University of Wollongong

Research Online

Faculty of Science, Medicine and Health - Papers: part A

Faculty of Science, Medicine and Health

1-1-2014

Investigation of genetic variants in ubiquitin enzyme genes involved in the modulation of neurodevelopmental processes: a role in schizophrenia susceptibility?

Jessica L. Andrews University of Wollongong, ja393@uowmail.edu.au

Francesca Fernandez-Enright University of Wollongong, fernande@uow.edu.au

Follow this and additional works at: https://ro.uow.edu.au/smhpapers

Part of the Medicine and Health Sciences Commons, and the Social and Behavioral Sciences Commons

Recommended Citation

Andrews, Jessica L. and Fernandez-Enright, Francesca, "Investigation of genetic variants in ubiquitin enzyme genes involved in the modulation of neurodevelopmental processes: a role in schizophrenia susceptibility?" (2014). *Faculty of Science, Medicine and Health - Papers: part A*. 2457. https://ro.uow.edu.au/smhpapers/2457

Research Online is the open access institutional repository for the University of Wollongong. For further information contact the UOW Library: research-pubs@uow.edu.au

Investigation of genetic variants in ubiquitin enzyme genes involved in the modulation of neurodevelopmental processes: a role in schizophrenia susceptibility?

Abstract

Despite extensive research during the last few decades, the etiology of schizophrenia remains unclear. Evidence of both genetic and environmental influences in the developmental profile of schizophrenia has grown, and due to the complexity of this disorder, a polygenic aspect has been associated with this neuropsychiatric pathology. Unfortunately, no diagnostic strategies based on biological measurement or genetic testing is currently available for schizophrenia. Gene-expression profiling and recent protein studies have shown a decrease in the expression of ubiquitin pathway proteins in the prefrontal cortex of schizophrenia patients. We have examined single nucleotide polymorphisms (or SNPs) within three genes from the ubiquitin protein system: the ubiquitin conjugating enzyme E2D1 (UBE2D1) gene, the E3 SUMOprotein ligase protein inhibitor of activated STAT 2 (PIAS2) gene, and the E3 ubiquitin ligase F-box and leucine-rich repeat protein 21 (FBXL21) gene, in a Caucasian case-control population for schizophrenia. After Bonferroni correction for multiple testing was applied, no significant associations were reported for any of the tested SNPs. Additional genetic analyses will be necessary to fully explore the role of these three genes in schizophrenia. Regarding the rising interest in ubiquitin-related proteins as a therapeutic target in other pathologies such as cancer, further research into the role of ubiquitin pathways in schizophrenia seems topical and timely.

Disciplines

Medicine and Health Sciences | Social and Behavioral Sciences

Publication Details

Andrews, J. L. & Fernandez-Enright, F. (2014). Investigation of genetic variants in ubiquitin enzyme genes involved in the modulation of neurodevelopmental processes: a role in schizophrenia susceptibility?. Genetics Research, 96 e15-1-e15-9.

Investigation of genetic variants in ubiquitin enzyme genes involved in the modulation of neurodevelopmental processes: a role in schizophrenia susceptibility?

Jessica L. Andrews ^{a,c} and Francesca Fernandez-Enright ^{a,b,c}*

^a Centre for Translational Neuroscience, Illawarra Health and Medical Research Institute, Faculty of Science, Medicine and Health, University of Wollongong, New South Wales 2522, Australia ^b School of Daushelery, Faculty of Social Sciences, University of Wollongong, New South

^b School of Psychology, Faculty of Social Sciences, University of Wollongong, New South Wales 2522, Australia

^c Schizophrenia Research Institute, 405 Liverpool Street, Darlinghurst New South Wales, 2010, Australia

Running Title: Investigation of ubiquitin genes in schizophrenia

Submission Category: Research Paper

*Corresponding author:

Dr Francesca Fernandez-Enright, Illawarra Health and Medical Research Institute, University of Wollongong, Northfields Avenue, Wollongong, 2522, NSW, Australia E-mail: fernande@uow.edu.au, Tel: (+61 2) 4221 3494, Fax: (+61 2) 4221 8130

Summary

Despite extensive research during the last decades, the etiology of schizophrenia remains unclear. Evidence of both genetic and environmental influences in the developmental profile of schizophrenia has grown, and due to the complexity of this disorder, a polygenic aspect has been associated with this neuropsychiatric pathology. Unfortunately, no diagnostic strategies based on biological measurement or genetic testing is currently available for schizophrenia. Gene expression profiling and recent protein studies have shown a decrease in the expression of ubiquitin pathway proteins in the prefrontal cortex of schizophrenia patients. We have examined Single Nucleotide Polymorphisms (or SNPs) within three genes from the ubiquitin protein system: the ubiquitin conjugating enzyme E2D1 (UBE2D1) gene, the E3 SUMO-protein ligase protein inhibitor of activated STAT 2 (PIAS2) gene, and the E3 ubiquitin ligase F-box and leucine-rich repeat protein 21 (FBXL21) gene, in a Caucasian case-control population for schizophrenia. After Bonferroni correction for multiple testing was applied, no significant associations were reported for any of the tested SNPs. Additional genetic analyses will be necessary to fully explore the role of these three genes in schizophrenia. Regarding the rising interest of ubiquitin related proteins as a therapeutic target in other pathologies such as cancer, further research into the role of ubiquitin pathways in schizophrenia seem topical and timely.

