The clinical presentation of culture-positive and culture-negative, qPCR-attributable shigellosis in the

Global Enteric Multicenter Study and derivation of a Shigella severity score: implications

for pediatric Shigella vaccine trials

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# SUMMARY

Molecular methods identified *Shigella* more commonly than microbiologic culture in younger and stunted children. A simplified clinical score containing dehydration, hospitalization, and diarrhea duration could be used to stratify vaccine trial endpoints by severity based on its ability to predict death.

#### ABSTRACT

**Background:** *Shigella* is a leading cause of childhood diarrhea and target for vaccine development. Microbiologic and clinical case definitions are needed for pediatric field vaccine efficacy trials.

**Methods:** We compared characteristics of moderate to severe diarrhea (MSD) cases in the Global Enteric Multicenter Study (GEMS) between children with culture positive *Shigella* to those with culture-negative, qPCR-attributable *Shigella* (defined by an *ipaH* gene cycle threshold <27.9). Among *Shigella* MSD cases, we determined risk factors for death and derived a clinical severity score.

**Results:** Compared to culture-positive *Shigella* MSD cases (n=745), culture-negative/qPCRattributable *Shigella* cases (n=852) were more likely to be under 12 months, stunted, have a longer duration of diarrhea, and less likely to have high stool frequency or a fever. There was no difference in dehydration, hospitalization, or severe classification from a modified Vesikari score. Twenty-two (1.8%) *Shigella* MSD cases died within the 14-days after presentation to health facilities, and 59.1% of these deaths were in culture-negative cases. Age < 12 months, diarrhea duration prior to presentation, vomiting, stunting, wasting, and hospitalization were associated with mortality. A model-derived score assigned points for dehydration, hospital admission, and longer diarrhea duration but was not significantly better at predicting 14-day mortality than a modified Vesikari score.

**Conclusions**: A composite severity score consistent with severe disease or dysentery may be a pragmatic clinical endpoint for severe shigellosis in vaccine trials. Reliance on culture for microbiologic confirmation may miss a substantial number of *Shigella* cases but is currently required to measure serotype specific immunity.

#### MAIN BODY

#### Introduction

*Shigella* is a leading cause of diarrhea among children younger than 5 years in resource-limited settings, associated with over 60,000 deaths in this age group per year.[1] Several *Shigella* vaccines are currently in development[2-4] and vaccine efficacy will be determined by the number of clinically relevant, *Shigella*-attributable diarrhea cases prevented by the vaccine.[5]

Microbiological culture is the gold standard for *Shigella*-confirmation, however the application of highly sensitive molecular tools, such as quantitative PCR (qPCR), has revealed a large burden of *Shigella* infections undetected by culture.[6, 7] This increased sensitivity of qPCR could make vaccine trials more efficient, but the clinical significance of culture-negative and PCR-attributable shigellosis is unclear.

Rotavirus vaccine is most efficacious in preventing severe rotavirus diarrhea[8] and the Vesikari score is commonly used to stratify clinical endpoints in vaccine trials.[9] There is no universally accepted clinical severity score for *Shigella* diarrhea in children, and the Vesikari score does not include severity indicators that may be specific to shigellosis, such as dysentery.[5] Identifying the ideal severity score to use in *Shigella* vaccine trials requires an empiric assessment of the performance of existing and new severity scores in disaggregating severe vs. non-severe cases of *Shigella* diarrhea.

The Global Enteric Multicenter Study (GEMS) was a multi-country case-control study which enrolled children seeking care for moderate-to-severe (MSD) diarrhea and matched controls.[10] Utilizing clinical and laboratory data from GEMS cases, we sought to inform a microbiologic and clinical case definition for severe, laboratory-confirmed *Shigella* diarrhea by answering three questions: 1) Does the clinical presentation of *Shigella* differ by culture vs. qPCR? 2) What are risk factors for death among children

### Methods

#### Parent Study

Children aged 0-59 months presenting to health centers in Bangladesh, India, Kenya, Mali, Mozambique, Pakistan and The Gambia with diarrhea were screened for eligibility into GEMS between 2007 and 2011 as described elsewhere.[10] Eligible children were those with an acute episode of MSD defined as one or more of the following: dehydration (sunken eyes, loss of skin turgor, intravenous rehydration recommended), dysentery, or hospital admission.[11] At enrollment, clinical history and sociodemographic information were ascertained using a standardized questionnaire, whole stool samples were collected, and a physical exam was performed. Length/height, weight, and mid-upper arm circumference (MUAC) were measured at enrollment and at a single follow-up visit that was performed 60 days later (acceptable range 50-90 days) at which time vital status was also ascertained.

Stool samples were originally processed using conventional enteric pathogen detection methods as described elsewhere[12, 13] and a portion of samples stored at -80°C. For *Shigella* diagnosis by bacterial culture, stool samples were transported in cold storage in buffered glycerol saline (BGS) transport media and were inoculated onto MacConkey and xylose lysine desoxycholate agar. Suspected *Shigella* colonies were confirmed using triple-sugar iron, motility indole ornithine (MIO lysine decarboxylase media), citrate and urea biochemical typing media. A random subset of stored stool samples and samples from all fatal cases (if not included in the random subset) were also tested by qPCR using a 32-enteropathogen TaqMan Array Card.[7] Cycle thresholds ( $C_t$ ) required to detect the pathogen gene target, which are inversely related to nucleic acid quantity, of 35 or greater were deemed negative. The gene amplified for

#### Nested Study

In this secondary data analysis, we excluded controls as well as cases who did not have both culture and qPCR results available then grouped MSD cases by Shigella culture results. Among culture-negative children, we further divided low and high quantity infections using the *ipaH*  $C_t$  cut-off (<27.9) associated with an odds ratio of 2.0 in the original qPCR GEMS analysis.[7] This grouping led to four mutually exclusive categories: 1) Shigella negative (no detection by culture or qPCR); 2) culture-negative/qPCRunattributable (culture negative and  $27.9 \le ipaH C_t < 35$ ), 3) culture-negative/qPCR-attributable (culture negative and *ipaH* Ct <27.9); and 4) culture-positive (culture-positive Shigella irrespective of qPCR value). The clinical and demographic characteristics of these four categories were compared using prevalence ratios determined from Poisson regression with culture-positive Shigella as the reference group and each dichotomous covariate of interest modelled separately in a model including site (indicator variable) and age (continuous variable). Characteristics of interest, ascertained at MSD presentation, included age, site, sex, dysentery (visibly bloody stool reported by caregiver, clinician, or laboratory technician), mucoid stool, caregiver-reported number of days of diarrhea prior to presentation, caregiver reported number of loose stools in previous 24-hours, axillary temperature, caregiver-reported vomiting, clinician-determined dehydration status (according to World Health Organization [WHO] IMCI guidelines[15]), stunting (length for age z-score [LAZ] <-2), wasting (mid-upper arm circumference [MUAC] <12.5cm among children 6 months or older) and admission to hospital. We utilized clinical signs at presentation to recreate the MVS generated previously with this data. [16] This MVS totaled 16 points and was categorized as mild (1-5 points), moderate (6-8 points), and severe (9-16 points). To establish the likelihood of causes of diarrhea other than *Shigella* in each of the four diagnostic categories, we considered site-and age-adjusted attributable fractions  $\geq 0.5$ , derived from qPCR C<sub>1</sub>-values.[17] These

