



Hepcidin Response to Exercise: A Review

Egzersize Hepsidin Yanıtı: Derleme

Raúl Domínguez, Davinia Vicente-Campos*, Davinia Vicente-Campos**, José Chicharro**

Alfonso X University Faculty of Physical Activity and Sports Sciences, Villanueva de la Cañada, Madrid, Spain

*Francisco de Vitoria University Faculty of Physical Activity and Sports Sciences, Pozuelo de Alarcon, Madrid, Spain

**Complutense University, FEBIO Group, Madrid, Spain

Abstract

Given the multiple functions of iron in the body, any state of iron deficiency will induce a series of secondary effects that could compromise sports performance. Low serum iron levels are commonly observed in athletes during the course of a training period, especially in those performing aerobic exercises and resistance training. Sometimes, body iron levels will even fall below those detected in sedentary individuals, and we could go as far as to say that iron deficiency is the most frequently observed nutrition disorder among athletes of any sport. Hepsidin, a hormone secreted by hepatocytes whose principal mechanism of action is the degradation of ferroportin (the main iron exporter from macrophages and the basolateral membrane of duodenal enterocytes), has been proposed as the main regulator of the body's iron reserves. Thus, elevated serum hepsidin levels lead to diminished iron absorption and recycling, while lower levels of the hormone will cause greater iron absorption. Among the factors that affect the hepsidin response produced, we should highlight an individual's total iron levels, erythropoietic demands, state of hypoxia, dietary iron, inflammation and physical exercise. Given the important role played by iron regulatory mechanisms in physical performance, this report reviews our current understanding of the physiological response of hepsidin to different sports intensities and modalities. *Turk Jem 2014; 18: 84-91*

Key words: Iron, iron deficiency, inflammation, cytokines, aerobic exercise, hepsidin

Özet

Demirin vücuttaki bir çok fonksiyonu göz önüne alındığında, demir eksikliğinin herhangi bir derecesi spor performansını azaltacak bir çok ikincil etkiye neden olur. Atletlerde hazırlık dönemlerinde ve özellikle aerobik egzersiz ve direnç çalışması yapanlarda daha sık olmak üzere antrenman dönemlerinde düşük serum demir düzeyleri sıklıkla gözlenmektedir. Bazen demir düzeyleri sedanter hayat tarzı olan insanların seviyesine bile düşmektedir. Bu şekilde demir eksikliğinin her spor türü ile uğraşan atletlerde görülebilen en sık beslenme bozukluğu olduğu söylenebilir. Hepsidin ferroportin (Makrofaj ve duodenal enterosit bazolateral membranlarındaki ana demir taşıyıcısı) yıkımından sorumlu olan hepatositlerden salınan bir hormon olup vücut demir depolarının ana düzenleyicisi olduğu düşünülmektedir. Artmış hepsidin düzeyleri azalmış demir emilimi ve döngüsüne, düşük düzeyleri ise artmış demir emilimine neden olmaktadır. Hepsidin yanıtını değerlendirirken, bir bireyin total demir düzeyi, eritropoetin ihtiyacı, hipoksi durumu, diet demiri, inflamasyon ve fiziksel egzersiz göz önünde tutulmalıdır. Fiziksel performans sırasında demirin düzenleyici rolü göz önüne alınarak, bu bildiride hepsidin değişik spor türleri ve yoğunluklarına olan fizyolojik yanıtı hakkındaki güncel bilgi düzeyimiz gözden geçirilmiştir. *Turk Jem 2014; 18: 84-91*

Anahtar kelimeler: Demir, demir eksikliği, inflamasyon, sitokinler, aerobik egzersiz, hepsidin

Introduction

Iron deficiency is the most frequently observed nutritional disorder in athletes performing aerobic exercise and resistance training (1,2), especially affecting women (3,4) and adolescents (2,5). Estimates of the incidence of this disorder in men and women athletes run at 11% and 35%, respectively (6,7). Studies have shown that physical exercise modifies several variables related to iron metabolism (8), and mean haemoglobin and ferritin concentrations have been generally found to be lower in athletes than in untrained subjects (9,10). Significant variations have also been detected in such variables during the course of a sports season (11).

Among the roles played by iron, we should highlight oxygen transport and storage (forming part of haemoglobin and myoglobin), a role in the electron transport chain and DNA synthesis, and as a catalyst in free radical production from oxygen as a beneficial pro-oxidant function (12,13,14). The nature of these functions makes iron the most important factor related to human sports performance. Iron depletion has the following physiological effects: it reduces performance capacity and causes fatigue, exercise intolerance, altered immune function, short bursts of attention loss, irritability, loss of visual perception, impaired temperature regulation in cold, and a reduced capacity to adapt to high altitudes (15,16,17,18,19). Hence, to a large extent, good sports performance will depend on maintaining adequate iron

levels. A major goal for any athlete should therefore be to keep iron levels stable throughout an entire sports season (20).

Although attempts have been made to relate the intake of iron in athletes to their body iron stores, numerous studies have detected no such correlation (21,22,23), mainly because absorption mechanisms may be affected by different physical activities (21). Thus, as a negative iron regulator in the duodenum (24) and the main iron metabolism regulator (25,26,27), understanding hepcidin response associated with physical exercise may help explain anaemia and iron deficiency that so frequently affect athletes, especially during resistance training (28).

In this report, we update the current state of the topic by reviewing the literature addressing the hepcidin response associated with exercise.

Physiology of Hepcidin

Hepcidin is a peptide hormone produced mainly by hepatocytes (29,30) and was given this name because its mRNA was found to be highly expressed in the liver and it showed weak microbicidal activity *in vitro* (31). Up to three different isoforms of hepcidin exist each with four disulphide bridges: one consisting of 25 amino acids (hepcidin-25) (27) and two smaller isoforms hepcidin-22 and hepcidin-20, comprising 22 and 20 amino acids, respectively (32,33). Blood and urine concentrations of hepcidin-20 and hepcidin-22 are generally low, and higher levels are only observed in some physiological processes associated with elevated hepcidin-25 concentrations, such as acute myocardial infarction, sepsis, anaemia of chronic disease, metabolic syndrome and chronic renal disease (34,35,36,37,38,39).

Mechanism of Action

According to Ganz (40), the main function of hepcidin is related to immunity since iron is essential for the survival of invading pathogens (41). In effect, a positive relationship has been observed between bacterial virulence and iron availability (42). Hepcidin regulates iron by hindering its absorption (30,43,44,45). Several studies have shown that overexpression of hepcidin leads to iron deficiency both in humans (46) and mice (47), while hepcidin deficiency gives rise to excessive deposition of iron (44,48). Thus, it seems that both iron deficiency (49) and iron overload (48) are influenced to a large extent by an individual's hepcidin response. Hepcidin's mechanism of action is based on degradation of ferroportin (50,51,52). *In vitro*, it has been observed that when hepcidin binds to its receptor ferroportin, the two proteins are degraded via endocytosis in lysosomes (53,54). As it is well-known, ferroportin exports iron from macrophages and the basolateral membrane of duodenal enterocytes (55,56,57). Hence, hepcidin -besides reducing the absorption of dietary iron- also blocks the recycling of iron released by haemolysis (53,54). Further, although still unconfirmed, it appears that hepcidin also exerts some effects on other means of iron transport, such as the divalent metal ion transporter (Figure 1).

