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Article

Azulene Functionalization by Iron-Mediated Addition to a Cyclohexadiene Scaffold

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■ INTRODUCTION

conjugated ketone and a fused tetracycle.

Azulene is a bicyclic nonalternant aromatic hydrocarbon, isomeric with naphthalene. The π -electrons of azulene are polarized toward the five-membered ring, resulting in a relatively large inherent dipole moment and a deep blue color.¹ Their unusual electronic properties make azulenes interesting as photocatalysts² and colorimetric indicators³ and for optoelectronics such as solar cells,⁴ photoswitches,⁵ and organic electronics.⁶ Azulene derivatives have also found use in medicine in the form of antiulcer,⁷ antidiabetic,⁸ and anticancer⁹ agents. The ability to functionalize the azulene scaffold is thus of great interest.

Azulenes are potent nucleophiles for electrophilic aromatic substitution (S_EAr), with the 1- and 3-positions being the most reactive toward electrophiles.¹⁰ Reported methods for derivatizing azulenes include (a) the Michael addition,¹¹ (b) Friedel–Crafts^{11a,12} and Vilsmeier–Haack reactions,^{11a,13} (c) azulene addition to heteroaromatic triflates,¹⁴ and (d) the formation of diazo compounds (Scheme 1).¹⁵ Azulenes are also susceptible to (e) electrophilic halogenations.^{10e,16} In addition, cross-coupling reactions have been reported.¹⁷ While these latter reactions are attractive for coupling the azulene to sp²- and sp-carbons, they generally rely on the use of precious metal catalysts and require prefunctionalization of the azulene skeleton.

Another class of electrophile that could potentially react with nucleophilic azulenes are cationic η^5 -iron carbonyl dienyl complexes (Scheme 2). Such complexes are capable of reacting with a wide range of nucleophiles to form new carbon–carbon or carbon–heteroatom bonds.¹⁸ Nucleophilic addition proceeds in a highly selective manner at the opposite face to the iron carbonyl moiety and at one of the ends of the conjugated system.^{18b,19} Substituents on the cation can act to direct the nucleophilic addition to either termini of the dienyl system;

Scheme 1. Electrophilic Aromatic Substitution Reactions for Functionalizing Azulenes







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electron-withdrawing groups will generally favor addition in the meta position, while electron-donating groups will often favor the para or ipso addition.²⁰ The cationic complexes can be formed via hydride abstraction (Scheme 2, path a) or by acid treatment of a complex possessing a leaving group (Scheme 2, path b). Once formed, the complex can be isolated as a shelf-stable fine powder.⁴²

Aromatic nucleophiles that can react with cationic iron carbonyl dienyl complexes include furan,²¹ indole,^{21,22} aniline,²³ and oxygenated arenes.²⁴ The scope was recently increased by us to include the selective C- or O-addition of phenolic nucleophiles.²⁵ Although azulenes, with a nucleophilicity similar to that of indoles,^{10d} can be expected to be efficient nucleophiles, no previous report of their use in this context has been reported. Herein, we present a convenient iron-mediated electrophilic C-H functionalization of azulenes (Scheme 1). The products of this process are valuable in two contexts. First, they are stimuli-responsive substances with multimodal outputs (colorimetric, fluorescence, and infrared spectroscopic). Second, they are versatile substrates for further synthetic transformations such as cyclizations. The cyclohexadiene also constitutes a proaromatic substituent, which may be readily converted to a phenyl ring, giving a formal product of azulene electrophilic arylation, for which there is no direct S_EAr equivalent.

RESULTS AND DISCUSSION

To investigate if azulenes could react with cationic iron carbonyl dienyl complexes, we selected guaiazulene 1 as the nucleophile, due to its low cost and availability from natural sources. As the electrophile, we selected iron complex 2, which we have used in earlier studies.²⁵ Optimization of the reaction conditions for this addition was then performed (Table 1).

Table 1. Optimization of the Addition of Guaiazulene (1) toCationic Complex 2^a



entry	solvent	base	yield ^b 3 (%)
1	EtOH		41
2	H ₂ O		72
3	acetone/ H_2O (3:1)		85
4	MeOAc	Et ₃ N	75
5	acetone	Et ₃ N	73
6	acetone	NaHCO ₃	97

^{*a*}Product **3** is racemic; relative stereochemistry is shown. ^{*b*}Yields are determined by quantitative ¹H NMR using *p*-xylene as the internal standard.

In our previous studies involving phenolic nucleophiles, we performed the reaction in ethanol in the absence of base at an ambient temperature.²⁵ Similar conditions here afforded the desired addition product **3**, albeit in a modest yield (Table 1, entry 1). Using water as the solvent afforded a heterogeneous reaction mixture where both guaiazulene and iron complex **2** were insoluble. Nevertheless, the yield was increased to 72%

(entry 2). To investigate if solubility issues might play a role in limiting the yield, the reaction was performed in a 3:1 mixture of acetone and water, resulting in an enhanced yield of 85% (entry 3). Adding an amine base such as triethylamine (Et_3N) was not beneficial, neither for methyl acetate as the solvent (entry 4) nor for acetone (entry 5). A near-quantitative yield could, however, be attained using acetone with sodium bicarbonate as the base (entry 6). This last protocol thus provides a set of green and benign reaction conditions for the addition reaction.

Azulene itself possesses two nucleophilic carbons, and a mixture of mono- and disubstituted products is thus to be expected. To see if azulene could be selectively functionalized to form either one of these products, the reaction was first performed using a slight excess of complex 2 relative to azulene. This afforded an equimolar mixture of the diastereomeric disubstituted azulenes 4 and 4' in an 81% yield (Scheme 3). Using a large excess of azulene instead, a mixture of mono- and disubstituted products was formed, where the monosubstituted product 5 could be isolated in a yield of 48%.







^{*a*}Products are racemic with the relative stereochemistry shown. ^{*b*}Isolated yields. ^{*c*}Excess of 2 (2.2 equiv), 4 equiv NaHCO₃. ^{*d*}Excess of azulene (5 equiv), 2 equiv NaHCO₃.

The scope of the transformation was then explored further. In addition to the activated complex (2), electronically neutral 6 and deactivated complex 7 were also selected for evaluation as electrophiles in the reaction (Figure 1).

Azulenes with extended π -systems are of interest for optoelectronic applications, as the properties of azulene can be tuned by extension of the conjugated system.^{4c} Routes to



Figure 1. Cationic iron carbonyl dienyl complexes 2, 6, and 7, selected as electrophilic coupling partners.

functionalizing these types of motifs could thus be valuable. A selection of 1-substituted azulene nucleophiles (8-11, Figure 2), as well as a 6-substituted azulene (12), was prepared and explored in reactions with iron complexes 6 and 7.



Figure 2. Scope of azulene nucleophiles with extended π -systems.

Azulene is significantly more expensive than guaiazulene. As nucleophiles 8-11 were prepared from azulene itself, they were used as the limiting reagent in the ensuing reactions. The results of their reaction with electrophiles 6 or 7 are shown in Scheme 4. Treating 1-phenylazulene 8 with 1.1 equiv of iron complex 6 resulted in the formation of 13 in an excellent yield of 95%. The less activated iron complex 7 was equally effective with the same nucleophile, resulting in 94% of adduct 14. Azulene 9, with a thienyl functionality, reacted with 6 to form compound 15 in a 73% yield. No other coupling products were detected, indicating that the thiophene unit does not compete with azulene as a nucleophile in this case. Azulene 10, with a pendant alkyne group, and formylated azulene 11 afforded adducts 16 and 17 in 70 and 50% yields, respectively. These products contain functional handles, which can be utilized for further derivatization. As in the case of azulene, 6-phenylazulene 12 possesses two nucleophilic sites and could be doubly alkylated using an excess of the cationic iron complex 6, giving rise to an equimolar mixture of diastereomers 18 and 18' in an 83% yield.

Naturally occurring guaiazulene (1) is an FDA-approved additive in cosmetics.²⁶ Being a large-scale commercial product, it is inexpensive and readily available and therefore of special interest for synthetic applications. In addition to guaiazulene itself, seven guaiazulene derivatives (19-25) with different substitution patterns on the five-membered ring were prepared to be used as nucleophiles in the addition reaction. Substituents included both electron-withdrawing and electron-donating groups, along with some potentially useful synthetic handles in the form of a boronic ester or a halogen (Figure 3).

