

University of Wollongong Research Online

Faculty of Science, Medicine and Health - Papers

Faculty of Science, Medicine and Health

2013

Rapid cascade synthesis of poly-heterocyclic architectures from indigo

Alireza Shakoori Ghasabi University of Wollongong, asg072@uowmail.edu.au

John Bremner
University of Wollongong, jbremner@uow.edu.au

Anthony C. Willis
Australian National University

Rachada Haritakun National Science and Technology Development Agency, Thailand

Paul A. Keller
University of Wollongong, keller@uow.edu.au

Publication Details

Shakoori, A., Bremner, J. B., Willis, A. C., Haritakun, R. & Keller, P. A. (2013). Rapid cascade synthesis of poly-heterocyclic architectures from indigo. Journal of Organic Chemistry, 78 (15), 7639-7647.

 $Research\ Online\ is\ the\ open\ access\ institutional\ repository\ for\ the\ University\ of\ Wollongong.\ For\ further\ information\ contact\ the\ UOW\ Library:\ research-pubs@uow.edu.au$

Rapid cascade synthesis of poly-heterocyclic architectures from indigo

Abstract

The base-induced propargylation of the dye indigo results in the rapid and unprecedented one-pot synthesis of highly functionalized representatives of the pyrazino [1,2-a:4,3-a'] diindole, pyrido [1,2-a:3,4-b'] diindole and benzo [b] indolo [1,2-h] naphthyridine heterocyclic systems, with the last two reflecting the core skeleton of the anticancer/antiplasmodial marine natural products fascaplysin and homofascaplysins and a ring B-homologue, respectively. The polycyclic compounds 6–8, whose structures were confirmed through single-crystal X-ray crystallographic analysis, arise from sequential inter/intramolecular substitution—addition reactions, and in some cases, ring rearrangement reactions. Preliminary studies on controlling the reaction path selectivity, and the potential reaction mechanisms, are also described. Initial biological activity studies with these new heterocyclic derivatives indicated promising in vitro antiplasmodial activity as well as good anticancer activity. The chemistry described is new for the indigo moiety and cascade reactions from this readily available and cheap starting material should be more broadly applicable in the synthesis of additional new heterocyclic systems difficult to access by other means.

Keywords

CMMB

Disciplines

Medicine and Health Sciences | Social and Behavioral Sciences

Publication Details

Shakoori, A., Bremner, J. B., Willis, A. C., Haritakun, R. & Keller, P. A. (2013). Rapid cascade synthesis of poly-heterocyclic architectures from indigo. Journal of Organic Chemistry, 78 (15), 7639-7647.

Rapid Cascade Synthesis of Poly-Heterocyclic Architectures from Indigo

Alireza Shakoori[†], John B. Bremner[†], Anthony C. Willis[‡], Rachada Haritakun[§] and Paul A. Keller[†]*

- [†] School of Chemistry, University of Wollongong, Wollongong, NSW, 2522, Australia
- [‡] School of Chemistry, The Australian National University, Canberra, ACT 0200, Australia
- § National Centre for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency (NSTDA), 113 Phaholyothin Road, Klong1, Klong Luang, Pathumanthani 12120 Thailand

Corresponding author email: keller@uow.edu.au

Table of Contents Graphic

ABSTRACT

The base-induced propargylation of the dye, indigo, results in the rapid and unprecedented one-pot synthesis of highly functionalised representatives of the pyrazino[1,2-a:4,3-a]diindole, pyrido[1,2-a:3,4-b]diindole and benzo[b]indolo[1,2-h]naphthyridine heterocyclic systems, the last two reflecting the core skeleton of the anti-cancer / anti-plasmodial marine natural products, fascaplysin and homofascaplysins and a ring B-homolog respectively. The polycyclic compounds 6 - 8, whose structures were confirmed through single crystal X-ray crystallographic analysis, arise from sequential inter/intramolecular substitution-addition reactions, and in some cases, ring rearrangement reactions.

Preliminary studies on controlling the reaction path selectivity, and the potential reaction mechanisms, are also described. Initial biological activity studies with these new heterocyclic derivatives indicated promising *in vitro* anti-plasmodial activity as well as good anti-cancer activity. The chemistry described is new for the indigo moiety and cascade reactions from this readily available and cheap starting material should be applicable more broadly in the synthesis of additional new heterocyclic systems difficult to access by other means.

INTRODUCTION

One of the current goals in organic synthesis is the controlled construction of complex molecules, in particular, through the use of cascade reaction sequences.¹ Such molecular explorations, yielding novel architectures, is of particular interest for the investigation of new bioactive agents with possible new modes of action, which could be subsequently elaborated in medicinal chemistry programs.² Approaches to the realization of these synthetic goals have often been explored in the context of complex multistep syntheses of natural product targets.³ Our focus has been on the smaller, though versatile canvas, of the abundant and cheap natural product, indigo 1 (Scheme 1).

There is significant advantage in starting with a readily available advanced precursor like indigo: its reported chemistry is very limited despite the presence of an array of closely positioned functionality in the 2,2′-diindolic unit which allows for cascade reaction paths. It also provides a unique opportunity in providing an advanced starting material for the potential rapid synthesis of the diindolic system of natural products analogs. Such biologically active natural products (*e.g.* with anti-cancer or antimicrobial activity) include fascaplysin⁴; homofascaplysin⁵; iheyamines⁶; staurosporine⁷ and rebeccamycin⁸ (Figure 1).

Figure 1. Some biologically active natural products with an embedded 2,2′-diindolic unit.

To explore our synthetic approach, we initially investigated the base-mediated reactions of indigo with allylic halides, which in the case of allylic bromide resulted in the rapid synthesis of the multicyclic compounds 2 and 3 (Scheme 1). These compounds represent new ring systems with functionality suitable for further elaboration of molecular complexity.

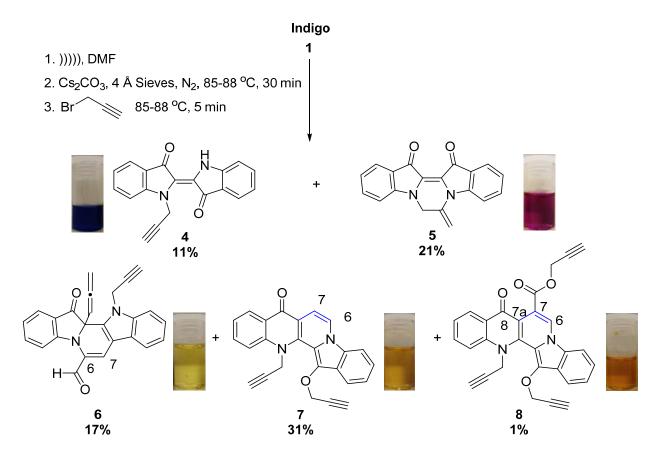
In an extension of this work and in an investigation of reaction directing elements, particularly those modulating nucleophilic-electrophilic reactivities, we have studied the base induced reactions of indigo with the simple alkyne analog, propargyl bromide. New facile routes to the core heterocyclic system of fascaplysin, homofascaplysin and the B-ring homolog of this system were discovered and the results are reported in this paper together with some initial *in vitro* anti-plasmodial and anti-cancer activity data.

Scheme 1. Allylation of indigo

RESULTS AND DISCUSSION

Synthesis and Structural Elucidation

Our previous experience in the chemistry of indigo⁹ revealed that relatively small changes to reaction conditions could have major impacts on the product outcome. Therefore, our attempts at the propargylation of indigo paid particular attention to stringent and repeatable reactions conditions. In this context, a solution of indigo in DMF was generated through sonication for 30 mins at room temperature and then transferred to a septum-equipped flask which contained pre-dried molecular sieves and cesium carbonate under an inert atmosphere. The flask was then plunged into a preheated oil-bath (strictly 85-87 °C) and stirred for 30 mins, followed by the addition of propargyl bromide and the reaction mixture heated at this temperature for 5 mins. Following quenching, a sequence of separations yielded the five new products 4 - 8 in a combined yield of 81% (Scheme 2).



Scheme 2. Base-mediated propargylation of indigo to produce the heterocyclic compounds **4 - 8**. Adjacent to each structure is illustrated a solution, highlighting the color.

The expected mono *N*-substituted indigo derivative (**4**) was isolated as a deep blue, papery solid in 11% yield after silica-gel column chromatography. Data from the HRESI mass spectrum was consistent with the molecular formula of **4** and was indicative of the addition of one propargyl unit. ¹H NMR NOE analysis also confirmed the presence of the expected *trans* isomer. In a separate reaction, under analogous conditions but with a very short reaction time (<1 min) and a stoichiometric quantity of propargyl bromide, compound **4** could be isolated in a much improved 93% yield (Scheme 3, Step **A**). The increased solubility in a range of organic solvents (e.g. THF, CH₂Cl₂) enables this monopropargylated material to be used as a starting material for subsequent reactions, including

cyclisations. This increased flexibility in the use of solvents allows for a greater variation in reaction conditions to be used.

Scheme 3. Synthesis of 4 and subsequent conversion to 5

The pyrazinodiindole **5** (Scheme 2) was isolated in 21% yield as a red-burgundy solid. The ¹H NMR spectrum showed singlets at 5.04 and 5.38 ppm, assigned to the exocyclic methylene protons whereas the ¹³C NMR spectrum showed 2 peaks at 179.7 and 180.8 ppm, assigned to two non-equivalent carbonyls. Data from the HRESI mass spectrum was supportive of the molecular formula of **5** and was indicative of the addition of one propargyl unit.

The UV-vis spectrum of 5 had a strong adsorption band with a maximum at 324 nm ($\varepsilon = 13,088$). The burgundy color was suggestive that the central double bond of indigo remained intact, with all other indigo derivatives in which this bond had been converted to a single bond appearing as yellow compounds. Simple modelling studies (Spartan, Wavefunction) indicated that compound 5, while planar through the indigo moiety, positioned the hydrogen atoms of the endocyclic methylene group above and below the plane of the molecule with the exocyclic methylene twisted out of the molecule

plane.¹⁰ This product **5** can also be synthesized in high yield (98%) by the reaction of **4** in DMF in the presence of Cs₂CO₃ (Scheme 3, Step **B**).

The pyridodiindole **6** (Scheme 2) was isolated in 17% yield as yellow-orange crystals. Analysis of the 1 H NMR indicated a peak at 9.62 ppm assigned to the aldehyde proton, and a doublet at 4.79 ppm (J = 6.5 Hz) assigned to the terminal protons of the allene moiety and a singlet at 2.33 assigned to the terminal alkyne proton. The 13 C NMR spectrum showed peaks at 90.4, 208.4 and 80.4 ppm, assigned sequentially to the three allene carbons from the CH. A peak at 195.7 ppm was assigned to the aldehyde. The HRESI mass spectrum of **6** was consistent with a molecular formula of $C_{25}H_{16}N_{2}O_{2}$. The molecular structure was confirmed as **6** by X-ray crystallographic analysis, together with the disposition of the allene group over the pyrido ring rather than pointing away from it (Scheme 2). There is one stereogenic carbon present at C12a, but in the absence of any chiral element during the reaction, the stereochemical outcome was a racemic mixture, as confirmed by optical rotation analysis. The structure of **6** poses interesting mechanistic questions. The addition of three propargyl units is evidenced by the presence of the allene, the *N*-propargyl unit and the three carbon moiety encompassing the aldehyde, and C6 and C7. Interestingly, this three carbon unit is attached to the indigoyl N at the middle carbon, and not through the propargyl bromide methylene or the terminal alkynic positions.