Keywords: Case-control association; Schizophrenia; Single Nucleotide Polymorphisms; Ubiquitin related genes

1. Introduction

Episodes of mental illness may appear and disappear throughout a person's life, affecting around one in five individuals. Schizophrenia is a mental disorder affecting approximately one percent of the general population and is characterized by symptoms such as hallucinations, delusions, disorganized communication, poor planning, reduced motivation, and blunted affect (Lewis and Lieberman, 2000; Lewis and Levitt, 2002). Despite 50 years of research in schizophrenia, no effective approach for prevention or cure has been produced. The etiology of this disorder remains unclear, although research strongly points towards the interaction between genetic and environmental influences (Tsuang, 2000; Aukes et al., 2008). Recent gene expression and protein studies have reported an alteration of ubiquitin pathways in the brains from schizophrenia sufferers compared to controls (Bousman et al., 2010; Rubio et al., 2013). The ubiquitin protein system plays an essential role in the regulation of membrane and cellular proteins, and has often been referred to as the "kiss of death" due to its labeling of proteins for degradation by proteases (Petroski, 2008; Tai and Schuman, 2008). The major function of the ubiquitin protein complex is to assure intracellular protein degradation by ubiquitination; a highly complex process involving a set of successive enzymes: E1 (ubiquitin activating enzymes), E2 (ubiquitin-conjugating enzymes), E3 (ubiquitin protein ligases), the 20S proteasome, and deubiquitinating enzymes (Yi and Ehlers, 2007). Ubiquitin is first activated by E1 ubiquitin-activating enzymes, before being transferred to its active site, the amino acid cysteine. This transfer requires ATP, making the process energy-dependent. The ubiquitin molecule is then passed on to the second enzymes within the complex, E2 (ubiquitin-conjugating enzymes), before reaching the final group of enzymes, the E3 ubiquitin protein ligases, which recognize and bind the target substrate and labels it with the ubiquitin. This process can be

Andrews and Fernandez-Enright

repeated until a short chain is formed, with three or more ubiquitin molecules usually targeting the protein to the proteasome, where the degradation occurs. Both E2 and E3 proteins exist as large families: more than 35 E2s and 600 E3s have been identified so far, resulting in highly complex combinations of E2s with different E3 proteins defining the substrate specificity. Defects in this ubiquitin-dependent protein degradation have been implicated in the etiology of neurodegenerative diseases, metabolic disorders, cancer (Weathington and Mallampalli, 2014), developmental deficiency, immunity pathologies (Sakamoto, 2002; Pagano and Benmaamar, 2003) and schizophrenia.

We have focused our analysis on three genes coding for ubiquitin related proteins (the ubiquitin conjugating enzyme E2D1 (UBE2D1), the E3 SUMO-protein ligase protein inhibitor of activated STAT 2 (PIAS2) and the E3 ubiquitin ligase F-box and leucine-rich repeat protein 21 (FBXL21), which is a SKP1-cullin-F-box (SCF) protein, implicated in the regulation of the p53 pathway (Figure 1). The p53 pathway plays an essential role in the modulation of neurodevelopmental processes (including cerebral vascularization and neurogenesis) and/or to neurodevelopmental disorders such as schizophrenia. Due to their previous association in different reports and populations (Chen et al., 2008; Middleton et al., 2002), we analyzed a set of potential Single Nucleotide Polymorphisms (SNPs) in the *UBE2D1* (coding for E2D1 protein), *PIAS2* (coding for the protein inhibitor of activated STAT 2) and *FBXL21* gene (coding for F-box and leucine-rich repeat protein 21) in the largest schizophrenia case-control Caucasian population collected in Australia to examine their potential associations with this devastating neurodevelopmental disorder.

2. Materials and methods

(*i*) DNA samples

DNA samples were obtained from the Australian Schizophrenia Research Bank (ASRB). Subjects with schizophrenia were identified using the *Diagnostic and Statistical Manual of Mental Disorders IV* criteria. All samples were from Caucasian volunteers. Subjects were matched for gender and age. The complete sample consisted of 268 schizophrenia cases, comprised of 186 males and 82 females, with an average age of 38.86±11.01 years; and 268 matched controls, comprised of 169 males and 99 females, with an average age of 38.56±12.57 years, with no prior history of mental disorders. After a complete description of the study to the subjects, written informed consent was obtained. This study was approved by, and conducted according to the guidelines of the University of Wollongong Human Research Ethics Committee (HE10/161).

(ii) SNP genotyping

SNPs within the *UBE2D1* (rs11006122 and rs1905455), *PIAS2* (rs8094449, rs10502878, rs11876274, and rs56352844) and *FBXL21* (rs1859427 and rs6861170) genes were tested in our Caucasian schizophrenia case-control population. The selection of these SNPs was based on their previous associations with schizophrenia and/or other disorders, and on their Minor Allele Frequencies (MAF) reported in Caucasian populations (MAF>15%) using HapMap data (http://hapmap.ncbi.nlm.nih.gov). High-throughput SNP genotyping was performed using the MassARRAY® genotyping assay (Sequenom, Inc., San Diego, CA, USA), with the analysis performed by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS). PCR

and extension primer design, selection and multiplexing were performed using MassARRAY® Designer Software (Sequenom, Inc., San Diego, CA, USA).

(iii) Statistical analysis

Power calculations indicated that for the smaller of the two cohorts, a sample size of 204 cases (408 alleles) with a minor allele frequency of 0.2; has >90% a priori power to detect a significant allelic association conferring an odds ratio of 1.5 or greater. The distribution of all tested SNPs did not deviate significantly from Hardy Weinberg Equilibrium (HWE) (p>0.05), except for *UBE2D1* (rs1905455, p=1.88x10⁻¹⁷) which was then excluded from further analysis. To detect associations between each SNP and schizophrenia, chi-square (χ^2) analysis was performed to test for significant differences in allele and genotype frequencies between the case and control groups. The significance for all statistical tests was set to p<0.05 and values p<0.10 were described as trends. Data are expressed as specific counts for alleles and genotypes. Due to multiple testing in the SNP analysis, a standard Bonferroni-corrected p-value of 0.007 (0.05/7) was required to give a 95% probability of correctly concluding not to reject the null hypothesis in the χ^2 test.

3. Results

Two SNPs within *UBE2D1*, four SNPs in *PIAS2* and two SNPs within *FBXL21* were analyzed for association with schizophrenia in a large Australian Caucasian population (268 schizophrenia patients *versus* 268 matched controls with no prior history of mental disorders).

