

Title	The aetiology of idiopathic Parkinson's disease
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Abstract

Agents potentially involved in the aetiology of idiopathic Parkinson's disease are discussed. These include factors regulating dopaminergic neurogenesis (Nurr 1, Ptx-3, and Lmx1b) and related proteins, together with genes involved in familial Parkinson's disease (α synuclein, parkin, and ubiquitin carboxy terminal hydroxylase L1), and endogenous and environmental agents.

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Keywords: Parkinson's disease; environmental agents; transcription factors; α synuclein; parkin; ubiquitin carboxy terminal hydroxylase L1

Idiopathic Parkinson's disease appears not to be a single entity but rather a spectrum of conditions resulting from the death of the pigmented dopaminergic neurones of the substantia nigra, pars compacta, which ultimately leads to the one single, fatal endpoint. As such, this spectrum is unlikely to have a single cause. Despite an intense research effort over many years, these causes still await elucidation. Various factors contribute to the difficulties in the search. Some obvious ones are the long period between the initiation of the disease process and the manifestation of clinical symptoms, the lack of any distinctive blood biochemistry with which to trace the disease process, and the inadequacy of current animal models. Quite when idiopathic Parkinson's disease begins is uncertain. Based on the premise that clinical symptoms appear when approximately 50% of pigmented dopaminergic neurones are dead and the surviving ones can supply the striatum with only about 20-30% of its dopamine demand, mathematical models of neuronal death rates suggest that there may be as small a gap as three years or as large a one as 20. However, beyond the strictly mathematical, clinical case histories of twins suggest the presence of a "parkinsonian" personality that, in hindsight, was present in very early life. Whether such a personality exists and, if so, whether it implies the presence of the disease process so early in life remain obscure. Nevertheless, the concept that early life events might be of crucial importance has been moved from the realms of fancy with the recognition of the role of specific transcription factors in brain development, particularly the proteins Nurr 1 (and its related subfamily members), Lmx1b, and Ptx-3. These are transcription factors that are involved in determining neurogenesis in the basal ganglia. It has been hypothesised that immature neurones die by apoptosis because of neurotrophic factor deprivation, as a result of failing to make adequate contacts in their target sites, whereas mature neurones die because of toxic insult. However, the quality of contact and/or degree of neurotrophic support in early life may be of importance in determining the length of survival. In addition, these proteins are expressed throughout life in the basal ganglia, which suggests that they have roles in maintaining the continuing health of specific neurones and makes them worthy of consideration in terms of the aetiology of idiopathic Parkinson's disease. These factors often have multiple names, which sometimes relate to the different species that research groups were investigating, or simply to different phenomena that were being considered at the time. For the sake of simplicity only one name will be used here in the main body of the text, but alternative names are listed.

Factors involved in dopaminergic neurogenesis

NUR RELATED FACTOR 1 (NURR 1) (ALSO KNOWN AS NOT, RNR-1, AND HZF-3)

Nurr 1 is a member of the superfamily of "zinc finger" transcription factors.1 Over 90 members of the superfamily have been described. Included among these are the hormone receptors for triiodothyronine, oestradiol, cortisol, aldosterone, vitamin D, all trans retinoic acid, and 9-cis retinoic acid. Increasingly, molecules that at one time were thought of as unlikely candidate ligands for these transcription factors are now being recognised as such. In the past few years, it has been established that bile acids modulate gene transcription via farnesyl receptors.² Nevertheless, many members of the superfamily have no known ligands-so called "orphan" receptors (reviewed by Enmark and Gustafsson³)—but despite this, the functions of these receptors are becoming recognised. Most recently, cholesterol entry and egress from cells has been shown to be regulated via such orphans.4 5 Also among this class of receptors is Nurr 1.

Nurr 1 was first recognised as a transcription factor that was primarily expressed in the brain⁶ and the regenerating liver⁷ (some details of the gene, cDNA, and protein are given in table 1, together with those for Ptx-3 and Lmx1b). Its close structural relation to Nur 77 led to its identification in stimulated T cells, where it functions as an intermediate early response gene, with stimulation causing a rapid increase in mRNA without the need for de novo protein synthesis.¹ Inhibition of Nurr 1 binding to its response element led to inefficient clonal deletion of self reactive T cells.34 It also functions as an intermediate early gene in the central nervous system (CNS), being rapidly induced in response to focal injury,³ but an appreciation of its crucial importance in brain development came later, after it was

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Nurr 1

- Member of "zinc finger" superfamily of receptors; its subfamily includes nerve growth factor inducible B (NGFI-B) and neurone derived orphan receptor 1 (NOR-1)⁸
- Human gene on 2q22–23; eight exons, spanning 8 kb. Murine, rat, and human genes have similar structures⁹⁻¹²
- Full length mRNA has 3427 bases, 1794 are translated; the protein has 598 amino acids (66 kDa)¹, splice variants exist—for example, Nurr 2—a novel cryptic exon located upstream in the Nurr 1 promoter region—and alternative splicing at exons 1, 2, and 6¹³
- Potential regulatory region contains consensus binding sites for nuclear factor κB (NF-κB), cAMP response element binding protein (CREB), and Sp1^{9 11 14}
 In the central nervous system (CNS) Nurr 1 expression occurs in the postmitotic late
- In the central nervous system (CNS) Nurr 1 expression occurs in the postmitotic late differential phase of dopaminergic precursor neuronal development,¹⁵¹⁶ detectable from rat embryonic day 10.5¹⁷
- Nurr 1 response elements: a single half site (AAAGGTCA) binds Nurr 1 as monomer; this is the same sequence as the NGFI-B response element^{7 8}; a direct repeat (DR5) (AGGTCANNNAAAGGTCA) binds Nurr 1/RXR as a heterodimer, Nurr 1 homodimers, and the Nurr 1/NGFI-B heterodimer¹⁸⁻²⁰
- Nurr 1 response elements are found in tyrosine hydroxylase (AAAGGTCA),²¹ the dopamine transporter,²² and other molecules such as proopiomelanocortin²³

Ptx-3

- Structurally related to pentaxins—for example, C reactive protein and serum amyloid P; subfamily includes Ptx-1 and Ptx-2, and homeobox proteins such as Otx-1 and Otx-2^{24 25}
- Human gene on chromosome 10.q25. has three exons²⁶
 cDNA has 1861 bp; protein has 381 amino acids²⁵; Ptx-3 is also a secreted, acute phase protein
- mRNA induced in endothelial, hepatic, and fibroblastic cells by interleukin 1 β (IL-1 β) and tumour necrosis factor *a*, but not by IL-6 and interferon γ ; raised serum concentrations after bacterial lipopolysaccharide injection^{25 27 28}
- Expression is restricted in the CNS²⁹
- The Ptx-3 response element is GGCTTT
- There is a Ptx-3 response element in the tyrosine hydroxylase gene³⁰
- Mutations are associated with autosomal dominant cataracts²

Lmx1b

- This transcription factor is related to members of the LIM family of homeobox proteins³¹
- The human gene is on 9q34 and has eight exons³²
 Genetic mutations are associated with nail-patella syndrome³³
 - shown to be expressed strongly in the midbrain region.^{16 36} In mice in which the expression of Nurr 1 is prevented (-/- knockout), there is a failure of development of midbrain dopaminergic neurones, with a 98% reduction in striatal dopamine and a 30% reduction in noradrenaline.^{37 38} In the olfactory bulb, another area that is important in idiopathic Parkinson's disease (70% of patients with idiopathic Parkinson's disease have olfactory bulb dysfunction, and anosia may precede other signs of the disease), Nurr 1 (-/-) mice have a 60% reduction in dopamine,³⁷ although claims concerning the details of Nurr 1 expression in the olfactory bulb are conflicting.^{39 40} The importance of Nurr 1 for dopaminergic neurones can be seen in relation to tyrosine hydroxylase expression, as shown by the degree of coexpression of the two proteins in the adult mouse brain, namely: substantia nigra (96%), ventral tegmental area (95%), retrorubral field (91%), olfactory bulb (85%), linear nucleus raphe (91%), central grey (61%), paraventricular and periventricular hypothalamic nucleus (few), and arcuate nucleus and zona incerta (0%).⁴¹ In the absence of the growth of the nigral dopaminergic neurones, the animals fail to thrive and die shortly after birth. In contrast, heterozygous animals (+/- Nurr 1 mice) are apparently healthy but have reduced midbrain dopamine values.³⁷ In humans, a similar role for Nurr 1 in dopaminergic neurogenesis is assumed.

Nurr 1 is not only expressed throughout life in midbrain neurones, but also in other regions unconnected with idiopathic Parkinson's disease,⁴² suggesting that it plays an important role in maintaining the continuing health of all these cells. Exactly what the role of Nurr 1 is in the development of idiopathic Parkinson's disease remains uncertain. Treatment of Nurr 1 deficient (-/+) animals with MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) showed that they were more susceptible to the effects of the neurotoxin than normal (+/+) mice.⁴³ The identification of such a factor, which acts from early fetal life onwards, raises the possibility that differences in the amount of expression and/or the timing of the onset of expression in the fetal period may be important in predisposing these cells to an earlier that normal death and, thence, the development of idiopathic Parkinson's disease. Other intriguing possibilities that surround Nurr 1 centre on its potential ligand binding ability and its immune responsiveness. As mentioned earlier, Nurr 1 is classed as an orphan receptor. It may act as a transcription factor without an endogenous natural ligand,⁴⁴ either binding as a monomer to its response element, or as a dimer.⁴⁵ However, this does not of necessity rule out the possibility that it can bind ligands, and binding of any such ligand may alter its function beneficially or deleteriously for the health of the cell. Until developments in molecular biology led to a re-evaluation of the evidence in the past decade, idiopathic Parkinson's disease was thought to result exclusively from non-genetic causes. However, it is now generally accepted that genetics play a part. Nevertheless, as will be reviewed later, environmental causes are still thought to be of great importance. Any ability of Nurr 1 either to bind endogenous or exogenous ligands or to change in concentration in response to exposure to viruses or other noxious stimuli offers a portal to see how the health of these cells might be uniquely adversely affected by environmental factors. Furthermore, not only does Nurr 1 offer the possibility of theoretical insights, but also that of practical exploration of dopaminergic neuronal cell biology and replacement tissue for implantation. In the past it was possible to culture neuronal cells, but these cultures were highly heterogeneous mixtures of cells, making culture to culture comparisons extremely difficult. However, a culture system has been described that overcomes this problem. This involves three steps, namely: (1) the transfection of neuronal stem cells from mouse cerebellum with a Nurr 1 expression plasmid so that they overexpress Nurr 1; (2) the propagation of the cells in the presence of basic fibroblast growth factor; and (3) co-culture of the cells in the presence of type 1 astrocytes to supply other growth factors. The protocol leads to the development of a dopaminergic phenotype in approximately 80% of the resultant neurones.⁴⁶ Such cultures should provide a model in vitro system to enable a detailed exploration of the physiology of these unique cells. Further advances in cell culture techniques, such as that described by Kawasaki et al, which allow a slightly simpler method of inducing a dopaminergic phenotype from a modified embryonic stem cell line, will also greatly aid this exploration.47 However, the close structural relation of Nurr 1 to other

members of its subfamily suggests that it should not be considered in isolation.

