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1	The effects of gradual vs. rapid weight loss on serum concentrations of myokines and body composition in
2	overweight and obese females
3 4	Reza Bagheri ^{1,*} , Damoon Ashtary-Larky ² , Bradley T Elliott ³ , Darryn S. Willoughby ⁴ , Mehdi Kargarfard ¹ , Meysam Alipour ² , Nasrin Lamuchi-Deli ² , Wesam Kooti ⁵ , Omid Asbaghi ⁶ , Alexei Wong ^{7,*}
5	1- Department of Exercise Physiology, University of Isfahan, Isfahan, Iran.
6 7	2- Nutrition and Metabolic Diseases Research Center, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.
8 9 10	 Translational Physiology Research Group, School of Life Sciences, University of Westminster, London, UK. School of Exercise and Sport Science, University of Mary Hardin-Baylor, Belton, TX, United States.
11 12	5- Lung Diseases & Allergy Research Center, Research Institute for Health Development, Kurdistan University of Medical Sciences, Sanandaj, Iran.
13	6- Student Research Committee, Lorestan University of Medical Sciences, Khorramabad, Iran.
14	7- Department of Health and Human Performance, Marymount University, Arlington, United States.
15	
16	Correspondence: 1- Reza Bagheri, Email: <u>Will.fivb@yahoo.com</u>
17	2-Alexei Wong, Email: <u>Awong@marymount.edu</u>
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- 33 Context: Research has shown the modulations of Follistatin (FST) and Myostatin (MST) following weight loss
- 34 **Objective:** We evaluated the effects of gradual weight loss (GWL) and rapid weight loss (RWL) on serum MST,
- 35 FST concentrations, and body composition in overweight and obese females.
- 36 Materials and Methods: Thirty-six overweight and obese females successfully completed the study interventions:
- 37 GWL (n = 18) or RWL (n= 18). Serum MST and FST concentrations, as well as anthropometric measurements,
- 38 were collected at baseline and at the conclusion of each weight loss intervention.
- **Results:** MST significantly (p<0.05) concentration decreased in the GWL; while FST, body fat percentage and
- 40 skeletal muscle mass significantly declined in both conditions. The loss in skeletal muscle mass was significantly
- 41 greater in RWL relative to GWL.
- 42 Discussion and Conclusion: GWL was more effective than RWL in preserving skeletal muscle mass in
- 43 overweight and obese females. Moreover, GWL leads to declines in MST concentrations.
- 44 Keywords: body composition, diet, insulin resistance, obesity, weight loss.
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62 Introduction

63 Excess caloric intake along with increased sedentarism have been reported as the most immediate contributors to 64 the increasing occurrence of obesity in society, as well as the subsequent development of several non-65 communicable diseases such as metabolic syndrome, type II diabetes mellitus, cardiovascular diseases, and 66 specific types of cancers(Middelbeek & Breda, 2013). Given the importance of obesity-related health conditions, 67 numerous strategies have been implemented aiming to reduce fat mass (FM). Dietary interventions leading to 68 body mass (BM) loss have been suggested as the first stage to alleviate obesity and allied health risk factors 69 (Jakicic et al., 2001). Correspondingly, diverse dietary interventions have been proposed for FM loss(Freire, 70 2020). Most of the proposed strategies are based off the division of macronutrients (e.g. low-carbohydrate/high-71 fat, high-carbohydrate/low-fat, or low-carbohydrate/high protein diet) (Floegel & Pischon, 2012; Yancy, Olsen, 72 Guyton, Bakst, & Westman, 2004), manipulation of total energy balance to induce either gradual weight loss 73 (GWL), or rapid weight loss (RWL) (Ashtary-Larky et al., 2018). Even though RWL approaches may be 74 attractive to obese individuals, it has been theorized that GWL may yield finer alterations in body composition 75 and anthropometric variables and be better maintained chronically (Ashtary-Larky et al., 2018). Indeed, prior 76 work (Hill, 2008; Lutes et al., 2008; Sbrocco, Nedegaard, Stone, & Lewis, 1999) has advised a GWL method, 77 contending that GWL may generate superior long-term FM levels compared with RWL, which is unlikely to be 78 maintained. However, others (Astrup & Rössner, 2000; Carels, Cacciapaglia, Douglass, Rydin, & O'Brien, 2003; 79 Elfhag & Rössner, 2005; Nackers, Ross, & Perri, 2010) have pointed out that larger calorie deficits and 80 subsequent RWL are more likely to reinforce the BM-change process and produce superior long-term FM loss 81 results. Accordingly, ambiguity continues concerning the optimal energy intake and rate of weight loss 82 demanded most favorable for obesity management.

83 Some myokines such as myostatin (MST) and follistatin (FST) have been reported to be affected by both BM 84 and FM alterations in humans (Flanagan et al., 2009; Hittel, Berggren, Shearer, Boyle, & Houmard, 2009; Milan 85 et al., 2004). MST serves as a negative regulator of skeletal muscle mass (SMM) growth and differentiation, 86 while FST acts as the regulator of MST through binding to the active form of MST and inhibiting the binding of 87 MST to the activin IIB receptor(Amthor et al., 2004). Consequently, FST over-expression and MST inhibition is 88 associated with increased SMM (Wagner, Liu, Chang, & Allen, 2005; Wagner, McPherron, Winik, & Lee, 2002) 89 and decreased FM (McPherron & Lee, 1997; Yang et al., 2001). On the contrary, increased MST concentration 90 have been reported to increase FM (Allen, Hittel, & McPherron, 2011; Reisz-Porszasz et al., 2003). FST was 91 found to be more expressed in lean compared with obese females, and weight loss resulted in increased FST in

