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## The Kynurenine Pathway of Tryptophan Catabolism and AIDS-associated Kaposi's Sarcoma in Africa

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#### **Abstract**

**Background**—Other than Kaposi's sarcoma (KS)-associated herpesvirus and CD4+ T cell lymphopenia, the mechanisms responsible for KS in the context of HIV are poorly understood. One recently explored pathway of HIV pathogenesis involves induction of the enzyme indoleamine 2,3 dioxygenase-1 (IDO), which catabolizes tryptophan into kynurenine and several other immunologically active metabolites that suppress T cell proliferation. We investigated the role of IDO in the development of KS in HIV disease.

**Methods**—In a case-control study among untreated HIV-infected Ugandans, cases were adults with KS and controls were without KS. IDO activity was assessed by the ratio of plasma kynurenine to tryptophan levels (KT ratio), measured by liquid chromatography tandem mass spectrometry.

**Results**—We studied 631 HIV-infected subjects: 222 KS cases and 409 controls. Non-KS controls had a higher median plasma KT ratio (130, IQR: 90 to 190 nM/ $\mu$ M) than cases (110, IQR: 90 to 150 nM/ $\mu$ M) (p = 0.004). After adjustment for age, sex, CD4 count and plasma HIV RNA level, subjects with the highest (fourth quartile) plasma KT ratios had a 59% reduction (95% CI: 27% to 77%) in the odds of KS compared to those with the lowest (first quartile) levels. KS was also independently associated with lower CD4+ count, higher plasma HIV RNA, and men.

**Conclusions**—Among HIV-infected individuals, greater activity of the kynurenine pathway of tryptophan catabolism, as evidenced by higher levels of plasma KT ratio, was associated with

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**Previous presentation:** This work was previously presented in part at the 14th International Conference on Malignancies in AIDS and Other Acquired Immunodeficiencies in Bethesda, Maryland, on November 12-13, 2013 and at the 21<sup>St</sup> Conference on Retroviruses and Opportunistic Infections in Boston, Massachusetts, on March 3-6, 2014.

lower occurrence of KS. Some consequences of immune activation in HIV infection might actually suppress certain cancers.

#### **Keywords**

tryptophan; kynurenine; indoleamine 2,3-dioxygenase-1; HIV; Kaposi's sarcoma; plasma HIV RNA; Africa

Kaposi's sarcoma (KS) was a harbinger of the HIV epidemic [1] and, despite the marked reduction in its incidence since the advent of effective antiretroviral therapy (ART) [2, 3], remains the most common malignancy in people with HIV worldwide [4]. In sub-Saharan Africa, the extent of the HIV epidemic has resulted in KS becoming one of the most frequently reported cancers among all adults [4]. Although the discovery of the causative viral agent for KS, Kaposi's sarcoma-associated herpesvirus (KSHV, also known as human herpesvirus 8), was a landmark accomplishment in the etiology of KS [5], this virus is not sufficient to cause KS. In fact, other than KSHV [5] and CD4+ T cell lymphopenia [3, 6], the specific biological mechanisms responsible for the development of KS, particularly in sub-Saharan Africa, are poorly understood.

A potential mechanism for KS that has recently received attention in HIV pathogenesis is the kynurenine pathway of tryptophan catabolism [7-10]. The induction of indoleamine 2,3-dioxygenase-1 (IDO) in activated monocytes and dendritic cells by interferon-γ and other inflammatory mediators in HIV-infected individuals causes catabolism of tryptophan into kynurenine and several other downstream catabolites with immunologic properties [11, 12]. Depletion of tryptophan [13] and the accumulation of the catabolites kynurenine and picolinic acid [14] directly inhibit T cell proliferation, potentially contributing to immune deficiency. In addition to its role in HIV infection, the induction of the kynurenine pathway has been implicated in maternal tolerance of the fetal allograft [15] and in the evasion of host immune responses by several cancers [16]. In KS etiology, we hypothesized that the kynurenine pathway might promote KS by decreasing immune surveillance. Alternatively, given the inflammatory histopathological nature of KS lesions, it is conceivable that the induction of the kynurenine pathway may inhibit emergence of KS by suppression of lymphocyte proliferation.

To begin to address the role of the kynurenine pathway in the development of KS, we studied it directly in HIV-infected adults newly diagnosed with KS in sub-Saharan Africa. IDO activity can be quantified by measuring the ratio of plasma kynurenine to tryptophan (KT ratio), where a high KT ratio reflects heightened IDO induction. We also took this opportunity to formally investigate, in Africa, the relationship between KS and other factors that have been studied in other populations, such as CD4+ T cell count, plasma HIV RNA, sex and age. The overarching objective was to identify causal determinants of KS in order to inform both the development of interventions to prevent KS and target groups for earlier ART initiation.

