

1 Estimation of sinking velocities using free-falling dynamically scaled  
2 models: foraminifera as a test case

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12 [Summary Statement](#)

13 We developed a novel method to determine the sinking velocity of biologically important micro-  
14 scale particles using 3D printed scale models.

## 15 Abstract

16 The velocity of settling particles is an important determinant of distribution in extinct and extant  
17 species with passive dispersal mechanisms, such as plants, corals, and phytoplankton. Here we adapt  
18 dynamic scaling, borrowed from engineering, to determine settling velocities. Dynamic scaling  
19 leverages physical models with relevant dimensionless numbers matched to achieve similar dynamics  
20 to the original object. Previous studies have used flumes, wind tunnels, or towed models to examine  
21 fluid flows around objects with known velocities. Our novel application uses free-falling models to  
22 determine the unknown sinking velocities of planktonic foraminifera – organisms important to our  
23 understanding of the Earth’s current and historic climate. Using enlarged 3D printed models of  
24 microscopic foraminifera tests, sunk in viscous mineral oil to match their Reynolds numbers and drag  
25 coefficients, we predict sinking velocities of real tests in seawater. This method can be applied to study  
26 other settling particles such as plankton, spores, or seeds.

## 27 Introduction

28 The transport of organisms and biologically derived particles through fluid environments strongly  
29 influences their spatiotemporal distributions and ecology. In up to a third of terrestrial plants (Willson  
30 et al., 1990), reproduction is achieved through passive movement of propagules (e.g., seeds) on the  
31 wind. In aquatic environments, propagules of many sessile groups from corals (Jones et al., 2015) to  
32 bivalves (Booth, 1983) are dispersed by ambient currents, eventually settling out of the water column  
33 to their final locations. Furthermore, most dead aquatic organisms (from diatoms to whales) sink,  
34 transporting nutrients to deeper water and contributing to long term storage of carbon (De La Rocha  
35 & Passow, 2007). In the case of microfossils, sinking dynamics of the original organisms even  
36 influences our reconstructions of the Earth’s paleoclimate (Van Sebille et al., 2015). Crucially, the  
37 horizontal distances over which all these biological entities are transported, and therefore their  
38 distributions, are affected by their settling velocities (Ali et al., 2011).

39 Measuring the individual settling velocities of small particles directly is challenging, especially when  
40 they are too small to be imaged easily without magnification (e.g. Walsby and Holland, 2006). Here  
41 we apply dynamic scaling, an approach commonly used in engineering, to circumvent this difficulty  
42 and accurately quantify the kinematics of sub-millimeter scale free-falling particles using enlarged  
43 physical models. We use scaled-up physical models in a high-viscosity fluid, enabling easy  
44 measurements of settling speed, orientation, and other parameters using inexpensive standard high-  
45 definition web cameras. While dynamically scaled models have previously been employed to study a  
46 number of problems in biological fluid mechanics (e.g. Vogel, Ellington and Kilgore, 1973; Vogel, 1987;  
47 Vogel, 1994; Koehl, 2003), the study of freely-falling particles of complex shape – for which settling  
48 speed is the key unknown parameter – presents a unique challenge to experimental design that we  
49 overcome in this work.

50 Engineering problems such as aircraft and submarine design often are approached using scaled-down  
51 models in wind tunnels or flumes to examine fluid flows around the model and the resulting fluid  
52 dynamic forces it is subjected to. To ensure that the behaviour of the model system is an accurate

53 representation of real life, similarity of relevant physical phenomena must be maintained between  
54 the two. If certain dimensionless numbers (i.e., ratios of physical quantities such that all dimensional  
55 units cancel) that describe the system are equal between the life-size original and the scaled-down  
56 model, “similitude” is achieved and all parameters of interest (e.g., velocities and forces) will be  
57 proportional between prototype and model (Zohuri, 2015). Intuitively, the model and real object must  
58 be geometrically similar (i.e., have the same shape), so that the dimensionless ratio of any length  
59 between model and original,  $Length_{model}/Length_{real}$ , is constant – this is the scale factor ( $S$ ) of the  
60 model. Less obvious is the additional requirement of dynamic similarity, signifying that the ratios of  
61 all relevant forces are constant. For completely immersed objects sinking steadily at terminal velocity  
62 (achieved quickly for most small particles, see Time to Terminal Velocity), dynamic similarity is  
63 achieved by matching the Reynolds number ( $Re$ ).

64  $Re$  is a measure of the ratio of inertial to viscous forces in the flow (Batchelor, 2000; within a biological  
65 context Vogel, 1994), and is typically defined as:

66 1

67 
$$Re = \frac{L U \rho_{fluid}}{\mu}$$

68 where  $\rho_{fluid}$  is density of the fluid ( $\text{kg m}^{-3}$ );  $L$  is a characteristic length (m) of the object;  $U$  is the  
69 object’s velocity ( $\text{m s}^{-1}$ ); and  $\mu$  is the dynamic viscosity ( $\text{N s m}^{-2}$ , or  $\text{Pa s}$ ) of the fluid. In cases where  
70  $L U \rho_{fluid}$  is large compared to  $\mu$ , e.g. whales, birds, and fish ( $Re \approx 3 \times 10^9 - 3 \times 10^6$ , Vogel, 1994),  
71 inertial forces dominate. In cases where  $L U \rho_{fluid}$  is relatively small compared to  $\mu$ , e.g. sperm,  
72 bacteria ( $Re \approx 3 \times 10^{-2} - 1 \times 10^{-5}$ ), viscous forces dominate. Finally, when  $L U \rho_{fluid}$  is of comparable  
73 magnitude to  $\mu$ ,  $Re$  is intermediate and one cannot discount either inertial or viscous forces. If the  
74 scaled model and original system exhibit identical  $Re$ , the relative importance of inertial versus viscous  
75 forces is matched between the two and any qualitative features of the flows (e.g. streamlines) will  
76 also be identical.

77 Dynamically scaled physical models exhibiting the same  $Re$  as the original systems have been used in  
78 a number of biological studies. Vogel and La Barbera (1978) outline the principles of dynamic scaling:  
79 to obtain the same  $Re$  when enlarging small organisms, the fluid flow must be slower and/or the fluid  
80 more viscous, and when making smaller models of large organisms, the fluid flow must be faster  
81 and/or the fluid less viscous. For instance, Vogel (1987) used air in place of water flowing at lower  
82 speeds when investigating the refilling of the squids mantle during swimming by scaling a model up  
83 1.5 times relative to the animal’s actual size. More recently, Stadler et al (2016) investigated sand  
84 inhalation in skinks with 3D-printed enlarged models, using helium instead of air (thereby increasing  
85 viscosity) as the experimental fluid. Koehl and colleagues have studied crustacean antennule flicking  
86 (lobsters (Reidenbach et al., 2008), mantis shrimp (Stacey et al., 2002) and crabs (Waldrop et al.,  
87 2015)) as well as the movements of copepod appendages (Koehl, 1995) with enlarged models, using  
88 mineral oil in place of water. Finally, perhaps the largest change in scale was employed by Kim et al  
89 (2003), who modelled the bundling of *E. coli* flagella at a scale factor of  $\sim 61,000$ , submerged in silicone

90 oil ( $10^5$  times more viscous than water), and rotated at 0.002 rpm compared to the 600 rpm observed  
91 in real bacteria (Sowa & Berry, 2008).

92 In all the above studies, basic kinematics such as speeds in the original system were relatively easy to  
93 measure, and the experiments aimed to reveal the forces involved (e.g. hydrodynamic drag) or details  
94 of the fluid flow such as the pattern of streamlines. Since the representative speed  $U$  of the original  
95 system was known, designing experiments to achieve similitude was relatively straightforward  
96 because the  $Re$  was also known *a priori* – in these cases, the model size, speed, and working fluid  
97 properties were simply interrelated through  $Re$  (Eqn 1). For instance, once a working fluid and the  
98 model size were chosen, the required towing speed was obvious. However, in the case of  
99 sedimentation of small particles (e.g. spores, seeds, plankton), the sinking speed ( $U$ ) is the key  
100 unknown. With an unknown sinking speed, the operating  $Re$  is also unknown, so it is not  
101 straightforward to design experiments that achieve similitude with the original system. Here we  
102 present an iterative methodology leveraging 3D printed dynamically scaled models that allows  
103 determination of the sinking speeds of small objects of arbitrarily complex shape.

104 We use planktonic foraminifera as an example of a small (200 – 1500  $\mu\text{m}$ ) biological particle for which  
105 the settling velocity is important and typically unknown. Foraminifera are a phylum of marine ameboid  
106 protists (B. K. Sen Gupta (ed.) , 2002; Schiebel & Hemleben, 2005). By secreting calcium carbonate,  
107 foraminifera produce a multi-chambered shell (test) which, in planktonic foraminifera, can grow up to  
108 1500  $\mu\text{m}$  in diameter, and which frequently exhibits complex shape (Table 1). Once the organism dies  
109 or undergoes reproduction, the empty test sinks to the ocean floor, and so oceanic sediments contain  
110 substantial numbers of foraminifera tests. Foraminifera account for 23-56% of the oceans production  
111 of carbonate ( $\text{CO}_3$ ) (Schiebel, 2002), an important factor in climate change models (Passow & Carlson,  
112 2012). Of particular interest for climate predictions is calculating the flux of tests reaching the ocean  
113 floor (Schiebel, 2002; Jonkers & Kučera, 2015). While there are more than 30 extant species and over  
114 600 species in the fossil record, settling velocities are known for only 14 species of foraminifera (Fok-  
115 Pun & Komar, 1983; Takahashi & Be, 1984; Caromel et al., 2014,  $3.41 \times 10^{-4}$  -  $6.8 \times 10^{-2} \text{ m s}^{-1}$ ,  $Re \approx 18$  –  
116 55).