92 obese females (Flanagan et al., 2009). For instance, some studies have reported MST (Allen, Cleary, Lindsay, 93 Loh, & Reed, 2010; Allen et al., 2008; Gonzalez-Cadavid et al., 1998; Milan et al., 2004; Park, Berggren, 94 Hulver, Houmard, & Hoffman, 2006) and FST (Allen et al., 2008; Flanagan et al., 2009) concentration to be 95 changed following weight loss. Nevertheless, most studies investigated the intracellular expression (Allen et al., 2010; Allen et al., 2008; Gonzalez-Cadavid et al., 1998; Milan et al., 2004; Park et al., 2006) or protein(Allen et 96 97 al., 2010) in animals (Allen et al., 2010; Allen et al., 2008), and only two studies conducted on obese 98 humans(Milan et al., 2004; Park et al., 2006). Although these previously mentioned studies were conducted on 99 weight loss, fewer studies have examined the alterations of FST and MST concentrations following the rate of 100 weight loss. Only Motevalli et al. (2015) reported a significant increase in MST and a decrease in FST 101 concentration following RWL compared to GWL in competitive wrestlers(Motevalli et al., 2015). 102 Combined, these studies suggest a relationship between BM and/or FM reductions and the MST 103 signaling pathway. However, little attention has focused on MST and FST following weight loss in sedentary 104 females. Given the effects of MST and FST in regulating SMM and FM, we hypothesized that FST and MST 105 could be affected by the rate of weight loss. Accordingly, as the primary purpose, this paper aimed to evaluate 106 alterations in MST and FST concentrations following different models of FM loss (RWL vs GWL) in 107 overweight and obese females. As a secondary aim, we assessed body composition changes after RWL and

108 GWL.

109 Materials and Methods

110 Participants

111 Thirty-six overweight and obese (body mass index, $BMI > 25 \text{ kg/m}^2$) female participants who were referred to 112 our nutrition clinic [(Ahvaz, Iran); (age: 35 ± 11 y, weight: 83.7 ± 13.6 kg, BMI: 33.1 ± 6.9 kg/m²)] successfully 113 completed this study. The study protocol was approved by the Ethics Committee of Jundishapur University of 114 Medical Sciences and conducted in accordance with the Declaration of Helsinki and was registered in Iranian 115 Registry of Clinical Trials (IRCT2016010424699N2). Procedures were explained to participants and written 116 informed consent was obtained before participation. Inclusion criteria were as follows: not to contribute in any 117 physical activity (PA) at least in the last year (self-reported), not to use alcohol or smoke, not to use herbal 118 supplements and vitamins, and a lack of BM changes in the last 6 months (self-reported). Exclusion criteria were 119 non-willingness to continue nutritional protocols, participation in other dietary weight loss procedures, 120 pregnancy, or breastfeeding, use of effective drugs on metabolism, lipid and glycemic profile, eating disorder, 121 diabetes, cardiovascular disease, kidney problems, thyroid, digestive, and respiratory diseases. In addition,

122 participants consuming more than 300 mg of caffeine daily (described as caffeine users (Shirali et al., 2016))

123 were excluded from the study.

124 Study design

This double-blinded clinical trial was conducted on overweight and obese female participants. All participants were initially screened for enrolment by phone and social media (Instagram or Telegram) and scheduled for a preliminary visit at nutrition clinic (preliminary visit). The preliminary session included measures of height, weight, waist circumference and questionnaires that confirm their eligibility for the study. Subsequently, Participants were randomly assigned into a GWL (n = 18; 34.4 ± 7.4 y, 83.6 ± 13.5 kg, 44 ± 6.4 body fat percentage (BFP]) or RWL (n = 18; 33.1 ± 11.7 y; 83.7 ± 14.1 kg; 40.8 ± 9.6 BFP) condition. Participants were asked not to change their PA levels throughout the study.

132 Diet protocol

133 The methods and design of were previously reported in detail elsewhere (Ashtary-Larky et al., 2017). Briefly, RWL and GWL, based on the lost weight (at least 5 %), were defined over a period of 5 weeks and 15 weeks, 134 135 respectively. Dietary restrictions were calculated from individual daily energy expenditure requirements. Total energy expenditure was estimated from the Dietary Reference Intake (DRI) for non-obese adult women(Table, 136 137 2005). The DRI prediction formula requires an estimate of the level of PA, age, weight, and height. To achieve 138 and maintain the dietary energy restriction, the food exchange system from the Academy of Nutrition and Dietetics 139 and American Diabetes Association was used (Wheeler, Daly, & Evert, 2014). The prescribed calorie-restricted 140 diet contained 15% protein, 30% to 35% fat, and 50% to 55% carbohydrate, on average, in order to provide weight 141 loss. In general, the meal plans included 3 main meals (breakfast, lunch, and dinner) and three snacks (mid-142 morning, mid-afternoon, and bedtime), and low saturation and trans fats, cholesterol, salt (sodium), and added 143 sugars. All diets were designed according to Dietary Guidelines for Americans, 2010 (Motevalli et al., 2015). Low-144 calorie diets produced an energy deficit of 500 to 750 for the 15-week duration GWL, and 1000 to 1500 kcal/d for 145 the 5-week duration for the RWL.

146 Anthropometric measurements

147 Prior to arriving to the laboratory, participants fasted for 12 hours (overnight, with an aim of 8 hours sleep),

148 refrained from consuming alcohol for 48 hours caffeinated beverages, and other diuretics before assessments.

