

# Antibodies to nodal/paranodal proteins in paediatric immune-mediated neuropathy

Desiree De Simoni, MD, Gerda Ricken, MSc, Michael Winklehner, MD, Inga Koneczny, PhD, Michael Karenfort, MD, Ulf Hustedt, MD, Ulrich Seidel, MD, Omar Abdel-Mannan, MD, Pinki Munot, MD, Simon Rinaldi, MD, Claudia Steen, MD, Michael Freiling, MD, Markus Breu, MD, Rainer Seidl, MD, Markus Reindl, PhD, Julia Wanschitz, MD, Cinta Lleixà, PhD, Günther Bernert, MD, Klaus-Peter Wandinger, MD, PhD, Ralf Junker, MD, Luis Querol, MD, PhD, Frank Leypoldt, MD, PhD, Kevin Rostásy, MD,\* and Romana Höftberger, MD\*

## Correspondence

Dr. Rostásy  
k.rostasy@kinderklinik-datteln.de  
or Dr. Höftberger  
romana.hoeftberger@  
meduniwien.ac.at

*Neurol Neuroimmunol Neuroinflamm* 2020;7:e763. doi:10.1212/NXI.0000000000000763

Patients with nodal/paranodal antibodies represent a specific subgroup of inflammatory peripheral neuropathies, whose clinical presentation with a prolonged subacute phase, additional symptoms such as ataxia and tremor, and poor treatment response to IV immunoglobulin (IVIG) often differs from classic Guillain-Barré syndrome (GBS) or chronic inflammatory demyelinating polyneuropathy (CIDP).<sup>1</sup>

Previous studies on nodo/paranodopathies mainly focused on adult patients, whereas the clinical spectrum of pediatric patients is less well established. We reviewed the clinical presentation of 54 children with GBS (n = 42) and CIDP (n = 12) and retrospectively screened for antibodies against neurofascin155 (NF155), NF186, NF140, contactin-1 (CNTN1), contactin-associated protein1 (CASPR1), and glycine-receptor (GlyR) using cell-based assays<sup>2,3</sup>; 1 patient was additionally tested with CNTN1-ELISA.<sup>4</sup> All cases with sufficient serum were tested for ganglioside-IgG-, IgA-, and IgM-antibodies against GM1 (n = 42), GD1a (n = 18), GD1b (n = 23), and GQ1b (n = 21).<sup>5</sup> Clinical and paraclinical information of all patients is summarized in the table. The study was approved by the ethics committee (EK1773/2016).

## Children with classic GBS

Of 42 children with GBS, 26 were classified as acute inflammatory demyelinating polyneuropathy (AIDP), 7 as acute motor/motor-sensory axonal neuropathy (AMAN/AMSAN) by nerve conduction velocity according to Hadden criteria,<sup>6</sup> 4 as Miller-Fisher syndrome (MFS), and 2 as MFS/GBS overlap. Three patients with GBS could not be classified because of lack of nerve-conduction studies. In 25 of 35 patients (71.4%), an infection was reported within 4 weeks before symptom onset (13 gastrointestinal, 4 respiratory, and 8 unspecified). Eight patients had IgG-ganglioside antibodies (19.0%), 6 IgM (14.2%), and 1 IgA (2.4%). Nodal/paranodal antibodies were not detected. Patients with AMAN/AMSAN (5/7 with reported infection: 1 campylobacter jejuni, 1 varicella-zoster virus, and 3 unspecified) were more often ganglioside antibody positive (6/7) than patients with AIDP (4/26; likelihood ratio 12.419) or MFS (2/4).

\*These authors contributed equally to this work.

