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Assessment of a multi-species in situ FTIR for precise atmospheric greenhouse gas observations

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Abstract

We thoroughly evaluate the performance of a multi-species, in situ Fourier transform infrared (FTIR) analyser with respect to high-accuracy needs for greenhouse gas monitoring networks. The in situ FTIR analyser is shown to measure CO₂, CO, CH₄ and N₂O mole fractions continuously, all with better reproducibility than the inter-laboratory compatibility (ILC) goals, requested by the World Meteorological Organization (WMO) for the Global Atmosphere Watch (GAW) programme. Simultaneously determined $\delta^{13}\text{C}_{\text{CO}_2}$ reaches reproducibility as good as 0.03‰. Second-order dependencies between the measured components and the thermodynamic properties of the sample, (temperature, pressure and flow rate) and the cross sensitivities among the sample constituents are investigated and quantified. We describe an improved sample delivery and control system that minimises the pressure and flow rate variations, making post-processing corrections for those quantities non-essential. Temperature disequilibrium effects resulting from the evacuation of the sample cell are quantified and improved by the usage of a faster temperature sensor. The instrument has proven to be linear for all measured components in the ambient concentration range. The temporal stability of the instrument is characterised on different time scales. Instrument drifts on a weekly time scale are only observed for CH₄ (0.04 nmol mol⁻¹ day⁻¹) and $\delta^{13}\text{C}_{\text{CO}_2}$ (0.02‰ day⁻¹). Based on 10 months of continuously collected quality control measures, the long-term reproducibility of the instrument is estimated to $\pm 0.016 \mu\text{mol mol}^{-1}$ CO₂, $\pm 0.03\text{‰}$ $\delta^{13}\text{C}_{\text{CO}_2}$, $\pm 0.14 \text{ nmol mol}^{-1}$ CH₄, $\pm 0.1 \text{ nmol mol}^{-1}$ CO and $\pm 0.04 \text{ nmol mol}^{-1}$ N₂O. We propose a calibration and quality control scheme with weekly calibrations of the instrument that is sufficient to reach WMO-GAW inter-laboratory compatibility goals.

Keywords

observations, greenhouse, gas, atmospheric, precise, ftir, situ, species, multi, assessment

Disciplines

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Assessment of a multi-species in situ FTIR for precise atmospheric greenhouse gas observations

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Abstract. We thoroughly evaluate the performance of a multi-species, in situ Fourier transform infrared (FTIR) analyser with respect to high-accuracy needs for greenhouse gas monitoring networks. The in situ FTIR analyser is shown to measure CO₂, CO, CH₄ and N₂O mole fractions continuously, all with better reproducibility than the inter-laboratory compatibility (ILC) goals, requested by the World Meteorological Organization (WMO) for the Global Atmosphere Watch (GAW) programme. Simultaneously determined $\delta^{13}\text{CO}_2$ reaches reproducibility as good as 0.03 ‰. Second-order dependencies between the measured components and the thermodynamic properties of the sample, (temperature, pressure and flow rate) and the cross sensitivities among the sample constituents are investigated and quantified. We describe an improved sample delivery and control system that minimises the pressure and flow rate variations, making post-processing corrections for those quantities non-essential. Temperature disequilibrium effects resulting from the evacuation of the sample cell are quantified and improved by the usage of a faster temperature sensor. The instrument has proven to be linear for all measured components in the ambient concentration range. The temporal stability of the instrument is characterised on different time scales. Instrument drifts on a weekly time scale are only observed for CH₄ (0.04 nmol mol⁻¹ day⁻¹) and $\delta^{13}\text{CO}_2$ (0.02 ‰ day⁻¹). Based on 10 months of continuously collected quality control measures, the long-term reproducibility of the instrument is estimated to $\pm 0.016 \mu\text{mol mol}^{-1} \text{CO}_2$, $\pm 0.03 \text{‰} \delta^{13}\text{CO}_2$, $\pm 0.14 \text{ nmol mol}^{-1} \text{CH}_4$, $\pm 0.1 \text{ nmol mol}^{-1} \text{CO}$ and $\pm 0.04 \text{ nmol mol}^{-1} \text{N}_2\text{O}$. We propose a calibration and quality control scheme with weekly calibrations of the instrument that is sufficient to reach WMO-GAW inter-laboratory compatibility goals.

1 Introduction

The globally distributed in situ greenhouse gas (GHG) monitoring network is one of the mainstays of modern climate research. Only a few continuous atmospheric CO₂ records go back to the 1950s (Keeling et al., 1976), but nowadays many stations monitor nearly all long-lived GHGs with in situ instrumentation (Worthy, 2003; Messenger et al., 2008). The required accuracy and precision for measurements of the most important GHG species have been set to limits that allow extracting the required biogeochemical information from spatial differences that are needed for quantifying continental scale GHG fluxes and their inter-annual changes (WMO report No. 5, 1981, cited in Francey and Steele, 2003). Over the most recent decades, non-dispersive infrared (NDIR) analysis of CO₂ and gas chromatography (GC) of CO₂ and all other long-lived GHGs has been proven to provide this accuracy and precision; they have thus become standard techniques for GHG monitoring. Both techniques require special care, maintenance, frequent calibration and quality control measures to guarantee data quality; they are thus labour intensive in their day-to-day operation. In recent years, optical techniques like Cavity Ring-Down Spectroscopy (CRDS), Off-Axis Integrated Cavity Output Spectroscopy (OA-ICOS) or Fourier transform infrared spectroscopy (FTIR) have reached similar or even better precisions than traditional GC systems. Contrary to the discrete samples measured with GC systems, these techniques offer real continuous data acquisition and are in general less labour intensive (Winderlich et al., 2010). Optical techniques can be divided into two fundamentally different methods: (1) laser-based methods and (2) broadband infrared spectroscopy. The major distinctive feature between them is the range of the recorded and evaluated absorption spectrum.

Whereas laser-based instruments are tuned to a narrow absorption window with ideally little interference from other species, the FTIR scans a broad IR range, thereby offering the possibility to measure a large number of species simultaneously. The laser-based techniques like CRDS and OA-ICOS have been extensively and successfully studied by the GHG measurement community; some instruments have already found their way into today's observational networks (Winderlich et al., 2010).