#### Methods

#### **Overall Design**

We conducted a case-control study of untreated HIV-infected adults ( 18 years old) in Uganda to investigate the relationship between plasma KT ratio and KS independent of known confounders.

#### **Study Population**

Cases were individuals with KS sampled between April 2007 and February 2011 throughout Uganda, who were enrolled in the Antiretrovirals for Kaposi's Sarcoma (ARKS) study based at the Infectious Diseases Institute, Kampala, Uganda [17]. To be eligible for ARKS, subjects had to have histologically-confirmed KS, with the exception of a few patients with highly characteristic oral lesions that could not be easily biopsied. No subjects had what was considered advanced KS by virtue of having an urgent indication for chemotherapy. Specifically, no individuals had any functionally disabling complication of KS. Subjects also could not have evidence of current active untreated opportunistic infection (e.g., tuberculosis or cryptococcosis) or other malignancy. While ARKS was a randomized trial of different ART regimens for treatment of KS, the patients in the current case-control study were studied at baseline prior to initiating ART. All ARKS enrollees with available plasma specimens were included as cases in the present analysis.

Controls were individuals without KS enrolled in the Uganda AIDS Rural Treatment Outcomes (UARTO) Cohort. UARTO is a consecutive sample of ambulatory HIV-infected adults residing within 60 kilometers of and starting ART at the Immune Suppression Syndrome (ISS) Clinic at Mbarara Regional Referral Hospital in southwestern Uganda. ART at this clinic is prescribed on the basis of CD4+ T cell count or clinical conditions, according to Ugandan national antiretroviral treatment guidelines [18]. For the present study, we included all UARTO participants without KS who were consecutively enrolled between June 2005 and April 2010 and who had available plasma specimens, and we studied them prior to ART initiation. The absence of KS in the controls was based on patient self-report as well as review of medical records at the ISS Clinic. The ISS Clinic has an active skin punch biopsy service for histological confirmation of KS sponsored by the International Epidemiologic Databases to Evaluate AIDS (IeDEA) Consortium.

Amongst both cases and controls, we excluded any woman with a history of pregnancy in the prior 12 months given the influence of pregnancy on the kynurenine pathway [19] and the fact that pregnant women were excluded from the ARKS trial.

#### Measurements

**Questionnaire-based**—The ARKS and UARTO studies used the same instruments for all measurements of socio-demographic characteristics, medical history, and clinical assessment. Within these instruments, we used the Medical Outcomes Study to determine physical health status and mental health status [20] and the Filmer-Pritchett asset index, as previously described [21], to characterize socioeconomic status.

**Laboratory**—All biological tests for the cases and controls were performed in the same laboratories on samples obtained prior to the start of ART. Tryptophan and kynurenine levels were measured on cryopreserved plasma samples by liquid chromatography tandem mass spectrometry as previously described [22]. Plasma HIV RNA level was measured using the Amplicor HIV Monitor version 1.5 or the Cobas Taqman HIV-1 version 1.0 assays (Roche, Branchburg, NJ). CD4+ T cell counts were assessed using the FACSCalibur system (Becton Dickinson, San Jose, CA). Two enzyme immunoassays [23, 24] and one direct immunofluorescence assay [25, 26] were used to detect antibodies to KSHV, and the results of these three assays were interpreted according to a previously described algorithm [27].

#### **Statistical Analysis**

Multivariable logistic regression was used to examine the relationship between plasma KT ratio and KS. In our base-case analysis, we were guided by our pre-specified perception of the biological system and hence adjusted for CD4+ T cell count, plasma HIV RNA, age and sex [3, 6, 28, 29]. Because of ample sample size and number of cases, we did not attempt to reduce the number of covariates. In additional analyses, we also adjusted for other proxies of HIV disease progression not captured by the aforementioned variables, including body mass index, hemoglobin, mental health status, physical health status and socioeconomic status [30-32]. In sensitivity analyses, we first restricted KS cases to those recruited from areas with moderate to high malaria endemicity, and then we also separately restricted the non-KS controls to those without suspicion of TB within the first 6 months after initiation of ART. Finally, we also performed an analysis in which we restricted the non-KS control group to subjects who were KSHV-antibody-positive.

We assessed for non-linearity in continuous predictor variables using splines, and variables were transformed or categorized where necessary. Influential observations were checked using post-estimation plots of dfbetas, followed by sensitivity analyses on data points with high dfbetas. We also assessed for possible interactions with the primary predictor variable, KT ratio, and used a p value of 0.05 to guide reporting of interaction. The Hosmer-Lemeshow test was used to examine goodness of fit. We used multiple imputation with iterative chained equations for missing values [33]. All analyses were performed using STATA 13 (College Station, TX, USA).