From the Division of Neuropathology and Neurochemistry (D.D.S., G.R., M.W., I.K., R.H.), Department of Neurology, Medical University of Vienna, Austria; Department of Neurology (D.D.S.), University Hospital St. Poelten, Austria; Department of General Pediatrics, Neonatology and Pediatric Cardiology (M.K.), University Children's Hospital, Heinrich Heine University Duesseldorf, Germany; Department of Neuropediatric Rehabilitation (U.H.), Vamed Clinic Hattingen, Germany; Department of Neuropediatrics (U.S.), Charité University, Berlin, Germany; Paediatric Neurology (O.A.-M.), Great Ormond Street Hospital for Children, London, United Kingdom; Dubowitz Neuromuscular Centre (P.M.), Great Ormond Street Hospital for Children, London, United Kingdom; Nuffield Department of Clinical Neurosciences (S.R.), University of Oxford and Oxford University Hospitals NHS Foundation Trust; Department of Paediatric and Adolescent Medicine (C.S.), St Joseph Hospital, Berlin, Germany; Department of Pediatrics and Adolescent Medicine (M.F., M.B., R.S.), Medical University of Vienna, Austria; Department of Neurology (M.R., J.W.), Medical University of Innsbruck, Austria; Neuromuscular Diseases Unit (C.L., L.Q.), Hospital de la Santa Creu i Sant Pau, Universitat Autònoma de Barcelona, Spain; SMZ Süd (G.B.), Kaiser-Franz Josef Hospital with Gottfried von Preyer Children Hospital, Vienna, Austria; Institute of Clinical Chemistry (K.-P.W., R.J., F.L.), University Hospital Schleswig-Holstein, Kiel/Lübeck, Germany; Department of Neurology (F.L.), University Hospital Schleswig-Holstein, Kiel, Germany; and Department of Pediatric Neurology (K.R.), Witten/Herdecke University, Children's Hospital Datteln, Germany.

Go to [Neurology.org/NN](https://Neurology.org/NN) for full disclosures. Funding information is provided at the end of the article.

The Article Processing Charge was funded by Austrian Science Fund.

This is an open access article distributed under the terms of the Creative Commons Attribution License 4.0 (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Table** Clinical and paraclinical data of patients with GBS and CIDP

	GBS (42)		CIDP (12)	
	Ganglioside abs pos. (IgG/IgM/IgA)	Seronegative	Nodal/paranodal antibodies pos.	Seronegative
<b>No. of patients</b>	15	27	5	7
<b>Age mean (range)</b>	11.6 (4–17)	10.22 (1–18)	7.9 (3–11)	10.4 (4–18)
<b>Gender m:f</b>	9:6	15:12	3:2	4:3
<b>AIDP</b>	4	22		
<b>AMAN/AMSAN</b>	6	1		
<b>MFS/MFS overlap</b>	3/1	1/1		
<b>GBS no NCS</b>	1	2		
<b>GM1+</b>	6	—	0	0
<b>GD1a+</b>	1		0	0
<b>GD1b+</b>	1		0	0
<b>GM1+GD1b+</b>	4		0	0
<b>GQ1b+</b>	1		0	0
<b>GM1+GD1a+ GQ1b+</b>	1			
<b>GD1a+GQ1b+</b>	1			
<b>Pan-neurofascin+</b>	—	—	2	0
<b>NF155+</b>			1	0
<b>CNTN1+</b>			2	0
<b>CSF mean cell count/<math>\mu</math>L (range)</b>	3.15 (0–11)	2.92 (0–11)	4.6 (0–21)	3.8 (1–9)
<b>CSF mean total protein mg/dL (range)</b>	98.08 (10–250)	118.59 (19–401)	292.4 (75–619)	107.7 (24–288)
<b>Infection (data available from 45/54)</b>	11 (2 C. jejuni; 1 VZV; 8 unspecified)	14 (2 C. jejuni; 3 VZV; 1 EBV; 8 unspecified)	1	2
<b>GI</b>	6	7	1	0
<b>Respiratory</b>	2	2	0	2
<b>Other</b>	3	5	0	0
<b>Infection d prior mean (range)</b>	9 (1–14)	11.6 (3–28)	0	4.8 (1–10)
<b>Days hospitalization mean (range)</b>	13.43 (3–30)	20.73 (0–135)	13 (2–28)	10.4 (2–16)
<b>Cranial nerve involvement</b>	3	7	1	2
<b>Autonomic dysfunction</b>	1	2	0	0
<b>Tremor/ataxia</b>	3	3	5	1
<b>Outcome mRS (available from 32/42 GBS and 11/12 CIDP)</b>	11 mRS 0–1; 1 mRS 2–4; 1 mRS >1;	15 mRS 0–1; 4 mRS 2–4; 1 mRS 5–6; 5 mRS >1;	2 mRS 0–1; 3 mRS 2–4; 3 mRS >1;	2 mRS 0–1; 4 mRS 2–4; 4 mRS >1;
<b>Severity at nadir HS (available from 41/42 GBS and 12/12 CIDP)</b>	2 HS1; 8 HS2; 1 HS3; 2 HS4; 1 HS5	2 HS1; 5 HS2; 7 HS3; 6 HS4; 6 HS5; 1 HS6	3 HS3; 2 HS4	2 HS1; 1 HS2; 4 HS4