Another benefit of optical spectroscopy is the possibility of discriminating isotopologues, e.g. it allows for continuous measurement of $\delta^{13}\text{CO}_2$. The challenge of isotopologue-selective measurements is, however, to accurately determine the absolute sums of all individual isotopologues. To do so, either all relevant isotopologues have to be measured independently, as done by the in situ FTIR analyser, or assumptions on the mean relation between $^{13}\text{CO}_2$ and $^{12}\text{CO}_2$ in clean air have to be taken into account during evaluation and/or should be already incorporated in the calibration of the instrument (Chen et al., 2010).

So far, studies discussing the use of the in situ FTIR technique for GHG monitoring purposes are rare, although promising (Griffith et al., 2010). Therefore, detailed investigations of the potential and the possible shortcomings of this technique with respect to the accuracy goals set by WMO (World Meteorological Organization)/GAW (Global Atmosphere Watch) experts for clean background air monitoring should be performed before it is used widely within the monitoring community. In particular, questions regarding long-term stability, calibration frequency and cross sensitivity of different trace gases have to be addressed. In the present paper we report on experiments that were performed at the University of Heidelberg Institut für Umwelphysik (IUP), on instrument repeatability, parameter- and cross sensitivity, linearity and long-term stability of an in situ FTIR analyser that was designed and built at the University of Wollongong, Australia (UoW) (Griffith et al., 2012). These data have been gathered in the IUP laboratory as well as in the course of the ICOS (Integrated Carbon Observation System, <http://www.icos-infrastructure.eu/>) Demonstration Experiment, where the instrument was run at two field stations in Europe. The UoW FTIR instrument is subject to an ongoing development process, and many findings of this paper have already led to improvements in newer instrument versions. Nonetheless, our findings are generally applicable to any in situ FTIR instrument and can therefore be used as guideline for in situ FTIR users in order to raise awareness for high-end accuracy applications.

In the first part of this paper (Sect. 2), the initial instrumental setup and its subsequent modifications are introduced, along with a description of the spectroscopic retrieval technique and the standard operating conditions used. Section 3 discusses and quantifies second-order sensitivities of the measured mole fractions to the thermodynamic properties of the sample, such as temperature and pressure. The is-

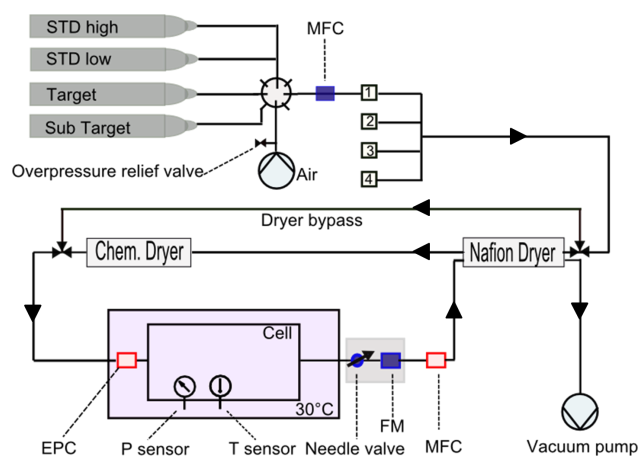


Fig. 1. Schematic setup of the in situ FTIR analyser. The blue parts have been replaced with the red parts in the final IUP setup. The modifications include mass flow controllers (MFC) and electronic pressure controllers (EPC) and replace the original needle valve–flow meter (FM) unit.

sue of inter-species cross sensitivities is discussed as well. This section is extended in Appendix A by an evaluation of the thermodynamic conditions in the cell after sample exchange. After characterising the instrument's sensitivities, its response function in the ambient concentration range is investigated in section 4, followed by an exhaustive study on instrument stability and performance on short (weekly) and long (monthly) time scales in Sect. 5. This section also includes an empirical determination of the required calibration frequency. An overall error assessment is given in Sect. 6. The paper concludes with a discussion of the general applicability of the in situ FTIR analyser for background greenhouse gas monitoring purposes, as well as a recommendation for further improvement of the instrument performance.

2 Instrumental setup and sample handling

Griffith et al. (2010, 2012) describe the improved in situ FTIR instrument used in the present work, which is based on the early version of the instrument as described by Esler et al. (2000a, b) and findings of the current study. The essential parts of the in situ FTIR analyser and the modifications introduced in Heidelberg, based on the first year's findings, are presented here. The different hardware configurations of the instrument throughout this study are specified in Sect. 2.1 below.

2.1 Instrument components and sample handling

The instrument consists of a commercially available FTIR interferometer (IRcube, Bruker Optics, Germany) and a 3.5 L multi-pass cell with 24 m optical path length (PA-24, Infrared Analysis, Anaheim, USA) (see Fig. 1). To avoid artefacts, the

transfer optics between the interferometer, the multi-pass cell and the interferometer housing itself are permanently flushed with high-purity nitrogen (99.999 %). The FTIR interferometer and the multi-pass cell are aligned via an optical bench situated in an actively temperature controlled enclosure. In its basic configuration the in situ FTIR analyser has four separate sample inlets and air is drawn through the instrument using an oil-free vacuum pump (model MV2NT, Vacuubrand, Germany) at the outlet of the instrument. More intake lines can be added via a multi-position valve (MWSD16 selection valve, Valco, USA) connected to any of the four standard inlets.

The in situ FTIR analyser has a built-in sample-drying system consisting of a 24 inch (0.6 m) Nafion[®] dryer (Permapure, Toms River, NJ, USA) operated in counter-flow mode, followed by a chemical dryer filled with magnesium perchlorate ($\text{Mg}(\text{ClO}_4)_2$) for uptake of residual water. The Nafion dryer is placed in a separate compartment of the instrument, along with tubing, valves, PC and power supplies. The drying system reaches a dew point of $\approx -65^\circ\text{C}$ and can be bypassed if measurement of H_2O isotopologues is of interest. The dew point is estimated based on the uncalibrated H_2O measurements of the in situ FTIR analyser. For more details on H_2O isotopologue measurements, refer to Parkes et al. (2013).