## 117 Materials and Methods

### 118 Similitude and Settling Theory

119 We assume that the size (i.e.,  $L$  – defined as the maximum length parallel to the settling direction,  $A$   
120 – defined as the projected frontal area, and  $V$  – the particle volume not including any fluid-filled  
121 cavities), 3D shape ( $\Psi$ , here treated as a categorical variable due to our consideration of arbitrarily  
122 complex morphologies, see Table 1), and density ( $\rho_{particle}$ ) of the original sinking particle are known,  
123 while the sinking speed ( $U$ ) is unknown. The properties of the fluid surrounding the original particle  
124 (i.e.  $\rho_{fluid}$ ,  $\mu$ ) are also known, and our goal is to design experiments in which we sink a scaled-up  
125 model particle in a working fluid of known  $\rho_{fluid}$  and  $\mu$  in order to determine the model particle's  
126 sedimentation speed and, via similitude,  $U$  of the original particle.

127 While previous work (Berger et al., 1972; Fok-Pun & Komar, 1983; Takahashi & Be, 1984; Caromel et  
 128 al., 2014) suggests that the  $Re$  of sinking forams should be  $10^0 - 10^2$ , the exact value of  $Re$  for  
 129 morphology  $\Psi$  is assumed to be unknown. Hence, it is not immediately clear what size the model  
 130 should be (i.e., the scale factor  $S = L_{model}/L_{real}$ ) in order to match this  $Re$  in the experiments and  
 131 ensure similitude. Solving for both  $Re$  and  $S$  simultaneously requires additional mathematical  
 132 relationships beyond Eqn 1.

133 Throughout, we use a superscript O to refer to the *Original* values of dimensioned variables at life size  
 134 (e.g.,  $L^O, V^O, A^O, \rho_{particle}^O, U^O$ ) and  $Re^O, C_D^O$  for the values of the dimensionless Reynolds number  
 135 and drag coefficient (defined below) corresponding to real particles sinking in the original fluid (e.g.,  
 136 seawater of  $\rho_{fluid}^O, \mu^O$ ). While the fluid dynamics of flow around a particle of particular shape  $\Psi$  can  
 137 be considered theoretically over a range of  $Re$ , only the dynamics at  $Re^O$  and  $C_D^O$  will represent the  
 138 “operating point” corresponding to the life size particle settling speed  $U^O$ .

139 When a particle is sinking steadily at its terminal velocity, the sum of the external forces acting on the  
 140 particle is zero (Eqn 2); that is, the upward drag force ( $F_{drag}$ , Eqn 3) and buoyant force ( $F_{buoyancy}$ ,  
 141 Eqn 4) must balance the weight of the particle ( $F_{weight}$ , Eqn 5):

142 2

$$\Sigma F = F_{drag} + F_{buoyancy} - F_{weight} = 0$$

143 where

144 3

$$F_{drag} = \frac{1}{2} C_D(\Psi, Re) \rho_{fluid} U^2 A$$

146 Eqn 3 introduces the drag coefficient  $C_D(\Psi, Re)$ , a dimensionless descriptor of how streamlined an  
 147 object is. Both  $C_D$  and  $Re$  must be matched to achieve similitude.  $C_D$  depends on the shape of the  
 148 object  $\Psi$ , including its orientation relative to the freestream flow – for instance,  $C_D$  of a flat plate  
 149 oriented parallel to laminar flow is as low as 0.003 while  $C_D$  of a flat plate oriented perpendicular to  
 150 the flow is  $\sim 2.0$  (Munson et al., 1994). However, in addition to object geometry,  $C_D$  also depends on  
 151 qualitative characteristics of the flow, such as whether it is laminar or turbulent – that is,  $C_D$  also  
 152 depends on  $Re$ .  $C_D$  of a sphere decreases from about 200 at  $Re = 0.1$  to about 0.5 at  $Re = 1000$ ;  $C_D$   
 153 generally decreases with  $Re$  for most shapes (Munson et al., 1994; Morrison, 2013). While  $C_D$  does  
 154 not depend on object size directly, larger objects generally experience higher drag forces and this is  
 155 captured by the inclusion of particle area ( $A$ ) in the expression for  $F_{drag}$  (Eqn 3). For brevity, we will  
 156 omit  $\Psi$  hereafter and write the drag coefficient as  $C_D(Re)$ .

158 The buoyant force ( $F_{buoyancy}$ , Eqn 4) and weight ( $F_{weight}$ , Eqn 5) are both expressed using particle  
 159 volume ( $V$ ), gravitational acceleration ( $g$ ), and density of the fluid ( $\rho_{fluid}$ ) or particle ( $\rho_{particle}$ ),  
 160 respectively:

161

4

162

$$F_{buoyancy} = V \rho_{fluid} g$$

163

5

164

$$F_{weight} = V \rho_{particle} g$$

165 Substituting Eqns 3, 4, and 5 into 2 and eliminating  $U$  via the definition of  $Re$  (Eqn 1) yields an  
166 expression for the drag coefficient obtained through a force balance (indicated by a superscript  $\mathcal{F}$ ):

167

6

$$168 \quad C_D^{\mathcal{F}}(Re) = \frac{2(\rho_{particle} - \rho_{fluid}) V g}{\rho_{fluid} U^2 A} = \left( \frac{2(\rho_{particle} - \rho_{fluid}) V g}{\rho_{fluid} A} \right) \left( \frac{\rho_{fluid} L}{Re \mu} \right)^2 = \frac{2(\rho_{particle} - \rho_{fluid}) \rho_{fluid} V g L^2}{A Re^2 \mu^2}$$

169 Note that this expression can be simplified further upon identification of the dimensionless  
170 Archimedes number  $Ar = g L_{Ar}^3 \rho_{fluid} (\rho_{particle} - \rho_{fluid}) / \mu^2$  if the cubed length scale  $L_{Ar}^3 =$   
171  $VL^2/A$ , yielding  $C_D^{\mathcal{F}}(Re) = Ar/Re^2$ , as previously highlighted by others (e.g. Karamanev, 1996).  
172 However, we will proceed with the original form of Eqn 6 to keep key variables such as  $L$  explicit.

173 If  $C_D$  were known for a particular morphology, we could simply substitute values corresponding to the  
174 original test in seawater into Eqn 6 and solve for  $Re = Re^0$  and thus  $U^0$  via Eqn 1, immediately solving  
175 the problem of unknown settling speed. Unfortunately, the complex shapes of foraminifera (Table 1)  
176 coupled with the implicit dependence of  $C_D$  on  $Re$  means that both variables are generally unknown,  
177 and thus far we have only one constraining relationship between  $C_D$  and  $Re$ . More information is  
178 required to determine where along this constraint curve the operating  $C_D^0$  and  $Re^0$  are located. This  
179 information can come from experiments in which the sinking speeds of scaled-up model particles of  
180 various sizes (i.e., scale factors  $S$ ) in a viscous fluid are measured directly, allowing us to calculate  $Re$   
181 via Eqn 1 and then  $C_D^{\mathcal{F}}(Re)$  via Eqn 6 for the models, with appropriate values substituted for each  
182 experiment. For clarity, we can rewrite Eqn 6 for a model in terms of  $S$  and the original test parameters  
183 ( $L^0, A^0, V^0$ ):

184

7

$$185 \quad C_D^{\mathcal{F}}(Re) = \frac{2(\rho_{particle} - \rho_{fluid}) \rho_{fluid} (V) g (S L^0)^2}{(S^2 A^0) Re^2 \mu^2}$$

186 where we use the fact that for a model,  $L = S L^0$  and  $A = S^2 A^0$ . While one would also expect  $V =$   
187  $S^3 V^0$  for 3D printed models, limitations of our 3D printer led to variation in  $V$  that we overcame using  
188 a more general empirical relationship between  $S$  and  $V$  based on mass measurements – see 3D Printer  
189 Limitations. Eqn 7 represents a constraining relationship between  $C_D$  and  $Re$  for the sinking particle,  
190 which we use to collect  $(Re, C_D)$  experimental data points at several  $S$ . Once sufficient data are  
191 collected, we can construct a new, empirical relationship (e.g., a cubic spline fit) between  $C_D$  and  $Re$   
192 for a particular particle shape, which we term  $C_D^E(Re)$ . Finally, we can solve for the operating  $Re^0$ ,  
193  $C_D^0$ , and  $U^0$  by finding the intersection point between the  $C_D^{\mathcal{F}}(Re)$  constraint curve specific to life-size  
194 particles sinking in seawater (i.e., Eqn 7 with  $S = 1$  and  $\rho_{particle}^0, \rho_{fluid}^0, \mu^0$ ) and our empirical  $C_D^E(Re)$

195 spline curve valid for a particular particle shape moving steadily through any fluid. MATLAB code can  
196 be downloaded from <https://github.com/matthewwalkerbio/Dynamic-scaling>.

## 197 Study Species

198 To construct an empirical  $C_D^E(Re)$  curve for a particular test morphology, we started with 3D scans of  
199 individual specimens from 30 different species (Table 1). The majority of the species were selected  
200 from the University of Tohoku museum's database, eforum Stock  
201 (<http://webdb2.museum.tohoku.ac.jp/e-foram/>), with a micro computed tomography ( $\mu$ CT) scan  
202 resolution between 2.5 and 3.6 pixels per  $\mu\text{m}$ , and were exported as 3D triangular mesh (STL format)  
203 files. Specimens of an additional three species were scanned using synchrotron radiation based  
204 micro-computed tomography (SR $\mu$ CT). Imaging was performed at the Imaging Beamline P05 (IBL)  
205 (Greving et al., 2014; Haibel et al., 2010; Wilde et al., 2016) operated by the Helmholtz-Zentrum-  
206 Geesthacht at the storage ring PETRA III (Deutsches Elektronen Synchrotron – DESY, Hamburg,  
207 Germany). Specimens were imaged at an photon energy of 14 keV and with a sample to detector  
208 distance of 17 mm. For each tomographic scan, 900 projections at equal intervals between 0 and  
209  $\pi$  were recorded. Tomographic reconstruction was done via a classical filtered back projection using  
210 the RECLBL library (Huesman et al., 1977). For processing, raw projections were binned two times  
211 resulting in an effective pixel size of the reconstructed volume of 1.44  $\mu\text{m}$ . These scans were  
212 segmented and rendered using SPIERS (Sutton et al., 2012), and again exported in STL format and are  
213 available from [Morpho Source](#). Meshes of all foraminifera were manually checked in Meshlab (Callieri  
214 et al., 2012) for integrity.