Abbreviations: AIDP = acute inflammatory demyelinating polyneuropathy; AMAN = acute motor axonal neuropathy; AMSAN = acute motor and sensory axonal neuropathy; C. jejuni = campylobacter jejuni; CIDP = chronic inflammatory demyelinating polyneuropathy; EBV = Epstein-Barr virus; GBS = Guillain-Barré syndrome; GI = gastrointestinal; HS = Hughes score; MFS = Miller-Fisher syndrome; mRS = modified Rankin Scale; NCS = nerve-conduction study; VZV = varicella-zoster-virus.

## Children with nodal/paranodal antibodies

Five of 12 children, who met the EFNS/PNS criteria for CIPD, had nodal/paranodal antibodies: 2 pan-neurofascin (NF155/NF186/140 triple positive), 1 NF155, and 2 CNTN1-antibodies. The IgG-subclass distribution was determined by flow cytometry analysis.<sup>7</sup> IgG4 was the predominant subclass in all patients and ranged from 75% to 100%. In addition, 1 patient with pan-neurofascin-antibodies tested positive for GlyR-antibodies but did not develop stiff-person syndrome or progressive encephalomyelitis with rigidity, and the significance of this finding needs further investigation. The mean age was 7.9 years (range 3–11), and the male:female ratio was 3:2. The median duration of hospitalization was 13 days (range 2–28). One pan-neurofascin-patient was initially diagnosed as GBS and reclassified as CIPD during disease course, the other patients had a chronic onset with slow progression over months or years. One child had a gastrointestinal infection before symptom onset. One CNTN1-patient showed cranial nerve involvement and optic neuritis during disease course. All children had ataxia, 4 neuropathic pain (all except 1 pan-neurofascin), and 3 (2 CNTN1, and 1 pan-neurofascin) tremor. At the peak of disease, 3 children needed a walking aid (Hughes 3) and 2 were bedridden (Hughes 4). None of the children had renal dysfunction. The mean CSF white cell count was 4.6  $\mu$ L (range 0–21), and the mean CSF protein was 292.4 mg/dL (range 75–619).

The mean time of follow-up was 32 months (range 17–57). The 2 CIPD patients with pan-neurofascin-antibodies initially showed no or only partial response to IVIG and therefore received corticosteroids, 1 along with plasma exchange and the other with mycophenolate. Both recovered only very slowly over up to 4 years with a modified Rankin Scale (mRS) score of 1 at the last follow-up. The NF155-patient did not respond to IVIG and corticosteroids and subsequently received immunoadsorption and rituximab, leading to significant clinical improvement. After 8 months, he relapsed in association with normalization of the CD19/20 ratio and again rapidly improved after another dose of rituximab, with a mRS score of 2 at the last follow-up. One patient with CNTN1-antibodies worsened despite monthly IVIG and corticosteroids given over 4 months. After treatment was switched to rituximab, he improved rapidly in the following weeks and remained stable since then. The second child with CNTN1-antibodies showed only partial response to IVIG with relapses in conjunction with infections. This child improved significantly after rituximab application with a mRS score of 2 at the last follow-up.

