. *New England Journal of Medicine* 1999; **341**: 936–942.

69. **Chang LY, et al.** Neurodevelopment and cognition in children after enterovirus 71 infection. *New England Journal of Medicine* 2007; **356**: 1226–1234.

70. **Huang MC, et al.** Long-term cognitive and motor deficits after enterovirus 71 brainstem encephalitis in children. *Pediatrics* 2006; **118**: e1785–1788.

71. **Kao SJ, et al.** Mechanism of fulminant pulmonary edema caused by enterovirus 71. *Clinical Infectious Diseases* 2004; **38**: 1784–1788.

72. **Chang LY, et al.** Clinical features and risk factors of pulmonary oedema after enterovirus-71-related hand, foot, and mouth disease. *Lancet* 1999; **354**: 1682–1686.

73. **Li CC, et al.** Clinical manifestations and laboratory assessment in an enterovirus 71 outbreak in southern Taiwan. *Scandinavian Journal of Infectious Diseases* 2002; **34**: 104–109.

74. **Yang TT, et al.** Clinical features and factors of unfavorable outcomes for non-polio enterovirus infection of the central nervous system in northern Taiwan, 1994–2003. *Journal of Microbiology, Immunology, and Infection* 2005; **38**: 417–424.

75. **Hsia SH, et al.** Predictors of unfavorable outcomes in enterovirus 71-related cardiopulmonary failure in children. *Pediatric Infectious Disease Journal* 2005; **24**: 331–334.

76. **Iturriza-Gómara M, Megson B, Gray J.** Molecular detection and characterization of human enteroviruses directly from clinical samples using RT-PCR and DNA sequencing. *Journal of Medical Virology* 2006; **78**: 243–253.

77. **Leitch EC, et al.** Direct identification of human enterovirus serotypes in cerebrospinal fluid by amplification and sequencing of the *VP1* region. *Journal of Clinical Virology* 2009; **44**: 119–124.

78. **Nix WA, Oberste MS, Pallansch MA.** Sensitive, seminested PCR amplification of VP1 sequences for direct identification of all enterovirus serotypes from original clinical specimens. *Journal of Clinical Microbiology* 2006; **44**: 2698–2704.

79. **Terletskaia-Ladwig E, et al.** A convenient rapid culture assay for the detection of enteroviruses in clinical samples: comparison with conventional cell culture and RT-PCR. *Journal of Medical Microbiology* 2008; **57**: 1000–1006.

80. **Tsao KC, et al.** Responses of IgM for enterovirus 71 infection. *Journal of Medical Virology* 2002; **68**: 574–580.

81. **Blomberg J, et al.** New enterovirus type associated with epidemic of aseptic meningitis and-or hand, foot, and mouth disease. *Lancet* 1974; **2**: 112.

82. **Hagiwara A, Tagaya I, Yoneyama T.** Epidemic of hand, foot and mouth disease associated with enterovirus 71 infection. *Intervirology* 1978; **9**: 60–63.

83. **Chumakov M, et al.** Enterovirus 71 isolated from cases of epidemic poliomyelitis-like disease in Bulgaria. *Archives of Virology* 1979; **60**: 329–340.

84. **Nagy G, et al.** Virological diagnosis of enterovirus type 71 infections: experiences gained during an epidemic of acute CNS diseases in Hungary in 1978. *Archives of Virology* 1982; **71**: 217–227.

85. **Chang LY, et al.** HLA-A33 is associated with susceptibility to enterovirus 71 infection. *Pediatrics* 2008; **122**: 1271–1276.

86. **Ho HY, et al.** Glucose-6-phosphate dehydrogenase deficiency enhances enterovirus 71 infection. *Journal of General Virology* 2008; **89**: 2080–2089.

87. **Le Jossec M, et al.** Genetic diversity patterns in the SR-BI/II locus can be explained by a recent selective sweep. *Molecular Biology and Evolution* 2004; **21**: 760–769.

88. **Mata LJ, Urrutia JJ, Lechtig A.** Infection and nutrition of children of a low socioeconomic rural community. *American Journal of Clinical Nutrition* 1971; **24**: 249–259.

89. **Beck MA, Matthews CC.** Micronutrients and host resistance to viral infection. *Proceedings of the Nutrition Society* 2000; **59**: 581–585.