pathogens were grouped as: viral (astrovirus, norovirus, rotavirus, sapovirus, adenovirus), parasitic (*Cryptosporidium, Entamoeba histolytica, Cyclospora, Isospora*), and other bacterial (*Campylobacter, Helicobacter pylori, Salmonella, Vibrio cholerae*, EAEC, ST-ETEC, LT-ETEC, tEPEC, STEC). The prevalence of other causes was compared between the four *Shigella* categories in age- and site-adjusted Poisson regression models.

To establish risk factors for death among children with *Shigella*-attributed diarrhea, we excluded children who were *Shigella* negative (by culture and qPCR) or had culture-negative/qPCR-unattributable *Shigella* and children without a 60-day follow-up visit in which vital status (and date of death, if applicable) was ascertained. Cox proportional hazards regression was used to identify univariate and adjusted risk factors for death in the first 14-days after presentation. We evaluated deaths in the 14-days to capture deaths most likely related to the MSD. Adjusted models included site (indicator variable) and age (continuous variable).

We derived a new severity score (model-derived score) based on risk of dying in the 14-days after MSD presentation using forward stepwise Cox proportional hazards regression and Akaike Information Criteria (AIC) for model-selection. The following clinical variables were considered in building this model: dysentery, mucoid stool, duration of diarrhea including and prior to the day of enrollment, maximum number of loose stools in last 24 hours, axillary temperature, caregiver-reported vomiting, WHO dehydration status, and clinician decision to hospitalize. Continuous variables were categorized to match that of the previously published MVS [16]. The final Cox model coefficients were used to calculate the new score using methods described elsewhere.[18] The total number of possible points were constrained to 16 and categorized as mild (<6), moderate (6-8), and severe (9+) to be consistent with the MVS.[16]

The model-derived score and the MVS were compared using the area under the curve (AUC) calculated from a logistic model containing deaths in the first 14 days as the outcome and the continuous score as the independent variable with bootstrapped standard errors and a chi-square statistic. Finally, both scores' ability to predict odds of death beyond 14-days (among those 14-day survivors) were also evaluated using logistic-regression based AUCs with bootstrapped standard errors.

To evaluate the robustness of our findings, we repeated all analyses in three subsets of data: 1) Excluding children with another possible etiology based on site and age-adjusted attributable fraction  $\geq 0.5$  in the culture-negative/qPCR-attributable and culture-positive groups; 2) excluding data from Bangladesh because of the uniquely high culture-positivity and dysentery rate at this site [7], 3) excluding the fatal cases that were enriched in the sample (not qPCR-tested randomly).

Analyses were conducted in Stata 14.0 (Stata Corp, College Station, TX) with an alpha of 0.05. The funder had no role in this manuscript's design, data collection, analysis, writing or submission.

#### Results

Of 9,439 MSD cases enrolled in GEMS, 5,670 had qPCR results and were included in the analysis of clinical presentation by *Shigella* diagnostic assay (Figure 1). Sixty percent (n=3,397) of children did not have *Shigella* detected by culture or by qPCR at any C<sub>t</sub> value. *Shigella* was isolated from culture in 745 children (13.1%), the majority (727 [97.5%]) of which were detected by qPCR (697 [95.9%] at qPCR-attributable levels and 30 [4.1%] qPCR-unattributable). Of the culture-positive *Shigella* cases, 65.4% were *S. flexneri*, 24.0% *S. sonnei*, 5.5% *S.dysenteriae*, and 5.1% *S. boydii*, as described elsewhere.[12] Of

4,925 culture-negative MSD cases, qPCR-attributable *Shigella* infections were identified in an additional 852 (17.3%) children and qPCR-unattributable infections in 676 (13.7%).

#### Culture vs. qPCR-based confirmation of Shigella diarrhea

Accounting for potential confounding by age and site, compared to children with culture-positive shigellosis (Table 1a & 1b), cases with culture-negative/qPCR-attributable shigellosis were more likely to be under one year of age, stunted, and have had more than three days of diarrhea; they were also less likely to be febrile or have passed more than 6 loose stools in a day. The prevalence of a concomitant attributable viral pathogen was similar between culture-positive (13.3%) and culture-negative/qPCR-attributable (15.6%) shigellosis as was the likelihood of dysentery, mucoid stool, severe dehydration, vomiting, hospital admission, and a "severe" classification by the MVS. When removing episodes with other potentially attributable pathogens detected, the observed differences in clinical presentation of culture-positive and culture-negative/qPCR-attributable shigellosis did not change for any manifestation other than diarrhea duration, which became a more pronounced difference between the two diagnostic categories (Supplementary Table 1 [S1]). Exclusion of the Bangladesh site (Table S2) resulted in a significantly lower prevalence of dysentery and mucoid stool among children with culture-negative/qPCR-attributable shigellosis, while removal of fatal cases that were enriched in the dataset (Table S3) did not meaningfully change any comparisons.

In contrast, children with culture-negative/qPCR-unattributable shigellosis were more likely than children with culture-positive shigellosis to present with vomiting, severe dehydration, to be hospitalized, and to have a viral etiology (Table 1a & 1b). Similar associations were found when comparing *Shigella* negative (no detection by culture or qPCR) cases to culture-positive shigellosis. Subset analyses revealed similar findings, except the analysis excluding Bangladesh data which found no differences in fever, severe

dehydration, or hospitalization when comparing culture-positive *Shigella* cases to the other two groups (Tables S1-S3).

Bacterial causes other than *Shigella* were more common in all three culture-negative groups (21.7% qPCR-attributable, 24.9% qPCR-unattributable, and 18.1% absent) compared to culture-positive *Shigella* (8.2%, Table 1a & 1b). Subset analyses led to similar findings (Tables S1-S3). These associations did not appear to be driven by a single bacterial pathogen (Table S4a/b).