215 For species where more than one scan was available, the scan that contained the best-preserved  
216 specimen was chosen. By only including one specimen per species, this approach neglects phenotypic  
217 plasticity which is demonstrated in planktonic foraminifera (e.g. Lohmann, 1983; Morard *et al.*, 2013),  
218 but was chosen due to limitations of  $\mu$ CT scan availability and time constraints on the project.

219

## 220 3D printing and model preparation

221 The 3D scans allowed us to easily fabricate scaled-up (scale factor  $S$ ) physical models of each specimen  
222 using a FormLabs Form1+ (Formlabs, Somerville, Massachusetts, USA) 3D printer, using FormLabs  
223 Clear Resin Version 2 (Formlabs, Somerville, Massachusetts, USA) with a layer thickness of 50  $\mu\text{m}$  (see  
224 **Error! Reference source not found.**D for examples) and x-y resolution of 200  $\mu\text{m}$ . Models were  
225 washed and flushed with isopropanol to remove excess resin following Formlabs' guide and allowed  
226 to air dry. Support material was removed (**Error! Reference source not found.**D.vi), and the models  
227 lightly sanded with 400 grit Wet 'n' Dry paper, followed by a final isopropanol wash to remove any  
228 remaining residue. Once dry, models were filled with mineral oil in preparation for sinking. Clear resin  
229 was chosen to allow each model to be checked for bubbles (which would increase the buoyancy of  
230 the model). Any bubbles were removed using a 30-gauge needle and syringe.



231 Following convention when defining the area  $A_{particle}$  used in the definition of  $C_D$  (Eqn 3), we  
232 measured projected area of the sinking foraminifera. Referring to high resolution images of the sinking  
233 model (**Error! Reference source not found.D**), a digital model of the foraminifera was manually  
234 aligned to measure the projected area in a plane perpendicular to the sinking direction (**Error!  
235 Reference source not found.D**). We used the same procedure to measure the maximum length  
236 parallel to the flow ( $L$ ) for the calculation of  $Re$  (**Error! Reference source not found.D**). These choices  
237 facilitated objective comparisons of  $C_D$  across morphologically diverse species, to be detailed in a  
238 future study.

### 239 3D Printer Limitations

240 Whilst in principle, the volume of a printed model should simply scale according to  $V = S^3 V^0$ , due to  
241 inherent limitations of the 3D printer as well as difficulty in removing excess resin from small models,  
242 we found that this expectation was usually not satisfied, and weighing the models showed that  
243  $M/\rho_{particle} > S^3 V^0$  where  $M$  is particle mass (**Error! Reference source not found.C**). Therefore, we  
244 estimated  $V$  of each model by weighing on an Entris 224-1S mass balance ( $\pm 0.001$  g) and assuming  
245  $\rho_{particle}$  was  $1121.43 \pm 13.73$  kg m<sup>-3</sup>, based on the average mass of five 1 cm<sup>3</sup> cubes of printed resin.  
246 Furthermore, whenever a predicted value for  $V$  at a given scale factor  $S$  was needed, i.e. in Eqn 7 (see  
247 Remaining iterations), we based this on cubic spline interpolation of our  $V(S)$  data for existing models  
248 when sufficient data were available, with extrapolation based on cubic scaling of  $V(S)$  if required (see  
249 **Error! Reference source not found.C**):

250

8

$$251 \quad V^{predict}(S) = \begin{cases} H(S), N \geq 3 \\ S^3 V^0, N < 3 \text{ or } S = 1 \end{cases}$$

252 where  $N$  is the number of existing volume measurements and  $H$  represents the cubic spline fit of  $V$   
253 vs  $S$ . Note that because we always directly measured  $V$  by weighing after printing each model, and it  
254 is not necessary to achieve the exact  $Re$  and  $C_D$  of the operating point ( $Re^0$  and  $C_D^0$ ) in the experiments  
255 (see Remaining iterations), the empirical spline-based volume prediction was not strictly required for  
256 our method to succeed. It merely aids in improving the rate of convergence of our iterative approach  
257 by reducing the difference between our anticipated and actual  $Re$ ,  $C_D$  for each experiment.

### 258 Settling tank

259 The models were released in a cylindrical acrylic tank (0.9 m in diameter and 1.2 m in height) of mineral  
260 oil ("Carnation" white mineral oil, Tennants Distribution Limited, Cheetham, Manchester, UK;  $\rho = 830$   
261 kg m<sup>-3</sup>,  $\mu = 0.022$  Pa S) filled to a depth of 1.18 m (approximately 750 L). The tank was fitted with a  
262 custom net and net retrieval system (**Error! Reference source not found.A**) to allow easy retrieval of  
263 the models after their descent, allowing each model to be sunk 5 times. Integrated to the net retrieval  
264 system was the release mechanism, which was held centrally over the tank, with the grasping parts  
265 submerged below the oil level. This ensured that each model was released in a controlled and  
266 repeatable fashion.

## 267 Particle imaging

268 To minimise reflections, the tank was surrounded by a black fabric tent-like structure. This also served  
269 as a dark background to facilitate visualisation of the model during descent. The tank was illuminated  
270 with a single 800 lumen LED spotlight placed underneath the tank and, as the Formlabs' Clear Resin is  
271 UV-fluorescent, two 20W "Blacklight" UV fluorescent tubes were placed above the tank.

272 The sinking models were recorded using two Logitech C920 HD webcams (Logitech, Lausanne,  
273 Switzerland), placed at 90° to each other (**Error! Reference source not found.A**) and recording at 960  
274 pixels x 720 pixels and ~30 frames per second, allowing monitoring of the position and orientation of  
275 the particle in 3D as it fell. As these consumer-grade webcams use a variable frame-rate system, a  
276 custom MATLAB script was used to initiate camera recording, recording both frames and frame  
277 timestamps. Videos were recorded for 500 frames (~17 s). Sinking velocity was calculated over a  
278 central 0.8 m depth range, ensuring the model was at terminal velocity (see Time to Terminal Velocity)  
279 whilst also avoiding end effects which could slow the model as it reached the bottom of the tank.  
280 Based on observations of suspended dust, there was no discernable convection in the tank during any  
281 trials that might potentially affect sinking velocities. The curved walls of the tank introduced distortion,  
282 which was removed using the MATLAB toolbox "Camera Calibrator" (Mcandrew, 2004). Pixel size was  
283 1.06 pixels per millimetre with a mean reprojection error of 0.5 pixels, therefore distance  
284 measurements (for calculating sinking velocities) were accurate to within 0.5 mm (0.06% of the  
285 traversed depth).

## 286 Velocity calculation

287 Models were tracked in distortion-corrected frames using a modified version of Trackbac (Guadayol  
288 et al., 2017; Guadayol, 2016). The per-frame centroid coordinates obtained were then paired with the  
289 timestamp values recorded to calculate average settling velocity components in 2D for each camera  
290 (below,  $U_x$  is horizontal speed from camera one,  $U_y$  is horizontal speed from camera two, and  $U_{z,1}$   
291 and  $U_{z,2}$  are the vertical speeds corresponding to the two cameras). A resultant velocity magnitude  
292 was then calculated for each camera, and these two values averaged to yield a single estimate for  $U$   
293 per experiment (Eqn 9).

294

9

$$295 \quad U = \frac{1}{2} \left( \sqrt{U_x^2 + U_y^2 + U_{z,1}^2} + \sqrt{U_x^2 + U_y^2 + U_{z,2}^2} \right)$$

296 Each model was sunk five times and a mean  $U$  was calculated from these replicates. Replicates beyond  
297 a threshold of  $\pm 5\%$  of the median sinking velocity were discarded from this average. Each model was  
298 dropped one additional time and photographed using a Canon 1200D DSLR camera (Tokyo, Japan)  
299 mounted on a tripod close to the tank, to obtain high resolution (18 megapixels) images which were  
300 used to determine model orientation (and thus  $L$  and  $A$ ) during settling (**Error! Reference source not  
301 found.D**).

## 302 Wall effects

303 At low  $Re$ , the effects of artificial walls in an experimental (or computational) system can be  
304 nonintuitively large and lead to substantial errors if not accounted for (Vogel, 1994). Acting as an  
305 additional source of drag, the walls several tens of particle diameters away can slow down a sinking  
306 particle and increase its apparent drag coefficient. We designed our experiments to minimise wall  
307 effects by using an 0.8 m diameter tank (**Error! Reference source not found.**A) and model diameters  
308 on the order of 1 cm. To reduce potential errors further, we applied the method of Fayon & Happel  
309 (1960; summarised in Clift et al., 1978) to convert between the apparent drag coefficient when walls  
310 are present ( $C_D^{wall}$ ) and the desired drag coefficient in an unbounded domain ( $C_D^\infty$ ):

311 10

$$312 \quad C_D^\infty \approx C_D^{walls} - \frac{24}{Re} (K(\lambda) - 1)$$

313 where

$$314 \quad K(\lambda) = \frac{1 - 0.75857 \cdot \lambda^5}{1 - 2.1050 \lambda + 2.0865 \lambda^3 - 1.7068 \lambda^5 + 0.72603 \lambda^6}$$

315 Here,  $\lambda = d/D$  where  $d$  is the diameter of the sinking particle and  $D$  is the tank diameter; we take  
316  $d = L$ . While Eqn 10 is not exact, it substantially reduces the error otherwise incurred if one were to  
317 neglect wall effects entirely. Note that Eqn 10 is only valid up to about  $Re = 50$ , beyond which  
318 different corrections can be used (Clift et al., 1978).

319 We applied this correction by taking any experimentally determined  $C_D$  to equal  $C_D^{walls}$ , and using  $C_D^\infty$   
320 estimated according to Eqn 10 for subsequent calculations as detailed below. In our experiments,  $\lambda$   
321 ranged from 0.0027 – 0.0173, yielding  $K$  between 1.0057 – 1.0377. Wall effects were therefore quite  
322 small, with  $C_D^\infty / C_D^{walls}$  ranging from 0.993 – 0.994.