Hepcidin Determination

No reference standard exists to determine hepcidin levels and neither is there a valid calibrator for its measurement, hindering

the comparison of results emerging from the studies conducted to date on this peptide (27). Notwithstanding, there appears to be no great difference in the methods usually employed, and for both serum and urine samples, mass spectrometry and immunochemical techniques are equally effective (58). We should, however, bear in mind the following basic principles:

- Hepcidin determination does not accurately reflect serum levels although correlation exists between the two variables both in healthy individuals and subjects with an iron metabolism disorder (27). We should also consider that being highly susceptible to oxidation (59), hepcidin-25 cannot be distinguished from the other isoforms of hepcidin in urine samples (58). Further aspects that impair the interpretation of urine hepcidin measurements are: their dependence on the glomerular filtration rate, tubular reabsorption and on the hepcidin production capacity of tubular epithelial cells (60).

- Serum hepcidin concentrations are affected by circadian rhythm (61,62,63,64,65). No differences have been detected according to age in men (63), nor according to sex (34,61,62,64,65,66), with the exception of women of fertile age. This population subset shows lower hepcidin levels than both postmenopausal women and men of the same age range (63). Good correlation has been reported between serum hepcidin and ferritin levels (63).

Regulating Hepcidin Synthesis

Given hepcidin's main role as a regulator of body iron homeostasis (67), *in vitro* studies have suggested the presence of iron sensors in hepatocytes along with the transduction apparatus required to modulate hepcidin synthesis (68). The different mechanisms controlling hepcidin synthesis proposed so far are:

a) Iron Status

Circulating transferrin can be detected as a hepatocellular complex, including the transferrin receptor protein transferrin 1 (TfR1), transferrin receptor protein 2 (TfR2) and human hemochromatosis protein (HFE). It has been proposed that reduced abundance of TfR2 and HFE receptors will lead to a drop

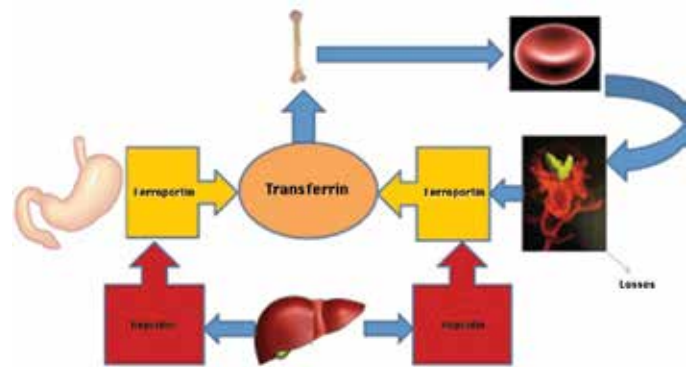


Figure 1. Mechanism of action of hepcidin. Hepcidin regulation of body iron. Hepcidin is released by hepatocytes into the circulation. Here it exerts its effects on serum iron levels by inhibiting iron absorption in the small intestine and iron release from macrophages, which recycle iron from senescent erythrocytes, by binding to its receptor ferroportin causing its degradation. Hepcidin only responds to iron when it is bound to its plasma transport protein transferrin.

in hepcidin levels via extracellular signals mediated by kinases (ERK/MAPK) and bone morphogenetic protein/Drosophila (BMP/SMAD) (27). This regulation of hepcidin synthesis serves to explain why serum hepcidin levels correlate negatively with transferrin levels (63,69). Accordingly, individuals with low total body iron levels show high hepcidin concentrations (70), while iron overload has been linked to low hepcidin concentrations (48).

In an effort to correlate serum ferritin concentrations with hepcidin response to iron supplementation, Borrione et al. (71) stratified a group of athletes according to their serum ferritin levels into: < 20 µg/dl; 20-30 µg/dl; 30-50 µg/dl or > 50 µg/dl. The results of this study revealed that subjects in the group featuring levels lower than 30 µg/dl of serum ferritin showed a diminished hepcidin response compared to the remaining groups. Thus, it seems that this level of 30 µg/dl of serum ferritin is the threshold below which iron reserves play an important role in regulating hepcidin synthesis.

b) Erythropoietic Demands

It has been proposed that stimulation of the bone marrow to upregulate erythropoiesis leads to increased iron absorption, driven by a reduced hepcidin response (40). In a cross-sectional study designed to obtain reference hepcidin values, it was observed that premenopausal women with higher erythropoietic demands due to menstrual blood loss showed significantly lower hepcidin levels than postmenopausal women (63). Similarly, other authors have reported that erythropoiesis inducing agents reduce hepcidin production (72,73).

The induction of erythropoietin has also been linked to a drop in hepcidin levels. Thus, administration of testosterone (enhancing erythropoiesis) to mice blocks hepcidin mRNA synthesis, which in turn, leads to increased iron import into red blood cells (74). Hence, the intake of erythropoiesis stimulating agents seems to reduce the hepcidin response (72,73). This is also true of anabolic substances (74), indicating a mechanism of action that improves aerobic performance in athletes consuming such performance-enhancing drugs.

c) Hypoxia

Hypoxia seems to be another factor able to modulate hepcidin, diminishing its serum levels (75,76). In exercise physiology, exposure to a high altitude has been the classic method of improving erythropoietin levels (EPO), as well as enhancing reticulocytes, haematocrit and haemoglobin (77,78,79). However, it is claimed that for a beneficial effect on the different haematological variables, a set of conditions has to be met including exposure to altitudes higher than 2000-2200 metres for at least 12 hours per day (80).

Serum testosterone levels are thought to be one of the factors responsible for the haematological modifications produced at high altitude. Several studies have observed elevated testosterone levels after a period at high altitude (81,82), probably attributable to its attenuating effects on altitude-related hyperventilation and respiratory alkalosis. Based on the finding that exogenous testosterone intake reduces hepcidin levels (83) by interfering with mRNA synthesis (74), it could be speculated that the hypoxia response of improved haematological indicators is due in part to a decrease in hepcidin synthesis as a consequence of the testosterone response to high altitude.

d) Dietary Iron

As may be predicted, iron ingested through diet may also affect the hepcidin response such that a transient increase in its urine levels is produced 4-8 hours after iron intake (34,62). In subjects given dietary iron supplements, Lin et al. (84) observed an increase in urine hepcidin clearance proportional to serum transferrin saturation.

e) Inflammation

Inflammation has been proposed as another factor that modulates the synthesis of hepcidin (85). Interleukin-6 (IL-6) is thought to play an important role in this process (86,87,88,89), along with BMP-2 (90).

