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1	MOTMOT: models of trait macroevolution on trees (an update)			
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6	Abstract			
7	1. The disparity in species' traits arises through variation in the tempo and mode of evolution			
8	over time and between lineages. Understanding these patterns is a core goal in evolutionary			
9	biology.			
10	2. Here we present the comprehensively updated R package MOTMOT: Models Of Trait			
11	Macroevolution On Trees that contains methods to fit and test models of continuous			
12	trait evolution on phylogenies of extant and extinct species.			
13	3. MOTMOT provides functions to investigate a range of evolutionary hypotheses, including			
14	flexible approaches to investigate heterogeneous rates and modes of evolution, models of trait			
15	change under interspecific competition, and patterns of trait change across significant			
16	evolutionary transitions such as mass extinctions. We introduce and test novel algorithms of			
17	heterogeneous tempo and mode of evolution that allow for phylogeny-wide shifts in			
18	evolution at specific times on a tree. We use these new MOTMOT functions to highlight an			
19	exceptionally high rate of mammalian body mass evolution for 10 million years following the			
20	Cretaceous-Palaeogene mass extinction.			
21	4. These methods provide biologists and palaeontologists with the tools to analyse continuous			
22	trait data on phylogenies, including large trees of up to thousands of species.			
23	Keywords: phylogenetic comparative methods, phylogenetics, maximum likelihood,			
24	macroevolution, R CRAN Contact: marknputtick@gmail.com			

25 **1. INTRODUCTION**

26 Phylogenies provide a framework on which we can understand macroevolutionary 27 trait change. For comparative studies, phylogenies are necessary to both account for 28 statistical non-independence of taxa (Felsenstein, 1985; Grafen, 1989), and to act as a 29 framework on which to model the tempo and mode of evolution (Simpson, 1944). 30 Researchers have developed numerous methods to model the tempo and mode of evolution, 31 particularly tree-transformation models for continuous traits based on Brownian motion (BM) 32 (Hansen, 1997; Pagel, 1997, 1999; Blomberg, Jr, & Ives, 2003; Harmon et al., 2010; 33 Eastman, Alfaro, Joyce, Hipp, & Harmon, 2011; Ingram, 2011; Venditti, Meade, & Pagel, 34 2011; Thomas & Freckleton, 2012). Thomas and Freckleton (2012) introduced the R package MOTMOT to estimate parameters for many phylogenetic comparative methods, alongside 35 36 novel approaches to analyse heterogeneous rates of continuous trait evolution on 37 phylogenies. Here we update MOTMOT to flexible hypothesis testing by including novel 38 methods and functions previously only available as stand-alone code, alongside all its 39 original functions.

40

41 **2 DESCRIPTION**

42 **2.1 Overview**

We summarise the models of trait evolution and other functions introduced to the new release of MOTMOT in Table 1. Many of the methods available in MOTMOT have been described in detail elsewhere (see references in Table 1), and we provide a MOTMOT vignette with R code. Below we describe and assess the performance of a novel method to detect temporal shifts in tempo and mode of trait evolution and use these methods to study morphological evolution in Mammaliaformes using data from Slater (2013). containing fossils, except the OU model in transformPhylo.MCMC. The
transformPhylo.ML OU model is suitable for use with non-ultrametric trees (Slater,
2014). Uncertainty in trait values (Silvestro, Kostikova, Litsios, Pearman, & Salamin, 2015)
is incorporated using the *meserr* argument that takes a vector of trait errors. Finally, users can
simulate data, including export of node states, for the majority of models using

Nearly all methods in the package are usable with non-ultrametric phylogenies

55 transformPhylo.sim to test model fit and adequacy.

