



Title	SiRNA and shRNA screens advance key understanding of host factors required for HIV-1 replication
Author(s)	Kok, KH; Lei, T; Jin, DY
Citation	Retrovirology, 2009, v. 6, p. 78
Issued Date	2009
URL	http://hdl.handle.net/10722/68211
Rights	Creative Commons: Attribution 3.0 Hong Kong License

Commentary

Open Access

siRNA and shRNA screens advance key understanding of host factors required for HIV-1 replication

Kin-Hang Kok, Ting Lei and Dong-Yan Jin*

Address: Department of Biochemistry, the University of Hong Kong, 21 Sassoon Road, Pokfulam, Hong Kong, PR China

Email: Kin-Hang Kok - khkok@hkucc.hku.hk; Ting Lei - leiting@hku.hk; Dong-Yan Jin* - dyjin@hkucc.hku.hk

* Corresponding author

Published: 27 August 2009

Received: 19 July 2009

Retrovirology 2009, **6**:78 doi:10.1186/1742-4690-6-78

Accepted: 27 August 2009

This article is available from: <http://www.retrovirology.com/content/6/1/78>

© 2009 Kok et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

A recent RNAi screen used a genome-wide shRNA library to search for cellular factors required for HIV-1 replication. This work complements three other siRNA-based screening studies and potentially opens the door to the discovery of factors that are important for HIV-1 replication in physiological host cells such as T lymphocytes. shRNA screens can be further improved, and they could promise to unravel new pathways and new facets of virus-cell interactions.

Commentary

The advent of RNAi-based whole-genome screens in mammalian cells provides a new impetus to the search of host cell factors needed for HIV replication [1,2]. Three screens that used siRNA pools to identify cellular proteins important in HIV-1 replication were reported in 2008, and a meta-analysis of these studies has been published recently [3-6]. One shortcoming to these reported screens is the use of HeLa or HEK293T cells that are not physiological substrates for infection by HIV-1. In addition, the use of a pseudotyped virus or a mutated strain of HIV-1 also limits the interpretability of some of the results. With this backdrop, a recently published genome-wide shRNA-screening performed in Jurkat T lymphocytes for cellular genes that contribute to HIV-1 replication (Figure 1) advances the field by extending the functional screening for cellular factors from attached epithelial/fibroblast cells to suspension T-cells [7].

In the shRNA loss-of-function screen, Jurkat cells are transduced with a lentiviral vector-based shRNA library. The lentivector is derived from feline immunodeficiency virus and is pseudotyped with the vesicular stomatitis

virus G protein. A major advantage that makes this scheme attractive is its potential application to cells that can be physiologically infected by HIV-1, including primary T cells and macrophages. In addition, it is noteworthy that the transduced Jurkat cells have been selected for shRNA-expression for extended duration before being subject to HIV-1 infection. This pre-infection selection for shRNA-expression serves to eliminate those shRNA-cell clones which are silenced for a gene whose knock down dramatically affects cell growth or survival. The pre-selection procedure thus significantly reduces the number of false positive genes identified in the screening. Unlike siRNAs, the activity of shRNAs in the cell is not transient, but is long-lasting. Because shRNAs are stably expressed, infection of cell clones with HIV-1 can be initiated at any desired time. Thus, HIV-1 infection can be performed long after the shRNA has silenced transcripts coding for long-lived proteins; these long half-life proteins generally cannot be depleted efficiently by transiently-transfected siRNAs. For the above reasons, shRNA screens have certain inherent and nuanced advantages over siRNA screens in selected settings.