#### Results

#### **Characteristics of the Study Population**

A total of 631 untreated HIV-infected Ugandans were examined: 222 KS cases and 409 non-KS controls. Men comprised 56% of the cases and 33% of the controls (Table 1). Compared to the controls, cases had a slightly higher median body mass index (21.4 versus 21.1kg/m²), lower hemoglobin (11.6 versus 12.2 g/dl), and higher plasma HIV RNA levels (5.3 versus 5.1 copies/ml). Sixty nine percent and 75% of cases and controls, respectively, had a CD4+T cell count 200 cells/µl; 13% and 3% of cases and controls respectively, had CD4+T cell count >350 cells/µl. The cases had a wide spectrum of mucocutaneous KS ranging from oral lesions only to widespread cutaneous dissemination.

#### Relationship between Plasma KT Ratio and KS

The median plasma KT ratio amongst all subjects was 120 (IQR: 90 to 180) nM/µM, which is higher than in HIV-uninfected individuals, at least in resource-rich settings [34]. The distribution of plasma KT ratio was more markedly skewed to the right in the controls compared to the cases (Figure 1). In an unadjusted analysis, the non-KS controls had a higher median plasma KT ratio (130, interquartile range [IQR]: 90 to 190 nM/µM) than cases (110, IQR: 90 to 150 nM/ $\mu$ M) (p = 0.004). Specifically, there was a non-linear "threshold" relationship between plasma KT ratio and KS (Table 2). Compared to those with KT ratio in the lowest quartile (i.e., lowest values, KT ratio <90 nM/μM), there was no association with KS with increasing values of KT ratios until the highest values were reached. Those with KT ratio in the highest quartile (i.e., highest values, KT ratio >179 nM/μM) had a 50% reduction in the odds of KS. We observed similar results in a multivariable logistic regression model adjusting for age, sex, CD4+ T cell count and plasma HIV RNA level (Table 2). Compared to individuals with KT ratio in the lowest quartile, there was 59% reduction (95% CI: 27% to 77%; p = 0.002) in the odds of KS for those with KT ratio in the highest quartile. In an additional analysis, to attempt further adjust for the confounding effect of HIV disease progression, we also adjusted for body mass index, hemoglobin, mental health status, physical health status and asset index. In this analysis, we observed an even a stronger and more linear association between plasma KT ratio and KS. The association between KT ratio and KS was not modified by CD4+ T cell count, plasma HIV RNA level, sex or age (p value for interaction = 0.61, 0.84, 0.54, and 0.19 respectively).

#### Effects of Malaria and Tuberculosis on the Relationship between KT Ratio and KS

Given that Plasmodium species may induce IDO activity [35], and that all the non-KS controls were enrolled from areas with moderate to high malaria endemicity while 31% of the cases were enrolled from areas of low malaria endemicity [36], we examined whether the association between KT ratio and KS might be because of differences in malaria comorbidity between the cases and controls. In analyses restricted to the 153 KS cases recruited from areas with moderate to high malaria endemicity [36], we again observed, after adjustment for age, sex, CD4+ T cell count and plasma HIV RNA viral load, that individuals with KT ratio in the highest quartile had a reduction in the odds of KS compared to those in the lowest quartile (OR = 0.39; 95% CI: 0.20 to 0.77; Table 3). When we also adjusted for BMI, hemoglobin, mental health status, physical health status and asset index, a dose-response relationship was again observed. Imbalance in concurrent tuberculosis was also a concern because Uganda has a high prevalence of tuberculosis amongst patients initiating ART [37], Mycobacterium tuberculosis may induce IDO [38], and the ARKS study excluded persons with untreated tuberculosis while UARTO did not. To address this, we restricted our analyses to the non-KS controls without TB by excluding all subjects who were receiving any anti-tuberculosis medications at enrollment, or who were subsequently treated for tuberculosis within the first 6 months after ART initiation (n=62). Our findings of an independent inverse association between plasma KT ratio and KS remained consistent (Table 3).

#### Effect of KSHV Infection on the Relationship between KT Ratio and KS

Consistent with prior data from this region, 40% of the 409 subjects in the non-KS control group were KSHV-antibody-positive. In an additional analysis in which we restricted the non-KS control group to only participants who were KSHV-antibody-positive and, again, adjusted for age, sex, plasma HIV RNA level, and CD4 count, we observed a somewhat stronger negative association between KT ratio and KS than in our analysis including all 409 non-KS controls. Compared to individuals with KT ratio in the lowest quartile, there was 71% reduction (95% CI: 42% to 86%; p=0.002) in the odds of KS for those with KT ratio in the highest quartile.