NERVE GROWTH FACTOR INDUCIBLE B (NGFI-B) (ALSO KNOWN AS NUR 77/TR3/NAK1/N10/ST59/TIS) AND NEURONE DERIVED ORPHAN RECEPTOR 1 (NOR-1) (ALSO KNOWN AS MINOR/TEC/CHN/NOR2) Nurr 1, NGFI-B, and NOR-1 form a subfamily within the "zinc finger" nuclear receptor superfamily. Of the three, NGFI-B was the first to be characterised. Like Nurr-1, NGFI-B and NOR-1 play important roles in brain development, and both are expressed in tissues outside the CNS. The regional expression of NGBI-B and NOR-1 within the CNS is different from that of Nurr 1, with that of NGFI-B being wider, but the three do overlap, suggesting selective roles for these transcription factors in the regulation of motor function.48 49 Table 2 summarises the expression of the three subfamily members in the CNS. Of possible relevance to idiopathic Parkinson's disease are the facts that: (1) both NGFI-B and NOR-1 are expressed in the caudate/putamen, the target site for dopaminergic neurones from the substantia nigra, but are not expressed in the substantia nigra itself; and (2) NGFI-B, NOR-1, and Nurr 1 are involved in the regulation of dopamineric neurone formation in the olfactory bulb, where NGFI-B is strongly

layers.51 Human NGFI-B cDNA is 2498 bp in length, with an open reading frame of 1794 base pairs, which encodes a protein of 598 amino acids and a predicted molecular mass of 64 kDa.52 53 The NGFI-B structural gene is encoded on human chromosome 12q13.1,54 and the gene for NOR-1 is located on human chromosome 9q22, spans some 35 kb, and has eight exons.⁵⁵ The NOR-1 gene gives rise to two transcripts, which when translated result in a protein containing 626 amino acids, with an approximate molecular weight of 68 kDa. All three receptors bind to a common response element—AAAAGGTCA—as monomers.56 Nurr 1 and NGFI-B bind to AGGTCA repeats as heterodimers with RXR isoforms,¹⁷ as homodimers,⁵⁷ and as a heterodimer with each other.58 NOR-1 is unusual in that it does not heterodimerise with RXR. x Ray crystallography has shown that, when NGFI-B binds as a

expressed in the glomerular and granule cell

 Table 2
 Regional expression of Nurr 1, NGFI-B, and

 NOR-1 mRNA in rat and mouse brain¹² ¹⁸ ⁴⁸ ⁵⁰

	Nurr 1		NGFI-B		NOR-1	
Area of brain	Α	В	Α	В	A	В
Neocortex		++		++		+
Caudate putamen		_		++		++
Hippocampus		++		++		++
Olfactory bulb	+		++		++	
Ventral tegmental						
area		++		-		_
Substantia nigra		++		-		_
Subiculum		++		++		++
Locus coeruleus		-		-		_
Cerebellum	++		++		++	
Spinal cord	-		-		-	

A, early, neonatal life; B, later life.

++, strong expression; +, moderate expression; -, weak or no expression.

monomer, the DNA binding domain of the receptor not only interacts with the core section of the response element (AGGTCA) in the major groove, but also with the AAA extension via the minor groove.⁵⁹

Like Nurr 1, NGFI-B and NOR-1 are intermediate early genes. NGFI-B mRNA in brain was induced rapidly and transiently by growth stimulating agents,^{53 60} and in hippocampal neurones in response to NMDA and muscarinic receptor stimulation.⁶¹ Both NGFI-B and NOR-1 are expressed in the fetal brain and are important signals in mitogenesis⁶² and apoptosis pathways in tissues outside the CNS.^{63 64}

The role of these receptors in apoptotic signalling is complex. Both NGFI-B and NOR-1 generally are regarded as proapoptotic factors in tissue outside the CNS.⁶⁵ ⁶⁶ This appears to be true for NGFI-B in the adult human CNS too, with high amounts of the receptor being detectable in the brains of patients with Alzheimer's disease.⁶⁷. However, as indicated above, both NGFI-B and NOR-1 are constitutively expressed in some regions of the brain in adult life where high rates of apoptotic neural death are not occurring, so obviously they have other functions, and in a model system where overexpression of NGFI-B was induced, it inhibited ceramide induced apoptosis but not the Fas-Fas ligand pathway.68 Despite these somewhat contradictory strands of evidence, an understanding of the role of NGFI-B in apoptosis and gene transcription is beginning to emerge. An essential step in the initiation of apoptosis is the release of cytochrome c from mitochondria, which then activates the caspase cascade.⁶⁹ After exposure of the prostate cancer cell line LNCaP to 6-[3-(1-adamantyl)-4hydroxyphenyl]-3-chloro-2-naphthalene carboxylic acid and other proapoptotic agents, human NGFI-B was induced and the protein was shown to move from the nucleus to the mitochondrion to trigger cytochrome c release. Therefore, in its role in apoptosis, human NGFI-B is not required to initiate gene transcription. Signals directing movement out of the nucleus were contained in both the N-terminus and C-terminus of the molecule. Apoptosis was inhibited by antisense human NGFI-B mRNA. In contrast, epidermal growth factor (EGF)-a non-apoptotic stimulus-also induced human NGFI-B mRNA, but the protein produced stayed within the nucleus and was capable of initiating gene transcription.⁷⁰ Therefore, it seems that the contrasting actions of human NGFI-B are modulated by its intracellular localisation, which in turn is dependent upon the nature of the signal to which the cell is exposed.

Export of human NGFI-B from the nucleus is also a means of modulating gene transcription via interaction with the retinoid signalling system. After nerve growth factor induced phosphorylation of NGFI-B serine residue 105, the NGFI-B–RXR complex leaves the nucleus, reducing the availability of RXR for heterodimerisation with RAR, which thus reduces the transcriptional activity of the RAR–RAR complex.⁷¹ Export of the RXR– NGFI-B complex from the nucleus has also been suggested as the mechanism whereby retinoids inhibit the activation induced apoptosis of immature thymocytes. It was suggested that this occurred because of the reduced ability of NGFI-B to initiate gene transcription,⁷² but it may be the result of the inability of the RXR–NGFI-B heterodimer to interact with the mitochondrion, in contrast to NGFI-B alone.

The recent work on the role of NGFI-B in apoptosis has been carried out largely in non-CNS cells, and whether the same actions and mechanisms apply in the CNS remains to be seen. However, NGFI-B is inducible in the dopaminergic target areas in the striatum in response to burst stimulation of the medial forebrain bundle,73 and both NGFI-B and NOR-1 are inducible in response to therapeutic drugs (halopyridol)74 and substances of abuse (cocaine and morphine).75 The induction of NGFI-B and NOR-1 was suggested as a possible mechanism whereby retinoid signalling was disturbed, so that it might be of relevance in schizophrenia.74 In contrast, compulsive running was shown to downregulate striatal NGFI-B and NOR-1 expression.76 Thus, these two transcription factors appear to be involved in dopaminergic related personality characteristics. However it is still unclear whether Nurr 1 has similar properties to NGFI-B, and whether and how the expression of NGFI-B and NOR-1 changes and, if it does, how such changes coincide with any alterations in the expression of Nurr 1 in idiopathic Parkinson's disease. Because of the complex way that all three principle members of the NGFI-B family act as individuals and interact with each other and other liganded nuclear receptors, such as the retinoid and glucocorticoid receptors,77 78 these proteins have the capacity to modulate the transcription of many genes, only a few of which have been recognised, and to act as both survival and proapoptotic factors, not necessarily by modulating events in the cell nucleus. This complexity is increased by the fact that further proteins are formed from the same genes by either exon splicing or the use of different transcription start sites and promoters.79 These also have transactivator properties; the Nurr 1 related protein (TINUR) derived by differential splicing is induced in T cells undergoing apoptosis.80

The elucidation of the actions of all three major proteins should give insights into dopaminergic neuronal cell survival and death in the substantia nigra, either because the function of Nurr 1 in these cells has changed or because the functions of NGFI-B or NOR-1 in neurones in target sites have changed, causing a lack of support for the incoming dopaminergic neurones. In turn, such knowledge may give insights into the biochemical basis of some of the behavioural and personality features of the disease, in addition to the cause of the neuronal death.

However, zinc finger nuclear receptors are not the only transcription factors that are of interest in dopaminergic neurone development and survival. Two others which are members of different families are being increasing recognised as important. These are Lmx1b and Ptx-3. Both are homeobox proteins.

LMX1B AND PENTAXIN 3 (PTX-3)

Although Nurr 1 expression is essential for the final differentiation of stem cells into dopaminergic neurones, it cannot initiate and complete this process by itself.⁸¹ Neuronal development and differentiation occur as a consequence of the actions of successive waves of transcription factors. In the early phases of the transformation of stem cells sonic hedgehog and fibroblast growth factor 8 (FGF-8) appear to be principal players.^{82 83} Another transcription factor that acts at this early stage is Lmx1b. The expression of this factor is stimulated by FGF-8 and, as part of this cascade, Lmx1b in turn stimulates another transcription factor, wnt1.⁸⁴

Lmx1b is a member of the LIM homeodomain protein family and was first recognised because of its role in dorsal-ventral limb patterning. It is expressed in numerous tissues where it affects skeletal, cranial, renal, and eye structures,⁸⁵⁻⁸⁷ and the trajectory of motor neurones in limbs.88 Subsequent studies have shown that loss of function mutations in the human gene are responsible for the nail-patella syndrome.⁸⁹⁻⁹⁴ Its role in dopaminergic neuronal development is only just beginning to be elucidated. It is expressed in the neural tube at an early stage in the genesis of mesencephalic dopaminergic neurones in response to FGF-8 stimulation, where it maintains the expression of another transcription factor Wnt1,⁸⁴ and in the normal adult midbrain.95 Lmx1b knockout mice fail to develop the full repertoire of dopaminergic neurones and their gene products. Although Nurr 1 and tyrosine hydroxylase are still expressed, there is a failure of expression of Ptx-3.

