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Three new species of *Pyricularia* are isolated as zingiberaceous endophytes from Thailand

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Abstract: Pyricularia costina and three undescribed Pyricularia species were found as endophytes on wild ginger Amomum siamense and Alpinia malaccensis in Doi Suthep-Pui National Park, Chiang Mai, Thailand. Three new species, Pyricularia kookicola, P. longispora, and P. variabilis are described, illustrated and compared to similar Pyricularia species.

Key words: Alpinia malaccensis, Amomum siamense, mitosporic fungi, taxonomy

INTRODUCTION

Genus Pyricularia Sacc. was established by Saccardo (1880) with the single species, P. grisea Sacc., which commonly is found on tropical grasses. The name Pyricularia referred to the pyriform shape of the conidia. Pyricularia oryzae Cav. subsequently was described for a very similar fungus on rice (Cavara 1891), and currently there are 67 names for Pyricularia species in Funindex (http://194.131.255.3/cabipages/Names/NAMES.APS). Molecular studies have confirmed that P. grisea and P. oryzae are identical (Lebrun et al 1991), and the teleomorph, Magnaporthe grisea also has been well studied (Hebert 1971, Kato et al 1976, Yaegashi and Hebert 1976, Barr 1977). Genus Pyricularia, however, needs a monographic study.

Pyricularia was circumscribed by Ellis (1971), although the distinction between it and some Dactylaria spp. is not always clear (Goh and Hyde 1997). The presence of a separating cell was a feature used by Ellis (1976) to distinguish Pyricularia from the similar genus, Dactylaria. The conidiogenous cells of Pyricularia are polyblastic, integrated on the conidiophores and are sympodial, cylindrical, geniculate and denticulate. Each denticle is cylindrical, thin-walled and cut off by a septum to form a separating cell. The conidia secede in a rhexolytic manner. They are solitary, dry, acropleurogenous, simple, obpyriform, obturbinate or obclavate, hyaline to pale olivaceous brown, and often have a protuberant hilum (Ellis 1971). In contrast, each denticle of Dactylaria is cylindrical, flat-topped and continuous with the conidiogenous cells. The conidia secede in a schizolytic manner and are of various shapes, usually fusiform, naviculate or cylindrical. However, in some species they may be obpyriform, obclavate, fabiform, botuliform or other shapes (Ellis 1976, Goh and Hyde 1997).

Most species of Pyricularia are recorded from monocotyledonous plants (Nisikado 1927, Roldan 1938, Viégas 1946, Luttrell 1954, Thirumalachar et al 1956, Rao and Reddy 1958, Veeraraghavan and Padmanabhan 1965, Prasada and Goyal 1970, 1974, Hashioka 1971, 1973, Albuquerque and Duarte 1971, Siwasin and Giatgong 1971, Rathaiah 1980, Zucconi et al 1984, Gaikwad and D'Souza 1987, Ondřej 1988, Sarbajna 1990), especially Commelinaceae, Cyperaceae, Poaceae, and Zingiberaceae. However, Pyricularia caffera Matsush., P. lourinae F.C. Albuq. & L.R. Duarte, P. peruamazonica Matsush., P. subsigmoidea R.F. Castañeda & W.B. Kendr., and P. vandalurensis Subram. & Vittal were described from dicotyledonous plants (Albuquerque and Duarte 1971, Subramanian and Vittal 1974, Castañeda and Kendrick 1991, Matsushima 1993, 1996). Some species originally described in *Pyricularia* subsequently have been synonymized or transferred to other genera. For example, P. aquatica Ingold and P. juncicola MacGaevie were moved to *Tumularia aquatica* (Ingold) Descals & Marvanová and Dactylaria junci M.B. Ellis, respectively (Ingold 1943, Ellis 1971, 1976, Marvanová and Descals 1987). In addition, Pyricularia musae Hughes (1958) was placed in Pyriculariopsis because its broad

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denticles are not cut off by a septum to form a separating cell, as they are in *Pyricularia* (Ellis 1971). The conidia of *Pyriculariopsis parasitica* are versicolored, with hyaline end cells and darker intermediate cells.

During our investigation of endophytic fungi on wild ginger, Amomum siamense Craib. and Alpinia malaccensis (Burm.) Rosc., at Doi Suthep-Pui National Park, Chiang Mai, Thailand, we found 41 fungal taxa, including four Pyricularia species, P. costina Sarbajna and three undescribed species. Pyricularia species were isolated from 8.38% of 800 healthy leaves of Amomum siamense sampled, and 2.12% of 800 healthy leaves of Alpinia malaccensis sampled (Bussaban et al 2001a, b). In this paper, the new species, P. kookicola Bussaban sp. nov., P. longispora Bussaban sp. nov. and P. variabilis Bussaban sp. nov. are described and compared to similar species.