149 The stature was measured with a stadiometer (Race Industrialization, China) to the nearest 0.1 cm. BM, BMI,

150 BFP, waist-hip ratio (WHR), and SMM were evaluated by a multi-frequency bioelectrical impedance device

- 151 (Inbody 230, Biospace, Korea) (Bagheri, Rashidlamir, Motevalli, et al., 2019). The test-retest reliability of the
- bioelectrical impedance method was high (R = 0.95 to 0.98).

153 Blood sampling and laboratory analysis

- 154 Fasting blood samples (5 mL) were taken from the cubital vein using standard procedures following 12-hour
- 155 overnight fasting (Bagheri, Rashidlamir, Ashtary-Larky, et al., 2019). The initial collection occurred 48 hours
- 156 before initiation of dieting. Blood samples were clotted and stored for 20 minutes at room temperature before
- 157 being centrifuged at 3000 revolutions per minute for 20 minutes. Spun serum was removed from the centrifuge
- 158 and frozen at -80°C. Serum MST (human MST, Zellbio Gmbh, Germany) and FST (human FST, Zellbio
- 159 Gmbh, Germany) concentration were measured in duplicate using enzyme-linked immunosorbent assay
- 160 (ELISA) with a microplate reader (GDV, Germany) at a wavelength of 450 nm. The intra-assay and inter-assay
- 161 coefficient of variation were less than 10% and 12% for MST and FST, respectively.

162 Nutrient intake and dietary analysis

- 163 Participants in RWL submitted 3-day (2 weekdays and 1 weekend) food records at baseline and after week 3 and
- 164 5, while participants in GWL submitted their food records at baseline and after week 3, 6, 9, 12, and 15. Each
- item of food was individually entered into Diet Analysis Plus version 10 (Cengage, Boston, MA, USA) and total
- 166 energy consumption, and the amount of energy derived from proteins, fats, and carbohydrates was assessed
- 167 (Bagheri, Rashidlamir, Motevalli, et al., 2019).

168 Statistical analysis

- 169 An a priori sample size calculation was conducted using the G*Power analysis software (Faul, Erdfelder, Lang, 170 & Buchner, 2007). Our rationale for sample size was based on our prior data (Bagheri et al., 2020) and estimated 171 that 15 participants per condition (30 total participants) would provide 80% power (two-sided $\alpha = 0.05$) to detect 172 significant change in FST and MST concentrations. Shapiro-Wilk test was performed to assess the normality of 173 data. Student's t test was performed for condition comparisons at baseline. A 2×2 RM ANOVAs [time (pre-174 intervention vs. post-intervention) × condition (GWL vs. RWL)] with Bonferroni adjustments was used to 175 determine treatment differences. When appropriate, a one-way ANOVA across change scores was used to detect 176 between-condition differences. Following extra sum-of squares F test, Pearson's correlation was performed 177 between body composition and endocrine markers with data treated as one condition or separated by condition 178 (GWL vs RWL) as suitable. Statistical significance was set at p < 0.05. Cohen's d effect size (ES) was calculated
- as post effect mean minus pre effect mean/pooled pre effect standard deviation means(Cohen, 1992). An ES of

- 180 0.00-0.19 was considered trivial, 0.20-0.49 = small, 0.50-0.79 = moderate, and $\ge 0.80 = \text{large}$. All analyses were
- 181 performed using SPSS (version 25.0, IBM; Chicago, IL).
- 182 Results

Of sixty-five participants that assessed for eligibility, forty-one participants underwent evaluation. Twenty participants were assigned to RWL and 21 were assigned to GWL. Of these, 36 participants completed the study (18 participants per condition). During the study, two participants in RWL (inability for maintaining diet) and three in GWL (exercising, pregnancy, or medication) were excluded. No adverse events in the two study conditions were reported. Baseline characteristics of RWL and GWL conditions are presented in Table 1. No significant differences were observed at baseline between two conditions for any variable (Table 1).

189 Energy and macronutrients consumption

190 Energy and macronutrients of RWL and GWL are presented in Table 2. A significant main effect of time was 191 observed for kilocalories, protein, fat, and carbohydrate consumption in both conditions (P<0.05). Participants in 192 RWL condition consumed significantly less kilocalories, protein, fat, and carbohydrates during week 3 and 5 193 compared to baseline. Participants in GWL condition consumed significantly less kilocalories, protein, fat, and 194 carbohydrates during week 3, 6, 9, 12, and 15, compared to baseline (P<0.05). In addition, participants in GWL 195 condition consumed significantly less kilocalories comparted to week 3, 6, and 9 (P<0.05). In addition, participants 196 in GWL condition during week 12 consumed significantly less protein compared to week 3. Moreover, participants 197 in GWL condition during week 12 consumed less carbohydrate compared to week 6 and 9 (P<0.05).

198 Rate of weight loss alters body composition

- 199 A significant condition \times time interaction was detected for BFP [(p = 0.002), (Figure 1B)] and SMM [(p < 0.001),
- 200 (Figure 1C)]. BFP [GWL = -2.8 % (95% CI= -3.5 to -2.1), (d= 1.9) and RWL = -1.2 % (95% CI, -1.9 to -0.5), (d=
- 201 0.9)] and SMM [GWL = -0.4 kg (95% CI= -0.7 to -0.1), (d= 0.7) and RWL = -1.4 kg (95% CI, -1.7 to -1), (d= 2)]
- significantly decreased in both conditions over time. No condition \times time interaction was noted for BM (p = 0.637),
- BMI (p= 0.403) or WHR [p= (0.202), (Figure 1A)]. Each variable was noted to be significantly decreased with
- 204 time BM (p < 0.001, η 2 = 0.961), BMI, (p < 0.001, η 2 = 0.957), BFP (p < 0.001, η 2 = 0.679), WHR (p < 0.001, η 2 = 0.679), WHR (p < 0.001, η 2 = 0.961), BMI, (p < 0.001, η 2 = 0.961), BMI (p < 0.001, \eta2 = 0.961), BMI (p < 0.961
- 205 $\eta 2 = 0.573$), and SMM (p < 0.001, $\eta 2 = 0.680$); (Table 1). When SMM was examined as change in (post-pre,
- Δ SMM), significantly greater loss in SMM was noted in RWL relative to GWL (p < 0.001, Figure 1D).