Addition of the guaiazulene nucleophiles 1 and 19–25 was performed using the optimized conditions shown in Table 1. The results are summarized in Scheme 5. Addition of guaiazulene itself to electrophilic iron complex 2 proceeded to give addition product 3 in an excellent yield of 97%. Complexes 6 and 7 were less reactive, but upon extending the reaction time from 4 to 16 h, alkylated azulenes 26 and 27 were obtained in yields of 98% and 93%, respectively. A deactivating effect was seen for the more electron-deficient 1formyl-substituted guaiazulene derivative 19, where compound 28 was produced in a 56% yield. The coupling reaction proved to be sensitive toward substituents in the 2-position of the guaiazulene scaffold. Guaiazulene derivative 20, with a boronic



^{*a*}Products are racemic with the relative stereochemistry shown. ^{*b*}Isolated yields. ^{*c*}16 h reaction time. ^{*d*}2.2 equiv electrophile, 4 equiv NaHCO₃.

ester in the 2-position, reacted with iron complex 2 to produce 29 in a 43% yield. 2-Iodoguaiazulene (21) reacted sluggishly, but a 63% yield of 30 could be attained using a reaction time of 16 h and an excess of cation 2. The bromo-substituted guaiazulene 22 showed a higher reactivity, resulting in 79% of product 31 under the standard reaction conditions, with a 4 h reaction time. This suggests that steric factors may influence the reaction outcome in the case of 2-substituted azulenes. No reaction occurred with the tolyl-substituted derivative 23. For the coupling with 2-aminoguaiazulene (24), competing Naddition is a possibility.²³ The reaction was unsuccessful, however; while all starting material was consumed, neither Cor N-addition product could be isolated. To investigate if this reaction outcome was linked to instability introduced by the amino group, compound 24 was acetylated to amide 25 and the latter was subsequently used in a coupling reaction. While ¹H NMR analysis of the crude product looked promising, no product could be isolated in this case.



Figure 3. Guaiazulene (1) and guaiazulene derivatives evaluated as nucleophilic coupling partners in the addition to iron complexes 2, 6, and 7.

In our previous research on phenolic nucleophiles, we developed a shorter route for the coupling reaction starting from neutral iron cyclohexadiene complexes.²⁵ This method foregoes the formation and isolation of the cationic iron carbonyl complex by forming the electrophile in situ from a neutral precursor using a catalytic amount of acid. We applied this strategy here and found that guaiazulene could be alkylated directly from the neutral iron complex 32 (precursor to cationic complex 2) in the presence of tetrafluoroboric acid (Scheme 6). When a catalytic amount of acid was used, only a small amount of product was formed. Upon increasing the amount of tetrafluoroboric acid to 1.1 equiv, however, the desired addition product 3 was obtained in an 86% yield. Using an excess of acid (2 equiv) lowered the yield to 45%, which may be due to protonation of guaiazulene, reducing its nucleophilicity. The 86% yield obtained under the optimal conditions is higher than the combined 76% yield obtained when preforming the complex,²⁵ followed by the addition of guaiazulene, opening up for a shorter synthetic path. As the isolation of the cationic complex includes precipitation using large amounts of solvent, this variant of the reaction can significantly reduce the amount of waste created in the synthesis of products such as 3.

Iron carbonyl complexes are of interest as bioprobes,²⁷ indicating that the azulenes formed in this study could be useful in their own right. In addition to a strong and distinctive absorption in the IR region from the iron carbonyl moiety, the azulene fragment introduces both a fluorophore and a clearly visible colorimetric indicator, thus potentially allowing multimodal chemical sensing. Accordingly, we investigated the UV– vis, fluorescence, and IR spectroscopic properties of a representative selection of the novel compounds reported here.

Azulene derivatives are known sometimes to exhibit halochromism, i.e., to undergo changes in color upon protonation (at the azulene-1- or 3-position).^{28,29} Compounds 4/4', 14, 26, 33, and 34 were assessed for their colorimetric responses to trifluoroacetic acid (TFA). Exposure to excess TFA (1000 equiv) resulted in each case in a color change discernible to the naked eye (Figure S67, see the Supporting Information (SI)), which became more pronounced when TFA was used as cosolvent (Figure 4). UV–vis absorption spectra of the neutral and protonated azulenes were also

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^{*a*}Products are racemic with the relative stereochemistry shown. ^{*b*}Isolated yields. ^{*c*}4 h reaction time. ^{*d*}16 h reaction time. ^{*e*}Excess of 2 (1.1 equiv).

acquired (Figure S68). The relationship between azulene substitution pattern and halochromism has been studied and a trend can be determined. It has been shown that for some strongly halochromic azulenes with a particular substituent at the 2- or 6-position, moving the substituent to the 1- or 3-position may significantly attenuate the halochromic response.^{28b-g} This may be due to the change in connectivity resulting in a different preferred site of protonation. The tricarbonyliron(diene)-substituted azulenes 4/4', 14, and 26 exhibit pronounced halochromism regardless of the fact the substituents are at 1- and 3-positions of the azulene core remains the

Scheme 6. Formation of Addition Product Directly from Neutral Hydroxylated Iron Carbonyl Diene Complex 32^a





Figure 4. Azulenes (0.5 mM) in CHCl₃ (left) and TFA/CHCl₃ 1:9 v/v (right). Photo taken 30 min after sample preparation.

preferred site of protonation, as opposed to the tricarbonyliron(diene) motif.

It is known that for many azulenes, protonation can induce a significant fluorescence turn-on response,³⁰ whereas fluorescence properties of tricarbonyliron(diene) complexes have been studied only rarely.^{27a} Exposure to UV irradiation is an established method of cleaving Fe–C bonds in tricarbonyliron(diene) complexes (see below), but the reaction time required for this process is >3 orders of magnitude greater than the acquisition time for a fluorescence spectrum (days vs seconds). On the basis of this semiquantitative consideration, we reasoned that fluorescence spectra could nevertheless be acquired. Compounds 4/4', 14, 26, 33, and 34 were all found to exhibit significant fluorescence enhancement upon addition of TFA, with 4/4' showing the greatest turn-on response ($\lambda_{ex} = 266 \text{ nm}$, $\lambda_{em} = 336 \text{ nm}$; Figure 5), and our current findings show that the tricarbonyliron(diene) motif

does not quench the fluorescence of the pendent azulenium fluorophore. A titration of azulene 4/4' with TFA was also performed, monitored by UV-vis and fluorescence spectroscopy (Figures S69 and S70). A weak second emission maximum was observed at higher acid concentrations, implying a possible further reaction under these conditions.

Iron carbonyl diene complexes exhibit strong vibrational absorption bands in the $2100-1800 \text{ cm}^{-1}$ region, a window where most biological media are transparent.^{27a,31} This makes them interesting in applications such as bioimaging using mid-IR and Raman spectroscopy.^{31b} Iron carbonyl diene complexes substituted with fluorescent coumarin moieties have been suggested as IR-fluorescent probes,^{27a} and we therefore envision that the azulene-functionalized iron carbonyl complexes could be useful for similar applications. The IR-absorption spectra of the azulene addition products were measured using attenuated total reflection (ATR)-Fourier transform infrared (FTIR) in the neat state. As expected, characteristic strong peaks in the 2100–1900 cm⁻¹ region were observed for all products possessing the iron carbonyl moiety.

To increase the synthetic utility of the formed products, we investigated the oxidative removal of the iron carbonyl moiety. Several methods have been developed for this transformation, where strong oxidative conditions are generally used, including the use of hydrogen peroxide in aqueous sodium hydroxide,³ cerium ammonium nitrate,³³ trimethylamine N-oxide,³⁴ or cupric chloride.³⁵ Initial application of these methods, however, showed that these conditions were too harsh for these compounds, resulting in poor yields, partial degradation of the oxidatively sensitive azulenes, and in some cases partial aromatization of the formed cyclohexadiene. Successful demetallation of the addition products could, however, be achieved by applying mild, photolytic decomplexation conditions, using a modification of a procedure reported by Knölker.³⁶ Irradiation of an acetonitrile solution of 13 with a low-energy UV light (370 nm, 15 W) for 72 h, in the presence of air, yielded the free diene 33 in a 78% yield (Scheme 7, entry 1). An advantage of the current procedure is that the demetallation can be carried out without the need for specialized equipment. However, it is likely that the reaction time can be significantly shortened with a more powerful light source. Notably, this photolytic decomplexation is more tolerant toward oxidatively labile groups than traditional



Figure 5. Response of azulene 4/4' to the addition of TFA in CHCl₃. (Left) UV–vis spectra of 4/4' (50 μ M) before and after the addition of TFA in CHCl₃ (inset: visible region, 500 μ M 4/4'). (Right) Fluorescence response of 4/4' (5 μ M) to TFA. λ_{ex} = 266 nm (em slit: 5 nm), ex slit: 5 nm).

Scheme 7. Photolytic Decomplexation of Addition $\operatorname{Products}^{a}$



methods. The free diene could also be liberated without concomitant aromatization. This has previously proved to be difficult for iron complexes possessing an aromatic substituent, which is not conjugated to the diene.^{25,37} Photolytic demetallation of the methoxy-substituted adduct 14 instead resulted in the formation of unsaturated ketone 34 in a 74% yield (Scheme 7, entry 2). This type of reactivity is known upon decomplexation of 2-methoxy-substituted iron carbonyl cyclohexadiene complexes³⁸ and provides access to a different compound class in terms of substituted azulenes. Guaiazulene-derived adduct 27 could also be demetallated (Scheme 7, entry 3), affording the unsaturated ketone 35 in a 42% yield.

Another product class could be accessed by oxidative aromatization of diene **33** using 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ), producing 1,3-diphenylazulene (**36**) in an 82% yield (Scheme 8).