56 **2.2 timeSlice and modeSlice models**

49

57 Many phylogenetic rate heterogeneous algorithms allow for rate variation on branches 58 and lineages. Some rate-heterogeneous methods test for the presence of rate variation through 59 time or among lineages but not specific rate changes for the whole tree at a certain time. The 60 new *timeSlice* algorithm in transformPhylo.ML models changes in Brownian rates at 61 certain times, similar to approaches introduced by Slater (2013). Previous models only allowed a single shift at a fixed time, these are extended as *timeSlice* allows users to set one 62 63 or more shift time(s) of rate change. When users supply no split time, *timeSlice* will search 64 multiple potential shift times and identify the time point with the highest likelihood using a 65 stepwise approach similar to the *medusa* and *traitMedusa* approaches (Alfaro et al., 2009; 66 Thomas & Freckleton, 2012). In the first iteration, the function tests all shift points, identifies 67 and fixes the shift point leading to the highest likelihood one-shift model, then searches for 68 the shift point leading to the highest likelihood two-shift model. The algorithm optimises the 69 rate scalars for each time bin in each model, and sequentially fixes the best fitting shift time 70 from each iteration for consequent searches. Finally, a comparison is made between BM, one 71 shift, and the user defined *nth* model using AIC_c.

Unless stated, we summarise the output of the *timeSlice* model using the stepwise
approach discussed above, but we note it is also possible to summarise outputs using a model

averaging approach. For each iteration (i.e, a model with *n* shifts; it is not possible to compare a *n* and n + 1 shift model in this way), the model averaging summarises the relative fit of all shift positions based on their Akaike weights; and returns the weighted average rates through time.

We have also incorporated the new *modeSlice* model in transformPhylo.ML. *modeSlice* incorporates and extends the methods of Slater (2013) by allowing for multiple shifts in various modes of evolution (BM, OU, EB, and Kappa) at different times in the phylogeny's history. *modeSlice* is flexible as users can input multiple rate shift times with different combinations of modes. Furthermore, time bins with a BM mode can optionally vary in rate compared to the background variance (*rate.var* argument), and users can include a rate scalar alongside EB modes.

85 2.3 Simulations with extant and fossil data

86 On extant trees, the power and accuracy of the *timeSlice* algorithm (Figure 1) 87 increases when shifts are more recent, in trees with more tips, and with larger differences in 88 rates (see Supporting Information). On ultrametric trees, the number of branches decreases 89 exponentially with age, so our results indicate *timeSlice* is more accurate with larger trees or 90 trees including fossils.

91 2.4 Simulations with fossil data

The addition of fossils increases the power and accuracy of parameter estimation
under *timeSlice*. We simulated data under the *timeSlice* model using the total-evidence
Mammaliaformes tree (211 taxa, 153 extant) from Slater (2013) with rate shifts (2x,3x,5x
background rate) at 233, 177, 122, 65, and 10 Ma. We repeated these simulations on an
extant-only, Mammalia tree.

97 Correct support for the *timeSlice* model over BM is high for the Mammaliaformes
98 tree: for rate shifts 3x background and above, with shifts at 122 Ma or younger, there is 95%

99 correct *timeSlice* support (Supplementary Figure S2). The higher power and accuracy of the 100 *timeSlice* model at 65 Ma compared to 10 Ma is likely a consequence of the lack of time for 101 trait variance to accrue or the absence of fossils in the 10 Ma–present bin.

102 Accuracy and precision for the *timeSlice* model is higher for the fossil 103 Mammaliaformes analyses compared to the extant-only analyses (Supplementary Figure S3-104 4). The median error of rate estimates across all simulations is 0.63 for total-evidence 105 analyses compared to 0.97 for the extant-only analyses. No *timeSlice* model produces an 106 estimate of zero rates for any time bin on the fossil Mammaliaformes tree, only on the extant-107 only phylogeny (median 3.35% of models). When rates increase following a shift, there is 108 strong support for the correct timeSlice model on the Mammaliaformes tree (median 66.1% 109 correct support across analyses, >95% for some shifts) (Figure S5).

110 As a test of potential erroneous modelling of *timeSlice* process, we compared the fit of 111 single process OU and EB models on the Mammaliaforms and Mammalia trees with 112 *timeSlice* simulated data. In the Mammaliaformes tree the *timeSlice* pattern of high early rates 113 results in increased support for an EB model compared to the Mammalia tree (Figure S5). 114 This bias towards EB model support over timeSlice for timeSlice generated data occurs as 115 both models describe a process in which high rates decrease through time. When simulations 116 have higher rate differences between high ancient rate and subsequent lower rate, *timeSlice* 117 models are more accurate.