In summary, our study demonstrates that nodal/paranodal antibodies occur in a subgroup of paediatric patients with CIPD, but not GBS. Children with AMAN/AMSAN frequently have ganglioside antibodies. Children with CIPD and atypical/prolonged disease course with high Hughes score (>2), sensory ataxia, prominent neuropathic pain, and tremor may have nodal/paranodal antibodies. These patients often

do not sufficiently respond to IVIG, whereas in our case series, rituximab led to prompt improvement in 3 children. Optimal treatment strategies for children with nodal/paranodal antibodies have to be further determined in larger studies.

### Acknowledgment

The authors thank Valerie Pichler and Sophie Dürauer for technical assistance and Andreas Spittler from the Flow Cytometry Core facility of the MUV for technical support.

### Study funding

This work was partly supported by grants from the “Jubiläumsfonds der Österreichischen Nationalbank,” project 16919 (R. Höftberger), the GBS/CIDP Foundation International (J. Wanschitz), Austrian Science Fund FWF, DOC 33-B27 (R. Höftberger, M. Winklehner) and I3334-B27 (R. Höftberger), Hertha Firnberg project number T996-B30 (I. Koneczny), the PI16/000627 grant of the Fondo de Investigaciones Sanitarias—Instituto de Salud Carlos III (fondos FEDER) (L. Querol), personal grant SLT006/17/00131 of the Pla estratègic de Recerca i Innovació en Salut (PERIS), Departament de Salut, Generalitat de Catalunya (L. Querol), and the German Ministry of Education and Research (BMBF, 01 GM1908A).

### Disclosure

D. De Simoni, G. Ricken, M. Winklehner, I. Koneczny, M. Karenfort, U. Hustedt, U. Seidel, O. Abdel-Mannan, P. Munot, S. Rinaldi, C. Steen, M. Freilinger, M. Breu, R. Seidl, M. Reindl, J. Wanschitz, C. Lleixá, G. Bernert, K.P. Wandinger, R. Junker, L. Querol, F. Leypoldt, K. Rostásy, and R. Höftberger report no disclosures relevant to the manuscript. Go to [Neurology.org/NN](http://Neurology.org/NN) for full disclosures.

### Publication history

Received by *Neurology: Neuroimmunology & Neuroinflammation* March 8, 2020. Accepted in final form April 29, 2020.

### Appendix Authors

Name	Location	Contribution
<b>Desiree De Simoni, MD</b>	Medical University of Vienna, Austria; University Hospital St. Poelten, Austria	Acquisition of data, statistical analysis, execution, and interpretation of data
<b>Gerda Ricken, MSc</b>	Medical University of Vienna, Austria	Acquisition of data, execution, interpretation of data, and critical review for important intellectual content
<b>Michael Winklehner, MD</b>	Medical University of Vienna, Austria	Statistical analysis and critical review for important intellectual content
<b>Inga Koneczny, PhD</b>	Medical University of Vienna, Austria	Acquisition of data, execution, interpretation of data, and critical review for important intellectual content

Continued

## Appendix (continued)

Name	Location	Contribution
<b>Michael Karenfort, MD</b>	University Children's Hospital, Heinrich Heine University Duesseldorf, Germany	Acquisition of data and critical review for important intellectual content
<b>Ulf Hustedt, MD</b>	Vamed Clinic Hattingen, Germany	Acquisition of data and critical review for important intellectual content
<b>Ulrich Seidel, MD</b>	Charité University, Berlin, Germany	Acquisition of data and critical review for important intellectual content
<b>Omar Abdel-Mannan, MD</b>	Great Ormond Street Hospital for Children, London, United Kingdom	Acquisition of data and critical review for important intellectual content
<b>Pinki Munot, MD</b>	Great Ormond Street Hospital for Children, London, United Kingdom	Acquisition of data and critical review for important intellectual content
<b>Simon Rinaldi, MD</b>	University of Oxford and Oxford University Hospitals NHS Foundation Trust	Acquisition of data and critical review for important intellectual content
<b>Claudia Steen, MD</b>	St Joseph Hospital, Berlin, Germany	Acquisition of data and critical review for important intellectual content
<b>Michael Freilinger, MD</b>	Medical University of Vienna, Austria	Acquisition of data and critical review for important intellectual content
<b>Markus Breu, MD</b>	Medical University of Vienna, Austria	Acquisition of data and critical review for important intellectual content
<b>Rainer Seidl, MD</b>	Medical University of Vienna, Austria	Acquisition of data and critical review for important intellectual content
<b>Markus Reindl, PhD</b>	Medical University of Innsbruck, Austria	Acquisition of data, interpretation of data, and critical review for important intellectual content
<b>Julia Wanschitz, MD</b>	Medical University of Innsbruck, Austria	Acquisition of data and critical review for important intellectual content
<b>Cinta Lleixà, PhD</b>	Hospital de la Santa Creu i Sant Pau, Universitat Autònoma de Barcelona, Barcelona, Spain	Acquisition of data and interpretation of data