Later and at almost the same time as Nurr 1 is expressed, another transcription factor-Ptx-3—appears (embryonic day 11.5 in the rat).¹⁶ In the CNS, this factor is found only in mesencephalic dopaminergic neurones, unlike Nurr 1 and the other factors mentioned above, but its expression does not by itself lead to the formation of the final dopaminergic phenotype. In Nurr 1 deficient (-/-) mice, Ptx-3 is expressed normally in the progenitor cells but these do not survive.³⁷ However, the importance of Ptx-3 for these neurones can be seen by the fact that a Ptx-3 response element (GGCTTT) is present in the 5' flanking regions of the human, rat, and mouse tyrosine hydroxylase genes, and binding of the transcription factor to this element results in pronounced upregulation of transcription.³⁰ Ptx-3 expressing neurones are reduced in number in parkinsonian substantia nigra and in that of the 6-hydroxydopamine lesioned rat. However, whether the neuronal loss in these situations is related to Ptx-3 function, or the reduction in Ptx-3 is simply a consequence of the death of these neurones, is uncertain. Nevertheless, it is principally the Ptx-3 expressing neurones that die.94

Genetic and environmental aetiological factors

The above discussion has concentrated on transcription factors that are involved in dopaminergic neurogenesis and have functions, as yet ill defined, in the adult CNS. Whether they have a role in the aetiology of idiopathic Parkinson's disease is very uncertain, but they clearly have the potential to be involved in this. By definition, the causes of idiopathic Parkinson's disease are as yet unknown. Over the past two decades, scientific opinion has varied between two extreme poles: from there being only environmental causes, to the position where genetic factors are considered to be the dominant aetiological feature. One major stimulus to this refocusing of emphasis on to genetics has stemmed from the ability to visualise dopaminergic neurones in vivo by positron emission tomography (PET) scanning. Early twin studies, before PET scanning was available, showed no significant difference between the incidence of idiopathic Parkinson's disease in the second twin when monogyzotic and dizygotic pairs were compared. On PET scanning of small groups, second monozygotic twins were found to have a greater dopaminergic deficit than dizygotic partners, suggesting than genetic parameters are important.⁹⁶ A second stimulus is the early finding that continues to be re-emphasised with studies of increasing sophistication, in terms of the elimination of potential confounding factors, is that a considerable proportion of patients with apparently sporadic idiopathic Parkinson's disease have a family history of the disease.97-99 In addition, advances in molecular biological techniques, such as whole genome scanning, reviewed and illustrated in an earlier edition of this journal, have made the exploration of genetic factors possible.¹⁰⁰ This has been particularly important in elucidating the genes involved in familial Parkinson's disease.

In contrast to the twin study quoted above, other recent major studies not based on PET scanning have failed to detect a significant difference between the incidence of idiopathic Parkinson's disease in monozygotic compared with dizygotic twins, leading the authors to the conclusion that genetic factors are of minor importance in the aetiology of the disease.¹⁰¹ ¹⁰² The emphasis on causal environmental agents received a boost with the recognition of the selective toxicity of rotenone in mice.¹⁰³ This is quite widely used as a "safe" insecticide. The compound is also a potent inhibitor of mitochondrial complex 1 and thus shares a common mode of action with 1-methyl-4phenylpyridinium ion (MPP⁺) formed from in vivo oxidation of the protoxin, MPTP. The recognition that a new class of rotenone-like compounds might exist in the environment helps to counter one objection to the concept that environmental toxins might be involved in the aetiology of idiopathic Parkinson's disease, simply that MPTP is not encountered in nature. Although this is the case for MPTP, MPP⁺-like compounds may be, and this possibility is discussed in a later section.

Whether either of these opposing opinions or a combination of the two is correct remains to be seen, but because most effort has been directed to elucidating the genetic basis of familial Parkinson's disease. Consequently, these developments will be reviewed first.

FAMILIAL GENETIC LOCI

To date, the genetic loci that have been found to be associated with Parkinson's disease with fairly typical symptoms are:

- A locus on chromosome 4q, which encodes the protein *α* synuclein. The mutations involved are A30P and A53T.^{104 105}
- A locus on chromosome 6q, which encodes the protein parkin. Multiple mutations are involved.^{106–108}
- A locus on chromosome 4p, which encodes the protein ubiquitin C-terminal hydrolase (UCTH). The mutation involved is I93M.¹⁰⁹
- A locus on chromosome 4p, which encodes an unknown protein.¹¹⁰
- A locus on chromosome 2p13, which encodes two unknown proteins,^{111 112} and cytochrome b.¹¹³

In addition, there are other loci associated with atypical symptoms. For example, a locus on chromosome 17q21–22, which encodes the protein tau and which is associated with frontotemporal dementia,^{114–116} and a locus on chromosome 19q13, which is associated with rapid onset dystonia Parkinson's disease.¹¹⁷ There are other loci that are associated with other modes of inheritance than the autosomal dominant transmission described in the original family with the α synuclein mutation. These include:

- Autosomal recessive—several mutations in the parkin gene.^{106-108 118}
- Autosomal dominant inheritance because of parkin gene mutation.¹¹⁹
- Maternal transmission—mutations in mitochondrial DNA.¹²⁰

Other loci are associated with both unusual features and mode of transmission—for example, a 12S ribosomal RNA gene point mutation causing disrupted protein synthesis resulting in parkinsonism, deafness, and neuropathy.¹²¹

Despite the increasing number of kindred being described, familial parkinsonism accounts for a relatively small proportion of the total patient population; therefore, the reason for the interest in these cases, apart from that of knowing the details of the causal mutation in each, is that they may shed light on the aetiologies of sporadic cases. This is proving to be the case to a surprising extent, although the mutations in the familial cases are not found in sporadic idiopathic Parkinson's disease.122-125 (In fact, it has been suggested that another relatively common mutation in the UCTH gene, S18Y, is protective.¹²²) What has been highlighted from this work is the importance of two inter-related pathways: α synuclein fibril formation and ubiquitin targeted protein catabolism.

 α Synuclein is a member of a small family of proteins (other members being β synuclein and persyn). Under normal circumstances, it is strongly expressed in neurones in a limited

number of areas of the brain, including the dopaminergic neurones of the substantia nigra (a feature in common with parkin and in contrast to ubiquitin carboxy terminal hydrolase¹²⁶), and comprises about 2% of total brain protein.127 Its physiological role is ill defined, but it is present in nerve terminals. Its importance was recognised because of the fact that, together with proteasome and synphilin-1, and in whole and partially digested forms, it is a major component of Lewy bodies, which are a cardinal feature of the pathology of the parkinsonian brain. Of added importance are the facts that the proteins composing Lewy bodies are heavily ubiquitinylated and they are resistant to proteolysis.¹²⁸⁻¹³² The mutations discovered in the initial parkinsonian families were thought either to render α synuclein more resistant to proteolysis or to have a greater tendency to form fibrils, and in these ways accelerate Lewy body formation, thereby effectively clogging up the cytosol of the neurone

and killing it.133 In support of this relatively simple scenario was the discovery of the mutation in the ubiquitin C-terminal hydroxylase gene. The ubiquitin targeted pathway of intracellular protein catabolism utilises initially three classes of proteins, corresponding to the three steps in the first phase of the pathway-ubiquitin activation, ubiquitin conjugation, and target protein ligation-to attach the polyubiquitin tail to the protein to be destroyed. Ubiquitin C-terminal hydroxylases in the final step release ubiquitin from this polyubiquitin tail for re-use after digestion of the target protein by the 26S proteasome complex.¹³⁴ An inability to release ubiquitin from the polyubiquitin tail, as a result of deficient ubiquitin C-terminal hydrolase activity, with the resultant absent or incomplete digestion of the target protein, could allow the accumulation of the building blocks for neurotoxic fibrils.

The first difficulty with this simple scenario in which toxicity occurs because of cytosolic blockage is the Japanese families with parkin gene defects. These defects give rise to an early onset form of Parkinson's disease. In the few individuals who underwent necropsy no Lewy bodies were found. While the function of the parkin gene product was unknown, this difficulty could be explained away, but it is now clear that the gene gives rise to a ubiquitin ligase.135 These enzymes are of crucial importance for attaching the initial ubiquitin to the target protein and then extending the ubiquitin chain. Lack of Lewy bodies in individuals with gross genetic mutation that inactivate the enzyme presumably arises from the fact that α synuclein and other proteins are not ubiquitinylated, so they do not progress to the point where partially digested material is available for fibril formation. Hence, simple cytosol blockage by insoluble proteins is not the only cause of cell death, which, therefore, may be caused by the inability to clear soluble proteins in these cases.

Second, in most sporadic cases investigated no mutation in the α synuclein gene has been found and it is not clear from in vitro

experiments that the variant α synucleins have a significantly greater rate of fibril formation when compared with that of the wild-type, which nevertheless does form fibrils.¹³⁶ These facts pose questions of how Lewy body formation occurs in sporadic Parkinson's disease and how this is related to cell death. Factors that may be more relevant than the rate of fibril formation are interactions with other proteins, such as synphilin-1¹³⁷; the rates of formation of soluble oligomers of α synuclein, a step that precedes fibril formation,^{138 139} which affects mitochondrial function¹⁴⁰; and the nucleation of fibril formation.¹⁴¹ In the last instance, the role of iron as the instigator of fibril nucleation has been suggested to be important.142 In health, the pigmented dopaminergic neurones normally have relatively high concentrations of both iron and copper, and in the clinical phase of the disease increased amounts of low molecular weight iron and copper compounds are released in the cerebrospinal fluid.¹⁴³ Thus, the iron released from one dead or dving cell may be available to propagate fibril nucleation in other cells.