MATERIALS AND METHODS

Endophytic fungi were isolated from Amonum siamense (1222 isolates) and Alpinia malaccensis (1110 isolates), using a triple sterilization method, and cultured on cornmeal agar (Bussaban et al 2001a, b). Pyricularia species were isolated from leaves treated by surface sterilization in 95% alcohol for 15 s, followed by 5 min in 1% sodium hypochlorite and finally, 15 s in 95% alcohol. Slides of fungi were mounted in distilled water for observation, photomicrography and measurement. Living cultures have been deposited in the National Center of Genetic Engineering and Biotechnology Culture Collection, Thailand (BCC).

TAXONOMY

Pyricularia costina Sarbajna, Journal of Mycopathological Research 28: 162. 1990. FIGS. 1, 2, 14A *Colonies* on potato-dextrose agar reaching 9 cm in 10 d, effuse, grayish-brown. *Conidiophores* up to 500 μ m long, 3.7–5 μ m thick, usually branched, flexuous, geniculate, septate, pale brown to brown, smooth. *Conidiogenous cells* sympodial, cylindrical, geniculate, denticulate; each denticle cylindrical, thin-walled, mostly cut off by septum to form a separating cell. *Conidia* 22.5–37.5 × 7.5–10 μ m, solitary, dry, obpyriform or obturbinate, hyaline, smooth, 2-septate, hilum often protuberant.

Speciments examined: THAILAND. CHIANG MAI: Doi Suthep-Pui National Park, isolated as an endophyte from leaves of Amomum siamense, February 2000, B. Bussaban CMUZE0003 (BCC8219); August 1999, B. Bussaban CMU-ZE0006; isolated as an endophyte from leaves of Alpinia malaccensis, August 1999, B. Bussaban CMUZE0141; February 2000, B. Bussaban CMUZE0504; CMUZE0505.

Commentary: This species has been described as a pathogen (Sarbajna 1990), causing leaf spots on Cos-

tus speciosus Smith (Zingiberaceae), but no spots were seen on either Amomum siamense or Alpinia malaccensis, the fungus being isolated from healthy green leaves.

Pyricularia kookicola Bussaban, sp. nov.

FIGS. 3–5, 14B

Coloniae effusae, viridi-griseus. Conidiophora macronemata, usque ad 250 μ m longa, 2.5–4 μ m crassa, raro ramosa, flexuosa, septata, pallide brunnea vel brunnea, laevia. Cellulae conidiogenae sympodiales, cylindricis, geniculatae, denticulatae, denticulis quibusque cylindricis, tennuitunicatis cellulam intercalarem producentibus. Conidia 27–33.5 \times 7.9–10 μ m, solitaria, sicca, obpyriformia, hyalina, laevia, 2-septata, hilo distincte.

Colonies on potato-dextrose agar reaching 9 cm in 12 d, effuse, greenish-gray. Conidiophores up to 250 μ m long, 2.5–4 μ m thick, macronematous, rarely branched, flexuous, septate, pale brown to brown, smooth. Conidiogenous cells sympodial, cylindrical, geniculate, denticulate; each denticle cylindrical, thinwalled, mostly cut off by septum to form a separating cell. Conidia 27–33.5 \times 7.9–10 μ m, solitary, dry, obpyriform, hyaline, smooth, 2-septate, hilum protuberant.

Etymology: Kook, the local northern Thai name for *Amomum siamense*, and *icola* meaning "loving".