### Risk factors for death among children with Shigella-attributed diarrhea

The clinical and demographic features of the 1,481 children with either culture-negative/qPCRattributable or culture-positive *Shigella* with known vital status at follow-up are presented, by age, in Table 2. Children under 12 months comprised 14.9% of the *Shigella*-attributed cases and were less likely to present with dysentery and fever and more likely to present with vomiting and dehydration. This trend held true when limiting to qPCR-attributable cases only (Table S5) and culture-positive shigellosis (Table S6).

Forty-two children (2.8%) with *Shigella*-attributable diarrhea died during follow-up, of whom 22 (52.4%) died in the first two weeks. There was no difference in risk of death between culture negative/qPCR-attributable shigellosis and culture-positive shigellosis (aHR: 1.1, 95% CI: 0.5-2.6). Age under 6 months, duration of diarrhea greater than 3 days, vomiting greater than 3 times per day, stunting, MUAC under 12.5cm, and hospital admission were associated with risk of death in the first 14-days (Table 3). Severe dehydration was not statistically associated with death in adjusted models, but 20/22 (90.9%) of the deaths among *Shigella* cases that occurred in the first two weeks were in children with severe

dehydration. Dysentery was not associated with death in adjusted models, nor was presence of a second attributable etiology. When excluding episodes with additional potentially attributable pathogens (Table S7), we found all risk factors remained significantly associated with death, including vomiting. There were no differences in significant risk factors in the subset of children excluding Bangladesh (Table S8). Finally, excluding enriched fatal cases (Table S9), young age and diarrhea duration were no longer significantly associated with death, although the direction and magnitude of association were similar to primary analyses.

Based on model-fit, three clinical features maximally predicted death: clinician decision to hospitalize, dehydration status, and diarrhea duration prior to presentation (Table 4, Table S10). This model-derived score had an AUC of 0.85 for predicting 14-day mortality (95% CI: 0.76-0.91). The MVS had a similar AUC (AUC=0.80, 95% CI: 0.71-0.88, p-value<sub>AUC derived vs. AUC MVS</sub> = 0.077) (Figure 2). The model-derived score classified 564 (38.1%) of children as severe, 409 (27.6%) as moderate, and 729 (49.2%) as mild whereas MVS classified 559 (37.7%) as severe, 721(48.7%) as moderate, and 201 (13.6%) as mild. Table 5 displays the median model-derived score and MVS across various characteristics. Against the outcome of death after 14-days among the 1,459 14-day survivors, the two scores did not differ significantly (AUC<sub>model</sub>= 0.75, 95% CI: 0.59-0.87 vs. AUC<sub>MVS</sub>: 0.67, 95% CI: 0.53-0.80, p=0.064). Among children 12 months and older (n=1,261) or in those with culture-confirmed *Shigella* (n=707), the AUC values were not meaningfully different (Figures S1 and S2) other than a significantly higher AUC for the model-derived score in predicting 14-day mortality (p=0.048).

#### Discussion

Our secondary analysis of children under 5 years of age presenting to health centers with MSD found the clinical presentations of culture-positive shigellosis and culture-negative/qPCR-attributable shigellosis to differ in terms of fever, duration, and stool frequency but not in terms dehydration, vomiting, and MVS

severity. Presuming culture-negative/qPCR-attributable *Shigella* infections are indeed *Shigella* diarrhea, as we routinely assume with culture-positive *Shigella*, culture missed half of *Shigella*-attributed MSD cases in this study, with a higher likelihood of missing the diagnosis in children under 12 months and those who were stunted. Infants and malnourished children may require a lower inoculum to cause MSD which may go undetected by culture methods. Culture also missed over half of the *Shigella*-associated deaths. A simplified severity score, comprised of dehydration status, diarrhea duration prior to presentation, and clinician decision to hospitalize performed similarly to a MVS at predicting mortality in the two weeks following *Shigella*-diarrhea presentation.

While reliance on culture determination alone in vaccine trials will require larger trials, omission of culture-confirmation will not be conducive to *Shigella* serotyping, a critical consideration in assessing serotype-specific immunity. Antibiotic resistance determination in *Shigella* and other enteric bacteria, an important secondary endpoint of *Shigella* vaccine trials, will also require cultured isolates for interpretation. In the absence of methods for culture-independent serotyping and resistance testing, we suggest that vaccine trials be powered for culture-confirmation with a pre-specified secondary molecular microbiologic endpoint that disaggregates low and high concentrations of *Shigella* DNA.

Care-seeking for diarrhea has been shown to correlate with severity, linear growth faltering, and mortality.[19-21] Medically-attended diarrhea has been suggested as a key feature of vaccine trial endpoints[5] and practically, centralizes clinical assessments of dehydration and other indicators of severity. With a 14-day *Shigella* case fatality rate of approximately 1.5%, an MSD definition among children seeking care (as used in GEMS) may be sufficient for clinical endpoint severity classification. Alternatively, the MVS or the simplified model-derived severity score could be used to stratify severity among care-seeking diarrhea cases. The three factors included in our model-derived score are all included

in the MVS and the two scores performed similarly at predicting immediate deaths, despite our score being derived within the same dataset in which its performance was validated. Dysentery did not end up in the model-derived score, possibly because of antibiotic management of dysentery according to WHO guidelines. [22] Despite this lack of association, we advocate for dysentery to be included in a severe *Shigella* case definition because it is a sign of intestinal inflammation and epithelial destruction, consequences of *Shigella* that likely lead to its long-term impact on growth.

Risk factors analyses revealed the host factors of young age, stunting, and wasting to be independently associated with death, as has been found previously.[23-26] The clinical presentation of *Shigella* in infants more commonly included vomiting, dehydration, and absence of dysentery, which is consistent with previous findings among infants in Bangladesh using culture-based diagnosis.[23] Decisions about when to introduce a *Shigella* vaccine must weigh the lower burden, but higher risk, of *Shigella* infection among infants against the difficulty of including new vaccinations in the early infant immunization schedule. Moreover, vaccination prior to 6 months may be more effective by pre-dating nutritional deterioration attributed to *Shigella*.

