### University of Massachusetts Amherst ScholarWorks@UMass Amherst

Microbiology Department Faculty Publication Series

Microbiology

2020

# Draft Genome Sequence of a Terrestrial Planctomycete, *Singulisphaera* sp. Strain GP 187, Isolated from Forest Soil

Maureen A. Morrow

Grace Pold

Kristen M. DeAngelis

Follow this and additional works at: https://scholarworks.umass.edu/micro\_faculty\_pubs



## Draft Genome Sequence of a Terrestrial Planctomycete, *Singulisphaera* sp. Strain GP187, Isolated from Forest Soil

**Microbiology**<sup>®</sup>

**Resource Announcements** 

Maureen A. Morrow,<sup>a</sup> Grace Pold,<sup>b</sup> Kristen M. DeAngelis<sup>c</sup>

AMERICAN SOCIETY FOR

MICROBIOLOGY

<sup>a</sup>Department of Biology, State University of New York at New Paltz, New Paltz, New York, USA <sup>b</sup>Department of Natural Resources Management and Environmental Sciences, California Polytechnic State University, San Luis Obispo, California, USA <sup>c</sup>Department of Microbiology, University of Massachusetts, Amherst, Massachusetts, USA

**ABSTRACT** Here, we present the draft genome sequence of a novel species of the genus *Singulisphaera* (phylum *Planctomycetes*, family *Isosphaeraceae*) isolated from soil. *Singulisphaera* sp. strain GP187 has a relatively large mobilome and numerous novel genes that may contribute to the production of bioactive molecules.

Culture-independent analysis reveals that *Planctomycetes* is the fifth most abundant bacterial phylum in global soil samples (1), yet this phylum remains underrepresented in axenic cultures, and a large majority of these cultures are derived from aqueous environments (2). Aqueous planctomycetes are hypothesized to have evolved from terrestrial species (3).

*Singulisphaera* sp. strain GP187 was isolated on 3 June 2014 from the Harvard Forest, a temperate forest ecosystem in Petersham, MA (42.54°N, -72.18°W). Organic horizon soil was pretreated with 6% yeast extract plus 0.05% SDS (4), plated onto oat-meal medium, and incubated aerobically at 20°C, with colonies appearing after 8 days. GP187 was the only *Planctomycetes* strain of the hundreds of isolates from this site (5) and thus was subjected to whole-genome sequencing.

GP187 was grown aerobically on Reasoner's 2A (R2A) medium (pH 7) (6). Genomic DNA was purified using a modified cetyltrimethylammonium bromide (CTAB) procedure (7) but was not sheared or size selected. The draft genome sequence was generated at the DOE Joint Genome Institute (JGI). A PacBio SMRTbell library was constructed and sequenced on the PacBio RS platform, generating 407,937 reads ( $N_{50}$ , 3.6 kbp). The filtered raw reads (675.3 Mbp) were assembled using HGAP v2.3 \_p5 (protocol version, 2.3.0; method, RS HGAP Assembly.3, smrtpipe.py v1.87.139483) (8). The final draft assembly contained 5 contigs in 5 scaffolds ( $N_{50}$ , 6.278 Mbp), estimated as 99.61% complete and 5.81% contaminated using CheckM v1.0.18 (9) in KBase (10). The input read sequencing depth was 72.8×. Gene annotations were completed within the JGI's Integrated Microbial Genomes (IMG) platform (11) and KBase. Default parameters were used for all software.

The genome is 10,689,158 bp (G+C content, 63.07%) and is predicted to encode 8,388 proteins (36.6% without predicted function), 8 rRNA operons, and 110 RNA genes (24 rRNAs, 64 tRNAs). GP187 has the largest genome of cultured *lsosphaeraceae* strains and the second largest genome of cultured *Planctomycetes* strains (12).

Phylogenetically, the closest species to GP187 is the aquatic *Singulisphaera acidiphila* DSM 18658 (13). These strains share 98.84% homology for 16S rRNA genes (average as determined by searching public RNA isolates with IMG BLAST) and 86.7% whole-genome average nucleotide identity (ANI) (IMG pairwise ANI). *Isosphaeraceae* genome sequences characteristically carry large plasmids; GP187 harbors a putative plasmid of 63.8 kb (G+C content, 61.8%; a lower G+C content is typical of plasmids [14]). The subtilisin gene open reading frame spans, without gaps, the ends of contig5

Citation Morrow MA, Pold G, DeAngelis KM. 2020. Draft genome sequence of a terrestrial planctomycete, *Singulisphaera* sp. strain GP187, isolated from forest soil. Microbiol Resour Announc 9:e00956-20. https://doi.org/10.1128/ MRA.00956-20.