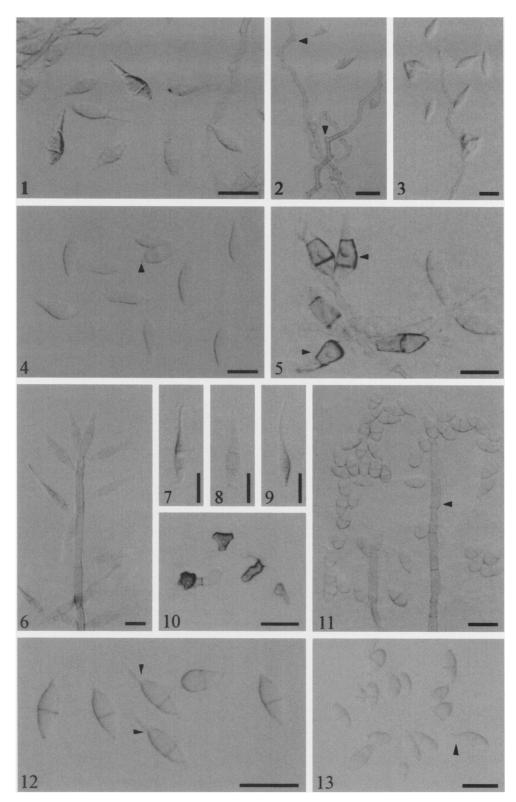
Speciments examined: THAILAND. CHIANG MAI: Doi Suthep-Pui National Park, isolated as an endophyte from leaves of Amomum siamense, February 2000, B. Bussaban CMUZE0501 (HOLOTYPE).

Commentary: This species is similar to Pyricularia angulata, P. cannaecola, P. distorta and P. zingiberi in having 2-septate conidia. However, the conidia of P. kookicola are larger. Pyricularia kookicola also is similar to P. guarmaicola in conidial shape and septation, but its conidia are narrower (27–33.5 \times 7.9–10 μ m vs 20– 28×10 –12 µm). Pyricularia kookicola has similar-size 2-septate conidia as P. caricis, P. costina, P. leersiae, P. penniseti and P. peruamazonica. Pyricularia kookicola has obpyriform conidia, whereas P. caricis and P. peruamazonica have obclavate and fusiform conidia, respectively. Moreover, conidiophores of P. caricis mostly are nonseptate. Pyricularia costina are bluntly rounded at the apex, and its apical cells are more slender than those of P. kookicola. Pyricularia leersiae and P. penniseti differ from P. kookicola in that they produce subglobose to ovate appressoria. Moreover, conidia of P. penniseti have slender apical cells and the percentage of the length of the apical cell to the length of a conidium reaches 42-44%, in comparison to P. kookicola, which occupies 30-35%.

Pyricularia longispora Bussaban, sp. nov.

FIGS. 6–10, 14C

Coloniae effusae, griseo-brunnae. Appressoria 12–15 μm per medius, angularis et irregularibus, stellatae, atro-brun-



FIGS. 1–13. Pyricularia costina, P. kookicola, P. longispora and P. variabilis. 1, 2. P. costina. 1. Conidia. 2. Conidiophores with denticles (arrowed) and conidia. 3–5. P. kookicola (from holotype). 3. Conidiophore and conidia. 4. Conidia with protuberant hilum (arrowed). 5. Matured conidia (arrowed). 6–10. P. longispora (from holotype). 6. Conidiophore and conidia. 7–9. Conidia. 10. Irregular shaped hyphopodia. 11–13. P. variabilis (from holotype). 11. Conidiophores with swollen intercalary nodes (arrowed) and conidia. 12, 13. One and two (arrowed) septate conidia. Scale bars = $20 \mu m$.

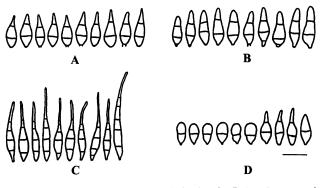


FIG. 14. Pyricularia costina, P. kookicola, P. longispora and P. variabilis. Diagrammatic representation of conidia. A. P. costina. B. P. kookicola. C. P. longispora. D. P. variabilis. Scale bars = $20 \mu m$.

nae. Conidiophora macronemata, usque ad 400 μ m longa, 3– 4.6 μ m crassa, raro ramosa, recta, septata, ad basem pallide brunnea, ad apicem plerumque subhyalina. Cellulae conidiogenae cylindricis, denticulis quibusque cylindricis, tennuitunicatis cellulam intercalarem producentibus. Conidia 47–72 × 5.6–7.6 μ m, solitaria, sicca, obclavata, hyalina vel pallide brunnea, laevia, 4(–5)-septata.

Colonies on potato-dextrose agar reaching 9 cm in 12 d, effuse, grayish-brown. Appressoria 12–15 μ m diam, angular to irregular, stellate, dark brown. Conidiophores up to 400 μ m long, 3–4.6 μ m thick, macronematous, rarely branched, straight, septate, pale brown near the base, often subhyaline at the apex. Conidiogenous cells cylindrical, denticulate; each denticle cylindrical, thin-walled, mostly cut off by septum to form a separating cell. Conidia 47–72 × 5.6–7.6 μ m, solitary, dry, obclavate, hyaline to pale brown, smooth, 4(–5)-septate.