The C-4 methyl group of guaiazulene is acidic. Upon deprotonation, the formed anion is stabilized via conjugation with the electron-poor seven-membered ring. This has been utilized for condensation reactions with aldehydes under basic conditions.^{28a,39} Limited examples of conjugate additions have also been reported in this context.⁴⁰ Demetallated guaiazulene derivative **35** possesses an unsaturated ketone moiety in close proximity to the C-4 methyl group, which could allow for an intramolecular conjugate addition. We were pleased to see that the treatment of **35** with *t*-BuOK in tetrahydrofuran (THF) at 0 °C yielded a tetracyclic product **37** in a 39% yield (Scheme 9). A strong nuclear Overhauser effect (NOE) interaction





^aIsolated yields. ^bProduct 37 is racemic with the relative stereochemistry shown.

between the two ring-junction protons indicates a cis-fused ring system. The formed tetracyclic compound possesses an unusual 6/6/5/7 ring system, similar to the carbon skeleton of swinhoeisterols, isolated from the marine sponge *Theonella swinhoei.*⁴¹ Swinhoeisterols have been reported to have cytotoxic properties⁴¹ and have recently been the target of a total synthesis.⁴² We therefore envision that tetracyclic structures of this type could be of interest in the synthesis of natural products and bioactive compounds. A photoswitch incorporating guaiazulene and cyclohexene was recently reported by Hecht and co-workers, and compounds such as **35** and **37** may find additional applications in this area.⁵

CONCLUSIONS

Azulenes are here shown to be competent nucleophiles in the addition to cationic iron carbonyl dienyl complexes with different electronic properties. The reaction proceeds smoothly at room temperature with acetone as the solvent, using an inexpensive base (NaHCO₃) as the only additive. Apart from azulene itself, functionalized azulenes with extended conjugated systems, as well as derivatives of naturally occurring guaiazulene, were evaluated as nucleophiles affording the targeted products in a 43-98% yield. Photolytic decomplexation allowed the oxidative removal of iron under mild conditions. The synthetic utility of the formed azulenes was demonstrated via further transformations, including the formation of a tetracyclic product. UV-vis and fluorescence properties of selected products were also explored, displaying some unusual halochromic properties. In conclusion, we believe that the reported methodology provides a valuable new method for azulene derivatization and could find applications in areas such as optoelectronics, sensors, pharmaceuticals, and natural product synthesis.

EXPERIMENTAL SECTION

General Considerations. 1-(Azulen-1-yl) tetrahydrothiophenium hexafluorophosphate,^{17d} neutral iron carbonyl complex 32,²⁵ cationic iron carbonyl complexes 2,²⁵ 6,⁴³ and 7^{44} (starting from commercial 1-methoxy-1,3-cyclohexadiene, technical grade 65%), and guaiazulene derivatives 11,⁴⁵ 12,⁴⁶ 19,¹² 20-22,⁴⁷ and 23^{48} were synthesized according to literature procedures. All other solvents and reagents were purchased from commercial suppliers and used without purification or drying, unless otherwise stated. Photocatalytic decomplexation was performed using a low-energy black light lamp (Velleman lighting, 15 W, 850 lm). Structural assignments were made with additional information from gradient correlation spectroscopy (gCOSY), gradient heteronuclear single-quantum coherence (gHSQC), nuclear Overhauser enhancement spectroscopy (NOESY), and gradient heteronuclear multiple-bond correlation (gHMBC) experiments.

Analytical Methods. Column chromatography was performed on a Biotage Isolera Spektra One with Biotage SNAP KP-sil (silica gel) or KP-C18-HS (C18, reverse phase) columns. NMR spectra, NMR and ¹³C NMR, were recorded on an Agilent 400 MHz (101 MHz for ¹³C NMR). The chemical shifts for ¹H and ¹³C NMR spectra are reported in parts per million (ppm) using the residual solvent peak for reference. The following abbreviations are used for reporting NMR peaks: singlet (s), doublet (d), triplet (t), quartet (q), heptet (hept), multiplet (m), broad (br), and apparent (app). All coupling constants (J) are reported in hertz (Hz). For diastereomeric mixtures, peaks that can be attributed to single diastereomers are labeled d_1/d_2 . ATR-FTIR spectra were recorded on a PerkinElmer Spectrum Frontier infrared spectrometer with a pike-GladiATR module and reported in wavenumber (cm^{-1}) . Melting points were recorded on a Büchi Melting Point B-545. High-resolution mass spectrometry (HRMS) was performed on an Agilent 1290 infinity LC system equipped with an autosampler tandem to an Agilent 6520 Accurate Mass Q-TOF LC/MS.^{29a} Fluorescence spectra were acquired on an Agilent Technologies Cary Eclipse fluorescence spectrophotometer. UV-vis spectra were acquired on a PerkinElmer Lambda20 Spectrophotometer, using a Starna Silica (quartz) cuvette with 10 mm path length, two faces polished.

General Procedure A: Addition of Guaiazulene-Derived Nucleophiles to Cationic Iron Carbonyl Complexes. Cationic iron carbonyl complex (1.0 equiv), guaiazulene derivative (1.1 equiv), NaHCO₃ (1.5 equiv), and acetone (2.0 mL) were added to a 5 mL microwave vial equipped with a stir bar. The vial was capped and stirred at room temperature for 4 or 16 h. The crude reaction mixture was diluted with diethyl ether (5.0 mL), filtered through a pad of basic aluminum oxide, concentrated under a stream of N₂ gas, and purified by reversed-phase flash column chromatography (MeOH/ H_2O).

General Procedure B: Addition of Azulene-Derived Nucleophiles to Cationic Iron Carbonyl Complexes. Azulene derivative (1.0 equiv), cationic iron carbonyl complex (1.1 equiv), NaHCO₃ (2.0 equiv), and acetone (1.0 mL) were added to a 5 mL microwave vial equipped with a stir bar. The vial was capped and stirred at room temperature for 5 h. The crude reaction mixture was diluted with diethyl ether (3.0 mL), filtered through a pad of silica, concentrated under a stream of N₂ gas, and purified by flash column chromatography.

Tricarbonyl[methyl 5-(5-isopropyl-3,8-dimethylazulen-1yl)cyclohexa-1,3-diene-1-carboxylate]iron (3). Starting from Cationic Iron Carbonyl Complex 2. Synthesized according to general procedure A using iron complex 2 (42.2 mg, 0.100 mmol), guaiazulene 1 (21.8 mg, 0.110 mmol), and NaHCO₃ (12.6 mg, 0.150 mmol), with a reaction time of 4 h. The product was purified by reversed-phase chromatography (MeOH/H₂O, 60:40 \rightarrow 85:15), yielding 3 as a blue solid (47.4 mg, 97%).

Starting from Neutral Iron Carbonyl Complex 32. Iron complex 32 (30.0 mg, 0.102 mmol) and guaiazulene 1 (30.3 mg, 0.153 mmol) were added to a 5 mL microwave vial equipped with a stir bar. Acetonitrile (1.0 mL) containing tetrafluoroboric acid diethyl ether complex (15.3 μ L, 0.112 mmol) was added, and the vial was capped

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and stirred at room temperature for 24 h. The reaction mixture was diluted with diethyl ether, filtered through a pad of basic aluminum oxide, and the solvent was evaporated under a stream of N₂ gas. Purification by flash column chromatography yielded 3 as a blue solid (41.7 mg, 86%); mp 46–47 °C; ¹H NMR (400 MHz, chloroform-*d*) δ 8.02 (d, J = 2.2 Hz, 1H), 7.46 (s, 1H), 7.24 (dd, J = 10.6, 2.2 Hz, 1H), 6.83 (d, J = 10.6 Hz, 1H), 6.31 (dd, J = 4.4, 0.8 Hz, 1H), 5.56 (dd, J = 6.4, 4.4 Hz, 1H), 4.56 (app dt, J = 11.3, 3.9 Hz, 1H), 3.73 (s, 3H), 3.44 (ddd, *J* = 6.5, 3.3, 1.4 Hz, 1H), 3.00 (hept, *J* = 6.9 Hz, 1H), 2.96 (s, 3H), 2.87 (dd, J = 15.2, 11.3 Hz, 1H), 2.59 (s, 3H), 1.60 $(ddd, J = 15.2, 4.3, 0.9 \text{ Hz}, 1\text{H}), 1.32 (d, J = 6.9 \text{ Hz}, 6\text{H}); {}^{13}\text{C}{}^{1}\text{H}$ NMR (101 MHz, chloroform-d) δ 172.6, 144.5, 139.7, 138.5, 136.4, 134.5, 133.7, 131.8, 130.9, 126.8, 124.9, 88.7, 84.7, 69.1, 62.8, 51.6, 39.4, 37.6, 33.7, 27.4, 24.5, 13.1. Signal for Fe-CO not seen; FTIR-ATR ν_{max}/cm^{-1} 2053 (Fe–CO), 1975 (Fe–CO), 1705 (C=O); HRMS (ESI+) m/z: $[M + H]^+$ calcd for C₂₆H₂₇FeO₅ 475.1208; found 475.1199.