Following Bonferroni correction for multiple comparisons there were no significant associations between the allelic frequency of any of the tested SNPs (*UBE2D1*

rs11006122, PIAS2 rs8094449, rs10502878, rs11876274, and rs56352844; FBXL21 rs1859427 and rs6861170) and schizophrenia (0.08 < p < 0.92; Tables 2, 3 and 4 respectively). In addition there were no genotypic associations between any of the tested SNPs and schizophrenia following Bonferroni correction (0.17<p<0.71: Tables 2. 3 and 4). Further, analysis of each of the genetic markers by gender revealed no significant allelic or genotypic associations with any of these genetic markers and either gender $(0.08 \le p \le 1.00;$ Tables 2, 3 and 4). Interestingly, the demographics of our tested population (Table 1) revealed that a large percentage of our schizophrenia subjects had a family history of mental disorders, twice as many as the control group. χ^2 analysis revealed that both of the FBXL21 genetic markers rs1859427 and rs6861170 had a significant genotypic association with schizophrenia in the subjects whose mother (rs1859427: χ^2 =8.35, p=0.01; rs6861170: χ^2 =8.20, p=0.01) and/or father (rs1859427: χ^2 =16.80, p<0.001; rs6861170: χ^2 =15.54, p<0.001) had a history of mental disorders. None of the other SNPs from any of the other genes studied had any significant associations with schizophrenia in subjects who had a family history of mental disorders $(0.10 \le \chi^2 \le 4.17, 0.12 \le p \le 0.95)$. In addition, a large percentage of subjects from both tested groups experienced some form of self-reported childhood trauma, including but not limited to neglect, physical abuse, sexual abuse, and post-traumatic stress, with a larger number of schizophrenia subjects (41.0%) having a traumatic childhood compared to the control group (22.4%). Again, χ^2 analysis revealed that both of the FBXL21 genetic markers rs1859427 and rs6861170 had a significant genotypic association with schizophrenia in the subjects who experienced childhood trauma (rs1859427: χ^2 =9.67, p=0.007; rs6861170: χ^2 =7.75, p=0.02). Again, none of the other SNPs analyzed had any significant associations with schizophrenia in subjects who experienced trauma during childhood $(1.49 \le \chi^2 \le 4.34, 0.11 \le p \le 0.47)$.

4. Discussion

We have investigated the association of three genes from the ubiquitin protein system involved in the regulation of the p53 pathway in a large Australian Caucasian case-control schizophrenia population. We did not report any significant associations with schizophrenia for any of the tested SNPs in the *UBE2D1*, *PIAS2* and *FBXL21* genes in our population.

There is little information in the literature regarding association studies for our tested genes. Only the *FBXL21* gene has been previously studied in two independent Irish populations, one corresponding to a high density of schizophrenia in Irish families (1,350 subjects from 273 families) and the other was a large Irish case-control population (814 cases *versus*. 625 controls) (Chen et al., 2008). Chen *et. al.* found rs1859427 and rs6861170 to be significantly associated with schizophrenia within their case-control population, and significance was maintained even after correction for multiple testing (p=0.01967 for both markers) (Chen et al., 2008). The MAF for both rs1859427 and rs6861170 *FBXL21* markers were very similar in the schizophrenia group for both the present study (MAF=0.32 for both markers) and in the Chen *et. al.* study (MAF=0.27). This suggests that there is likely to be a difference between the genotyping frequencies in the schizophrenia groups and/or frequencies in the control group between our study and the Chen *et. al.* study.

Interestingly, when we factor into our analysis the subjects who experienced some sort of trauma during childhood, the *FBXL21* SNPs showed a significant association with schizophrenia among the tested SNPs (Table 5), however none of the other tested SNPs in any of the other tested genes had any significant associations in schizophrenia subjects who self-reported any traumatic childhood experiences. Differential epigenetic

Andrews and Fernandez-Enright

gene regulation in relation to traumatic childhood experiences has only ever been reported for one ubiquitin E3 ligase (Mahogunin Ring Finger 1 or MGRN1) in women with fibromyalgia who had experienced traumatic childhood events (Menzies et al., 2013). This suggests that adverse childhood experiences are able to induce genetic and epigenetic variations in ubiquitin protein genes responding to stressful conditions such as Ring Type E3 ubiquitin ligase and F-box proteins (Hermand, 2006; Hua and Vierstra, 2011). A number of studies have shown strong associations between negative childhood experiences and adult psychiatric illnesses, in particular depression, psychosis and schizophrenia (Edwards et al., 2003; Kelleher et al., 2008; Lu et al., 2008). When we accounted for a family history of mental illness from either the mother and/or father within our tested population, we found a positive association between the FBXL21 SNPs and schizophrenia (Table 5); but again there were no significant associations with any of the other tested SNPs in the tested genes in schizophrenia subjects who had a family history of mental illness, suggesting an inheritance for the FBXL21 genetic markers in the context of psychiatric disorders. This is in line with the Chen et. al study which showed that the haplotype of FBXL21 markers rs1859472 and rs6861170 were over-transmitted in the Irish study of high density schizophrenia families.