207 GWL condition alters MST and FST while RWL only alters FST concentration

- 208 A significant condition \times time interaction was detected for MST [(p = 0.011), (Figure 1E)], FST [(p = 0.036),
- 209 (Figure 1F)], and FST/MST ratio [(p = 0.006), (Figure 1G)]. MST only decreased in GWL [GWL = -44.4 ng/l]

210 (95% CI= -67.4 to -21.5), (d= 0.9)] while FST [GWL = -3 ng/ml (95% CI= -4.6 to -1.5), (d= 0.3) and RWL= -5

211 ng/ml (95% CI, -6.2 to -3.9), (d= 2.1)] significantly decreased in both conditions. In addition, FST/MST ratio

212 [RWL= -0.01 (95% CI, -0.02 to -0.006), (d= 0.9)] significantly diminished in RWL.

213 Change in FST associates with change in BFP independent of weight loss model

Following sum-of-square F-test, weight loss condition was either ignored (Figure 2C and 2D), or considered (Figure 2B) when examining changes in endocrine markers as a function of change in body composition. The alteration in FST (Δ FST) was differentiated between GWL and RWL (p = 0.013), and the change in BFP (Δ BFP) in both conditions was associated with Δ FST (GWL r² = 0.531, p = 0.025; RWL r² = 0.342, p = 0.011; Figure 2B). No other condition was differentiated by sum-of-squares F test, nor considered to be associated by Pearson's correlation when considered as one condition (Figure 2A) r² = 0.023, p = 0.382; Figure 2C), r² = 0.098, p = 0.063;

220 Figure 2D), $r^2 = 0.020$, p = 0.413).

221 Discussion

The major findings were as follows: GWL resulted in a significant decrease in MST concentration, an effect not seen in the RWL group. In addition, both GWL and RWL resulted in a significant decrease in FST concentration. The FST/MST ratio significantly diminished in RWL. In regards to body composition, BFP significantly decreased in both conditions. Moreover, significantly greater loss in SMM was noted in RWL relative to GWL. Collectively, our results suggest a more catabolic environment in the RWL than the GWL condition.

227 MST is a member of transforming growth factor-beta (TGF- β) family member which inhibits muscle 228 differentiation and growth(Allen et al., 2011). Mice in which MST processing or signaling is disrupted exhibited 229 muscle mass gains (Matsakas et al., 2009; Yang et al., 2001), while MST over-expression resulted in a significant 230 decrements of muscle mass(Reisz-Porszasz et al., 2003). These results confirm MST's critical role in inhibiting 231 muscle mass gains. Follistatin is a member of the TGF- β superfamily (Görgens et al., 2013), which is ubiquitously 232 expressed in all tissues of the human body, including skeletal muscle, and has both paracrine and autocrine 233 influences. FST has shown to bind MST and inhibit its activity(Nakatani, Kokubo, Ohsawa, Sunada, & Tsuchida, 234 2011), but also can bind and inhibit other TGF- β family members (Tsuchida et al., 2000), suggesting a more 235 diverse physiological role. Obesity is associated with increased MST expression in both adipose and skeletal 236 muscle tissues(Allen et al., 2011), and MST mRNA levels decreased during weight loss following daily injection 237 of recombinant leptin in mice (Allen et al., 2008). Similarly, circulating FST concentration is also elevated in 238 obese individuals relative to normal weight controls (Maïmoun et al., 2020), concentration of which is subsequently reduced following bariatric surgery-induced weight loss (Wiewiora et al., 2020). These resultshighlight a role of weight loss to reduce MST and FST concentrations.

To date, only one study has been conducted comparing GWL and RWL on serum MST and FST concentrations. Motevalli et al. (2015) examined 8 weeks of weight loss in male competitive wrestlers, serum MST and FST concentrations remained unchanged in the GWL condition while a significant increase in serum MST and a significant decrease in FST concentration was observed in the RWL condition. In their study, serum MST/FST was significantly increased in the RWL condition (Motevalli et al., 2015), suggesting a more catabolic environment was present in the RWL compared to RWL condition, similar to the findings of the results we present herein.

247 At the end of our study, GWL experienced 711 kcal/d and 13.6 g/d of protein reduction in daily energy intake 248 while RWL experienced a quite severe 1339 kcal/d and 44.2 g/d of protein reduction. Moreover, FST/MST was 249 decreased following RWL, indicating a drop in positive muscle growth leading to their deficiency in the 250 homeostatic balance of the muscle. Considering the losses in SMM observed our RWL participants, ultimately 251 these changes may have led to a greater catabolic condition in RWL compared to GWL. The main mechanism of 252 this catabolic condition could be due the activation of AMP-activated protein kinase (AMPK), which is an 253 intracellular energy sensor and the activation of AMPK occurs when the ratio of ATP/ADP decreases in result of 254 caloric restriction. The activation of AMPK can directly activate Tuberous Sclerosis 2 (TSC2) signaling, which is 255 an antagonist of mTORC1 activation(Laplante & Sabatini, 2009), involved in the regulation of MST 256 signaling(Elliott, Renshaw, Getting, & Mackenzie, 2012). It could be speculated that these mechanisms are 257 responsible, at least in part, for inducing a catabolic intracellular condition witnessed herein; however, more 258 invasive measures such as muscle biopsies would be required to confirm this mechanism.