Hexacarbonyl[dimethyl 5,5'-(azulene-1,3-diyl)bis-(cyclohexa-1,3-diene-1-carboxylate)]diiron (4/4'). Synthesized according to general procedure B using azulene (12.9 mg, 0.101 mmol), iron complex 2 (93.4 mg, 0.221 mmol), and NaHCO₃ (33.8 mg, 0.403 mmol). The product was purified by reversed-phase flash chromatography (MeOH/H₂O 85:15), yielding an equimolar mixture of diastereomers 4 and 4' as a blue-green amorphous solid (55.1 mg, 81%); ¹H NMR (400 MHz, chloroform-d) δ 8.14 and 8.13 (d_1/d_2) (d, J = 9.6 Hz, 2H), 7.50 (app t, J = 9.8 Hz, 1H), 7.41 and 7.38 ($d_1/$ (d_1) (s, 1H), 7.02 and 7.02 (d_1/d_2) (app t, J = 9.9 Hz, 2H), 6.35–6.31 (m, 2H), 5.53 and 5.48 (d_1/d_2) (dd, J = 6.4, 4.4 Hz, 2H), 4.19–4.09 (m, 2H), 3.75 and 3.74 (d_1/d_2) (s, 6H), 3.44 and 3.40 (ddd, J = 6.4, 3.5, 1.4 Hz, 2H), 2.95 and 2.93 (d_1/d_2) (dd, J = 15.2, 11.4 Hz, 2H), 1.66 and 1.59 (d_1/d_2) (dd, J = 15.4, 4.0 Hz, 2H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 210.2 (br), 172.7, 172.6, 138.4, 138.4, 135.6, 135.5, 133.5, 133.5, 133.2, 133.1, 131.6, 131.4, 121.9, 121.8, 89.2, 89.2, 84.5, 67.6, 67.4, 63.0, 62.9, 51.9, 37.2, 37.1, 32.8, 32.6; FTIR-ATR ν_{max}/cm^{-1} 2048 (Fe-CO), 1967 (Fe-CO), 1703 (C=O); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₃₂H₂₅Fe₂O₁₀ 681.0147; found 681.0139.

Tricarbonyl[methyl 5-(azulen-1-yl)cyclohexa-1,3-diene-1carboxylate]iron (5). Synthesized according to general procedure B, using iron complex 2 (21.1 mg, 0.0500 mmol) and azulene (32.0 mg, 0.250 mmol). The product was purified by reversed-phase flash chromatography (methanol/water, $80:20 \rightarrow 90:10$), yielding 5 as a blue sticky solid (9.7 mg, 48%); ¹H NMR (400 MHz, chloroform-d) δ 8.26 (d, J = 9.1 Hz, 1H), 8.23 (d, J = 8.7 Hz, 1H), 7.70 (d, J = 3.9 Hz, 1H), 7.55 (app t, J = 9.9 Hz, 1H), 7.29 (d, J = 4.0 Hz, 1H), 7.12 (app t, J = 9.8 Hz, 1H), 7.08 (app t, J = 9.7 Hz, 1H), 6.30 (ddd, J = 4.4, 1.4, 0.9 Hz, 1H), 5.51 (dd, J = 6.4, 4.4 Hz, 1H), 4.20 (app dt, J = 11.4, 3.7 Hz, 1H), 3.73 (s, 3H), 3.46 (ddd, J = 6.4, 3.5, 1.4 Hz, 1H), 2.97 (ddd, J = 15.3, 11.4, 0.9 Hz, 1H), 1.67 (ddd, J = 15.4, 3.9, 0.9 Hz, 1H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 172.6, 141.1, 137.9, 137.0, 134.7, 134.7, 134.4, 133.1, 122.8, 122.0, 117.2, 89.3, 84.6, 67.7, 63.1, 51.8, 37.2, 32.9. Signal for Fe–CO not seen; FTIR-ATR $\nu_{\rm max}/$ 2047 (Fe-CO), 1969 (Fe-CO), 1711 (C=O); HRMS (ESI+) cm^{-1} m/z: $[M + H]^+$ calcd for $C_{21}H_{17}FeO_5$ 405.0425; found 405.0424.

1-Phenylazulene (8). Synthesized according to a literature procedure.^{17d} To a Radleys Carousel reaction tube equipped with a stirrer was added 1-(azulen-1-yl) tetrahydrothiophenium hexafluorophosphate (500 mg, 1.39 mmol), phenylboronic acid (203 mg, 1.67 mmol), K₃PO₄ (589 mg, 2.78 mmol), Xphos (60.2 mg, 0.139 mmol), and Pd(OAc)₂ (12.5 mg, 0.0557 mmol). The vial was capped, evacuated, and refilled with argon. Dimethylformamide (DMF) (10 mL) was added, and the vial was heated in a heating block at 75° for 6 h. The crude product was diluted with water and extracted with petroleum ether, and the organic phase was washed with brine and 5% aqueous lithium chloride. The solvent was evaporated under reduced pressure, and the crude product was purified by flash column chromatography (100% petroleum ether), yielding 8 as a blue solid (166.7 mg, 59%). ¹H NMR (400 MHz, chloroform-d) δ 8.57 (d, J = 9.8 Hz, 1H), 8.36 (d, J = 9.4 Hz, 1H), 8.03 (d, J = 3.8 Hz, 1H), 7.65-7.57 (m, 3H), 7.53-7.48 (m, 2H), 7.45 (d, J = 3.9 Hz, 1H), 7.38-

7.33 (m, 1H), 7.19–7.12 (m, 2H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-*d*) δ 141.8, 138.3, 137.7, 137.4, 137.3, 135.8, 135.4, 131.5, 129.9, 128.8, 126.4, 123.5, 123.2, 117.6. Analysis data are in accordance with published data for this compound.⁴⁹

1-(2-Thienyl)-azulene (9). Synthesized according to a literature procedure.^{17d} To a Radleys Carousel reduced volume reaction tube equipped with a stirrer was added 1-(azulen-1-yl) tetrahydrothiophenium hexafluorophosphate (250 mg, 0.694 mmol), 2-(thiopheneboronic acid)pinacol ester (175 mg, 0.833 mmol), K₃PO₄ (295 mg, 1.39 mmol), Xphos (33.1 mg, 0.0694 mmol), and Pd(OAc)₂ (6.2 mg, 0.028 mmol). The vial was capped, evacuated, and refilled with argon. DMF (5.0 mL) was added, and the vial was heated in a heating block at 75 °C for 6 h. The crude product was diluted with water and extracted with petroleum ether, and the organic phase was washed with brine and 5% aqueous lithium chloride. The solvent was evaporated under reduced pressure, and the crude product was purified by flash column chromatography (100% petroleum ether), yielding 9 as a blue-green oil (77.5 mg, 53%). ¹H NMR (400 MHz, chloroform-d) δ 8.77 (d, J = 9.8 Hz, 1H), 8.32 (d, J = 9.4 Hz, 1H), 8.07 (d, J = 4.0 Hz, 1H), 7.64-7.67 (m, 1H), 7.41 (d, J = 4.0 Hz, 1H), 7.38–7.35 (m, 1H), 7.32–7.29 (m, 1H), 7.23–7.15 (m, 3H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 142.3, 139.8, 138.7, 137.5, 137.4, 135.9, 135.1, 127.8, 124.7, 124.5, 123.8, 123.7, 123.7, 117.9. Analysis data are in accordance with published data for this compound.⁵

(Azulen-1-ylethynyl)triisopropylsilane (10). To a microwave reaction vial was added 1-(azulen-1-yl) tetrahydrothiophenium hexafluorophosphate (100 mg, 0.278 mmol), CuI (5.3 mg, 0.028 mmol), K₃PO₄ (64.8 mg, 0.305 mmol), t-BuXphos (11.8 mg, 0.0278 mmol), and Pd(OAc)₂ (3.1 mg, 0.014 mmol). The vial was capped, evacuated, and refilled with argon. (Triisopropylsilyl)acetylene (94 μ L, 0.419 mmol) and 2.0 mL DMF were added, and the vial was heated in a heating block at 80 °C for 16 h. The crude product was diluted with water and extracted with petroleum ether. The organic phase was then washed with brine and 5% aqueous lithium chloride. The solvent was evaporated under reduced pressure, and the crude product was purified by flash column chromatography (100% petroleum ether), yielding 10 as a blue oil (23.7 mg, 27%). ¹H NMR (400 MHz, chloroform-d) δ 8.59 (d, J = 9.6 Hz, 1H), 8.28 (d, J = 9.4 Hz, 1H), 7.98 (d, I = 4.0 Hz, 1H), 7.63 (app t, I = 9.9 Hz, 1H), 7.30-7.18 (m, 3H), 1.22-1.19 (m, 21H); ¹³C{¹H} NMR (101 MHz, chloroform-d) & 142.0, 141.3, 140.0, 138.7, 137.3, 136.5, 124.7, 124.2, 117.5, 111.4, 103.3, 94.8, 19.0, 11.7; FTIR-ATR ν_{max}/cm^{-1} 2133 $(C \equiv C)$; HRMS (ESI+) m/z: $[M + H]^+$ calcd for $C_{21}H_{29}Si$ 309.2033; found 309.2048.