The major product of the reaction was the benzoindolonaphthyridinone **7** (scheme 2) isolated in 31% yield as a yellow solid which was also highly fluorescent in CH_2Cl_2 solution with a brilliant yellow color under UV light (365 nm). The ¹H NMR showed two pairs of doublets at 8.01 and 7.33 ppm (J = 7.4 Hz) assigned to H6 and H7 respectively. Two singlets at 2.14 and 2.30 ppm were assigned to the terminal acetylenic protons with the corresponding propargyl methylenes assigned to the doublets at 4.74 and 5.49 ppm (J = 2.4 Hz). A comparative analysis of the NMR and mass spectra indicated that although two substituent propargyl units were present, the molecular ion indicated the addition of three propargyl units, the last being incorporated into a ring (Scheme 2, **7**, blue). The structure of **7** was

confirmed by X-ray crystallographic analysis. The ring expansion of the indigo 5-membered ring into a 6-membered ring is new chemistry, with the three additional carbons of the indolo[1,2-h][1,7]naphthyridine parent structure being sourced from the additional propargyl unit. Retrosynthetically, sourcing the C6-C7-C7a moiety from a propargyl unit with the remaining skeleton from indigo isn't intuitive and highlights the novelty of this new chemistry of indigo. The final product, isolated in very low yield (1%), was the yellow benzoindolonaphthyridinone **8** (Scheme 2), with the structure confirmed by single-crystal X-ray analysis.

Color is an important qualitative element in the structural elucidation of these polycyclic compounds. The disappearance of the blue and emergence of yellow appears to indicate the loss of H-bonding between the indigo carbonyl and the NH, along with loss of unsaturation in the indigo central bond and the presence of either sp³ hybridised carbon atoms (*e.g.* 6), or extended fused ring systems *e.g.* heterocycles 7 or 8. The mono-*N*-propargylated 4, with both structural elements still present, maintains the deep blue intensity whereas the cyclized structure 5, which still contains the central double bond but has lost the H-bonding, is a burgundy color (Scheme 2).

Mechanistic and Reaction Discussion

The proposed mechanisms for the propargylation of indigo are summarised in Schemes 4-6 and involve five key pathways. The pyrazinodiindole **5** is derived from the monopropargylated indigo **4** (blue, **Path I**, Scheme 4, and Scheme 3) after *N*-deprotonation, followed by delocalisation allowing rotation around the central bond to the *cisoid* conformation. Subsequent intramolecular nucleophilic addition to the propargyl C2 position yields **5** (Scheme 4).

$$\begin{array}{c} \text{Br} \\ \text{indigo} \\ \textbf{1} \end{array} \begin{array}{c} \text{Cs}_2\text{CO}_3 \\ \text{Path I} \end{array}$$

Scheme 4. Proposed mechanism for formation of **4** and **5**. Structures that are colored indicate common intermediates in the overall mechanism and are the same within Schemes 5 and 6.

Path II starts with the identical key intermediate **4** (blue, Scheme 5) undergoing prototropic tautomerism (**A**) which subsequently *N*-alkylates (**B**). Deprotonation of the *N*-methylene generates a stabilised ylid, which allows cyclisation onto the carbonyl generating an activated cyclic allene intermediate (**C**). Under standard conditions, an 'alkyne nucleophile' is insufficiently strong to attack an electrophilic carbonyl in the absence of a metal (*e.g.* Au, Ru) or an activating influence. In this instance, the anion from the ylid serves as a formal negative charge allowing this cyclisation to the 7-ring allene to occur. A comprehensive review on allenes from 1989¹¹ reported the isolation of an eightmembered carbocyclic allene, however, the corresponding six-membered rings have been plausibly demonstrated as reactive intermediates.¹² Further, with the seven-membered carbocyclic allene, isodesmic reaction energy calculations indicate¹³ an allene strain component of 13.5 - 14.3 kcal/mol, consistent with its ready preparation and trapping. Heterocyclic allenes have also been isolated as small as eight-membered rings, with a mixed 'P' and 'B' heteroatom ylid.¹⁴ Therefore the postulated cyclic allene intermediate **C** (Scheme 5) is reasonable.

The cyclic allene could then undergo a ring-expansion reaction, to produce the benzo[b]indolo[1,2-h][1,7]naphthyridin-8-(13H)-one ring structure **D** (Scheme 5). The proposed driving force behind this ring-expansion is relief of ring strain of the 7-membered allenic ring - therefore, there is a favourable energy balance between the 7-membered allenic ring formation, and its subsequent role in providing a driving force for ring expansion. An additional crucial component of this step is the presence of an electrophile (E) - the major product arising from the reaction, **7**, requires $E = H^+$, whereas the minor product **8** requires $E = CO_2$, probably generated, on the basis of the results noted in Table 1, from carbonic acid decomposition, the acid in turn resulting ultimately from the Cs_2CO_3 base via bicarbonate (see below for a greater discussion). Once the carboxylate unit is incorporated, a further propargyl moiety could be added via nucleophilic displacement to produce the ester substituent of **8**. Subsequent dehydrogenation, followed by base-induced aromatisation allows for O-propargylation in a cascade process, yielding the final products **7** (**Path III**) and **8** (**Path IV**)(Scheme 5).

Scheme 5. Proposed mechanism for formation of **7** and **8**.

The addition of a 1-carbon unit is novel and imposes the question as to the origin of this carbon, even though **8** is isolated in very low yield. Two possibilities arise and involve either the well-known degradation of DMF¹⁵ to produce a "C=O" fragment that could be incorporated, or it could arise from the generated bicarbonate anion that is present in the solution. Table 1 summarizes experiments to determine the source of the additional carbon atom in **8**. Entry 1 is the standard reaction as previously outlined, whereas entry 2 describes replacing the DMF solvent with DMSO - this resulted in an increase from <1% to a 5% yield suggesting that DMF was not the source of the carboxylate of **8**. Bubbling CO₂ gas through a standard reaction (entry 3) resulted in a 6% yield of **8**, however, the most significant outcome from this reaction is the notably reduced yields of **4**, **5**, **6** and **7**, and a dramatic increase in the production of non-characterisable baseline material. This suggests that the presence of

significant quantities of electrophiles could be reacting with different indigo-based nucleophiles as they are being generated resulting in mixtures of products. Entry 4 describes the experiment replacing the Cs_2CO_3 with K_3PO_4 as base, to eliminate the presence of a bicarbonate source. However, the lack of solubility of this base in DMF is the likely reason for the outcome of mostly unreacted indigo being isolated from the reaction.

Table 1: Experiments to probe the source of the ester moiety **8**.

Entry	Solvent	Other Reaction	4	5	6	7	8	Baseline	Recovered
		Conditions	Yield	Yield	Yield	Yield	Yield	material	Indigo
			%	%	%	%	%	(%mass)	(%mass)
1	DMF	Cs ₂ CO ₃ , N ₂	11	21	17	31	1	11	-
2	DMSO	Cs ₂ CO ₃ , N ₂	-	-	12	13	5	65	-
3	DMF	Cs ₂ CO ₃ , CO ₂	13	-	7	10	6	61	-
4	DMF	K_3PO_4 , N_2	21	-	-	-	-	-	60
5	DMF	Cs ₂ CO ₃ , Ar	10	15	14	28	1	*	-

^{*} not isolated

Path V (Scheme 6) describes a possible mechanism to the allene 6 and diverges from the same intermediate **B** (red, Scheme 5). In this instance it is the iminium indigo moiety that activates the adjacent carbonyl, allowing sufficient electrophilicity to attract the relatively weak nucleophilic alkyne to undergo a cyclisation reaction, promoted by the initial attack of the other indigo nitrogen lone electron pair onto the propargyl C2 in a concerted process yielding the strained fused-aziridine **E**. Carbonate could then act as a nucleophile in a ring-opening of the aziridine and subsequent aromatisation of the central pyridinyl ring to give **F**. *O*-Propargylation of **F** could then afford intermediate **G**. The acidic proton in the CH₂OH group α to the iminium ion in **G** may then be removed under the influence of base and further OH proton loss would yield the aldehyde moiety in the

intermediate **H**. A Claisen rearrangement of the propargyloxy group with the indolic C2-C3 bond could then give rise to product **6**.

Scheme 6. Proposed mechanism for formation of **6**.

The outcomes from this propargylation reaction are repeatable and preliminary investigations also indicate reliable scalability up to at least double quantities. Further, our mechanistic proposals for the formation of $\bf 6 - 8$ all start from N,N-dipropargylated intermediates, rather than an intermediate that had cyclised initially from a N-monopropargylated molecule. In support of this proposal were the outcomes from an experiment where a DMF solution of $\bf 4$ was dripped into a mixture of Cs_2CO_3 and propargyl bromide in DMF over 3 mins before quenching after 2 mins. The result was formation of $\bf 5$ (<3%), $\bf 6$

(11%), **7** (25%) and **8** (<1%) with complete consumption of the starting material. The poor return of **5** with respect to **6** - **8** suggests that the second *N*-propargylation is a more competitive reaction than cyclisation, and lends support to our proposed dipropargylated compounds as intermediates.

It is also relevant to compare the mechanistic outcomes of this propargylation reaction with the outcomes of the corresponding allylation reaction (Scheme 1). Although cyclisation onto the indigo carbonyl of the unsaturated moiety occurred in both instances, reaction onto the indigo C2 position to form a spiro compound only occurs in the case of the alkene. There was no evidence suggesting an equivalent mechanism pathway in the presence of the alkyne. Presumably, the linear alkyne is not able to approach the indigo C2 position whereas the 'bent' nature of the alkene makes this cyclisation reasonable. Further, a significant by-product of the allylation reaction (Scheme 1) was *N*-allylisatin (structure not illustrated), derived by oxidative cleavage of the central indigo double bond. Under propargylation conditions, there was no *N*-propargylisatin present, as evidenced by TLC analysis of the reaction mixture against an authentic sample.

We have also undertaken some simple experiments to ascertain that the reactions are likely to proceed through nucleophilic mechanisms rather than through radical-based sequences. Previous studies have shown that radical reactions with indigo will proceed exceptionally slowly at room temperature and in 4 hours at 100 °C in the presence of oxygen and with irradiation. In contrast, we have repeated our propargylation reaction (as shown in Scheme 2) in the absence of oxygen and light (under argon) with these conditions realizing the same product outcome after 5 mins of reaction at 86 °C. Radical reactions are unlikely to proceed under these conditions, let alone to produce a total yield of 84% of products in 5 mins of reaction time.

The heterocycles **7** and **8** are analogs of the marine natural products fascaplysin and homofascaplysin (Figure 2), initially isolated from the sponge *Fascaplysinopsis reticulata* and noted for its ATP-competitive inhibitor of Cdk4/D1 (IC₅₀ = 0.35 μ M)^{3,17} activity. There have been reported syntheses of these natural products including the silver catalysed cascade synthesis of their parent compound and

analogs, with this approach involving the initial intermolecular reaction of two components of similar molecular complexity, however, this necessitated the prior synthesis of these two components. The Bring homolog of fascaplysin has also been synthesised starting from 4(1*H*-indol-1-yl)butanoic acid in a 2 step synthesis in a best yield of 42%. 19

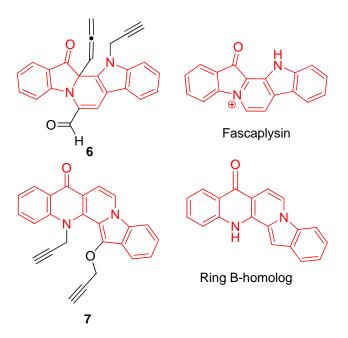


Figure 2: Representation of compounds **6** and **7** as analogs of fascaplysin/homofascaplysin and fascaplysin/homofascaplysin ring B-homolog respectively.