This erroneous OU support on the extant tree (median 18.8% across all analyses) is
likely a consequence of the OU model lengthening recent and reducing ancient branch
lengths, mimicking simulated parameters (Cooper, Thomas, Venditti, Meade, & Freckleton,
2015). This bias is apparent on the extant-only tree as the root-to-tip distance is equal for all
taxa (Supplementary Figure S5).

2.5 Cretaceous-Palaeogene shifts in evolution

125	We analysed rates of morphological evolution in extinct and extant Mammaliaformes				
126	using data from Slater (2013) using timeSlice. Slater showed a high Cenozoic rate of body				
127	mass evolution in Mammaliaformes that resulted from an OU to BM shift in mode at the				
128	Cretaceous-Palaeogene boundary (K-Pg) 66 Ma. Here, we analyse these data using				
129	MOTMOT functions; after testing a number of hypothesis-based and exploratory models we				
130	find best relative support for an increase in mammalian body mass evolution in the 10 million				
131	years following the K-Pg mass extinction.				
132	We extracted the mean mammal body mass and error measurement from the Slater				
133	(2013) dataset, and then matched these data to the phylogeny using sortTraitData				
134	(Figure 2a).				
135 136 137 138 139 140 141 142 143	<pre>> data(mammals) > attach(mammals) > trait.phy <- sortTraitData(phy = mammal.phy, y = as.matrix(mammal.mass), data.name = c("mean", "sem"), log.trait = FALSE) > phy <- trait.phy\$phy > y <- as.matrix(trait.phy\$trait[, 1]) > errors <- as.numeric(trait.phy\$trait[, 2])</pre>				
144	With these body mass and error data data, we tested the relative fit of BM, OU, and				
145	Early Burst using transformPhylo. As with Slater's K-Pg Shift model, estimated				
146	Cenozoic rates are higher than Mesozoic rates, but <i>timeSlice</i> is not supported over BM as				
147	shown by the <i>ModelFit</i> output the function timeSliceSummary (Table 2; Figure 2b).				
148 149 150 151 152 153	<pre>> time.slice.66.model <- transformPhylo.ML(y = y, phy = phy, Model = "timeSlice", splitTime = 66, meserr = errors) > plot.timeSlice.ML(time.slice.66.model, phylo.plot = FALSE) [c("ModelFit", "Rates")] \$ModelFit</pre>				
154 155 156 157 158	[1] "BM" \$Rates lnL AIC AICc sigma.sq.1 anc.state.1 -466.62552164 937.25104327 937.30873558 0.09924604 4.28252379				

159 However, we found support for a low background Mesozoic rate that accelerated

160 between 66-56 Ma (9.5x background) before reducing to a lower rate (56-0 Ma, 1.34x)

```
161 (Figure 2b).
```

```
162
     > time.slice.66.model.multi <- transformPhylo.ML(y = y,
       phy = phy, model = "timeSlice", splitTime = c(66, 56),
163
164
       meserr = errors)
165
     > plot.timeSlice.ML(time.slice.66.model.multi,
       show.tip.label = FALSE, edge.col = "white", edge.width = 1,
166
167
       cex = 1.3)[c("ModelFit", "Rates")]
     [1] "split 1"
168
169
170
     $Rates
171
               lnL
                              AIC
                                           AICc
                                                   sigma.sq.1
172
     anc.state.1
                          rates1
173
     -460.0902328
                     930.1804656
                                    930.4731485
                                                     0.3589563
174
     4.2811433
                 0.1785373
175
                           rates3 time.split1
                                                  time.split2
            rates2
176
         1.6957191
                       0.2400284
                                     66.0000000
                                                   56.000000
177
178
           Here we fit a more naïve model that searches for all shifts in 1 Ma increments from 50
179
     million years after the root age to 20 million years before the present. The best relative fit of
```

180 these models as judged by AIC_c shows an ancient rate acceleration commencing 170 million

181 years ago. The two-shift model is not supported, and the single shift model has a poor relative182 fit (Figure 2c).