Tricarbonyl[1-(cyclohexa-2,4-dien-1-yl)-3-phenylazulene]iron (13). Synthesized according to general procedure B using azulene derivative 8 (20.6 mg, 0.101 mmol), iron complex 6 (40.5 mg, 0.111 mmol), and NaHCO₃ (16.9 mg, 0.202 mmol). The product was purified by flash chromatography (100% petroleum ether), yielding 13 as a blue-green solid (40.6 mg, 95%); mp 51-55 °C; ¹H NMR (400 MHz, chloroform-d) δ 8.41 (dd, J = 9.7, 0.9 Hz, 1H), 8.23 (d, J = 9.5 Hz, 1H), 7.84 (s, 1H), 7.61-7.56 (m, 2H), 7.55-7.46 (m, 3H), 7.35 (app ddt, J = 7.9, 6.8, 1.3 Hz, 1H), 7.06 (app t, J = 9.8 Hz, 1H), 7.03 (app t, J = 9.7 Hz, 1H), 5.59 (ddd, J = 6.5, 3.3, 2.0 Hz, 1H), 5.46 (ddd, J = 5.9, 4.1, 1.5 Hz, 1H), 4.06 (dt, J = 11.2, 3.6 Hz, 1H), 3.33 (ddd, J = 6.4, 3.6, 1.4 Hz, 1H), 3.31–3.27 (m, 1H), 2.57 (ddd, J = 15.2, 11.1, 3.8 Hz, 1H), 1.90-1.83 (m, 1H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 212.3, 138.6, 137.4, 136.3, 136.1, 135.8, 134.9, 134.9, 133.5, 130.3, 129.9, 128.8, 126.5, 123.1, 122.0, 86.4, 84.7, 66.9, 60.8, 35.7, 34.3; FTIR-ATR ν_{max} /cm⁻¹ 2035 (Fe–CO), 1946 (Fe– CO); HRMS (ESI+) m/z: $[M + H]^+$ calcd for C₂₅H₁₉FeO₃ 423.0683; found 423.0695.

Tricarbonyl[1-(4-methoxycyclohexa-2,4-dien-1-yl)-3-phenylazulene]iron (14). Synthesized according to general procedure B using azulene derivative 8 (100 mg, 0.490 mmol), iron complex 7 (212 mg, 0.539 mmol), and NaHCO₃ (82.3 mg, 0.979 mmol) in 4.0 mL acetone. The product was purified by flash chromatography (petroleum ether/ethyl acetate, 98:2), yielding 14 as a blue-green solid (209 mg, 94%); mp 53–58 °C; ¹H NMR (400

MHz, chloroform-*d*) δ 8.43 (d, J = 9.7 Hz, 1H), 8.23 (d, J = 9.7 Hz, 1H), 7.83 (s, 1H), 7.63–7.58 (m, 2H), 7.57–7.49 (m, 3H), 7.41–7.35 (m, 1H), 7.07 (app t, J = 9.8 Hz, 1H), 7.04 (app t, J = 9.8 Hz, 1H), 5.22 (dd, J = 6.5, 2.2 Hz, 1H), 3.90 (app dt, J = 11.1, 3.5 Hz, 1H), 3.72 (s, 3H), 3.56 (app dtd, J = 3.4, 2.3, 0.8 Hz, 1H), 2.94 (dd, J = 6.5, 3.5 Hz, 1H), 2.60 (ddd, J = 14.8, 11.1, 3.7 Hz, 1H), 1.99 (app dt, J = 15.2, 2.9 Hz, 1H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 140.4, 138.6, 137.5, 136.4, 136.0, 135.8, 135.0, 134.7, 133.6, 130.3, 130.0, 128.8, 126.5, 123.1, 122.0, 66.8, 56.0, 54.6, 53.8, 35.4, 35.1, signal for Fe–CO not seen; FTIR-ATR ν_{max} /cm⁻¹ 2035 (Fe–CO), 1946 (Fe–CO); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₂₆H₂₁FeO₄ 453.0789; found 453.0787.

Tricarbonyl[2-(3-(cyclohexa-2,4-dien-1-yl)azulen-1-yl)thiophene]iron (15). Synthesized according to general procedure B using azulene derivative 9 (10.5 mg, 0.0499 mmol), iron complex 6 (20.0 mg, 0.0550), and NaHCO₃ (8.4 mg, 0.10 mmol) in 0.5 mL acetone. The product was purified by flash chromatography (petroleum ether/EtOAc 99:1), yielding 15 as a sticky green solid (15.6 mg, 73%); ¹H NMR (400 MHz, chloroform-d) δ 8.61 (d, J = 9.6 Hz, 1H), 8.18 (d, J = 9.6 Hz, 1H), 7.86 (s, 1H), 7.53 (app t, J =9.8 Hz, 1H), 7.36 (dd, J = 5.1, 1.2 Hz, 1H), 7.25 (dd, J = 3.6, 1.2 Hz, 1H), 7.18 (dd, J = 5.1, 3.5 Hz, 1H), 7.07 (app t, J = 9.8 Hz, 1H), 7.06 (app t, J = 9.8 Hz, 1H), 5.63–5.58 (m, 1H), 5.48 (ddd, J = 6.0, 4.1, 1.5 Hz, 1H), 4.02 (app dt, J = 11.1, 3.6 Hz, 1H), 3.35–3.24 (m, 2H), 2.56 (ddd, J = 15.1, 11.1, 3.8 Hz, 1H), 1.83 (dddd, J = 15.2, 3.5, 2.2, 1.1 Hz, 1H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 212.2, 139.5, 139.0, 137.0, 135.9, 135.9, 135.2, 134.9, 133.7, 127.8, 124.9, 124.7, 123.4, 122.6, 122.5, 86.4, 84.7, 66.7, 60.7, 35.7, 34.1; FTIR-ATR ν_{max}/cm^{-1} 2035 (Fe–CO), 1946 (Fe–CO); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₂₃H₁₇FeO₃S 429.0248; found 429.0246.

Tricarbonyl[((3-(cyclohexa-2,4-dien-1-yl)azulen-1-yl)ethynyl)triisopropylsilane]iron (16). Synthesized according to general procedure B using azulene derivative 10 (11.2 mg, 0.0363 mmol), iron complex 6 (14.5 mg, 0.0398 mmol), and NaHCO₃ (6.1 mg, 0.073 mmol) in 0.5 mL acetone. The product was purified by reversed-phase flash chromatography (MeOH/H₂O 70:30 \rightarrow 100:0), yielding 16 as a green sticky solid (13.3 mg, 70%); ¹H NMR (400 MHz, chloroform-d) δ 8.45 (d, J = 9.6 Hz, 1H), 8.15 (d, J = 9.6 Hz, 1H), 7.81 (s, 1H), 7.56 (app t, J = 9.8 Hz, 1H), 7.15 (app t, J = 9.9Hz, 1H), 7.12 (app t, J = 9.9 Hz, 1H), 5.61–5.57 (m, 1H), 5.48 (ddd, J = 6.0, 4.1, 1.5 Hz, 1H), 3.96 (app dt, J = 11.2, 3.6 Hz, 1H), 3.27-3.23 (m, 2H), 2.51 (ddd, J = 15.1, 11.2, 3.8 Hz, 1H), 1.79–1.72 (m, 1H), 1.21–1.19 (m, 21H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 212.2, 142.7, 139.0, 137.2, 136.5, 136.3, 135.1, 133.7, 123.8, 123.7, 110.1, 103.1, 95.2, 86.5, 84.8, 77.5, 77.2, 76.8, 66.4, 60.6, 35.6, 34.0, 19.0, 11.7; FTIR-ATR $\nu_{\rm max}/{\rm cm}^{-1}$ 2133 (C=C), 2039 (Fe-CO), 1954 (Fe-CO); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₃₀H₃₅FeO₃Si 527.1705; found 527.1727.

Tricarbonyl[3-(cyclohexa-2,4-dien-1-yl)azulene-1carbaldehyde]iron (17). Synthesized according to general procedure B using azulene derivative 11 (26.4 mg, 0.169 mmol), iron complex 6 (67.7 mg, 0.186 mmol), and NaHCO₃ (28.4 mg, 0.338 mmol), with an extended reaction time of 16 h. The product was purified by reversed-phase flash chromatography (MeOH/H2O 86:14), yielding 17 as a green solid (31.8 mg, 50%); mp 131-134 °C; ¹H NMR (400 MHz, chloroform-d) δ 10.87 (s (br), 1H), 9.44 (d, J = 9.0 Hz, 1H), 8.39 (d, J = 9.7 Hz, 1H), 8.17 (s, 1H), 7.83 (app t, J = 9.6 Hz, 1H), 7.58-7.42 (m, 2H), 5.65-5.57 (m, 1H), 5.55-5.47 (m, 1H), 3.98 (app dt, J = 11.2, 3.4 Hz, 1H), 3.30-3.23 (m, 2H), 2.56 (ddd, J = 15.1, 11.2, 3.7 Hz, 1H), 1.76 (d, J = 15.4 Hz, 1H); $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR (101 MHz, DMSO- d_{6}) δ 212.8, 186.2, 141.2, 141.1, 141.0, 137.8, 137.2, 136.9, 136.8, 129.6, 128.5, 124.4, 87.3, 85.4, 66.9, 61.9, 34.9, 34.0; FTIR-ATR $\nu_{\text{max}}/\text{cm}^{-1}$ 2032 (Fe–CO), 1944 (Fe– CO), 1638 (C=O); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₂₀H₁₅FeO₄ 375.0320; found 375.0320.

