

2006

Synthesis, characterisation and application of inherently conducting polymer nanoparticles

Orawan Ngamna
University of Wollongong

Follow this and additional works at: <https://ro.uow.edu.au/theses>

University of Wollongong

Copyright Warning

You may print or download ONE copy of this document for the purpose of your own research or study. The University does not authorise you to copy, communicate or otherwise make available electronically to any other person any copyright material contained on this site.

You are reminded of the following: This work is copyright. Apart from any use permitted under the Copyright Act 1968, no part of this work may be reproduced by any process, nor may any other exclusive right be exercised, without the permission of the author. Copyright owners are entitled to take legal action against persons who infringe their copyright. A reproduction of material that is protected by copyright may be a copyright infringement. A court may impose penalties and award damages in relation to offences and infringements relating to copyright material.

Higher penalties may apply, and higher damages may be awarded, for offences and infringements involving the conversion of material into digital or electronic form.

Unless otherwise indicated, the views expressed in this thesis are those of the author and do not necessarily represent the views of the University of Wollongong.

Recommended Citation

Ngamna, Orawan, Synthesis, characterisation and application of inherently conducting polymer nanoparticles, PhD thesis, Intelligent Polymer Research Institute, Department of Chemistry, University of Wollongong, 2006. <http://ro.uow.edu.au/theses/767>

Research Online is the open access institutional repository for the University of Wollongong. For further information contact the UOW Library: research-pubs@uow.edu.au

NOTE

This online version of the thesis may have different page formatting and pagination from the paper copy held in the University of Wollongong Library.

UNIVERSITY OF WOLLONGONG

COPYRIGHT WARNING

You may print or download ONE copy of this document for the purpose of your own research or study. The University does not authorise you to copy, communicate or otherwise make available electronically to any other person any copyright material contained on this site. You are reminded of the following:

Copyright owners are entitled to take legal action against persons who infringe their copyright. A reproduction of material that is protected by copyright may be a copyright infringement. A court may impose penalties and award damages in relation to offences and infringements relating to copyright material. Higher penalties may apply, and higher damages may be awarded, for offences and infringements involving the conversion of material into digital or electronic form.

**SYNTHESIS, CHARACTERISATION AND
APPLICATION OF INHERENTLY CONDUCTING
POLYMER NANOPARTICLES**

A thesis submitted in fulfilment of the requirements for the award of the degree

DOCTOR OF PHILOSOPHY

from the

UNIVERSITY OF WOLLONGONG

by

ORAWAN NGAMNA, B.Sc.(Food Technology)

INTELLIGENT POLYMER RESEARCH INSTITUTE

DEPARTMENT OF CHEMISTRY

February 2006

This work is dedicated to my dad and mom, Mr. Surin and Mrs Pratin Ngamna for their unconditional love and support.

CERTIFICATION

I, Orawan Ngamna, declare that this thesis, submitted in fulfilment of the requirements for the award of Doctor of Philosophy, in the Department of Chemistry, University of Wollongong, is wholly my own work unless otherwise referenced or acknowledged. The document has not been submitted for qualifications at any other academic institution.

Orawan Ngamna

February 2006

ACKNOWLEDGEMENTS

Firstly I would like to thank my family: dad, mom, P’Nong and Ekkapol for giving me life as well as unconditional love and support.

Thanks also to Unilever Thai Holdings Ltd. for financial support during my Master’s degree. P’Noo (PYT) and Rene (RL) who encouraged and supported me to undertake further study and to pursue a PhD as well as P’Tee, P’Pensri, K. Adisai (AMV) and all friends in Unilever are really appreciated.

My life in IPRI would not be possible without the great help and support from both of my supervisors- Prof. Gordon G. Wallace and Dr. Simon E. Moulton. I appreciate the time that Simon has given to my work and thesis, and also his encouragements throughout my time in IPRI. Gordon is an absolutely exceptional professor who cares not only about the research outcome, but also for the well-being of every one under his reign, and is always understanding and helpful. I have never felt left behind or unappreciated whilst working in his ‘Evil Empire’. Thanks to Peter (and Uncle Pete’s magic toys), Dr. Chee, Violeta (for the nice TEM and AFM images) and Elvis, who always has something to do for other people in IPRI. Thanks to all those involved in collaborative works: Siree, Aoife and Jenny. Thanks to P’Yingpit, Maria, Fatemeh, Vahid, Brianna, Carol, Kerry, Aoife, Su Ryon, Changkee and Eve for all of their encouragements and for always making my work in the lab and my personal life fruitful. My gratitude to Leon, Bjørn, Chee, Peter, Toni and George for comments/proof reading my final thesis chapters. Last but not least, I would like to thank Ngae for being such an unbelievable friend throughout both good and tough times and always be there for me.