There were a number of limitations to this analysis. Because of its exploratory nature, the limited number of deaths, as well as the use of AIC, rather than p-values, for developing a severity score, we did not account for multiple comparisons. We were limited by which and how clinical data was collected to inform the severity scores. For example, two previously validated MVS[27, 28] could not be generated with this data due to the unavailability or lack of finer categorization of some symptoms. Validation of our model-derived *Shigella* severity score in other cohorts where death or other poor outcomes are ascertained, would strengthen this score's utility and generalizability. We chose 14-day mortality as the gold standard measure of severity; however children may have severe shigellosis and survive. We found

bacteria other than *Shigella* to be more common in children with culture-negative/qPCR-attributable *Shigella* (but not viral etiologies) and cannot exclude the possibility that a small subset of this group of children had a bacterial etiology other than *Shigella*. Ultimately, vaccine trials that include both culture-and qPCR-based diagnostics can confirm culture-negative qPCR-attributable episodes are indeed *Shigella*.

The GEMS study design and extensive diagnostic testing provided a unique opportunity to compare clinical features of *Shigella* by diagnostic categories and to examine risk factors for death. A composite severity score consistent with severe disease or dysentery may be a pragmatic clinical endpoint (confirmed by culture and secondarily, by qPCR) in vaccine trials.

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#### References

- 1. Khalil IA, Troeger C, Blacker BF, et al. Morbidity and mortality due to shigella and enterotoxigenic Escherichia coli diarrhoea: the Global Burden of Disease Study 1990-2016. Lancet Infect Dis **2018**.
- 2. Mani S, Wierzba T, Walker RI. Status of vaccine research and development for Shigella. Vaccine **2016**; 34(26): 2887-94.
- 3. Barry E, Cassels F, Riddle M, Walker R, Wierzba T. Vaccines Against Shigella and Enterotoxigenic Escherichia coli: A summary of the 2018 VASE Conference. Vaccine **2019**; 37(34): 4768-74.
- 4. Anderson JDt, Bagamian KH, Muhib F, et al. Potential impact and cost-effectiveness of future ETEC and Shigella vaccines in 79 low- and lower middle-income countries. Vaccine X **2019**; 2: 100024.
- 5. Porter CK, Gutierrez RL, Kotloff KL. Clinical endpoints for efficacy studies. Vaccine **2019**; 37(34): 4814-22.
- 6. Liu J, Kabir F, Manneh J, et al. Development and assessment of molecular diagnostic tests for 15 enteropathogens causing childhood diarrhoea: a multicentre study. The Lancet Infectious diseases **2014**; 14(8): 716-24.
- 7. Liu J, Platts-Mills JA, Juma J, et al. Use of quantitative molecular diagnostic methods to identify causes of diarrhoea in children: a reanalysis of the GEMS case-control study. Lancet **2016**; 388(10051): 1291-301.
- 8. Soares-Weiser K, Bergman H, Henschke N, Pitan F, Cunliffe N. Vaccines for preventing rotavirus diarrhoea: vaccines in use. Cochrane Database Syst Rev **2019**; 3: CD008521.
- 9. Ruuska T, Vesikari T. Rotavirus disease in Finnish children: use of numerical scores for clinical severity of diarrhoeal episodes. Scand J Infect Dis **1990**; 22(3): 259-67.
- 10. Kotloff KL, Nataro JP, Blackwelder WC, et al. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. The Lancet **2013**; 382(9888): 209-22.
- 11. Kotloff KL, Blackwelder WC, Nasrin D, et al. The Global Enteric Multicenter Study (GEMS) of diarrheal disease in infants and young children in developing countries: epidemiologic and clinical methods of the case/control study. Clin Infect Dis **2012**; 55 Suppl 4: S232-45.
- 12. Livio S, Strockbine NA, Panchalingam S, et al. Shigella Isolates From the Global Enteric Multicenter Study Inform Vaccine Development. Clin Infect Dis **2014**.
- 13. Panchalingam S, Antonio M, Hossain A, et al. Diagnostic microbiologic methods in the GEMS-1 case/control study. Clin Infect Dis **2012**; 55 Suppl 4: S294-302.
- 14. Liu J, Almeida M, Kabir F, et al. Direct Detection of Shigella in Stool Specimens by Use of a Metagenomic Approach. J Clin Microbiol **2018**; 56(2).
- 15. World Health Organization. Chart Booklet: Integrated Management of Childhood Illness. Geneva, Switzerland. WHO 2014.
- Kotloff KL, Platts-Mills JA, Nasrin D, Roose A, Blackwelder WC, Levine MM. Global burden of diarrheal diseases among children in developing countries: Incidence, etiology, and insights from new molecular diagnostic techniques. Vaccine 2017; 35(49 Pt A): 6783-9.

- 17. Platts-Mills JA, Liu J, Rogawski ET, et al. Use of quantitative molecular diagnostic methods to assess the aetiology, burden, and clinical characteristics of diarrhoea in children in low-resource settings: a reanalysis of the MAL-ED cohort study. Lancet Glob Health **2018**; 6(12): e1309-e18.
- 18. Sullivan LM, Massaro JM, D'Agostmo RB. Presentation of multivariate data for clinical use: The Framingham Study risk score functions. Stat Med **2004**; 23(10): 1631-60.
- Lamberti LM, Fischer Walker CL, Taneja S, Mazumder S, Black RE. The Influence of Episode Severity on Caregiver Recall, Care-seeking, and Treatment of Diarrhea Among Children 2-59 Months of Age in Bihar, Gujarat, and Uttar Pradesh, India. Am J Trop Med Hyg 2015; 93(2): 250-6.
- 20. Lamberti LM, Fischer Walker CL, Black RE. Systematic review of diarrhea duration and severity in children and adults in low- and middle-income countries. BMC Public Health **2012**; 12: 276.
- 21. Kotloff KL, Nasrin D, Blackwelder WC, et al. The incidence, aetiology, and adverse clinical consequences of less severe diarrhoeal episodes among infants and children residing in low-income and middle-income countries: a 12-month case-control study as a follow-on to the Global Enteric Multicenter Study (GEMS). Lancet Glob Health **2019**; 7(5): e568-e84.
- 22. World Health Organization. The Treatment of Diarrhoea. A manual for physicians and other senior health workers. Geneva, Switzerland. WHO 2007.
- 23. Huskins WC, Griffiths JK, Faruque AS, Bennish ML. Shigellosis in neonates and young infants. J Pediatr **1994**; 125(1): 14-22.
- 24. Bennish ML, Harris JR, Wojtyniak BJ, Struelens M. Death in shigellosis: incidence and risk factors in hospitalized patients. J Infect Dis **1990**; 161(3): 500-6.
- 25. Bennish ML, Wojtyniak BJ. Mortality due to shigellosis: community and hospital data. Rev Infect Dis **1991**; 13 Suppl 4: S245-51.
- 26. Levine MM, Nasrin D, Acacio S, et al. Diarrhoeal disease and subsequent risk of death in infants and children residing in low-income and middle-income countries: analysis of the GEMS case-control study and 12-month GEMS-1A follow-on study. The Lancet Global health **2020**; 8(2): e204-e14.
- 27. Freedman SB, Eltorky M, Gorelick M, Pediatric Emergency Research Canada Gastroenteritis Study G. Evaluation of a gastroenteritis severity score for use in outpatient settings. Pediatrics **2010**; 125(6): e1278-85.
- 28. Schnadower D, Tarr PI, Gorelick MH, et al. Validation of the modified Vesikari score in children with gastroenteritis in 5 US emergency departments. J Pediatr Gastroenterol Nutr **2013**; 57(4): 514-9.