**Editor** Julie C. Dunning Hotopp, University of Maryland School of Medicine

**Copyright** © 2020 Morrow et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Maureen A. Morrow, morrowm@newpaltz.edu.

Received 13 August 2020 Accepted 18 November 2020 Published 10 December 2020

		No. (%) of:			
Strain	Genome size (Mbp)	Protein coding genes <sup>a</sup>	Genes without predicted function <sup>a</sup>	Giant genes without predicted function <sup>6</sup>	Genes without predicted function in BGCs <sup>c</sup>
S. acidiphila DSM 18658	9.76	7,576 (98.6)	2,578 (33.6)	7 (20.6)	76 (29.8)
S. acidiphila sp. GP187	10.69	8,388 (98.7)	3,110 (36.6)	14 (35.0)	104 (36.4)

TABLE 1 Differences in select characteristics between S. acidiphila DSM 18658 and S. acidiphila sp. GP187

<sup>a</sup> Percentage of total protein coding genes (IMG).

<sup>b</sup> Percentage of giant genes (KBase).

<sup>c</sup> Percentage of total genes in biosynthetic gene clusters (BCGs) (IMG).

and shares 82% homology with the plasmid-encoded subtilisin gene of *S. acidiphila* DSM 18658, suggesting that contig5 is a circular plasmid.

GP187 contains 60% more mobilome-associated genes (186 [2.2% of proteinencoding genes] versus 117 [1.5%], respectively) and 78% more genomic islands (41 versus 23, respectively) than *S. acidiphila* DSM 18658, as predicted by IslandViewer4 (15). GP187 has a greater potential to synthesize specialized metabolites, given that novel genes of this species are categorized as giant genes ( $\leq$ 5,000 bp with Kbase RAST annotation) (16) or are located in biosynthesis gene clusters (17) more often than in *S. acidiphila* DSM 18658 (Table 1).

This genome sequence will contribute to our understanding of terrestrial species of *Planctomycetes*, a phylum abundant in soil but underrepresented in isolate genome analysis. Analysis of this genome sequence may elucidate its ecological role in terrestrial ecosystems, identify evolutionary relationships between terrestrial and aquatic *Singulisphaera* species, and contribute to the discovery of novel secondary metabolites.

**Data availability.** This whole-genome sequence was deposited at DDBJ/EMBL/ GenBank under the accession number NZ\_FSRB00000000.1. The raw data were deposited in the JGI GOLD under the project number Gp0151081 and in the Sequence Read Archive under the accession number SRX2158412. The JGI annotation is found at https:// img.jgi.doe.gov/cgi-bin/m/main.cgi?section=TaxonDetail&page=taxonDetail&taxon\_oid= 2695420965#.

### **ACKNOWLEDGMENTS**

This work was supported by the National Science Foundation under contract DEB-1749206 (K.M.D. and M.A.M.) and the DOE Genomic Sciences Program under contract DE-SC0016590 (K.M.D.). DNA sequencing and library preparation were supported by the U.S. Department of Energy Joint Genome Institute (JGI) Community Science Program, project CSP506489, under K.M.D. JGI, a DOE Office of Science User Facility, is supported by the Office of Science of the U.S. Department of Energy under contract DE-AC02-05CH11231.

We thank Mallory Choudoir for helpful comments on the manuscript and Daniel Freedman for help with the statistical analysis.

#### REFERENCES

- Delgado-Baquerizo M, Oliverio AM, Brewer TE, Benavent-González A, Eldridge DJ, Bardgett RD, Maestre FT, Singh BK, Fierer N. 2018. A global atlas of the dominant bacteria found in soil. Science 359:320–325. https:// doi.org/10.1126/science.aap9516.
- Wiegand S, Jogler M, Boedeker C, Pinto D, Vollmers J, Rivas-Marín E, Kohn T, Peeters SH, Heuer A, Rast P, Oberbeckmann S, Bunk B, Jeske O, Meyerdierks A, Storesund JE, Kallscheuer N, Lücker S, Lage OM, Pohl T, Merkel BJ, Hornburger P, Müller R-W, Brümmer F, Labrenz M, Spormann AM, Op den Camp HJM, Overmann J, Amann R, Jetten MSM, Mascher T, Medema MH, Devos DP, Kaster A-K, Øvreås L, Rohde M, Galperin MY, Jogler C. 2020. Cultivation and functional characterization of 79 planctomycetes uncovers their unique biology. Nat Microbiol 5:126–140. https:// doi.org/10.1038/s41564-019-0588-1.
- Andrei A-Ş, Salcher MM, Mehrshad M, Rychtecký P, Znachor P, Ghai R. 2019. Niche-directed evolution modulates genome architecture in

freshwater Planctomycetes. ISME J 13:1056–1071. https://doi.org/10.1038/ s41396-018-0332-5.