As mentioned above, the *FBXL21* gene encodes for an E3 ubiquitin ligase F-box protein in the SKP1-cullin-F-box (SCF) complex. The FBXL21 protein is known to be able to stabilize cryptochrome (CRY) proteins CRY1 and CRY2, which are implicated in regulating mammalian circadian rhythms (Hirano et al., 2013). In fact, the FBXL21 protein was reported to antagonize the FBXL3 protein, another F-box-type E3 ligase, which ubiquitinates CRY proteins and mediates their degradation (Hirano et al., 2013). By attenuating the destabilizing action of FBXL3 on CRY proteins, FBXL21 expression allows for an adapted regulation of circadian rhythm gene transcription by CRY

proteins according to the circadian cycle. Furthermore, the circadian pattern of FBXL21 expression in the mouse suprachiasmatic nucleus (region responsible for controlling circadian rhythm) is reminiscent of the expression pattern seen for other circadian pacemaker genes such as *Period 1 (PER1)* (Dardente et al., 2008). Mutations in either the FBXL21 or FBXL3 genes can lead to a dysfunction of circadian rhythm oscillations and lead to significant behavioral disturbances in individuals and alterations in their sleeping patterns; moreover the absence of *FBXL21* causes a short-period phenotype in both mice and cells (Hirano et al., 2013; Yoo et al., 2013). Interestingly, schizophrenia patients have been reported to have intrinsically unstable circadian oscillators. A study by Bromundt et. al. recently provided important new information concerning the link between impairments in neuropsychological function and disrupted circadian rhythms in schizophrenia. (Bromundt et al., 2011); which is further supported by a microarray study which found a significant downregulation of the circadian pacemaker gene PER1 in the postmortem temporal cortex of schizophrenia patients compared to healthy controls (Aston et al., 2004). Furthermore, an animal model study has shown dysfunction in the synchronization of circadian rhythms between brain cell networks involved in sleep-wake regulation and cognition (Dudley et al., 2003). Overall this suggests that alterations in circadian rhythms are present in schizophrenia, and that polymorphisms in the FBXL21 gene in addition to the FBX3 gene, could be involved in the circadian cycle disturbances that have been observed in schizophrenia (Mansour et al., 2009).

The *FBXL21* gene is located on Chromosome 5q31, a region that has previously been shown to contribute to the susceptibility for schizophrenia in both German and Israeli pedigree families (lod score 1.8) (Schwab et al., 1997). Putative loci associated with psychosis in bipolar disorder pedigrees were characterized in the chromosomic region

Andrews and Fernandez-Enright

18q21, which includes the locus for the PIAS2 gene (Park et al., 1995). This region has also been suggested to be an influential genetic loci, common to both schizophrenia and bipolar disorders, depending on polygenic influence and critical environmental factors (Mors et al., 1997). Unfortunately, we did not report any significant associations between the tested SNPs (promoter and intron variants) in the PIAS2 gene with schizophrenia, suggesting that other genetic markers within this gene may be involved. Similarly, the genetic polymorphisms analyzed within the UBE2D1 gene (located in the promoter region) were not associated with schizophrenia in our tested population. The UBE2D1 is located the chromosomic region 10q21.1 gene at (http://www.ncbi.nlm.nih.gov/gene/7321). This region includes the Ankyrin 3 (ANK3) gene, previously associated with schizophrenia by a number of studies that found the rs10761482 SNP to be associated with schizophrenia in a large European population as well as Han Chinese populations (Gella et al., 2011; Yuan et al., 2012).

Due to its early expression in brain development and its key role in genomic stability as well as apoptotic process in brain cells, the p53 pathway plays an essential role in the modulation of neurodevelopmental processes, including those within the schizophrenia pathophysiology. Interestingly a reduced risk of cancer has been observed in individuals with schizophrenia during the last decade; considering the significant role p53 plays in the progression of cancer, this suggests a significant role of p53 in schizophrenia. Previous p53 polymorphisms were found to be associated with schizophrenia in both Chinese and Caucasian case-control populations and in family studies (Yang et al., 2004; Ni et al., 2005). It is thought that the regulation of p53 expression by ubiquitin degradation may play an important role in schizophrenia pathophysiology. Although the present study did not examine polymorphisms within p53, all of the genes examined are involved in p53 signaling. While we did not find any significant associations between

any of the tested polymorphisms and schizophrenia, replicating our study in a larger population and/or testing additional polymorphisms within the same genes will add to, and allow for further exploration of the role of the tested ubiquitin related genes in the genetic vulnerability of schizophrenia.

Our study reports the analysis of potential SNPs in three candidate genes from the ubiquitin protein system in a large Australian case-control schizophrenia population. Due to the limited information available on the role of E2 ubiquitin conjugating enzymes and E3 ubiquitin ligases in schizophrenia, in addition to the increasing variety of these groups of proteins, additional studies will be necessary to further examine the role of these ubiquitin proteins in the genetics of schizophrenia. A growing interest has recently emerged which is targeting ubiquitin related proteins in the treatment of cancer and inflammatory related diseases. Keeping in mind the paradoxical relationship between cancer and schizophrenia, the ubiquitin protein systems seem to be a good candidate to further analyze in the genetics and therapy for schizophrenia.

Acknowledgements

This work was supported by the Schizophrenia Research Institute, utilizing infrastructure and funding from NSW Health.

This study was supported by the Australian Schizophrenia Research Bank (ASRB), which is supported by the National Health and Medical Research Council of Australia, the Pratt Foundation, Ramsay Health Care, the Viertel Charitable Foundation and the Schizophrenia Research Institute.

We thank Pavel Bitter and Shalima Nair from the Australian Cancer Research Foundation at The Garvan Institute of Medical Research in Sydney for their help in the MassARRAY genotyping assay of our samples.

Jessica L. Andrews is supported by an Ian Scott Scholarship from Australian Rotary Health.

This research received no specific grant from any funding agency, commercial or notfor-profit sectors.