The modifications of the instrumental setup performed in Heidelberg are all related to sample handling as well as controlling and measuring sample conditions. In the following the different instrumental setups are introduced:

1. Initial UoW setup: the sample flow through the system is adjusted with a needle valve (NV) and monitored using a flow meter (FM) mounted at the outlet of the cell (see Fig. 1, blue parts). The multi-pass cell is equipped with an in situ PT100 resistance temperature detector (RTD) and with a pressure sensor (HPM-760s, Teledyne Hastings, USA) to determine the thermodynamic sample properties. Both sensors are assumed to be linear and calibrated using a simple two point calibration.
2. ICOS Demonstration Experiment setup: for additional stabilisation of the sample flow and pressure in the cell, the ICOS Demonstration Experiment setup used an external add-on mass flow controller (MFC) (2 slpm, MKS Instruments, USA) in between the outlet of the multi-port valve and the instrument air inlet (see Fig. 1). Pressure and temperature sensors remain unchanged.
3. Intermediate setup: the intermediate setup has one built-in mass flow-controller (Model 3660, Kofloc, Japan), replacing the needle-valve flow meter unit as well as the external MKS flow controller (compare Fig. 1). The internal MFC is located at the outlet of the cell and can be operated in flow control mode or in pressure control mode when combined with a software embedded feedback loop to the pressure sensor. The RTD temperature

sensor in the cell was also replaced by a faster responding J-type thermocouple. The thermocouple was centred in the cell to get a more representative temperature measurement. The temperature sensor of the enclosure temperature control was removed from the cell wall. This version was equivalent to that originally provided by Ecotech (Spectronus GHG analyser, Knoxfield, Australia) in the first commercially available versions of the analyser.

4. IUP setup: in addition to the intermediate setup, the sample pressure in the cell is controlled by an electronic pressure controller (EPC) (P-602CV EL-Press, Bronkhorst, The Netherlands) mounted at the cell inlet (see, Fig. 1). This configuration is functionally equivalent to the current version described by Griffith et al. (2012), with both pressure and flow control and available commercially from Ecotech.

In all configurations the cell is operated at slight overpressure to increase signal-to-noise ratio and to ease leak detection. A diaphragm pump fitted with an EPDM membrane (N 86 KN.18, KNF Neuberger, Germany) is used to pressurise ambient air up to 1800 hPa. Long-term GC experience has shown that these pumps have negligible effects on the measured species. Nevertheless, each pump is tested for contamination prior to its use by sucking air from a free-flowing cylinder and directing it to the GC. The free flowing cylinder air is alternately analysed by the GC with and without the pump to account for the fast drainage effects of cylinders and regulators (Hammer et al., 2012).

2.2 Spectroscopic concentration retrieval

The spectral range of the IRCube is $1800\text{--}7500\text{ cm}^{-1}$ with a 1 cm^{-1} resolution. The in situ FTIR analyser records and stores a broadband absorption spectrum from $1800\text{--}5000\text{ cm}^{-1}$ for each measurement. The recorded spectra are analysed online by non-linear least squares fitting of sections of the measured spectrum with a modelled spectrum calculated from the HITRAN database of absorption line parameters (Rothman et al., 2005). The theoretical spectrum is calculated by MALT (multiple atmospheric layer transmission) as described elsewhere (Griffith, 1996; Griffith et al., 2003, 2012). Three separate spectral regions are fitted for each spectrum: $2150\text{--}2320\text{ cm}^{-1}$ for $^{13}\text{CO}_2$, $^{12}\text{CO}_2$, CO and N_2O , $3001\text{--}3150\text{ cm}^{-1}$ for CH_4 and $3520\text{--}3775\text{ cm}^{-1}$ for CO_2 (all isotopologues) and residual H_2O . The spectral analysis determines the molar concentrations (c_i [mol m^{-3}]) of each gas species. To convert molar concentrations into mole fractions (x_i [mol mol^{-1}]), sample pressure and temperature need to be taken into account:

$$x_{i(\text{wet})} = c_i / (p/RT), \quad (1)$$

where p is the absolute cell pressure, T the absolute temperature, R the universal gas constant and i the investigated

species. Since residual water is determined from the spectrum for each sample, dry-air mole fractions x_i can easily be derived from

$$x_{i(\text{dry})} = x_{i(\text{wet})} / (1 - x_{\text{H}_2\text{O}}). \quad (2)$$

2.3 Standard operating conditions (SOC)

The standard operating conditions (SOC) of the in situ FTIR comprises the instrument settings, measurement mode and interval as well as sample change-over strategies.

- Instrument settings: the enclosure temperature is set to 30.0 °C. It is stable within ± 0.06 °C, which leads to a stability of the cell temperature of ± 0.02 °C for moderately stable laboratory conditions of ± 1 °C. All samples are dried using the built-in drying system (Fig. 1). The sample flow rate is set to 1 ± 0.02 slpm. For the ICOS Demonstration Experiment setup, the cell pressure was set via sample delivery pressure and was kept at 1100 ± 8 hPa. In the modified IUP setup, sample pressure and flow are controlled separately to better than ± 0.1 hPa and ± 0.008 slpm, respectively.
- Measurement mode: the in situ FTIR analyser offers static and dynamic measurement modes. In the static measurement mode, the cell is filled with the sample, sample flow is shut off and the sample is subsequently measured for a certain measurement interval. The dynamic mode measures the sample for a certain interval while it is continuously flushed through the cell. Since systematic differences between the two measurement modes were found (see Sect. 3.5), we chose to flush both sample types, ambient air and air from high-pressure cylinders, continuously through the cell with the same flow rate of 1.00 ± 0.02 slpm. This approach is taken to assure comparability for both sample types.
- Measurement interval: the measurement interval is set to 3 min, in which a 2.5 min spectra collection period is followed by 0.5 min online analysis with the installed PC. This time interval was chosen as a compromise between instrument precision (increasing with averaging time; see Sect. 5.1) and smoothing out natural variability in the ambient air, which itself blurs the averaged spectra. For example, in Heidelberg CO values can change by more than $100 \text{ nmol mol}^{-1}$ within 30 min during rush-hour situations (Hammer et al., 2009). In addition, the 3 min measurement interval equals the approximate turn over time of the sample in the cell at 1 slpm flow rate.
- Sample change-over strategy: under SOC each sample change-over, i.e. changing from ambient to cylinder measurements and vice versa, involves a two-step evacuation of the cell that is described in detail in Appendix A. Possible disadvantages of evacuating the cell,

i.e. by disturbance of moisture or temperature equilibrium, will be discussed in Sect. 3.6 and Appendix A. In SOC each cylinder measurement is performed over 30 min, including the sample change-over, and requires a total gas volume of about 27 L of air.