#### Other Determinants of KS

We observed that KS was also independently associated with male sex, higher BMI, lower hemoglobin, lower mental health status score, lower CD4+ T cell count, and higher plasma HIV RNA level (Table 2). The correlation with plasma HIV RNA was most notable with a dose-response relationship seen throughout most of the range of exposure and over a 5-fold greater odds of KS observed amongst those with RNA > 100,000 copies/ml compared to those with < 10,000 copies/ml.

#### Discussion

Other than KSHV infection and CD4+ T cell lymphopenia, the specific biological mechanisms responsible for the development of AIDS associated KS remain poorly understood. In this case-control study of untreated HIV-infected Ugandans, we have described a potentially new mechanism affecting the occurrence of KS. Specifically, greater activity of the kynurenine pathway of tryptophan catabolism was associated with a lower occurrence of KS. These data provide the first evidence suggesting a role of the kynurenine pathway of tryptophan catabolism in KS pathogenesis and may explain why some KSHV-infected individuals with advanced AIDS fail to develop KS. We have also added to the knowledge base for how HIV infection causes KS by finding a strong independent association between plasma HIV RNA level and KS.

Our findings add to the growing literature regarding the kynurenine pathway of tryptophan catabolism in carcinogenesis [16]. In the majority of work to date, increased kynurenine pathway activity has been positively associated with the occurrence of cancer, which is generally attributed to the escape of tumor from immune surveillance [16]. IDO is expressed in both tumor cells and antigen-presenting cells in tumor-draining lymph nodes, and by fostering immune suppression, IDO activity facilitates the survival and growth of tumor cells expressing unique antigens that would normally be recognized as foreign [39, 40]. Indeed, increased serum KT ratio has been shown to correlate with disease progression and poor prognosis in certain cancers [41, 42]. In contrast, in our study, higher plasma KT ratio (reflecting higher levels of IDO activity) appeared protective against KS. Our results suggest that intact lymphocyte proliferation is necessary for the development of KS. The *in vitro* observation that malignant B lymphocyte cell growth is suppressed by IDO activity also supports this hypothesis [43]. A protective effect of the highest levels of IDO activity against KS is also compatible with the observation that KS often occurs at local sites of

inflammation [44] and the inflammatory nature histologically of KS lesions (i.e., an immune cell infiltrate). In addition, several inflammatory cytokines are increased in KS lesions and are capable of increasing lytic replication of KSHV [45]. Th1 cytokines from activated infiltrating T and B cells may also trigger KS lesion formation by promoting spindle cell proliferation and angiogenesis [45, 46]. That an inflammatory microenvironment fosters KS lesions suggests that IDO-derived local immune suppression could impair the development of KS. Alternatively, high levels of KSHV infection might suppress interferon-gamma-induced IDO and lower KT levels as previously observed in other viral infections [47]. Experiments on KS tissue, and appropriate controls, to directly examine KT levels would be a natural next step to evaluate the biological plausibility of our finding.

We observed a stronger inverse association between plasma KT ratio and KS — in fact, a dose-response effect — when we also adjusted for other markers of HIV disease progression, namely BMI, mental health score, physical health score, asset index, and hemoglobin. Further interrogation of the regression models revealed that this change in the shape of the KT-KS relationship was caused solely by adjustment for hemoglobin (data not shown). We cannot, however, determine whether this further adjustment is warranted and hence whether the stronger association is valid. Specifically, because this is a cross-sectional study, we cannot distinguish between whether low hemoglobin is a common outcome of both KT ratio and KS (also known as a "collider" in epidemiologic parlance), or whether it is a proxy for a confounder (causally responsible for both KT ratio and KS), or a mediator (intermediary along the pathway) for the relationship between KT ratio and KS, and hence we presented the results of both adjusted models 1 and 2. While the latter two scenarios warrant adjustment for hemoglobin when trying to understand the direct causal effect of KT ratio on KS, the first scenario does not. In any case, whether or not hemoglobin is adjusted for, we consistently observed that having the highest level plasma KT ratio was associated with lower occurrence of KS.

As is often true in case-control studies, the principal limitation of our work concerns the selection of the controls. Because of the relative rarity of KS and the lack of a populationbased surveillance system in Uganda for the identification of KS as it occurs in the community, we were unable to assemble unassailable case and control groups sampled from a primary study base [48]. Instead, we took advantage of a large group of recently diagnosed cases of KS, compared them to a representative sample of persons without KS who were similarly advanced in their HIV infection by virtue of an indication for ART, and accommodated for any differences between the groups by statistical adjustment. Importantly, our case and control groups were not independently constructed and compared in retrospect with a piecemeal assemblage of measurements. Rather, the ARKS and UARTO studies were prospectively performed in parallel with identical questionnaire-based and laboratory measurements in anticipation of comparative research like the present study. While it remains possible that the UARTO-based control group was systematically enriched for patients with higher values of KT ratio, we feel our various analyses render this unlikely. First, the most obvious difference between the groups, their geographic residence, was addressed by restricting analysis in the case group to those living in malarious areas. In this sensitivity analysis, our primary inference was unchanged. Second, the potential for the control group to harbor patients with a greater prevalence of co-infections which are