We report here a facile synthesis of the benzoindolo[1,2-h][1,7]naphthyridine (fascaplysin) heterocyclic skeleton from indigo in the presence of propargyl bromide and base in a 17% unoptimised yield. This provides direct access to this skeleton starting from exceptionally cheap starting materials and reagents, and provides a convenient access to this system for further elaboration in structure-activity studies.

The reaction of indigo with propargyl bromide in the presence of base produces new complex heterocycles *via* a multi-step, one-pot, cascade-type series of reactions. This is new science for the indigo moiety and represents an exciting untapped area of heterocyclic chemistry. Our proposed mechanisms (Schemes 4 - 6) reveal that at different stages of the cascade reaction, the reactivity of

different sites on the indigo moiety changes, and additionally, different types of reactivity can occur at different times, including sites acting as nucleophiles or electrophiles, with bond breakage leading to ring expansion and dehydration (Figure 3). A key element to the reactivity is the ability of the indigo heterocycle to tautomerise under different circumstances (*e.g.* upon generation of the N-centered anion with base) to shift reactivity to a different site. This constantly changing reactivity leads to the variety of outcomes as demonstrated. Further, this range of different chemistry all occurs in a tight cluster in the centre of the indigo structure and this combination of properties is the key to the variety of unusual heterocycles that arises. Further, the unexpected outcomes of this initial study not only shows us that there is highly unusual and significant chemistry of indigo, but that there is much yet to be explored in this area.

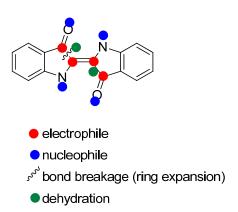


Figure 3: Graphical representation of the changing reactivity of the indigo skeleton with sites of nucleophilicity, electrophilicity, dehydration and bond breakage illustrated. The degree of reactivity at each of these points changes as the molecule(s) progress through the cascade reaction.

Biological Testing

As an initial screen for biological activity, compounds **4** to **7** were subjected to *in vitro* anti-plasmodial and anti-cancer testing (Table 2). All four compounds revealed notable anti-plasmodial²⁰ activity against a drug resistant strain in the micromolar range, sufficient for all to be considered lead compounds for further development.

Table 2. Anti-plasmodial and anti-cancer activity of the indigo-derived heterocyles 4 - 7.

		Plasmodium	NCI-H187	KB - Oral	Cytotoxicity
		falciparum#	Lung	cavity	Vero cell
Entry	Compound	Anti-plasmodial	Anti-cancer	Anti-cancer	
		$IC_{50} \mu g/mL (\mu M)$	$IC_{50} \mu g/mL$	$IC_{50} \mu g/mL$	$IC_{50} \mu g/mL$
			(μM)	(μM)	(μM)
1	4	0.33(1.1)	4.79 (15.9)	8.95 (29.8)	16.26 (54.2)
2	5	0.26 (0.85)	1.88 (6.24)	1.31 (4.36)	1.93 (6.43)
3	6	3.34 (8.9)	4.79 (12.7)	7.81 (20.7)	^
4	7	3.75 (9.9)	17.3 (46.0)	8.44 (2234)	16.4 (43.6)
5	Dihydroartemisinine /	0.24 / 0.02			
5	Mefloquine	0.24 / 0.03	-	-	-
6	Ellipticine /		1 21 / 0 100	1.04/0.674	
	Doxorubicin	-	1.21 / 0.108	1.04 / 0.674	-

[#] drug resistant K1 strain; ^ non-cytotoxic

More significantly, **4** (anti-plasmodial IC₅₀ 0.33 μ g/mL, 1.1 μ M) and **5** (anti-plasmodial IC₅₀ 0.256 μ g/mL, 0.85 μ M) are an order of magnitude more potent than compounds **6** and **7**. In the case of **4**, there was also a significant difference between the level of mammalian (Vero) cell toxicity (IC₅₀ 16.26 μ g/mL) and anti-plasmodial activity (Table 1). With respect to the activity against cancer cells²⁰, compounds **4 - 7** showed notable activity against the NCI-H187 lung cancer and KB-oral cavity cell lines. In particular, **7** was equipotent with the positive control ellipticine. Cytotoxicity²⁰ studies were particularly interesting for **5** and **6** with the latter being non-cytotoxic to normal mammalian cells but the former was toxic to these cells. Clearly, significant medicinal chemistry studies would need to be undertaken to decrease the toxicity to normal mammalian cells and increase the required selectivity.

However, as a random screening process, all these compounds showed activity that could be considered as new lead compounds for a variety of targets.

CONCLUSION

We have reported here new cascade reactions of indigo *via* base-mediated propargylation. The resulting shifting of reactivity during each step in the process generates, and then propagates, a cascade reaction, producing a range of poly-heterocyclic compounds in unprecedented outcomes. Some of these compounds also hold promise as leads for the development of new anti-malarial and anti-cancer agents. The scope for this reaction currently remains untapped, however, these initial studies suggest that the chemistry of indigo has significant potential for further development towards novel polycyclic compounds. The exploration of a variety of reaction types, inclusive of cascade processes, using indigo as a starting material remains an additional exciting area of synthetic chemistry to investigate. Interestingly, the possibility of using indirubin and isoindigo, as indigo analogs, in comparable cascade reactions remains unexplored and these starting materials also offer possibilities for the generation of new heterocycles in a one-pot process. These studies are currently underway in our laboratories.

EXPERIMENTAL SECTION

General Experimental Information

Reagents and solvents were purchased reagent grade and used without further purification unless otherwise stated. All reactions were performed in standard oven dried glassware under a nitrogen atmosphere unless otherwise stated. Melting points temperatures are expressed in degrees Celsius ($^{\circ}$ C) and are uncorrected. 1 H and 13 C NMR spectra (CHCl₃) solutions) were recorded at 500 and 125 MHz with chemical shifts (δ) reported in parts per million relative to TMS (δ = 0 ppm) or CDCl₃ (δ = 77.0 ppm) as internal standards. Coupling constants (J) are reported in Hertz (Hz). Multiplicities are reported as singlet (s), broad singlet (bs), doublet of doublets (dd) or multiplet (m). Electron impact (EI) mass spectra (MS) and electrospray (ESI - single quadrupole) mass spectra have their ion mass to

charge (m/z) values stated with their relative abundances as a percentage in parentheses. Peaks assigned to the molecular ion are denoted by M^+ or M+1. Infrared (IR) spectra were recorded on neat samples. UV-Visible spectra were recorded with solutions of samples in CH_2Cl_2 . Images from crystals were captured using stereo microscopes. All the images were provided from X-ray quality single crystals. Optical rotations were measured in CH_2Cl_2 solution at 25 °C. Thin Layer Chromatography (TLC) was performed using Silica Gel F254 aluminium sheets. Column chromatography was performed under gravity using Silica Gel 60 (0.063-0.200 mm). Eluents are in volume to volume (v:v) proportions. Solvent extracts or chromatographic fractions were concentrated by rotary evaporation *in vacuo*. Indigo (dye content 95%) was used without further purification. Petroleum spirit had a b.p. range of 40-60 °C.

Products from the Propargylation of Indigo

A suspension of powdered indigo (250 mg, 0.95 mmol) in anhydrous DMF (40 mL) was sonicated for 30 min and stirred vigorously under N_2 overnight. The resulting suspension was added to pre-dried anhydrous cesium carbonate (1.20 g, 3.71 mmol) and molecular sieves (4 Å) while being stirred and warmed to 80-85 °C under a N_2 atmosphere. After 30 mins propargyl bromide (0.595 mg 5.0 mmol) was added and the reaction mixture was heated at 82-85 °C for 5 min. The mixture was then filtered hot and then the solution was concentrated by rotary evaporation and the residue applied to a short plug of silica gel/celite (1:1) and washed consecutively with 4 different solvent mixtures, 1: 70:30 CH₂Cl₂/petroleum spirit (250 mL), 2: CH₂Cl₂, 3: 50:50 CH₂Cl₂/EtOAc and 4: 95:5 EtOAc/MeOH (250 mL). Four fractions were collected from the four different elutions. **Fraction 1** was concentrated and slowly recrystallised in 9:1 petroleum spirit/EtOAc then filtered to yield **1-(prop-2-yn-1-yl)-[2,2'-biindolinylidene]-3,3'-dione 4** (33.00 mg 11%) as a blue, papery solid. R_f (7:3 CH₂Cl₂/petroleum spirit) = 0.53, m.p: 267-269 °C; λ_{max} /nm (ε , M⁻¹cm⁻¹) 291 (10447), 634 (6381). IR (neat) ν_{max} 3278 (m), 1605 (s), 1463 (s), 1297 (s), 1066 (s), 1027 (s), 927 (m), 743 (m) cm⁻¹. ¹H NMR δ 2.17 (1H, s, H3"), 5.41 (1H, s, H1"), 6.96 (1H, t, J = 7.1 Hz, H5'), 6.99 (1H, d, J = 8.1 Hz, H7'), 7.09 (1H, t, J = 7.5 Hz, H6'), 7.69 (1H, t)

d, J = 7.5 Hz, H4'), 7.77 (1H, d, J = 7.5 Hz, H4) 10.60 (1H, s, H1). ¹³C NMR (CDCl₃) δ 37.1 (C1"), 72.5 (C3"), 78.4 (C2"), 111.5 (C7'), 111.9 (C7), 120.4 (C3'a), 120.9 (C5'), 121.5 (C5), 122.0 (C2'), 122.4 (C2), 125.0 (C4'), 125.1 (C4), 126.2 (C3a), 135.9 (C6), 136.4 (C6), 151.8 (C7'a), 152.6 (C7a), 187.6 (C3), 189.6 (C3'). MS (EI), m/z 300 (100%, M⁺), 271 (53), 262 (32). HRMS (ESI) [M+H]⁺ calcd for $C_{19}H_{13}N_2O_2$ 301.0977, found 301.0964.