Finally, I would like to thank Prof. Gordon Wallace again for giving me the IPRI scholarship throughout my PhD studies.

PUBLICATIONS

1. S.E. Moulton*, P.C. Innis, L.A.P. Kane-Maguire, O. Ngamna, and G.G. Wallace, “Polymerisation and characterisation of conducting polyaniline nanoparticle dispersions”, *Current Applied Physics*, 2004, **4** (2-4), 402-406.
2. A. Morrin, O. Ngamna, A.J. Killard, S.E. Moulton, M.R. Smyth*, and G.G. Wallace*, “An Amperometric Enzyme Biosensor Fabricated from Polyaniline Nanoparticles”, *Electroanalysis*, 2005, **17** (5-6), 423-430.
3. O. Ngamna, A. Morrin, S.E. Moulton, A.J. Killard, M.R. Smyth, and G.G. Wallace*, “An HRP based biosensor using sulphonated polyaniline”, *Synthetic Metals*, 2005, **153** (1-3), 185-188.
4. A. Morrin, F. Wilbeer, O. Ngamna, S.E. Moulton, A.J. Killard, G.G. Wallace, and M.R. Smyth*, “Novel biosensor fabrication methodology based on processable conducting polyaniline nanoparticles”, *Electrochemistry Communications*, 2005, **7** (3), 317-322.
5. J.M. Pringle*, O. Ngamna, J. Chen, G.G. Wallace, M. Forsyth and D.R. MacFarlane, “Conducting Polymer Nanoparticles Synthesized in an Ionic Liquid by Chemical Polymerisation”, *Synthetic Metals*, in press.

6. O. Ngamna, S.E. Moulton and G.G. Wallace, “Incorporation of Dye into Conducting Polyaniline Nanoparticles”, *Polymer*, accepted.

TABLE OF CONTENTS

CERTIFICATION	I
ACKNOWLEDGEMENTS	II
PUBLICATIONS	III
TABLE OF CONTENTS	V
LIST OF ABBREVIATIONS	XVI
ABSTRACT	XXI

PART I - General Introduction and Characterisation Techniques

Chapter 1 – General Introduction

1.1 Inherently Conducting Polymer (ICPs)	1
1.1.1 Polyaniline (PANI)	3
1.1.2 Poly (2-methoxyaniline-5-sulphonic acid)	7
1.1.3 Polypyrrole	10
1.1.4 Polythiophene	13
1.2 Conducting Polymer Nanoparticles	16
1.2.1 Emulsion polymerisation	18
1.2.2 Use of steric stabilisers	20
1.3 Polymer-modified Amperometric Biosensors	21
1.3.1 Horseradish peroxidase and hydrogen peroxide system	22
1.3.2 Fabrication methods	24
1.3.2.1 Electrodeposition	24
1.3.2.2 Evaporative casting	25

1.3.2.3	Ink-jet printing	25
1.4	Structure of the Thesis and the Aim of the Chapter	26
1.5	References	29
Chapter 2 – Instrumental and Characterisation Techniques		
2.1.	Spectroscopy	37
2.1.1.	Ultraviolet-visible (UV-visible) spectroscopy	37
2.1.2.	Raman spectroscopy	37
2.2	Microscopic and Light Scattering Methods	39
2.2.1	Transmission electron microscopy (TEM)	39
2.2.2	Atomic force microscopy (AFM)	41
2.2.3	Light scattering	44
2.3	Electrochemical Techniques	48
2.3.1	Open circuit potential (OCP)	48
2.3.2	Cyclic voltammetry (CV)	49
2.3.3	DC amperometry	51
2.4	Conductivity Measurement	53
2.5	Contact Angle Measurement	54
2.6	References	57