In many chronic diseases, moderate anaemia is produced by inflammation. This anaemia of chronic disease is characterized by low serum iron concentrations (91). In patients on dialysis, alterations have been observed in levels of hepcidin, interleukin-6 and C-reactive protein (92). These same modifications have been observed in acute stage malaria (93), tuberculosis (94), inflammatory disorders (62,95), multiple myeloma (90), Hodgkin's disease (96), Castleman's disease (88), and in some patients with tumours (97).

Obesity is another disorder in which there is inflammation (98) and, a direct relationship has been noted between obesity and anaemia (99). In a group of morbidly obese subjects, it was observed that a reduction in the body mass index of 47.5 kg/m² to 39.5 kg/m² led to improved inflammation variables as well as hepcidin and haemoglobin levels and haematocrit (100).

f) Exercise

Exercise affects several of the factors mentioned above given; it promotes erythropoiesis (101) and induces an acute inflammatory response -similar to that observed in infection or inflammatory states (102). It has also been well established that free iron levels rise during post-exercise recovery (103,104,105,106,107,108,109). In the pursuit of ergogenic effects, athletes often take iron supplements (110,111,112). In addition, several studies have shown that exercise affects the hepcidin response increasing its synthesis or release (3,4,41,113,114,115,116,117,118,119,120).

Since inflammation is one of the factors that mediate the hepcidin response (34,86,88,89) and given that exercise directly affects cytokine levels (121), it has been proposed that the hepcidin response to exercise is a consequence of the inflammatory and haemolytic state associated with exercise (122,123). Although the regulation of hepcidin synthesis by means of modulating the levels of the variables related to inflammation has been confirmed in patients with chronic inflammatory processes (100), it has not been possible to reproduce this response following exercise (41,118). Recently, Sim et al. (119) observed differences in serum levels of interleukin-6 when running at 85% VO_{2max} versus the same intensity and volume of exercise performed on a cycle ergometer, though no changes in hepcidin levels were observed. These results suggest that other factors associated with physical exercise act as precursors of the hepcidin response to exercise, and it is unlikely that the inflammatory response induced by exercise plays a major contributing role.

Hepcidin Response to Exercise

Roecker et al. (117) were the first to report changes in urine hepcidin levels in women who had completed a marathon. The main findings of studies analysing the urine and/or serum hepcidin response to different exercise protocols are summarized in Tables 1 and 2. The main observation in all these investigations, except for the study by Troadec et al. (120), is that hepcidin levels rise significantly after exercise (3,4,41,113,114,115,116,117,118,119,120). Below we summarize the results obtained from different studies that have examined the effects of several exercise-related variables on the hepcidin response.

a) Exercise Intensity

- An intensity of exercise corresponding to 60% of the heart rate reserve or under does not seem to be associated with a significant increase in blood or urine hepcidin levels as a response to exercise (120).

- Exercise conducted at an intensity corresponding to 65% of VO_{2max} seems to be linked to an increased serum hepcidin concentration (113,119).

- Interval exercise performed at 85% VO_{2max} does not produce a greater response of serum hepcidin than that recorded for a continuous exercise protocol at 65% VO_{2max} (113).

- The same exercise volume (10 km) performed continuously at a relative intensity of 70% VO_{2max} induces a reduced response compared to the same volume performed at 90%-95% VO_{2max} in intervals (114). In this study, it was also observed that while at the lower intensity (70% VO_{2max}) hepcidin levels returned to normal after 12 hours, in the high intensity (90%-95% VO_{2max}) elevated levels of the hormone persisted for 24 hours.

In summary, exercise performed at relative intensities of 65% VO_{2max} or greater induces an increase in serum hepcidin levels (113,119), with maximum levels reached when intensities approach the individual's VO_{2max} (90%-95% VO_{2max}) (114).

b) Volume

In the only study that has examined the hepcidin response to training volume, it was observed that 120 minutes of exercise performed at a relative intensity of 65% VO_{2max} on a treadmill by physically active women triggered a significantly higher response

Table 1. Studies examining the acute hepcidin response to exercise in urine

Reference	Subjects	Experimental conditions (C)	Procedure	Main results
Roecker et al. (116)	14 female runners	Marathon	Pre and post-run, 24 h of recovery. Hepcidin	Hepcidin increased at 24 h of recovery*. 6/14 classified as non-responders.
Peeling et al. (113)	10 highly-trained triathletes	C1. 10 km at 70% VO_{2max} . C2. 10 km at 70% VO_{2max} + 10 x 1 km at 90% VO_{2max} (12 h recovery).	Pre, post-run and 3 h, 24 h of recovery. Hepcidin, IL-6, serum ferritin, SF, Hapt.	Hepcidin increased at 3 h of recovery*, IL-6 post-run* and 3 h of recovery*. Greatest increases produced in 10 x 1 at 90% VO_{2max} . No cumulative effect. Fes*, Fe* and Hapt* increased in post-run and 3 h of recovery (same as hepcidin and IL-6), but with a cumulative effect.
Peeling et al. (114)	10 trained male runners	C1. 10 km at 75-80% VO_{2max} on grass. C2. 10 km at 75-80% VO_{2max} on asphalt. C3. 10 x 1 km at 90-95% VO_{2max} on grass.	Pre, post-run, 3 h, 24 h of recovery. Hepcidin, IL-6; free Hb; Hapt.	Hepcidin increased at 3h of recovery*. No differences between groups. IL-6 and free Hb: increases post-run*, no differences between C1 and C2, but C3 showed an increase * versus C1 and C2. Hapt: post-run reduction*.
Peeling et al. (115)	11 trained male runners	60' Cr: 15' at 75-80% peak HR + 45' at 85%-90% peak HR.	Pre, post-run, 3 h, 6 h, 12 h, 24 h of recovery. Hepcidin, IL-6, PCR, serum ferritin, serum Fe.	Hepcidin: increases at 3 h*, 6 h*, 12 h* and at 24 h of recovery*. Peak at 3 h and 6 h. IL-6: increased at post-run* and 3 h of recovery*. Post peak. CRP: increased at 6 h*, 12 h* and 24 h of recovery*. Peak at 24 h of recovery. Ferritin and serum Fe: increased at post-run *. 3 h of recovery* lowest vs Pre.
Troadec et al. (119)	14 sedentary men	45' HRR cycle ergometer.	Pre, post-exercise, 0.5 h, 1 h, 2 h, 4 h, 6 h, 12 h, 24 h of recovery. Hepcidin, IL-6, serum ferritin, serum Fe, Hapt, CRP.	No changes detected in any variable examined.