3 Residual sensitivities to sample properties and inter-species cross sensitivities

3.1 Origin of residual and cross sensitivities

3.1.1 Residual sensitivities to sample properties

The line shapes of the investigated species are dependent on pressure- and Doppler broadening and thus depend on sample properties like pressure and temperature. For both broadening effects, temperature- and pressure-dependent line widths are tabulated in the HITRAN 2004 database (Rothman et al., 2005); they are considered by the FTIR spectra evaluation program MALT (Griffith, 1996), using the measured sample temperature and pressure. However, the line shape parameters themselves are subject to ongoing improvement: as an example, for the CO₂ line parameters, differences of up to a few percent are reported in recent studies (Long et al., 2011; Nakamichi et al., 2006). Small errors in the HITRAN parameters lead to systematic biases in the retrieved molar concentrations. We will refer to this error contribution as the *line shape error*.

Furthermore, the retrieved molar concentrations are biased by potential offsets in the measured sample temperature and pressure since these quantities are directly used in MALT to select the tabulated line shape parameters. In the following, these introduced retrieval biases are referred to as the *introduced spectroscopic error*.

In addition to the line shape and the introduced spectroscopic error, the conversion from molar concentrations to mole fractions (see Eq. 1) constitutes another direct link to the measured sample properties and their precision. This direct link is established through the sample density (dependent on temperature and pressure) and is thus approximately the same for all species on a percentage basis. This emphasises the importance of accurate sample temperature- and pressure measurements to minimise the *density error*. Accurate and temporally stable sensor calibration down to a level of 0.01 % is thus important to reach the required accuracy and precision for the greenhouse gas measurements. Determining the average sample temperature is challenging since temperature is not homogeneous within the cell. In all current setups temperature is measured in one location only, assuming a constant temperature distribution. However, the *true* temperature distribution in the cell depends on sample flow rate and injection (see Appendix A). The measured mole fractions may thus implicitly depend on the flow rate as well.

The combination of the line shape, the introduced spectroscopic and the density error are thus the underlying cause of *residual* pressure and temperature sensitivities. The term *residual* is used as the first-order changes of these parameters are already accounted for and only the deviations between the measured and the *true* sample and spectroscopic properties cause these effects. Since all error contributions are coupled, it is experimentally not possible to disentangle the error contributions. However, in a synthetic MALT study it is possible to investigate the introduced spectroscopic and the density error. This approach will be discussed in Sects. 3.3 and 3.4.

3.1.2 Inter-species cross sensitivities

Apart from residual pressure, temperature and flow sensitivities, additional inter-species cross sensitivities exist, which are caused by overlapping spectral absorption regions of different trace species. Generally, the MALT least-squares fit is able to disentangle the contributions to absorption at each wave number and to attribute their shares to the different species. Nevertheless, the MALT algorithm, the measured spectra and the HITRAN data are not perfect, and small inter-species cross sensitivities remain. Since H₂O absorption in the infrared region occurs at many different wavelengths, the inter-species sensitivity to residual water vapour is noticeable for all investigated species, apart from CH₄. The second-strongest absorber in ambient air is CO₂. Since the ¹³CO₂, and N₂O absorptions in the 2150–2320 cm⁻¹ region overlap with strong absorption of ¹²CO₂, measurable inter-species cross sensitivities to CO₂ also exist.

3.2 Sensitivity experiments

We carried out a series of dedicated experiments to detect and quantify the residual sensitivities to sample properties (pressure, temperature and flow) and the cross sensitivities (H₂O and CO₂) of the in situ FTIR analyser. If applicable, these measurements were used to define correction functions for each species. In all experiments the investigated sample property or species was systematically varied, while all other parameters or species were kept as constant as possible. The experiments have been repeated several times over the course of 1 yr to investigate temporal stability of the sensitivities. For each test we used ambient Heidelberg air collected in high-pressure cylinders with a diving compressor (model P3W, Bauer, Germany) and dried to a dew point of approximately -40 °C. The cylinders (40L L6X aluminium, Luxfer, UK) and pressure regulators (model 14A, Scott Specialty Gases, USA) have proven to be suitable for high-precision GHG measurements by GC analysis (Hammer, 2008). The GHG concentrations in each test cylinder were checked for drifts by GC analysis before and after use.

To investigate the residual sensitivities against temperature, pressure and flow, the respective parameter was tuned at the FTIR analyser itself. Determining the inter-species cross

sensitivities involved a custom-made mixing device, consisting of two mass flow controllers (MFC) and a scrubbing agent, either Ascarite[®] for CO₂, or Mg(ClO₄)₂ for H₂O. The mixing device divides the sample stream into two branches, one of them containing the scrubbing agent. The flow controllers are used to provide varying flow ratios in the two branches. After scrubbing, both branches are re-combined and the mixed gas is injected into the in situ FTIR analyser.

A residual- or cross-sensitivity experiment determines the concentrations of a fixed sample with respect to at least four different settings of the investigated sensitivity parameter. For each setting we allowed sufficient time to re-establish equilibrium in the whole system, i.e. until the variability in CO₂ was on the order of the instrument's repeatability, and then held constant for at least 30 min. The averaged dry-air mole fractions for each equilibrium setting were then used to quantify the sensitivity. In the following sections, the results of the experiments will be discussed in detail with respect to their significance and temporal stability. As a reference for the required precision to monitor natural variability at clean-air background sites, we will refer to the inter-laboratory compatibility (ILC) goals as defined by the WMO-GAW expert group (WMO, 2011). We are aware that *precision* and *compatibility* are two different concepts. However, since to our knowledge no explicit precision goals for GHG measurements are defined, we use those of the inter-laboratory compatibility instead, which are as follows: 0.1 or 0.05 μmol mol⁻¹ for CO₂ in the Northern and Southern hemispheres, respectively, 0.01 ‰ for δ¹³CO₂, 2 nmol mol⁻¹ for CO and CH₄ and 0.1 nmol mol⁻¹ for N₂O. A compilation of all residual and cross-sensitivity parameters is given in Table 1.