PART II – Synthesis and Characterisation of Conducting Polymer

Nanoparticles

Chapter 3 – Synthesis and Characterisation of Polyaniline

Nanoparticles

3.1	Introduction	60
3.2	Experimental	63
3.2.1	Chemicals	63
3.2.2	Equipment/instrumentation	63
3.2.3	Methods	64
3.2.3.1	Polymerisation and purification	64
3.2.3.2	Characterisation of PANI nanoparticles and dispersions	65
3.2.3.3	Stability test of <i>nano</i> PANI-DBSA	65
3.3	Results and Discussion	66
3.3.1	<i>nano</i> PANI-DBSA	66
3.3.1.1	Polymerisation	66
3.3.1.1.1	Open circuit potential (OCP) measurement	67
3.3.1.1.2	UV-visible spectroscopy during polymerisation	68
3.3.1.1.3	Particle size measurement during polymerisation by DLS	69
3.3.1.2	Characterisation of nanoparticles and dispersions	71
3.3.1.2.1	Transmission electron microscopy (TEM), mass of product and conductivity	71
3.3.1.2.2	pH and redox switching	73

3.3.1.2.3	Raman Spectroscopy	76
3.3.1.2.4	Cyclic voltammetry (CV)	77
3.3.1.2.5	Stability test	79
3.3.1.3	Further processing and characterization	82
3.3.1.3.1	Preparation method for the sPANI/DBSA	83
3.3.1.3.2	UV-visible spectroscopy	83
3.3.1.3.3	Cyclic Voltammetry (CV)	83
3.3.1.3.4	Particle size and morphology	85
3.3.2	PANI-DBSA-RM	87
3.3.2.1	Polymerisation	87
3.3.2.2	Characterisations of PANI-DBSA-RM nanodispersions	89
3.3.2.2.1	UV-visible spectroscopy and redox switching	89
3.3.2.2.2	Raman spectroscopy	91
3.3.2.2.3	Cyclic Voltammetry (CV)	92
3.3.2.2.4	Conductivity, particle size and morphology	93
3.4	Conclusions	96
3.5	References	98

Chapter 4 – Incorporation of Dye into Polyaniline Nanoparticles

4.1	Introduction	100
4.2	Experimental	101
4.2.1	Chemicals	101
4.2.2	Equipment/instrumentation	101
4.2.3	Methods	102

4.2.3.1	Dye purification	102
4.2.3.2	Polymerisation and purification	102
4.2.3.3	Characterisation of nanoparticles and dispersions	103
4.3	Results and Discussion	104
4.3.1	Polymerisation	104
4.3.2	Characterisations of nanoparticles and dispersions	109
4.3.2.1	Transmission electron microscopy (TEM), conductivity and mass of product	109
4.3.2.2	pH switching	111
4.3.2.3	Redox switching	113
4.3.2.4	Raman spectroscopy	116
4.3.2.5	Cyclic voltammetry (CV)	120
4.3.2.6	Stability test	122
4.4	Conclusions	124
4.5	References	125
 Chapter 5 – Synthesis and Characterisation of Polypyrrole Nanoparticles		
5.1	Introduction	127
5.2	Experimental	128
5.2.1	Chemicals	128
5.2.2	Equipment/instrumentation	128
5.2.3	Methods	129
5.2.3.1	Polymerisation and purification	129

5.2.3.2	Characterisation of dispersions	130
5.2.3.3	Stability test of PPy-DS-PVA nanodispersions	130
5.3	Results and Discussion	131
5.3.1	PPy-DS polymerisation	131
5.3.2	PPy-DS Characterisation	132
5.3.2.1	UV-visible spectroscopy	132
5.3.2.2	Raman spectroscopy	133
5.3.2.3	Cyclic voltammetry (CV)	134
5.3.2.4	Mass of product, conductivity, particle size and morphology	135
5.3.3	PPy-DS polymerisation in the presence of PVA	136
5.3.3.1	Effect of PVA concentration on nanoparticle formation	136
5.3.3.2	Effect of polymerisation temperature on nanoparticle formation	138
5.3.3.3	Effect of SDS concentration on nanoparticle formation	140
5.3.3.4	Effect of monomer concentration on nanoparticle formation	142
5.3.4	Characterisation of PPy-DS-PVA nanodispersions	144
5.3.4.1	UV-visible spectroscopy	144
5.3.4.2	Raman spectroscopy	144
5.3.4.3	Cyclic voltammetry (CV)	146
5.3.4.4	Mass of product, conductivity, particle size and morphology	146