Third, it is worth repeating that none of the genetic abnormalities seen in the familial cases is seen in most of the patients. Therefore, if the inability to catabolise α synuclein and other proteins is not only the basis of Lewy body formation but the ultimate reason why the neurone dies, a defect must exist that precedes and precipitates this. Such a conclusion is supported by the existence of the well characterised familial cases that result from mitochondrial DNA mutations where genomic DNA mutations can be discounted.¹²⁰ This leads back to themes that are familiar to those in the immediate field: oxidative stress, mitochondrial complex 1 underactivity and energy production, excitotoxicity, and dopamine metabolism. In addition, as pointed out by Goldberg and Lansbury, one consequence of events before fibril formation occurring being of prime importance may be that a therapeutic approach directed solely at inhibiting the polymerisation of protein into fibrils might not be beneficial.138

ENDOGENOUS GENERATION OF SELECTIVE NEUROTOXINS

The evidence that the substantia nigra in idiopathic Parkinson's disease experiences free radical mediated oxidative stress has been reviewed exhaustively on numerous occasions,144 and the abnormally low mitochondrial complex 1 activity well documented previously,¹⁴⁵ so neither will be discussed in detail here. That overexcitation of receptors such as the α 1B-adrenergic receptor might lead to neurodegenerative disease also continues to receive support, as in the recent findings in multiple system atrophy.146 However, in idiopathic Parkinson's disease the things that are not clear are: (1) whether oxidative stress precedes complex 1 inhibition or vice versa, and (2) how excitotoxicity relates to the two. The MPTP model of idiopathic Parkinson's disease would suggest that underactivity of complex 1 leads to oxidative stress. This

reduced activity could occur because of the presence of some inhibitory neurotoxin, or as a result of the accumulation of random replication errors in mitochondrial DNA. In terms of neurotoxins, two classes of compounds have been investigated extensively: cysteine-dopamine reaction products and MPP⁺-like molecules.

The salient feature of MPP⁺ is the aromatic quaternary N-methyl, which allows it to bind to complex 1 but, because such a charged molecule would not pass the blood-brain barrier, intra CNS routes of synthesis are required, as in the case of the conversion of MPTP to MPP⁺ by monoamine oxidase B. One such route is N-methylation, with simple pyridines, β carbolines, and tetrahydroisoquinolines as substrates. Members of these classes of compounds are present in the food chain, the general environment, and in the human brain and cerebrospinal fluid.¹⁴⁷ Enzymes that can convert them into their N-methylpyridinium analogues are also present in the mammalian brain.148 In an apparently analogous fashion to MPP⁺, the N-methyl derivatives are selectively neurotoxic, interact with the dopamine uptake system, and inhibit mitochondrial respiration. In the case of β carbolines, the molecule can be N-methylated at two sites, and the kinetics of the two reactions are different. However, the enzyme(s) responsible has not been isolated and cloned, so it is not clear whether the two activities observed represent different actions of the same enzyme or two entirely different ones. Following on from earlier work by Matsubara *et al*,¹⁴⁷ who showed that β carboline concentrations in the cerebrospinal fluid of patients with Parkinson's disease were higher than those of control subjects, Gearhart et al

have shown increased β carboline 9*N*methyltransferase activity in the frontal cortex of parkinsonian brain compared with that of non-parkinsonian brain.¹⁴⁹ Thus, there appears to be both higher amounts of substrate and one form of enzyme activity in the disease population.

Our own work in this area has concentrated nicotinamide-N-methyltransferase on (NNMT), which, apart from catabolising the amide form of vitamin B3, has a wide substrate specificity that enables it to generate a variety of potentially toxic N-methylpyridinium ions. Its expression is dominated by an as yet undetermined, non-coding, genetic polymorphism, which results in a quarter of the general population having high hepatic enzymic protein and activity values.¹⁵⁰ Because nicotinamide is an integral part of NADH, which is the electron and hydrogen donor for complex 1, catabolism of nicotinamide could conceivably have an effect on intracellular NADH values. Thus, NNMT links, albeit tenuously, the ability to produce N-methylpyridinium ions, the metabolism of NADH, complex 1 activity, and the genetics of 25% of the general population. In exploring the hypothesis that NNMT may be an aetiological factor in idiopathic Parkinson's disease, some of our initial findings are that the enzyme is expressed in neurones and in higher amounts in two regions of the brain (caudate nucleus and cerebellum) in Parkinson's disease compared with control brain (cerebellum, fig 1). The enzyme is expressed in the substantia nigra,¹⁵¹ but the destruction of so many neurones that has occurred in this area by the time subjects reach necropsy makes comparisons between parkinsonian and nonparkinsonian material difficult to interpret.

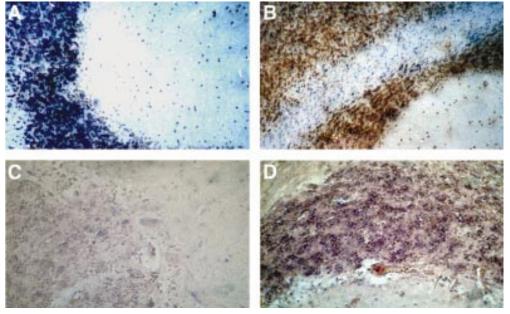


Figure 1 Expression of nicotinamide-N-methyltransferase (NNMT) protein and mRNA in control (non-Parkinsonian) and Parkinsonian cerebellum. (A) Protein expression in non-Parkinsonian cerebellum. Counterstain only (blue) is present. (B) Protein expression in the Parkinsonian cerebellum. Brown staining indicating the presence of NNMT protein is present solely in the neurones of the granular layer, with little or no staining in the white matter and the molecular layer. (C) mRNA expression in the non-Parkinsonian cerebellum. Little expression indicated by blue/purple staining was present in the granular layer. (D) mRNA expression in the Parkinsonian cerebellum. Pronounced blue/purple staining indicating the presence of NNMT mRNA is present solely in the granular layer, with no staining in either the white matter or the molecular layer.

How NNMT relates to β carboline *N*-methyltransferases and tetrathydroisoquinoline *N*-methyltransferase is unclear, but as Gearhart *et al* pointed out, these may all be the same entity.¹⁴⁹ If so, the collective evidence from the three areas would present a strong case for *N*-methylpyridinium ion formation being an important aetiological factor.

A second area of biochemistry where the generation of neurotoxins has been explored is that of dopamine metabolism. The arguments as to whether dopamine itself is deleterious to the neurone when considered in relation to L-DOPA treatment have been well rehearsed previously and will not be repeated here. One aspect of endogenous dopamine metabolism that has not received wide attention is that of its interaction with cysteine. Work in this area has been carried out in the main by the group led by Dryhurst. This group has shown that under mild oxidising conditions cysteine and dopamine or other catecholamines react together to form a variety of compounds, including dihydrobenzothiazines, which are potent inhibitors of complex 1, and free radical species.¹⁵²⁻¹⁵⁸ If the products of these reactions are involved in the aetiology of the disease, because the neurone has evolved to generate dopamine, factors that control the intracellular concentration of the other reactant (cvsteine) would be important. Within the hepatocyte, the enzyme cysteine dioxygenase is one such factor. Therefore, we explored the possibility that this enzyme, which has been shown to be expressed in brain previously,159 is expressed in dopaminergic neurones. To date, we have shown that, in the rat, the enzyme is present in numerous brain regions and the highest unstimulated activity can be seen in the basal ganglia and olfactory bulb.160 Inspection of chromosome 5 sequence data in GenBank shows a potential Ptx-3 response element within 2 kb of the transcription start site of the gene, suggesting that this exclusively dopaminergic transcription factor may be involved in the regulation of expression. Initial immunohistochemistry shows that the enzyme is present in neurones,¹⁶¹ and further work is under way to characterise expression in the human substantia nigra. Relatively little work has been done on the genetics of cysteine dioxygenase regulation, but some phenotyping studies suggest that most of the population has a low to medium degree of constitutive activity, with a small proportion being in a high activity group.¹⁶² A low activity would favour a high intracellular cysteine concentration and hence the formation of toxic reaction products.

THE GENETICS OF "NON-FAMILIAL" PARKINSON'S DISEASE: CANDIDATE GENES

The work outlined in the previous section may be categorised as a "candidate toxin" approach to elucidate the aetiology of idiopathic Parkinson's disease. However, the general emphasis on genetics has led to other themes such as xenobiotic metabolism and links with Alzheimer's disease. Polymorphic alleles are sought that are more or less common in the

idiopathic Parkinson's disease population compared with the control population, controls being either age and sex matched to varying extents, or sibling pairs. Many genes have been investigated and a few phenotypic studies carried out. Examples of genes investigated in some recent studies are CYP1A1,¹⁶³ the dopamine D2 receptor gene,¹⁶⁴ tau,¹⁶⁵ and apolipoprotein E.¹⁶⁶ In addition, associations between Parkinson's disease and combinations of genetic loci, such as α synuclein and apo lipoprotein E alleles¹⁶⁷ have been investigated. On the whole, the results of such studies have been unconvincing. One group will present evidence of an allele being positively associated with the development of idiopathic Parkinson's disease, only for another group either to fail to find the correlation with the disease or to give another interpretation. An example of this is the case of the NAT2 alleles, where Bandmann and colleagues¹⁶⁸¹⁶⁹ claim a positive association between the "slow acetylator" genotype and the development of idiopathic Parkinson's disease in what they call a "familial" Parkinson's disease group—a group composed of individuals with at least one affected relative-as opposed to the classic, "large family" studies, which identified the mutant α synuclein and parkin genes referred to earlier. This was suggested in a smaller scale phenotypic study too.¹⁷⁰ In contrast, Harhangi et al had failed to find this connection in randomly selected patients with idiopathic Parkinson's disease and claim that the genotype is a marker of mortality in the general population.¹⁷¹ We also failed to find this connection,163 although patient selection was not identical to that of Bandmann et al.

The question arises, therefore, of how to interpret the results of such studies. Where two or more groups have put forward opposing findings, has one side simply got it wrong? If so, how does one tell which? In the case of CYP2D6 alleles, which were suggested to be important by both separate, early phenotyping¹⁷² and genotyping¹⁷³ results, the weight of evidence from repeat studies by other groups has tipped the scales against these conclusions.¹⁶³ ¹⁷⁴⁻¹⁷⁶ Nevertheless, it would be depressing to think that such a process would have to be gone through for every candidate gene that is dreamt up. It may be argued that whole genome scanning will obviate this problem. However, in the case where it has been most successful in identifying susceptibility loci in a common multigene disease-diabetes mellitus, type 1-the genetic link in families is stronger than in idiopathic Parkinson's disease, much larger population groups and family clusters were studied, and controls were carefully matched. Thus, if one accepts the multigene hypothesis, it presents huge logistical problems in terms of sample acquisition and classification to achieve adequate statistical power to recognise small differences in allelic frequencies. Moreover, the case for a genetically based aetiology is still strongly challenged,¹⁰⁰ and although the multigene hypothesis may form a convenient way of explaining a baffling problem, it is not necessarily correct.