183

```
184 > time.slice.66.model_naive <- transformPhylo.ML(y = y,</pre>
```

```
185 phy = phy, model = "timeSlice", nSplits = 2,
```

```
186 boundaryAge = c(50, 20), meserr = errors, testAge = 1)
```

```
187 > model.averaged.out <-</pre>
```

```
188 plot.timeSlice.ML(time.slice.66.model naive, model.average =
```

189 TRUE)

We compared this *timeSlice* model to shifts in modes using the approach of Slater
(2013), now implemented in the *modeSlice* algorithm in transformPhylo.ML. The
'release and radiate' model (Mesozoic OU shifts to BM at K-Pg) has a superior relative fit
compared to the *timeSlice* models, but we find a superior fit for a *modeSlice* model with low
Mesozoic rates (OU model) that accelerated to a high post-K-Pg rate (BM, rate scalar =
3.73), before shifting back to OU at 56 Ma (Table 2). The relative support for these models is
shown in Figure 2d.

```
199
     > release.model <- transformPhylo.ML(y = y, phy = phy,</pre>
200
        model = "modeslice", mode.order = c("ou", "bm"),
201
        splitTime = 66, meserr = errors)
202
     > release.radiate.model <- transformPhylo.ML(y = y, phy = phy,</pre>
203
        Model = "modeslice", mode.order = c("ou", "bm"),
204
        splitTime = 66 , meserr = errors, rate.var = TRUE)
205
     > release.radiate.recapture.model <- transformPhylo.ML(y = y,</pre>
206
        Phy = phy, model = "modeslice",
207
        mode.order = c("ou", "bm", "ou"), splitTime = c(66, 56),
208
        meserr = errors, rate.var = TRUE)
209
     $MaximumLikelihood
210
      [1] -453.9791
211
     $brownianVariance
212
     [1] 0.1395728
213
     $root.state
214
      [1] 4.259928
215
     $mode.1.ou
216
                 alpha
                               LCI
                                            UCI
217
      [1,] 0.02158316 0.01138688 0.03497292
218
     $mode.2.bm
219
           BM.rate
                         LCI
                                    UCI
220
      [1,] 4.69979 1.667876 10.93732
221
     $mode.3.ou
222
                 alpha
                                LCI
                                             UCI
223
      [1,] 0.01343794 0.003040442 0.02588539
224
     $AIC
225
     [1] 917.9582
226
     $AICc
227
      [1] 918.2509
228
229
230
           A high post-K-Pg rate of body mass evolution is congruent with data from the fossil
231
     record (Alroy, 1999; Raia et al., 2013), and may represent an evolutionary 'release' following
```

232 the extinction of non-avian dinosaurs at the end of Cenozoic (Slater, 2013). Although body 233 size evolution may be a poor proxy for functional traits such as dental characteristics, the 234 high rates of evolution for mammals in the earliest Cenozoic may be indicative of clades 235 movements into high level niches (Slater et al., 2019). Our new modelling framework, that 236 allows flexibility in the estimation of the time, tempo, and mode of trait evolution, therefore 237 provides new insight into body size evolution in the Mammaliaformes. More generally, we 238 expect this approach to add potentially important nuance to our understanding of phenotypic 239 macroevolutionary trends.

240

241 ADDITIONAL INFORMATION

The package motmot is available on CRAN and can also be installed directly from GitHub (github.com/PuttickMacroevolution/motmot). There is a full explanation of all functions and arguments as part of the R documentation, and a comprehensive vignette of the package is available online (github.com/PuttickMacroevolution/motmot).

247

248 CONCLUSIONS

MOTMOT provides a range of functions to analyse continuous trait evolution, with the main extensions summarised here. More details can be found in the package documentation and vignette available on CRAN.

252

253 AUTHOR CONTRIBUTIONS

254 MNP and GHT conceived the ideas and designed methodology; code was written by MNP,

255 GHT, MC, and TI. MNP analysed the data; MNP led the writing of the manuscript. All

authors contributed critically to the drafts and gave final approval for publication.

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- 261 funding.
- 262

263 DATA AVAILABILITY

- 264 The R MOTMOT package is available on CRAN (https://cran.r-
- 265 project.org/web/packages/motmot/index.html) and the Mammaliaformes data can be accessed
- 266 directly from the package. The code to generate simulations and analyses Mammaliaformes
- body mass evolution is available at Figshare (doi: 10.6084/m9.figshare.11337050).