5.4	Conclusions	149
5.5	References	150

Chapter 6 – Synthesis and Characterisation of Polyterthiophene Nanoparticles

6.1	Introduction	152
6.2	Experimental	154
6.2.1	Chemicals	154
6.2.2	Equipment/instrumentation	155
6.2.3	Methods	156
6.2.3.1	Electrochemical synthesis of PTTh-DBSA	156
6.2.3.2	Chemical synthesis of PTTh-DBSA nanoparticles	156
6.2.3.3	Chemical synthesis of PTTh nanoparticles in IL	156
6.2.3.4	Characterisation of PTTh nanoparticles	157
6.3	Results and Discussion	157
6.3.1	PTTh-DBSA	157
6.3.1.1	Electrochemical synthesis of PTTh-DBSA	157
6.3.1.2	Chemical synthesis of PTTh-DBSA nanoparticles	162
6.3.1.2.1	Effect of oxidant concentration	162
6.3.1.2.2	Effect of surfactant concentration	168
6.3.1.2.3	Electroactivity study	172
6.3.1.2.4	Colloidal stability	175
6.3.1.2.5	Size and morphology study by TEM	176
6.3.2	PTTh nanoparticles synthesised in IL	176

6.3.2.1	Chemical synthesis of PTh nanoparticles in emiTfSA	176
6.3.2.1.1	Effect of oxidant concentration	176
6.3.2.1.2	Effect of solvent	179
6.3.2.1.3	Electroactivity study	181
6.3.2.1.4	Colloidal stability	184
6.3.2.1.5	Size and morphology study by TEM	184
6.4	Conclusions	185
6.5	References	186

PART III – Applications of Conducting Polymer Nanodispersions

Chapter 7 – Polyaniline-Dedecylbenzene Sulphonic acid-

Horseradish Peroxidase Biosensors

7.1	Introduction	188
7.2	Experimental	189
7.2.1	Chemicals	189
7.2.2	Equipment/instrumentation	190
7.2.3	Methods	190
7.2.3.1	Preparation of <i>nano</i> PANI-DBSA modified electrodes	190
7.2.3.2	Characterisations of the polymer modified electrodes	192
7.2.3.3	Colorimetric enzyme assay	192
7.3	Results and Discussion	193
7.3.1	Preparation of <i>nano</i> PANI-DBSA films onto glassy carbon electrodes	193

7.3.2	Characterisations of electrodeposited <i>nano</i> PANI-DBSA films	196
7.3.3	Optimisation of the mass of HRP on the electrode surface	200
7.3.4	Optimisation of electrodeposition	204
7.3.5	Colorimetric enzyme assay	205
7.4	Conclusions	206
7.5	References	207
Chapter 8 – Poly(2-methoxyaniline-5-sulphonic acid)- Horseradish Peroxidase-Poly(L-lysine) Biosensors		
8.1	Introduction	208
8.2	Experimental	210
8.2.1	Chemicals	210
8.2.2	Equipment/instrumentation	211
8.2.3	Methods	211
8.2.3.1	Preparation of polymer modified electrodes	211
8.2.3.2	Characterisation of polymer modified electrodes	212
8.3	Results and Discussion	213
8.3.1	Complexation and fabrication methods	213
8.3.2	Optimisation of PMAS concentration	215
8.3.3	Optimisation of PLL concentration	216
8.3.4	Optimisation of the mass of HRP on the electrode surface	219
8.3.5	Characterisations of the PMAS-HRP-PLL sensors	221
8.3.5.1	Sensor response to various applied potentials and buffer pH	221

8.3.5.2	Linearity and sensitivity	223
8.3.5.3	Interferent test	224
8.3.5.4	Long-term stability test	225
8.4	Conclusions	226
8.5	References	227

Chapter 9 – Inherently Conducting Polymer Nanodispersion

Ink-jet Printed Biosensors

9.1	Introduction	228
9.2	Experimental	232
9.2.1	Chemicals	232
9.2.2	Substrate	233
9.2.3	Equipment/instrumentation	233
9.2.4	Methods	233
9.2.4.1	Viscosity measurements and pH adjustment of the ICPs nanoparticles	233
9.2.4.2	Printing method for Epson C45 printer	234
9.2.4.3	Film preparation	235
9.2.4.4	Real-time monitoring of enzyme/substrate interactions in a batch cell	235
9.3	Results and Discussion	235
9.3.1	Instrumentation	235
9.3.2	Substrate characterisation	236
9.3.2.1	Electrochemistry	236

9.3.2.2	Hydrophilicity of substrates	238
9.3.3	Properties of ICP nanoparticles	239
9.3.3.1	Particle size in concentrated solution	239
9.3.3.2	Wetting ability and viscosity	242
9.3.4	Ink-jet printing of ICP nanoparticles	243
9.3.4.1	Printing the PPy-DS-PVA nanodispersions	243
9.3.4.2	Printing the PMAS and PLL solutions	244
9.3.4.3	Printing the sPANI-DBSA	251
9.3.4.4	Printing the PANI-DBSA-RM nanodispersions	254
9.4	Conclusions	258
9.5	References	260
	PART IV – Conclusions and Future Studies	261