3.3 Residual pressure sensitivity (RPS):

As the accuracy of the piezo-resistive pressure transducer is 0.25 %, and thus much larger than the required 0.01 %, significant calibration offsets can be expected. The magnitude of residual pressure sensitivity (RPS) depends on the accuracy and calibration of the pressure sensor as well as errors in the Hitran pressure-dependent line widths and MALT model. Consequently, all five species – CO₂, δ¹³CO₂, CO, CH₄ and N₂O – show a significant residual sensitivity to cell pressure. Two different pressure ranges, one from 800 to 1200 hPa and a sub-range around the operating pressure from 1085 to 1115 hPa were tested. The observed RPSs were linear and compatible for both pressure ranges as displayed in Fig. 2a for CO₂. This allows the use of a linear correction function to account for the RPS. The slope of the correction function was determined by a weighted-total least-squares fit, accounting for errors in pressure and the investigated species mole fractions (Krystek and Anton, 2007). During the year of our investigations, nine RPS experiments were conducted for all species. The temporal evolution of the derived pressure sensitivity slopes for CO₂ is shown in Figure 2b. The observed slopes vary between 0.0078 and 0.0092 [μmol mol⁻¹ hPa⁻¹].

Table 1. Summary of all residual and cross-sensitivity experiments for all species. For each investigated sensitivity (except for the flow rate sensitivity) three rows are given: The first row states the averaged slopes (dx/dy) of the linear regression together with their 1σ uncertainties. Where x denotes the gas species and y denotes the sensitivity parameter. The second row gives an example of the magnitude of the sensitivity for a typical range of parameter variations. The last row categorises the temporal stability of the sensitivities over the course of 1 yr. In the case of a temporally unstable sensitivity, the largest sensitivity value (dx/dy) is given as an upper estimate and in those cases no uncertainty estimates are tabulated. For the flow rate sensitivity, an additional row is given stating the difference between a measurement performed at a flow rate of 1 slpm and a static measurement. For the CO_2 cross sensitivity, only the linear cross sensitivities for unpolluted CO_2 levels are given. The temperature disequilibrium sensitivities (TDS) that were derived from cylinder measurements in the ICOS Demonstration Experiment setup are given as well. The last three rows summarise the effect of a 1°C temperature bias as result of the theoretical MALT study. The correction factors are given independently for the density and spectroscopic error. The bold font of some numbers is to highlight their exceedance of WMO inter-laboratory compatibility goals.

	CO_2 [$\mu\text{mol mol}^{-1}$]	Error [%]	$\delta^{13}\text{CO}_2$ [nmol mol^{-1}]	Error [nmol mol^{-1}]	CO [nmol mol^{-1}]	Error	CH_4	Error	N_2O	Error
dx/dp : [unit hPa $^{-1}$]	0.0085	0.0004	0.005	0.002	0.006	0.002	0.031	0.003	0.007	0.001
Typical variation for: 20 hPa	0.17	0.01	0.10	0.04	0.12	0.04	0.62	0.06	0.14	0.02
Temporal development	stable		linearly decreasing		stable		linearly decreasing		stable	
dx/dT [unit $^\circ\text{C}^{-1}$]	<0.8	–	0.6	0.2	<1	–	<1.6	–	0.6	0.2
Typical variation for: 0.1 $^\circ\text{C}$	<0.08	–	0.06	0.02	0.10	–	0.16	–	0.06	0.02
Temporal development	unstable		rel. stable		unstable		unstable		rel. stable	
$dx/d\text{Flow}$ [unit slpm $^{-1}$]	0.15	0.001	–0.9	0.6	<2	–	<4	–	<–0.8	–
Typical variation for: 0.03 slpm	0.00	0.00	–0.03	0.02	<0.1	–	<0.1	–	<–0.02	–
Static vs. 1 slpm	0.25		0.60		0.10		1.40		0.28	
Temporal development	stable		unstable		unstable		unstable		unstable	
$dx/d\text{H}_2\text{O}$ [unit ppm $^{-1}$ (H_2O)]	0.04	0.02	–	–	<0.2	–	<0.2	–	–	–
Typical variation for: 2 $\mu\text{mol mol}^{-1}$ (H_2O)	0.08	0.04	–	–	<0.4	–	<0.4	–	–	–
Temporal development	stable				unstable		unstable			
$dx/d\text{CO}_2$ [unit $\mu\text{mol mol}^{-1}$]	–	–	0.006	0.0003	<0.015	–	–	–	0.008	0.0008
Typical variation for: 50 $\mu\text{mol mol}^{-1}$ (CO_2)	–	–	0.3	0.02	0.75	–	–	–	0.4	0.04
Temporal development			stable		unstable				stable	
dx/dT (TDS) [unit $^\circ\text{C}^{-1}$]	2.07	0.05	4.1	0.1	–4.6	0.3	10.2	0.3	3.2	0.1
Typical variation for: 0.1 $^\circ\text{C}$	0.2	0.01	0.41	0.01	0.50	0.03	1.0	0.03	0.32	0.01
Temporal development	stable		stable		stable		stable		stable	
dx/dT [unit $^\circ\text{C}^{-1}$] 1 $^\circ\text{C}$ temperature bias MALT experiment	2.0	4.45	–4.0	7.8	1.7					
dx/dT [unit $^\circ\text{C}^{-1}$] related to the density error	1.3	0.0	0.3	5.0	1.0					
dx/dT [unit $^\circ\text{C}^{-1}$] related to spectroscopic error	0.7	4.45	–4.3	2.8	0.7					

The uncertainties of the slopes depend on the investigated pressure range. Since the RPS has proven to be linear, a larger pressure range is preferable to determine the sensitivity since the fit is more stable and the experiment is easier to conduct. Table 1 summarises the averaged slopes and their 1σ standard deviation. For CO_2 , CH_4 and N_2O , the residual pressure sensitivity slopes were temporally stable within their uncertainties and had a standard deviation of 10%. $\delta^{13}\text{CO}_2$ and CO showed a slight temporal change leading to a standard deviation of 30% for the correction functions for these two components.

Comparing the RPS on a percentage basis shows that the correction factors for the different trace gases differ only by ca. 25%. This would be in accordance with substantial contribution from a common density error (see Sect. 3.1.1). Re-analysing the measured spectra with different pressures allowed investigation of the effect of the density error and the introduced spectroscopic error separately. CH_4 and N_2O did not show spectroscopic error contributions as both species do not have individually resolved lines. For CO_2 the effect of the density error and the introduced spectroscopic error are of similar magnitude, whereas for CO the introduced spectroscopic error is three times larger. Since $\delta^{13}\text{CO}_2$ is calcu-

lated as a ratio, the density effect and most of the introduced spectroscopic effect cancel.