stimulating IDO induction was addressed in our main analysis by extensive adjustment for determinants of co-infections (e.g., CD4+ T cell count) as well as manifestations of co-infections (e.g., BMI). Furthermore, when we limited the control group to those without tuberculosis, we again saw no change in our inference. Although we were unable to directly examine the effect of potential differences in prevalence of soil transmitted helminthes, the species that may induce IDO [49, 50] are more prevalent in the geographic areas from which the cases were recruited [51]. Therefore, we would expect a bias towards the null. Nonetheless, as with any observational study, our findings require replication.

Consistent with prior research from resource-rich settings [29, 52], we found in untreated HIV-infected Africans that higher plasma HIV RNA levels were strongly associated with KS even after adjustment for CD4+ T cell count and other factors. The independent role of plasma HIV RNA provides clinical augmentation of earlier laboratory research suggesting a causative role of HIV per se in KS etiology [53]. CD4+ T cell lymphopenia was also independently associated with KS, as has been previously well established. Of note, that a CD4+ T cell count > 350 cells/µl appeared to carry a greater risk of KS than a CD4+ T cell count < 50/µl is likely an artifact of the patient sample and not a biologic phenomenon. That is, the non-KS controls were HIV-infected patients about to initiate ART according to local ART guidelines, which, for much of the period of the study, required a CD4+ T cell count < 200 cells/µl or a World Health Organization stage IV condition [18]. Therefore, the majority of the controls had a CD4+ T cell count < 200 cells/µl. On the other hand, there was no restriction on the CD4+ T cell count for the KS cases, and, as seen elsewhere [3, 54], a relevant proportion of KS cases had a CD4+ T cell count > 350 cells/µl. This therefore explains why the KS cases are enriched, in comparison with the controls, for high CD4+ T cell counts. This difference in CD4+ T cell count distribution between cases and controls, however, does not explain the association between KT ratio and KS because CD4+ T cell count was statistically controlled for when assessing the KT ratio and KS relationship, and, furthermore, there was no statistical interaction between KT ratio and CD4+ T cell count. We also observed that lower hemoglobin was associated with KS which we speculate reflects anemia of chronic inflammation driven by elevated levels of inflammatory cytokines [55, 56] and/or KS involvement of the gastrointestinal tract with occult hemorrhage [57]. Similarly, we believe that the association between BMI and KS is also an example of reverse causality, explained by KS-induced lymphedema and associated weight gain. Like prior studies in this setting, we observed that male sex was independently associated with KS [28].

We note that the explosion of human subjects-based biological discovery in HIV/AIDS (sometimes called "translational" research) has not occurred in the study of KS [58]. The absence of translational research in KS is particularly regrettable in Africa where KS is among the most common cancers in the entire general population. The differences between Africa and settings like the U.S. in terms of human host, causative viral pathogen, and environment suggest that African KS science cannot simply survive by extrapolating from the translational work done in resource-rich settings. Now that biological measurement tools and research infrastructure such as ARKS, UARTO, and the AIDS Malignancy Consortium

are in place in sub-Saharan Africa, we hope that studies like ours will spur further Africanbased translational work in KS etiopathogenesis.

In summary, we have demonstrated that higher IDO activity, as evidenced by higher plasma KT ratio, is associated with lower occurrence of KS amongst HIV-infected adults in Africa. The inverse relationship between KT ratio and KS is consistent with the inflammatory nature of KS lesions and suggests that intact lymphocyte proliferation is required for KS lesion development. Results from this study imply that some consequences of immune activation in HIV might actually suppress or mask certain cancers.

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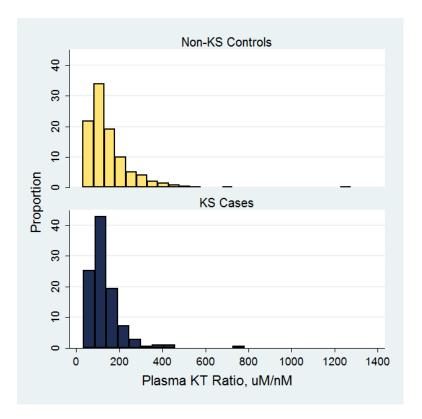
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**Figure 1.** Distribution of plasma KT ratio in KS cases and non-KS controls