ABBREVIATIONS

μ	micro
μ	viscosity
λ	wavelength
k_B	Boltzman's constant
γ	surface tension
Ω	Ohm
ν	frequency
$^{\circ}\text{C}$	degree Celsius
Θ	angle
η	refractive index
σ	conductivity
ρ	resistivity
ν	scan rate
A	ampere
\AA	angstrom
A^-	anion
A_i	activity of species i
ABTS	2,2'-azino-bis(3-ethylbenzthiazoline-6-sulphonic acid)
AFM	atomic force microscopy
Ag/AgCl	silver/silver chloride reference electrode
Ag/Ag ⁺	silver/silver ion reference electrode
APS	ammonium peroxydisulphate

BSA	bovine serum albumin
BFEE	boron-fluoride ethyl ether
C	Cunningham slip correction factor
ca.	approximately
CB	Carbolan Blue
CCD	charge-coupled device
cm	centrimeter
cm ⁻¹	wave number
CMC	critical micelle concentration
CV	cyclic voltammetry
d	diameter
Da	dalton
D _{AB}	diffusion coefficient
DBSA	dodecylbenzene sulphonic acid
DLS	dynamic light scattering
E	potential
E ⁰	standard reduction potential
EB	emeraldine base
emiTFSA	1-ethyl-3-methylimidazolium- bis(trifluoromethane-sulfonyl)amide
ES	emeraldine salt
F	Faraday constant
g	gram
GC	glassy carbon

HCSA	camphorsulphonic acid
HPLC	high performance liquid chromatography
HRP	horseradish peroxidase
h	hour
ICP	inherently conducting polymer
IL	ionic liquid
ITO	indium tin oxide
k	thousand
L	litre
LB	leucoemeraldine base
m	metre
M	molar
MAS	2-methoxyaniline-5-sulphonic acid
mg	milligram
min	minute
ml	millilitre
mM	millimolar
MoAb	monoclonal antibody
mS	millisiemen
Mw	molecular weight
nm	nanometre
OCP	open circuit potential
PANI	polyaniline
PB	perniganiline base

PBS	phosphate buffer saline
PEI	poly(ethyleneimine)
PLL	poly(L-lysine)
PMAS	poly(2-methoxyaniline-5-sulphonic acid)
PPy	polypyrrole
PS	perniganiline salt
PSS	poly(sodium 4-styrene sulfonate)
Pt	platinum
PTh	polythiophene
PTTh	polyterthiophene
PVA	poly(vinyl alcohol)
PVP	poly(4-vinylpyridine)
PVS	polyvinylsulphonate
Py	pyrrole
r	radius
RM	rapid mixing
RMS	root mean square
rpm	round per minute
S	siemen
SCE	saturated calomel electrode
SDS	sodium dodecylsulfate
SEM	scanning electron microscopy
SHE	standard hydrogen electrode
SPM	scanning probe microscopy

T	temperature
TBAP	tetrabutylammonium perchlorate
TEM	transmission electron microscopy
Th	thiophene
TTh	terthiophene
V	volt
vs.	versus
w/v	weight by volume
w/w	weight by weight

ABSTRACT

Synthesis of inherently conducting polymers (ICPs) nanoparticles is an option to improve the processability and conductivity of ICPs. In this thesis, the synthesis and application of ICPs nanoparticles has been demonstrated. Various polymerisation methods, such as emulsion polymerisation, use of steric stabiliser and synthesis in ionic liquid (IL) media, have been used to synthesise polymer nanoparticles. These synthesis methods render the ICPs nanoparticles stable as dispersions which are more processable and contain peculiar and fascinating properties superior to their bulk counterparts. These nanoparticles are further applied as mediators for biosensors. They have been fabricated into sensors using electrodeposition, evaporative casting, or ink-jet printing methods. Electrodeposition method results in formation of ultra thin nanostructured polymeric films that enhance sensor performance. Evaporative casting method is an easy one-step method, but precision is hard to achieve and dense films with rough morphology are formed. Ink-jet printing can be used to produce precise and accurate patterns and also this approach is amenable to mass production.