## Appendix (continued)

Name	Location	Contribution
<b>Günther Bernert, MD</b>	Kaiser-Franz Josef Hospital with Gottfried von Preyer children Hospital, Vienna, Austria	Acquisition of data and critical review for important intellectual content
<b>Klaus-Peter Wandinger, MD, PhD</b>	University Hospital Schleswig-Holstein, Kiel/Lübeck, Germany	Acquisition of data and critical review for important intellectual content
<b>Ralf Junker, MD</b>	University Hospital Schleswig-Holstein, Kiel/Lübeck, Germany	Acquisition of data and critical review for important intellectual content
<b>Luis Querol, MD, PhD</b>	Hospital de la Santa Creu i Sant Pau, Universitat Autònoma de Barcelona, Barcelona, Spain	Acquisition of data, execution, interpretation of data, and critical review for important intellectual content
<b>Frank Leyboldt, MD, PhD</b>	University Hospital Schleswig-Holstein, Kiel/Lübeck, Germany	Conception and design, acquisition of data, execution, interpretation of data, and critical review for important intellectual content
<b>Kevin Rostásy, MD</b>	Witten/Herdecke University, Children's Hospital Datteln, Germany	Conception and design, acquisition of data, statistical analysis, execution, interpretation of data, and critical review for important intellectual content
<b>Romana Höftberger, MD</b>	Medical University of Vienna, Austria	Conception and design, acquisition of data, execution, interpretation of data, and critical review for important intellectual content

## References

1. Querol L, Devaux J, Rojas-Garcia R, Illa I. Autoantibodies in chronic inflammatory neuropathies: diagnostic and therapeutic implications. *Nat Rev Neurol* 2017;13:533–547.
2. Martin-Aguilar L, Pascual-Goni E, Lleixa C, et al. Antibodies against nodo-paranodal proteins are not present in genetic neuropathies. *Neurology Epub* 2020 Feb 26.
3. Martinez-Hernandez E, Sepulveda M, Rostásy K, et al. Antibodies to aquaporin 4, myelin-oligodendrocyte glycoprotein, and the glycine receptor  $\alpha 1$  subunit in patients with isolated optic neuritis. *JAMA Neurol* 2015;72:187.
4. Mathey EK, Garg N, Park SB, et al. Autoantibody responses to nodal and paranodal antigens in chronic inflammatory neuropathies. *J Neuroimmunol* 2017;309:41–46.
5. Yuki N, Tagawa Y, Irie F, et al. Close association of Guillain-Barré syndrome with antibodies to minor monosialogangliosides GM1b and GM1a. *J Neuroimmunol* 1997;74:30–34.
6. Hadden R, Cornblath D, Hughes R, et al. Electrophysiological classification of Guillain-Barré syndrome: clinical associations and outcome. *Ann Neurol* 1998;44:780–788.
7. Ogata H, Yamasaki R, Hiwatashi A, et al. Characterization of IgG4 anti-neurofascin 155 antibody-positive polyneuropathy. *Ann Clin Transl Neurol* 2015;2:960–971.