259 Participants in GWL gradually reduced calories at a rate of 500-750 kcal/d for 15 weeks, while participants 260 in RWL reduced calories at a rate of 1000-1500 kcal/d for 5 weeks. Both conditions significantly decreased BM, 261 BFP, WHR, and SMM. It was not surprising, however, that SMM was decreased to a greater extent in RWL 262 relative to GWL due to the respective reduction of daily protein intake of 44.2 g/d and 13.6 g/d. In a systematic 263 review and meta-analysis study, we have shown the beneficial effects of GWL compared to RWL on BFP in 264 overweight individuals, which is in line with the outcomes of the current study. However, our review also showed 265 that fat-free mass was not different following both rates of weight loss (Ashtary Larky, Bagheri, Abbasnezhad, & 266 M Tinsley, 2020). Additionally, previous studies showed that RWL diets are suboptimal for lean body mass preservation (Ashtary-Larky et al., 2017; Peos, Norton, Helms, Galpin, & Fournier, 2019; Vink, Roumans, 267 268 Arkenbosch, Mariman, & van Baak, 2016). It has been shown that lean body mass contributes approximately 269 ranged between 20-30% to total weight loss in individuals with overweight or obesity(Tinsley & Willoughby, 270 2016). Our BFP and SMM results agree with the study of Motevalli et al. (2015) who observed a significant BFP 271 reduction and lean body mass in both RWL and GWL conditions. In our study, GWL experienced a decrease in 272 BFP by 3.2 % more than RWL, likely due to the longer caloric restriction. Indeed, our results revealed an 273 interesting correlation between the changes in BFP and those of FST concentration in both conditions, with a 274 higher r^2 value (= 0.531) with GWL. These findings indicate that participants with smaller deteriorations in FST 275 were the ones with higher decreases in BFP. Although it is not possible to infer causality from correlation, i.e., 276 whether the smaller decrement of FST leads to a higher decrease in BFP or vice versa, the present findings are 277 consistent with the model proposed by Brown et al. (2012) in which the declines in FST concentration may explain 278 increases in fat mass in mice(Brown et al., 2011). The authors proposed that a deterioration in FST concentration 279 could lead to an impaired glucose homeostasis and adipose tissue accumulation due to its important physiological 280 roles in regulating these processes (Brown et al., 2011).

281 It is important to mention that our investigation possesses several limitations. First, we did not measure tissue 282 level expression of MST and FST, which may give further insight into some of our results if collected at both 283 muscle and sub-cutaneous adipose sites. However, it has been proposed that increases in circulating concentrations 284 of cell-signaling molecules enhances the likelihood of receptor interaction and consequently improve the 285 probability of a physiological effect within these tissues (Kraemer et al., 1990; Patel & Demontis, 2014). Second, 286 we used bioelectrical impedance to measure body composition, which is more feasible in field-based settings but 287 not as accurate as other options such as DEXA. However, previous studies have shown that it is a valid and reliable 288 method (Jackson, Pollock, Graves, & Mahar, 1988; Ling et al., 2011). Since we evaluated MST and FST, as well 289 as body composition markers following RWL and GWL in overweight female participants, our results may not be 290 considered universal to all population types. Finally, we used self-reported dietary intake (via food record), which 291 has been shown to be vulnerable to social desirability bias with under-reporting of energy intake and over-reporting 292 of fruit and vegetable intake(Schoeller, 1995).

This study was the first to evaluate the effects of GWL and RWL on serum FST and MST in overweight and obese females. Since both options resulted in matched weight loss, and RWL also resulted in reductions in MST and SMM, we would suggest the RWL condition induced a more catabolic environment for SMM and muscle protein. As a result, we conclude that GWL was more effective than RWL in preserving skeletal muscle mass in overweight and obese females. Future efforts should investigate if these differences in serum MST and FST and differences in SMM loss are maintained in the presence of caloric deficit but maintenance of dietary protein intake.