## **Figure Legends**

Figure 1. Participant flow of children with MSD included in each analysis

Figure 2. ROC curves of model-derived score & modified Vesikari score predicting death in first 14-days and within 60-days (range 50-90 days) among 14-day survivors. Model-derived score:  $AUC_{0-14}$  of 0.85 (95% CI: 0.77-0.92);  $AUC_{15-90}$  0.75 (95% CI: 0.60-0.87). Modified Vesikari score:  $AUC_{0-14}$ : 0.80 (95% CI: 0.70-0.88);  $AUC_{15-90}$ : 0.67 (95% CI: 0.53-0.80)

	Model-derived score [14-day]	
_ <b> _</b> .	Model-derived score [60-day]	2
	Modified Vesikari score [14-day]	

Table 1a. Frequencies of sociodemographic, clinical, and patho	gen characte	eristics by diagnostic cat	egories		
	Shigella Culture		Absent		Present
Characteristic	Shigella qPCR	Absent <sup>i</sup> (n=3,397)	qPCR-unattributable <sup>ii</sup> (n=676)	qPCR-attributable <sup>iii</sup> (n=852)	Any qPCR value <sup>iv</sup> (n=745)
		n (%)	n (%)	n (%)	n (%)
Sociodemographic					
Age					-
1m to 11m		1,684(49.6)	129(19.1)	149(17.5)	91 (12.2)
12m to 23m		977(28.8)	271(40.1)	405(47.5)	301 (40.4)
24m to 59m		/36(21.7)	276(40.8)	298(35.0)	353 (47.4)
Female sex		1,422(42.5)	270(40.8)	371(43.3)	339 (45.5)
Bangladash		357(10.5)	35(5.2)	98(11.5)	410 (55 0)
India		567(16.7)	123(18.2)	138(16.2)	58 (7.8)
Kenva		650(19.3)	107(15.8)	83(9.7)	72 (9.7)
Mali		556(16.4)	150(22.2)	163(19.1)	18 (2.4)
Mozambique		337(9.9)	73(10.8)	85(10.0)	24 (3.2)
Pakistan		481(14.2)	110(16.3)	153(18.0)	86 (11.5)
The Gambia		449(13.2)	78(11.5)	132(15.5)	77 (10.3)
Clinical characteristics at enrollment					
Dysentery		459(13.5)	91(13.5)	355(41.7)	534 (71.7)
Caregiver reported mucoid stool		765(22.5)	148(21.9)	255(29.9)	364 (48.9)
Duration of diarrhea (including day of presentation) $\geq 3$ days	5	1,748(51.5)	348(51.5)	489(57.4)	351 (47.1)
$\geq$ 7 loose stools child in 24 hour period		1,213(35.7)	218(32.3)	303(35.6)	424 (56.9)
Temperature ≥38°C		735(21.6)	129(19.1)	140(16.4)	238 (32.0)
Caregiver reported vomiting > 3 times per day		1,509(44.4)	289(42.8)	222(20.1) 427(51.2)	102 (21.7)
Stunted (LAZ = 2)		0/2 (27.0)	<u> </u>	43/(31.3)	221 (30.0)
Simile $(LAL - 2)$		743(47.7)	240(33.7)	510(50.7)	214 (20.7)

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411 (14.3	3) 99 (15.3)	124(15.0)	66 (9.1)
819 (24.1	1) 158 (23.4)	145(17.0)	209 (28.1)
1,544 (45.5	5) 314 (46.5)	298(35.0)	304 (40.8)
1,169 (34.4	4) 172 (25.4)	133(15.6)	99 (13.3)
280 (8.2)	) 41 (6.1)	54(6.4)	15 (2.0)
615 (18.1	1) 168 (24.9)	185(21.7)	61 (8.2)
	411 (14.: 819 (24.: 1,544 (45.: 1,169 (34.: 280 (8.2) 615 (18.:	411 (14.3) 99 (15.3)   819 (24.1) 158 (23.4)   1,544 (45.5) 314 (46.5)   1,169 (34.4) 172 (25.4)   280 (8.2) 41 (6.1)   615 (18.1) 168 (24.9)	$\begin{array}{ c c c c c c c c c c c c c c c c c c c$

i.  $ipaH C_t$  value  $\geq 35$ 

ii.  $27.9 \le ipaH C_t < 35$ 

iii.  $ipaH C_t$  value < 27.9

iv. Absent by TAC (n=18 {2.4%}), present below diarrhea-associated quantity (n=30 {4.0%}), present at, or above, diarrhea associated quantity (n=697 {93.6%})

v. Among those  $\geq 6$  months of age (in whom MUAC is validated)

vi. As derived in Kotloff et al., Vaccine, 2017

vii. Site and age-adjusted attributable fraction  $\geq$ .5 for any of the following: adenovirus, astrovirus, norovirus, rotavirus, sapovirus, adenovirus

viii. Site and age-adjusted attributable fraction  $\geq$ .5 for any of the following: *Cryptosporidium*, *Entamoeba histolytica*, *Cyclospora*, *Isospora* 

ix. Site and age-adjusted attributable fraction 2.5 for any of the following: H. pylori, Campylobacter, Aeromonas, Salmonella, V. cholerae, EAEC, St-ETEC, Lt-ETEC,

tEPEC, STEC

Accept





Table 1b. Sociodemographic, clinical, and pathogen factors associated with Shigella diagnostic categories