The mother liquors from **fraction 1** were combined with **fraction 2** and then subjected to a silica gel column and elution with 7:3 CH₂Cl₂/petroleum spirit gave **13-oxo-12-(prop-2-yn-1-yl)-12b** (**propa1,2-dien-1-yl)-12b,13-dihydro-12***H***-pyrido[1,2-***a***:3,4-***b***']diindole-6-carbaldehyde 6** as yellow-orange crystals (63.92 mg, 17%). X-ray quality crystals were grown through slow crystallization from petroleum spirit/EtOAc (5:3). R_f (CH₂Cl₂) = 0.26, m.p. 229-231 °C; UV-Vis (CH₂Cl₂) λ_{max} /nm (ϵ , M⁻¹cm⁻¹) 386 (12825), 244 (34151). IR (neat) ν_{max} 3278 (m), 1604 (s), 1463 (s), 1297 (m), 1065 (s), 1027 (s), 1015 (s), 743 (s) cm⁻¹. ¹H NMR δ 2.33 (1H, s, H3'), 4.79 (2H, d, J = 6.5 Hz, H3"), 5.42 (1H, t, J = 6.5 Hz, H1"), 5.51-5.66 (2H, m, H1'), 6.82 (1H, d, J = 8.4 Hz, H11), 6.94 (1H, t, J = 7.2 Hz, H3), 7.27 (1H, t, J = 7.6 Hz, H2), 7.35 (1H, t, J = 8.0 Hz, H9), 7.49 (1H, t, J = 8.0 Hz, H8), 7.54 (1H, d, J = 8.4 Hz, H10), 7.64-7.68 (2H, m, H1, H4), 7.65 (1H, s, H7), 9.62 (1H, s, 1"'). 13°C NMR (CDCl₃) δ 36.2 (C1'), 71.3 (C12b), 73.6 (C3'), 77.8 (C2'), 80.4 (C3"), 90.4 (C1"), 110.7 (C7a), 111.1 (C13a), 111.5 (C9), 113.6 (C1), 117.2 (C12a), 118.6 (C11), 121.1 (C3), 122.6 (C8), 124.1 (C10), 124.5 (C11a), 124.9 (C4), 131.8 (C7), 137.7 (C7b), 138.2 (C4a), 138.5 (C2), 158.6 (C6), 185.3 (C1"'), 195.7 (C13), 208.4 (C2"). MS (EI) m/z 376 (5, M⁺), 371 (100%), 298 (24). HRMS (ESI) [M+H1⁺ calcd for C₂₅H₁₇N₂O₂ 377.1298, found 377.1285.

Fraction 3 was recrystallised from CH₂Cl₂/petroleum spirit (1:9) giving 13-(prop-2-yn-1-yl)-14-(prop-2-yn-1-yloxy)benzo[b]indolo[1,2-h][1,7]naphthyridin-8-(13H)-one 7 (116.6 mg, 31%) as orange crystals. X-ray quality crystals were grown through slow crystallization from petroleum spirit : ethyl acetate (4:3). R_f (9:1 CH₂Cl₂/EtOAc) = 0.52, m.p: 218-220 °C; UV-Vis (CH₂Cl₂) λ_{max} /nm (ε , M⁻¹cm⁻¹) 324 (27663), 441 (10360). IR (neat) ν_{max} 3205 (w), 1588 (m), 1485 (m), 1349 (m), 1260 (m),

1059 (m), 725 (s) cm⁻¹. ¹H NMR δ 2.14 (1H, t, J = 2.4 Hz, H3'), 2.30 (1H, t, J = 2.4 Hz, H3"), 4.74 (2H, d, J = 2.4 Hz, H1'), 5.49 (2H, d, J = 2.4 Hz, H1"), 7.33 (1H, d, J = 7.4 Hz, H7), 7.45 (3H, m, H1, H2, H3), 7.74-7.77 (1H, t, J = 7.8 Hz, H11), 7.85-7.90 (3H, m, H4, H10, H12), 8.01 (1H, d, J = 7.4 Hz, H6), 8.71 (1H, dd, J = 8.1, 1.4 Hz, H9). ¹³C NMR (CDCl₃) δ 44.0 (C1"), 63.7 (C1'), 74.5 (C3'), 76.5 (C3"), 78.5 (C2"), 78.6 (C2'), 104.0 (C7), 110.9 (C3), 117.9 (C7a), 118.6 (C12), 118.9 (C13b), 119.1 (C10), 119.9 (C6), 121.6 (C4a), 123.3 (C2), 123.9 (2C, C1, C4), 126.4 (C8a), 126.9 (C9), 129.3 (C14a), 132.6 (C11), 133.2 (C14), 139.1 (C13a), 143.5 (C12a), 175.8 (C8). MS (EI), m/z 376 (6, M⁺), 337 (100%), 309 (20), 298 (75). HRMS (ESI) [M+H]⁺ calcd for C₂₅H₁₇N₂O₂ 377.1285, found 377.1302.

The mother liquor from the recrystallisation of **7** was concentrated and then subjected to silica gel column chromatography and elution with CH₂Cl₂/EtOAc (92:8) resulted in **prop-2-yn-1-yl 8-oxo-13-(prop-2-yn-1-yl)-14-(prop-2-yn-1-yloxy)-8,13-dihydrobenzo[***b***]indolo[1,2***h***][1,7]naphthyridine-7-carboxylate 8** (2 mg, >1%) as bright orange crystals. X-ray quality crystals were grown through slow crystallization from chloroform. R_f (9:1, CH₂Cl₂/EtOAc) = 0.59, m.p.: 258-260 °C; IR (neat) v_{max} 3278 (m), 2925 (m), 1718 (s), 1595 (s), 1482 (m), 1237 (m), 1062 (m), 964 (m), 749 (s), 643 (s) cm⁻¹. ¹H NMR δ 2.14 (1H, bs, H3"), 2.30 (1H, bs, H3"), 2.53 (1H, bs, H3'), 4.76 (2H, d, J = 2.0 Hz, H1"), 5.06 (2H, d, J = 1.7 Hz, H1"), 5.43 (2H, d, J = 1.7 Hz, H1'), 7.43-7.50 (3H, m, H1, H2, H3), 7.75 (1H, t, J = 7.8 Hz, H11), 7.82-7.90 (3H, m, H4, H10, H12), 8.01 (1H, s, H6), 8.48-8.50 (1H, d, J = 7.8, H9). ¹³C NMR (CDCl₃) δ 44.1 (C1"), 53.4 (C1'), 63.5 (C1"), 74.7 (C3"'), 75.1 (C3'), 76.5 (C3"'), 77.9 (C2"'), 78.1 (C2'), 78.2 (C2"''), 110.7 (C7a), 110.8 (C7), 112.2 (C3), 115.1 (C13b), 118.1 (C12), 118.7 (C10), 118.8 (C6), 122.0 (C4a), 123.8 (C2), 124.1 (C1), 124.7 (C4), 126.7 (C8a), 126.8 (C9), 130.0 (C14a), 133.6 (C11), 134.5 (C14), 139.2 (C13a), 142.9 (C12a), 167.1 (ester C=O) 174.5 (C8). MS (EI), m/z 458 (5, M+), 419 (100%), 380 (10), 375 (5), 337 (20), 298 (45). HRMS (ESI) [M+H]⁺ calcd for C₂₉H₁₉N₂O₄ 459.1345, found 459.1363.

Fraction 4 was purified by preparative TLC using CH₂Cl₂/EtOAc (88:12) as the developing solvent and gave 6-methylene-6,7-dihydropyrazino[1,2-a:4,3-a']diindole-13,14-dione 5 was isolated as a dark burgundy powder (63 mg, 21%. R_f (8.5:1.5, CH₂Cl₂/EtOAc) = 0.51, m.p: 280-284 °C. UV-Vis (CH₂Cl₂) λ_{max} /nm (ε, M⁻¹cm⁻¹) 324 (13088), 573 (6430). IR (neat) ν_{max} 1701 (m), 1604 (m), 1470 (m), 1298 (m), 1185 (m), 1122 (s), 742 (s) cm⁻¹. ¹H NMR δ 4.41 (2H, s, H7), 5.04 (1H, H1'a), 5.38 (1H, H1'b), 6.95-7.00 (2H, m, H9, H11), 7.09 (1H, t, J = 7.4 Hz, H2), 7.48-7.55 (3H, m, H3, H4, H10), 7.73 (1H, d, J = 7.5 Hz, H12), 7.82 (1H, d, J = 7.5 Hz, H1). ¹³C NMR (CDCl₃) δ 45.9 (C7), 97.9 (C1'), 109.5 (C12), 112.7 (C4), 121.4 (C13a), 121.8 (C11), 122.7 (C2), 123.0 (C12a), 124.6 (C14a), 125.3 (C1), 125.5 (C6), 131.6 (C13b), 135.1 (C10), 135.5 (C3), 147.0 (C4a), 150.0 (C8a), 179.7 (C14), 180.8 (C13). MS (EI), m/z 300 (14), 207 (100%). HRMS (ESI) [M+H]⁺ calcd for C₁₉H₁₃N₂O₂ 301.0972, found 301.0960.

1-(Prop-2-yn-1-yl)-[2,2'-biindolinylidene]-3,3'-dione (4)

A suspension of powdered indigo (500 mg, 1.90 mmol) in anhydrous DMF (50 mL) was sonicated for 60 min and stirred vigorously under N₂ overnight. The resulting suspension was added to pre-dried anhydrous cesium carbonate (2.4 g, 7.42 mmol) and the mixture was stirred and warmed to 80-85 °C under a N₂ atmosphere. After 30 mins propargyl bromide (1.90 mg 10.0 mmol) was added and the reaction mixture was heated at 82-85 °C for 5 seconds. The mixture was then poured into ice water and the resulting precipitate was filtered and recrystallised from petroleum spirit/EtOAc (90:10) to furnish 1-(prop-2-yn-1-yl)-[2,2'-biindolinylidene]-3,3'-dione 4 (279.00 mg 93%) as a blue fluffy solid.

6-Methylene-6,7-dihydropyrazino[1,2-a:4,3-a']diindole-13,14-dione (5)

A solution of 6-methylene-6,7-dihydropyrazino[1,2-a:4,3-a']diindole-13,14-dione **4** (100 mg, 0.33 mmol) in anhydrous DMF (20 mL) was stirred and warmed to 80-85 °C under a N₂ atmosphere for 20 min and the solution was then added to pre-dried anhydrous cesium carbonate (107 mg, 0.33 mmol) and was stirred and warmed at 80-85 °C under a N₂ atmosphere for 10 min. The mixture was then poured into ice water and the resulting precipitate separated and subjected to silica gel short column

chromatography of and eluted with CH₂Cl₂/EtOAc (85:15) to give 6-methylene-6,7-dihydropyrazino[1,2-a:4,3-a]diindole-13,14-dione **5** as a dark burgundy powder (98 mg, 98%).

Preparation of *N*-Propargylisatin

To a solution of isatin (147 mg, 1.00 mmol) in dry DMF (40 mL) was added cesium carbonate (650 mg, 2.00 mmol). The resulting brown suspension was stirred at 80-85 °C for 30 min and then propargyl bromide (119 mg, 1.00 mmol) was added under a N₂ atmosphere. The resulting mixture was stirred at 80-85 °C for 30 min, poured into ice water and the suspension partitioned between CH₂Cl₂ (20 mL) and water (20 mL). The aqueous layer was washed with CH₂Cl₂ (4×5 mL), and the combined organic layers washed with water (3×10 mL), dried (MgSO₄) and concentrated. The residue was recrystallized from chloroform/hexane (1:6) to give *N*-propargylisatin (133 mg, 91%) as an orange solid. ¹H NMR (CDCl₃) δ 2.32 (2H, d, J = 1.7 Hz, H3'), 4.54 (2H, d, J = 1.9, H1'), 7.16 (2H, m, H5, H7), 7.66 (2H, m, H4, H6). ¹³C NMR (CDCl₃) δ 29.8 (C3'), 73.7 (C2'), 76.0 (C1'), 111.4 (C7), 118.0 (C3a), 124.5 (C5), 125.8 (C4), 138.8 (C6), 149.9 (C7a), 157.5 (C2), 182.9 (C3). MS (EI) m/z 185 (85%, M⁺), 129 (100%) consistent with literature values. ²¹

Antiplasmodial Assay

The compounds and extracts were tested *in vitro* against *Plasmodium falciparum*, K1CB1 (K1), which is a multidrug resistant (chloroquine and antifolate resistant) strain, received as a generous gift from Professor Sodsri Thaithong, Chulalongkorn University, Bangkok, Thailand. The parasites were maintained in human red-blood cells in RPMI 1640 medium supplemented with 25 mM HEPES, 0.2% sodium bicarbonate and 8% human serum, at 37 °C in a 3% carbon dioxide gas incubator (Trager and Jensen, 1976). Samples were made up in DMSO solution and the *in vitro* antimalarial activity testing was carried out using the microdilution radioisotope technique. The test sample (25 μL, in the culture medium) was placed in triplicate in a 96-well plate where parasitized erythrocytes (200 μL) with a cell suspension (1.5%) of parasitemia (0.5–1%) were then added to the wells. The ranges of the final

concentrations of the samples were varied from 2×10^{-5} to 1×10^{-7} M with 0.1% of the organic solvent. The plates were then cultured under standard conditions for 24 h after which 3 H-hypoxanthine (25 μ L, 0.5 mCi) was added. The culture was incubated for 18–20 h after which the DNA from the parasite was harvested from the culture onto glass fibre filters and a liquid scintillation counter used to determine the amount of 3 H-hypoxanthine incorporation. The inhibitory concentration of the sample was determined from its dose-response curves or by calculation.