90. **Beck MA, Williams-Toone D, Levander OA.** Coxsackievirus B3-resistant mice become susceptible in Se/vitamin E deficiency. *Free Radical Biology and Medicine* 2003; **34**: 1263–1270.

91. **Cermelli C, et al.** Selenite inhibition of Coxsackie virus B5 replication: implications on the etiology of Keshan disease. *Journal of Trace Elements in Medicine and Biology* 2002; **16**: 41–46.

92. **Chan KP, et al.** Epidemic hand, foot and mouth disease caused by human enterovirus 71, Singapore. *Emerging Infectious Diseases* 2003; **9**: 78–85.

93. **Ooi MH, et al.** Human enterovirus 71 disease in Sarawak, Malaysia: a prospective clinical, virological, and molecular epidemiological study. *Clinical Infectious Diseases* 2007; **44**: 646–656.

94. **Chinese Center for Disease Control and Prevention and the Office of the World Health Organization in China.** Report on the hand, foot and mouth disease outbreak in Fuyang City, Anhui Province and the prevention and control in China, May 2008 (http://www.wpro.who.int/NR/rdonlyres/591D6A7B-FB15-4E94-A1E9-1D3381847D60/0/HFMDCCDC20080515ENG.pdf). Accessed 29 July 2009.

95. **Chung PW, et al.** Duration of enterovirus shedding in stool. *Journal of Microbiology, Immunology, and Infection* 2001; **34**: 167–170.

96. **Tosato G, Rocchi G, Archetti I.** Epidemiological study of a 'hand-foot-and-mouth disease' outbreak observed in Rome in the fall of 1973. *Zentralblatt für Bakteriologie, Parasitenkunde, Infektionskrankheiten und Hygiene. Erste Abteilung Originale. Reihe A: Medizinische Mikrobiologie und Parasitologie* 1975; **230**: 415–421.

97. **Wang JR, et al.** An outbreak of enterovirus 71 infection in Taiwan, 1998. II. Laboratory diagnosis and genetic analysis. *Journal of Clinical Virology* 2000; **17**: 91–99.

98. **Ooi MH, et al.** Evaluation of different clinical sample types in diagnosis of human enterovirus 71-associated hand-foot-and-mouth disease. *Journal of Clinical Microbiology* 2007; **45**: 1858–1866.

99. **Kuramitsu M, et al.** Non-polio enterovirus isolation among families in Ulaanbaatar and Tov province, Mongolia: prevalence, intrafamilial spread, and risk factors for infection. *Epidemiology and Infection* 2005; **133**: 1131–1142.

100. **Welch JB, et al.** Detection of enterovirus viraemia in blood donors. *Vox Sanguinis* 2001; **80**: 211–215.

101. **Donoso Mantke O, et al.** High prevalence of cardiotropic viruses in myocardial tissue from explanted hearts of heart transplant recipients and heart donors: a 3-year retrospective study from a German patients' pool. *Journal of Heart and Lung Transplantation* 2005; **24**: 1632–1638.

102. **Chow KC, et al.** Congenital enterovirus 71 infection: a case study with virology and immunohistochemistry. *Clinical Infectious Diseases* 2000; **31**: 509–512.

103. **Nishikii Y, et al.** Favorable outcome in a case of perinatal enterovirus 71 infection. *Pediatric Infectious Disease Journal* 2002; **21**: 886–887.

104. **Hsu BM, Chen CH, Wan MT.** Prevalence of enteroviruses in hot spring recreation areas of Taiwan. *FEMS Immunology and Medical Microbiology* 2008; **52**: 253–259.

105. **Chen CH, Hsu BM, Wan MT.** Molecular detection and prevalence of enterovirus within environmental water in Taiwan. *Journal of Applied Microbiology* 2008; **104**: 817–823.

106. **Faustini A, et al.** An outbreak of aseptic meningitis due to echovirus 30 associated with attending school and swimming in pools. *International Journal of Infectious Diseases* 2006; **10**: 291–297.

107. **Centers for Disease Control and Prevention (CDC).** Aseptic meningitis outbreak associated with echovirus 9 among recreational vehicle campers – Connecticut, 2003. *Morbidity and Mortality Weekly Report* 2004; **53**: 710–713.