Declaration of Interest

None

References

- Aston, C, Jiang, L, Sokolov, BP. 2004. Microarray analysis of postmortem temporal cortex from patients with schizophrenia. Journal of Neuroscience Research 77: 858–866.
- Aukes, MF, Alizadeh, BZ, Sitskoorn, MM, Selten, J-P, Sinke, RJ, Kemner, C, Ophoff, RA & Kahn, RS. (2008). Finding suitable phenotypes for genetic studies of schizophrenia: heritability and segregation analysis. *Biol. Psychiatry* 64: 128– 136.
- Bousman, CA, Chana, G, Glatt, SJ, Chandler, SD, Lucero, GR, Tatro, E, May, T, Lohr, JB, Kremen, WS, Tsuang, MT & Everall, IP. (2010). Preliminary evidence of ubiquitin proteasome system dysregulation in schizophrenia and bipolar disorder: convergent pathway analysis findings from two independent samples. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 153B: 494–502.
- Bromundt, V, Köster, M, Georgiev-Kill, A, Opwis, K, Wirz-Justice, A, Stoppe, G, Cajochen, C. 2011. Sleep-wake cycles and cognitive functioning in schizophrenia. Br J Psychiatry 198: 269–276.
- Chen, X, Wang, X, Sun, C, Chen, Q, O'Neill, FA, Walsh, D, Fanous, A & Kendler, KS. (2008). FBXL21 association with schizophrenia in Irish family and case-control samples. Am. J. Med. Genet. B Neuropsychiatr. Genet. 147B: 1231–1237.
- Dardente, H, Mendoza, J, Fustin, J-M, Challet, E, Hazlerigg, DG. 2008. Implication of the F-Box Protein FBXL21 in Circadian Pacemaker Function in Mammals. PLoS ONE 3: e3530.
- Dudley, CA, Erbel-Sieler, C, Estill, SJ, Reick, M, Franken, P, Pitts, S, McKnight, SL. 2003. Altered patterns of sleep and behavioral adaptability in NPAS2-deficient mice. Science 301: 379–383.
- Edwards, VJ, Holden, GW, Felitti, VJ & Anda, RF. (2003). Relationship between multiple forms of childhood maltreatment and adult mental health in community respondents: results from the adverse childhood experiences study. *Am J Psychiatry* 160: 1453–1460.
- Gella, A, Segura, M, Durany, N, Pfuhlmann, B, Stöber, G & Gawlik, M. (2011). Is Ankyrin a genetic risk factor for psychiatric phenotypes? *BMC Psychiatry* 11: 103.

Hermand, D. (2006). F-box proteins: more than baits for the SCF? Cell Div 1: 30.

Hirano, A, Yumimoto, K, Tsunematsu, R, Matsumoto, M, Oyama, M, Kozuka-Hata, H, Nakagawa, T, Lanjakornsiripan, D, Nakayama, KI, Fukada, Y. 2013. FBXL21 regulates oscillation of the circadian clock through ubiquitination and stabilization of cryptochromes. Cell 152: 1106–1118.

- Hua, Z & Vierstra, RD. (2011). The cullin-RING ubiquitin-protein ligases. *Annu Rev Plant Biol* 62: 299–334.
- Kelleher, I, Harley, M, Lynch, F, Arseneault, L, Fitzpatrick, C & Cannon, M. (2008). Associations between childhood trauma, bullying and psychotic symptoms among a school-based adolescent sample. *Br J Psychiatry* 193: 378–382.
- Lewis, DA & Levitt, P. (2002). Schizophrenia as a disorder of neurodevelopment. *Annu Rev Neurosci* 25: 409–32.
- Lewis, DA & Lieberman, JA. (2000). Catching up on schizophrenia: natural history and neurobiology. *Neuron* 28: 325–334.
- Lu, W, Mueser, KT, Rosenberg, SD & Jankowski, MK. (2008). Correlates of adverse childhood experiences among adults with severe mood disorders. *Psychiatr Serv* 59: 1018–1026.
- Mansour, HA, Talkowski, ME, Wood, J, Chowdari, KV, McClain, L, Prasad, K, Montrose, D, Fagiolini, A, Friedman, ES, Allen, MH, Bowden, CL, Calabrese, J, El-Mallakh, RS, Escamilla, M, Faraone, SV, Fossey, MD, Gyulai, L, Loftis, JM, Hauser, P, Ketter, TA, Marangell, LB, Miklowitz, DJ, Nierenberg, AA, Patel, J, Sachs, GS, Sklar, P, Smoller, JW, Laird, N, Keshavan, M, Thase, ME, Axelson, D, Birmaher, B, Lewis, D, Monk, T, Frank, E, Kupfer, DJ, Devlin, B, Nimgaonkar, VL. 2009. Association study of 21 circadian genes with bipolar I disorder, schizoaffective disorder, and schizophrenia. Bipolar Disord 11: 701– 710.
- Menzies, V, Lyon, DE, Archer, KJ, Zhou, Q, Brumelle, J, Jones, KH, Gao, G, York, TP & Jackson-Cook, C. (2013). Epigenetic alterations and an increased frequency of micronuclei in women with fibromyalgia. *Nurs Res Pract* 2013: 795784.
- Middleton, FA, Mirnics, K, Pierri, JN, Lewis, DA & Levitt, P. (2002). Gene expression profiling reveals alterations of specific metabolic pathways in schizophrenia. *J. Neurosci.* 22: 2718–2729.
- Mors, O, Ewald, H, Blackwood, D & Muir, W. (1997). Cytogenetic abnormalities on chromosome 18 associated with bipolar affective disorder or schizophrenia. *Br J Psychiatry* 170: 278–280.
- Ni, X, Trakalo, J, Valente, J, Azevedo, MH, Pato, MT, Pato, CN, Kennedy, JL. 2005. Human p53 tumor suppressor gene (TP53) and schizophrenia: case-control and family studies. Neurosci. Lett. 388: 173–178.
- Pagano, M & Benmaamar, R. (2003). When protein destruction runs amok, malignancy is on the loose. *Cancer Cell* 4: 251–256.
- Park, S, Holzman, PS & Goldman-Rakic, PS. (1995). Spatial working memory deficits in the relatives of schizophrenic patients. *Arch. Gen. Psychiatry* 52: 821–828.