Studies examining the acute hepcidin response to exercise in urine. Cr: continuous running, HR: heart rate, Fe: iron; h: hour, Hapt: haptoglobin, Hb: haemoglobin, IL-6: interleukin-6, CRP: C-reactive protein, HRR: heart rate reserve, *: statistical significance ($p < 0.05$)

to that observed by the same women performing the same intensity of exercise (65% VO_{2max}) for a duration of 60 minutes (113). These results suggest that training volume may also influence the hepcidin response to exercise.

c) Exercise Modality

To determine if the modality of aerobic resistance exercise could affect the hepcidin response, Sim et al. (119) compared this response in a group of triathletes performing two forms of exercise (cycle ergometry vs running) at two different relative intensities (65% VO_{2max} vs 85% VO_{2max}). Results revealed no significant differences according to exercise modality or intensity despite the detection of some differences in serum iron or interleukin-6 levels. In a study performed in runners, Peeling et al. (115) compared the hepcidin response to footstrike as a cause of haemolysis in a protocol carried out at a fixed intensity of 70% VO_{2max} on different running surfaces (grass vs. asphalt). Results indicated no significant differences in serum hepcidin levels between the two conditions.

Thus, the limited data available suggest that neither the exercise modality (running vs cycling) nor the running surface (grass vs. asphalt) significantly affect the hepcidin response to exercise.

Adaptations of the Hepcidin Response to Exercise

Although attractive, the working hypothesis suggesting a possible relationship between modified hepcidin levels and the onset of anaemia or iron deficiency in athletes (28) has not yet provided conclusive results, and few studies have addressed this hypothesis to date (3,4). In an initial study, Auersperger et al. (3) investigated the effects of long-term endurance exercise on hepcidin concentrations and inflammation and iron status parameters. These authors designed a training protocol consisting of two 3-week progressive overload periods, each followed by a week's recovery and runners were assigned to either a continuous or interval training group. As the study's main finding, significant correlation ($p < 0.05$) was detected between total body iron levels and C-reactive protein or serum hepcidin concentrations. Also, higher hepcidin levels were observed after the first/second overload period compared to baseline and a significant decrease was produced after the first/final recovery period. In a later study by the same authors (4), an increased incidence of modified iron metabolism-related variables was noted in response to a similar training program (from 5/10 to 7/10), including reduced

Table 2. Studies examining the acute hepcidin response to exercise in serum

Reference	Subjects	Experimental conditions (C)	Procedure	Main results
Troadec et al. (119)	14 sedentary men	45' HRR cycle ergometer	Pre, post-exercise, 0.5 h, 1 h, 2 h, 4 h, 6 h, 12 h, 24 h of recovery. Hepcidin, IL-6, serum ferritin, serum Fe, Hapt, CRP.	No change* in any variable examined.
Robson-Ansley et al. (41)	9 trained men	120' Cr at 60% VO_{2max} +5 km trial C1. Personalized hydration using a drink enriched with CH. C2. Control.	Pre, post-run, 24 h of recovery. Hepcidin, IL-6, serum Fe.	Hepcidin: elevated in C1 and C2 post-run*, no differences between groups. IL6: increase post-run* and C2* greater than C1.
Sim et al. (117)	11 highly-trained triathletes	90' Cr at 75% VO_{2max} . C1. Personalized hydration using a drink enriched with CH. C2. Control.	Pre, post-run, 3 h, 24 h of recovery. Hepcidin; IL-6; free Hb; serum ferritin, serum Fe; Trans.	Hepcidin and IL-6: post-run increase*, no differences between C1 and C2. Hapt: post decrease*. Free Hb: post decrease*. Serum ferritin, serum Fe and Trans: post-run increase* and decrease 24 h of recovery* vs Pre.
Newlin et al. (112)	12 physically active women	C1. 60' Cr at 65% VO_{2max} . C2. 120'Cr at 65% VO_{2max} .	Pre, post-run, 3 h, 6 h, 9 h, 24 h of recovery. Hepcidin; IL-6.	Hepcidin: increases post-run* and 3 h of recovery*. C2 greater* vs C1. IL-6: post-run increase*.
Sim et al. (118)	10 highly-trained triathletes	C1. 60' Cr at 60-65% peak VO_2 . C2. 60' cycle ergometer at 60%-65% peak VO_2 .	Pre, post-exercise, 3 h of recovery. Hepcidin; IL-6; serum ferritin; serum Fe.	Hepcidin: post-exercise increase* in all groups with no differences between groups. IL-6: post-exercise increase* in all groups and increase* in C4 versus C3. Serum ferritin: post increase* in C3 and C4 versus C1 and C2. Serum Fe: post increase* in C1, C3 and C4.

Studies examining the acute hepcidin response to exercise in serum. Cr: continuous running, HR: heart rate, Fe: iron; h: hour, Hapt: haptoglobin, Hb: haemoglobin, CH: carbohydrates, IL-6: interleukin-6, CRP: C-reactive protein, Trans: transferrin, HRR: heart rate reserve, *: statistical significance ($p < 0.05$)

serum hepcidin concentrations at the end of the study. This finding was attributed to a need to increase iron reabsorption to avoid compromising iron stores. However, it was observed that mean serum ferritin levels in the group of subjects without iron deficiency was 28 µg/dl. According to Borrione et al. (71), beyond 30 µg/dl of serum ferritin, the hepcidin response to dietary iron and probably also to exercise may be modified. Hence, the group initially classified as "without iron deficiency" could be considered equally deficient in physiological terms.

Among the limitations of studies addressing hepcidin adaptations to exercise, we should mention that if baseline samples are taken after 24 h of rest following exercise, hepcidin concentrations are likely to be normal considering that it is an acute phase hormone.

Conclusions

- The hepcidin response to exercise seems to be dependent on a minimum intensity of exercise (~65% VO_{2max}), with maximal levels of the hormone recorded in response to intensities approaching VO_{2max} (90%-95% VO_{2max}). Exercise duration and load also seem to affect the hepcidin response, and higher hepcidin concentrations have been detected for longer exercise durations. Finally, although only scarcely addressed, it does not seem that the modality of exercise is too important in the hepcidin response to exercise.

- The few investigations that have tried to examine possible adaptations of serum hepcidin levels to training have not provided sufficiently clear results to draw any firm conclusions. The reason for this is interference with other variables (diet, iron status) that have shown an effect on hepcidin regulation.

Future Studies

Given the key role played by hepcidin in whole body iron regulation and that exercise is able to modulate hepcidin synthesis, the response of hepcidin to exercise (modality, intensity, duration, frequency) needs to be further examined in detail. The information emerging from future studies should help modify training loads and/or establish dietary-nutritional regimens designed to maintain adequate body iron levels during training periods.

Conflicts of Interest

There are no conflicts of interest.