Etymology: Referring to the length of conidia.

Speciments examined: THAILAND. CHIANG MAI: Doi Suthep-Pui National Park, isolated as an endophyte from leaves of Amomum siamense, February 2000, B. Bussaban CMUZE0005 (HOLOTYPE: BCC11377); August 1999, B. Bussaban CMUZE0502; isolated as an endophyte from leaves of Alpinia malaccensis, August 1999, B. Bussaban CMUZE0503.

Commentary: This species differs from other Pyricularia spp. in its long, 4-septate conidia. Pyricularia angulata (ovate-pyriform, $18.2-28 \times 4.9-9.1 \mu m$, 2septate), P. leersiae (obpyriform, $27-37.5 \times 6-9.7 \mu m$, 2-septate) and P. penniseti (obclavate, $18.4-36.7 \times 7.4-11 \mu m$, 2-septate) also form appressoria in culture. However, conidia are very different in shape, size and septation, when compared to P. longispora (obclavate, $47-72 \times 5.6-7.6 \mu m$, 4(-5)-septate).

Pyricularia variabilis Bussaban, sp. nov.

FIGS. 11–13, 14D

Coloniae effusae, griseo-brunnae. Conidiophora macronemata, usque ad 700 μ m longa, 3.8–5 μ m crassa, septata, recta vel flexuosa, atro-brunnea, ad apicem subhyalina, laevia, ad apicem et intercalarem 5–7 μ m inflata. Cellulae conidiogenae sympodiales, geniculatae, denticulatae, denticulis quibusque cylindricis, tennuitunicatis cellulam intercalarem producentibus. Conidia 15.7–28.6 × 6.4–9.3 μ m, solitaria, obovatae vel obpyriformia, hyalina vel subhyalina, laevia, 1– 2-septata, hilo distincte.

Colonies on potato-dextrose agar reaching 9 cm in 7 d, effuse, grayish-brown, hairy. Conidiophores up to 700 μ m long, 3.8–5 μ m thick, macronematous, septate, straight or flexuous, dark brown, subhyaline at the apex, smooth, terminal and intercalary nodes swelling 5–7 μ m diam. Conidiogenous cells sympodial, cylindrical, geniculate, denticulate; each denticle cylindrical, thin-walled, mostly cut off by septum to form a separating cell. Conidia 15.7–28.6 × 6.4–9.3 μ m, solitary on denticles arising from terminal and intercalary swellings, obovoid or obpyriform, hyaline to subhyaline, smooth, 1–2-septate, hilum protuberant.

Etymology: Referring to variation of shape of conidia.

Speciments examined: THAILAND. CHIANG MAI: Doi Suthep-Pui National Park, isolated as an endophyte from leaves of Amomum siamense, February 2000, B. Bussaban CMUZE0229 (HOLOTYPE: BCC8210); August 1999, B. Bussaban CMUZE0506.

Commentary: Pyricularia variabilis differs from other species in the genus in its variable conidial shape. It also has swollen, terminal and intercalary nodes on the conidiophores, a feature not found in other species of *Pyricularia*.

Five species of Pyricularia have been found to be parasitic on Zingiberaceae. Nisikado (1927) isolated P. zingiberi Nisik. from common ginger (Zingiber officinale Rosc.) and Japanese wild ginger (Z. mioga Rosc.). Hashioka (1971) described a new species, P. distorta Hashioka, as a blast fungus on Catymbium sp. and Alpinia sp. in Thailand, while Siwasin and Gaitgong (1971) reported P. globbae Siwasin & Giatgong, infecting Globba sp. in Thailand. In India, P. costina Sarbajna and P. curcumae Rathaiah were reported as pathogens, causing leaf spots of Costus speciosus Smith and leaf blast of turmeric, Curcuma longa L., respectively (Rathaiah 1980, Sabajna 1990). This study is the first report of Pyricularia costina and three new Pyricularia spp. as endophytes of wild ginger.

Some endophytes are thought to benefit host plants by protecting them against insect pests and plant pathogens, conferring drought tolerance, or enhancing absorption of soil nutrients (Webber 1981, Funk et al 1983, Carroll 1986, 1988, Thomson et al 1986, Clay 1989, Breen 1993, 1994, Stone et al 2000). Endophytic fungi may develop as saprobes, once a plant senesces or a leaf dies. However, they also might be latent pathogens and develop to cause plant diseases under some conditions (Latch 1993, Brown et al 1998). Photita et al (2001) found *Pyriculariopsis parasitica* to be a common endophyte of *Musa acuminata* Colla, but it also is pathogenic on bananas (Meredith 1962, Stover 1972). In *Amomum siamense* and *Alpinia malaccensis*, *Pyricularia* species were isolated from leaves, not from pseudostems or rhizomes (Bussaban et al 2001a, b). They might be pathogenic or saprobic, and it is unknown what role endophytic species isolated from ginger in Thailand might play.