- Petroski, MD. (2008). The ubiquitin system, disease, and drug discovery. *BMC Biochem.* 9 Suppl 1: S7.
- Rubio, MD, Wood, K, Haroutunian, V & Meador-Woodruff, JH. (2013). Dysfunction of the ubiquitin proteasome and ubiquitin-like systems in schizophrenia. *Neuropsychopharmacology* 38: 1910–1920.
- Sakamoto, KM. (2002). Ubiquitin-dependent proteolysis: its role in human diseases and the design of therapeutic strategies. *Mol. Genet. Metab.* 77: 44–56.
- Schwab, SG, Eckstein, GN, Hallmayer, J, Lerer, B, Albus, M, Borrmann, M, Lichtermann, D, Ertl, MA, Maier, W & Wildenauer, DB. (1997). Evidence suggestive of a locus on chromosome 5q31 contributing to susceptibility for schizophrenia in German and Israeli families by multipoint affected sib-pair linkage analysis. *Mol. Psychiatry* 2: 156–160.
- Tai, H-C & Schuman, EM. (2008). Ubiquitin, the proteasome and protein degradation in neuronal function and dysfunction. *Nat. Rev. Neurosci.* 9: 826–838.
- Tsuang, M. (2000). Schizophrenia: genes and environment. Biol Psychiatry 47: 210-20.
- Weathington, NM & Mallampalli, RK. (2014). Emerging therapies targeting the ubiquitin proteasome system in cancer. J. Clin. Invest. 124: 6–12.
- Yang, Y, Xiao, Z, Chen, W, Sang, H, Guan, Y, Peng, Y, Zhang, D, Gu, Z, Qian, M, He, G, Qin, W, Li, D, Gu, N, He, L. 2004. Tumor suppressor gene TP53 is genetically associated with schizophrenia in the Chinese population. Neurosci. Lett. 369: 126–131.
- Yi, JJ & Ehlers, MD. (2007). Emerging Roles for Ubiquitin and Protein Degradation in Neuronal Function. *Pharmacol Rev* 59: 14–39.
- Yoo, S-H, Mohawk, JA, Siepka, SM, Shan, Y, Huh, SK, Hong, H-K, Kornblum, I, Kumar, V, Koike, N, Xu, M, Nussbaum, J, Liu, X, Chen, Z, Chen, ZJ, Green, CB, Takahashi, JS. 2013. Competing E3 Ubiquitin Ligases Govern Circadian Periodicity by Degradation of CRY in Nucleus and Cytoplasm. Cell 152: 1091– 1105.
- Yuan, A, Yi, Z, Wang, Q, Sun, J, Li, Z, Du, Y, Zhang, C, Yu, T, Fan, J, Li, H & Yu, S. (2012). ANK3 as a risk gene for schizophrenia: new data in Han Chinese and meta analysis. Am. J. Med. Genet. B Neuropsychiatr. Genet. 159B: 997–1005.

	Control subjects n (%)	Schizophrenia subjects n (%)
Gender		
Female	99 (37%)	82 (30.6%)
Male	169 (63%)	186 (69.4%)
Age at assessment (years)		
Female	32.19	40.48
Male	42.22	38.14
Family history of mental disorders		
Mother	41 (15.3%)	95 (35.5%)
Father	49 (18.3%)	106 (39.6%)
Traumatic childhood experience	60 (22.4%)	110 (41.0%)

Table 1. Subject Demographics for Control (n = 268) and Schizophrenia Subjects (n =
268).

	Ν	Alle	eles	Genotypes			
UBE2D1	(alleles) rs11006122	С	Т	CC	СТ	TT	
Schizophrenia	264	150 (56.8%)	114 (43.2%)	45 (34.1%)	60 (45.5%)	27 (20.4%)	
Male	190	106 (55.8%)	84 (44.2%)	30 (31.6%)	46 (48.4%)	19 (20.0%)	
Female	74	44 (59.5%)	30 (40.5%)	15 (40.5%)	14 (37.8%)	8 (21.6%)	
Control	368	217 (59.0%)	151 (41.0%)	63 (34.2%)	91 (49.5%)	30 (16.3%)	
Male	232	134 (57.8%)	98 (42.2%)	40 (34.5%)	54 (46.6%)	22 (18.9%)	
Female	136	83 (61.0%)	53 (39.0%)	23 (33.9%)	37 (54.4%)	8 (11.7%)	
Total case vs. control		$\chi^2 = 0.29$	p = 0.58		$\chi^2 = 0.99$	p = 0.61	
Male case vs. control		$\chi^2 = 0.17$	p = 0.68		$\chi^2 = 0.20$	p = 0.90	
Female case vs. control		$\chi^2 = 0.05$	p = 0.82		$\chi^2 = 3.18$	p = 0.20	

Table 2. Allelic and genotypic distributions for UBE2D1 genetic marker in schizophrenia subjects and controls.