References

1. Beard J, Tobin B. Iron status and exercise. *Am J Clin Nutr* 2000;72:594-597.
2. Zoller H, Vogel W. Iron supplementation in athletes--first do no harm. *Nutrition* 2004;20:615-619.
3. Auersperger I, Knap B, Jerin A, Blagusc R, Lainscak M, Skitek M, Skof B. The effects of 8 weeks of endurance running on hepcidin concentrations, inflammatory parameters, and iron status in female runners. *Int J Sports Nutr & Exerc Metab* 2012;22:55-63.
4. Auersperger I, Skof B, Leskosek B, Knap B, Jerin A, Lainscak M. Exercise-induced changes in iron status and hepcidin response in female runners. *PLoS ONE* 2013;8:3.
5. Anttila R, Cook JD, Siimes MA. Body iron stores decrease in boys during pubertal development: the transferrin receptor-ferritin ratio as an indicator of iron status. *Pediatric Res* 1991;41:224-228.
6. Malczewska J, Raczynski G, Stupnicki R. Iron status in female endurance athletes and in non-athletes. *Int J Sport Nutr & Exerc Metab* 2000;10:260-276.
7. Malczewska J, Szczepanska B, Stupnicki R, Sendek W. The assessment of frequency of iron deficiency in athletes from the transferrin receptor-ferritin index. *Int J Sport Nutr & Exerc Metab* 2001;11:42-52.
8. Tsalis G, Nikolaidis MG, Mougios V. Effects of iron intake through food supplementation on iron status and performance of healthy adolescent swimmers during a training season. *Int J Sport Med* 2004;25:306-313.
9. Roecker L, Hinz K, Holla K, Gunga HC, Vogelgesang J, Kiesewetter H. Influence of endurance exercise (triathlon) on circulating transferrin receptors and other indicators of iron status in female athletes. *Clin Lab* 2002;48:307-312.
10. Shaskey DJ, Green GA. *Sports Haematology*. *Sport Med* 2000;29:27-38.
11. Reinke S, Taylor WR, Duda GM, Von Haehling S, Reinke P, Volk HD, Anker SD, Doehner W. Absolute and functional iron deficiency in professional athletes during training and recovery. *Int J Cardiol* 2012;156:186-191.
12. Beard JL. Iron biology in immune function, muscle, metabolism, and neuronal functioning. *J Nutr* 2001;131:568-579.
13. Brody T. *Nutritional Biochemistry*. San Diego: Academic Press; 1999.
14. Wilmore JH, Costill DL. *Fisiología del esfuerzo y del Deporte*. Barcelona: Editorial Paidotribo; 2004.
15. Deakin V. Iron depletion in athletes. In: Burke L, Deakin V. *Clinical Sports Nutrition*. Sydney: McGraw-Hill; 2006. P. 174-199.
16. Murray-Kolb LE, Beard JL. Iron treatment normalizes cognitive functioning in young women. *Am J Clin Nutr* 2007;85:778-787.
17. Schumacher YO, Chmid A, Granthwohl D, Bültermann D, Berg A. Hematological indices and iron status in athletes of various sports and performance. *Med & Sci Sports Exerc*. 2012;34:869-875.
18. Stray-Gundersen J, Hochstein A, deLemos D, Levine BD. Failure of red cell volume to increase to altitude exposure in iron deficient runners. *Med & Sci Sport & Exerc* 2002;24:90.
19. Williams MH. Dietary supplements and sports performance, minerals. *J Int Society Sport Nutr* 2005;2:43-49.
20. Ostojic, S. Ahmetovic, Z. Indicators of iron status in elite soccer players during the sport season. *Int J Lab Hematol* 2008;31:447-452.
21. Nuviala RJ, Castillo MC, Lapieza MG, Escanero JF. Iron Nutritional Status in Female Karatekas, Handball and Basketball Players, and Runners. *Physiol Behav* 1995;59:449-453.
22. Telford RD, Cunningham RB, Deakin V, Kerr DA. Iron status and diet in athletes. *Med & Sci Sport & Exerc* 1993;25:796-800.
23. Weight LM, Klein M, Noakes TD, Jacobs P. Sports anemia-a real or apparent phenomenon in endurance-trained athletes. *Int J Sport Med* 1992;13:344-347.
24. Gardenghi S, Marongiu MF, Ramos P, Guy E, Breda L, Chadburn A, Liu Y, Amariglio N, Rechavi G, Rachmilewitz EA, Breuer W, Cabantchik ZI, Wrighting DM, Andrews NC, de Sousa M, Giardina PJ, Grady RW, Rivella S. Ineffective erythropoiesis in β -thalassemia is characterized by increased absorption mediated by down-regulation of hepcidina and up-regulation of ferroportin. *Blood* 2007;109:5027-5035.
25. Barrios Y, Espinoza M, Barón MA. Pro-hepcidina, su relación con indicadores del metabolismo del hierro y de inflamación en pacientes hemodializados tratados o no con eritropoyetina recombinante. *Nutr Hosp* 2010;25:555-560.
26. Ganz T. Molecular control of iron transport. *J Am Soc Nephrol* 2007 18:394-400.
27. Kroot JJ, Tjalsma H, Fleming RE, Swinkels DW. Hepcidin in human iron disorders: diagnostic implications. *Clin Chem* 2011;57:1650-1669.
28. Liu YQ, Chang YZ, Zhao B, Wang HT, Duan XL. Does hepatic hepcidin play an important role in exercise-associated anemia in rats? *Int J Sport Nutr & Exerc Metab* 2011;21:19-26.
29. Krause A, Neitz S, Magert HJ, Schulz A, Forssmann WG, Schulz-Knappe P, Adermann K. LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. *FEBS Letters* 2000;480:147-150.
30. Pigeon C, Ilyin G, Courselaud B, Leroyer P, Turlin B, Brissot P, Loreal O. A new mouse liver-specific gene, encoding a protein homologous to human antimicrobial peptide hepcidin, is overexpressed during iron overload. *J Biol Chem* 2001;276:7811-7819.
31. Park CH, Valore EV, Waring AJ, Ganz T. Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *J Biol Chem* 2001;276:7806-7810.
32. Schranz M, Bakry R, Creus M, Bonn G, Vogel W, Zoller H. Activation and inactivation of the iron hormone hepcidin: biochemical characterization of prohepcidin cleavage and sequential degradation to N-terminally truncated hepcidin isoforms. *Blood Cell Mol Dis* 2009;43:169-179.
33. Valore EV, Ganz T. Posttranslational processing of hepcidin in human hepatocytes is mediated by the prohormone convertase furin. *Blood Cell Mol Dis* 2008;40:132-138.