Hexacarbonyl[1,3-di(cyclohexa-2,4-dien-1-yl)-6phenylazulene]diiron (18, 18'). Synthesized according to general procedure A using azulene (21.0 mg, 0.103 mmol), iron complex 6 (82.3 mg, 0.226 mmol), and NaHCO₃ (34.5 mg, 0.411 mmol). The product was purified by reversed-phase flash chromatography (MeOH/H₂O 95:5), yielding an equimolar mixture of diastereomers **18** and **18**' as a green sticky solid (54.8 mg, 83%); ¹H NMR (400 MHz, chloroform-*d*) δ 8.14 (d, *J* = 9.5 Hz, 2H), 7.64–7.59 (m, 2H), 7.52–7.40 (m, 4H), 7.19 (d, *J* = 10.2 Hz, 2H), 5.68–5.60 (m, 2H), 5.53–5.43 (m, 2H), 4.01 (app dt, *J* = 11.2, 3.6 Hz, 2H), 3.35–3.24 (m, 4H), 2.60–2.46 (m, 2H), 1.86–1.75 (m, 2H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 212.3, 151.7, 145.2, 134.9, 134.9, 134.3, 134.3, 132.4, 132.2, 132.1, 128.8, 128.4, 128.1, 122.0, 86.3, 86.2, 84.7, 84.7, 67.1, 67.1, 60.8, 60.8, 36.0, 36.0, 34.0, 33.9; FTIR-ATR $\nu_{max}/$ cm⁻¹ 2037 (Fe–CO), 1942 (Fe–CO); HRMS (ESI+) *m/z*: [M + H]⁺ calcd for C₃₄H₂₅Fe₂O₆ 641.0349; found 641.0340.

7-Isopropyl-1,4-dimethylazulen-2-amine (24). To a microwave reaction vial equipped with a magnetic stirring bar was added 2-(7-isopropyl-1,4-dimethyl-azulen-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane 20 (111.7 mg, 0.344 mmol), hydroxylamine-O-sulfonic acid (81.2 mg, 0.718 mmol), acetonitrile (2.0 mL), and 1 M aqueous NaOH (2.0 mL). The vial was capped and stirred at room temperature for 24 h. The reaction mixture was diluted with 2 mL water and extracted with EtOAc (2×10 mL), and the organic phase was dried over Na₂SO₄ and concentrated under reduced pressure. The crude product was purified using flash column chromatography (petroleum ether/EtOAc, 100:0 \rightarrow 95:5), yielding 24 as a yellowbrown viscous liquid (26.9 mg, 37%). ¹H NMR (400 MHz, chloroform-d) δ 7.81 (d, J = 1.8 Hz, 1H), 7.12 (dd, J = 10.6, 1.8 Hz, 1H), 6.99 (d, J = 10.6 Hz, 1H), 6.59 (s, 1H), 4.39 (s, 2H), 3.05 (hept, J = 6.9 Hz, 1H), 2.73 (s, 3H), 2.39 (d, J = 0.6 Hz, 3H), 1.35 (d, I = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 153.4, 141.5, 138.0, 137.3, 135.8, 128.3, 126.7, 126.3, 107.4, 99.7, 38.7, 25.0, 24.2, 8.9; FTIR-ATR $\nu_{\rm max}/{\rm cm}^{-1}$ 3471 (N – H), 3355 (N – H), 1617 (N - H); HRMS (ESI+) m/z: $[M + H]^+$ calcd for $C_{15}H_{20}N$ 214.1596; found 214.1605.

N-(7-Isopropyl-1,4-dimethylazulen-2-yl)acetamide (25). To a microwave reaction vial equipped with a magnetic stirring bar was added 2-(7-isopropyl-1,4-dimethyl-azulen-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane 20 (213.0 mg, 0.657 mmol), hydroxylamine-Osulfonic acid (223.0 mg, 1.97 mmol), acetonitrile (3.0 mL), and 1 M aqueous NaOH (4.0 mL). The vial was capped and stirred at room temperature for 24 h. The reaction mixture was diluted with water (5 mL) and extracted with dichloromethane. The organic phase was dried over Na2SO4, and the solvent was evaporated under reduced pressure. The crude aminoazulene 24 was then redissolved in dichloromethane (10.0 mL). Acetic anhydride (0.5 mL) and pyridine (0.5 mL) were added, and the reaction was stirred at room temperature for 14 h. The reaction mixture was reduced in vacuo, and the crude product was purified using flash column chromatography (20% EtOAc in petroleum ether), yielding 25 as a blue crystalline solid (60.0 mg, 36%); mp 153-154 °C; ¹H NMR (400 MHz, chloroform-d) δ 8.07 (d, J = 1.9 Hz, 1H), 7.91 (s, 1H), 7.66 (s (br), 1H), 7.34 (dd, J = 10.8, 1.9 Hz, 1H), 7.09 (d, J = 10.7 Hz, 1H), 3.09 (hept, J = 6.9 Hz, 1H), 2.84 (s, 3H), 2.51 (s, 3H), 2.32 (s, 3H), 1.37 (d, J = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, DMSO- d_6) δ 168.6, 143.9, 140.4, 140.1, 136.2, 134.6, 131.5, 129.9, 126.1, 111.3, 104.7, 37.6, 24.7, 23.9, 23.7, 9.4; FTIR-ATR $\nu_{\rm max}/{\rm cm}^{-1}$ 3302 (N -H), 1665 (C=O); HRMS (ESI+) m/z: $[M + H]^+$ calcd for C₁₇H₂₂NO 256.1701; found 256.1711.

Tricarbonyl[3-(cyclohexa-2,4-dien-1-yl)-7-isopropyl-1,4-dimethylazulene]iron (26). Synthesized according to general procedure A using guaiazulene **1** (21.8 mg, 0.110 mmol), iron complex **6** (36.4 mg, 0.100 mmol), and NaHCO₃ (12.6 mg, 0.150 mmol), with a reaction time of 16 h. The product was purified by reversed-phase chromatography (MeOH/H₂O, 60:40 \rightarrow 85:15), yielding **26** as a blue sticky solid (41.6 mg, 98%); this product was also synthesized on a 1 mmol scale, yielding 392 mg of **26** (94%); ¹H NMR (400 MHz, chloroform-*d*) δ 8.00 (d, *J* = 2.1 Hz, 1H), 7.53 (s, 1H), 7.21 (dd, *J* = 10.6, 2.2 Hz, 1H), 6.79 (d, *J* = 10.6 Hz, 1H), 5.63–5.57 (m, 1H), 5.49 (ddd, *J* = 5.9, 4.1, 1.5 Hz, 1H), 4.39 (app dt, *J* = 11.0, 3.7 Hz, 1H), 3.30–3.22 (m, 2H), 2.99 (hept, *J* = 6.9 Hz, 1H), 2.90 (s, 3H), 2.59 (d, *J* = 0.7 Hz, 3H), 2.42 (ddd, *J* = 15.2, 11.0, 4.0 Hz, 1H), 1.73 (dddd, *J* = 15.2, 3.7, 2.2, 1.1 Hz, 1H), 1.31 (d, *J* = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 212.3, 144.5,

139.5, 138.4, 136.7, 134.4, 133.5, 132.8, 130.9, 126.5, 124.8, 85.8, 84.8, 68.5, 60.5, 38.1, 37.6, 35.0, 27.3, 24.5, 13.1; FTIR-ATR ν_{max}/cm^{-1} 2040 (Fe–CO), 1957 (Fe–CO); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₂₄H₂₅FeO₃ 417.1153; found 417.1155.

Tricarbonyl[7-isopropyl-3-(4-methoxycyclohexa-2,4-dien-1yl)-1,4-dimethylazulene]iron (27). Synthesized according to general procedure A using guaiazulene 1 (21.8 mg, 0.110 mmol), iron complex 7 (39.4 mg, 0.100 mmol), and NaHCO₃ (12.6 mg, 0.150 mmol), with a reaction time of 16 h. The product was purified by reversed-phase chromatography (MeOH/H₂O, $60:40 \rightarrow 85:15$), yielding 27 as a blue solid (41.4 mg, 93%); mp 101-103 °C; ¹H NMR (400 MHz, chloroform-d) δ 8.01 (d, J = 2.2 Hz, 1H), 7.49 (s, 1H), 7.22 (dd, J = 10.6, 2.2 Hz, 1H), 6.79 (d, J = 10.7 Hz, 1H), 5.26 (dd, J = 6.6, 2.4 Hz, 1H), 4.22 (app dt, J = 11.0, 3.6 Hz, 1H), 3.74 (s, 3H), 3.50 (app dt, J = 3.9, 2.3 Hz, 1H), 2.99 (hept, J = 6.9 Hz, 1H), 2.90 (s, 3H), 2.85 (dd, J = 6.6, 3.4 Hz, 1H), 2.60 (s, 3H), 2.44 (ddd, J= 14.8, 10.9, 3.9 Hz, 1H), 1.85 (ddd, J = 14.7, 3.7, 2.3 Hz, 1H), 1.32 (d, J = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 211.7 (br), 144.7, 140.3, 139.6, 138.5, 136.6, 134.6, 133.7, 133.0, 131.1, 126.7, 124.9, 67.2, 57.5, 54.6, 53.6, 38.0, 37.7, 36.2, 27.5, 24.7, 13.2; FTIR-ATR ν_{max}/cm^{-1} 2036 (Fe–CO), 1958 (Fe–CO); HRMS (ESI +) m/z: $[M + H]^+$ calcd for C₂₅H₂₇FeO₄ 447.1258; found 447.1258.