## 323 Iterative approach

### 324 *First iteration*

325 To construct an empirical cubic spline  $C_D^E(Re)$  needed to solve for  $U^O$ , at least three experimental  
326 data points (corresponding to three scale factors) are needed. These first three  $S$  were chosen by using  
327 an existing empirical  $C_D(Re)$  relationship for a sphere, valid for  $0 < Re < 10^6$  (Morrison, 2013, Fig. 8.13,  
328 page 625):

329 11

$$330 \quad C_D^M(Re) = \frac{24}{Re} + \frac{2.6 \left(\frac{Re}{5.0}\right)}{1 + \left(\frac{Re}{5.0}\right)^{1.52}} + \frac{0.411 \left(\frac{Re}{263,000}\right)^{-7.94}}{1 + \left(\frac{Re}{263,000}\right)^{-8.00}} + \left(\frac{Re^{0.80}}{461,000}\right)$$

331 While morphologically complex particles such as foraminifera tests (Table 1) are not expected to  
332 behave like ideal spheres, Eqn 11 should be sufficient to provide initial guesses, after which we iterate  
333 to find the solution. We note that if the particle shapes of interest were all most similar to some other

334 well-studied geometry (e.g., cylinders, discs, etc), using a known  $C_D(Re)$  relationship for that shape  
335 could provide better initial guesses and faster convergence.

336 Substituting Eqn 11 into Eqn 7 (with  $S = 1$ ,  $V = V^O$ , and  $\rho_{particle}^O, \rho_{fluid}^O, \mu^O$  substituted) and moving  
337 all terms to one side, we can numerically solve (MATLAB: *fzero* function) for our first estimate of the  
338 operating  $Re^O$ . Substituting this  $Re$  back into Eqn 7 or Eqn 11 yields an estimate of the operating  $C_D^O$ .  
339 We aimed to reproduce this  $Re$  and  $C_D$  in the first experiment, excepting that we accounted for wall  
340 effects by distinguishing between  $C_D^\infty$  and  $C_D^{walls}$  expected to occur in the tank. Hence, we could again  
341 substitute this  $Re$  into Eqn 7 but now with  $\rho_{particle}$  corresponding to the resin model and  $\rho_{fluid}$  and  
342  $\mu$  corresponding to mineral oil, and combine this expression with Eqn 10, assuming our estimated  
343  $C_D = C_D^\infty$ ,  $C_D^F = C_D^{walls}$ , and  $\lambda = S L^O / D$ . The resulting expression can be solved numerically for the  
344 first scale factor, termed  $S_1$ . Two more scale factors ( $S_2$  and  $S_3$ ), one smaller and one larger than  $S_1$ ,  
345 were chosen to span expected  $Re$  values for forams from published literature (e.g. Fok-Pun & Komar,  
346 1983; Takahashi & Be, 1984; Caromel et al., 2014) as well as  $Re^O$  for other species which had reached  
347 convergence. This procedure was intended to bound the correct  $S$  value that reproduces the operating  
348  $Re^O$  and  $C_D^O$  of the settling particle. The three models were printed, their actual volumes  $V$  measured  
349 via weighing, and their settling velocities  $U$  experimentally measured as detailed in Appendix 3.

350 An empirical cubic spline curve  $C_D^E(Re)$  can now be fitted (D'Errico, 2009) to these three initial ( $Re, C_D$   
351 ) data points, constrained to be monotonically decreasing and concave up within the limits of the data  
352 to match expectations for drag on objects at low to moderate  $Re$ . Three optimally spaced spline knots  
353 were used since this yielded excellent fits to the data as the number of data points increased. These  
354 details of the spline as well as its order (i.e., cubic vs linear) are somewhat arbitrary but we ensured  
355 that our results were sufficiently converged as to be insensitive to them (see Remaining iterations).

356 The operating point ( $Re^O, C_D^O$ ) corresponding to the particle settling in the natural environment can  
357 be visually represented as the intersection point of the  $C_D^F(Re)$  curve defined by Eqn 7 (with  $S = 1$   
358 and  $\rho_{particle}^O, \rho_{fluid}^O, \mu^O$ ) and the empirical  $C_D^E(Re)$  relationship based on our experimental data.  
359 Algebraically, the operating point is the solution to  $C_D^F(Re) = C_D^E(Re)$ . We solved for  $Re^O$  numerically  
360 using a root finding algorithm (MATLAB's *fzero*) on the objective function  $C_D^F(Re) - C_D^E(Re) = 0$  and  
361 then obtained  $C_D^O$  by substituting  $Re^O$  into Eqn 7. Finally,  $U^O$  was easily determined from the definition  
362 of  $Re^O$  (Eqn 1 with  $U^O, L^O$ , and  $\rho_{fluid}^O$  substituted).

363 Since our first three empirical data points and fitted spline  $C_D^E$  corresponded to guessed model scale  
364 factors  $S$ , our initial operating point prediction ( $Re^O, C_D^O$ ) often was not located near any of these  
365 initial points or sometimes even within the bounds of these data (in which case linear extrapolation  
366 of  $C_D^E$  was used to estimate the operating point). Therefore, to ensure the accuracy of our predicted  
367  $U^O$ , we continued iterating with additional experiments.

### 368 *Remaining iterations*

369 The model scale factor for the  $N^{\text{th}}$  experiment was chosen by combining Eqns 7, 8, and 10 with  
370  $Re = Re^O$  and  $C_D^{walls} = C_D^O$ , (from the previous iteration),  $C_D^F = C_D^\infty$ , and  $V = V^{\text{predict}}$ , and

371 numerically solving for  $S$ . A model close to this new scale was printed and sunk, its settling velocity  $U$   
372 recorded and  $Re$  and  $C_D$  computed, and a more accurate spline  $C_D^E$  constructed by including this new  
373 data point. The calculation of  $(Re^O, C_D^O)$  detailed in the previous section was then repeated, yielding  
374 a more accurate operating point. Overall, the aim was to tightly bound the predicted operating point  
375 with experimental data to maximize confidence in the fitted spline in this region.

376 The iterative process (visualized as a flowchart, **Error! Reference source not found.B**, with a specific  
377 example of convergence given in Fig. 2 C & D) was repeated until:

- 378 1) the predicted operating point was not extrapolated beyond our existing data,
- 379 2) the variation in calculated  $U^O$  between the fitting of a linear spline and cubic spline was no  
380 greater than 5%, and
- 381 3) the variation between the predicted  $Re^O$  and the closest experimentally measured  $Re$  was  
382 less than 15%.

383 In many cases, the difference between results based on four versus three data points was very small  
384 (Fig. 2 C, D), indicating rapid convergence and the possibility of streamlining the method further in  
385 the future. Through this method we calculated the sinking velocities of 30 species of planktonic  
386 foraminifera (Table 1).

### 387 Method Validation

388 Our basic methodology was first validated by 3D printing a series of spherical models (10 – 20 mm in  
389 diameter) for which the theoretical  $C_D(Re)$  relationship is already well-known. In order to achieve low  
390 density (and thus low sinking velocity and low  $Re$ ), these spheres were hollow and filled with oil via  
391 two small holes (of diameter 0.8% of the sphere diameter). Our empirically generated  $C_D^E(Re)$  curve  
392 compares favourably with the theoretical  $C_D^M(Re)$  curve (Morrison, 2010) ( $R^2=0.875$ , **Error! Reference  
393 source not found.A**), with the distance between the curves approximately constant above  $Re \approx 25$ .  
394 While the error grows larger at lower  $Re$ , we expected most foraminifera species to operate at  $Re \approx$   
395 18 – 55 based on previous work (Berger et al., 1972; Fok-Pun & Komar, 1983; Takahashi & Be, 1984;  
396 Caromel et al., 2014).

397 To quantify errors in our approach even more directly, we then considered hypothetical hollow  
398 spherical particles with the same material density ( $\rho_{particle}^O$ ) as foraminifera tests and a range of sizes  
399 ( $L^O = 750 - 1150 \mu\text{m}$ , similar to the species we studied) settling in seawater. This size range  
400 corresponds to  $Re = 12 - 27$ , the area where our  $C_D^E(Re)$  curve is most divergent from  $C_D^M(Re)$ . We  
401 compared predictions of the operating  $U^O$  based on our empirical  $C_D^E(Re)$  curve versus the theoretical  
402  $C_D^M(Re)$  curve for spheres as outlined above, substituting Eqn 11 for  $C_D^E(Re)$  in the latter case.  
403 Maximum relative error in predicted  $U^O$  was 11.5% at  $Re = 16$  (corresponding to a sphere 860  $\mu\text{m}$  in  
404 diameter) while the minimum difference was 6.5% at  $Re = 27$  (corresponding to a sphere of 1150  $\mu\text{m}$   
405 in diameter, **Error! Reference source not found.A**). This level of error is much smaller than the  
406 variation in  $U^O$  we predicted across the 30 foraminifera species we investigated (Table 1).

407 Time to Terminal Velocity

408 This study was concerned with predicting steady sinking speeds, but in our experiments, each model  
 409 foraminifera took a finite amount of time to accelerate from rest at the point of release to its  
 410 terminal sinking velocity. Since this transient portion of the sinking trajectory could introduce errors  
 411 into our analysis, it is important to determine whether it affected any of our recorded data.