Table 1
Characteristics of the cases and controls

Characteristic	KS Cases (n=222)	Non-KS Controls (n=409)
Age, years	34 (28 to 40)*	35 (29 to 40)
Male sex	56%	33%
Physical health status $^{\dagger}$	53.3 (36.4 to 58.2)	54.0 (45.0 to 58.6)
Mental health status $\dot{\tau}$	48.6 (37.7 to 56.7)	52.6 (45.9 to 58.7)
Asset index <sup>‡</sup>	0.18 (-1.80 to 1.78)	-0.58 (-1.82 to 0.86)
Body mass index, kg/m <sup>2</sup>	21.4 (19.4 to 23.1)	21.1 (19.4 to 23.4)
Hemoglobin, g/dl	11.6 (10.3 to 13.2)	12.2 (10.6 to 13.7)
HIV RNA, $\log_{10}$ plasma copies/ml	5.3 (5.0 to 5.6)	5.1 (4.6 to 5.6)
CD4+ T cells, count/μl		
< 50	35%	16%
51-100	11%	18%
101-200	23%	41%
201-350	18%	21%
> 350	13%	3%
Plasma KT Ratio, nM/uM, by quartile		
quartile 1 (34-89)	27%	24%
quartile 2 (90-120)	28%	23%
quartile 3 (121-179)	29%	23%
quartile 4 (180-1369)	16%	30%

<sup>\*</sup> Median (interquartile range) unless indicated

<sup>†</sup>Mental and Physical Health Status scores were derived using the Medical Outcomes Study-HIV survey. Responses to questions on 5 scales regarding vitality, cognitive function, quality of life, health distress and mental health are summarized in the Mental Health Status score, and responses to questions on 6 scales regarding general health, vitality, pain, physical, role and social function are summarized in the Physical Health Status score. Responses to individual questions are aggregated, and scores are converted to a 0-100 point scale, with 100 representing the best mental or physical health status.<sup>20</sup>

The Filmer Pritchet index is a measure of socioeconomic status based on self-reported asset ownership and housing characteristics. Appropriate weights for individual questions are determined using the statistical method of principal components, and responses are aggregated to create the asset index score, which may be positive or negative with a median of zero. A higher score represents a higher socioeconomic status.

Unadjusted and adjusted logistic regression evaluating factors associated with Kaposi's sarcoma