108. **Hauri AM, et al.** An outbreak of viral meningitis associated with a public swimming pond. *Epidemiology and Infection* 2005; **133**: 291–298.

109. **Amvrosieva TV, et al.** Enteroviral infection outbreak in the Republic of Belarus: principal characteristics and phylogenetic analysis of etiological agents. *Central European Journal of Public Health* 2006; **14**: 67–73.

110. **Lee SH, Kim SJ.** Detection of infectious enteroviruses and adenoviruses in tap water in urban areas in Korea. *Water Research* 2002; **36**: 248–256.

111. **Vivier JC, Ehlers MM, Grabow WO.** Detection of enteroviruses in treated drinking water. *Water Research* 2004; **38**: 2699–2705.

112. **Ehlers MM, Grabow WO, Pavlov DN.** Detection of enteroviruses in untreated and treated drinking water supplies in South Africa. *Water Research* 2005; **39**: 2253–2258.

113. **Croci L, et al.** Determination of enteroviruses, hepatitis A virus, bacteriophages and *Escherichia coli* in Adriatic Sea mussels. *Journal of Applied Microbiology* 2000; **88**: 293–298.

114. **Beuret C, Baumgartner A, Schluep J.** Virus-contaminated oysters: a three-month monitoring of oysters imported to Switzerland. *Applied and Environmental Microbiology* 2003; **69**: 2292–2297.

115. **Dubois E, et al.** Diversity of enterovirus sequences detected in oysters by RT-heminested PCR. *International Journal of Food Microbiology* 2004; **92**: 35–43.

116. **Gabrieli R, et al.** Enteric viruses in molluscan shellfish. *New Microbiologica* 2007; **30**: 471–475.

117. **Le Guyader FS, et al.** Aichi virus, norovirus, astrovirus, enterovirus, and rotavirus involved in clinical cases from a French oyster-related gastroenteritis outbreak. *Journal of Clinical Microbiology* 2008; **46**: 4011–4017.

118. **Ambert-Balay K, et al.** Prevalence and genetic diversity of Aichi virus strains in stool samples from community and hospitalized patients. *Journal of Clinical Microbiology* 2008; **46**: 1252–1258.

119. **Goyer M, et al.** Seroprevalence distribution of Aichi virus among a French population in 2006–2007. *Archives of Virology* 2008; **153**: 1171–1174.

120. **Saoji VA.** Hand, foot and mouth disease in Nagpur. *Indian Journal of Dermatolology, Venereology and Leprology* 2008; **74**: 133–135.

121. **Centre for Health Protection, Department of Health, Hong Kong.** EV scan (http://www.chp.gov.hk/guideline1_year.asp?lang=en&id=502&pid=441&ppid=134&pppid=29). Accessed 28 July 2009.

122. **Centre for Health Protection, Department of Health, Hong Kong.** Guidelines on prevention of

communicable diseases in schools/kindergartens/ kindergartens-cum-child care centres/child care centres (revised January 2009) (http://www.chp.gov.hk/ files/pdf/School_full_eng_20090115.pdf). Accessed 28 July 2009.

123. **Wang JR, et al.** Change of major genotype of enterovirus 71 in outbreaks of hand-foot-and-mouth disease in Taiwan between 1998 and 2000. *Journal of Clinical Microbiology* 2002; **40**: 10–15.

124. **Podin Y, et al.** Sentinel surveillance for human enterovirus 71 in Sarawak, Malaysia: lessons from the first 7 years. *BMC Public Health* 2006; **6**: 180.

125. **Huang SC, et al.** Appearance of intratypic recombination of enterovirus 71 in Taiwan from 2002 to 2005. *Virus Research* 2008; **131**: 250–259.

126. **Chan YF, AbuBakar S.** Phylogenetic evidence for inter-typic recombination in the emergence of human enterovirus 71 subgenotypes. *BMC Microbiology* 2006; **6**: 74.

127. **Shih SR, et al.** Genetic analysis of enterovirus 71 isolated from fatal and non-fatal cases of hand, foot and mouth disease during an epidemic in Taiwan, 1998. *Virus Research* 2000; **68**: 127–136.