412 During the transient acceleration phase, Eqn 2 does not hold. Instead, we can revert to the more  
 413 general form of Newton's second law:

414 12

415 
$$\Sigma F = F_{drag} + F_{buoyancy} - F_{weight} = -Ma = -M \frac{dU}{dt}$$

416 where  $M = V\rho_{particle}$  is particle mass, and the acceleration  $a$  can be equated to the time derivative  
 417 of instantaneous velocity  $\frac{dU}{dt}$ . A negative sign appears on the right-hand side so that we can define  
 418 the downward movement as positive for convenience. We can then substitute expressions for each  
 419 force as before:

420 13

421 
$$1/2 C_D(\Psi, Re) \rho_{fluid} U^2 A + V \rho_{fluid} g - V \rho_{particle} g = -M \frac{dU}{dt}$$

422 While thus far we have not assumed anything about the particle shape, to proceed further we require  
 423 knowledge of  $C_D(\Psi, Re)$  from vanishingly small  $Re$  (when the particle is at rest) up to the terminal  
 424 velocity. Hence we will assume a spherical particle as an approximation to the model forams, so that  
 425 Morrison's empirical equation (Eqn 11) can then be substituted for  $C_D(\Psi, Re)$ :

426 14

427 
$$- \left\{ 1/2 \left[ \frac{24}{Re} + \frac{2.6 \left(\frac{Re}{5.0}\right)}{1 + \left(\frac{Re}{5.0}\right)^{1.52}} + \frac{0.411 \left(\frac{Re}{263,000}\right)^{-7.94}}{1 + \left(\frac{Re}{263,000}\right)^{-8.00}} + \left(\frac{Re^{0.80}}{461,000}\right) \right] \rho_{fluid} U^2 A + V \rho_{fluid} g - V \rho_{particle} g \right\} / M = \frac{dU}{dt}$$

428 Here we have isolated  $\frac{dU}{dt}$  on the right-hand side. If the definition of  $Re = \frac{LU \rho_{fluid}}{\mu}$  is inserted into 14  
 429 (not shown for brevity), one obtains an ordinary differential equation (ODE) for the unsteady velocity  
 430  $U(t)$ . The depth of the sphere  $Z(t)$  can then be obtained by solving a second much simpler ODE:

431 15

432 
$$U = \frac{dZ}{dt}$$

433 Both ODEs are easily solved numerically by e.g. MATLAB's *ode45* subject to the initial conditions  
 434  $U(t = 0) = 0$  and  $Z(t = 0) = 0$ .

435 It is well known that as  $Re$  approaches zero in the limit of inertia-less Stokes flow, unsteadiness can  
 436 only occur due to time-varying boundary conditions. Thus, a microorganism that stops actively  
 437 swimming will almost instantly come to a stop, and a heavy micro-particle released from rest will

438 almost instantly begin sinking at its terminal velocity (Purcell, 1977). As  $Re$  increases and inertia  
439 becomes increasingly important, the transient period of acceleration becomes longer. Therefore, a  
440 reasonable worst-case to examine here is the foraminifera model that sank at the highest  $Re$ .

441 We found *Globorotalia (Truncorotalia) truncatulinoides* to operate at  $Re = 42$  (Table 1) but here we  
442 conservatively chose the largest scale model used to generate its  $C_D^F(Re)$  spline for which  $S = 16$  and  
443  $Re = 90$ . Inserting this model's length  $L$ , area  $A$ , and measured volume  $V$  into 14, we obtain solutions  
444 for the time varying speed and depth of a sphere approximating this model's geometry (**Error!**  
445 **Reference source not found.**B). The depth corresponding to where speed equals 99.9% of the terminal  
446 velocity is approximately 4.6 cm, which is much smaller than the 19 cm between where the models  
447 were released and the edge of the cameras' fields of view for data collection. Hence, the transient  
448 acceleration of each model foraminifera should have had no effect on our data or results. Most of our  
449 models should have reached terminal velocity even sooner since they sank at lower  $Re$ , e.g. within 2.2  
450 cm for *C. dissimilis* operating at  $Re = 36$  (Table 1).

## 451 Results & Discussion

452 Here we present a novel method of determining settling speed by leveraging dynamically scaled  
453 models falling under gravity rather than being towed at a controlled speed. Applying our method to  
454 foraminifera-inspired spherical particles (**Error! Reference source not found.**A), we predict settling  
455 speeds within 11.5% of theoretical expectations (**Error! Reference source not found.**E). In **Error!**  
456 **Reference source not found.**C & D we present an example of convergence of our method to the  
457 operating  $Re^0$ ,  $C_D^0$ , and  $U^0$  of a typical foraminifera species. There was little variation in the number  
458 of iterations required to reach convergence (mean 4, range 3-6, see Table 1), despite the  
459 morphological complexity of some species (e.g. *Globigerinoidesella fistulosa*). We suspect the higher  
460 end of this range was due to these species having forms that were particularly challenging to clean  
461 residual resin from, or the incomplete removal of air bubbles once submerged in oil.

462 *Table 1. Predicted sinking speeds  $U^0$  for the 30 species of planktonic foraminifera included in this study, with each species*  
463 *shown in both spiral view and 90° rotation (so that the spiral view is facing to the left). <sup>1</sup> indicates species scanned high*  
464 *resolution. Scans of the remaining 27 species were obtained from The University of Tohoku museum's database. Operating*  
465  *$Re^0$  and  $C_D^0$  predicted for each species are also presented, and for comparison, the theoretical  $C_D^M$  of a sphere at the same*  
466  *$Re^0$ . The number of model iterations required to achieve convergence of  $U^0$  is listed (iter). Species synonyms checked at*  
467 *World Foraminifera Database (Hayward et al., 2018).*



Image	Species	$L^0$ ( $\mu\text{m}$ )	$U^0$ ( $\text{cm s}^{-1}$ )	$Re^0$	$C_D^0$	$C_D^M$ ( $Re^0$ )	Iter	Image	Species	$L^0$ ( $\mu\text{m}$ )	$U^0$ ( $\text{cm s}^{-1}$ )	$Re^0$	$C_D^0$	$C_D^M$ ( $Re^0$ )	Iter
	<i>Neogloboquadrina acostaensis</i> (Blow, 1959)	307	3.6	9.3	2.5	4.2	4		<i>Sphaeroidinella dehiscentes</i> (Parker & Jones, 1865)	460	5.1	32.8	1.7	1.9	4
	<i>Globigerinella adamsi</i> (Banner & Blow, 1959)	596	3.0	9.6	2.6	4.1	4		<i>Sphaeroidinella dehiscentes</i> <sup>1</sup> (Parker & Jones, 1865)	269	2.5	8.3	5.0	4.5	4
	<i>Dentoglobigerina altispira</i> (Cushman & Jarvis, 1936)	282	6.1	37.3	1.6	1.8	5		<i>Catapsydrax dissimilis</i> (Cushman & Bermúdez, 1937)	380	6.2	36.4	1.7	1.8	4
	<i>Globoturborotalita apertura</i> (Cushman, 1918)	650	2.8	7.4	4.2	4.8	3		<i>Neogloboquadrina dutertrei</i> (d'Orbigny, 1839)	259	4.0	17.6	2.0	2.8	3
	<i>Globigerina bulloides</i> (d'Orbigny, 1826)	296	3.3	13.4	2.4	3.3	5		<i>Globigerinoidesella fistulosa</i> (Schubert, 1910)	392	2.6	11.2	3.0	3.7	4
	<i>Globigerinoides conglobatus</i> <sup>2</sup> (Brady, 1879)	445	1.8	5.8	5.9	5.7	4		<i>Globorotaloides hexagonus</i> (Natland, 1938)	321	2.7	8.3	3.4	4.5	4
	<i>Globorotalia (Truncorotalia) crassaformis</i> (Galloway and Wissler, 1927)	250	3.3	9.5	2.3	4.1	4		<i>Globigerinella obesa</i> (Bolli, 1957)	539	2.8	9.4	3.2	4.1	4
	<i>Neogloboquadrina humerosa</i> (Takayanagi & Saito, 1962)	360	5.0	22.7	2.0	2.4	4		<i>Globorotalia praemenardii</i> (Cushman & Stainforth, 1945)	569	3.9	14.0	2.0	3.2	4
	<i>Globoconella inflata</i> (d'Orbigny, 1839)	422	3.9	21.4	2.5	2.5	4		<i>Globoconella puncticulata</i> (d'Orbigny in Deshayes, 1832)	269	4.0	14.5	2.8	3.1	3
	<i>Fohsella lobata</i> (Bermúdez, 1949)	312	4.2	13.1	1.5	3.3	3		<i>Fohsella robusta</i> (Bolli, 1950)	312	3.5	11.2	2.3	3.7	4
	<i>Globorotalia margaritae</i> (Bolli & Bermúdez, 1965)	258	2.9	8.1	2.8	4.6	3		<i>Dentoglobigerina tripartita</i> (Koch, 1926)	303	5.7	33.6	2.0	1.9	3
	<i>Paragloborotalia mayeri</i> (Cushman & Ellisor, 1939)	298	3.1	8.1	3.2	4.5	4		<i>Paragloborotalia siakensis</i> (LeRoy, 1939)	366	3.4	10.4	2.6	3.9	4
	<i>Globoturborotalita nepenthes</i> (Todd, 1957)	353	4.2	16.4	2.4	2.9	3		<i>Globoconella sphericomiozea</i> (Walters, 1965)	627	4.3	18.8	2.3	2.7	4
	<i>Pulleniatina obliquiloculata</i> (Parker & Jones, 1865)	350	4.1	24.6	2.1	2.3	3		<i>Globorotalia (Truncorotalia) truncatulinoides</i> (d'Orbigny, 1839)	325	6.3	42.4	1.5	1.6	6
	<i>Fohsella peripheroronda</i> (Blow & Banner, 1966)	422	3.6	10.1	2.4	4.0	3		<i>Orbulina universa</i> <sup>2</sup> (d'Orbigny, 1839)	567	1.0	3.3	10.7	8.7	6
	<i>Praeorbulina curva</i> (Blow, 1956)	433	4.8	17.8	2.2	2.8	3								

469 Our predicted sinking speeds of foraminifera fall within aggregated existing data for 14 species (**Error!**  
470 **Reference source not found.**, Fok-Pun & Komar, 1983; Takahashi & Be, 1984; Caromel et al., 2014)  
471 and compare well with known speeds for other particles of comparable size and density (e.g. faecal  
472 pellets, (Table 3, Iversen & Ploug, 2010), phytoplankton (Fig. 1 in Smayda, 1971)). However, it should  
473 be noted that our predicted speeds are higher than published values for five out of the seven  
474 foraminifera species for which direct comparisons are possible (**Error! Reference source not found.**).  
475 This could be due to our ability to observe enlarged models of sinking foraminifera more accurately  
476 compared to actual specimens, and the lack of control for wall effects in previous work, which would  
477 tend to underestimate sinking speeds. There could also be considerable natural variation, which our  
478 single specimen per species (excluding *S. dehiscens*) does not capture.