	<i>Shigella</i> Culture		Present		
Characteristic	Shigella qPCR	Absent <sup>i</sup> (n=3,397)	qPCR- unattributable <sup>ii</sup> (n=676)	qPCR- attributable <sup>iii</sup> (n=852)	Any qPCR value <sup>iv</sup> (n=745)
		aPR'(95%CI)	aPR <sup>v</sup> (95%CI)	aPR <sup>v</sup> (95%CI)	aPR'(95%CI)
Sociodemographic					
Age < 12 m		4.5 2(3.63-5.63)	1.76(1.33-2.32)	1.57 (1.20-2.05)	Ref
Female sex		0.89(0.78-1.01)	0.85(0.72-1.01)	0.91 (0.78-1.07)	Ref
Clinical characteristics at enrollment					
Dysentery		0.37(0.32-0.42)	0.40(0.31-0.50)	1.06 (0.91-1.22)	Ref
Caregiver reported mucoid stool		0.66(0.57-0.77)	0.72(0.58-0.89)	0.95 (0.80-1.13)	Ref
Duration of diarrhea $\geq 3$ days		1.04(0.92-1.19)	1.09(0.93-1.28)	1.20 (1.04-1.39)	Ref
$\geq$ 7 loose stools child in 24 hour period	od	0.83(0.73-0.94)	0.80(0.67-0.95)	0.82 (0.70-0.96)	Ref
Temperature ≥38°C		0.78(0.66-0.92)	0.71(0.56-0.90)	0.62 (0.50-0.78)	Ref
Caregiver reported vomiting > 3 time	es per	1.04(1.55.2.20)			Ref
day Severe dehydration		1.84(1.55-2.20)	1.85(1.51-2.27)	1.13 (0.91-1.39)	Dof
Severe denyuration		1.22 1.41)	1.22 (1.03-1.45)	1.14 1.34)	Kei
Stunted (LAZ<-2)		(0.89- 1.05 1.24)	1.21 (0.99-1.48)	(1.05- 1.27 1.53)	Ref
Wasted <sup>vi</sup> (MUAC < 12.5cm)		(0.65- 0.861.14)	1.19 (0.85-1.64)	(0.79- 1.08 1.48)	Ref
Hospitalized	Ċ	(1.08- 1.29 1.54)	1.37 (1.10-1.71)	(0.72- 0.90 1.13)	Ref
Severe by modified Vesikari score <sup>vii</sup>		(0.95- 1.111.25)	1.16 (0.98-1.37)	(0.74- 0.87 1.03)	Ref
Other etiologies					
Viral <sup>viii</sup>		(1.80-		(0.91-	Ref
	-	2.24 2.79)	2.56 (1.53-2.56)	1.18 1.55)	
Parasitic <sup>ix</sup>		(1.14-	2 02 (0 88 2 02)	(0.97-	Ref
Other bacteria <sup>x</sup>		(1.69-	2.93 (0.88-2.93)	(1.88-	Ref
		2.24 2.97)	3.88 (2.09-3.88)	2.54 3.44)	

- i.  $ipaHC_t$  value  $\geq 35$
- ii.  $27.9 \le ipaH C_t < 35$
- iii.  $ipaHC_t$  value < 27.9
- iv. Absent by TAC (n=18 {2.4%}), present below diarrhea-associated quantity (n=30 {4.0%}), present at, or above, diarrhea associated quantity (n=697 {93.6%})
- v. Adjusted prevalence ratios (aPR) from relative risk regression assuming Poisson distribution adjusting for site (considered as an indicator variable) and age (considered continuously) except age model adjusted only for site.
- vi. Among those≥6 months of age (in whom MUAC is validated)
- vii. As derived in Kotloff et al., Vaccine, 2017
- viii. Site and age-adjusted attributable fraction ≥.5 for any of the following: astrovirus, norovirus, rotavirus, sapovirus, adenovirus
- ix. Site and age-adjusted attributable fraction ≥.5 for any of the following: Cryptosporidium, Entamoeba histolytica, Cyclospora, Isospora
- x. Site and age-adjusted attributable fraction  $\geq$  .5 for any of the following: *H. pylori*, *Campylobacter*, *Aeromonas*, *Salmonella*, *V*.
- cholerae, EAEC, St-ETEC, Lt-ETEC, tEPEC, STEC

Table 2. Age-stratified characteristics of Sha	gella MSD cases defined as culture	positive or qPCR-attributable (n=1,481)
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		0-5m		6-11m		23m	24-59m	
Characteristic	(n=	35)	( <b>n</b> =	185)	( <b>n</b> =	654)	(n=0	<b>507</b> )
	n	(%)	n	(%)	n	(%)	n	(%)
Sociodemographic								
Female	14	(40.0)	74	(40.0)	307	(46.9)	263	(43.3)
Site								
Bangladesh	3	(8.6)	49	(26.5)	198	(30.3)	252	(41.5)
India	5	(14.3)	19	(10.3)	62	(9.5)	101	(16.6)
Kenya	9	(25.7)	26	(14.1)	58	(8.9)	56	(9.2)
Mali	1	(2.9)	16	(8.7)	87	(13.3)	55	(9.1)
Mozambique	0	(0)	11	(6.0)	49	(7.5)	35	(5.8)
Pakistan	15	(42.9)	32	(17.3)	92	(14.1)	60	(9.9)
The Gambia	2	(5.7)	32	(17.2)	108	(16.5)	48	(7.9)
Clinical characteristics					•			
Dysentery	13	(37.1)	88	(47.6)	345	(52.8)	390	(64.3)
Caregiver reported mucoid stool	13	(37.1)	70	(37.8)	253	(38.7)	247	(40.7)
Duration of diarrhea (including day of								
presentation)								
1-3	19	(54.3)	128	(69.2)	460	(70.3)	448	(73.8)
4-5	14	(40.0)	35	(18.9)	143	(21.9)	124	(20.4)
6+	2	(5.7)	22	(11.9)	51	(7.8)	35	(5.8)
Max # of loose stools child passed in 24 hour								
period								
$\leq 6$	20	(57.1)	95	(51.4)	361	(55.2)	321	(52.9)
7-10	11	(31.4)	66	(35.7)	196	(30.0)	182	(30.0)
>10	4	(11.4)	24	(13.0)	97	(14.8)	104	(17.1)
Axillary temperature at presentation								
<38°C	30	(85.7)	152	(82.2)	510	(78.0)	431	(71.0)
38-38.9°C	3	(8.6)	24	(13.0)	80	(12.2)	106	(17.5)
≥39°C	2	(5.7)	9	(4.9)	64	(9.8)	70	(11.5)
Caregiver reported vomiting $\geq 3$ times per day	12	(34.3)	63	(34.1)	159	(24.3)	124	(20.4)
WHO-defined dehydration categories								
None	11	(31.4)	53	(28.7)	208	(31.8)	255	(42.0)
Some	5	(14.3)	46	(24.9)	165	(25.2)	141	(23.2)
Severe	19	(54.3)	86	(46.5)	281	(43.0)	211	(34.8)
Modified Vesikari Score <sup>i</sup>								
Mild	6	(17.1)	24	(13.0)	78	(11.9)	93	(15.3)
Moderate	15	(42.9)	81	(43.8)	326	(49.9)	299	(49.3)
Severe	14	(40.0)	80	(43.2)	250	(38.2)	215	(35.4)
Stunted (LAZ $\leq$ -2)	14	(40.0)	46	(25.1)	197	(30.3)	220	(36.4)
Wasted (MUAC $< 12.5$ cm) <sup>ii</sup>			50	(27.0)	97	(14.8)	27	(4.5)