To better judge the relevance of the residual sensitivities, an example based on typical ranges of the investigated parameter is given in Table 1 as well. Pressure variations of ± 10 hPa typical of the initial UoW hardware version translate to $0.17 \mu\text{mol mol}^{-1}$ CO_2 variations; they are thus larger than the inter-laboratory compatibility (ILC) target for CO_2 . The same is true for $\delta^{13}\text{CO}_2$ and N_2O (see Table 1). All three components thus need to be corrected. The uncertainty introduced by the residual pressure correction is negligible for CO_2 and N_2O . In the case of $\delta^{13}\text{CO}_2$, a temporal trend in the RPS leads to a larger uncertainty in the averaged correction factor. This error can be reduced by introducing a temporally changing RPS correction. Even assuming a constant RPS reduces the pressure-induced deviation by a factor of 2.5 compared to the uncorrected values. For CO and CH_4 the RPS corrections are smaller than the required ILCs for the assumed 10 hPa pressure change. Still, for CH_4 in contrast to CO the RPS is significant and its correction will improve the precision of the measurements. Apart from $\delta^{13}\text{CO}_2$, the stability of the correction parameters indicates that annual determination of the residual pressure sensitivity should be

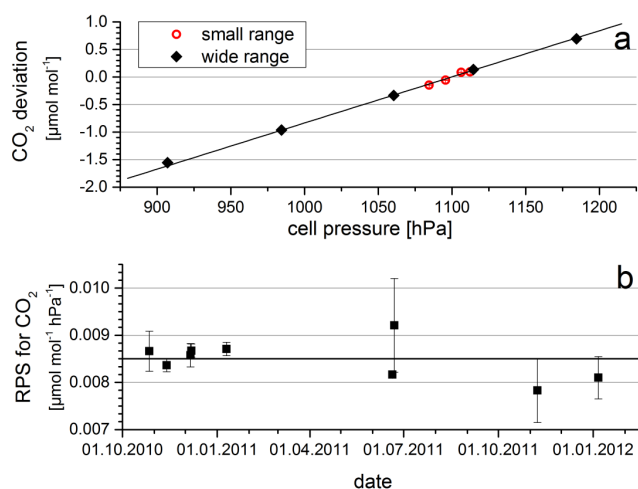


Fig. 2. (a) Residual pressure sensitivity (RPS) for CO₂ measured on 20 and 22 June 2011. Small and wide pressure ranges are shown together with a common linear fit. Individual errors are smaller than the symbols. (b) Temporal stability of the slope of the residual pressure sensitivity for CO₂. The mole fractions of the used cylinders were all in the range of ambient concentrations in Heidelberg.

sufficient to take into account systematic long-term drifts of the RPSs. The underlying cause of the drift in the $\delta^{13}\text{CO}_2$ RPS correction parameter is not yet understood. Temporal drifts in the calibration of the pressure sensor would cause a systematic drift in the correction parameters of all species except $\delta^{13}\text{CO}_2$.

For the instrument versions after the ICOS demonstration setup the pressure is controlled to $\ll 1$ hPa and pressure corrections become very small or negligible. In the IUP instrumental setup, a potential drift of the pressure sensor can additionally be controlled by the second pressure sensor in the additional EPC.

3.4 Temperature sensitivities

3.4.1 Residual temperature sensitivity (RTS)

Compared to the RPS, the residual temperature sensitivity (RTS) has the additional complication that we do not measure the *true* mean sample temperature with one temperature sensor (see Appendix A). The RTD sensor used in the ICOS Demonstration Experiment setup was placed near the outlet of the cell, whilst the thermocouple used from the intermediate setup onwards was located in the middle of the cell. The sample temperature distribution within the cell depends on at least three parameters: (a) the set temperature of the enclosure, (b) the sample temperature when entering the cell, and (c) the sample flow rate. In total, six dedicated RTS experiments were performed for all species. In order to investigate the RTS, the temperature of the cell enclosure was varied either in its normal operational range, i.e. between 29.9 °C and 30.1 °C, or in a wider range from 29.7 °C to

30.3 °C. For each set temperature step we allowed enough time for the spectrometer and the sample cell to equilibrate. In our setups this took roughly 60 to 90 min, based on the CO₂ variability. For CO₂, N₂O and $\delta^{13}\text{CO}_2$, linear residual temperature sensitivities were observed in each experiment. Although each experiment showed good linear relations for CO₂, the slopes varied substantially and even changed sign after six months. For N₂O and $\delta^{13}\text{CO}_2$ the RTS was stable within 30 % for all experiments. The residual temperature sensitivity for CO and CH₄ was weak, not temporally stable and only for some experiments a distinct relation to cell temperature was observed. The averaged RTS slopes and their standard deviations are given in Table 1. For better classification of the results, an example based on observed peak-to-peak temperature variability of 0.1 °C is listed in Table 1 as well. When considering the ILC targets, only the observed RTS for $\delta^{13}\text{CO}_2$ is significant and needs to be corrected.

In order to investigate the effect of the initial sample temperature on the cell temperature, we conducted an experiment where a cylinder was first measured at room temperature before the entire cylinder was cooled to zero degrees Celsius and measured again. The cell temperature as well as the measured mole fractions did, however, not change in this experiment. This implies that the residence time of the sample in the inlet and drying system, i.e. the Nafion dryer, is sufficient to compensate for at least a 30 °C temperature difference of the incoming sample.

3.4.2 Temperature disequilibrium sensitivity (TDS)

The temperature disequilibrium sensitivity (TDS) described in this subsection is related to the RTD temperature sensor used in the UoW and the ICOS Demonstration Experiment setup. These findings resulted in a replacement of the RTD temperature sensor with a J-type thermocouple from the intermediate setup onwards.

Using the RTD sensor, stronger temperature sensitivities were present in all cylinder gas measurement records, implying that temperature is one of the key parameters to perform precise calibration measurements. In the course of the ICOS Demonstration Experiment, the in situ FTIR analyser was set up in different laboratories under different environmental conditions, i.e. averaged laboratory temperatures ranged from 17 to 27 °C. Although the enclosure temperature was stable at 30.00 ± 0.05 °C at all locations, cell temperature varied slightly and was anti-correlated with laboratory temperature. Figure 3 shows the deviations from the averaged mole fraction of the 24- to 48-hourly measured sub-target tank with respect to the cell temperature. The other simultaneously measured cylinders, two calibration gases and the regular target gas, show similar temperature dependencies. We will further refer to this effect as the temperature disequilibrium sensitivity (TDS).