479 Sedimentation of microscale plankton has been measured both *in situ* (e.g. Waniek, Koeve and Prien,  
480 2000) and in the laboratory (e.g. Smayda, 1971; Miklasz and Denny, 2010). By settling dense  
481 suspensions of microorganisms, these studies provided a population sinking rate (Bienfang, 1981)  
482 which could be two to three times lower than the settling velocity of an isolated particle in the typically  
483 dilute ocean (Miklasz & Denny 2010). Other studies have, like us, used enlarged models of microscale  
484 plankton to facilitate observations. Padisak et al. (2003) used handmade models of plankton to  
485 examine drag, but there was no attempt at accurately matching  $Re$ . Holland (2010) used mechanical  
486 pencil leads as models of sinking diatom chains, keeping  $Re < 1$  in an improvement over Padisak et al.  
487 (2003). However, neither authors calculated sinking velocities for real organisms. Our dynamic scaling  
488 approach ensures that we accurately recreate the fluid flows around settling organisms – a  
489 requirement for the correct prediction of sinking speeds. We also improve on previous methodologies  
490 by effectively eliminating wall effects, basing our models on  $\mu$ CT scans, and using inexpensive cameras  
491 to observe natural sinking orientation.

492 By design, our dynamic scaling approach yields an interpolated  $C_D(Re)$  curve that describes the flow  
493 dynamics (and thus sinking speeds) that would occur if various fluid and/or particle parameters were  
494 varied, offering a degree of flexibility not seen in other studies. For example, phytoplankton blooms  
495 can increase both the density and viscosity of water due to exudates (Jenkinson et al., 2015), while  
496 increasing global temperatures have the opposite effect. The density and viscosity of seawater also  
497 naturally vary with latitude. Understanding how these variations affect sinking rates can offer insights  
498 into the evolutionary pressures on plankton. Our approach also allows us to isolate the effects of  
499 shape on sinking, even across species of widely varying size, density, etc, by comparing  $C_D$  of different  
500 species all hypothetically sinking at the same  $Re$ ; a manuscript focused on such biological questions  
501 relating to foraminifera is currently in preparation. Differential settling speeds of foraminifera also

502 have implications for nutrient cycling, paleoclimate reconstruction (Kucera, 2007), and the marine  
503 calcite budget (Schiebel, 2002).

504 Our method can easily be modified to study sedimenting particles operating at any  $Re$ , providing the  
505 system's  $Re$  range can be experimentally replicated. Other sinking marine particles include diatoms  
506 ( $Re \approx 10^{-2} - 1$ , Botte, Ribera D'Alcalà and Montresor, 2013) and radiolaria ( $Re \approx 10 - 200$ , Takahashi and  
507 Honjo, 1983), for which one could use digital models as we have in conjunction with a suitably viscous  
508 fluid (high viscosity silicone oil, see SI Further Applications) to enable sufficiently large models to be  
509 produced (25 cm, see SI: Further Applications). The method can also be applied to terrestrial systems  
510 such as settling spores ( $Re \approx 50$  e.g. Noblin, Yang and Dumais, 2009) and dispersing seeds ( $Re \approx 10^3$   
511 Azuma and Yasuda, 1989), again by using 3D printed models based on (often existing)  $\mu$ CT data.

512 Whilst our method pertains to settling in a quiescent fluid, one could conduct similar experiments  
513 using a flume to calculate threshold resuspension velocities (i.e. the horizontal flow speed required to  
514 lift a particle off the substrate), important in the study of wind erosion and particle transport and  
515 deposition (Bloesch, 1995; Bagnold, 1971). Similarly, studying particles suspended in shear flow could  
516 be achieved using a treadmill-like device (e.g. Durham et al (2009)) or a Taylor-Couette apparatus (e.g.  
517 Karp-Boss & Jumars (1998)). While additional dimensionless groups beyond  $Re$  and  $C_D$  would need to  
518 be matched to achieve similitude in these systems, we hope that our study provides a starting point  
519 for the experimental study of these and other more complex problems.

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## 529 Competing interests

530 The authors have no competing interests to declare.

## 531 Author contributions

532 SH & MW were responsible for conceptualisation of the project, MW, SH and RS contributed to  
533 developing the methodology, MW collected data and JUH & FW scanned the three foraminifera at  
534 high resolution (See Table 1). RS & MW were responsible for MATLAB code, data curation and  
535 visualisation, formal validation of the method and the initial draft of the manuscript. TH segmented  
536 the high resolution scans, generated models used for 3D printing, and assisted with 3D printing of all  
537 models. MW, SH & RS contributed to editing and reviewing the manuscript, FW & JUH provided details  
538 on methodology of high resolution scanning. SH & RS supervised MW for the duration of the project,  
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## 544 References

- 545 Ali, A., Thiem, Ø. & Berntsen, J. (2011). Numerical modelling of organic waste dispersion from fjord  
546 located fish farms. *Ocean Dynamics*. [Online]. 61 (7). p.pp. 977–989. Available from:  
547 <http://link.springer.com/10.1007/s10236-011-0393-8>.
- 548 Azuma, A. & Yasuda, K. (1989). Flight performance of rotary seeds. *Journal of Theoretical Biology*.  
549 [Online]. 138 (1). p.pp. 23–53. Available from:  
550 <https://linkinghub.elsevier.com/retrieve/pii/S0022519389801766>.
- 551 Bagnold, R.A. (1971). *The Physics of Blown Sand and Desert Dunes*. Dover Earth Science. [Online].  
552 Dordrecht: Springer Netherlands. Available from: [http://link.springer.com/10.1007/978-94-](http://link.springer.com/10.1007/978-94-009-5682-7)  
553 [009-5682-7](http://link.springer.com/10.1007/978-94-009-5682-7).
- 554 Batchelor, G.K. (2000). *An Introduction to Fluid Dynamics*. [Online]. Cambridge University Press.  
555 Available from:  
556 <https://www.cambridge.org/core/product/identifier/9780511800955/type/book>.
- 557 Berger, W.H., & Piper, D.J.W.W., (1972). Planktonic Foraminifera: Differential Settling, Dissolution,  
558 and Redeposition. *Limnology and Oceanography*. [Online]. 17 (2). p.pp. 275–287. Available  
559 from: <http://doi.wiley.com/10.4319/lo.1972.17.2.0275>.
- 560 Bienfang, P.K. (1981). SETCOL — A Technologically Simple and Reliable Method for Measuring  
561 Phytoplankton Sinking Rates. *Canadian Journal of Fisheries and Aquatic Sciences*. [Online].  
562 38 (10). p.pp. 1289–1294. Available from:  
563 <http://www.nrcresearchpress.com/doi/10.1139/f81-173>.
- 564 Bloesch, J. (1995). Mechanisms, measurement and importance of sediment resuspension in lakes.  
565 *Marine and Freshwater Research*. [Online]. 46 (1). p.p. 295. Available from:  
566 <http://www.publish.csiro.au/?paper=MF9950295>.
- 567 Booth, J.D. (1983). Studies on twelve common bivalve larvae, with notes on bivalve spawning  
568 seasons in New Zealand. *New Zealand Journal of Marine and Freshwater Research*. [Online].  
569 17 (3). p.pp. 231–265. Available from:  
570 <http://www.tandfonline.com/doi/abs/10.1080/00288330.1983.9516001>.
- 571 Botte, V., Ribera D'Alcalà, M. & Montresor, M. (2013). Hydrodynamic interactions at low Reynolds  
572 number: An overlooked mechanism favouring diatom encounters. *Journal of Plankton*  
573 *Research*. [Online]. 35 (4). p.pp. 914–918. Available from:  
574 [http://academic.oup.com/plankt/article/35/4/914/1529648/Hydrodynamic-interactions-at-](http://academic.oup.com/plankt/article/35/4/914/1529648/Hydrodynamic-interactions-at-low-Reynolds-number)  
575 [low-Reynolds-number](http://academic.oup.com/plankt/article/35/4/914/1529648/Hydrodynamic-interactions-at-low-Reynolds-number).

576 Callieri, M., Ranzuglia, G., Dellepiane, M., Cignoni, P. & Scopigno, R. (2012). Meshlab as a Complete  
577 Open Tool for the Integration of Photos and Colour with High- Resolution 3D Geometry Data.  
578 Computer Applications and Quantitative Methods in Archaeology. p.pp. 406–416.

579 Caromel, A.G.M., Schmidt, D.N., Phillips, J.C. & Rayfield, E.J. (2014). Hydrodynamic constraints on the  
580 evolution and ecology of planktic foraminifera. *Marine Micropaleontology*. [Online]. 106.  
581 p.pp. 69–78. Available from:  
582 <http://linkinghub.elsevier.com/retrieve/pii/S037783981400005X>. [Accessed: 28 September  
583 2014].

584 Clift, R., Grace, J.R. & Weber, M.E. (1978). *Bubbles, Drops and Particles*. New York: Academic Press.

585 D’Errico, J. (2009). SLM - Shape Language Modeling. [Online]. 2009. MATLAB Central File Exchange.  
586 Available from: [http://www.mathworks.co.uk/matlabcentral/fileexchange/24443-slm-](http://www.mathworks.co.uk/matlabcentral/fileexchange/24443-slm-shape-language-modeling)  
587 [shape-language-modeling](http://www.mathworks.co.uk/matlabcentral/fileexchange/24443-slm-shape-language-modeling).

588 Durham, W.M., Kessler, J.O. & Stocker, R. (2009). Disruption of Vertical Motility by Shear Triggers  
589 Formation of Thin Phytoplankton Layers. *Science*. [Online]. 323 (5917). p.pp. 1067–1070.  
590 Available from: <https://www.sciencemag.org/lookup/doi/10.1126/science.1167334>.

591 Fayon, A.M. & Happel, J. (1960). Effect of a cylindrical boundary on a fixed rigid sphere in a moving  
592 viscous fluid. *AIChE Journal*. [Online]. 6 (1). p.pp. 55–58. Available from:  
593 <http://doi.wiley.com/10.1002/aic.690060111>.