Tricarbonyl[methyl 5-(3-formyl-5-isopropyl-8-methylazulen-1-yl)cyclohexa-1,3-diene-1-carboxylate]iron (28). Synthesized according to general procedure A using azulene derivative 19 (23.4 mg, 0.110 mmol), iron complex 2 (42.2 mg, 0.100 mmol), and NaHCO₂ (12.6 mg, 0.150 mmol), with a reaction time of 16 h. The product was purified by reversed-phase chromatography (MeOH/ $H_2O 60:40 \rightarrow 85:15$), yielding 28 as a purple solid (27.3 mg, 56%); mp 74-76 °C; ¹H NMR (400 MHz, chloroform-d) δ 10.27 (s, 1H), 9.60 (s, 1H), 8.10 (s, 1H), 7.57 (dd, J = 10.8, 2.2 Hz, 1H), 7.35 (d, J = 10.7 Hz, 1H), 6.33 (d, J = 4.3 Hz, 1H), 5.66 (dd, J = 6.3, 4.4 Hz, 1H), 4.48 (app dt, J = 11.4, 4.0 Hz, 1H), 3.72 (s, 3H), 3.44 (ddd, J = 6.4, 3.2, 1.4 Hz, 1H), 3.15 (hept, J = 6.9 Hz, 1H), 3.07 (s, 3H), 2.91 (dd, J = 15.1, 11.3 Hz, 1H), 1.54 (dd, J = 15.1, 4.4 Hz, 1H), 1.37 (d, J = 6.9 Hz, 6H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 185.9, 172.3, 149.0, 148.0, 141.6, 141.3, 139.4, 137.3, 137.2, 135.3, 133.0, 124.1, 89.1, 84.8, 67.7, 62.4, 51.7, 39.3, 37.9, 33.8, 27.5, 24.4. Signal for Fe-CO not seen; FTIR-ATR $\nu_{\rm max}/{\rm cm}^{-1}$ 2050 (Fe–CO), 1974 (Fe–CO), 1707 (C=O), 1643 (C=O); HRMS (ESI+) m/z: $[M + H]^+$ calcd for C₂₆H₂₅FeO₆ 489.1000; found 489.1016.

Tricarbonyl[methyl 5-(5-isopropyl-3,8-dimethyl-2-(4,4,5,5tetramethyl-1,3,2-dioxaborolan-2-yl)azulen-1-yl)cyclohexa-1,3-diene-1-carboxylate]iron (29). Synthesized according to general procedure A using azulene derivative 20 (35.7 mg, 0.110 mmol), iron complex 2 (42.2 mg, 0.100 mmol), and NaHCO₃ (12.6 mg, 0.150 mmol), with a reaction time of 16 h. The product was purified by reversed-phase chromatography (MeOH/H₂O 60:40 \rightarrow 90:10), yielding 29 as a green oil (26.5 mg, 43%); ¹H NMR (400 MHz, chloroform-*d*) δ 8.01 (d, *J* = 2.0 Hz, 1H), 7.20 (dd, *J* = 10.6, 2.1 Hz, 1H), 6.83 (d, J = 10.6 Hz, 1H), 6.30 (d, J = 4.3 Hz, 1H), 5.39 (dd, J = 6.5, 4.3 Hz, 1H), 4.82 (ddd, J = 11.0, 5.7, 2.6 Hz, 1H), 3.73 (s, 3H), 3.37 (ddd, J = 6.4, 2.6, 1.2 Hz, 1H), 3.00 (s, 3H), 2.97 (hept, J = 6.9 Hz, 1H), 2.76 (dd, J = 15.0, 11.1 Hz, 1H), 2.64 (s, 3H), 1.88 (dd, J = 15.0, 5.8 Hz, 1H), 1.48 (s, 6H), 1.44 (s, 6H), 1.31 (d, J = 6.9 Hz, 6H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 210.8 (br), 173.0, 144.6, 140.0, 139.3, 135.0, 134.8, 133.6, 133.2, 131.2, 127.6, 87.9, 85.6, 84.0, 71.4, 62.9, 51.7, 41.4, 37.6, 29.83, 28.55, 25.90, 25.48, 24.58, 13.12. Signal for C-B not seen; FTIR-ATR ν_{max}/cm^{-1} 2045 (Fe-CO), 1968 (Fe-CO), 1708 (CO); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₃₂H₃₈BFeO₇ 601.2059; found 601.2049.

Tricarbonyl[methyl 5-(2-iodo-5-isopropyl-3,8-dimethylazulen-1-yl)cyclohexa-1,3-diene-1-carboxylate]iron (30). Synthesized according to general procedure A using azulene derivative 21 (28.6 mg, 0.088 mmol), iron complex 2 (40.9 mg, 0.097 mmol), and NaHCO₃ (11.1 mg, 0.132 mmol), with a reaction time of 16 h. The product was purified by reversed-phase chromatography (MeOH/ H_2O , 60:40 \rightarrow 90:10), yielding 30 as a blue oil (33.4 mg, 63%); although stable in neat form, the product was found to be somewhat unstable in chloroform-*d* solution. Increased stability was observed when the CDCl₃ was filtered through a plug of basic aluminum oxide prior to use; ¹H NMR (400 MHz, chloroform-*d*) δ 8.13 (d, *J* = 2.1 Hz, 1H), 7.34 (dd, *J* = 10.7, 2.1 Hz, 1H), 6.98 (d, *J* = 10.7 Hz, 1H), 6.33 (d, *J* = 4.3 Hz, 1H), 5.66 (dd, *J* = 6.5, 4.3 Hz, 1H), 4.97 (ddd, *J* = 11.0, 6.4, 2.5 Hz, 1H), 3.74 (s, 3H), 3.13 (d, *J* = 5.6 Hz, 1H), 3.09–2.97 (m, 4H), 2.65 (dd, *J* = 14.8, 11.2 Hz, 1H), 2.59 (s, 3H), 1.97 (dd, *J* = 14.8, 6.3 Hz, 1H), 1.33 (d, *J* = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 210.9 (br), 173.0, 143.3, 141.8, 136.2, 134.9, 133.8, 133.2, 129.0, 128.8, 126.4, 105.7, 88.9, 86.4, 67.5, 63.2, 51.8, 40.5, 37.7, 29.0, 26.8, 24.7, 16.3; FTIR-ATR ν_{max} /cm⁻¹ 2048 (Fe–CO), 1972 (Fe–CO), 1706 (C=O); HRMS (ESI+) *m*/*z*: [M + H]⁺ calcd for C₂₆H₂₆FeIO₅ 601.0174; found 601.0177.

Tricarbonyl[methyl 5-(2-bromo-5-isopropyl-3,8-dimethylazulen-1-yl)cyclohexa-1,3-diene-1-carboxylate]iron (31). Synthesized according to general procedure A using azulene derivative 22 (10.0 mg, 0.0361) mmol, iron complex 2 (13.9 mg, 0.0329), and NaHCO₃ (5.5 mg, 0.066 mmol) in 0.5 mL acetone, with a reaction time of 4 h. The product was purified by reversed-phase chromatography (MeOH/H₂O 60:40 \rightarrow 90:10), yielding 31 as a blue solid (14.4 mg, 79%). Although stable in neat form, the product was found to be somewhat unstable in chloroform-d solution. Increased stability was observed when the CDCl₃ was filtered through a plug of basic aluminum oxide prior to use; mp 147-149 °C; ¹H NMR (400 MHz, chloroform-d) δ 8.10 (d, I = 2.2 Hz, 1H), 7.33 (dd, *J* = 10.7, 2.1 Hz, 1H), 6.99 (d, *J* = 10.7 Hz, 1H), 6.30 (dd, *J* = 4.3, 1.1 Hz, 1H), 5.61 (dd, J = 6.4, 4.4 Hz, 1H), 4.90 (ddd, J = 10.9, 5.9, 2.6 Hz, 1H), 3.73 (s, 3H), 3.15 (ddd, J = 6.5, 2.7, 1.3 Hz, 1H), 3.09–2.97 (m, 4H), 2.65 (dd, J = 14.7, 11.3 Hz, 1H), 2.55 (s, 3H), 1.93 (ddd, J = 14.8, 6.0, 0.9 Hz, 1H), 1.33 (d, J = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, methylene chloride- d_2) δ 173.1, 144.3, 142.3, 136.5, 135.1, 133.4, 133.2, 129.6, 129.4, 125.1, 124.7, 88.2, 87.2, 67.6, 63.8, 51.9, 40.0, 38.1, 29.0, 27.6, 24.8, 12.7. Signal for Fe-CO not seen; FTIR-ATR ν_{max}/cm^{-1} 2047 (Fe–CO), 1968 (Fe–CO), 1705 (C= O); HRMS (ESI+) m/z: $[M + H]^+$ calcd for $C_{26}H_{26}BrFeO_5$ 553.0313; found 553.0324.