The observed TDSs for cylinder measurements are much larger than the RTSs, which were derived from the dedicated

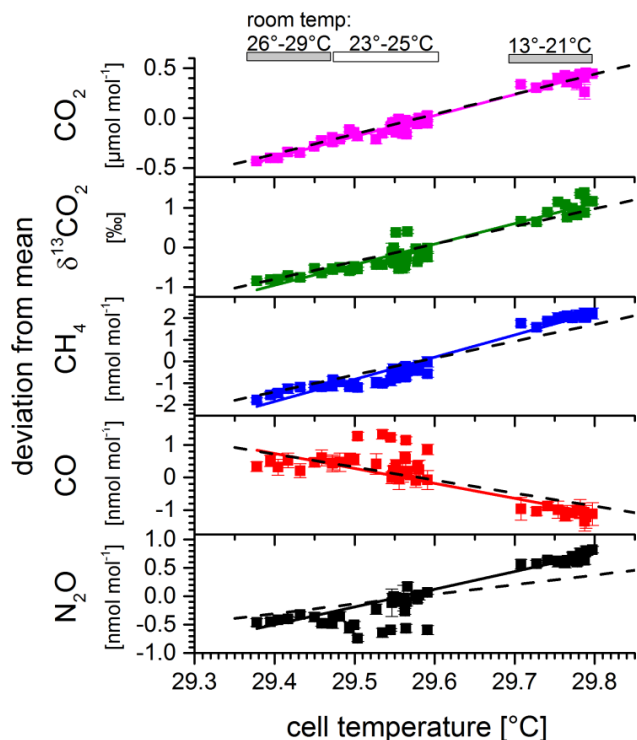


Fig. 3. Temperature disequilibrium sensitivity (TDS) of the sub-target measurements, shown as deviation from mean mole fractions. Values have been corrected for all residual sensitivities except for residual temperature, but not calibrated. The three different temperature ranges in the cell result from different laboratory temperatures at the remote field stations and in the IUP laboratory that are indicated in the top panel. The dashed lines correspond to a TDS for a 1 °C bias in sample temperature determination, based on the density and the introduced spectroscopic error only. Please refer to the text for more details.

temperature experiments described in Sect. 3.4.1. The slopes of the linear TDS and their uncertainties are summarised in Table 1 as well. For the typical temperature range of 0.1 °C, the effect is significantly larger than the ILC goals for CO₂, δ¹³CO₂ and N₂O. A possible cause for the difference between the two temperature sensitivity results may arise from different conditions of the instrument. While in the dedicated RTS experiments sample air was continuously flushed through the cell, each cylinder measurement, performed under standard operating conditions, comprises evacuation of the cell prior to the measurement in flow mode (Appendix A). The evacuation of the cell causes adiabatic cooling on the order of 5 °C. Although this adiabatic cooling effect is compensated for during the re-filling of the cell, the slow response of the RTD sensor, caused by its large thermal mass, leads to a biased temperature measurement, even after the sample temperature has recovered. In the UoW and ICOS demonstration experiment setup the evacuation-induced temperature deviation was further amplified by the fact that the sensor of the active enclosure temperature control was mounted directly on

the cell walls. The adiabatic cooling thus disturbed the temperature equilibrium in the enclosure, leading to a feedback-loop of the active temperature control.

To investigate if the TDS can be explained by a biased sample temperature measurement we examined the influence of a 1 °C temperature bias on the retrieved concentrations of a synthetic absorption spectrum. This allows separating the impact of the density error from the introduced spectroscopic error. The line shape error does not show up in such a study. The theoretical temperature bias correction parameters are listed in Table 1 and shown in Fig. 3 as dashed black lines. The theoretical temperature sensitivity of an assumed temperature bias of 1 °C agrees well with the observed TDS for all species. Only for N₂O the theoretical temperature sensitivity is smaller than the observed TDS.

This result gives strong evidence for biased sample temperature measurements after an evacuation process. The TDS was the main reason to replace the RTD sensor with a faster responding thermocouple. Such thermocouples have become the standard temperature sensor from the intermediate setup onwards. In addition, we moved the sensor of the active enclosure temperature control out of direct contact with the cell wall. With these measures, the TDS was no longer observed for cylinder measurements.

For all cylinder measurements performed with the ICOS Demonstration Experiment setup, we use the TDS correction parameters derived from the sub-target gas measurements to correct all our standard and target gas measurements. The anti-correlated relation between room and cell temperature is most likely caused by an overcompensation of the active enclosure temperature control.

3.5 Flow rate sensitivity

Neither the spectroscopic nor the mole fraction determinations have a direct link to the sample flow rate through the cell. The flow rate has only an indirect effect through the temperature distribution in the cell. Thus we expect the flow rate sensitivity to be small compared to the temperature sensitivity. Nevertheless, the flow rate sensitivity was investigated in three dedicated experiments, mainly motivated from the observed difference between a constantly flushed and a closed-off cell. Apart from CO, the measured mole fractions are always higher in a closed off cell than in a constantly flushed cell. Under our standard operating conditions this effect can be as large as 0.25 μmol mol⁻¹ for CO₂, 0.6 ‰ for δ¹³CO₂, -0.1 nmol mol⁻¹ for CO, 1.4 nmol mol⁻¹ for CH₄ and 0.28 nmol mol⁻¹ for N₂O. We hypothesise that the observed difference between static and dynamic measurements is caused by different temperature distributions in the cell.