128. **Li ML, et al.** The 3C protease activity of enterovirus 71 induces human neural cell apoptosis. *Virology* 2002; **293**: 386–395.

129. **Hagiwara A, Yoneyama T, Takami S, Hashimoto I.** Genetic and phenotypic characteristics of enterovirus 71 isolates from patients with encephalitis and with hand, foot and mouth disease. *Archives of Virology* 1984; **79**: 273–283.

130. **Pérez-Vélez CM, et al.** Outbreak of neurologic enterovirus type 71 disease: a diagnostic challenge. *Clinical Infectious Diseases* 2007; **45**: 950–957.

131. **Smith TJ, et al.** The site of attachment in human rhinovirus 14 for antiviral agents that inhibit uncoating. *Science* 1986; **233**: 1286–1293.

132. **Shia KS, et al.** Design, synthesis, and structure-activity relationship of pyridyl imidazolidinones: a novel class of potent and selective human enterovirus 71 inhibitors. *Journal of Medicinal Chemistry* 2002; **45**: 1644–1655.

133. **Nolan MA, et al.** Survival after pulmonary edema due to enterovirus 71 encephalitis. *Neurology* 2003; **60**: 1651–1656.

134. **Prager P, et al.** Neurogenic pulmonary edema in enterovirus 71 encephalitis is not uniformly fatal but causes severe morbidity in survivors. *Pediatric Critical Care Medicine* 2003; **4**: 377–381.

135. **Li ZH, et al.** Ribavirin reduces mortality in enterovirus 71-infected mice by decreasing viral replication. *Journal of Infectious Diseases* 2008; **197**: 854–857.

136. **Liu ML, et al.** Type I interferons protect mice against enterovirus 71 infection. *Journal of General Virology* 2005; **86**: 3263–3269.

137. **McKinney Jr. RE, Katz SL, Wilfert CM.** Chronic enteroviral meningoencephalitis in agammaglobulinemic patients. *Reviews of Infectious Diseases* 1987; **9**: 334–356.

138. **Cheng MF, et al.** Clinical application of reverse-transcription polymerase chain reaction and intravenous immunoglobulin for enterovirus encephalitis. *Japanese Journal of Infectious Diseases* 2008; **61**: 18–24.

139. **Moschovi MA, et al.** Enteroviral infections in children with malignant disease: a 5-year study in a single institution. *Journal of Infection* 2007; **54**: 387–392.

140. **Jantausch BA, et al.** Maternal plasma transfusion in the treatment of disseminated neonatal echovirus 11 infection. *Pediatric Infectious Disease Journal* 1995; **14**: 154–155.

141. **Wang SM, et al.** Clinical spectrum of enterovirus 71 infection in children in southern Taiwan, with an emphasis on neurological complications. *Clinical Infectious Diseases* 1999; **29**: 184–190.

142. **Frange P, et al.** Enterovirus 71 meningoencephalitis during chemotherapy in a child with metastatic osteosarcoma. *Journal of Pediatric Hematology Oncology* 2007; **29**: 566–568.

143. **Wang SM, et al.** Modulation of cytokine production by intravenous immunoglobulin in patients with enterovirus 71-associated brainstem encephalitis. *Journal of Clinical Virology* 2006; **37**: 47–52.

144. **Galama JM, et al.** Antibodies against enteroviruses in intravenous Ig preparations: great variation in titres and poor correlation with the incidence of circulating serotypes. *Journal of Medical Virology* 1997; **53**: 273–276.

145. **Wang SM, et al.** Therapeutic efficacy of milrinone in the management of enterovirus 71-induced pulmonary edema. *Pediatric Pulmonology* 2005; **39**: 219–223.

146. **Wang JN, et al.** Critical management in patients with severe enterovirus 71 infection. *Pediatrics International* 2006; **48**: 250–256.

147. **Wu CN, et al.** Protection against lethal enterovirus 71 infection in newborn mice by passive immunization with subunit VP1 vaccines and inactivated virus. *Vaccine* 2001; **20**: 895–904.

148. **Chen HF, et al.** Oral immunization of mice using transgenic tomato fruit expressing VP1 protein from enterovirus 71. *Vaccine* 2006; **24**: 2944–2951.