299	Conflict of interest
300	The authors declare no conflicts of interest.
301 302	Data availability: Data sharing is applicable.
303 304 305 306 307	References:
308 309 310	Allen, D. L., Cleary, A. S., Lindsay, S. F., Loh, A. S., & Reed, J. M. (2010). Myostatin expression is increased by food deprivation in a muscle-specific manner and contributes to muscle atrophy during prolonged food deprivation in mice. <i>Journal of applied physiology</i> , 109(3), 692-701.
311 312 313 314 215	 Allen, D. L., Cleary, A. S., Speaker, K. J., Lindsay, S. F., Uyenishi, J., Reed, J. M., Mehan, R. S. (2008). Myostatin, activin receptor IIb, and follistatin-like-3 gene expression are altered in adipose tissue and skeletal muscle of obese mice. <i>American Journal of Physiology-Endocrinology and Metabolism, 294</i>(5), E918-E927. Allen, D. L. Littel, D. S., & McPharman, A. C. (2011). Expression and function of muscle time in chesity, disketen
315 316 317 318	 Allen, D. L., Hittel, D. S., & McPherron, A. C. (2011). Expression and function of myostatin in obesity, diabetes, and exercise adaptation. <i>Medicine and science in sports and exercise</i>, 43(10), 1828. Amthor, H., Nicholas, G., McKinnell, I., Kemp, C. F., Sharma, M., Kambadur, R., & Patel, K. (2004). Follistatin complexes Myostatin and antagonises Myostatin-mediated inhibition of myogenesis. <i>Developmental</i>
319 320 321 322 323	 biology, 270(1), 19-30. Ashtary-Larky, D., Daneghian, S., Alipour, M., Rafiei, H., Ghanavati, M., Mohammadpour, R., Afrisham, R. (2018). Waist Circumference to Height Ratio: Better Correlation with Fat Mass Than Other Anthropometric Indices During Dietary Weight Loss in Different Rates. International journal of endocrinology and metabolism, 16(4).
324 325 326	Ashtary-Larky, D., Ghanavati, M., Lamuchi-Deli, N., Payami, S. A., Alavi-Rad, S., Boustaninejad, M., Alipour, M. (2017). Rapid weight loss vs. slow weight loss: which is more effective on body composition and metabolic risk factors? <i>International journal of endocrinology and metabolism</i> , 15(3).
327 328 329	Ashtary Larky, D., Bagheri, R., Abbasnezhad, A., & M Tinsley, G. (2020). Effects of gradual weight loss vs rapid weight loss on body composition and resting metabolic rate: A systematic review and meta-analysis. <i>British Journal of Nutrition.</i>
330 331 332	 Astrup, A., & Rössner, S. (2000). Lessons from obesity management programmes: greater initial weight loss improves long-term maintenance. <i>obesity reviews</i>, 1(1), 17-19. Bagheri, R., Moghadam, B. H., Jo, E., Tinsley, G. M., Stratton, M. T., Larky, D. A., Wong, A. (2020).
333 334 335	 Digheri, R., Roghadani, D. H., 50, E., Thisey, O. W., Statton, M. T., Earky, D. A., T. Wong, A. (2020). Comparison of whole egg vs. egg white ingestion during 12 weeks of resistance training on skeletal muscle regulatory markers in resistance-trained men. <i>British Journal of Nutrition</i>, 1-20. Bagheri, R., Rashidlamir, A., Ashtary-Larky, D., Wong, A., Alipour, M., Motevalli, M. S., Zouhal, H. (2019).
336 337	Does Green Tea Extract Enhance the Anti-inflammatory Effects of Exercise on Fat Loss? <i>British journal of clinical pharmacology</i> .
338 339 340	Bagheri, R., Rashidlamir, A., Motevalli, M. S., Elliott, B. T., Mehrabani, J., & Wong, A. (2019). Effects of upper- body, lower-body, or combined resistance training on the ratio of follistatin and myostatin in middle-aged men. <i>European journal of applied physiology</i> , 1-11.
341 342 343	Brown, M. L., Bonomi, L., Ungerleider, N., Zina, J., Kimura, F., Mukherjee, A., Schneyer, A. (2011). Follistatin and follistatin like-3 differentially regulate adiposity and glucose homeostasis. <i>Obesity</i> , <i>19</i> (10), 1940-1949.
344 345 346	 Carels, R. A., Cacciapaglia, H. M., Douglass, O. M., Rydin, S., & O'Brien, W. H. (2003). The early identification of poor treatment outcome in a women's weight loss program. <i>Eating Behaviors</i>, 4(3), 265-282. Cohen, J. (1992). A power primer. <i>Psychological bulletin</i>, 112(1), 155.
347 348 349	 Elfhag, K., & Rössner, S. (2005). Who succeeds in maintaining weight loss? A conceptual review of factors associated with weight loss maintenance and weight regain. <i>obesity reviews</i>, 6(1), 67-85. Elliott, B., Renshaw, D., Getting, S., & Mackenzie, R. (2012). The central role of myostatin in skeletal muscle and
350 351 352 353 354	 whole body homeostasis. <i>Acta physiologica</i>, 205(3), 324-340. Faul, F., Erdfelder, E., Lang, AG., & Buchner, A. (2007). G* Power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. <i>Behavior research methods</i>, 39(2), 175-191. Flanagan, J. N., Linder, K., Mejhert, N., Dungner, E., Wahlen, K., Decaunes, P., Bhasin, S. (2009). Role of follistatin in promoting adipogenesis in women. <i>The Journal of Clinical Endocrinology & Metabolism</i>,
355	<i>94</i> (8), 3003-3009.