General Procedure for the Photocatalytic Decomplexation, Exemplified by the Formation of 1-(Cyclohexa-2,4-dien-1-yl)-3phenylazulene (33). To a 100 mL borosilicate glass round-bottom flask equipped with a magnetic stirring bar, 25.0 mg (0.0592 mmol) of 13 was added and dissolved in 24 mL of acetonitrile. The flask was sealed with a rubber septum, which was pierced with a syringe needle open to the atmosphere. The flask was placed 5 cm from a low-energy black light lamp (Velleman lighting, 15 W, 850 lm) in a container lined with aluminum foil. The reaction mixture was irradiated (emission maximum of 370 nm; for emission spectrum, see Figure S71) without the use of any filter under stirring in room temperature, cooled by a stream of air, for 72 h. The solvent was removed under reduced pressure, and the residue was redissolved in dichloromethane, filtered through a plug of silica, and the solvent was evaporated under a stream of nitrogen. The crude product was purified by reversedphase flash chromatography (methanol/water, 84:16), yielding 33 as a green sticky solid (13.1 mg, 78%); ¹H NMR (400 MHz, chloroformd) δ 8.49 (d, J = 9.8 Hz, 1H), 8.36 (d, J = 9.7 Hz, 1H), 8.04 (s, 1H), 7.64-7.61 (m, 2H), 7.57-7.45 (m, 3H), 7.40-7.32 (m, 1H), 7.14-7.02 (m, 2H), 6.16-6.04 (m, 2H), 6.04-5.99 (m, 1H), 5.94-5.88 (m, 1H), 4.34 (app ddt, J = 12.5, 9.2, 3.0 Hz, 1H), 2.68 (dddd, J = 17.3, 9.2, 4.9, 1.3 Hz, 1H), 2.57 (dddd, J = 17.2, 13.3, 3.8, 2.3 Hz, 1H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 138.4, 137.6, 136.8, 136.6, 136.0, 135.6, 133.7, 132.9, 131.1, 130.1, 129.9, 128.7, 126.4, 126.2, 124.3, 124.2, 123.0, 122.0, 32.1, 31.6; HRMS (ESI+) m/z: [M + H]⁺ calcd for C₂₂H₁₉ 283.1486; found 283.1483.

4-(3-Phenylazulen-1-yl)cyclohex-2-en-1-one (34). Synthesized according to the general procedure, using 25.0 mg (0.0553 mmol) of compound 14 dissolved in 25 mL of acetonitrile. The crude product was purified by flash chromatography (petroleum ether/EtOAc 93:7), yielding 34 as a green sticky solid (12.2 mg, 74%); ¹H NMR (400 MHz, chloroform-*d*) δ 8.53 (d, *J* = 10.0 Hz, 1H), 8.36 (d, *J* = 9.4 Hz, 1H), 7.87 (s, 1H), 7.65–7.56 (m, 3H), 7.52–7.46 (m, 2H), 7.38–7.33 (m, 1H), 7.19–7.11 (m, 3H), 6.23 (ddd, *J* = 10.1, 2.5, 0.6 Hz, 1H), 4.46 (ddt, *J* = 8.3, 5.3, 2.8 Hz, 1H), 2.69–2.47 (m,

3H), 2.36–2.25 (m, 1H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 199.7, 153.9, 138.9, 137.1, 136.7, 136.3, 136.2, 136.1, 133.7, 130.3, 129.9, 129.6, 129.4, 128.8, 126.7, 123.6, 122.5, 37.3, 35.2, 31.9; FTIR-ATR ν_{max} /cm⁻¹ 1674 (C=O); HRMS (ESI+) *m*/*z*: [M + H]⁺ calcd for C₂₂H₁₉O 299.1436; found 299.1440.

4-(5-lsopropyl-3,8-dimethylazulen-1-yl)cyclohex-2-en-1one (35). Synthesized according to the general procedure, using 41.4 mg (0.0928 mmol) of compound 27, dissolved in 42 mL of acetonitrile in a 500 mL round-bottom flask. The crude product was purified by flash chromatography (petroleum ether/EtOAc 93:7), yielding 35 as a blue sticky oil (11.4 mg, 42%); ¹H NMR (400 MHz, chloroform-d) δ 8.13 (d, J = 2.2 Hz, 1H), 7.47 (s, 1H), 7.33 (dd, J =10.7, 2.2 Hz, 1H), 7.10 (ddd, J = 10.1, 3.0, 1.2 Hz, 1H), 6.92 (d, J = 10.7 Hz, 1H), 6.16 (dd, J = 10.1, 2.5 Hz, 1H), 4.76 (app ddt, J = 7.9, 5.0, 2.8 Hz, 1H), 3.05 (hept, J = 6.9, 1H) 3.02 (s, 3H), 2.65–2.57 (m, 4H), 2.55-2.44 (m, 2H), 2.23-2.11 (m, 1H), 1.35 (d, J = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, chloroform-*d*) δ 199.9, 155.4, 144.7, 140.1, 138.1, 137.8, 135.2, 134.1, 131.6, 128.8, 128.0, 127.3, 125.0, 37.8, 37.4, 37.0, 33.9, 27.5, 24.7, 13.1; FTIR-ATR $\nu_{\rm max}/{\rm cm}^{-1}$ 1679 (C=O); HRMS (ESI+) m/z: [M + H]⁺ calcd for C₂₁H₂₅O 293.1905; found 293.1904.

1,3-Diphenylazulene (36).^{16b} To a 5 mL microwave vial equipped with a stir bar, containing 27.4 mg (0.0970 mmol) of compound 33, was added 24.2 mg (0.107 mmol) DDQ. The vial was put under an atmosphere of nitrogen. Toluene (1.0 mL) was added, and the solution was stirred at room temperature for 40 min. The reaction mixture was diluted with ethyl acetate (20 mL) and washed with 1 M aqueous NaOH $(3 \times 20 \text{ mL})$ followed by brine $(1 \times 20 \text{ mL})$ mL). The organic phase was dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The product was purified by flash chromatography (petroleum ether/EtOAc 99:1), yielding 36 as a blue-green solid (22.3 mg, 82%); ¹H NMR (400 MHz, chloroform-*d*) δ 8.57 (dd, J = 9.8, 1.2 Hz, 2H), 8.15 (s, 1H), 7.69–7.66 (m, 4H), 7.60 (t, J = 9.8, 1.2 Hz, 1H), 7.56-7.50 (m, 4H), 7.40 (app ddt, J = 7.9, 6.9, 1.3 Hz, 2H), 7.14 (dd, J = 9.8, 9,8 Hz, 2H); ${}^{13}C{}^{1}H$ NMR (101 MHz, chloroform-d) δ 139.1, 137.4, 137.3, 136.8, 136.3, 130.7, 130.0, 128.8, 126.6, 123.6. Analysis data are in accordance with published data for this compound.¹⁶¹

(7aS,11aR)-4-Isopropyl-2-methyl-7,7a,8,10,11,11a-hexahydro-9H-naphtho[1,2,3-cd]azulen-9-one (37). In a 2 mL microwave vial equipped with a stir bar, compound 35 (9.3 mg, 0.032 mmol) was dissolved in THF (1 mL). The vial was capped and cooled in an ice bath. *t*-BuOK was added as a 1 M solution in THF (100 μ L, 0.100 mmol). The solution was left to stir at 0 °C for 20 min, after which it was poured into 1 M aqueous HCl (10 mL) and extracted with EtOAc $(3 \times 10 \text{ mL})$. The combined organic extracts were dried over Na₂SO₄, filtered, and the solvent was removed under a stream of nitrogen. The product was purified by flash chromatography (petroleum ether/EtOAc 90:10), yielding 37 as a blue sticky oil (3.6 mg, 39%); ¹H NMR (400 MHz, chloroform-d) δ 8.09 (d, J = 1.8 Hz, 1H), 7.54 (s, 1H), 7.36 (dd, J = 10.5, 1.8 Hz, 1H), 6.77 (d, J = 10.5 Hz, 1H), 3.61 (app dt, J = 5.7, 4.5 Hz, 1H), 3.25 (dd, J = 16.8, 4.4 Hz, 1H), 3.04 (hept, J = 6.9 Hz, 1H), 2.91 (dd, J = 16.8, 6.6 Hz, 1H), 2.84-2.75 (m, 1H), 2.67 (s, 3H), 2.49-2.42 (m, 2H), 2.41-2.36 (m, 2H), 2.32 (dd, J = 14.3, 8.6 Hz, 1H), 2.25-2.18 (m, 1H), 1.35 (d, J = 6.9 Hz, 6H); ¹³C{¹H} NMR (101 MHz, chloroform-d) δ 212.3, 144.0, 139.3, 135.8, 134.2, 133.4, 133.3, 133.0, 125.8, 124.5, 122.7, 44.6, 38.7, 38.7, 38.3, 37.7, 34.7, 30.4, 24.90, 24.91, 12.9; FTIR-ATR *v*max/cm⁻¹ 1709 (C=O); HRMS (ESI+) *m*/*z*: [M + H]⁺ calcd for C₂₁H₂₅O 293.1905; found 293.1904.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.0c01412.

Spectroscopic data for all products; NMR assignments and stereochemical elucidation of **3**7; colorimetric response to TFA and UV–vis/fluorescence spectra for

selected compounds; and emission spectrum of lamp used for photolytic decomplexations (PDF)

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Notes

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