34. Kemna EH, Tjalsma H, Podust VN, Swinkels DW. Mass spectrometry-based hepcidin measurements in serum and urine: analytical aspects and clinical implications. *Clin Chem* 2007;53:620-628.
35. Kroot JJ, Laarakkers CM, Geurts-Moespot AJ, Grebenchtchikov N, Pickkers P, van Ede AE, Peters HP, van Dongen-Lases E, Wetzels JF, Sweep FC, Tjalsma H, Swinkels DW. Immunochemical and mass-spectrometry based serum hepcidina assays for iron metabolism disorders. *Clin Chem* 2010;56:1570-1579.
36. Peters HP, Laarakkers CM, Swinkels DW, Wetzels JF. Serum hepcidin-25 levels in patients with chronic kidney disease are independent of glomerular filtration rate. *Nephrol Dial Transplant* 2010;25:848-853.
37. Suzuki H, Toba K, Kato K, Ozawa T, Tomosugi N, Higuchi M, Kusuyama T, Iso Y, Kobayashi N, Yokoyama S, Fukuda N, Saitoh H, Akazawa K, Aizawa Y. Serum hepcidin-20 is elevated during the acute phase of myocardial infarction. *Tokohu J Exp Med* 2009;218:93-98.
38. Tessitore N, Girelli D, Campostrini N, Bedogna V, Pietro Solero G, Castagna A, Melilli E, Mantovani W, De Matteis G, Olivieri O, Poli A, Lupo A. Hepcidin is not useful as a biomarker for iron needs in haemodialysis patients on maintenance erythropoiesis stimulating agents. *Nephrol Dial Transplant* 2010;25:3996-4002.
39. Tomosugi N, Kawabata H, Wakatabe R, Higuchi M, Yamaya H, Umehara H, Ishikawa I. Detection of serum hepcidin in renal failure and inflammation by using protein Chip System. *Blood* 2006;108:1381-1387.
40. Ganz T. Hepcidin, a key regulator of iron metabolism and mediator of anemia of inflammation. *Blood* 2003;102:783-788.
41. Robson-Ansley P, Walshe I, Ward D. The effect of carbohydrate ingestion on plasma interleukin-6, hepcidina and iron concentrations following prolonged exercise. *Cytokine* 2011;53:196-200.
42. Wright AC, Simpson LM, Oliver JD. Role of iron in the pathogenesis of *Vibrio vulnificus* infections. *Infect Immun* 1981;34:503-507.
43. Fleming RE, Sly WS. Hepcidin: a putative iron-regulatory hormone relevant to hereditary hemochromatosis and the anemia of chronic disease. *Proceedings Nat Acad Sci USA* 2001;98:8160-8162.
44. Nicolas G, Bennoun M, Devaux I, Beaumont C, Grandchamp B, Kahn A, Vaulont S. Lack of hepcidina gene expression and severe tissue iron overload in upstream stimulatory factor 2 (USF2) knockout mice. *Proceedings Nat Acad Sci USA* 2001;98:8780-8785.
45. Knutson MD. Iron-sensing proteins that regulate hepcidin and enteric iron absorption. *Ann Rev Nutr* 2010;30:149-171.
46. Rivera S, Nemeth E, Gabayan V, Lopez MA, Farshidi D, Ganz T. Synthetic hepcidin causes rapid dose-dependent hypoferrremia and is concentrated in ferroportin-containing organs. *Blood* 2005;106:2196-2199.
47. Nicolas G, Bennoun M, Porteu A, Mafivet S, Beaumont C, Grandchamp B, Sirito M, Sawadogo M, Kahn A, Vaulont S. Severe iron deficiency anemia in transgenic mice expressing liver hepcidina. *Proc Natl Acad Sci U S A* 2002;99:4596-4601.
48. Papanikolaou G, Tzilianos M, Christakis JI, Bogdanos D, Tsimirika K, MacFarlane J, Goldberg YP, Sakellaropoulos N, Ganz T, Nemeth E. Hepcidin in iron overload disorders. *Blood* 2005;105:4103-4105.
49. Ganz T. Hepcidin-a regulator of intestinal iron absorption and iron recycling by macrophages. *Best Pract Res Clin Haematol* 2005;18:171-182.
50. Preza GC, Pinon R, Ganz T, Nemeth E. Cellular catabolism of the iron-regulatory peptide hormone hepcidin. *PLoS One* 2013;8:58934.
51. Fleming MD. The regulation of hepcidina and its effect on systemic and cellular iron metabolism. *Hematol Am Soc Hematol Educ Program* 2008;151-158.
52. Lymoussaki A, Pignatti E, Montosi G, Garuti C, Haile DJ, Pietrangelo A. The role of iron responsive element in the control of ferroportin/IREG1/MPT1 gene expression. *J Hepatol* 2013;39:710-715.
53. Nemeth E, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, Ganz T. IL-6 mediates hypoferrremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest* 2004;113:1271-1276.
54. Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, Kaplan J. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* 2004;306:2090-2093.
55. Abboud S, Haile DJ. A novel mammalian iron regulated protein involved in intracellular iron metabolism. *J Biol Chem* 2000;275:19906-19912.
56. Donovan A, Brownlie A, Zhou Y, Shepard J, Pratt SJ, Moynihan J, Paw BH, Drejer A, Barut B, Zapata A, Law TC, Brugnara C, Lux SE, Pinkus GS, Pinkus JL, Kingsley PD, Palis J, Fleming MD, Andrews NC, Zon LI. Positional cloning of zebrafish ferroportin 1 identifies a conserved vertebrate iron exporter. *Nature* 2000;275:19906-19912.