- Floegel, A., & Pischon, T. (2012). Low carbohydrate-high protein diets. In: British Medical Journal Publishing
 Group.
- Freire, R. (2020). Scientific evidence of diets for weight loss: Different macronutrient composition, intermittent
 fasting, and popular diets. *Nutrition, 69*, 110549.
- Gonzalez-Cadavid, N. F., Taylor, W. E., Yarasheski, K., Sinha-Hikim, I., Ma, K., Ezzat, S., ... Mamita, M. (1998).
 Organization of the human myostatin gene and expression in healthy men and HIV-infected men with muscle wasting. *Proceedings of the National Academy of Sciences*, *95*(25), 14938-14943.
- Görgens, S. W., Raschke, S., Holven, K. B., Jensen, J., Eckardt, K., & Eckel, J. (2013). Regulation of follistatin like protein 1 expression and secretion in primary human skeletal muscle cells. *Archives of physiology and biochemistry*, 119(2), 75-80.
- Hill, J. O. (2008). Can a small-changes approach help address the obesity epidemic? A report of the Joint Task
 Force of the American Society for Nutrition, Institute of Food Technologists, and International Food
 Information Council. *The American journal of clinical nutrition*, 89(2), 477-484.
- Hittel, D. S., Berggren, J. R., Shearer, J., Boyle, K., & Houmard, J. A. (2009). Increased secretion and expression of myostatin in skeletal muscle from extremely obese women. *Diabetes*, 58(1), 30-38.
- Jackson, A., Pollock, M. L., Graves, J. E., & Mahar, M. (1988). Reliability and validity of bioelectrical impedance
 in determining body composition. *Journal of applied physiology*, 64(2), 529-534.
- Jakicic, J. M., Clark, K., Coleman, E., Donnelly, J. E., Foreyt, J., Melanson, E., . . . Volpe, S. L. (2001). Appropriate
 intervention strategies for weight loss and prevention of weight regain for adults. *Medicine & Science in Sports & Exercise*, *33*(12), 2145-2156.
- Kraemer, W. J., Marchitelli, L., Gordon, S. E., Harman, E., Dziados, J. E., Mello, R., . . . Fleck, S. J. (1990).
 Hormonal and growth factor responses to heavy resistance exercise protocols. *Journal of applied physiology*, 69(4), 1442-1450.
- 279 Laplante, M., & Sabatini, D. M. (2009). mTOR signaling at a glance. *Journal of cell science*, *122*(20), 3589-3594.
- Ling, C. H., de Craen, A. J., Slagboom, P. E., Gunn, D. A., Stokkel, M. P., Westendorp, R. G., & Maier, A. B.
 (2011). Accuracy of direct segmental multi-frequency bioimpedance analysis in the assessment of total
 body and segmental body composition in middle-aged adult population. *Clinical nutrition*, 30(5), 610 615.
- Lutes, L. D., Winett, R. A., Barger, S. D., Wojcik, J. R., Herbert, W. G., Nickols-Richardson, S. M., & Anderson,
 E. S. (2008). Small changes in nutrition and physical activity promote weight loss and maintenance: 3 month evidence from the ASPIRE randomized trial. *Annals of Behavioral Medicine*, *35*(3), 351-357.
- Maïmoun, L., Mura, T., Attalin, V., Dupuy, A. M., Cristol, J.-P., Avignon, A., . . . Sultan, A. (2020). Modification
 of Muscle-Related Hormones in Women with Obesity: Potential Impact on Bone Metabolism. *Journal of Clinical Medicine*, 9(4), 1150.
- Matsakas, A., Foster, K., Otto, A., Macharia, R., Elashry, M. I., Feist, S., ... Walsh, F. (2009). Molecular, cellular
 and physiological investigation of myostatin propeptide-mediated muscle growth in adult mice.
 Neuromuscular Disorders, 19(7), 489-499.
- 393 McPherron, A. C., & Lee, S.-J. (1997). Double muscling in cattle due to mutations in the myostatin gene.
 394 *Proceedings of the National Academy of Sciences*, 94(23), 12457-12461.
- 395 Middelbeek, L., & Breda, J. (2013). Obesity and sedentarism: Reviewing the current situation within the WHO
 396 European Region. *Current Obesity Reports*, 2(1), 42-49.
- Milan, G., Dalla Nora, E., Pilon, C., Pagano, C., Granzotto, M., Manco, M., . . . Vettor, R. (2004). Changes in muscle myostatin expression in obese subjects after weight loss. *The Journal of Clinical Endocrinology* & *Metabolism*, 89(6), 2724-2727.
- Motevalli, M. S., Dalbo, V. J., Attarzadeh, R. S., Rashidlamir, A., Tucker, P. S., & Scanlan, A. T. (2015). The
 effect of rate of weight reduction on serum myostatin and follistatin concentrations in competitive
 wrestlers. *International journal of sports physiology and performance*, 10(2), 139-146.
- 403 Nackers, L. M., Ross, K. M., & Perri, M. G. (2010). The association between rate of initial weight loss and long 404 term success in obesity treatment: does slow and steady win the race? *International journal of behavioral* 405 *medicine*, 17(3), 161-167.
- 406 Nakatani, M., Kokubo, M., Ohsawa, Y., Sunada, Y., & Tsuchida, K. (2011). Follistatin-derived peptide expression
 407 in muscle decreases adipose tissue mass and prevents hepatic steatosis. *American Journal of Physiology-* 408 *Endocrinology and Metabolism, 300*(3), E543-E553.
- Park, J.-J., Berggren, J. R., Hulver, M. W., Houmard, J. A., & Hoffman, E. P. (2006). GRB14, GPD1, and GDF8
 as potential network collaborators in weight loss-induced improvements in insulin action in human
 skeletal muscle. *Physiological genomics*, 27(2), 114-121.
- 412 Patel, V. K., & Demontis, F. (2014). GDF11/myostatin and aging. Aging (Albany NY), 6(5), 351.
- Peos, J. J., Norton, L. E., Helms, E. R., Galpin, A. J., & Fournier, P. (2019). Intermittent dieting: theoretical considerations for the athlete. *Sports*, 7(1), 22.