ars Odds Ratio (95% CT)   P value   Odds Ratio (95% CT)   P value    ars 0.08 (0.97-1.01)   0.08   0.79 (0.62-1.01)   0.059    Bef. 2.69 (1.92-3.76)   <0.001   2.42 (1.63-3.58)   <0.001    b status† 0.98 (0.97-0.99)   <0.001      1.06 (1.01-1.14)   0.044      1.06 (1.01-1.14)   0.044      1.06 (1.01-1.14)   0.064      1.06 (1.01-1.14)   0.064      1.07 (1.01-1.14)   0.064      1.08 (0.97-0.99)   0.001      1.09 (0.85-0.96)   0.001      1.00 (0.85-0.96)   0.001      1.00 (0.85-0.96)   0.001      2.01 (1.14-7.43)   0.02   0.55 (1.80-17.1)   0.001    2.01 (1.14-7.43)   0.02   0.30 (0.16-0.55)   0.001    2.01 (1.14-7.43)   0.001   0.30 (0.16-0.55)   0.001    2.01 (0.023 (0.15-0.37)   0.001   0.34 (0.16-0.55)   0.001    2.01 (0.024 (0.17-0.34)   0.002   0.33 (0.116-0.55)   0.001    2.01 (0.024 (0.17-0.34)   0.007   0.34 (0.31-0.94)   0.002    2.01 (0.024 (0.17-0.34)   0.007   0.34 (0.31-0.94)   0.002    2.01 (0.024 (0.17-0.34)   0.007   0.34 (0.31-0.94)   0.001    2.01 (0.024 (0.17-0.34)   0.007   0.34 (0.31-0.94)   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.01   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001    2.01 (0.007-1.158)   0.001   0.001    2.01 (0.007-1.1		Unadjusted		Adjusted Model 1		Adjusted Model 2	2
Ref.	Characteristic	Odds Ratio (95% CI)	P value	* Odds Ratio (95% CI)	P value	* Odds Ratio (95% CI)	P value
Ref.  2.69 (1.92-3.76) < 0.0011	Age, per 10 years	0.98 (0.97-1.01)	0.08	0.79 (0.62-1.01)	0.059	0.84 (0.64-1.09)	0.19
Ref.       Ref.         2.69 (1.92-3.76)       -0.001       2.42 (1.63-3.58)       <0.001	Sex						
2.69 (1.92-3.76)	Women	Ref.		Ref.		Ref.	
status† 0.98 (0.97-0.99)	Men	2.69 (1.92-3.76)	<0.001	2.42 (1.63-3.58)	<0.001	4.50 (2.60-7.79)	<0.001
status† 0.96 (0.95-0.98) < 0.001	Physical health status $^{\dagger}$	0.98 (0.97-0.99)	<0.001	1		1.00 (0.98-1.03)	0.83
1.06 (1.01-1.14)   0.044       sma copies/ml   Ref.   Ref.     0.90 (0.85-0.96)   0.001       1.95 (0.73-5.23)   0.18   2.61 (0.84-8.08)   0.006     0.00	Mental health status $^{\dagger}$	0.96 (0.95-0.98)	<0.001	I		0.97 (0.95-1.00)	0.041
lex, kg/m2\$  1.01 (0.97-1.04)  9.06  9.07  9.08-0.96  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.85-0.96)  9.09 (0.84-8.08)  9.09 (0.84-8	Asset index <sup>‡</sup>	1.06 (1.01-1.14)	0.044	I		1.04 (0.95-1.14)	0.42
ml  Ref.  1.95 (0.73-5.23) 0.18 2.61 (0.84-8.08) 0.096 3.05 (1.18-7.91) 0.021 4.07 (1.34-12.3) 0.013 7.37 (3.01-18.1) 0.026 5.55 (1.80-17.1) 0.003  Ref.  0.23 (0.15-0.37) 0.001 0.30 (0.16-0.55) 0.001 0.24 (0.17-0.34) 0.001 0.33 (0.21-0.54) 0.029 1.50 (0.96-2.35) 0.075 2.60 (1.15-5.87) 0.021  Liby quartile  Ref.	Body mass index, kg/m <sup>2</sup> §	1.01 (0.97-1.04)	99.0	I		1.11 (1.04-1.18)	0.002
Ref.         Ref.       Ref.         1.95 (0.73-5.23)       0.18       2.61 (0.84-8.08)       0.096         3.05 (1.18-7.91)       0.021       4.07 (1.34-12.3)       0.013         7.37 (3.01-18.1)       0.026       5.55 (1.80-17.1)       0.001         Lyµl       Ref.       Ref.       Ref.         0.23 (0.15-0.37)       <0.001	Hemoglobin, g/dl <sup>§</sup>	0.90 (0.85-0.96)	0.001	ı		0.73 (0.60-0.89)	0.002
Ref.       Ref.         (0.73-5.23)       0.18       2.61 (0.84-8.08)       0.096         (1.18-7.91)       0.021       4.07 (1.34-12.3)       0.013         (3.01-18.1)       <0.001	HIV RNA, plasma copies/ml						
(0.73-5.23)     0.18     2.61 (0.84-8.08)     0.096       (1.18-7.91)     0.021     4.07 (1.34-12.3)     0.013       (3.01-18.1)     <0.001	10,000	Ref.		Ref.		Ref.	
(J.18-7.91) 0.021 4.07 (J.34-12.3) 0.013 (3.01-18.1) <0.0001 9.64 (3.35-27.7) <0.001 (J.14-7.43) 0.026 5.55 (J.80-17.1) 0.003  Ref. (0.15-0.37) <0.001 0.30 (0.16-0.55) <0.001 (0.17-0.34) <0.001 0.33 (0.21-0.54) 0.029 (0.96-2.35) 0.075 2.60 (J.15-5.87) 0.021  Ref. (0.77-1.58) 0.61 1.02 (0.61-1.72) 0.93 (0.77-1.58) 0.58 0.86 (0.51-1.45) 0.58	10,001-50,000	1.95 (0.73-5.23)	0.18	2.61 (0.84-8.08)	960.0	2.43 (0.75-7.92)	0.14
(3.01-18.1)       <0.0001	50,001-100,000	3.05 (1.18-7.91)	0.021	4.07 (1.34-12.3)	0.013	4.05 (1.27-12.8)	0.018
Ref.         Ref.         Ref.           (0.15-0.37)         <0.001	100,001-500,000	7.37 (3.01-18.1)	<0.001	9.64 (3.35-27.7)	<0.001	9.26 (3.09-27.8)	<0.001
Ref.       Ref.         (0.15-0.37)       <0.001	> 500,000	2.91 (1.14-7.43)	0.026	5.55 (1.80-17.1)	0.003	5.17 (1.60-16.8)	0.006
Ref.       Ref.         (0.15-0.37)       <0.001	CD4+ T cells, count/µl						
(0.15-0.37)     <0.001	< 50	Ref.		Ref.		Ref.	
(0.17-0.34)     <0.001	51-100	0.23 (0.15-0.37)	<0.001	0.30 (0.16-0.55)	<0.001	0.25 (0.13-0.49)	<0.001
(0.23-0.50) <0.001 0.54 (0.31-0.94) 0.029 (0.96-2.35) 0.075 2.60 (1.15-5.87) 0.021 Ref. Ref. (0.77-1.58) 0.61 1.02 (0.61-1.72) 0.93 (0.77-1.58) 0.58 0.86 (0.51-1.45) 0.58	101-200	0.24 (0.17-0.34)	<0.001	0.33 (0.21-0.54)	<0.001	0.34 (0.20-0.57)	<0.001
(0.96-2.35) 0.075 2.60 (1.15-5.87) 0.021  Ref. (0.77-1.58) 0.61 1.02 (0.61-1.72) 0.93 (0.77-1.58) 0.58 0.86 (0.51-1.45) 0.58	201-350	0.34 (0.23-0.50)	<0.001	0.54 (0.31-0.94)	0.029	0.52 (0.28-0.95)	0.033
Ref. (0.77-1.58) 0.61 1.02 (0.61-1.72) 0.93 (0.77-1.58) 0.58 0.86 (0.51-1.45) 0.58	> 350	1.50 (0.96-2.35)	0.075	2.60 (1.15-5.87)	0.021	2.55 (1.08-7.04)	0.033
Ref. 1.10 (0.77-1.58) 0.61 1.02 (0.61-1.72) 0.93 1.11 (0.77-1.58) 0.58 0.86 (0.51-1.45) 0.58	Plasma KT Ratio, nM/uM, b	y quartile					
1.10 (0.77-1.58)     0.61     1.02 (0.61-1.72)     0.93       1.11 (0.77-1.58)     0.58     0.86 (0.51-1.45)     0.58	quartile 1 (34-89)	Ref.		Ref.		Ref.	
1.11 (0.77-1.58) 0.58 0.86 (0.51-1.45) 0.58	quartile 2 (90-120)	1.10 (0.77-1.58)	0.61	1.02 (0.61-1.72)	0.93	0.74 (0.41-1.36)	0.34
	quartile 3 (121-179)	1.11 (0.77-1.58)	0.58	0.86 (0.51-1.45)	0.58	0.51 (0.26-1.00)	0.049

