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| Author(s) | Schanen, NC; Scherer, SW; Tsui, LC; Francke, U |
| Citation | Cytogenetics And Cell Genetics, 1996, v. 72 n. 2-3, p. 187-188 |
| Issued Date | 1996 |
| URL | http://hdl.handle.net/10722/42533 |
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Assignment* of the 5-hydroxytryptamine (serotonin) receptor 5A gene (HTR5A) to human chromosome band 7q36.1

N.C. Schanen, ¹ S.W. Scherer, ³ L.-C. Tsui, ³ and U. Francke ^{1,2}

Departments of Genetics and Pediatrics, and

Rationale and significance

Serotonin (5-hydroxytryptamine) plays a role in the regulation of diverse processes including sleep, appetite, aggression, and anxiety. It acts through a variety of distinct receptor subtypes which are differentially expressed in both the central and peripheral nervous systems (Peroutka, 1988). A novel receptor subtype, HTR5A, was recently identified that appears to be expressed uniquely in the central nervous system (Rees et al., 1994). We used PCR analysis of somatic cell hybrid panels and a collection of chromosome 7-specific yeast artificial chromosome (YAC) clones to map the HTR5A gene to chromosome 7, sub-band q36.1.

Materials and methods

The primary chromosome assignment to 7q was done using a panel of 19 human × Chinese hamster hybrid cell lines derived from several fusion experiments (Francke et al., 1986). Further sublocalization to 7q32 -> qter was achieved using a regional mapping panel (Tsui et al., 1995). PCR screening of YACs from the region assigned HTR5A to a YAC contig.

| Primer names | Primer sequences: (from GenBank X81412) | | | | | | |
|--------------|---|--|--|--|--|--|--|
| 5HTR5A-F | 5' CCTGTCAGTGTCCAGGCTCA 3' (nt 6–25) | | | | | | |
| 5HTR5A-R | 5' GCGGACCGTGAACACCAT 3' (nt 125–108) | | | | | | |

Amplicon: 120 bp

PCR Conditions: 1 mM MgCl2

Cycle: Denaturation at 95°C for 5 min, followed by 28 cycles at 95°C for 30 s, 62 °C for 30 s, and 72 °C for 30 s with a final extension at 72 °C for 10 min. Under these conditions, no amplification was obtained from rodent parental DNA.

Results

Table I. Segregation of HTR5A with human chromosomes in somatic cell hybrids

| Gene/ | Chr | ome | som | ie ni | ımbe | r | | | | | | | | | | | | | | | | | | |
|--------------|-----|-----|-----|-------|------|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| chromosome | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | Х | Y |
| | | | | | | | _ | | | _ | | | | | 3 | 0 | 0 | 2 | ı | 2 | 1 | 2 | 2 | 0 |
| +/+ | 0 | 0 | 1 | 1 | 0 | 1 | 3 | 1 | 2 | 1 | 1 | 1 | 1 | | _ | - | 3 | 1 | 2 | ī | 2 | 0 | 0 | 2 |
| +/ | 3 | 3 | 2 | 1 | 2 | 2 | 0 | 2 | 1 | 2 | 2 | 2 | l | 1 | • | 3 | - | _ | - | ż | 10 | 10 | 2 | 3 |
| | 6 | 4 | 9 | 6 | 5 | 10 | 0 | 9 | 2 | 2 | 7 | 7 | 10 | 12 | 6 | 9 | 2 | 7 | 10 | • | | | 4 | 13 |
| -/+ | - | | - | - | - | 5 | 14 | 6 | 12 | 14 | 7 | 9 | 5 | 3 | 10 | 5 | 14 | 9 | 5 | 9 | 5 | O | 4 | 13 |
| -/- | 9 | 11 | 5 | 8 | 11 | 3 | 14 | U | 12 | 14 | | | | | | | | | | | | | | |
| % discordant | 50 | 39 | 65 | 44 | 39 | 67 | 0 | 61 | 18 | 21 | 53 | 47 | 65 | 76 | 32 | 71 | 26 | 42 | 67 | 42 | 67 | 56 | 20 | 28 |

Data for chromosomes with rearrangements or present at very low copy (<0.1) were excluded. The % discordance is calculated as the sum of the discordant hybrids over the total number of informative hybrids for each chromosome

² Howard Hughes Medical Institute, Stanford University Medical Center, Stanford CA (USA), and

³ Department of Genetics, The Hospital for Sick Children, Toronto, Ontario (Canada)

This work was supported by NIH Grants K08HD01103 (N.C.S.) and R01HG00298 (U.F.), and the Howard Hughes Medical Institute (U.F.).

Address communications to: Dr. Uta Francke, Howard Hughes Medical Institute, Stanford University Medical Center, Stanford CA 94305-5428 (USA); telephone: 415-725-8089; fax: 415-729-8112; e-mail: Francke@cmgm.stanford.edu

Received 5 September 1995; accepted 11 September 1995.

To our knowledge this is the first time this gene has been mapped

Regional mapping results

| Hybrid designation | Chromosome region present | HTR5A present (+) or absent (-) | | | | | | |
|--------------------|---------------------------|---------------------------------|--|--|--|--|--|--|
| XXI-23A-2c | 7p | _ | | | | | | |
| Ru-Rag6-19 | 7 a | + | | | | | | |
| 2068Rag22-2 | 7q22-qter | + | | | | | | |
| GM1059-Rag5 | 7pter-q22::q32-qter | + | | | | | | |

Three YAC clones were identified that contained the gene, HSC7E769, CEPH clone 744a8, and CEPH clone 868g12. These clones are part of a YAC contig anchored by the Généthon microsatellite marker D7S637 (unpublished data) which is localized to 7q36.1 (Thacker et al., 1995). In addition, YAC clone HSC7E769 was previously mapped to 7q36 by FISH (Kunz et al., 1993).

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