Ν		Alle	les	Genotypes		
PIAS2	(alleles) rs8094449	G	А	GG	ĠĂ	AA
Schizophrenia	442	423 (95.7%)	19 (4.3%)	202 (91.4%)	19 (8.6%)	0 (0%)
Male	314	300 (95.5%)	14 (4.5%)	143 (91.1%)	14 (8.9%)	0 (0%)
Female	128	123 (96.1%)	5 (3.9%)	59 (92.2%)	5 (7.8%)	0 (0%)
Control	498	471 (94.6%)	27 (5.4%)	223 (89.6%)	25 (10.0%)	1 (0.4%)
Male	314	300 (95.5%)	14 (4.5%)	143 (91.1%)	14 (8.9%)	0 (0%)
Female	184	171 (92.9%)	13 (7.1%)	80 (87.0%)	11 (11.9%)	1 (1.1%)
Total case <i>vs</i> . control	101	$\chi^2 = 0.63$	p = 0.42	00 (07.070)	$\chi^2 = 1.19$	p = 0.55
Male case <i>vs.</i> control		$\chi^2 = 0.00$	p = 1.00		$\chi^2 = 0.00$	p = 1.00
Female case vs. control		$\chi^2 = 0.39$	p = 0.24		$\chi^2 = 1.44$	p = 0.48
	27	/5	<u> </u>		<i>,</i> ,,	p 0.10
DIACO	$\frac{N}{(2112122)}$	Alle	T	00	Genotypes	TT
PIAS2	(alleles) rs10502878	С		CC	СТ	TT
Schizophrenia	204	160 (78.4%)	44 (21.6%)	62 (60.8%)	36 (35.3%)	4 (3.9%)
Male	150	117 (78.0%)	33 (22.0%)	46 (61.3%)	25 (33.3%)	4 (5.3%)
Female	54	43 (79.6%)	11 (20.4%)	16 (59.3%)	11 (40.7%)	0 (0%)
Control	324	273 (84.3%)	51 (15.7%)	113 (69.8%)	47 (29.0%	2 (1.2%
Male	210	173 (82.4%)	37 (17.6%)	69 (65.7%)	35 (33.3%)	1 (0.9%
Female	114	100 (87.7%)	14 (12.3%)	44 (77.2%)	12 (42.1%)	1 (1.7%
Total case vs. control		$\chi^2 = 2.88$	p = 0.08		$\chi^2 = 3.53$	$p = 0.1^{2}$
Male case vs. control		$\chi^2 = 1.07$	p = 0.30		$\chi^2 = 3.15$	p = 0.20
Female case vs. Control		$\chi^2 = 1.89$	p = 0.16		$\chi^2 = 3.89$	p = 0.14
	Ν	Alle	les		Genotypes	
PIAS2	(alleles) rs11876274	Т	С	TT	TC	CC
Schizophrenia	278	260 (93.5%)	18 (6.5%)	121 (87.1%)	18 (12.9%)	0 (0%)
Male	196	184 (93.9%)	12 (6.2%)	86 (87.8%)	12 (12.2%)	0 (0%)
Female	82	76 (92.7%)	6 (7.3%)	35 (85.4%)	6 (14.6%)	0 (0%)
Control	382	354 (92.7%)	28 (7.3%)	164 (85.9%)	26 (13.6%)	1 (0.5%
Male	238	224 (94.1%)	14 (5.9%)	105 (88.2%)	14 (11.8%)	0 (0%)
Female	144	130 (90.3%)	14 (9.7%)	59 (81.9%)	12 (16.7%)	1 (1.4%
Total case vs. control		$\chi^2 = 0.18$	p = 0.67		$\chi^2 = 0.77$	p = 0.6
Male case <i>vs.</i> control		$\chi^2 = 0.01$	p = 0.91		$\chi^2 = 0.01$	p = 0.9
Female case <i>vs</i> . control		$\chi^2 = 0.37$	p = 0.54		$\chi^2 = 0.67$	p = 0.7
	Ν	Alle			Genotypes	
		Α	G	AA	AG	GG
PIAS2	(alleles) rs56352844	Α	-			
PIAS2 Schizophrenia		A 193 (93.7%)	13 (6.3%)	90 (87.4%)	13 (12.6%)	0 (0%)
	rs56352844			90 (87.4%) 68 (88.3%)	13 (12.6%) 9 (11.7%)	· · · ·
Schizophrenia	rs56352844 206	193 (93.7%)	13 (6.3%)		· · · ·	0 (0%)
Schizophrenia Male	rs56352844 206 154 52	193 (93.7%) 145 (94.2%)	13 (6.3%) 9 (5.8%) 4 (7.3%)	68 (88.3%)	9 (11.7%) 4 (15.4%)	0 (0%) 0 (0%)
Schizophrenia Male Female	rs56352844 206 154	193 (93.7%) 145 (94.2%) 48 (92.3%)	13 (6.3%) 9 (5.8%)	68 (88.3%) 22 (84.6%)	9 (11.7%) 4 (15.4%) 18 (11.0%)	0 (0%) 0 (0%) 1 (0.6%
Schizophrenia <i>Male</i> <i>Female</i> Control	rs56352844 206 154 52 328	193 (93.7%) 145 (94.2%) 48 (92.3%) 308 (93.9%)	13 (6.3%) 9 (5.8%) 4 (7.3%) 20 (6.1%) 12 (5.7%)	68 (88.3%) 22 (84.6%) 145 (88.4%) 94 (88.7%)	9 (11.7%) 4 (15.4%)	0 (0%) 0 (0%) 1 (0.6%) 0 (0%)
Schizophrenia Male Female Control Male Female	rs56352844 206 154 52 328 212	193 (93.7%) 145 (94.2%) 48 (92.3%) 308 (93.9%) 200 (94.3%) 108 (93.1%)	13 (6.3%) 9 (5.8%) 4 (7.3%) 20 (6.1%) 12 (5.7%) 8 (6.9%)	68 (88.3%) 22 (84.6%) 145 (88.4%)	9 (11.7%) 4 (15.4%) 18 (11.0%) 12 (11.3%) 6 (10.3%)	0 (0%) 0 (0%) 1 (0.6% 0 (0%) 1 (1.7%
Schizophrenia Male Female Control Male	rs56352844 206 154 52 328 212	193 (93.7%) 145 (94.2%) 48 (92.3%) 308 (93.9%) 200 (94.3%)	13 (6.3%) 9 (5.8%) 4 (7.3%) 20 (6.1%) 12 (5.7%)	68 (88.3%) 22 (84.6%) 145 (88.4%) 94 (88.7%)	9 (11.7%) 4 (15.4%) 18 (11.0%) 12 (11.3%)	$\begin{array}{c} 0 \ (0\%) \\ 0 \ (0\%) \\ 0 \ (0\%) \\ 1 \ (0.6\%) \\ 0 \ (0\%) \\ 1 \ (1.7\%) \\ p = 0.66 \\ p = 0.99 \end{array}$

Table 3. Allelic and genotypic distributions for *PIAS2* genetic markers in schizophrenia subjects and controls.