	Unadjusted		Adjusted Model 1	1	Adjusted Model 2	2
Characteristic	Odds Ratio (95% CI) P value	P value	Odds Ratio * (95% CI)	P value	CI) P value Odds Ratio (95% CI) P	P value
quartile 4 (180-1369)	0.50 (0.33-0.76)	0.001	0.41 (0.23-0.73)	0.002	0.21 (0.09-0.46)	<0.001

\* all variables adjusted for all other variables in column

 $\vec{\tau}$  per 1 unit increase in score derived from Medical Outcomes Study-HIV survey  $^{20}$ 

 $^{\sharp}_{\mathrm{per}\,1}$  unit increase in Filmer-Pritchett index $^{21}$ 

§ per 1 unit increase in respective native scale

Fable 3

Unadjusted and adjusted logistic regression evaluating factors associated with Kaposi's sarcoma

			Adjusted Model 1	1	Adjusted Model 2'	2,
Characteristic	Odds Ratio (95% CI) P value	P value	Odds Ratio (95% CI) P value	P value	Odds Ratio (95% CI) P value	P value
Restricted to subjects from moderate to high malaria endemic areas	n moderate to high malar	ria endemi	areas			
Plasma KT ratio, nM/uM, by quartile	'uM, by quartile					
quartile 1 (34-89)	Ref		Ref		Ref	
quartile 2 (90-120)	1.12 (0.70-1.77)	0.64	1.14 (0.63-2.06)	0.67	0.72 (0.36-1.43)	0.34
quartile 3 (121-179)	1.09 (0.68-1.74)	0.73	1.02 (0.57-1.83)	96.0	0.51 (0.24-1.07)	0.073
quartile 4 (180-1369)	0.48 (0.28-0.81)	0.006	0.39 (0.20-0.77)	0.006	0.15 (0.06-0.38)	<0.001
Restricted to subjects with no suspicion of TB	no suspicion of TB					
Plasma KT ratio, nM/uM, by quartile	'uM, by quartile					
quartile 1 (34-89)	Ref		Ref		Ref	
quartile 2 (90-120)	1.22 (0.73-2.03)	0.45	1.08 (0.64-1.85)	0.77	0.75 (0.40-1.40)	0.37
quartile 3 (121-179)	1.26 (0.75-2.09)	0.38	0.86 (0.50-1.49)	0.59	0.50 (0.25-0.99)	0.046
quartile 4 (180-1369)	0.43 (0.24-0.78)	0.006	0.43 (0.23-0.79)	0.007	0.22 (0.10-0.50)	<0.001

 $^{\ast}$  adjusting for plasma HIV RNA, CD4+ T cell count, age and sex

† djusting for plasma HIV RNA, CD4+ T cell count, age, sex, BMI, hemoglobin, physical health status, mental health status and asset index