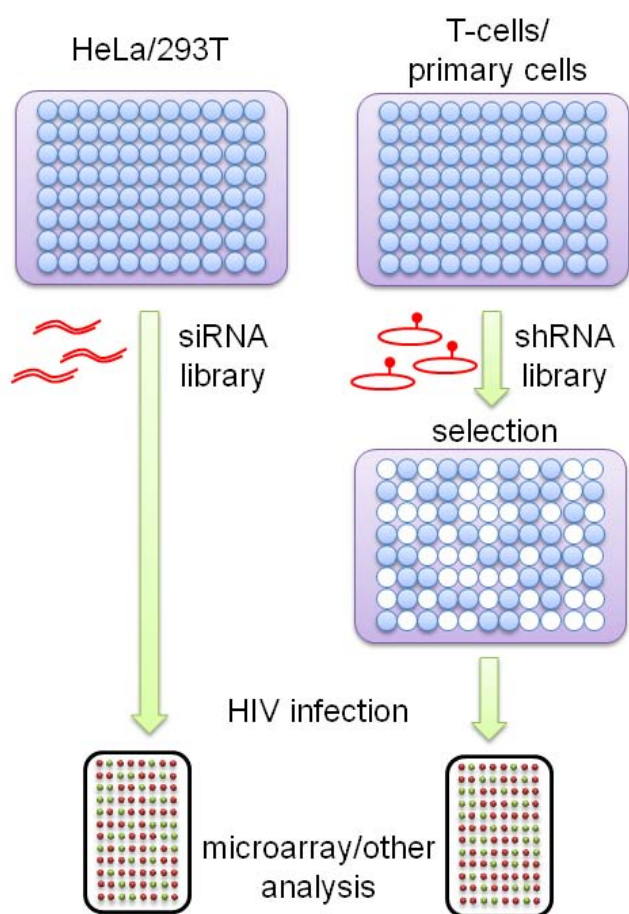


Figure 1
A comparison between siRNA- and shRNA-based screens for HIV-1 replication cofactors. See text for additional details.

Although four screens have been performed to date [3-7], the search for HIV-1 replication cofactors appears to be far from saturated. Overlaps among the genes identified in the four screens are scarce [7]; and some well known HIV-1 cofactors such as the Sp1 transcription factor, which drives LTR-dependent expression of viral genes, were not identified [6]. This lends credence to the notion that the approaches were not exhaustive. The goal of siRNA and shRNA screens is to identify all possible HIV-1 cofactors. For productive replication to occur, HIV-1 has to switch on and off many cellular pathways. Plausibly, at different stages of infection HIV-1 would hijack different cofactors to modulate the same cellular function for its own benefit. Thus, even an essential cofactor could become non-essential in a different experimental context (e. g., at an exceedingly high multiplicity of infection). That is to say, the quantitative differences in critical assay parameters such as multiplicity and duration of infection might actually affect the qualitative outcomes of a siRNA or shRNA

screen. In addition, off-target effects, differences in cell types and differences in how the primary data sets are filtered have also been suggested to account for the identification of the many different HIV-1 cofactors in the screens [1].

In the shRNA screen of Yeung et al., although the removal of cell clones that did not survive puromycin selection helped to reduce the number of false positive genes whose loss would globally inhibit cell growth or survival, rather than specifically affect HIV-1 replication, this step which eliminated more than 80% of the transduced cell clones might falsely miss those HIV-1 cofactors that are also simultaneously important for cell growth and/or survival. This issue has to be addressed in future screens. Thus, as the search continues, many more HIV-1 cofactors from different cell types are likely to emerge from further targeted si-/sh-RNA screens. Collectively, the four initial RNAi screens have already implicated new pathways that were not known previously to play critical roles in HIV-1 replication [8]. For example, HIV-1 cofactors have been identified as components of the mediator complex that regulates transcription [9], the nuclear pore complex that regulates macromolecular entry and egress, and the Golgi apparatus that specifies protein processing. While the mediator complex is thought to support Tat-dependent transcriptional activation of HIV-1 long terminal repeats (LTR), the nucleoporins and Golgi proteins could be critically involved in the intracellular transport and processing of viral nucleic acids and proteins (Table 1). Given the wide ranging functions of host cofactors identified in the four screens, the jury is still out as to which newly identified factors are of preeminence in the HIV-1 life cycle. As mentioned above, it would not be surprising that depending on the cell type and the experimental context, different cellular factors may be implicated as being rate-determining for HIV-1 replication. For example, the identification of different nucleoporins in different screens suggests that HIV-1 might employ different components of the nuclear pore complex to facilitate nuclear import/export of viral nucleic acids and proteins under different circumstances.