Cancer Growth Inhibition and Vero Cell Toxicity Assay

Cancer growth inhibition assay and the *Vero* cell assay were performed using the Resazurin microplate assay (REMA) Method as described by O'Brien *et al.*²³ In brief, cells at a logarithmic growth phase were harvested and diluted to 2.2×104 cells/mL for KB and 3.3×104 cells/mL for NCI-H187, in fresh medium. Successively, 5 μ l of test sample diluted in 5% DMSO, and 45 μ l of cell suspension were added to 384-well plates, incubated at 37 °C in 5% CO₂ incubator. After the incubation period (3 days for KB, and 5 days for NCI-H187), 12.5μ l of 62.5μ g/mL resazurin solution was added to each and the plates were then incubated at 37 °C for 4 hours. Fluorescence signal was measured using SpectraMax M5 multi-detection microplate reader (Molecular Devices, USA) at the excitation and emission wavelengths of 530 nm and 590 nm. Percent inhibition of cell growth was calculated by the following equation: % Inhibition = $[1-(FU_T/FU_C)] \times 100$ whereas FU_T and FU_C are the mean fluorescent unit from treated and untreated conditions, respectively. Dose response curves were plotted from 6 concentrations of 3-fold serially diluted test compounds and the sample concentrations that inhibit cell growth by 50% (IC_{50}) been derived using the SOFTMax Pro software (Molecular Devices, USA). Ellipticine and doxorubicin were used as a positive control, and 0.5% DMSO and water were used as a negative control.²³

ASSOCIATED CONTENTS

Supporting Information

Copies of ¹H and ¹³C NMR for compounds **4 - 8** and UV-vis spectra for compounds **4 - 7**, ORTEP plots and CIF files for **6**, **7**, and **8**, images of fluorescence emission for **7** and **8**, and a computed model for **5**. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

Paul A. Keller, Tele: +61 2 4221 4692, Fax: +61 2 4221 4287, email: keller@uow.edu.au

Notes

The authors declare no competing financial interests.

ACKNOWLEDGMENTS

Financial support from the University of Wollongong, through the Centre for Medicinal Chemistry and a UPA scholarship to A.S. is gratefully acknowledged. We also thank Dr. Solomon Beckman for his help in the preparation of microscopic images of the crystals, and the Australian National University and BIOTEC for their support.

REFERENCES AND FOOTNOTES

- (1) (a) Anderson, E. A. *Org. Biomol. Chem.* **2011**, *9*, 3997-4006; (b) Shao, Z.; Peng, F. Z., *Curr. Org. Chem.* **2011**, *15*, 4144-4160.
- (2) Dančík, V.; Seiler, K. P.; Young, D. W.; Schreiber, S. L.; Clemons, P. A. J. Am. Chem. Soc. 2010, 132, 9259-9261.
- (3) Liu, W.; Khedkar, V.; Baskar, B.; Schürmann, M.; Kumar, K. *Angew. Chem. Int. Ed.* **2011**, *50*, 6900-6905.
- (4) (a) Gribble, G. W.; Pelcman, B. *J. Org. Chem.* **1992**, *57*, 3636-3642; (b) Carter, D. S.; Vranken, D. L. V. *J. Org. Chem.* **1999**, *64*, 8537-8545; (c) Segraves, N. L.; Robinson, S. J.; Garcia, D.; Said, S. A.; Fu, X.; Schmitz, F. J.; Pietraszkiewicz, H.; Valeriote, F. A.; Crews, P. *J. Nat. Prod.* **2004**, *67*, 783-792; (d) Dubovitskii, S. V. *Tetrahedron Lett.* **1996**, *37*, 5207-5208.

- (5) Bergman, J.; Koch, E.; Pelcman, B. *Tetrahedron* **1995**, *51*, 5631-5642.
- (6) Sasaki, T.; Ohtani, I. I.; Tanaka, J.; Higa, T. Tetrahedron Lett. 1999, 40, 303-306.
- (7) Lawrie, A. M.; Noble, M. E. M.; Tunnah, P.; Brown, N. R.; Johnson, L. N.; Endicott, J. A. *Nat. Struct. Mol. Biol.* **1997**, *4*, 796-801.
- (8) Bush, J. A.; Long, B. H.; Catino, J. J.; Bradner, W. T.; Tomita, K. J. Antibiot. 1987, 40, 668-678.
- (9) Abdel-Hamid, M. K.; Bremner, J. B.; J. B.; Coates, J.; Keller, P. A.; Miländer, C.; Torkamani, Y. S.; Skelton, B. W.; White, A. H.; Willis, A. C. *Tetrahedron Lett.* **2009**, 50, 6947-6950.
- (10) See supplementary information, Figure S5 for illustrations of conformation.
- (11) Johnson, R. P. Chem. Rev. 1989, 89, 1111-1124.
- (12) Krause, A. N.; Hashmi, S. K. *Modern Allene Chemistry*. 1st ed.; Wiley-VCH: Weinheim, **2004**; Vol. 1.
- (13) Daoust, K. J.; Hernandez, S. M.; Konrad, K. M.; Mackie, I. D.; Winstanley, J.; Johnson, R. P. J. Org. Chem. **2006**, 71, 5708-5714.
- (14) Moemming, C. M.; Kehr, G.; Wibbeling, B.; Froehlich, R.; Schirmer, B.; Grimme, S.; Erker, G. Angew. Chem. Int. Ed. 2010, 49, 2414-2417.
- (15) Muzart, J. Tetrahedron 2009, 65, 8313-8323.
- (16) (a) Smith, B. D.; Alonso, D.; Bien, J. T.; Zielinski, J.; Smith, S. L.; Haller, K. J. J. Org. Chem.,
 1993, 58, 6493-6496. (b) Smith, B. D.; Haller, K. J.; Shang, M. J. Org. Chem., 1993, 58, 6905-6908 (c)
 Smith, B. D.; Alonso, D.; Bien, J. T.; Metzler, E. C.; Shang, M.; Roosenburg, J. M. J. Org. Chem. 1994,
 59, 8011-8014.
- (17) Yan, Xiaojun; Chen, Haimin; Lu, Xiaoling; Wang, Feng; Xu, Weifeng; Jin, Haixiao; Zhu, Peng. Eur. J. Pharm. Sci. 2011, 42, 251-259
- (18) Waldmann, H.; Eberhardt, L.; Wittstein, K.; Kumar, K. Chem. Commun. 2010, 7, 4622-4624.
- (19) Baranova, O. V.; Zhidkov, M. E.; Dubovitskii, S. V. Tetrahedron Lett. 2011, 52, 2397-2398.

- (20) (a) Changsen, C.; Franzblau, S. G.; Palittapongarnpim, P. Antimicrob. Agents Chemother. 2003, 47, 3682-3687. (b) Desjardins, R. E.; Canfield, C. J.; Haynes, J. D.; Chulay, J. D. Antimicrob. Agents Chemother. 1979, 16, 710–718. (c) O'Brien, J.; Wilson, I.; Orton, T.; Pognan, F. Eur. J. Biochem. 2000, 267, 5421-5426.
- (21) Radul, O. M.; Zhungietu, G. I.; Rekhter, M. A.; Bukhanyuk, S. M. *Khimiya Geterotsiklicheskikh Soedinenii*, **1983**, 3, 353-355.
- (22) (a) Changsen, C.; Franzblau, S. G..; Palittapongarnpim, P. Antimicrob. Agents Chemother. **2003**, 47, 3682–3687. (b) Desjardins, R. E.; Canfield, C. J.; Haynes, J. D.; Chulay, J. D. Antimicrob. Agents Chemother. **1979**, 16, 710-718.
- (23) O'Brien, J.; Wilson, I.; Orton, T.; Pognan, F. Eur. J. Biochem. 2000, 267, 5421-5426.

Rapid Cascade Synthesis of Poly-Heterocyclic Architectures from Indigo

Alireza Shakoori¹, John B. Bremner¹, Anthony C. Willis², Rachada Haritakun³ and Paul A. Keller¹*

Supporting Information

Table of Contents

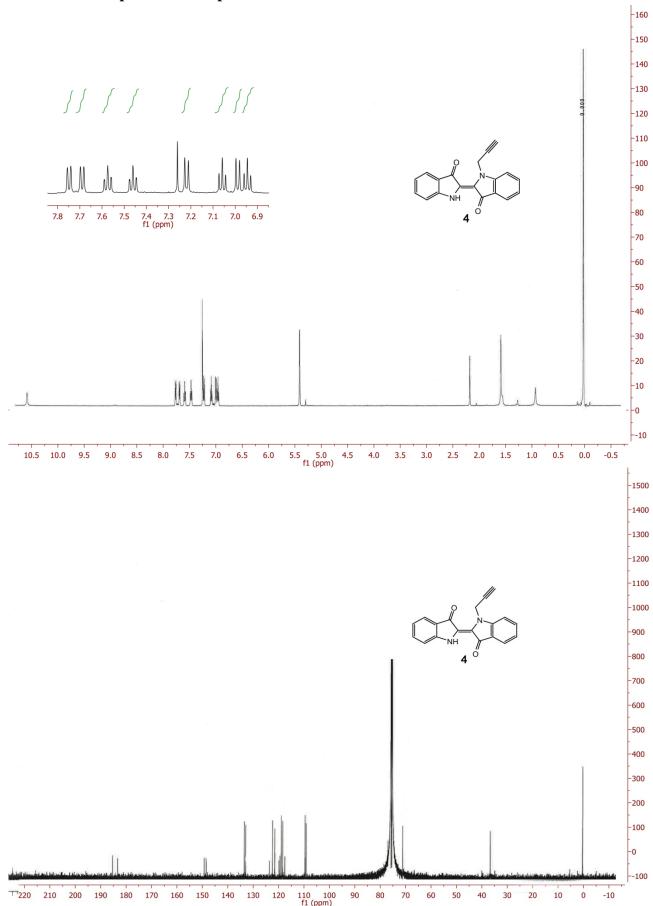
Selected NMR Spectra of Compounds 4-8	S2
Selected UV-Vis. Spectra of Compounds 4-7	S13
ORTEP Plots for the Three X-ray Structures Reported in this Manuscript (larger scale)	S15
Figure S1 and S2.Figure S3.	
Supplementary Pictures and Figures	S18
 Figure S4: The Fluorescence Emission of Compounds 7 and 8 Figure S5:Computational Modeling for Compound 5 with Spartan 10, Geometry Optimi 	