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LITERATURE CITED

- Albuquerque FC, de Duarte LR. 1971. Duas novas espécies de *Piricularia* coletadas na Amazônia. Pesq Agropecu Brasil 6:177–181.
- Barr ME. 1977. Magnapothe, Telimenella and Hyponectria (Physosporellceae). Mycologia 69:952–956.
- Brown KB, Hyde KD, Guest DI. 1998. Preliminary studies on endophytic fungal communities of *Musa acuminata* species complex in Hong Kong and Australia. Fungal Diversity 1:26–50.
- Breen JP. 1993. Enhanced resistance to fall armyworm (Lepidoptera: Noctuidae) in *Acremonium* endophyte-infected turfgrasses. J Eco Entomol 86:621–629.
 - ——. 1994. Acremonium-endophyte interactions with enhanced plant resistance to insects. Ann Rev Entomol 39:401–423.
- Bussaban B, Lumyong S, Lumyong P, McKenzie EHC, Hyde KD. 2001a. Endophytic fungi from *Amomum siamense*. Canad J Microbiol 47:943–948.
 - ——. 2001b. Endophytic fungi from Zingiberaceae: Alpinia malaccensis. Phytopathology 91:S104. Publication no. P-2001-0023-MSA.
- Carroll G. 1986. The biology of endophytism in plants with particular reference to woody perennials. In: Fokkema NJ, Van der Heavel J. eds. Microbiology of the Phylloplane. London: Cambridge University Press. p 205– 222.
 - ——. 1988. Fungal endophytes in stems and leaves: from latent pathogens to mutualistic symbionts. Ecology 69: 2–9.
- Castañeda RF, Kendrick WB. 1991. Ninety-nine conidial fungi from Cuba and three from Canada. Univ. Waterloo Biol. Ser. 35:1–132.

- Cavara F. 1891. Fungi Longobardiae exsiccati sive mycetum specimina in *Longobardia collecta*, exsiccata et speciebus novis vel criticis, iconibus illustrata. Pugillus I no. 49 (Cited in Padwick GW. 1950 p. 18).
- Clay K. 1989. Clavicipitaceous endophytes of grasses: their potential as biocontrol agents. Mycol Res 92:1–12.
- Ellis MB. 1971. Dematiaceous Hyphomycetes. England: Commonwealth Mycological Institute. 680 p.
- ——. 1976. More Dematiaceous Hyphomycetes. England: Commonwealth Mycological Institute. 507 p.
- Funk CR, Halisky PM, Johnson MC, Siegel MR, Stewart AV, Ahmad S, Hurley RH, Harvey IC. 1983. An endophytic fungus and resistance to sod webworms: association in *Lolium perenne*. Bio/Technology 1:189–191.
- Gaikwad AP, D'Souza TF. 1987. A comparative study on *Pyr-icularia* spp. J. Maharashtra Agric. Univ. 12:134–135.
- Goh TK, Hyde KD. 1997. A revision of *Dactylaria*, with description of *D. tunicata* sp. nov. from submerged wood in Australia. Mycol Res 101:1265–1272.
- Hashioka Y. 1971. Notes on *Pyricularia* I. Three species parasitic to Musaceae, Cannaceae and Zingiberaceae. Trans Mycol Soc Japan 12:126–135.
- ———. 1973. Notes on *Pyricularia* II. Four species and one variety parasitic to Cyperaceae, Gramineae and Commelinaceae. Trans Mycol Soc Japan 14:256–265.
- Hebert TT. 1971. The perfect stage of *Pyricularia grisea*. Phytopathology 61:83-87.
- Hughes SJ. 1958. Revisiones Hyphomycetum aliquot cum appendice de nominibus rejiciendis. Can J Bot 36:727-836.
- Ingold CT. 1943. Further observations on aquatic hyphomycetes of decaying leaves. Trans Brit Mycol Soc 26: 105–115.
- Kato H, Yamaguchi T, Nishihara N. 1976. The perfect stage of *Pyricularia oryzae* Cav. in culture. Ann Phytopathol Soc Japan 42:507–510.
- Latch GCM. 1993. Physiological interactions of endophytic fungi and their hosts; biotic stress tolerance imparted to grasses by endophytes. Agric Ecosyst Environ 44:143–156.
- Lebrun MH, Capy MP, Garcia N, Brygoo Y, Boccara M, Notteghem JL, Vales M. 1991. Biology and genetics of *Pyricularia oryzae* and *P. grisea* populations: current situation and development of RFLP markers. In: Rice Genetics 2. Proceedings of the Second International Rice Genetics Symposium. IRRI. Los Banos, Laguna, The Philippines. p 487–497.
- Luttrell ES. 1954. An undescribed species of *Pyricularia* on sedges. Mycologia 46:810–814.
- Marvanová L, Descals E. 1987. New taxa and new combinations of 'aquatic hyphomycetes'. Trans Brit Mycol Soc 89:499–507.
- Matsushima T. 1993. Matsushima Mycol. Mem. 7. Japan: Matsushima, Kobe. 75 p.
- ——. 1996. Matsushima Mycol. Mem. 9. Japan: Matsushima, Kobe. 40 p.
- Meredith DS. 1962. *Pyricularia musae* Hughes in Jamaica. Trans Brit Mycol Soc 45:137–142.
- Nisikado Y. 1927. Studies on the rice blast disease. Jap J Bot 3:239–244.