HIV-1 infection has been suggested to exert a suppressive effect on miRNA processing and RNA-silencing [10-16]. Indeed, the forced overexpression of shRNA and siRNA in cells might also have the potential to exhaust the cellular machinery for RNA silencing [17], complicating the interpretation of biological outcomes. Whether these influences might significantly affect the siRNA and shRNA screens for HIV-1 cofactors remains to be determined. We note, however, that the expression of a single shRNA from a lentiviral vector is unlikely to overwhelm the cell's RNAi machinery. Moreover, when shRNA-cell clones are infected by HIV-1, the shRNA-silencing of targeted mRNAs would have already been completed and would

Table 1: Selected HIV-1 cofactors identified in the siRNA and shRNA screens

Gene name	Protein function	Possible role in HIV-1 life cycle
MED4	mediator complex subunit	Tat-dependent activation of LTR
MED7	mediator complex subunit	Tat-dependent activation of LTR
MED14	mediator complex subunit	Tat-dependent activation of LTR
MED28	mediator complex subunit	Tat-dependent activation of LTR
NUP98	nucleoporin	nuclear import/export of viral nucleic acids and proteins
NUP155	nucleoporin	nuclear import/export of viral nucleic acids and proteins
NUP210	nucleoporin	nuclear import/export of viral nucleic acids and proteins
GCC1	peripheral Golgi protein	processing of viral proteins
GOLM1	Golgi transmembrane protein	processing of viral proteins
GOSR2	Golgi membrane protein	processing of viral proteins

be unlikely influenced by the effects of infection on RNAi activities.

One important direction for improving the shRNA approach is to use inducible expression systems to express the shRNAs. In this regard, a doxycycline-inducible retroviral vector has already been used successfully to construct a shRNA expression library [18]. Inducible expression of shRNAs might also help overcome potential adaptation by the cell to shRNAs, and the premature elimination of many shRNA-expressing cell clones. A further improvement to control for the possibility that some HIV-1 cofactors might also be required for the full expression of the feline immunodeficiency virus derived lentiviral vector would be to use alternate expression formats such as adenoviral vectors.

The technology of shRNA-based screening is still in its infancy [19,20]. These early reports of siRNA/shRNA screens for HIV-1 replication cofactors are therefore not the beginning of the end, but the end of the beginning of a new era in the search for host factors required for HIV-1 replication. Future screens will improve the stringency of the selection, expand upon the cell types being analyzed, and devise better strategies to address false positive/negative candidates. Furthermore, more specific questions in HIV-1 life cycle might also be addressed with siRNA and shRNA screens. For example, screens can be used to identify all the cofactors required for a specific process in HIV-1 replication such as fusion, viral entry, Tat-dependent transcription or integration. Synthetic lethal screens may also be employed to shed light on the functional interaction between different cofactors. In the years to come, the four recently reported complementary siRNA and shRNA approaches will likely be regarded as setting an important milestone for our understanding of host cell - HIV-1 interaction.