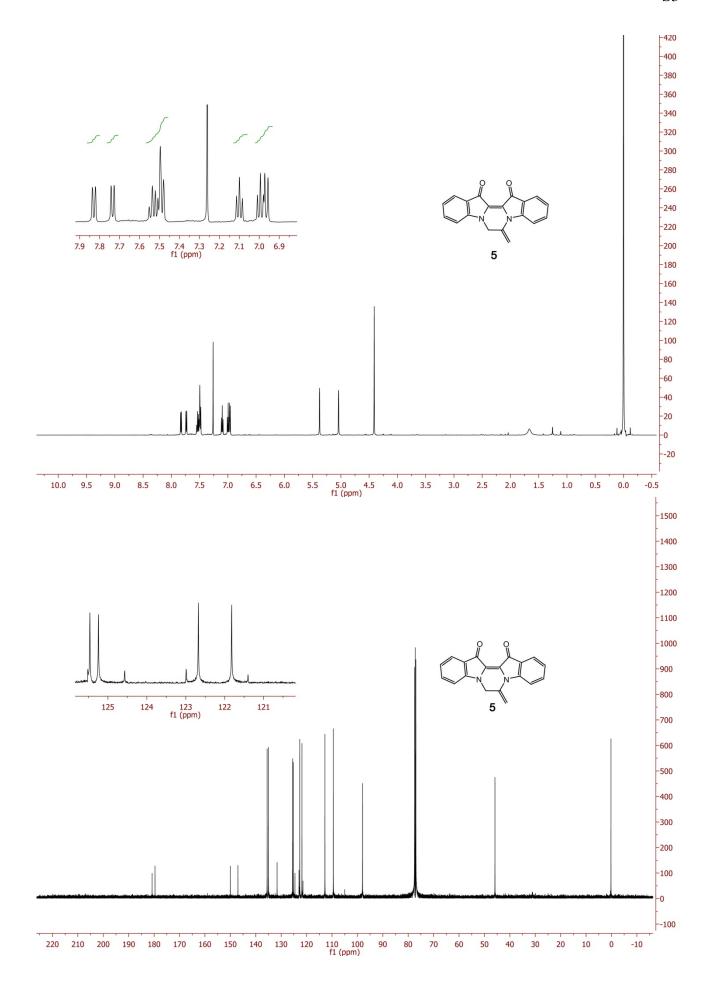
¹ School of Chemistry, University of Wollongong, Wollongong, NSW, 2522, Australia

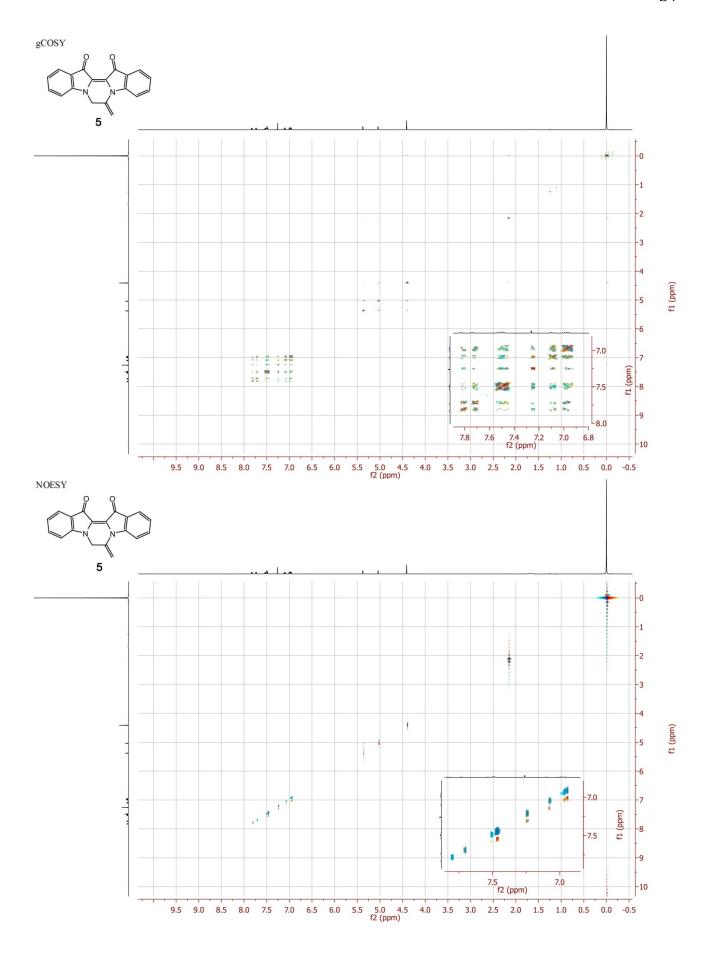
² School of Chemistry, The Australian National University, Canberra, ACT 0200, Australia

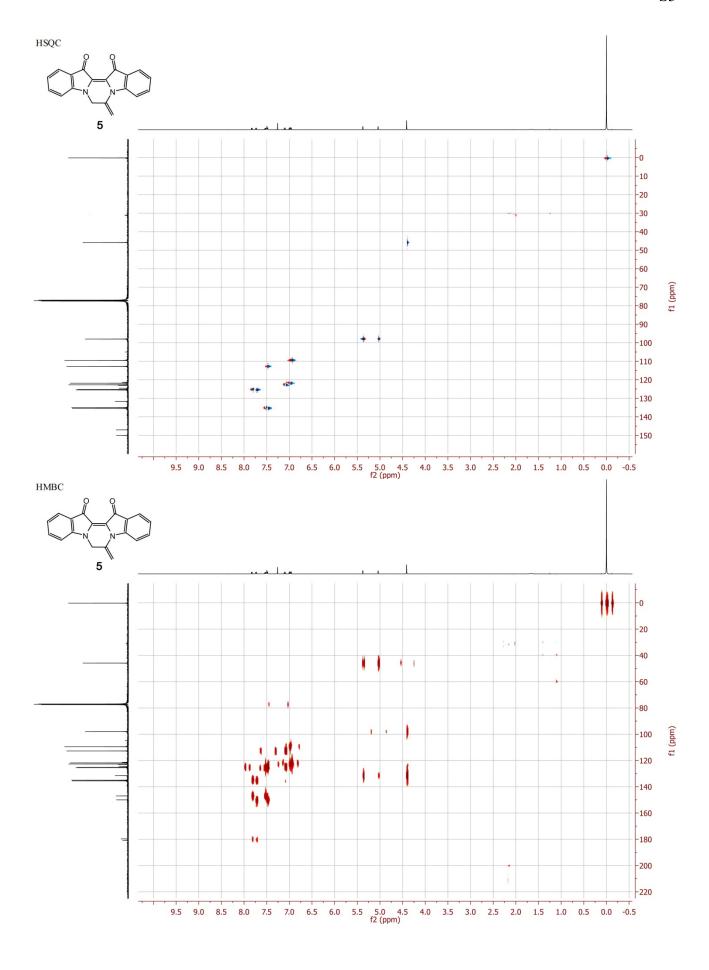
³ National Centre for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency (NSTDA), 113 Paholyothin Road, Klong 1, Klong Laung, Pahmanthani 12120 Thailand