- Ondřej M. 1988. *Pyricularia luzulae* Ondřej sp. n. Česká Mykol 42:81–83.
- Photita W, Lumyong S, Lumyong P, Hyde KD. 2001. Endophytic fungi of wild banana (*Musa acuminata*) at Doi Suthep Pui National Park, Thailand. Mycol Res 105:1508–1513.
- Prasada R, Goyal JP. 1970. A new species of *Pyricularia* on Bajra. Curr Sci 39:287–288.
- Prasada R. 1974. A new species of *Pyricuaria* inciting leaf spot disease of Bajra (*Pennisetum typhoides* Stapf. and Hubbard). Beih Nova Hedwigia 47:621-623.
- Rao PG, Reddy TCV. 1958. A new *Pyricularia* from India. Sci & Cult 24:133–135.
- Rathaiah Y. 1980. Leaf blast of turmeric. Pl Dis 64:104-105.
- Roldan EF. 1938. New or noteworthy lower fungi of the Philippine Islands II. Philipp J Sci 66:7–13.
- Saccardo PA. 1880. Fungorum extra-europaeorum Pugillus. Michelia 2:136–149.
- Sarbajna KK. 1990. New species of *Mycovellosiella* and *Pyricularia* from West Bengal. J Mycopathol Res 28:159– 164.
- Siwasin C, Giatgong P. 1971. Cytological study and cross inoculation of *Pyricularia* spp. Newslett Int Rice Commiss 20:13–19.
- Stone JK, Bacon CW, White JF. 2000. An overview of endophytic microbes: endophytism defined. In: Bacon

CW, White JF, eds. Microbial endophytes. New York: Jr. Marcel Dekker. p 1–29.

- Stover RH. 1972. Banana, Plantain and ABACA Diseases. England: Commonwealth Mycological Institute. 316 p.
- Subramanian CV, Vittal BPR. 1974. Hyphomycetes on litter from India I. Proc Indian Acad Sci B 80:216-221.
- Thirumalachar MJ, Kulkarni NB, Patel MK. 1956. Two new records of *Pyricularia* species from India. Indian Phytopathol 9:48–51.
- Thomson BD, Robson AD, Abbott LK. 1986. Effects of phosphorus and the formation of mycorrhyzas by *Gigaspora calospora* and *Glomus fasiculatum* in relation to root carbohydrates. New Phytol 103:751–765.
- Veeraraghavan J, Padmanabhan SY. 1965. Studies on the host range of *Pyricularia oryzae* Cav. causing blast disease of rice. Proc Indian Acad Sci B 61:109–120.
- Viégas AP. 1946. Alguns fungos do Brasil XIII. Hifomicitos. Bragantia 6:353–442.
- Webber J. 1981. A natural control of Dutch elm disease. Nature, London 292:449–451.
- Yaegashi H, Hebert TT. 1976. Perithecial development and nuclear behavior in *Pyricularia*. Phytopathology 66: 122–126.
- Zucconi L, Onofri S, Persiani AM. 1984. Hyphomycetes rari o interessanti della foresta tropicale II. Pyricularia fusispora comb. nov., nuova combinazione per la specie Nakataea fusispora. Micol Ital 2:7–10.