268

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- 337

Table 1. A summary of the new and main functions included in MOTMOT. Multiple

New models				
Model	Description	Function(s)		
timeSlice*	Identifies shifts in the tree-wide rate of evolution at discrete times. <i>timeSlice</i> takes user-supplied split times or the function searches and finds the highest- likelihood shift time(s). The function timeSliceSummary can summarise,	transformPhylo.ML timeSliceSummary		
modeSlice*	plot, and calculate model averaging for <i>timeSlice</i> outputs. Estimates models with shifts in the tree- wide mode of evolution at specified times. Modes can shift between BM, Early Burst (EB), OU, and Kappa models. The BM modes can optionally here different rates (manifed aging the	transformPhylo.ML		
	have different rates (specified using the <i>rate.var</i> argument), and a rate scalar can be set to the EB model.	tuon of our Dhulo MI		
nested modes*	Calculates parameters for a shift from a Brownian motion to a different evolutionary mode (λ , δ , κ , OU, ACDC, or ϕ) within a subclade of a phylogeny (Puttick, 2018).	CTANSIOIMPHYIO.ML		
character displacement	Simulation of data under BM and trait change under intra-specific competition (Clarke, Thomas, & Freckleton, 2017).	chr.disp.param chr.disp.lrt		
Bayesian estimation of parameters*	Bayesian MCMC estimation of λ , δ , κ , OU, ACDC, or ϕ models.	transformPhylo.MCMC mcmc.plot		
φ and multi-φ*	Estimation of the relative contributions of separational and gradual evolution to trait evolution; fit as a whole-tree process (ϕ) or with different values estimated in subclades (multi- ϕ) (Ingram, 2011; Ingram et al., 2016).	transformPhylo.ML traformPhylo.MCMC		
Pagel's λ*	Measure of phylogenetic signal, can be estimated simultaneously alongside δ , κ , OU, ACDC, and ϕ models (Pagel, 1997, 1999).	transformPhylo.ML transformPhylo.MCMC		
Phylogenetic Generalised Least Squares (PGLS)	Phylogenetic regression model with continuous traits estimated using contrasts, faster than using variance- covariance matrices (Felsenstein, 1973, 1985; Grafen, 1989; Freckleton, 2012).	pic.pgls		
Acceleration-	Exponential change in evolutionary rate	transformPhylo.ML		

traits can be analysed in models marked by an asterisk (*). 339

Deceleration	through time. If the upperBound	transformPhylo.MCMC
(ACDC) /	argument is set to zero, ACDC becomes	
Early Burst*	the Early Burst model (Blomberg et al.,	
	2003; Harmon et al., 2010)	
trend*	Time-dependent change in character	transformPhylo.ML
	values; only applicable for non-	
	ultrametric trees (Pagel, 2002)	
utility functions	Functions to facilitate analyses of trait	contemporaryPhy
	selectivity of mass extinction in the fossil	addFossilToPhy
	record (Puttick et al., 2017; Allen,	
	Stubbs, Benton, & Puttick, 2018). Also	
	functions to add fossils to ultrametric	
	phylogenies.	

Table 2. Modes of evolution fit to Mammaliaformes body mass evolution used to investigate
a shift in evolution at the Cretaceous-Palaeogene boundary 66 Ma. The *modeSlice* model
with best relative fit is shown in **bold**. The table summarises the MOTMOT function used for
each analysis, the estimated parameters with 95% confidence intervals in brackets, *n*

346 parameters, and AIC_c, and AIC_c weights.