<sup>i</sup>As derived in Kotloff et al., Vaccine, 2017 <sup>i</sup>Among children older than 6 months

Table 3. Characteristics of Shigella MSD cases defined as culture positive or qPCR-attributable (N=1,481) w	vho died	vs. those who
survived in the 14 days post enrollment		

		Died		vived	Hazard Ratio <sup>ii</sup>	aHazard Ratio	
Characteristic		(n=22)	( <b>n</b> =	1,459)	(95% CI)	(95% CI) <sup>iii</sup>	
	n	$(\%)^{1}$	n	$(\%)^{1}$			
Sociodemographic							
Age							
Om to 5m	3	(13.6)	32	(2.2)	10.7 (2.6-45.0)	6.3 (1.4-27.5)	
6m to 11m	4	(18.2)	181	(12.4)	2.6 (0.7-9.8)	1.6 (0.4-5.8)	
12m to 23m	10	(45.5)	644	(44.1)	1.9 (0.6-5.4)	1.3 (0.4-3.8)	
24m to 59m	5	(22.7)	602	(41.3)	Ref	Ref	
Sex	_						
Female	7	(31.8%)	651	(44.6%)	0.6 (0.2-1.4)	0.6 (0.2-1.4)	
Male	15	(68.2%)	808	(55.4%)	Ref	Ref	
Clinical characteristics							
Dysentery	_						
Present	7	(31.8%)	829	(56.8%)	0.4 (0.2-0.9)	0.6 (0.2-1.6)	
Absent	15	(68.2%)	630	(43.2%)	Ref	Ref	
Caregiver reported mucoid stool	0			(20.40/)	0.0 (0.4.2.1)	12(0520)	
Present	8	(36.4%)	5/5	(39.4%)	0.9 (0.4-2.1)	1.2 (0.5-3.0)	
Absent	14	(63.6%)	884	(60.6%)	Ref	Ref	
Duration of diarrhea (including day of							
presentation)	10	$(0 \land 40)$	757	(51.00/)	5 9 (1 7 10 ()	4 4 (1 2 15 2)	
$\geq 3$	19	(86.4%)	/5/	(51.9%)	5.8 (1.7-19.6)	4.4 (1.3-15.3)	
	3	(13.6%)	702	(48.1%)	Ref	Ref	
Max # of loose stools child passed in 24 hour							
period	-	(21.00())		(16 10/)	0.5 (0.0.1.0)	11(0400)	
$\geq 1$	1.7	(31.8%)	6//	(46.4%)	0.5 (0.2-1.3)	1.1 (0.4-2.9)	
	15	(68.2%)	782	(53.6%)	Ref	Ref	
1 emperature	0	(10.00())	240	(22.00())	22(0051)	21(0051)	
≥38°C	12	(40.9%)	1110	(23.9%)	2.2 (0.9-5.1)	2.1 (0.9-5.1)	
< so C	15	(39.1%)	1110	(70.1%)	Kel	Kei	
Caregiver reported volinting	10	(5160/)	216	(22.70/)	20(1600)	25(1150)	
> 3 times per day	12	(34.0%)	1112	(25.7%)	5.0 (1.0-0.0)	2.3 (1.1-3.9)	
$\leq 3$ times per day (or none)	10	(45.5%)	1113	(76.3%)	Ref	Ref	
who-defined denydration categories		(00.00/)	577	(20.70/)	17.0 (2.4	7.0.(0.9.70.7)	
Severe	20	(90.9%)	5//	(39.7%)	17.9 (2.4-	7.9 (0.8-79.7)	
		(A, CO())	250	(24.40/)	155.5)	1 2 (0 07 22 5)	
Some	1	(4.0%)	330	(24.4%)	1.5(0.09-	1.5 (0.07-25.5)	
N	1	(1 < 0/)	576	(26.10/)	23.0) Def	Def	
	1	(4.0%)	320	(30.1%)	Kel	Kei	
Chronic Mainutrition	11	(57.00/)	100	(22,10/)	20(1272)	20(1277)	
Stunted (LAZ<-2)	11	(57.9%)	400	(52.1%)	2.9 (1.2-7.2)	3.0(1.2-7.7)	
Non-stunied	8	(42.1%)	988	(08.0%)	Kel	Rei	
Acute manufation $MUAC < 12.5 \text{ cm}$	o	(42.10%)	166	(11.60%)	5 4 (2 2 12 4)	22(1286)	
MUAC < 12.5 cm	0	(42.1%)	1 261	(11.0%)	5.4 (2.2-15.4)	5.5 (1.2-6.0) Pof	
$MUAC \ge 12.5 cm$	11	(37.9%)	1,201	(00.4%)		Kei	
Admission status at enrollment visit	16	(70 70()	210	(01.00/)	0.2(2.6,02,0)	145 (51 41 0)	
Hospitalized	16	(12.1%)	319	(21.9%)	9.3 (3.6-23.8)	14.5 (5.1-41.2)	
Seen as outpatient	6	(27.5%)	1,140	(78.1%)	Ket	Ket	
Modified Vesikari Score'		101 5	<b>_</b>				
Severe	18	(81.8%)	541	(37.1%)	5.9 (2.0-17.4)	4.4 (1.5-12.9)	
Moderate	4	(18.2%)	717	(49.1%)	Ref	Ref	
Mild	0	(0)	201	(13.8%)	Not estimable	Not estimable	