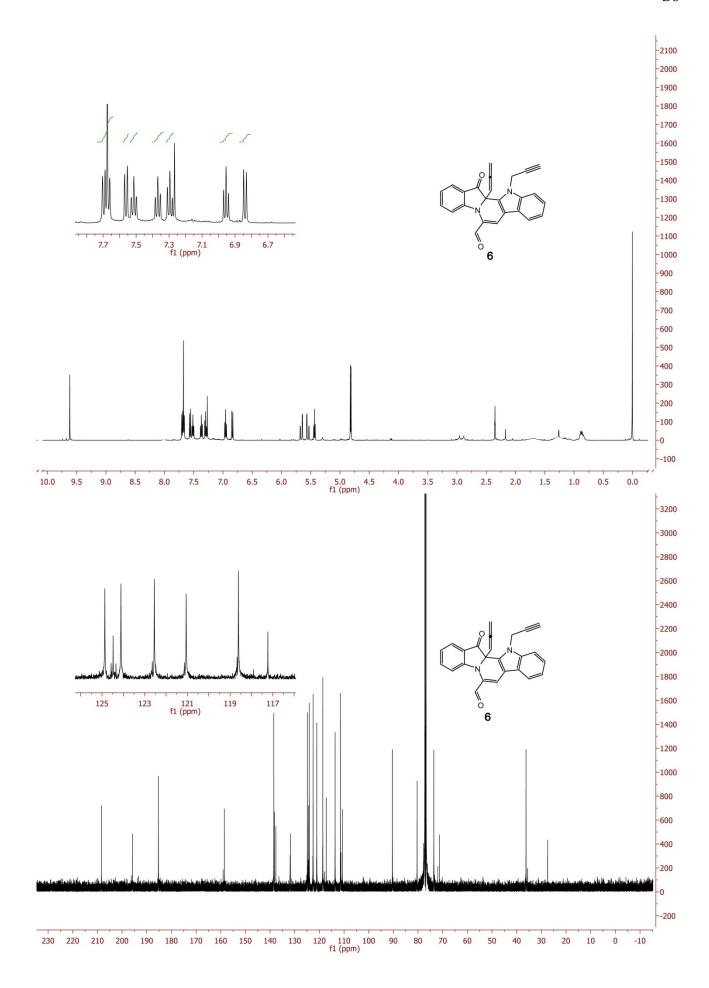
Selected NMR Spectra of Compound 4-8

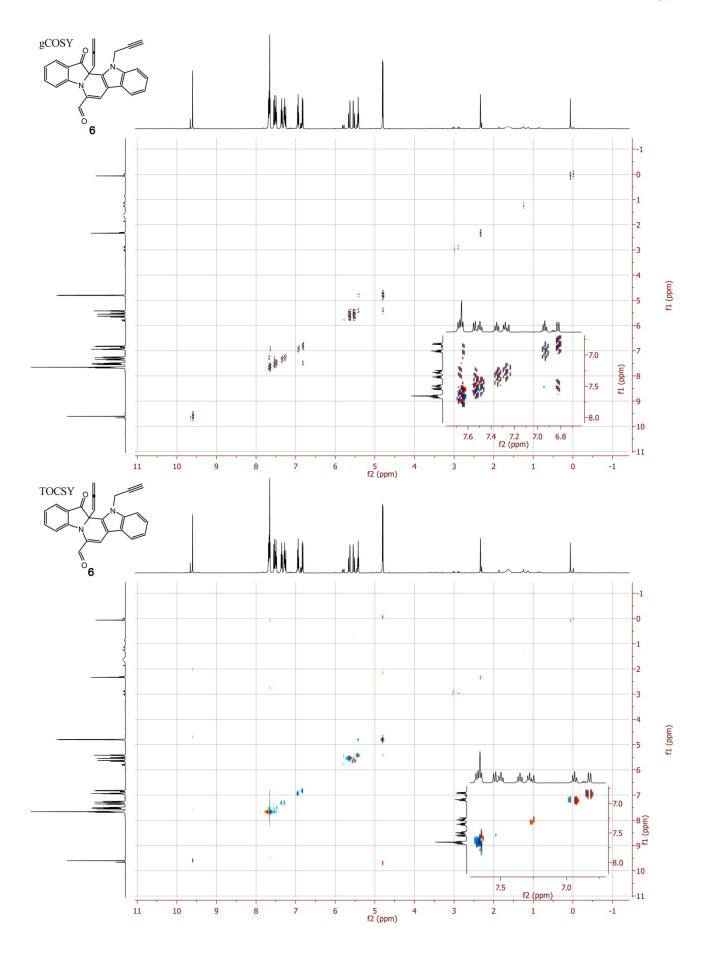


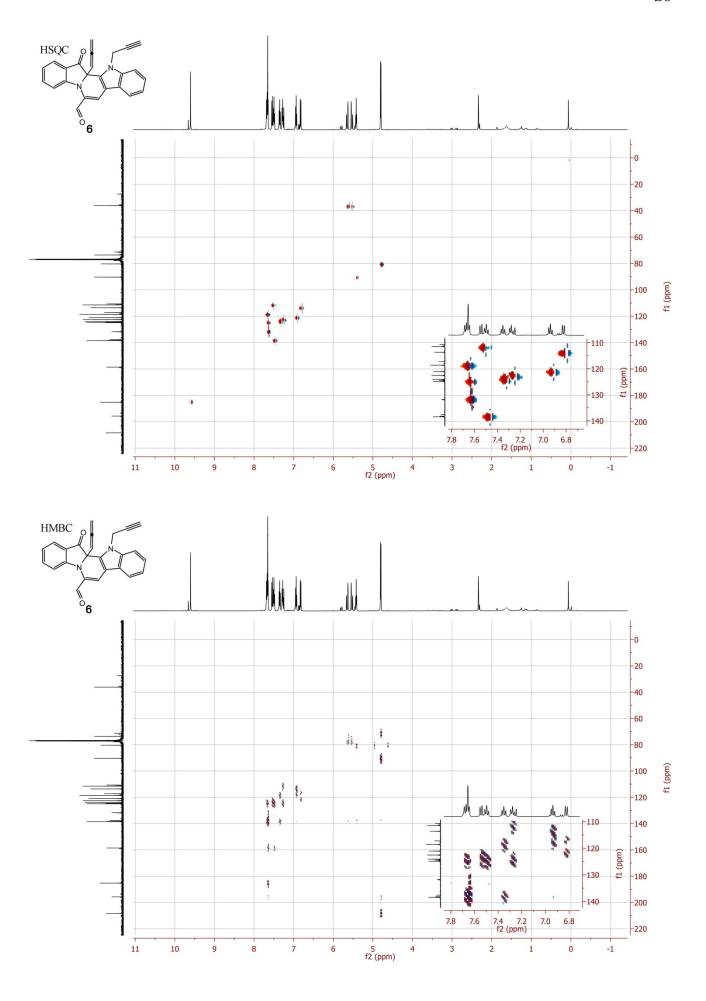


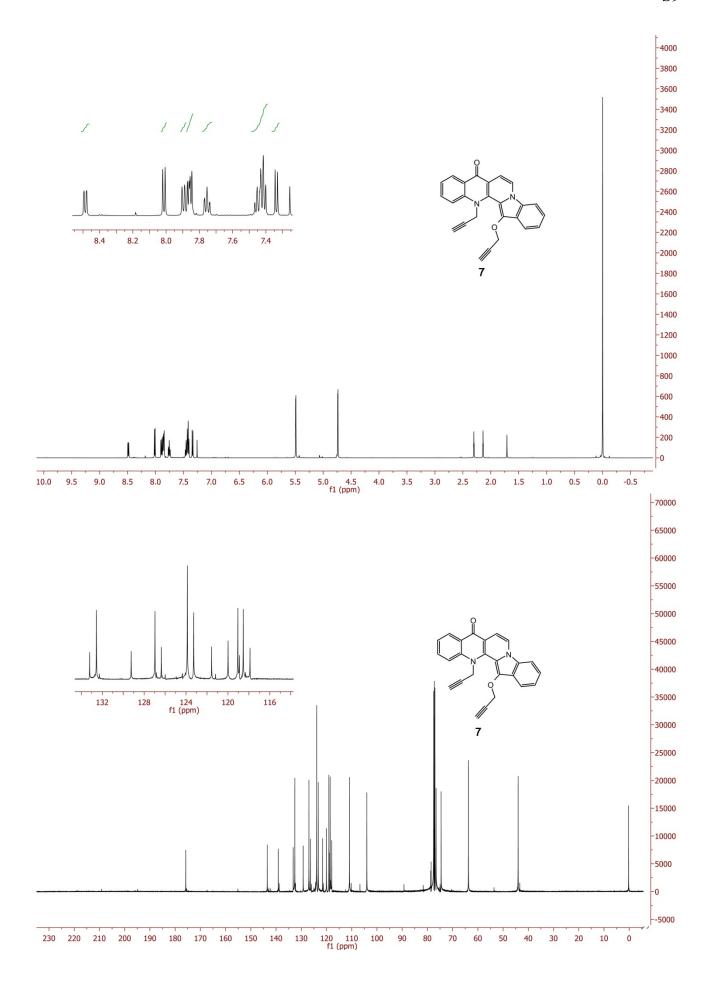


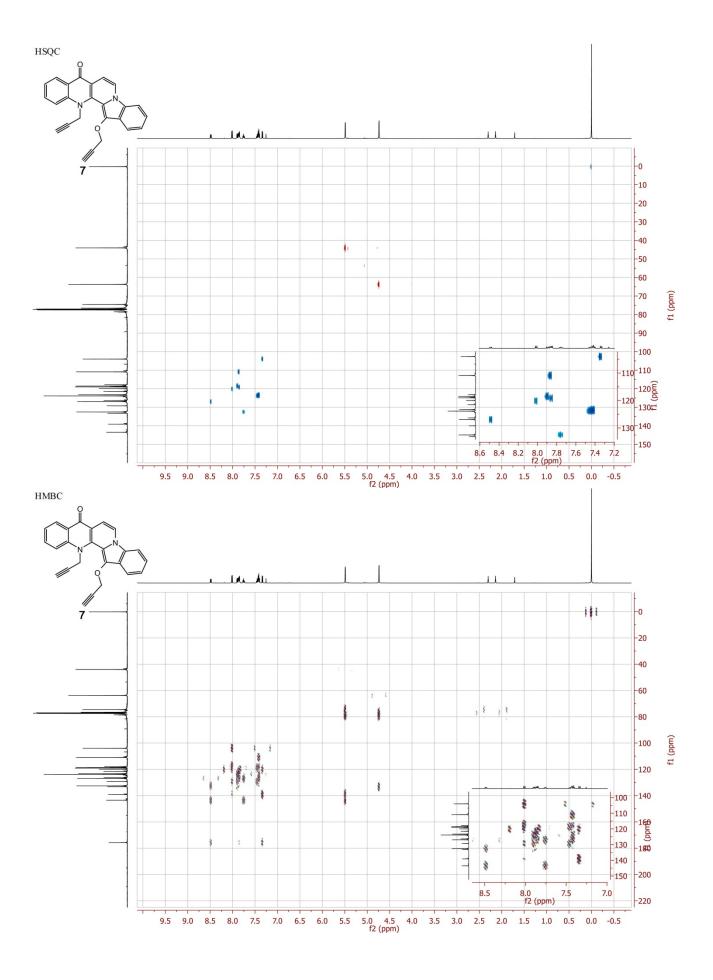


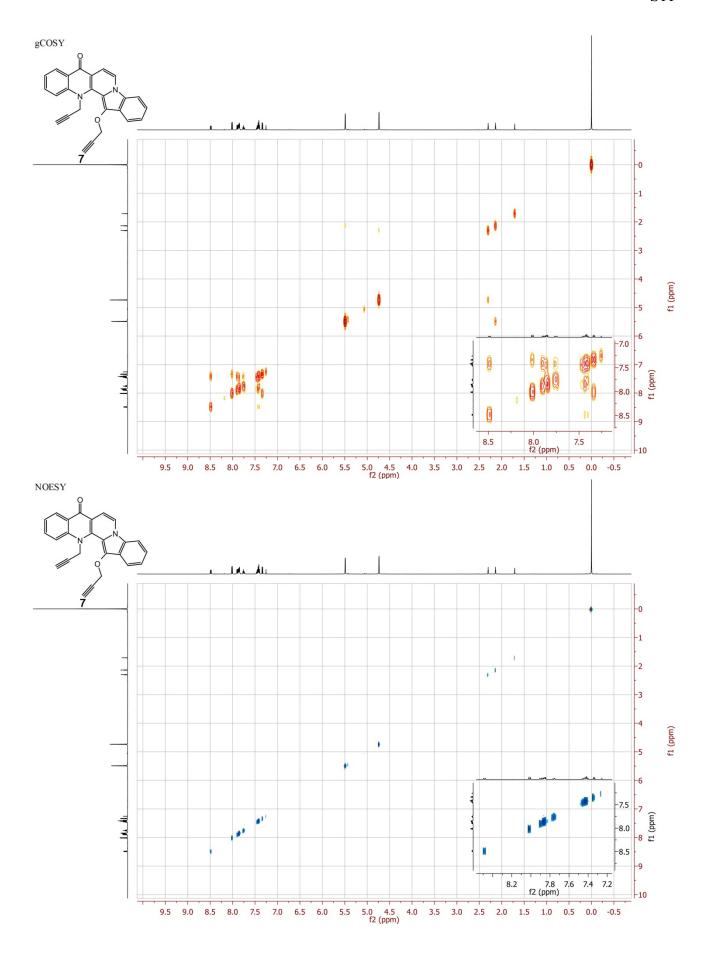


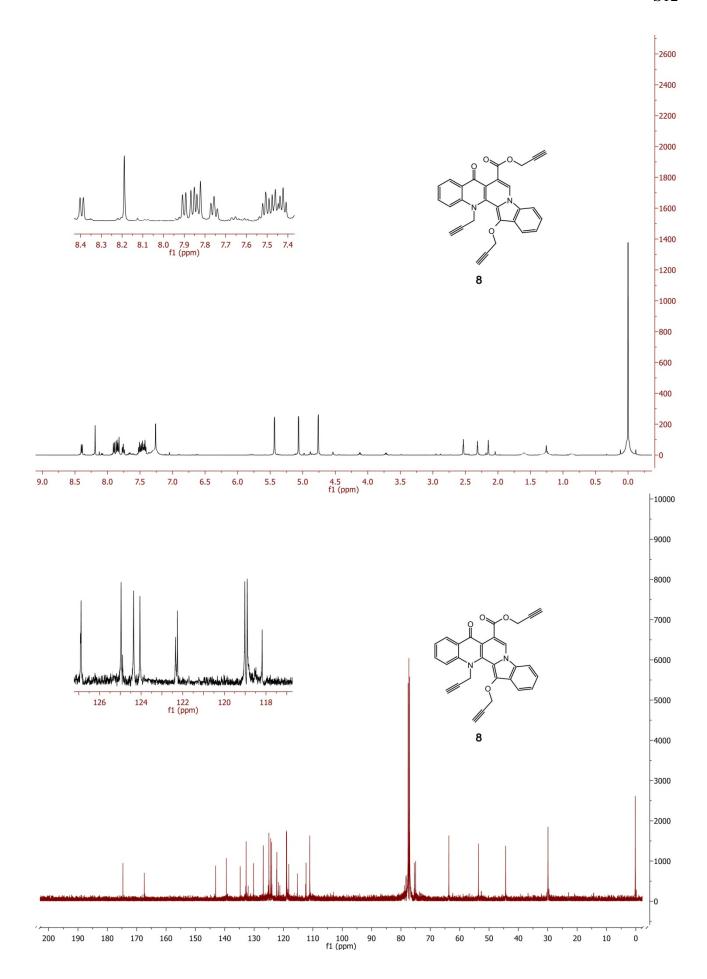




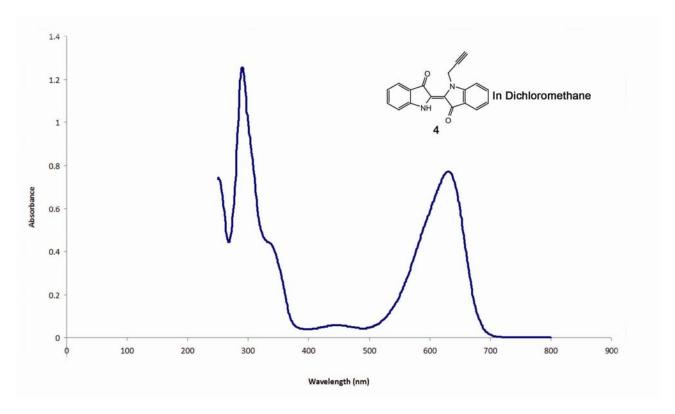


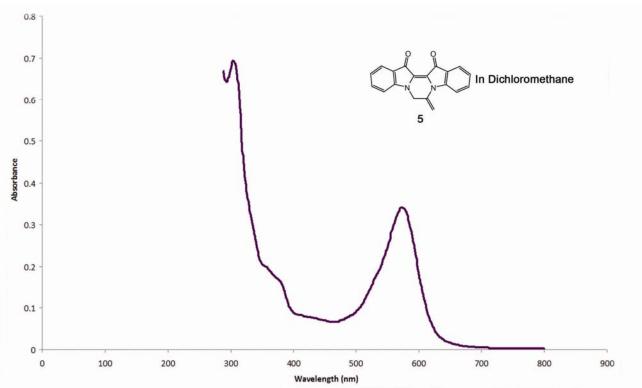


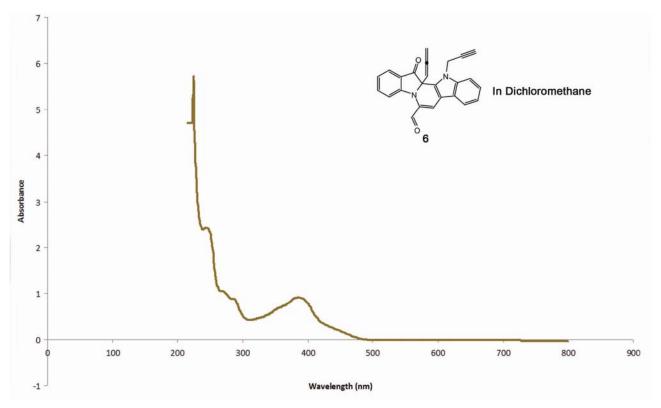


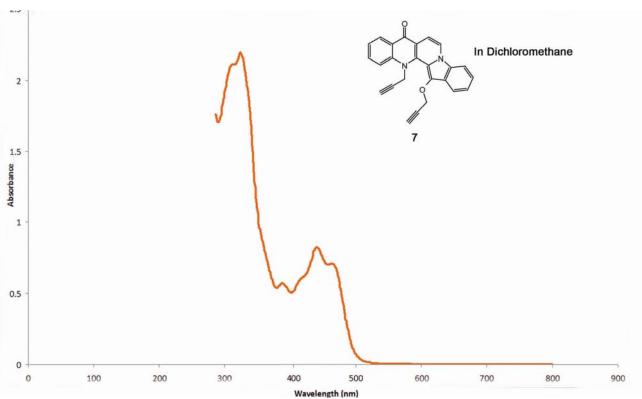


Selected UV-Vis. Spectra of Compound 4-7









Ortep Plots for the Three X-ray Structures Reported in this Manuscript (larger scale)

Crystallographic Studies

Crystallographic Data for Compounds 6, 7 and 8

Compound 6: C₂₅H₁₆N₂O₂, M = 376.41, T = 200 K, monoclinic, space group $P2_1/n$, Z = 4, a = 8.4251(2), b = 10.9245(3), c = 20.7242(4) Å, $\beta = 100.7475(15)$ °; V = 1874.00(8) Å³, $D_x = 1.334$ g cm⁻³, 4284 unique data $(2\theta_{\text{max}} = 50 \text{ °})$, R = 0.039 [for 3584 reflections with $I > 2.0\sigma(I)$]; Rw = 0.104 (all data), S = 0.99.

Compound 7: C₂₅H₁₆N₂O₂, M = 376.41, T = 200 K, monoclinic, space group $P2_1/n$, Z = 4, a = 8.8893(2), b = 21.6743(3), c = 10.0894(4) Å, $\beta = 106.0180(9)$ °; V = 1868.45(7) Å³, $D_x = 1.338$ g cm⁻³, 4287 unique data $(2\theta_{\text{max}} = 50 \text{ °})$, R = 0.040 [for 3143 reflections with $I > 2.0\sigma(I)$]; Rw = 0.098 (all data), S = 0.96.

Compound 8: $C_{29}H_{18}N_2O_2$, M = 458.47, T = 200 K, triclinic, space group P-1, Z = 2, a = 7.5927(2), b = 9.4172(3), c = 16.2992(4) Å, $\alpha = 76.3835(14)$, $\beta = 88.6282(18)$, $\gamma = 78.0391(19)$ °; V = 1107.70(6) Å³, $D_x = 1.375$ g cm⁻³, 5070 unique data $(2\theta_{\text{max}} = 50)$ °), R = 0.046 [for 4074 reflections with $I > 2.0\sigma(I)$]; Rw = 0.125 (all data), S = 0.99.

Structure Determination