594 Fok-Pun, L. & Komar, P.D. (1983). Settling velocities of planktonic foraminifera; density variations  
595 and shape effects. *The Journal of Foraminiferal Research*. [Online]. 13 (1). p.pp. 60–68.  
596 Available from: <https://pubs.geoscienceworld.org/jfr/article/13/1/60-68/76175>.

597 Gómez-Noguez, F., Pérez-García, B., Mehlreter, K., Orozco-Segovia, A. & Rosas-Pérez, I. (2016).  
598 Spore mass and morphometry of some fern species. *Flora: Morphology, Distribution,*  
599 *Functional Ecology of Plants*. [Online]. 223. p.pp. 99–105. Available from:  
600 <http://dx.doi.org/10.1016/j.flora.2016.05.003>.

601 Greving, I., Wilde, F., Ogurreck, M., Herzen, J., Hammel, J.U., Hipp, A., Friedrich, F., Lottermoser, L.,  
602 Dose, T., Burmester, H., Müller, M. & Beckmann, F. (2014). P05 imaging beamline at PETRA  
603 III: first results. In: S. R. Stock (ed.). *Developments in X-Ray Tomography IX*. [Online]. 12  
604 September 2014, San Diego, CA: SPIE, p. 92120O. Available from:  
605 <https://doi.org/10.1117/12.2061768>.

606 Guadayol, Ò. (2016). Trackbac. [Online]. Available from:  
607 <https://zenodo.org/record/45559#.W6z6c2j0nIU>.

608 Guadayol, Ò., Thornton, K.L. & Humphries, S. (2017). Cell morphology governs directional control in  
609 swimming bacteria. *Scientific Reports*. [Online]. 7 (1). p.p. 2061. Available from:  
610 <http://www.nature.com/articles/s41598-017-01565-y>.

611 B. K. Sen Gupta (ed.) (2002). *Modern Foraminifera*. Kluwer Academic Publishers.

612 Haibel, A., Ogurreck, M., Beckmann, F., Dose, T., Wilde, F., Herzen, J., Müller, M., Schreyer, A.,  
613 Nazmov, V., Simon, M., Last, A. & Mohr, J. (2010). Micro- and nano-tomography at the GKSS  
614 Imaging Beamline at PETRA III. In: S. R. Stock (ed.). [Online]. 19 August 2010, p. 78040B.  
615 Available from:  
616 <http://proceedings.spiedigitallibrary.org/proceeding.aspx?doi=10.1117/12.860852>.

617 Happel, J. & Brenner, H. (1983). *Low Reynolds number hydrodynamics with special applications to*  
618 *particulate media*. Third. The Hague: Martin Nijhoff Publishers a member of Kluwer  
619 Academic Publishers Group.

620 Hayward, B.W., Le Coze, F. & Gross, O. (2018). *World Foraminifera Database*. [Online]. 2018.  
621 Available from: <http://www.marinespecies.org/foraminifera/>. [Accessed: 23 September  
622 2020].

623 Holland, D.P. (2010). Sinking rates of phytoplankton filaments orientated at different angles: theory  
624 and physical model. *Journal of Plankton Research*. [Online]. 32 (9). p.pp. 1327–1336.  
625 Available from: [https://academic.oup.com/plankt/article-](https://academic.oup.com/plankt/article-lookup/doi/10.1093/plankt/fbq044)  
626 [lookup/doi/10.1093/plankt/fbq044](https://academic.oup.com/plankt/article-lookup/doi/10.1093/plankt/fbq044).

627 Huesman, R.H., Gullberg, W.L., Greenberg, W.L. & Budinger, T.F. (1977). *RECLBL Users Manual:*  
628 *Donner Algorithms for Reconstruction Tomography*. [Online]. Available from:  
629 <https://escholarship.org/uc/item/1mz679x3#main>.

630 Iversen, M.H. & Ploug, H. (2010). Ballast minerals and the sinking carbon flux in the ocean: Carbon-  
631 specific respiration rates and sinking velocity of marine snow aggregates. *Biogeosciences*. 7  
632 (9). p.pp. 2613–2624.

633 Jenkinson, I.R., Sun, X. & Seuront, L. (2015). Horizons: Thalassorheology, organic matter and  
634 plankton: Towards a more viscous approach in plankton ecology. *Journal of Plankton*  
635 *Research*. 37 (6). p.pp. 1100–1109.

636 Jones, R., Ricardo, G.F. & Negri, A.P. (2015). Effects of sediments on the reproductive cycle of corals.  
637 *Marine Pollution Bulletin*. [Online]. 100 (1). p.pp. 13–33. Available from:  
638 <http://dx.doi.org/10.1016/j.marpolbul.2015.08.021>.

639 Jonkers, L. & Kučera, M. (2015). Global analysis of seasonality in the shell flux of extant planktonic  
640 Foraminifera. *Biogeosciences*. [Online]. 12 (7). p.pp. 2207–2226. Available from:  
641 <http://www.biogeosciences.net/12/2207/2015/>.

642 Karamanev, D.G. (1996). Equations for calculation of the terminal velocity and drag coefficient of  
643 solid spheres and gas bubbles. *Chemical Engineering Communications*. 147. p.pp. 75–84.

644 Karp-Boss, L. & Jumars, P.A. (1998). Motion of diatom chains in steady shear flow. *Limnology and*  
645 *Oceanography*. [Online]. 43 (8). p.pp. 1767–1773. Available from:  
646 <http://doi.wiley.com/10.4319/lo.1998.43.8.1767>.

647 Kim, M.J., Bird, J.C., Van Parys, A.J., Breuer, K.S. & Powers, T.R. (2003). A macroscopic scale model of  
648 bacterial flagellar bundling. *Proceedings of the National Academy of Sciences of the United*  
649 *States of America*. 100 (26). p.pp. 15481–15485.

650 Koehl, M.A.R. (1995). Fluid flow through hair-bearing appendages: feeding, smelling, and swimming  
651 at low and intermediate Reynolds number. In: C. P. Ellington & T. J. Pedley (eds.). *Biological*  
652 *Fluid Dynamics*. Cambridge: The Company of Biologists Limited, pp. 157–182.

653 Koehl, M.A.R. (2003). Physical modelling in biomechanics J. van Leeuwen & P. Aerts (eds.).  
654 *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*.  
655 [Online]. 358 (1437). p.pp. 1589–1596. Available from:  
656 [http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1693254&tool=pmcentrez&ren](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1693254&tool=pmcentrez&rendertype=abstract)  
657 [dertype=abstract](http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=1693254&tool=pmcentrez&rendertype=abstract). [Accessed: 27 October 2014].

658 Kucera, M. (2007). Chapter Six Planktonic Foraminifera as Tracers of Past Oceanic Environments. In:  
659 *Developments in Marine Geology*. [Online]. London: Elsevier, pp. 213–262. Available from:  
660 <http://linkinghub.elsevier.com/retrieve/pii/S1572548007010111>.

661 De La Rocha, C.L. & Passow, U. (2007). Factors influencing the sinking of POC and the efficiency of  
662 the biological carbon pump. *Deep Sea Research Part II: Topical Studies in Oceanography*.  
663 [Online]. 54 (5–7). p.pp. 639–658. Available from:  
664 <https://linkinghub.elsevier.com/retrieve/pii/S0967064507000392>.

665 Lohmann, G.P. (1983). Eigenshape analysis of microfossils: A general morphometric procedure for  
666 describing changes in shape. *Journal of the International Association for Mathematical*  
667 *Geology*. 15 (6). p.pp. 659–672.

668 McAndrew, A. (2004). *An Introduction to Digital Image Processing with Matlab Notes for SCM2511*  
669 *Image Processing 1 Semester 1 , 2004*.

670 Miklasz, K.A. & Denny, M.W. (2010). Diatom sinkings speeds: Improved predictions and insight from  
671 a modified Stokes' law. *Limnology and Oceanography*. [Online]. 55 (6). p.pp. 2513–2525.  
672 Available from: <http://doi.wiley.com/10.4319/lo.2010.55.6.2513>.

673 Morard, R., Quillévéré, F., Escarguel, G., de Garidel-Thoron, T., de Vargas, C. & Kucera, M. (2013).  
674 Ecological modeling of the temperature dependence of cryptic species of planktonic  
675 Foraminifera in the Southern Hemisphere. *Palaeogeography, Palaeoclimatology,*



676 Palaeoecology. [Online]. 391. p.pp. 13–33. Available from:  
677 <http://dx.doi.org/10.1016/j.palaeo.2013.05.011>.

678 Morrison, F.A. (2013). *An Introduction to Fluid Mechanics*. Cambridge: Cambridge University Press.

679 Morrison, F.A. (2010). Data correlation for drag coefficient for sphere. Michigan Technology  
680 University, Houghton, MI. [Online]. 6 (April). p.pp. 1–2. Available from:  
681 <http://www.chem.mtu.edu/~fmorriso/DataCorrelationForSphereDrag2013.pdf>.

682 Munson, B.R., Young, D.F. & Okiishi, T.H. (1994). *Fundamentals of Fluid Mechanics*. Second. New  
683 York: John Wiley and Sons, Inc.

684 Noblin, X., Yang, S. & Dumais, J. (2009). Surface tension propulsion of fungal spores. *Journal of*  
685 *Experimental Biology*. 212 (17). p.pp. 2835–2843.

686 Oskui, S.M., Bhakta, H.C., Diamante, G., Liu, H., Schlenk, D. & Grover, W.H. (2017). Measuring the  
687 mass, volume, and density of microgram-sized objects in fluid. *PLoS ONE*. 12 (4). p.pp. 1–17.

688 Padisák, J., Soróczki-Pintér, É. & Rezner, Z. (2003). Sinking properties of some phytoplankton shapes  
689 and the relation of form resistance to morphological diversity of plankton – an experimental  
690 study. *Hydrobiologia*. [Online]. 500 (1–3). p.pp. 243–257. Available from:  
691 <http://link.springer.com/10.1023/A:1024613001147>.