149. **Tung WS, et al.** DNA vaccine constructs against enterovirus 71 elicit immune response in mice. *Genetic Vaccines and Therapy* 2007; **5**: 6.

150. **Chung YC, et al.** Immunization with virus-like particles of enterovirus 71 elicits potent immune responses and protects mice against lethal challenge. *Vaccine* 2008; **26**: 1855–1862.

151. **Chen HL, et al.** Expression of VP1 protein in the milk of transgenic mice: a potential oral vaccine protects against enterovirus 71 infection. *Vaccine* 2008; **26**: 2882–2889.

152. **Arita M, et al.** An attenuated strain of enterovirus 71 belonging to genotype a showed a broad spectrum of antigenicity with attenuated neurovirulence in cynomolgus monkeys. *Journal of Virology* 2007; **81**: 9386–9395.

153. **Mizuta K,** *et al.* Cross-antigenicity among EV71 strains from different genogroups isolated in Yamagata, Japan, between 1990 and 2007. *Vaccine* 2009; **27**: 3153–3158.

154. **Lin TY, Chu C, Chiu CH.** Lactoferrin inhibits enterovirus 71 infection of human embryonal rhabdomyosarcoma cells *in vitro*. *Journal of Infectious Diseases* 2002; **186**: 1161–1164.

155. **Weng TY,** *et al.* Lactoferrin inhibits enterovirus 71 infection by binding to VP1 protein and host cells. *Antiviral Research* 2005; **67**: 31–37.

156. **Chen HL,** *et al.* Recombinant porcine lactoferrin expressed in the milk of transgenic mice protects neonatal mice from a lethal challenge with enterovirus type 71. *Vaccine* 2008; **26**: 891–898.

157. **Keswick BH, Gerba CP, Goyal SM.** Occurrence of enteroviruses in community swimming pools. *American Journal of Public Health* 1981; **71**: 1026–1030.

158. **Begier EM,** *et al.* An outbreak of concurrent echovirus 30 and coxsackievirus A1 infections associated with sea swimming among a group of travelers to Mexico. *Clinical Infectious Diseases* 2008; **47**: 616–623.

159. **Chan YF, AbuBakar S.** Virucidal activity of Virkon S on human enterovirus. *Medical Journal of Malaysia* 2005; **60**: 246–248.

160. **Abad FX, Pintó RM, Bosch A.** Disinfection of human enteric viruses on fomites. *FEMS Microbiology Letters* 1997; **156**: 107–111.

161. **Chambon M,** *et al.* Virucidal efficacy of glutaraldehyde against enteroviruses is related to the location of lysine residues in exposed structures of the VP1 capsid protein. *Applied and Environmental Microbiology* 2004; **70**: 1717–1722.

162. **Zoni R,** *et al.* Investigation on virucidal activity of chlorine dioxide. experimental data on feline calicivirus, HAV and Coxsackie B5. *Journal of Preventive Medicine and Hygiene* 2007; **48**: 91–95.

163. **Kawana R,** *et al.* Inactivation of human viruses by povidone-iodine in comparison with other antiseptics. *Dermatology* 1997; **195** (Suppl. 2): 29–35.

164. **Kampf G, Kramer A.** Epidemiologic background of hand hygiene and evaluation of the most important agents for scrubs and rubs. *Clinical Microbiology Reviews* 2004; **17**: 863–893.

165. **Wutzler P, Sauerbrei A.** Virucidal efficacy of a combination of 0·2% peracetic acid and 80% (v/v) ethanol (PAA-ethanol) as a potential hand disinfectant. *Journal of Hospital Infection* 2000; **46**: 304–308.

166. **Kramer A,** *et al.* Virucidal activity of a new hand disinfectant with reduced ethanol content: comparison with other alcohol-based formulations. *Journal of Hospital Infection* 2006; **62**: 98–106.

167. **Macinga DR, Sattar SA, Jaykus LA, Arbogast JW.** Improved inactivation of nonenveloped enteric viruses and their surrogates by a novel alcohol-based hand sanitizer. *Applied and Environmental Microbiology* 2008; **74**: 5047–5052.

168. **Scientific Committee on Enteric Infections and Foodborne Diseases, Centre for Health Protection, Hong Kong.** Strategies for the Prevention and Control of EV71 Infection in Hong Kong (http://www.chp.gov.hk/files/pdf/sas4_ev71_20050927.pdf). Accessed 24 July 2009.