57. McKie AT, Marciani P, Rolfs A, Brennan K, Wehr K, Barrow D, Miret S, Bomford A, Peters TJ, Farzaneh F, Hediger MA, Hentze MW, Simpson RJ. A novel duodenal iron-regulated transporter, IREG1, implicated in the basolateral transfer of iron to the circulation. *Mol Cell* 2000;5:299-309.
58. D'Angelo G. Role of hepcidin in the pathophysiology and diagnosis of anemia. *Blood Res* 2013;48:10-15.
59. Swinkels DW, Girelli D, Laarakkers C, Kroot J, Campostrini N, Kemna EH, Tjalsma H. Advances in quantitative hepcidin measurements by time-of-flight mass spectrometry. *PLoS One* 2008;3:2706.
60. Peters HP, Laarakkers CM, Pickkers P, Masereeuw R, Boerman OC, Eek A, Cornelissen EA, Swinkels DW, Wetzels JF. Tubular reabsorption and local production of urine hepcidin-25. *BMC Nephrol* 2013;14:70.
61. Busbridge M, Griffiths C, Ashby D, Gale D, Jayantha A, Sanwaiya A, Chapman RS. Development of a novel immunoassay for the iron regulatory peptide hepcidin. *Br J Biomed Sci* 2009;66:150-7.
62. Ganz T, Olbina G, Girelli D, Nemeth E, Westerman M. Immunoassay for human serum hepcidina. *Blood* 2008;112:4292-4297.
63. Galesloot TE, Vermeulen SH, Geurts-Moespot AJ, Klaver SM, Kroot JJ, van Tienoven D, Wetzels JF, Kiemeneij LA, Sweep FC, den Heijer M, Swinkels DW. Serum hepcidin: reference ranges and biochemical correlates in the general population. *Blood* 2011;117:218-225.
64. Grebenchtchikov N, Geurts-Moespot AJ, Kroot JJ, Den Heijer M, Tjalsma H, Swinkels DW, Sweep FG. High-sensitive radioimmunoassay for human serum hepcidina. *Br J Haematol* 2009;146:317-325.
65. Kroot JJ, Hendriks JC, Laarakkers CM, Klaver SM, Kemnsa EH, Tjalsma H, Swinkels DW. (Pre) analytical imprecision, between-subject variability, and daily variations in serum and urine hepcidina implications for clinical studies. *Anal Biochem* 2009;389:124-129.
66. Murphy AT, Witcher DR, Luan P, Wroblewski VJ. Quantitation of hepcidina from human and mouse serum using liquid chromatography tandem mass spectrometry. *Blood* 2007;110:1048-1054.
67. Collins JF, Wessling-Resnick M, Knutson MD. Hepcidin regulation of iron transport. *J Nutr* 2008;138:2284-2288.
68. Ganz T. Hepcidin and iron metabolism, 10 years later. *Blood* 2012;117:4425-4433.
69. Nemeth E, Valore EV, Territo M, Schiller G, Lichtenstein A, Ganz T. Hepcidin, a putative mediator of anemia of inflammation is a type II acute-phase protein. *Blood* 2003;101:2461-2463.
70. Nicolas G, Chauvet C, Viatte L, Danan JL, Bigard X, Devaux I, Beaumont C, Kahn A, Vaulont S. The gene encoding the iron regulatory peptide hepcidina is regulated by anemia, hypoxia, and inflammation. *J Clin Invest* 2002;110:1037-1044.
71. Borriore P, Spaccamiglio A, Rizzo M, Termine A, Chierito E, Campostrini N, Quaranta F, Di Gianfrancesco A, Pigozzi F. Urinary hepcidin identifies a serum ferritin cut-off for iron supplementation in young athletes: a pilot study. *J Biol Regul Homeost agent* 2001;25:427-434.
72. Ashby DR, Gale DP, Busbridge M, Murphy KG, Duncan ND, Cairns TD, Taube DH, Bloom SR, Tam FW, Chapman RS, Maxwell PH, Choi P. Plasma hepcidin levels are elevated but responsive to erythropoietin therapy in renal disease. *Kidney Int* 2009;75:976-981.
73. Banfi G, Lombardi G, Colombini A, Lippi G. A world apart: Inaccuracies of laboratory methodologies in antidoping testing. *Clin Chem Acta* 2010;411:1003-1008.
74. Guo W, Bachman E, Li M, Roy CN, Bluszajin J, Wong S, Chan SY, Serra C, Jasuja R, Trivison TG, Muckenthaler MU, Nemeth E, Bhasin S. Testosterone administration inhibits hepcidina transcription and is associated with increased iron incorporation into red blood cells. *Aging Cell* 2013;12:280-291.
75. Nemeth E, Ganz T. Regulation of iron metabolism by hepcidin. *Ann Rev Nutr* 2006;26:323-342.
76. Darshan D, Anderson GJ. Interacting signals in the control of hepcidin expression. *Biomaterials* 2009;22:77-87.
77. Friedmann B, Jost J, Rating T, Weller E, Werle E, Eckardt KU, Bärtisch P, Mairbörl H. Effects of iron supplementation on total body hemoglobin during endurance training at moderate altitude. *Int J Sport Med* 1999;20:78-85.
78. Saunders PU, Telford RD, Pyne DB, Hahn AG, Gore CJ. Improved running economy and increased hemoglobin mass in elite runners after extended moderate altitude exposure. *J Sci Med Sport* 2009;12:62-72.
79. Wehrli JP, Zuest P, Hallén J, Marti B. Live high-train low for 24 days increase hemoglobin mass and red cell volume in elite endurance athletes. *J Appl Physiol* 2006;100:1938-1945.
80. Rusko HK, Tikkanen HO, Peltonen JE. Altitude and resistance training. *J Sport Sci* 2004;22:928-944.
81. Gonzales GF, Rodríguez L, Valera J, Sandoval E, García-Hjarles M. Prevention of high altitude-induced testicular disturbances by previous treatment with cyproheptadine in male rats. *Arch Androl* 1990;24:201-205.