In an essential monogenic disease such as familial amyloidotic polyneuropathy a single mutation in a single gene can give rise to greatly different phenotypic forms of expression. In the case of the Met30 variant transthyretin, the Portugese expression is one of lower limb involvement and renal failure,¹⁷⁷ whereas in a Danish family the heart is the organ primarily affected.¹⁷⁸ Presumably, this arises in part from the interaction of different levels of gene products in the two populations. A similar scenario (in this case a combination of apolipoprotein E and α synuclein alleles¹⁷⁹), compounded by the effects of exposure to varieties of natural (such as homocysteine¹⁸⁰) and man made toxicants (such as lead¹⁸¹), might also explain the multifarious features of the entity we call idiopathic Parkinson's disease.

Conclusions

We are some way from unravelling the conundrum of the aetiology of idiopathic Parkinson's disease, although two big advances have been made. The first is the elucidation of the genetic defects in large affected kindreds, which has pointed to the importance of defects in ubiquitin targeted α synuclein catabolism for Lewy body formation, and as a likely end reason for death of the dopaminergic neurone in idiopathic Parkinson's disease. However, the initial precipitating problem is uncertain. Whether it is falling ATP concentrations, the accumulative effects of the inadequate clearance of oxidative free radicals, overexcitation of the neurone, or low amounts of neurotrophic support remains to be resolved.

The second major advance is the increasing recognition of the factors involved in dopaminergic neurogenesis, which, in addition to the intrinsic importance of the knowledge, will allow the application of these factors to produce good model in vitro culture systems. Hopefully, such in vitro cultures, coupled with hybrid systems, which were first described a decade ago and that are beginning to elucidate mitochondrial defects,182 183 will provide the tools that are needed to enable the conundrum to be explained.

- 1 Mages HW, Rilke O, Bravo R, et al. NOT, a human immediate-early response gene closely related to the steroid/thyroid hormone receptor NAK1/TR3. Mol Endocrinol 1994;8:1583-91.
- 2 Walters JR. Bile acids are physiological ligands for a nuclear receptor. *Gut* 2000;**46**:308–9. 3 Enmark E, Gustafsson J-A. Orphan nuclear receptors. The
- Finnark P., Gustatsson J-A. Orphan nuclear receptors. The first eight years. Mol Endocrinol 1996(10:1293–307.
 Repa JJ, Turley SD, Lobaccaro JA, et al. Regulation of adsorption and ABC-1 mediated efflux of cholesterol by RXR heterodimers. Science 2000;289:1524–9.
 Tall AR, Costet P, Luo Y. "Orphans" meet cholesterol. Nat Mod 2006;e1104 5.
- Tall AR, Costet P, Luo Y. "Orphans" meet cholesterol. Nat Med 2000;6:1104-5.
 Law SW, Conneely OM, DeMayo FJ, et al. Identification of a new brain-specific transcription factor, NURR1. Mol Endocrinol 1992;6:2129-35.
 Scearce LM, Laz TM, Hazel TG, et al. RNR-1, a nuclear receptor in the NGFI-B/Nur77 family that is rapidly induced in regenerating liver. J Biol Chem 1993;268:8855-61
- Baulsen RF, Granas K, Johnsen H, et al. Three related brain nuclear receptors, NGFI-B, Nurr1, and NOR-1, as transcriptional activators. *J Mol Neurosci* 1995;6:249–55.
 Castillo SO, Xiao Q, Lyu MS, et al. Organization, sequence, chromosomal localization, and promoter identification of the mouse orphan nuclear receptor Nurr1 gene. Genomics
- 1997;**41**:250–7. 10 Saucedo-Cardenas O, Kardon R, Ediger TR, *et al.* Cloning and structural organization of the gene encoding the murine nuclear receptor transcription factor, NURR1. Gene 1997;187:135-9.

- 11 Torii T, Kawarai T, Nakamura S, et al. Organization of the human orphan nuclear receptor Nurr1 gene. Gene 1999:230.225-32
- 12 Maruyama K, Tsukada T, Bandoh S, et al. Expression of the putative transcription factor NOR-1 in the nervous, the endocrine and the immune systems and the developing brain of the rat. Neuroendocrinology 1997;65:2-8.
- Ohkura N, Hosono T, Maruyama K, et al. An isoform of
- Nurri Functions as a negative inhibitor of the NGFI-B family signaling. *Biochim Biophys Acta* 1999;1444:69–79.
 Ichinose H, Ohye T, Suzuki T, *et al.* Molecular cloning of the human NurrI gene: characterization of the human gene and cDNAs. *Gene* 1999;230:233–9.
 Wallen A, Zetterstrom RH, Solomin L, *et al.* Fate of mesencable AHD2 averaging the paragraphic sells.
- cephalic AHD2-expressing dopamine progenitor cells in NURR1 mutant mice. *Exp Cell Res* 1999;**253**:737-46.
- 16 Saucedo-Cardenas O, Quintana-Hau JD, Le WD, et al. Nurr1 is essential for the induction of the dopaminergic phenotype and the survival of ventral mesencephalic late dopaminergic precursor neurons. *Proc Natl Acad Sci U S A* 1998;**95**:4013–18.
- Zetterstrom RH, Solomin L, Mitsiadis T, et al. Retinoid X receptor heterodimerization and developmental expression distinguish the orphan nuclear receptors NGFI-B, Nurr1, and Nor1. Mol Endocrinol 1996;10:1656-66. 18 Zetterstrom RH, Solomin L, Mitsiadis T, et al. Retinoid X
- receptor heterodimerization and developmental expression distinguish the orphan nuclear receptors NGFI-B, Nurr1, and Nor1. *Mol Endocrinol* 1996;10:1656–66. Perlmann T, Jansson L. A novel pathway for vitamin A sign-
- aling mediated by RXR heterodimerization with NGFI-B and NURR1. Genes Dev 1995;9:769-82.
- and rooman BM, Umesono K, Chen J, et al. Unique response pathways are established by allosteric interactions among nuclear hormone receptors. Cell 1995;81:541–50.
 Sakurada K, Ohshima-Sakurada M, Palmer TD, et al.
- Nurr1, an orphan nuclear receptor, is a transcriptional activator of endogenous tyrosine hydroxylase in neural progenitor cells derived from the adult brain. *Development* 1999;**126**:4017–26.
- Sacchetti P, Brownschidle LA, Granneman JG, et al. Characterization of the 5'-flanking region of the human dopamine transporter gene. Brain Res Mol Brain Res 1999; 74:167-74
- 23 Picon A, Bertagna X, de Keyzer Y. Analysis of proop omelanocortin gene transcription mechanisms in bronchial tumour cells. *Mol Cell Endocrinol* 1999;**147**:93–102.
- Simeone A, Acampora D, Gulisano M, et al. Nested expression domains of four homeobox genes in developing rostral brain. *Nature* 1992;**358**:687–90.
- brain. Nature 1992;358:687-90.
 25 Breviario F, d'Aniello EM, Golay J, et al. Interleukin-1-inducible genes in endothelial cells. Cloning of a new gene related to C-reactive protein and serum amyloid P compo-nent. *β Biol Chem* 1992;267:22190-7.
 26 Semina EV, Ferrell RE, Mintz-Hittner HA, et al. A novel homeobox gene PTX3 is mutated in families with autosomal-dominant cataracts and ASMD. Nat Genet 1998;19:167-70. 1998;19:167-70.
- 27 Altmeyer A, Klampfer L, Goodman AR, et al. Promoter structure and transcriptional activation of the murine TSG-14 gene encoding a tumor necrosis factor interleukin-1-inducible pentraxin protein. J Biol Chem 1995;270:25584-90.
- Goodman AR, Levy DE, Reis LF, et al. Differential regula-
- Goodman AK, Levy DE, Keis LF, et al. Differential regula-tion of TSG-14 expression in murine fibroblasts and peri-toneal macrophages. *J Leukoc Biol* 2000;67:387–95.
 Smidt MP, van Schaick HS, Lanctot C, et al. A homeodomain gene Ptx3 has highly restricted brain expression in mesencephalic dopaminergic neurons. *Proc* Natl Acad Sci U S A 1997;94:13305–10.
 Carned R, D Stridt MD, OWHER KI, and A Sarangara.
- 30 Cazorla P, Smidt MP, O'Malley KL, *et al.* A response element for the homeodomain transcription factor Ptx3 in the tyrosine hydroxylase gene promoter. J Neurochem 2000; 4:1829-37.
- Sanchez-Garcia I, Rabbitts TH. The LIM domain: a new structural motif found in zinc-finger-like proteins. Trends
- Genet 1994;10:315–20.
 32 McIntosh I, Clough MV, Schaffer AA, et al. Fine mapping of the nail-patella syndrome locus at 9q34. Am J Hum Genet 1997:60-133-42
- 33 Dreyer SD, Morello R, German MS, et al. LMX1B transac tivation and expression in nail-patella syndrome. Hum Mol Genet 2000;9:1067-74.
- 34 Zhou T, Cheng J, Yang P, *et al.* Inhibition of Nur77/Nur11 leads to inefficient clonal deletion of self-reactive T cells. *3* Exp Med 1996;183:1879-92.
- 25 Horkariemi J, Sagar SM, Pyykonen I, et al. Focal brain injury induces multiple immediate early genes encoding zinc finger transcription factors. Brain Res Mol Brain Res 1995;28:157–63.
- 36 Zetterstrom RH, Williams R, Perlmann T, et al. Cellular expression of the immediate early transcription factors Nurr1 and NGFI-B suggests a gene regulatory role in several brain regions including the nigrostriatal dopamine system. Brain Res Mol Brain Res 1996;41:111-20.
- Zetterstrom RH, Solomin L, Jansson L, et al. Dopamine neuron agenesis in Nurr1-deficient mice. Science 1997;276: 248 - 50.
- 38 Le W, Conneely OM, Zou L, et al. Selective agenesis of mesencephalic dopaminergic neurons in Nurr1-deficient mice. Exp Neurol 1999;159:451–8.
- 39 Liu N, Baker H. Activity-dependent Nurr1 and NGFI-B ene expression in adult mouse olfactory bulb. Neuroreport 1999;10:747-51.