169. **Kao RY,** *et al.* Identification of novel small-molecule inhibitors of severe acute respiratory syndrome-associated coronavirus by chemical genetics. *Chemistry and Biology* 2004; **11**: 1293–1299.

170. **Herrero LJ,** *et al.* Molecular epidemiology of enterovirus 71 in peninsular Malaysia, 1997–2000. *Archives of Virology* 2003; **148**: 1369–1385.

171. **Chua KB,** *et al.* Genetic diversity of enterovirus 71 isolated from cases of hand, foot and mouth disease in the 1997, 2000 and 2005 outbreaks, Peninsular Malaysia. *Malaysian Journal of Pathology* 2007; **29**: 69–78.

172. **Abubakar S,** *et al.* Molecular detection of enteroviruses from an outbreak of hand, foot and mouth disease in Malaysia in 1997. *Scandinavian Journal of Infectious Diseases* 1999; **31**: 331–335.

173. **Liu CC,** *et al.* An outbreak of enterovirus 71 infection in Taiwan, 1998: epidemiologic and clinical manifestations. *Journal of Clinical Virology* 2000; **17**: 23–30.

174. **Chen KT,** *et al.* Epidemiologic features of hand-foot-mouth disease and herpangina caused by enterovirus 71 in Taiwan, 1998–2005. *Pediatrics* 2007; **120**: e244–252.

175. **Sanders SA,** *et al.* Molecular epidemiology of enterovirus 71 over two decades in an Australian urban community. *Archives of Virology* 2006; **151**: 1003–1013.

176. **McMinn P, Stratov I, Nagarajan L, Davis S.** Neurological manifestations of enterovirus 71 infection in children during an outbreak of hand, foot, and mouth disease in Western Australia. *Clinical Infectious Diseases* 2001; **32**: 236–242.

177. **AbuBakar S,** *et al.* Enterovirus 71 outbreak, Brunei. *Emerging Infectious Diseases* 2009; **15**: 79–82.

178. **Ministry of Health, People's Republic of China.** http://www.moh.gov.cn/publicfiles/business/htmlfiles/mohbgt/s3582/200902/39079.htm. Accessed on 22 July 2009.

179. **Yang F,** *et al.* Enterovirus 71 outbreak in the People's Republic of China in 2008. *Journal of Clinical Microbiology* 2009; **47**: 2351–2352.

180. **Ministry of Health, People's Republic of China.** March (http://www1.www.gov.cn/gzdt/2009-04/10/content_1282418.htm); April (http://www1.www.gov.cn/gzdt/2009-05/11/content_1311009.htm); May (http://www1.www.gov.cn/gzdt/2009-06/11/content_1337632.htm); June (http://www1.www.gov.cn/gzdt/2009-07/15/content_1366225.htm). Accessed 22 July 2009.

181. **Li L,** *et al.* Genetic characteristics of human enterovirus 71 and coxsackievirus A16 circulating from 1999 to 2004 in Shenzhen, People's Republic of China. *Journal of Clinical Microbiology* 2005; **43**: 3835–3839.

182. **Lin KH,** *et al.* Evolution of EV71 genogroup in Taiwan from 1998 to 2005: an emerging of sub-genogroup C4 of EV71. *Journal of Medical Virology* 2006; **78**: 254–262.

183. **Hosoya M, *et al.*** Genetic diversity of enterovirus 71 associated with hand, foot and mouth disease epidemics in Japan from 1983 to 2003. *Pediatric Infectious Disease Journal* 2006; **25**: 691–694.

184. **Tu PV, *et al.*** Epidemiologic and virologic investigation of hand, foot, and mouth disease, southern Vietnam, 2005. *Emerging Infectious Diseases* 2007; **13**: 1733–1741.

185. **Huang YP, *et al.*** The circulation of subgenogroups B5 and C5 of enterovirus 71 in Taiwan from 2006 to 2007. *Virus Research* 2008; **137**: 206–212.

186. **Ang LW, *et al.*** Epidemiology and control of hand, foot and mouth disease in Singapore, 2001–2007. *Annals of the Academy of Medicine Singapore* 2009; **38**: 106–112.