Laboratory						
Shigella culture results						
Culture positive	9	(40.9%)	698	(47.8%)	0.8 (0.3-1.8)	1.1 (0.4-2.6)
Culture negative	13	(59.1%)	761	(52.2%)	Ref	Ref
Shigella qPCR $C_t$ values <sup>vi</sup>						
<20.8	11	(50.0%)	635	(44.9%)	1.5 (0.5-4.7)	1.9 (0.6-6.0)
20.8-24.34	7	(31.8%)	436	(30.8%)	1.4 (0.4-4.7)	1.7 (0.5-5.8)
24.35-27.89	4	(18.2%)	343	(24.3%)	Ref	Ref
Other potential etiology <sup>vii</sup>						
Yes	8	(36.4%)	429	(29.4%)	1.4 (0.6-3.3)	1.4 (0.6-3.3)
No	14	(63.6%)	1,030	(70.6%)	Ref	Ref

i. Column percentages

- ii. From Cox proportional hazards regression including only the variable of interest in the model
- iii. From Cox proportional hazards regression including the variable of interest, site as an indicator variable, and age as a continuous variable except age model adjusted only for site
- iv. Among those≥6 months of age in whom MUAC is validated
- v. As derived in Kotloff et al., Vaccine, 2017
- vi. Among those with qPCR attributable-*Shigella* (n=1436)

CC'

vii. Based on site and age-adjusted attributable fraction ≥0.5 for any of the following pathogens: astrovirus, norovirus, rotavirus, sapovirus, adenovirus, *Cryptosporidium*, *E. histolytica*, *Cyclospora*, *Isospora*, *isospora*, *H. pylori*, *Campylobacter*, *Salmonella*, *V. cholerae*, EAEC, St-ETEC, Lt-ETEC, tEPEC, STEC

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	Model-	Modified
Predictor	derived score	Vesikari Score <sup>i</sup>
Days diarrhea prior to presentation (including day of presentation)		
$\geq 6$	3	3
4-5	2	2
1-3	0	1
Max # of loose stools child passed in 24 hour period		
>10		3
7-10		2
$\leq 6$		1
Max # of vomiting episodes in 24 hour period		
> 3 times per day		2
$\leq$ 3 times per day (or none)		1
WHO-defined dehydration categories		
Severe	8	3
Some	4	2
None	0	0
Temperature		
≥39°C		3
38.5-38.9°C		2
37.1-38.4°℃		1
Hospitalized <sup>ii</sup>		
Yes	5	2
No	0	0

Table 4. Shigella model-derived score and modified Vesikari score

i. As derived in Kotloff et al., Vaccine, 2017

ii. 28 participants received IV rehydration in a short-stay ward and these were upgraded to "hospitalized"

	Model-deri	ved score	Modified Vesikari		
Characteristic			SCO	re <sup>i</sup>	
	Median	(IQR)	Median	(IQR)	
Sociodemographic					
Age					
Om to 5m	8	(2-10)	8	(6-11)	
6m to 11m	8	(4-9)	8	(6-10)	
12m to 23m	6	(4-8)	8	(6-9)	
24m to 59m	5	(2-8)	8	(6-9)	
Sex					
Female	6	(2.0)		$(\boldsymbol{\zeta}, \boldsymbol{\Omega})$	
Male Clinical characteristics	6	(3-8)	8	(6-9)	
Dysontery	0	(3-0)	0	(0-9)	
Present	1	(0.8)	7	(6.0)	
Absent	8	(5-10)	8	(0-7) (7-10)	
Caregiver reported mucoid stool		(510)	0	(7 10)	
Present	5	(2-8)	8	(6-9)	
Absent	8	(4-9)	8	(7-9)	
Duration of diarrhea (including day of presentation)					
>3	6	(3-10)	8	(6-9)	
$\overline{3}$	5	(4-8)	8	(6-10)	
Max # of loose stools child passed in 24 hour period		()	-	()	
≥7	6	(4-8)	9	(7-10)	
<7	5	(2-8)	7	(6-9)	
Temperature					
≥38°C	6	(4-9)	10	(8-11)	
<38°C	6	(2-8)	7	(6-9)	
Caregiver reported vomiting					
> 3 times per day	8	(5-10)	10	(8-11)	
$\leq$ 3 times per day (or none)	5	(2-8)	7	(6-9)	
WHO-defined dehydration categories					
Severe	8	(8-13)	9	(8-11)	
Some	4	(4-6)	7	(6-9)	
None	2	(0-3)	6	(5-8)	
Chronic Malnutrition	-	(2.0)	0	( <b>7</b> 0)	
Stunted (LAZ<-2)	1	(3-9)	8	(7-9)	
Non-stunted	5	(3-8)	8	(6-9)	
Acute main unition $MUAC < 12.5 \text{ cm}$	0	(7, 11)	0	(7, 11)	
MUAC < 12.5 cm	o 5	(7-11) (7-8)	9	(7-11) (6-9)	
Admission status at annollment visit	5	(2-0)	0	(0-))	
Hospitalized	0	(5 - 13)	10	(0, 12)	
Seen as outpatient	1	(2-8)	10	(5-12) (6-8)	
Modified Vesikari score <sup>i</sup>	4	(2-0)	1	(0-0)	
Severe	0	(8.13)	10	(0, 11)	
Severe Moderate	9	(3-13)	10	(9-11)	
Mild	4	(3-8)	7	(0-8)	
	0	(0-0)	5	(4-3)	
Snigena culture results	-	(2, 9)	0	(6, 10)	
Culture positive	5	(2-8)	8 0	(0-10)	
Culture negative	8	(4-9)	8	(0-9)	
Snigella qPCR $C_t$ values "	_	(2,0)	6	(6.10)	
<20.8	5	(3-8)	8	(6-10)	
20.8-24.34	6	(2-8)	8	(6-9)	
24.35-27.89	8	(4-9)	8	(6-9)	
Other potential etiology <sup>1</sup>					

Table 5. Median and interquartile range (IQR) of model-derived and modified Vesikari scores by participant characteristic among 1,481 children with *Shigella*-attributed diarrhea

Yes	7 (4-9)	8	(7-9)
No	5 (2-8)	8	(6-9)

- i. As derived in Kotloff et al., Vaccine, 2017
- ii. Among those≥6 months of age in whom MUAC is validated
- iii. Among those with qPCR attributable Shigella (n=1,436)

iv. Based on site and age-adjusted attributable fraction ≥.5 for any of the following pathogens: astrovirus, norovirus, rotavirus, sapovirus, adenovirus, *Cryptosporidium, E. histolytica, Cyclospora, Isospora, H. pylori, Campylobacter, Salmonella, V. cholerae*, EAEC, St-ETEC, Lt-ETEC, tEPEC, STEC