- 40 Baffi JS, Palkovits M, Castillo SO, et al. Differential expres-sion of tyrosine hydroxylase in catecholaminergic neurons of neonatal wild-type and Nurr1-deficient mice. *Neuro-science* 1999;93:631-42.
- Backman C, Perlmann T, Wallen A, et al. A selective group of dopaminergic neurons express Nurr1 in the adult mouse brain. Brain Res 1999;85:125–32.
- 42 Tocco G, Feldman JD, Herschman HR, et al. Nurr1 mRNA expression in neonatal and adult rat brain following kainic acid-induced seizure activity. Brain Res Mol Brain Res 1998:59.178-88
- 43 Le W, Conneely OM, He Y, et al. Reduced Nurr1 expression increases the vulnerability of mesencephalic dopamine neurons to MPTP-induced injury. J Neurochem 1999;73: 2218 - 21
- 44 Castro DS, Arvidsson M, Bondesson Bolin M, et al. Activity of the Nurr1 carboxyl-terminal domain depends on cell type and integrity of the activation function 2. J Biol Chem
- 1999;274:37483–90. Castillo SO, Xiao Q, Kostrouch Z, et al. A divergent role of COOH-terminal domains in Nurr1 and Nur77 transactiva-tion. Gene Expr 1998;7:1–12.
- Biotechnol 1999;17:653-9 Kawasaki H. Mizueli V. Stational March 1997 46 Wagner J, Akerud P, Castro DS, et al. Induction of a
- Kawasaki H, Mizuseki K, Nishiwaka, et al. Induction of midbrain neurones from ES cells by stromal cell-derived inducing factor. *Neuron* 2000;28:31–40.
 Xiao Q, Castillo SO, Nikodem VM. Distribution of messen-
- ger RNAs for the orphan nuclear receptors Nurr1 and Nur77 (NGFI-B) in adult rat brain using in situ hybridiza-
- Num V (NCM-D) in adult at the num using in start hydrauz-tion. Neuroscience 1996;5:221–30.
 Saucedo-Cardenas O, Conneely OM. Comparative distri-bution of NURR1 and NUR77 nuclear receptors in the mouse central nervous system. J Mol Neurosci 1996;7:51– 63
- 50 Bandoh S, Tsukada T, Maruyama K, et al. Differential expression of NGFI-B and RNR-1 genes in various tissues and developing brain of the rat: comparative study by quantitative reverse transcription-polymerase chain reaction. \mathcal{J} Neuroendocrinol 1997;9:3–8.
- 51 Liu N, Baker H. Activity-dependent Nurr1 and NGFI-B gene expression in adult mouse olfactory bulb. Neuroreport 1999:10:747-51.
- 52 Chang C, Kokontis J, Liao SS, et al. Isolation and characterization of human TR3 receptor: a member of steroid receptor superfamily. J Steroid Biochem 1989;34: 391 - 5
- 53 Nakai A, Kartha S, Sakurai A, et al. A human early response gene homologous to murine nur77 and rat NGFI-B, and related to the nuclear receptor superfamily. *Mol Endocrinol* 1990;4:1438-43.
- 54 Homo sapiens chromosome 12 working draft sequence, 2001, GenBank, NT 009609.
 55 Ohkura N, Ito M, Tsukada T, *et al.* Structure, mapping and
- expression of a human NOR-1 gene, the third member of the Nur77/NGFI-B family. *Biochim Biophys Acta* 1996; 1308:205-14
- 56 Murphy EP, Dobson AD, Keller C, et al. Differential regulation of transcription by the NURR1/NUR77 subfamily of
- nuclear transcription factors. *Gene Expr* 1996;5:169–79. 57 Philips A, Lesage S, Gingras R, *et al.* Novel dimeric Nur77 *Gell Biol* 1997;17:5946–51.
- 58 Maira M, Martens C, Philips A, et al. Heterodimerization between members of the Nur subfamily of orphan nuclear receptors as a novel mechanism for gene activation. Mol
- *Cell Biol* 1999;19:7549–57.
 Meinke G, Sigler PB. DNA-binding mechanism of the monomeric orphan nuclear receptor NGFI-B. *Nat Struct* Biol 1999;6:471-7
- 60 Rivest S, Laflamme N. Neuronal activity and neuropeptide gene transcription in the brains of immune-challenged rats. *J Neuroendocrinol* 1995;7:501–25.
- 61 Dragunow M, Abraham W, Hughes P. Activation of NMDA and muscarinic receptors induces nur-77 mRNA in hippocampal neurons. *Brain Res Mol Brain Res* 1996;36:
- 62 Jiang H, Movsesyan V, Fink DW, Jr, et al. Expression of human p140trk receptors in p140trk-deficient, PC12/ endothelial cells results in nerve growth factor-induced sig-rel transformed DNA methods. *Cell Biology* 1007 (2019) 1007 nal transduction and DNA synthesis. J Cell Biochem 1997; 66·229-44
- 63 Uemura H, Mizokami A, Chang C. Identification of a new
- 63 Uemura H, Mizokami A, Chang C. Identification of a new enhancer in the promoter region of human TR3 orphan receptor gene. A member of steroid receptor superfamily. *J Biol Chem* 1995;270:5427–33.
 64 Ohkubo T, Ohkura N, Maruyama K, *et al.* Early induction of the orphan nuclear receptor NOR-1 during cell death of the human breast cancer cell line MCF-7. *Mol Cell Endocrinol* 2000;162:151–6.
 65 Chem LF, Che JK, Cola D, et al. Engine and death of the formation of the orphan the formation of the orphan death of the second s
- Cheng LE, Chan FK, Cado D, et al. Functional redundancy of the Nur77 and Nor-1 orphan steroid receptors in T-cell apoptosis. *EMBO J* 1997;**16**:1865–75.
- 66 Ohkubo T, Ohkura N, Maruyama K, et al. Early induction of the orphan nuclear receptor NOR-1 during cell death of the human breast cancer cell line MCF-7. Mol Cell Endocrinol 2000;162:151-6.
 7 Neuron SL Back D, Coll D, and D, Coll D, and D. Coll D,
- 67 Newman SJ, Bond B, Crook B, et al. Neuron-specific locali-sation of the TR3 death receptor in Alzheimer's disease. Brain Res 2000;857:131–40. Bras A, Albar JP, Leonardo E, et al. Ceramide-induced cell
- 68 death is independent of the Fas/Fas ligand pathway and is

prevented by Nur77 overexpression in A20 B cells. *Cell Death Differ* 2000;7:262–71.

- 69 Martinou JC, Desagher S, Antonsson B. Cytochrome c release from mitochondria: all or nothing. Nat Cell Biol 2000:2:E41-3
- 70 Li H, Kolluri SK, Gu J, et al. Cytochrome c release and apoptosis induced by mitochondrial targeting of nuclear orphan receptor TR3. *Science* 2000;**289**:1159-64.
- Statagiri Y, Takeda K, Yu ZX, et al. Modulation of retinoid signalling through NGF-induced nuclear export of NGFI-B. Nat Cell Biol 2000;2:435–40.
- 72 Kang HJ, Song MR, Lee SK, et al. Retinoic acid and its receptors repress the expression and transactivation functions of Nur77: a possible mechanism for the inhibition of apoptosis by retinoic acid. *Exp Cell Res* 2000; **256**:545–54.
- 2 200.79 74. 73 Chergui K, Svenningsson P, Nomikos GG, et al. Increased expression of NGFI-A mRNA in the rat striatum following burst stimulation of the medial forebrain bundle. Eur J Neurosci 1997;9:2370–82.
- Werme M, Ringholm A, Olson L, et al. Differential patterns of induction of NGFI-B, Nor1 and c-fos mRNAs in striatal 74 subregions by halopridol and clozapine. Brain Res 2000; 863:112-19
- Werme M, Olson L, Brene S. NGFI-B and nor1 mRNAs are upregulated in brain reward pathways by drugs of abuse: 75 different effects in Fischer and Lewis rats. Brain Res Mol Brain Res 2000;76:18-24.
- Werme M, Thoren P, Olson L, et al. Addiction-prone Lewis but not Fischer rats develop compulsive running that coin-76 cides with downregulation of nerve growth factor inducible-B and neuron-derived orphan receptor 1. f Neurosci 1999;19:6169-74.
- rosci 1999;19:6169-74. Drouin J., Maira M, Philips A. Novel mechanism of action for Nur77 and antagonism by glucocorticoids: a conver-gent mechanism for CRH activation and glucocorticoid repression of POMC gene transcription. *J Steroid Biochem Mol Biol* 1998;65:59-63.
- Philips A, Maira M, Mullick A, et al. Antagonism between Nur77 and glucocorticoid receptor for control of transcrip-tion. Mol Cell Biol 1997;17:5952–9.
 Ohkura N, Ito M, Tsukada T, et al. Alternative splicing gen-erates isoforms of human neuron-derived orphan 1 receptor-1 (NOR-1) mRNA. Gene 1998;211:79–85. 78
- Received T, Takayanagi R, Imasaki K, *et al.* cDNA cloning of a NGFI-B/nur77-related transcription factor from an apoptotic human T cell line. *J Immunol* 1995;154:3871–9.
 81 Sakurada K, Ohshima-Sakurada M, Palmer TD, *et al.*
- Nurr1, an orphan nuclear receptor, is a transcriptional activator of endogenous tyrosine hydroxylase in neural progenitor cells derived from the adult brain. *Developr* 1999;**126**:4017–26.
- Hynes M, Porter JA, Chiang C, et al. Induction of midbrain dopaminergic neurons by Sonic hedgehog. Neuron 1995; 15-35-44
- Poulsen KT, Armanini MP, Klein RD, et al. TGF beta 2 and TGF beta 3 are potent survival factors for midbrain dopaminergic neurons. *Neuron* 1994;13:1245-52.
- Adams KA, Maida JM, Golden JA, et al. The transcription factor Lmx1b maintains Wnt1 expression within the isthmic organizer. *Development* 2000;27:1857–67.
- 85 Dreyer SD, Zhou G, Baldini A, et al. Mutations in LMX1B cause abnormal skeletal patterning and renal dysplasia in nail patella syndrome. *Nat Genet* 1998;19:47–50. Chen H, Ovchinnikov D, Pressman CL, *et al.* Multiple cal-
- varial defects in Lmx1b mutant mice. Dev Genet 1998;22: 314-20.
- Pressman CL, Chen H, Johnson RL. LMX1B, a LIM 87 homeodomain class transcription factor, is necessary for mormal development of multiple tissues in the anterior seg-ment of the murine eye. *Genesis: The Journal of Genetics and Development* 2000;26:15–25.
- 88 Kania Á, Johnson RL, Jessell TM. Coordinate roles for LIM homeobox genes in directing the dorsoventral trajectory of motor axons in the vertebrate limb. *Cell* 2000;**102**:161–73.
- 89 Hamlington JD, Clough MV, Dunston JA, et al. Deletion of a branch-point consensus sequence in the LMX1B gene causes exon skipping in a family with nail patella syndrome. Eur J Hum Genet 2000;8:311–14.
- Clough MV, Hamlington JD, McIntosh I. Restricted distri-bution of loss-of-function mutations within the LMX1B 90 genes of nail-patella syndrome patients. Hum Mutat 1999; 14:459-6
- 14:459-65.
 Seri M, Melchionda S, Dreyer S, et al. Identification of LMX1B gene point mutations in Italian patients affected with nail-patella syndrome. Int J Mol Med 1999;4:285-90.
 McIntosh I, Dreyer SD, Clough MV, et al. Mutation analy-sis of LMX1B gene in nail-patella syndrome patients. Am J Hum Genet 1998;63:1651-8.
 Velweich D, Leveich MV, et al. Leve et al.
- Vollrath D, Jaramillo-Babb VL, Clough MV, et al. Loss-of-function mutations in the LIM-homeodomain gene, 93 LMX1B, in nail-patella syndrome. Hum Mol Genet 1998;7: 1091 - 8
- 94 Chen H, Lun Y, Ovchinnikov D, et al. Limb and kidney defects in Lmx1b mutant mice suggest an involvement of LMX1B in human nail patella syndrome. Nat Genet 1998; 19:51-5
- 95 Smidt MP, Asbreuk CH, Cox JJ, et al. A second independent pathway for development of mesencephalic dopaminergic
- neurons requires Lmx1b. *Nat Neurosci* 2000;3:337–41. Piccini P, Burn DJ, Ceravolo R, *et al.* The role of inheritance in sporadic Parkinson's disease: evidence from a longitudi-96 nal study of dopaminergic function in twins. Ann Neurol 1999;45:577-82.