- Reisz-Porszasz, S., Bhasin, S., Artaza, J. N., Shen, R., Sinha-Hikim, I., Hogue, A., . . . Gonzalez-Cadavid, N. F.
 (2003). Lower skeletal muscle mass in male transgenic mice with muscle-specific overexpression of
 myostatin. American Journal of Physiology-Endocrinology and Metabolism, 285(4), E876-E888.
- 418 Sbrocco, T., Nedegaard, R. C., Stone, J. M., & Lewis, E. L. (1999). Behavioral choice treatment promotes
 419 continuing weight loss: Preliminary results of a cognitive-behavioral decision-based treatment for
 420 obesity. *Journal of Consulting and Clinical Psychology*, 67(2), 260.
- Schoeller, D. A. (1995). Limitations in the assessment of dietary energy intake by self-report. *Metabolism*, 44, 18 22.
- Shirali, S., Daneghian, S., Hosseini, S. A., Ashtary-Larky, D., Daneghian, M., & Mirlohi, M.-S. (2016). Effect of caffeine co-ingested with carnitine on weight, body-fat percent, serum leptin and lipid profile changes in male teen soccer players: A randomized clinical trial. *International Journal of Pediatrics*, 4(10), 3685-3698.
- Table, M. (2005). *Dietary reference intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein, and amino acids* (Vol. 5): National Academy Press: Washington, DC, USA.
- Tinsley, G. M., & Willoughby, D. S. (2016). Fat-free mass changes during ketogenic diets and the potential role of resistance training. *International journal of sport nutrition and exercise metabolism*, 26(1), 78-92.
- Tsuchida, K., Arai, K. Y., Kuramoto, Y., Yamakawa, N., Hasegawa, Y., & Sugino, H. (2000). Identification and
 characterization of a novel follistatin-like protein as a binding protein for the TGF-β family. *Journal of Biological Chemistry*, 275(52), 40788-40796.
- Vink, R. G., Roumans, N. J., Arkenbosch, L. A., Mariman, E. C., & van Baak, M. A. (2016). The effect of rate of weight loss on long-term weight regain in adults with overweight and obesity. *Obesity*, 24(2), 321-327.
- Wagner, K. R., Liu, X., Chang, X., & Allen, R. E. (2005). Muscle regeneration in the prolonged absence of
 myostatin. *Proceedings of the National Academy of Sciences*, 102(7), 2519-2524.
- Wagner, K. R., McPherron, A. C., Winik, N., & Lee, S. J. (2002). Loss of myostatin attenuates severity of muscular dystrophy in mdx mice. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*, 52(6), 832-836.
- Wheeler, M., Daly, A., & Evert, A. (2014). Choose your foods: Food lists for diabetes. *Chicago, IL: Academy of Nutrition and Dietetics/American Diabetes Association*.
- Wiewiora, M., Mertas, A., Gluck, M., Nowowiejska-Wiewiora, A., Czuba, Z., & Piecuch, J. (2020). Effect of
 Weight Loss Surgery on Biomarkers of Angiogenesis in Obese Patients. *Obesity Surgery*, 1-9.
- Yancy, W. S., Olsen, M. K., Guyton, J. R., Bakst, R. P., & Westman, E. C. (2004). A low-carbohydrate, ketogenic diet versus a low-fat diet to treat obesity and hyperlipidemia: a randomized, controlled trial. *Annals of internal medicine*, 140(10), 769-777.
- Yang, J., Ratovitski, T., Brady, J. P., Solomon, M. B., Wells, K. D., & Wall, R. J. (2001). Expression of myostatin
 pro domain results in muscular transgenic mice. *Molecular Reproduction and Development: Incorporating Gamete Research*, 60(3), 351-361.
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458

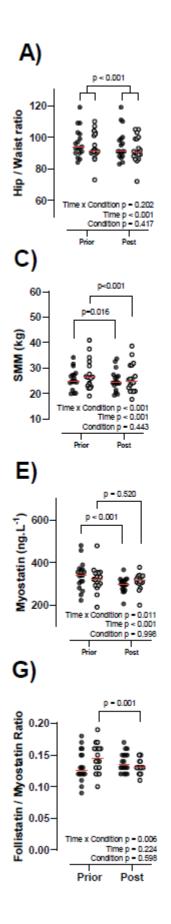
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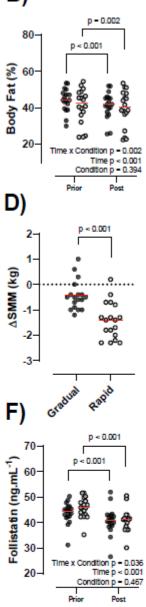
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464 Figure legends:







- 467 Figure 1: Alterations in Body Composition and endocrine markers with dietary condition. A) Hip / Waist ratio,
- **B** BFP (%), **C**) SMM (kg) as a function of time (prior or post). **D**) ΔSMM (kg) by condition (GWL or RWL), **E**)
- 469 Myostatin (ng.L⁻¹), **F**) Follistatin (ng.mL⁻¹), **G**) Follistatin/Myostatin ratio. Closed black circles indicate GWL and
- 470 open grey circles indicate RWL. Horizontal red lines indicate condition means. Statistical comparisons and p
- 471 values as indicated. N = 18 per condition and time point.
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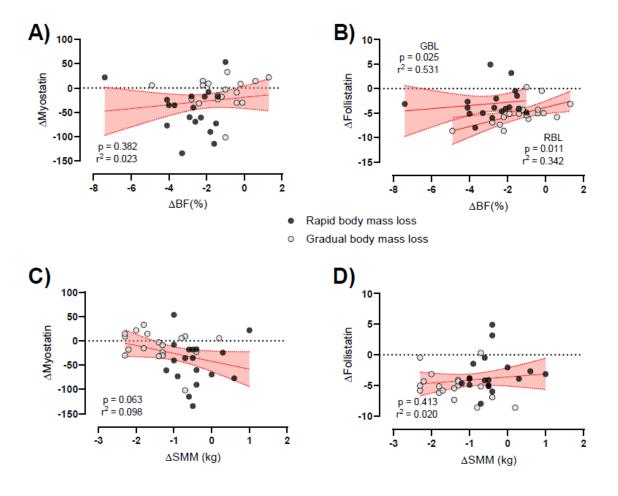




Figure 2: Correlations between change in (Δ) Endocrine marker and Δ body composition. **A**) Δ Myostatin, **B**) Δ Follistatin as a function of Δ BFP, **C**) Δ Myostatin, **D**) Δ Follistatin as a function of Δ SMM (kg). Closed black circles indicate RWL and open grey circles indicate GWL. Solid red line indicates linear regression; shaded red zone indicates 95% confidence interval. Figures **A**), **C**), and **D**) treated as one condition (sum-of-squares F test **A**) **p** = 0.505, **C**) **p** = 0.177), and **D**) **p** = 0.063), **Figure B**) treated as two distinct conditions (**p** = 0.013).