82. Gonzales GF, Chung FA, Miranda S, Valdez LB, Zaobornyj T, Bustamante J, Boveris A. Heart mitochondrial nitric oxide synthase in rats at high altitude. *Am J Physiol Heart Circ Physiol*. 2005;288:2568-2573.
83. Bachman E, Feng R, Trivison T, Li M, Olbina G, Ostland V, Ulloor J, Zhang A, Basaria S, Ganz T, Westerman M, Bhasin S. Testosterone suppresses hepcidin in men: a potential mechanism for testosterone-induced erythrocytosis. *J Clin Endocrinol Metab* 2010;95:4743-4737.
84. Lin L, Valore EV, Nemeth E, Goodnough JB, Gabayan V, Ganz T. Iron transferrin regulates hepcidin synthesis in primary hepatocyte culture through hemojuvelin and BMP2/4. *Blood* 2007;110:2182-2189.
85. Hoppe M, Lonnerdal B, Hossain B, Olsson S, Nilson F, Lundberg PA, Rödger S, Hulthen L. Hepcidin, interleukin-6 and hematological markers in males before and after heart surgery. *J Nutr Biochem* 2009;20:11-16.
86. Hashizume M, Uchiyama Y, Horai N, Tomosugi N, Mihara M. Tocilizumab anti-interleukin-6 receptor antibody, improved anemia in monkey arthritis by suppressing IL-6-induced hepcidin production. *Rheumatol Int* 2010;30:917-923.
87. Kemna E, Pickers P, Nemeth E, Van der Hoeven H, Swinkels D. Time-course analysis of hepcidin, serum iron, and plasma cytokine levels in humans injected with LPS. *Blood* 2005;106:864-866.
88. Song SN, Tomosugi N, Kawabata H, Ishikawa T, Nishikawa T, Yoshizaki K. Downregulation of hepcidin resulting from long-term treatment with an IL-6 receptor antibody (focilizumab) improves anemia of inflammation in multicentric Castleman disease. *Blood* 2010;116:3627-3634.
89. Wrighting DM, Andrews NC. Interleukin-6 induces hepcidin expression through STAT3. *Blood* 2006;108:3204-3209.
90. Maes K, Nemeth E, Roodman GD, Huston A, Esteve F, Freytes C, Callander N, Katodritou E, Tussing-Humphreys L, Rivera S, Vanderkerken K, Lichtenstein A, Ganz T. In anemia of multiple myeloma, hepcidin is induced by increased bone morphogenetic protein 2. *Blood* 2010;116:3635-3644.
91. Corwin HL, Hantz SB. Anemia of the critically ill: "acute" anemia of chronic disease. *Critical Care Med* 2010;28:3098-3099.
92. Khalil A, Goodhand JR, Wahed M, Yeung J, Ali FR, Rampton DS. Efficacy and tolerability of intravenous iron dextran and oral iron in inflammatory bowel disease: a case-matched study in clinical practice. *Eur J Gastroenterol Hepatol* 2011;23:1029-1035.
93. de Mast Q, Syafruddin D, Keijimel S, Reiekerink TO, Deky O, Asih PB, Swinkels DW, van der Ven AJ. Increased serum hepcidin and alterations in blood iron parameters associated with asymptomatic *P. falciparum* and *P. vivax* malaria. *Haematologica* 2010;95:1068-1074.
94. Rattledge C. Iron, mycobacteria and tuberculosis. *Tuberculosis (edinb)* 2004;84:110-130.
95. Butterfield AM, Luan P, Wichter DR, Manetta J, Murphy AT, Wroblewski VJ, Konrad RJ. A dual-monoclonal sandwich ELISA specific for hepcidin-25. *Clin Chem* 2010;56:1725-1732.
96. Hohaus S, Massini G, Giachella M, Vannata B, Bozzoli V, Cuccaro A, D'Alo' F, Larocca LM, Raymakers RA, Swinkels DW, Voso MT, Leone G. Anemia in Hodgkin's lymphoma: the role of interleukin-6 and hepcidin. *J Clin Oncol* 2010;28:2538-2543.
97. Sasu BJ, Cooke KS, Arvedson TL, Plewa C, Ellison AR, Sheng J, Winters A, Juan T, Li H, Begley CG, Molineux G. Antihepcidin antibody treatment modulates iron metabolism and is effective in a mouse model of inflammation-induced anemia. *Blood* 2010;115:3616-3624.
98. Pietiläinen KH, Kannisto K, Korshennikova E, Rissanen A, Kaprio J, Ehrenborg E, Hamsten A, Yki-Järvinen H. Acquired obesity increases CD68 and tumor necrosis factor- α and decrease adiponectin gene expression in adipose tissue: a study in monozygotic twins. *J Clin Endocrinol Metab* 2006;91:2776-2781.
99. Seltzer CC, Mayer J. Serum iron and iron-binding capacity in adolescents. *Am J Clin Nutr* 1963;1:354-361.
100. Tussing-Humphreys LM, Nemeth E, Fantuzzi G, Freels S, Holterman AX, Galvani C, Ayloo S, Vitello J, Braunschweig C. Decreased serum hepcidin and improved functional iron status 6 months after restrictive bariatric surgery. *Obesity (Silver Spring)* 2010;18 :2010-2016.
101. Weight LM, Byrne MJ, Jacobs P. Hemolytic effect of exercise. *Clin Sci (Lond)* 1991;81:147-152.
102. Pedersen BK, Hoffmann-Görtz L. Exercise and the immune system: regulation, integration and adaptation. *Physiol Rev* 2000;80:1055-1081.
103. Egan LM, Watts PB, Silta BC. Changes in serum haptoglobin as an acute response to a marathon road race. *J Sports Sci* 1987;5:55-60.
104. Pizza FX, Flynn MG, Boone JB, Rodriguez-Zayas JR, Andres FF. Serum haptoglobin and ferritin during a competitive running and swimming season. *Int J Sport Med* 1997;18:233-237.
105. Remacha AF, Ordoñez J, García-Die F, Estruch A, Gimferrer E. Hematologic changes induced by exertion during a long-distance race. *Sangre (Barc)* 1993;38:443-447.
106. Resina A, Gatteschi L, Giamberardino MA, Imreh F, Rubenni MG, Vecchiet L. Hematological comparison of iron status in trained top-level soccer players and control subjects. *Int J Sport Med* 1991;12:453-456.
107. Schobersberger W, Tschann M, Hasibeder W, Steidl M, Herold M, Nachbauer W, Koller A. Consequences of 6 weeks of strength training on red cells = 2 transport and iron status. *Eur J Appl Physiol Occup Physiol* 1990;60:163-168.
108. Selby GB, Eichner ER. Endurance swimming, intravascular hemolysis, anemia, and iron depletion: New perspective on athlete's anemia. *Am J Med* 1986;81:791-794.
109. Telford RD, Sly GJ, Hahn AG, Cunningham RB, Bryant C, Smith JA. Footstrike is the major cause of hemolysis during running. *J Appl Physiol* 2003;94:38-42.
110. Chicharro JL, Hoyos J, Gómez-Gallego F, Villa JG, Bandrés F, Celaya P, Jiménez F, Alonso JM, Córdova A, Lucia A. Mutations in the hereditary haemochromatosis gene HFE in professional endurance athletes. *Brit J Sport Med* 2004;38:418-421.
111. Mettler S, Zimmermann MB. Iron excess in recreational marathon runners. *Eur J Clin Nutr* 2010;64:490-494.
112. Nakanishi M, Ishii K, Watanabe A, Sugiura K, Kajiwara Y, Kobayashi K. Supplement intake in female university long-distance runners. *Japanese J Physiol Fitness Sport Med* 2003;52:631-637.
113. Newlin MK, Williams S, McNamara T, Tjalsma H, Swinkels DW, Haymes EM. The effects of acute exercise bouts on hepcidin in women. *Int J Sport Nutr Exercise Metab* 2012;22:79-88.
114. Peeling P, Dawson B, Goodman C, Landers G, Wiegerenck ET, Swinkels DW, Trinder D. Cumulative effects of consecutive running sessions on hemolysis, inflammation and hepcidin activity. *Eur J Appl Physiol* 2009;106:51-59.
115. Peeling P, Dawson W, Goodman C, Landers G, Wiegerenck ET, Swinkels DW, Trinder D. Training Surface and Intensity: Inflammation, Hemolysis, and Hepcidin Expression. *Med Sci Sports Exerc* 2009;41:1138-1145.
116. Peeling P, Dawson B, Goodman C, Landers G, Wiegerenck ET, Swinkels DW, Trinder D. Effects of exercise on hepcidin response and iron metabolism during recovery. *Int J Sport Nutr Exerc Metab* 2009;19:583-597.
117. Roecker L, Meier-Buttermilch R, Bretschel L, Nemeth E, Ganz T. Iron-regulatory protein hepcidin is increased in female athletes after a marathon. *Eur J Appl Physiol* 2005;95:569-571.
118. Sim M, Dawson B, Landers G, Wiegerenck ET, Swinkels DW, Townsend MA, Trinder D, Peeling P. The effects of carbohydrate ingestion during endurance running on post-exercise inflammation and hepcidin levels. *Eur J Appl Physiol* 2012;12:1289-1298.
119. Sim M, Dawson B, Landers G, Swinkels DW, Tjasma H, Trinder D, Peeling P. Effect of exercise modality and intensity on postexercise interleukin-6 and hepcidin levels. *Int J Sport Nutr Exerc Metab* 2013;23:178-186.
120. Troadec MB, Lainé F, Daniel V, Rochongar P, Ropert M, Cabillic F, Perrin M, Morcet J, Loréal O, Olbina G, Westerman M, Nemeth E, Ganz T, Brissot P. Daily regulation of serum and urinary hepcidin is not influenced by submaximal cycling exercise in humans with normal iron metabolism. *Eur J Appl Physiol* 2009;106:435-443.
121. Fisher CP. Interleukin-6 in acute exercise and training: What is the biological relevance? *Exerc Imm Rev* 2006;12:6-33.
122. Peeling P. Exercise as a mediator of hepcidin activity in athletes. *Eur J Appl Physiol* 2010;110:877-883.
123. Peeling P, Dawson B, Goodman C, Landers G, Trinder D. Athletic induced iron deficiency: new insights into the role of inflammation, cytokines and hormones. *Eur J Appl Physiol* 2008;103:381-391.