References

- Goff SP: **Knockdown screens to knockout HIV-1.** *Cell* 2008, **135**:417-420.
- Cherry S: **What have RNAi screens taught us about viral-host interactions?** *Curr Opin Microbiol* 2009, **12**:1-7.
- Brass AL, Dykxhoorn DM, Benita Y, Yan N, Engelman A, Xavier RJ, Lieberman J, Elledge SJ: **Identification of host proteins required for HIV infection through a functional genomic screen.** *Science* 2008, **319**:921-926.
- König R, Zhou Y, Elleder D, Diamond TL, Bonamy GM, Irelan JT, Chiang CY, Tu BP, De Jesus PD, Lilley CE, Seidel S, Opaluch AM, Caldwell JS, Weitzman MD, Kuhen KL, Bandyopadhyay S, Ideker T, Orth AP, Miraglia LJ, Bushman FD, Young JA, Chanda SK: **Global analysis of host-pathogen interactions that regulate early-stage HIV-1 replication.** *Cell* 2008, **135**:49-60.
- Zhou H, Xu M, Huang Q, Gates AT, Zhang XD, Castle JC, Stec E, Ferrer M, Strulovici B, Hazuda DJ, Espeseth AS: **Genome-scale RNAi screen for host factors required for HIV replication.** *Cell Host Microbe* 2008, **4**:495-504.
- Bushman FD, Malani N, Fernandes J, D'Orso I, Cagney G, Diamond TL, Zhou H, Hazuda DJ, Espeseth AS, König R, Bandyopadhyay S, Ideker T, Goff SP, Krogan NJ, Frankel AD, Young JA, Chanda SK: **Host cell factors in HIV replication: meta-analysis of genome-wide studies.** *PLoS Pathog* 2009, **5**:e1000437.
- Yeung ML, Houzet L, Yedavalli VS, Jeang KT: **A genome-wide short hairpin RNA screening of Jurkat T-cells for human proteins contributing to productive HIV-1 replication.** *J Biol Chem* 2009, **284**:19463-19473.
- Lama J, Planelles V: **Host factors influencing susceptibility to HIV infection and AIDS progression.** *Retrovirology* 2007, **4**:52.
- Conaway RC, Sato S, Tomomori-Sato C, Yao T, Conaway JW: **The mammalian Mediator complex and its role in transcriptional regulation.** *Trends Biochem Sci* 2005, **30**:250-255.
- Bennasser Y, Le SY, Benkirane M, Jeang KT: **Evidence that HIV-1 encodes an siRNA and a suppressor of RNA silencing.** *Immunity* 2005, **22**:607-619.
- Bennasser Y, Yeung ML, Jeang KT: **HIV-1 TAR RNA subverts RNA interference in transfected cells through sequestration of TAR RNA-binding protein, TRBP.** *J Biol Chem* 2006, **281**:27674-27678.
- Triboulet R, Mari B, Lin YL, Chable-Bessia C, Bennasser Y, Lebrigand K, Cardinaud B, Maurin T, Barbry P, Baillat V, Reynes J, Corbeau P, Jeang KT, Benkirane M: **Suppression of microRNA-silencing pathway by HIV-1 during virus replication.** *Science* 2007, **315**:1579-1582.
- Schnettler E, de Vries W, Hemmes H, Haasnoot J, Kormelink R, Goldbach R, Berkhout B: **The NS3 protein of rice hoja blanca virus complements the RNAi suppressor function of HIV-1 Tat.** *EMBO Rep* 2009, **10**:258-263.
- Qian S, Zhong X, Yu L, Ding B, de Haan P, Boris-Lawrie K: **HIV-1 Tat RNA silencing suppressor activity is conserved across kingdoms and counteracts translational repression of HIV-1.** *Proc Natl Acad Sci USA* 2009, **106**:605-610.
- Houzet L, Yeung ML, de sLame V, Desai D, Smith SM, Jeang KT: **MicroRNA profile changes in human immunodeficiency virus type 1 (HIV-1) seropositive individuals.** *Retrovirology* 2008, **5**:118.
- Yeung ML, Bennasser Y, Myers TG, Jiang G, Benkirane M, Jeang KT: **Changes in microRNA expression profiles in HIV-1-transfected human cells.** *Retrovirology* 2005, **2**:81.

17. Grimm D, Streetz KL, Jopling CL, Storm TA, Pandey K, Davis CR, Marion P, Salazar F, Kay MA: **Fatality in mice due to oversaturation of cellular microRNA/short hairpin RNA pathways.** *Nature* 2006, **441**:537-541.
18. Ngo VN, Davis RE, Lamy L, Yu X, Zhao H, Lenz G, Lam LT, Dave S, Yang L, Powell J, Staudt LM: **A loss-of-function RNA interference screen for molecular targets in cancer.** *Nature* 2006, **441**:106-110.
19. Bernards R, Brummelkamp TR, Beijersbergen RL: **shRNA libraries and their use in cancer genetics.** *Nat Methods* 2006, **3**:701-706.
20. Sharma S, Rao A: **RNAi screening: tips and techniques.** *Nat Immunol* 2009, **10**:799-804.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