- Hayakawa M, Nonomura H. 1989. A new method for the intensive isolation of actinomycetes from soil. Actinomycetologica 3:95–104. https://doi .org/10.3209/saj.3\_95.
- Pold G, Billings AF, Blanchard JL, Burkhardt DB, Frey SD, Melillo JM, Schnabel J, van Diepen LTA, DeAngelis KM. 2016. Long-term warming alters carbohydrate degradation potential in temperate forest soils. Appl Environ Microbiol 82:6518–6530. https://doi.org/10.1128/AEM .02012-16.
- van der Linde K, Lim BT, Rondeel JMM, Antonissen LPMT, de Jong GMT. 1999. Improved bacteriological surveillance of haemodialysis fluids: a comparison between Tryptic soy agar and Reasoner's 2A media. Nephrol Dial Transplant 14:2433–2437. https://doi.org/10.1093/ndt/14.10.2433.
- 7. Joint Genome Institute. 2012. Bacterial genomic DNA isolation using CTAB.

Joint Genome Institute, Walnut Creek, CA. https://jgi.doe.gov/wp-content/uploads/2014/02/JGI-Bacterial-DNA-isolation-CTAB-Protocol-2012.pdf.

- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10.1038/nmeth.2474.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. https:// doi.org/10.1101/gr.186072.114.
- Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, Sneddon MW, Henderson ML, Riehl WJ, Murphy-Olson D, Chan SY, Kamimura RT, Kumari S, Drake MM, Brettin TS, Glass EM, Chivian D, Gunter D, Weston DJ, Allen BH, Baumohl J, Best AA, Bowen B, Brenner SE, Bun CC, Chandonia J-M, Chia J-M, Colasanti R, Conrad N, Davis JJ, Davison BH, DeJongh M, Devoid S, Dietrich E, Dubchak I, Edirisinghe JN, Fang G, Faria JP, Frybarger PM, Gerlach W, Gerstein M, Greiner A, Gurtowski J, Haun HL, He F, Jain R, Joachimiak MP, Keegan KP, Kondo S, et al. 2018. KBase: the United States Department of Energy Systems Biology Knowledgebase. Nat Biotechnol 36:566–569. https://doi.org/ 10.1038/nbt.4163.
- 11. Chen I-MA, Chu K, Palaniappan K, Pillay M, Ratner A, Huang J, Huntemann M, Varghese N, White JR, Seshadri R, Smirnova T, Kirton E, Jungbluth SP, Woyke T, Eloe-Fadrosh EA, Ivanova NN, Kyrpides NC. 2019. IMG/M v.5.0: an integrated data management and comparative analysis system for mi-

- Ravin NV, Rakitin AL, Ivanova AA, Beletsky AV, Kulichevskaya IS, Mardanov AV, Dedysh SN. 2018. Genome analysis of Fimbriiglobus ruber SP5(T), a planctomycete with confirmed chitinolytic capability. Appl Environ Microbiol 84:e02645-17. https://doi.org/10.1128/AEM.02645-17.
- Guo M, Han X, Jin T, Zhou L, Yang J, Li Z, Chen J, Geng B, Zou Y, Wan D, Li D, Dai W, Wang H, Chen Y, Ni P, Fang C, Yang R. 2012. Genome sequences of three species in the family *Planctomycetaceae*. J Bacteriol 194:3740–3741. https://doi.org/10.1128/JB.00639-12.
- Nishida H. 2012. Comparative analyses of base compositions, DNA sizes, and dinucleotide frequency profiles in archaeal and bacterial chromosomes and plasmids. Int J Evol Biol 2012:342482.
- Bertelli C, Laird MR, Williams KP, Simon Fraser University Research Computing Group, Lau BY, Hoad G, Winsor GL, Brinkman FSL. 2017. Island-Viewer 4: expanded prediction of genomic islands for larger-scale datasets. Nucleic Acids Res 45:W30–W35. https://doi.org/10.1093/nar/gkx343.
- Reva O, Tümmler B. 2008. Think big—giant genes in bacteria. Environ Microbiol 10:768–777. https://doi.org/10.1111/j.1462-2920.2007.01500.x.
- 17. Hadjithomas M, Chen I-MA, Chu K, Ratner A, Palaniappan K, Szeto E, Huang J, Reddy TBK, Cimermančič P, Fischbach MA, Ivanova NN, Markowitz VM, Kyrpides NC, Pati A. 2015. IMG-ABC: a knowledge base to fuel discovery of biosynthetic gene clusters and novel secondary metabolites. mBio 6:e00932-15. https://doi.org/10.1128/mBio.00932-15.