	Ν	N Alleles			Genotypes			
FBXL21	(alleles) rs1859427	G	А	GG	GA	AA		
Schizophrenia	266	179 (67.3%)	87 (32.7%)	64 (48.1%)	51 (38.3%)	18 (13.6%)		
Male	192	129 (67.2%)	63 (32.8%)	45 (46.9%)	39 (40.6%)	12 (12.5%)		
Female	74	50 (67.6%)	24 (32.4%)	19 (51.4%)	12 (32.4%)	6 (16.2%)		
Control	374	264 (70.6%)	110 (29.4%)	98 (52.4%)	68 (36.4%)	21 (11.2%)		
Male	234	166 (70.9%)	68 (29.1%)	64 (54.7%)	38 (32.5%)	15 (12.8%)		
Female	140	98 (70%)	42 (30%)	34 (48.6%)	30 (42.8%)	6 (8.6%)		
Total case vs. control		$\chi^2 = 0.79$	p = 0.37		$\chi^2 = 0.70$	p = 0.70		
Male case vs. control		$\chi^2 = 0.70$	p = 0.40		$\chi^2 = 1.60$	p = 0.44		
Female case vs. control		$\chi^2 = 0.02$	p = 0.87		$\chi^2 = 1.97$	p = 0.37		
	Ν	All	eles		Genotypes			
FBXL21	(alleles) rs6861170	Т	G	TT	TG	GG		
Schizophrenia	268	182 (67.9%)	86 (32.1%)	66 (49.3%)	50 (37.3%)	18 (13.4%)		
Male	192	132 (68.8%)	60 (31.2%)	47 (49.0%)	38 (39.6%)	11 (11.4%)		
Female	76	50 (65.8%)	26 (34.2%)	19 (50%)	12 (31.6%)	7 (18.4%)		
Control	372	264 (71.0%)	108 (29.0%)	98 (52.7%)	68 (36.7%)	20 (10.6%)		
Male	234	167(71.4%)	67 (28.6%)	64 (54.7%)	39 (33.3%)	14(12.0%)		
Female	138	97 (70.3%)	41 (29.7%)	34 (49.3%)	29 (42.0%)	6 (8.7%)		
Total case vs. control		$\chi^2 = 0.69$	p = 0.41		$\chi^2 = 0.66$	p = 0.71		
Male case vs. control		$\chi^2 = 0.35$	p = 0.55		$\chi^2 = 0.92$	p = 0.63		
Female case vs. control		$\chi^2 = 0.46$	p = 0.49		$\chi^2 = 4.84$	p = 0.08		

Table 4. Allelic and genotypic distributions for *FBXL21* genetic marker inschizophrenia subjects and controls.

		Genotypes	
FBXL21	GG	GA	AA
rs1859427			
Schizophrenia	64 (48.1%)	51 (38.3%)	18 (13.6%)
Mother mental history	19 (46.3%)	19 (46.3%)	3 (7.4%)
Father mental history	29 (56.9%)	18 (35.3%)	4 (7.8%)
Traumatic childhood	25 (46.3%)	21 (38.9%)	8 (14.8%)
Control	98 (52.4%)	68 (36.4%)	21 (11.2%)
Mother mental history	8 (29.6%)	14 (51.9%)	5 (18.5%)
Father mental history	18 (52.9%)	13 (38.2%)	3 (8.8%)
Traumatic childhood	13 (37.1%)	19 (54.3%)	3 (8.6%)
Mother mental case vs. control	. ,	$\chi^2 = 8.35$	p = 0.01
Father mental case vs. control		$\chi^2 = 16.80$	p < 0.001
Traumatic childhood case vs. control		$\chi^2 = 9.67$	p = 0.007
		Genotypes	
FBXL21	TT	TG	GG
rs6861170			
Schizophrenia	66 (49.3%)	50 (37.3%)	18 (13.4%)
Mother mental history	20 (47.6%)	18 (42.9%)	4 (9.5%)
Father mental history	29 (54.7%)	18 (34.0%)	6 (11.3%)
Traumatic childhood	25 (44.6%)	21 (37.5%)	10 (17.9%)
Control	98 (52.7%)	68 (36.7%)	20 (10.6%)
Mother mental history	9 (34.6%)	13 (50.0%)	4 (15.4%)
Father mental history	18 (54.5%)	13 (39.4%)	2 (6.1%)
Traumatic childhood	13 (38.2%)	18 (52.9%)	3 (8.8%)
Mother mental case vs. control	```	$\chi^2 = 8.20$	p = 0.01
Father mental case vs. control		$\chi^2 = 15.54$	p < 0.001

Table 5. Genotypic distributions for *FBXL21* genetic markers in schizophrenia subjects and controls with respect to parental history of mental health issues and traumatic childhood experiences.

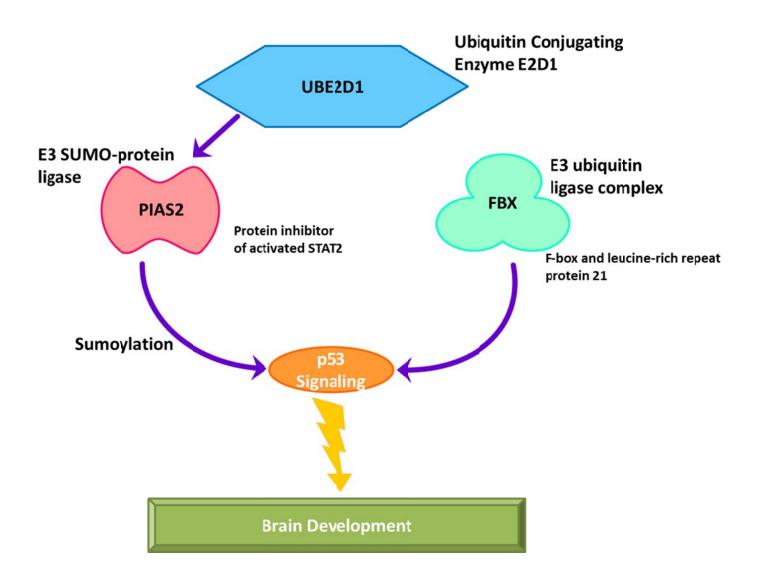


Figure 1. Schematic of the implication of the proteins coded by the genes of interest in brain development.