Images were measured on a Nonius Kappa CCD diffractometer (MoKa, graphite monochromator, λ = 0.71073 Å) and data extracted using the DENZO package. Structure solution was by direct methods (SIR92). The structures of compounds **6**, **7** and **8** were refined using the CRYSTALS program package. Atomic coordinates, bond lengths and angles, and displacement parameters for compounds **6**, **7** and **8** have been deposited at the Cambridge Crystallographic Data Centre (CCDC nos. 932222, 932223 and 932224, respectively). These data can be obtained free-of-charge via www.ccdc.cam.ac.uk/data_request/cif, by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

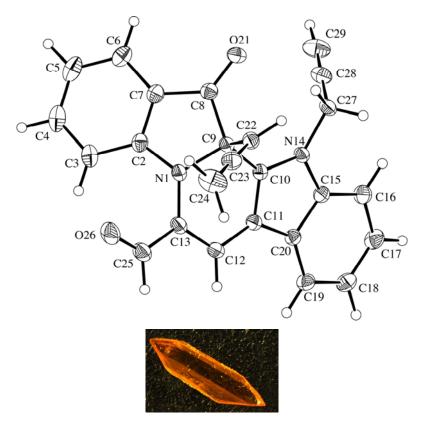


Figure S1: Structure of compound **6** with labeling of selected atoms. Anisotropic displacement ellipsoids show 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

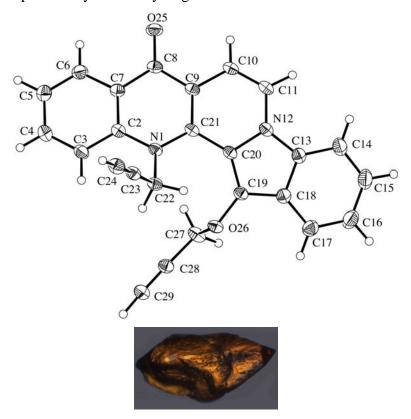


Figure S2: Structure of compound **7** with labeling of selected atoms. Anisotropic displacement ellipsoids show 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

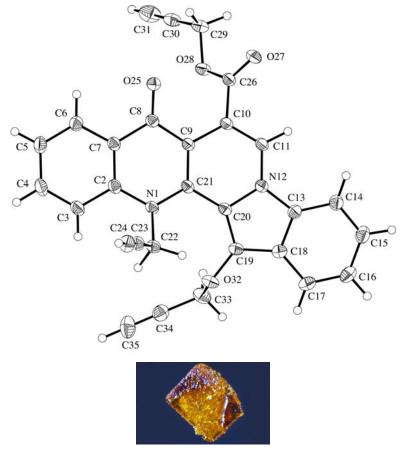
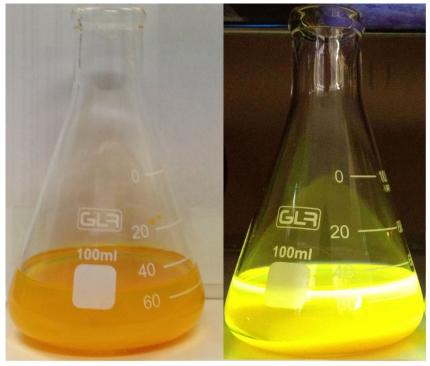


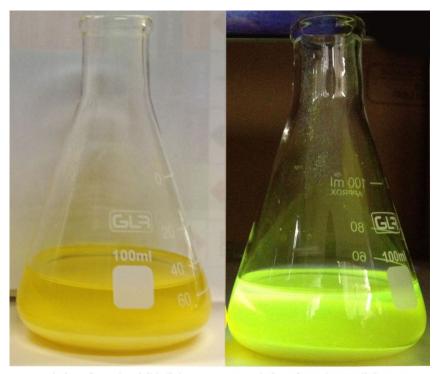
Figure S3: Structure of compound **8** with labeling of selected atoms. Anisotropic displacement ellipsoids show 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

Supplementary Pictures and Figures



A; Solution of 7 under visible light

B; Solution of 7 under UV-light 365 nm



 ${f C}$; Solution of ${f 8}$ under visible light

D; Solution of 8 under UV-light, 365 nm

Figure S4: Colour of the solutions of **7** and **8** in EtOAc under visible & UV-light and the fluorescence emission.

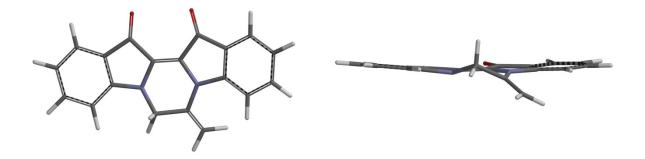


Figure S5: Modeling for the position of the hydrogen atoms of the *endo* cyclic methylene and the exocyclic methylene in **5** with Spartan 10, Geometry Optimised, 6-31G (D) Level.

References

- 1. DENZO–SMN. Otwinowski, Z.; Minor, W. Processing of X-ray diffraction data collected in oscillation mode. In *Methods in Enzymology, Volume 276: Macromolecular Crystallography, Part A*; Carter Jr., C. W.; Sweet, R. M., Eds.; Academic Press: New York, 1997; pp. 307–326.
- 2. SIR92. Altomare, A.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli, M. *J. Appl. Crystallogr.* **1994**, 27, 435.
- 3. Betteridge, P. W.; Carruthers, J. R.; Cooper, R. I.; Prout, K.; Watkin, D. J. J. Appl. Crystallogr. 2003, 36, 1487.