Model	MOTMOT function	estimated parameters	n	AICc	AICc Weights
Brownian motion	<pre>transformPhylo.ML(y = y, phy = phy, model = "BM")</pre>	σ ² 0.0992 μ 4.2825	2	937.3087	6.81e-05
Ornstein- Uhlenbeck	transformPhylo.ML(y = y, phy = phy, model = "OU")	σ2 0.1022 μ 4.4433 α 5.976e-4 (1e-8, 0.006)	3	939.3174	2.50e-05
Early Burst	transformPhylo.ML(y = y, phy = phy, model = "OU", upperBound = -1e-6)	σ ² 0.0967 μ 4.2825 <i>a</i> -1e-06 (-0.002, -1e-06)	3	938.3214	4.11e-05
split at K- Pg (66 Ma)	transformPhylo.ML(y = y, phy = phy, model = "timeSlice", splitTime = 66)	$\sigma^2 0.2938$ $\mu 4.2823$ Pre-Kg rate: 1 (0.611, 1.679) Post-K-Pg rate: 1.33 (0.776, 2.23)	4	937.3087	6.81e-05
<i>timeSlice</i> with split at K-Pg (66 Ma) and 56 Ma	transformPhylo.ML(y = y, phy = phy, model = "timeSlice", splitTime = c(66, 56))	$\sigma^2 0.3590$ $\mu 4.2811$ Pre-Kg rate: 1 (0.650, 1.59) 66-53 Ma rate: 9.50 (3.59, 21.53) 53-0 Ma rate: 1.34 (0.859, 2.09)	5	930.4731	2.08e-03
<i>timeSlice</i> naïve search for two best- fitting shifts between 215-20 myrs in 1 myr increment.	time.slice.66.model_naive <- transformPhylo.ML(y = y, phy. = phy, model = "timeSlice", nSplits = 2, boundaryAge = c(50, 20), meserr = errors, testAge = 1)	One split: 171 Ma σ^2 0.2979 μ 4.3561 Pre-171 Ma rate: 1 (0.413, 2.677) 171-0 Ma rate: 4.8589 (2.1035, 11.4792)	4	930.6699	1.88e-03
'Release' model OU to BM shift at K-	transformPhylo.ML(y = y, phy = phy, model = "modeslice", mode.order = c("ou", "bm"), splitTime = 66)	$\sigma^2 0. 1145$ $\mu 4.4345$ $\alpha 0.015 (0.007, 0.0258)$	3	925.7641	0.0219

Pg					
'Release and Radiate' (Slater model) OU to BM plus rate shift at K- Pg	transformPhylo.ML(y = y, phy = phy, model = "modeslice", mode.order = c("ou", "bm"), splitTime = c(66), rate.var = TRUE)	σ2 0.2146 μ 4.2652 α 0.0291 (0.0173, 0.0449) BM rate: 0.4629 (0.2785, 0.7607)	4	924.7107	0.0371
<i>modeSlice</i> pre-K-Pg OU 66-56 Ma BM with rate shift 56-0 Ma OU	transformPhylo.ML(y = y, phy = phy, model = "modeslice", mode.order = c("ou","bm", "ou"), splitTime = c(66, 56), rate.var = TRUE)	σ ² 0.140 μ 4.2599 OU (root-66 Ma): α 0.0216 (0.0114, 0.0350) BM rate (66-56 Ma): 4.670 (1.6679, 10.9373) α 0.0134 (0.0030, 0.0259)	5	918.2509	0.937

Figure 1. Relative support for the *timeSlice* model (light blue) and BM (dark blue) from fully simulated data. Data were generated under BM or with one shift to a rate higher or lower rate (scalar=2,3,5x background) and at various ages (0.1, 0.25, 0.5, 0.75, 0.9). 1000 replicates were generated on trees with 50, 100, and 500 tips with 1000 replicates. Model power of *timeSlice* increases with larger shift magnitudes, tree size, and when shifts are closer to the present.

355

356 Figure 2. Analysis of mammal body mass evolution during the pass 250 million years.

357 The output from sortTraitData showing the relative body mass for tips on the

358 phylogeny (a). The estimated rates of evolution for shifts in body mass evolution at 66 and 53

359 Ma from the *timeSlice* model in transformPhylo.ML with branches scaled to rates and

360 these same rate estimates and associated CIs plotted through time using

361 timeSliceSummary (b). The model-average rate estimates from a naïve *timeSlice* search

362 with two shifts identified as the best-fitting points from million-year increments between

363 215-20 Ma (the best fit relative fit supports a single shift model at 171 Ma) plotted using

364 timeSliceSummary (c). The Akaike weights for various *timeSlice* and *modeSlice* models

365 (d), showing the overwhelming support for the model of OU to 66 Ma followed by a BM

366 with a rate increase with a subsequent shift to a lower rate in another OU model. Full details

367 of each model are shown in Table 2.