- 97 Autere JM, Moilanen JS, Myllyla VV, et al. Familial aggrega-tion of Parkinson's disease in a Finnish population. J Neurol Neurosurg Psychiatry 2000;69:107–9. 98 Elbaz A, Grigoletto F, Baldereschi M, et al. Familial aggre-
- gation of Parkinson's disease: a population-based case-control study in Europe. EUROPARKINSON study group. Neurology 1999;52:1876–82. 99 Taylor CA, Saint-Hilaire MH, Cupples LA, et al. Environ-
- mental, medical, and family history risk factors for Parkin-son's disease: a New England-based case control study. Am f Med Genet 1999;**88**:742–9.
- 100 Bennett P. Microsatellites. J Clin Pathol: Mol Pathol 2000;53:177-83.
- 101 Tanner CM, Ottman R, Goldman SM, et al. Parkinson's disease in twins: an etiological study. JAMA 1999;281: 376 - 8
- 102 Vieregge P, Hagenah J, Heberlein I, et al. Parkinson's disease in twins: a follow-up study. Neurology 1999;53:566-
- 12. 12. 103 Ferrante RJ, Schulz JB, Kowall NW, et al. Systemic admin-istration of rotenone produces selective damage in the striatum and globus pallidus, but not in the substantia nigra. Brain Res 1997;753:157–62.
- 104 Polymeropoulos MH, Lavedan C, Leroy E, et al. Mutation in the alpha-synuclein gene identified in families with Par-kinson's disease. *Science* 1997;**27**6:2045–7. 105 Kruger R, Kuhn W, Muller T, *et al.* Ala30Pro mutation in
- the gene encoding alpha-synuclein in Parkinson's disease. Nat Genet 1998;**18**:106–8.
- 106 Kitada T, Asakawa S, Hattori N, et al. Mutations in the parkin gene cause autosomal recessive juvenile parkinson-ism. Nature 1998;392:605-8.
- 107 Matsumine H, Saito M, Shimoda-Matsubayashi S, et al. Localization of a gene for an autosomal recessive form of juvenile Parkinsonism to chromosome 6q25.2–27. Am J Hum Genet 1997;60:588–96.
- 108 Mizuno Y, Hattori N, Mori H. Genetics of Parkinson's disease. *Biomed Pharmacother* 1999;53:109–16. 109 Leroy E, Boyer R, Auburger G, *et al.* The ubiquitin
- pathway in Parkinson's disease [letter]. Nature 1998;395: 451-2.
- 110 Farrer M, Gwinn-Hardy K, Muenter M, et al. A chromosome 4p haplotype segregating with Parkinson's disease and postural tremor. Hum Mol Genet 1999;8:81-5.
- 111 Gasser T, Muller-Myhsok B, Wszolek ZK, et al. A susceptibility locus for Parkinson's disease maps to chromosome 2p13. Nat Genet 1998;18:262-5.
- 112 Gwinn-Hardy KA, Crook R, Lincoln S, et al. A kindred with Parkinson's disease not showing genetic linkage to established loci. *Neurology* 2000;54:504–7.
 113 Rana M, de Coo I, Diaz F, et al. An out of frame
- cytochrome b gene deletion from a patient with Parkinson's disease is associated with impaired complex III assembly and an increase in free radical production. Ann Neurol 2000;48:774-81.
- 114 Baker M, Kwok JB, Kucera S, et al. Localization of frontotemporal dementia with parkinsonism in an Australian kin-dred to chromosome 17q21-22. Ann Neurol 1997;42:794-
- 115 Hulette CM, Pericak-Vance MA, Roses AD, et al. Neuropathological features of frontotemporal dementia and parkinsonism linked to chromosome 17q21-22 (FTDP-17): Duke family 1684. *J Neuropathol Exp Neurol* 0, 2017 1999;**58**:859-66.
- 116 Sperfeld AD, Collatz MB, Baier H, et al. FTDP-17: an early-onset phenotype with parkinsonism and epileptic seizures caused by a novel mutation. Ann Neurol 1999;46: 708-15.
- 117 Kramer PL, Mineta M, Klein C, et al. Rapid-onset dystonia-parkinsonism: linkage to chromosome 19q13. Ann Neurol 1999;46:176-82.
- 118 Munoz E, Pastor P, Marti MJ, et al. A new mutation in the parkin gene in a patient with atypical autosomal recessive juvenile parkinsonism. *Neurosci Lett* 2000;289:66–8.
 Klein C, Pramstaller PP, Kis B, *et al.* Parkin deletions in a
- family with adult-onset, tremor-dominant parkinsonism: expanding the phenotype. *Ann Neurol* 2000;48:65–71.
 Swerdlow RH, Parks JK, Davis JN, 2nd, et al. Matrilineal inheritance of complex I dysfunction in a multigenerational
- Parkinson's disease family. Ann Neurol 1998;44:873-81. 121 Thyagarjan D, Bressman S, Bruno C, et al. A novel mitochondrial 12S rRNA point mutation in Parkinsonism, deafness and neuropathy. Ann Neurol 2000;48:730-6.
- 122 Maraganore DM, Farrer MJ, Hardy JA, *et al.* Case-control study of the ubiquitin carboxy-terminal hydrolase L1 gene in Parkinson's disease. *Neurology* 1999;**53**:1858–60. 123 Harhangi BS, Farrer MJ, Lincoln S, *et al.* The Ile93Met
- mutation in the ubiquitin carboxy-terminal-hydrolase-L1 gene is not observed in European cases with familial Parkinson's disease. Neurosci Lett 1999;270:1-4
- 124 Lincoln S, Vaughan J, Wood N, et al. Low frequency of pathogenic mutations in the ubiquitin carboxy-terminal hydrolase gene in familial Parkinson's disease. Neuroreport 1999;10:427-9.
- 125 Vaughan J, Durr A, Tassin J, et al. The alpha-synuclein Ala53Thr mutation is not a common cause of familial Parkinson's disease: a study of 230 European cases. European consortium on genetic susceptibility in Parkinson's disease. Ann Neurol 1998;44:270-3.
- 126 Solano SM, Miller DW, Augood SJ, et al. Expression of alpha-synuclein, parkin, and ubiquitin carboxy-terminal hydrolase L1 mRNA in human brain: genes associated with familial Parkinson's disease. Ann Neurol 2000;47:201-10.