692 Passow, U. & Carlson, C. a. (2012). The biological pump in a high CO<sub>2</sub> world. *Marine Ecology Progress*  
693 *Series*. 470 (2). p.pp. 249–271.

694 Purcell, E.M. (1977). Life at low Reynolds number. *American Journal of Physics*. 45 (1). p.pp. 3–11.

695 Reidenbach, M.A., George, N. & Koehl, M.A.R. (2008). Antennule morphology and flicking kinematics  
696 facilitate odor sampling by the spiny lobster, *Panulirus argus*. *Journal of Experimental*  
697 *Biology*. [Online]. 211 (17). p.pp. 2849–2858. Available from:  
698 <http://jeb.biologists.org/cgi/doi/10.1242/jeb.016394>.

699 Schiebel, R. (2002). Planktic foraminiferal sedimentation and the marine calcite budget. *Global*  
700 *Biogeochemical Cycles*. [Online]. 16 (4). p.pp. 3-1-3–21. Available from:  
701 <http://doi.wiley.com/10.1029/2001GB001459>.

702 Schiebel, R. & Hemleben, C. (2005). Modern planktic foraminifera. *Paläontologische Zeitschrift*.  
703 [Online]. 79 (1). p.pp. 135–148. Available from:  
704 <http://link.springer.com/10.1007/BF03021758>.

705 Van Sebille, E., Scussolini, P., Durgadoo, J. V., Peeters, F.J.C.C., Biastoch, A., Weijer, W., Turney, C.,  
706 Paris, C.B. & Zahn, R. (2015). Ocean currents generate large footprints in marine  
707 palaeoclimate proxies. *Nature Communications*. [Online]. 6 (1vm). p.p. 6521. Available from:  
708 <http://dx.doi.org/10.1038/ncomms7521>.

709 Smayda, T.J. (1971). Normal and accelerated sinking of phytoplankton in the sea. *Marine Geology*. 11  
710 (2). p.pp. 105–122.

711 Sowa, Y. & Berry, R.M. (2008). Bacterial flagellar motor. *Quarterly Reviews of Biophysics*. [Online]. 41  
712 (2). p.pp. 103–132. Available from:  
713 [https://www.cambridge.org/core/product/identifier/S0033583508004691/type/journal\\_arti](https://www.cambridge.org/core/product/identifier/S0033583508004691/type/journal_article)  
714 [cle](https://www.cambridge.org/core/product/identifier/S0033583508004691/type/journal_article).

715 Stacey, M.T., Mead, K.S. & Koehl, M.A.R. (2002). Molecule capture by olfactory antennules: Mantis  
716 shrimp. *Journal of Mathematical Biology*. [Online]. 44 (1). p.pp. 1–30. Available from:  
717 <http://link.springer.com/10.1007/s002850100111>.

718 Stadler, A.T., Vihar, B., Günther, M., Huemer, M., Riedl, M., Shamiyeh, S., Mayrhofer, B., Böhme, W.  
719 & Baumgartner, W. (2016). Adaptation to life in aeolian sand: how the sandfish lizard,  
720 *Scincus scincus*, prevents sand particles from entering its lungs. *The Journal of Experimental*  
721 *Biology*. [Online]. 219 (22). p.pp. 3597–3604. Available from:  
722 <http://jeb.biologists.org/lookup/doi/10.1242/jeb.138107>.

723 Sutton, M.D., Garwood, R.J., Siveter, D.J. & Siveter, D.J. (2012). SPIERS and VAXML; A software  
724 toolkit for tomographic visualisation and a format for virtual specimen interchange.  
725 *Paleontologica electronica*. 15 (2). p.pp. 1–15.

726 Takahashi, K. & Be, A.W.H. (1984). Planktonic foraminifera: factors controlling sinking speeds. *Deep*  
727 *Sea Research Part A. Oceanographic Research Papers*. [Online]. 31 (12). p.pp. 1477–1500.  
728 Available from: <https://linkinghub.elsevier.com/retrieve/pii/0198014984900839>.

729 Takahashi, K. & Honjo, S. (1983). Radiolarian skeletons: size, weight, sinking speed, and residence  
730 time in tropical pelagic oceans. *Deep Sea Research Part A. Oceanographic Research Papers*.  
731 [Online]. 30 (5). p.pp. 543–568. Available from:  
732 <https://linkinghub.elsevier.com/retrieve/pii/0198014983900882>.

733 Vogel, S. (1987). Flow-Assisted Mantle Cavity Refilling in Jetting Squid. *The Biological Bulletin*.  
734 [Online]. 172 (1). p.pp. 61–68. Available from:  
735 <https://www.journals.uchicago.edu/doi/10.2307/1541606>.

736 Vogel, S. (1994). *Life in Moving Fluids*. Second. Princeton, N.J., N.J.: Princeton University Press.

737 Vogel, S., Ellington, C.P. & Kilgore, D.L. (1973). Wind-induced ventilation of the burrow of the prairie-  
738 dog, *Cynomys ludovicianus*. *Journal of comparative physiology*. [Online]. 85 (1). p.pp. 1–14.  
739 Available from: <http://link.springer.com/10.1007/BF00694136>.

740 Vogel, S. & LaBarbera, M. (1978). Simple Flow Tanks for Research and Teaching. *BioScience*. [Online].  
741 28 (10). p.pp. 638–643. Available from: [https://academic.oup.com/bioscience/article-](https://academic.oup.com/bioscience/article-lookup/doi/10.2307/1307394)  
742 [lookup/doi/10.2307/1307394](https://academic.oup.com/bioscience/article-lookup/doi/10.2307/1307394).

743 Waldrop, L.D., Reidenbach, M.A. & Koehl, M.A.R. (2015). Flexibility of crab chemosensory sensilla  
744 enables flicking antennules to sniff. *Biological Bulletin*. 229 (2). p.pp. 185–198.

745 Walsby, A.E. & Holland, D.P. (2006). Sinking velocities of phytoplankton measured on a stable  
746 density gradient by laser scanning. *Journal of The Royal Society Interface*. [Online]. 3 (8).  
747 p.pp. 429–439. Available from:  
748 <https://royalsocietypublishing.org/doi/10.1098/rsif.2005.0106>.

749 Waniek, J., Koeve, W. & Prien, R.D. (2000). Trajectories of sinking particles and the catchment areas  
750 above sediment traps in the northeast Atlantic. *Journal of Marine Research*. [Online]. 58 (6).  
751 p.pp. 983–1006. Available from:  
752 <http://hermia.ingentaselect.com/vl=6505216/cl=63/nw=1/rpsv/cw/jmr/00222402/v58n6/s6>  
753 [/p983](http://hermia.ingentaselect.com/vl=6505216/cl=63/nw=1/rpsv/cw/jmr/00222402/v58n6/s6).

754 Wilde, F., Ogurreck, M., Greving, I., Hammel, J.U., Beckmann, F., Hipp, A., Lottermoser, L.,  
755 Khokhriakov, I., Lytaev, P., Dose, T., Burmester, H., Müller, M. & Schreyer, A. (2016). Micro-  
756 CT at the imaging beamline P05 at PETRA III. *AIP Conference Proceedings*. 1741 (July).

757 Willson, M.F., Rice, B.L. & Westoby, M. (1990). Seed dispersal spectra: a comparison of temperate  
758 plant communities. *Journal of Vegetation Science*. [Online]. 1 (4). p.pp. 547–562. Available  
759 from: <http://doi.wiley.com/10.2307/3235789>.

760 Zohuri, B. (2015). *Dimensional Analysis and Self-Similarity Methods for Engineers and Scientists*.  
761 [Online]. Cham: Springer International Publishing. Available from:  
762 <http://link.springer.com/10.1007/978-3-319-13476-5>

## 763 Figure & Table Legends

### 764 Figure 1

765 *Fig. 1.A) Diagram of relevant forces and parameters between the model (left) and real life (right).*  
766 *B) Summary of the full method; details are discussed in main text. Boxes with thicker lines represent a decision, square boxes*  
767 *are data inputs, rounded square boxes are manual processes, and circles are computational steps.*

### 768 Figure 2

769 *Fig. 2 A) Comparison of our empirically generated  $C_D^E(Re)$  curve for 3D printed spheres versus the theoretical  $C_D^M(Re)$  curve*  
770 *(Morrison, 2010) . Goodness of fit of  $C_D^E(Re)$  to  $C_D^M(Re)$  is  $R^2 = 0.857$ . Sphere diameter is indicated for each model.*  
771 *C & D) An example of our iterative solution process for *C. dissimilis* showing best estimates of operating values (including*  
772 *the required model scale  $S$  to achieve similitude) based on experimental data from 3 (C), vs 4 (D) models. For reference, the*  
773 *theoretical  $C_D^M(Re)$  curve (Morrison, 2010) for a sphere is also shown. In C,  $S$  corresponding to the operating point is*  
774 *estimated as 13.84. After an additional model was sunk at  $S = 13$  (D), slightly more accurate estimates of the operating  $S$ ,*  
775  *$Re^0$ , and  $C_D^0$  were obtained. When scaling the model for 3D printing only 1 decimal place was used rather than the 2 shown.*  
776 *D-I) Models of foraminifera 3D printed in clear and black resin. Models were used for public engagement but demonstrate*  
777 *the fidelity of the printer compared to the scan data shown Table 1.*  
778

### 779 Table 1

780 *Table 1. Predicted sinking speeds  $U^0$  for the 30 species of planktonic foraminifera included in this study, with each species*  
781 *shown in both spiral view and  $90^\circ$  rotation (so that the spiral view is facing to the left). <sup>1</sup> indicates species scanned high*  
782 *resolution. Scans of the remaining 27 species were obtained from The University of Tohoku museum's database. Operating*  
783  *$Re^0$  and  $C_D^0$  predicted for each species are also presented, and for comparison, the theoretical  $C_D^M$  of a sphere at the same*  
784  *$Re^0$ . The number of model iterations required to achieve convergence of  $U^0$  is listed (iter). Species synonyms checked at*  
785 *World Foraminifera Database (Hayward et al., 2018).*