Polyaniline (PANI) nanoparticles; *nano*PANI-dodecylbenzene sulphonic acid (DBSA) have been synthesised using emulsion polymerisation (Chapter 3). The *nano*PANI-DBSA obtained has a conductivity of 34 ± 7 S/cm with particle size in the range of 10 ± 2 nm. The *nano*PANI-DBSA has been used as a mediator layer in biosensor applications as demonstrated in Chapter 7. These nanoparticles were fabricated onto the conductive electrode using an electrodeposition method with subsequent immobilisation of the enzyme horseradish peroxidase (HRP). Sensor performance was examined using

amperometric method and HRP/hydrogen peroxide (H_2O_2) configuration as a model system. The nanodomain of the *nano*PANI-DBSA particles contributed to highly ordered nanostructure patterning on the electrode surface. This uniform surface showed improved enzyme deposition characteristics, a lower background signal and better sensor performance at a lower HRP loading when compared to the sensors fabricated from electropolymerisation of the bulk monomer.

*Nano*PANI-DBSA particles aggregate at high concentrations; hence they are not amenable to ink-jet printing. *s*PANI-DBSA was prepared from centrifugation of the *nano*PANI-DBSA dispersions and used as a material for ink-jet printing. HRP was pre-mixed with the *s*PANI-DBSA nanodispersions before fabrication onto ITO-coated mylar using ink-jet printing. The print quality from the *s*PANI-DBSA nanodispersions was inconsistent and the catalytic signal of this biosensor was very low. These resulted in no further ink-jet printing work for this material.

The PANI-DBSA-rapid mixing (RM) nanodispersions were synthesised using a RM method. These dispersions contained nanometre size PANI particles dispersed in aqueous media. These nanoparticles have been successfully printed using ink-jet printing as outline in Chapter 9. This work has demonstrated the ink-jet printability of conducting polymer nanoparticles and their use as working electrodes for biosensors. The sensor response from these ink-jet printed PANI-DBSA-RM was higher than the sensor response from evaporative casting of poly(2-methoxyaniline-5-sulphonic acid) (PMAS) in Chapter 8.

The addition of functional group into PANI nanoparticles was also investigated. Carbolan Blue (CB) dye was incorporated into the PANI backbone using emulsion polymerisation method as demonstrated in Chapter 4. The dye was proved to have strong interaction with PANI backbone using Raman spectroscopy and centrifugation test. The

distinct solution colour after a reduction process could lead the PANI-DBSA-CB to be a potential candidate of the material for electrochromic devices.

Synthesis of polypyrrole (PPy) nanoparticles is demonstrated in Chapter 5. Poly(vinyl alcohol) (PVA) was used as the steric stabiliser to produce PPy-DS-PVA nanoparticles. These nanoparticles were well dispersed in water with particle size in the order of 52 ± 5 nm. Aggregation was obvious in concentrated solutions and led to poor ink-jet printed quality of the PPy-DS-PVA nanoparticles.

The water soluble polymer, PMAS, was also used to fabricate biosensors using evaporative casting method in Chapter 8 and ink-jet printing in Chapter 9. In chapter 8, its solubility enabled PMAS to pre-mix with the HRP enzyme prior to complexing with the polycations poly(L-lysine) hydrochloride (PLL) and subsequently casting onto ITO coated mylar substrate. This biosensor format has proven ability to easily fabricate the conducting polymer nanoparticles by one-step evaporative casting. The optimised sensors exhibited good sensor response, high selectivity and very good long-term stability. The ink-jet printed films from PMAS and PLL solutions (Chapter 9) showed better electroactivity compared to the evaporative cast films which could lead to better sensor performance. However, the problem of PLL blocking the print head resulted in the discontinuation of its use.

The polyterthiophene (PTTh) aqueous dispersed nanoparticles were also successfully synthesised in the presence of surfactant (DBSA) and in ionic liquid; 1-ethyl-3-methylimidazolium bis(trifluoromethane-sulfonyl)amide (emiTFSA) as demonstrated in Chapter 6. The dispersion of PTTh-DBSA nanoparticles has shown poor colloidal stability and poor electroactivity. Although the PTTh nanoparticles synthesised in emiTFSA needed

2-3 minutes sonication to be dispersed in water, they have shown good electrochemistry and being test in another study in our laboratories for its use in photovoltaic devices.

These processable ICPs nanoparticles are promising materials for biosensor applications, electrochromic devices and solar cells. Assembly of these nanoparticles on to conductive substrates leads to highly ordered nanostructured ICPs on the surface and improves the biosensor performances. Also these nanoparticles prove their ability to be processable in mass production scale.