- 127 Doran JF, Jackson P, Kynoch PA, et al. Isolation of PGP 9.5, a new human neurone-specific protein detected by high-resolution two-dimensional electrophoresis. J Neuro chem 1983;40:1542-7.
- 128 Irizarry MC, Growdon W, Gomez-Isla T, et al. Nigral and cortical Lewy bodies and dystrophic nigral neurites in Par-kinson's disease and cortical Lewy body disease contain alpha-synuclein immunoreactivity. J Neuropathol Exp Neurol 1998;57:334-7.
- 129 Spillantini MG, Crowther RA, Jakes R, et al. Alpha-synuclein in filamentous inclusions of Lewy bodies from Parkinson's disease and dementia with Lewy bodies. Proc Natl Acad Sci USA 1998;95:6469-73.
- 130 Wakabayashi K, Matsumoto K, Takayama K, et al. NACP, a presynaptic protein, immunoreactivity in Lewy bodies in Parkinson's disease. *Neurosci Lett* 1997;**239**:45–8.
- 131 Wakabayashi K, Engelender S, Yoshimoto M, et al. Synphilin-1 is present in Lewy bodies in Parkinson's disease. Ann Neurol 2000;47:521–3.
- 132 Ii K, Ito H, Tanaka K, et al. Immunocytochemical co-localization of the proteasome in ubiquitinated structures in neurodegenerative diseases and the elderly. J Neuropathol Exp Neurol 1997;56:125-31.
- 133 El-Agnaf OM, Jakes R, Curran MD, et al. Aggregates from mutant and wild-type alpha-synuclein proteins and NAC peptide induce apoptotic cell death in human neuroblas-toma cells by formation of beta-sheet and amyloid-like filaments. FEBS Lett 1998;440:71-5
- 134 Hershko A, Ciechanover A. The ubiquitin system. Annu Rev Biochem 1998;7:425–79.
- 135 Shimura H, Hattori N, Kubo Si, et al. Familial Parkinson disease gene product, parkin, is a ubiquitin-protein ligase.
- Nat Genet 2000;25:302-5.
 136 Baba M, Nakajo S, Tu PH, et al. Aggregation of alpha-synuclein in Lewy bodies of sporadic Parkinson's disease and dementia with Lewy bodies. Am J Pathol 1998; 152:879-84.
- 137 Engelender S, Kaminsky Z, Guo X, et al. Synphilin-1 asso ciates with alpha-synuclein and promotes formation of cytosolic inclusions. Nat Genet 1999;22:110-14.
- 138 Goldberg MS, Lansbury PT, Jr. Is there a cause-and-effect relationship between alpha-synuclein fibrillization and Parkinson's disease? Nat Cell Biol 2000;2:E115-9.
- 139 Conway KA, Lee SJ, Rochet JC, et al. Acceleration of oligermerization, not fibrillation is a shared property of both alpha-synuclein mutations. Proc Natl Acad Sci U S A 2000;**97**:571–6.
- 140 Saha AR, Ninkina NN, Hanger DP, et al. Induction of neuronal death by alpha-synuclein. Eur J Neurosci 2000;12: 3073 - 7
- 141 Wood SJ, Wypych J, Steavenson S, et al. Alpha-synuclein fibrillogenesis is nucleation-dependent. Implications for the pathogenesis of Parkinson's disease. J Biol Chem 1999; 274:19509-12.
- 142 Ostrerova-Golts N, Petrucelli L, Hardy J, et al. The A53T alpha-synuclein mutation increases iron-dependent aggre-
- gation and toxicity. J Neurosci 2000;20:6048-54. 143 Dexter DT, Wells FR, Agid F, et al. Increased nigral iron content in post mortem parkinsonian brain. Lancet 1987;i: 1219 - 20
- 144 Owen AD, Schapira AH, Jenner P, et al. Indices of oxidative stress in Parkinson's disease, Alzheimer's disease and dementia with Lewy bodies. J Neural Transm Suppl 1997;51:167-73
- 145 Schapira AH. Mitochondrial complex I deficiency in Parkinson's disease. Adv Neurol 1993;60:288–91.
- 146 Zuscik MJ, Sands S, Ross SA, et al. Overexpression of the a1B-adrenergic receptor causes apoptotic neuro-degeration:multiple system atrophy. Nat Med 2000;6: 1388-94.
- 147 Matsubara K, Kobayashi S, Kobayashi Y, et al. Beta-carbolinium cations, endogenous MPP+ analogs, in the lumbar cerebrospinal fluid of patients with Parkinson's disse. Neurology 1995;45:2240-5
- 148 Collins MA, Neafsey EJ, Matsubara K, et al. Indole-N-methylated beta-carbolinium ions as potential brain-bioactivated neurotoxins. Brain Res 1992;570:154-60.
 149 Gearhart DA, Collins MA, Lee JM, et al. Increased beta-carboline N-methyltransferase activity in the frontal cortex in Parkinson's disease. Neurobiol Dis 2000;7:201-11.
 150 Strick M L Burgert D. Burgert M. D. Burgert M. A. disertion of the second sec
- 150 Smith M-L, Burnett D, Bennett P, et al. A direct correlation between nicotinamide N-methyltransferase activity and protein levels in human liver cytosol. *Biochim Biophys Acta* 1998;**1422**:238–44.
- 151 Williams AC, Smith M-L, Waring RH, et al. The aetiology of idiopathic Parkinson's disease: a genetic and environ-mental model. Adv Neurol 1999;80:215-18.
- 152 Zhang F, Dryhurst G. Effects of L-cysteine on the oxidation chemistry of dopamine: new reaction pathways of potential relevance to idiopathic Parkinson's disease. J Med Chem 1994;34:1084–98.
- 153 Li H, Shen XM, Dryhurst G. Brain mitochondria catalyze the oxidation of 7-(2-aminoethyl)-3,4-dihydro-5-hydroxy-2H-1,4-benzothiazine-3-carboxylic acid (DHBT-1) to in-termediates that irreversibly inhibit complex I and scavenge glutathione: potential relevance to the pathogenesis of Par-kinson's disease. J Neurochem 1998;71:2049-62.
- 154 Shen XM, Dryhurst G. Iron- and manganese-catalyzed autoxidation of dopamine in the presence of L-cysteine: possible insights into iron- and manganese-mediated dopaminergic neurotoxicity. *Chem Res Toxicol* 1998;11: 824-37.

- 155 Shen XM, Zhang F, Dryhurst G. Oxidation of dopamine in the presence of cysteine: characterization of new toxic products. *Chem Res Toxicol* 1997;10:147–55.
 156 Cheng FC, Kuo JS, Chia LG, et al. Elevated 5-S-cysteinyldopamine/homovanillic acid ratio and reduced homovanillic acid in cerebrospinal fluid: possible markers and netatinilic acid in the pathenticiloury of Backingane, and potential insights into the pathoetiology of Parkinson's disease. J Neural Transm 1996;103:433-46.
 157 Shen XM, Dryhurst G. Further insights into the influence of L-cysteine on the oxidation chemistry of dopamine:
- reaction pathways of potential relevance to Parkinson's dis-ease. Chem Res Toxicol 1996;9:751-63.
- 158 Xin W, Shen XM, Li H, et al. Oxidative metabolites of 5-S-cysteinylnorepinephrine are irreversible inhibitors of mitochondrial complex I and the alpha-ketoglutarate dehydro-genase and pyruvate dehydrogenase complexes: possible implications for neurodegenerative brain disorders. Chem Res Toxicol 2000;13:749-60.
- 159 Misra CH. In vitro study of cysteine oxidase in rat brain.
- Neurochem Res 1983;8:1497-508.
 Parsons RB, Barber PC, Waring RH, et al. Human cysteine dioxygenase type I (CDO-I; EC 1.13.11.20): regional expression of activity in rat brain. Neurosci Lett 1998;248:
- 161 Ramsden DB, Parsons RB, Ho SL, et al. Further studies in xenobiotic metabolism and Parkinson's disease. Adv Neurol 2001;86:105–13.
- Mitchell SC, Waring RH, Haley CS, et al. Genetic aspects of the polymodally distributed sulphoxidation of S-carboxymethyl-L-cysteine in man. Br J Clin Pharmacol 1984;18:507-21.
- 163 Nicholl DJ, Bennett P, Hiller L, et al. A study of five candidate genes in Parkinson's disease and related neurodegen-
- erative disorders. *Neurology* 1999;53;1415–21. 164 Oliveri RL, Annesi G, Zappia M, *et al.* The dopamine D2 receptor gene is a susceptibility locus for Parkinson's disease. *Mov Disord* 2000;15:127–31.
- Insease. Mor Disora 2000;15:121–51.
 Isopara 2000;15:121–51.
 Isopara 2000;15:121–51.
 Isopara 2000;15:121–51.
 Oliveri RL, Nicoletti G, Cittadella R, et al. Apolipoprotein
- E polymorphisms and Parkinson's disease. Neurosci Lett 1999;277:83-6.
 167 Kruger R, Vieira-Saecker AM, Kuhn W, et al. Increased susceptibility to sporadic Parkinson's disease by a certain

- susceptibility to sporadic Parkinson's disease by a certain combined alpha-synuclein/apolipoprotein E genotype. Ann Neurol 1999;45:611–17.
 168 Bandmann O, Vaughan J, Holmans PA, et al. Toxins, genetics, and Parkinson's disease: the role of N-acetyltransferase 2. Adv Neurol 1999;80:199–204.
 169 Bandmann O, Vaughan JR, Holmans P, et al. Detailed genotyping demonstrates association between the slow acetylator genotype for N-acetyltransferase 2 (NAT2) and familial Parkinson's disease. May Divard 2000:15:30-5 familial Parkinson's disease. Mov Disord 2000;15:30-5.

- 170 Milejski P, Orzechowska-Juzwenko K, Kamienowski J, et al. Clinical significance of oxidation and acetylation genetic polymorphism in patients with Parkinson's disease. Neurol Neurochir Pol 1999;33:1015-24.
- 171 Harhangi BS, Oostra BA, Heutink N-acetyltransferase-2 polymorphism in Heutink P, et al. Parkinson's disease: the Rotterdam study. J Neurol Neurosurg Psychiatry 1999:67:518-20.
- 172 Barbeau A, Cloutier T, Roy M, et al. Ecogenetics of Parkinson's disease: 4-hydroxylation of debrisoquine. Lancet 1985;2:1213-16.
- 173 Smith CA, Gough AC, Leigh PN, et al. Debrisoquine hydroxylase gene polymorphism and susceptibility to Parkinson's disease. *Lancet* 1992;**339**:1375–7.
- 174 Ho SL, Kung MH, Li LS, et al. Cytochrome P4502D6 (debrisoquine 4-hydroxylase) and Parkinson's disease in Chinese and Caucasians. Eur J Neurol 1999;6:323-9.
- 175 Christensen PM, Gotzsche PC, Brosen K. The sparteine/ debrisoquine (CYP2D6) oxidation polymorphism and the risk of Parkinson's disease: a meta-analysis. *Pharmacogenet*ics 1998;8:473–9.
- 176 Rostami-Hodjegan A, Lennard MS, Woods HF, et al. Meta-analysis of studies of the CYP2D6 polymorphism in relation to lung cancer and Parkinson's disease. Pharmacogenetics 1998:8:227-38
- Saraiva MJ, Costa PP, Goodman DS. Genetic expression 177 of a transthyretin mutation in typical and late-onset Portuguese families with familial amyloidotic polyneuropathy. Neurology 1986;36:1413-17.
- 178 Husby G, Ranlov PJ, Sletten K, et al. The amyloid in familial amyloid cardiomyopathy of Danish origin is related to pre-albumin. *Clin Exp Immunol* 1985;**60**:207–16.
- Kruger R, Vieira-Saecker AM, Kuhn W, et al. Increased susceptibility to sporadic Parkinson's disease by a certain combined alpha-synuclein/apolipoprotein E genotype. Ann Neurol 1999;45:611-17.
- 180 Kuhn W, Roebroek R, Blom H, et al. Elevated plasma levels of homocysteine in Parkinson's disease. Eur Neurol 1998;40:225-7.
- 181 Kuhn W, Winkel R, Woitalla D, et al. High prevalence of parkinsonism after occupational exposure to lead-sulfate batteries. *Neurology* 1998;50:1885-6.
- 182 Trimmer PA, Swerdlow RH, Parks JK, et al. Abnormal mitochondrial morphology in sporadic Parkinson's and Alzheimer's disease cybrid cell lines. *Exp Neurol* 2000;162: 37 - 50
- 183 Ghosh SS, Swerdlow RH, Miller SW, et al. Use of cytoplasmic hybrid cell lines for elucidating the role of mitochondrial dysfunction in Alzheimer's disease and Parkinson's disease. Ann N Y Acad Sci 1999;893:176-91.

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