The observed flow sensitivity in the range between 0.8 and 1.2 slpm could be linearly approximated, however the linearity broke down when approaching zero flow. As expected, the flow sensitivities are small. In addition to this, the gas-flow through the cell was very stable due to the additional sample

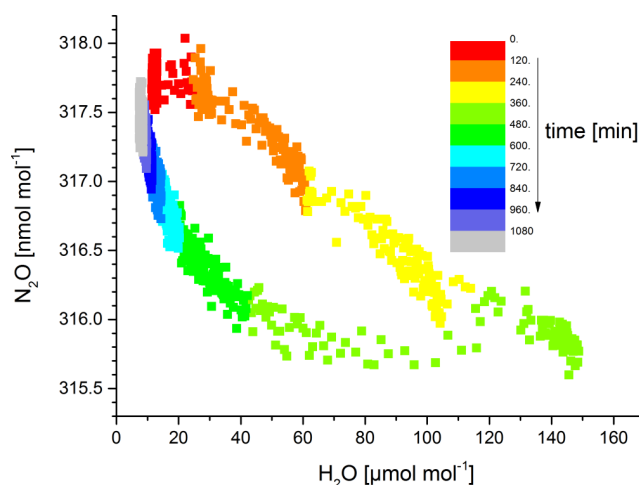


Fig. 4. H₂O cross sensitivity for N₂O. In the course of this experiment, the cell was first gradually humidified and dried afterwards. The colour code represents time since start of the experiment in minutes.

flow controller. Thus, the effects of the flow sensitivity are negligible for all trace gases (see, Table 1). For $\delta^{13}\text{CO}_2$ the flow sensitivity is larger than the ILC target; however, as the repeatability of $\delta^{13}\text{CO}_2$ measurements is still on the order $\pm 0.03\%$, this result is not crucial for the instrument performance. Nevertheless, we apply the flow rate sensitivity correction to our data.

3.6 H₂O cross sensitivity

Measuring the H₂O cross sensitivity is experimentally challenging and time consuming since it takes time until the moisture equilibrium between gas phase and instrument surfaces is established. The drying cartridge in the mixing device acts as an additional resistance, and it was not until our last experiments that we introduced a needle valve into the direct branch of the mixing device to counteract this resistance. Thus, for the first experiments the total resistance of the mixing device changed according to flow proportion in the course of the H₂O cross-sensitivity experiment. This led to a variation in cell pressure on the order of 10 hPa during the experiment. In the following, all results have been corrected for the residual pressure sensitivity before the H₂O cross sensitivity was determined.

A further challenge is a species-dependent hysteresis effect in the H₂O cross sensitivity. Figure 4 shows the H₂O cross sensitivity for N₂O and the very pronounced hysteresis effect. In contrast to the dedicated H₂O cross-sensitivity experiments performed later on, this test was conducted with gradually changing cell moisture. The H₂O cross sensitivity is linear while humidifying the cell (from red to yellow in Fig. 4), whereas a clear hysteresis appears during drying (from green to blue). The origin of the H₂O hysteresis is believed to be related to surface effects in the instrument, but

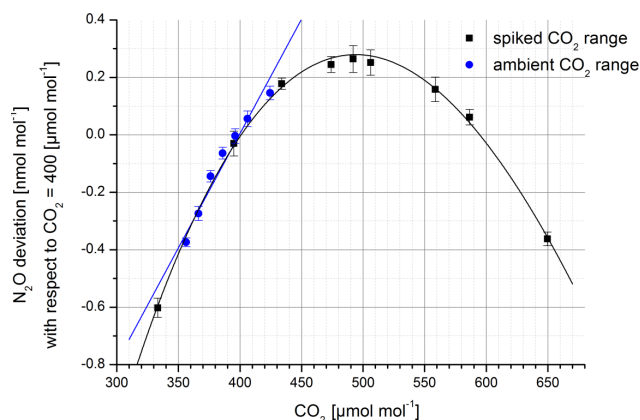


Fig. 5. CO₂ cross sensitivity for N₂O, shown as deviation to a reference CO₂ mole fraction of 400 $\mu\text{mol mol}^{-1}$. In blue a CO₂ cross-sensitivity experiment spanning the unpolluted CO₂ range is shown, together with its linear approximation. Results from a spiked CO₂ experiment and a cubic fit are displayed in black. Both experiments have been conducted with N₂O mole fractions of about 311 nmol mol^{-1} .

was not further investigated since such large moisture variations do not occur in our standard operation conditions. They may, however, become relevant, if also water and water isotopologues are to be measured with the instrument.

For the five dedicated H₂O cross-sensitivity experiments, the investigated H₂O range was restricted to moisture levels between 2 and 10 $\mu\text{mol mol}^{-1}$, i.e. those which may occur during the lifetime (typically 2 months) of one Mg(ClO₄)₂ drying cartridge. Individual H₂O concentrations were kept as constant as possible for each set point to minimise transient effects. Within the measurement precision, no significant H₂O cross sensitivity was observed for N₂O and $\delta^{13}\text{CO}_2$. This is explained by the weak H₂O absorption in the spectral region of N₂O, and for $\delta^{13}\text{CO}_2$ the H₂O cross sensitivity is low since all CO₂ isotopologues are subject to a similar H₂O cross sensitivity in this H₂O range. Although some experiments did show a clear relation between H₂O level and CH₄ or CO mole fractions, no unequivocal cross sensitivity could be determined for these two trace gases. Only for CO₂ a more or less temporally stable H₂O cross sensitivity was found with a 1σ standard deviation of $\pm 50\%$. The unstable results are most likely due to experimental problems. Precise adjustment and stability of moisture levels in the range between 2–10 $\mu\text{mol mol}^{-1}$ is difficult to achieve and in the initial setup of the mixing device small H₂O cross sensitivities have been superimposed by cell pressure variations. Further experiments with additional methods to stabilise cell pressure were needed and later on performed with the improved version of the in situ FTIR analyser. However, since the variations in the residual moisture level during standard operating conditions can be restricted to less than $\pm 2 \mu\text{mol mol}^{-1}$, the H₂O cross sensitivity is not very important. In

Table 1 the H₂O effect is shown in a calculation for a 2 $\mu\text{mol mol}^{-1}$ effect, which exemplifies that for all species the effect is smaller than the ILC target. Nevertheless, for CO₂ the H₂O cross sensitivity is larger than the instrument's repeatability and thus worth correcting. The CO₂ variability associated with the H₂O cross sensitivity could at least be halved by applying the H₂O correction.

3.7 CO₂ cross sensitivity

¹²CO₂ is the strongest absorber in the spectral range between 2150 and 2320 cm^{-1} and its absorption ranges do partly overlap with those of ¹³CO₂, N₂O and CO analysed in the same spectral range. Since MALT uses a broad spectral region for fitting, it is able to distinguish the different species; however, certain cross sensitivities to ¹²CO₂ remain for all mentioned species. For CH₄ the CO₂ cross-sensitivity effect is negligible since the CH₄ concentration is derived at 3001–3150 cm^{-1} , where no significant ¹²CO₂ absorption occurs. In contrast to all previously discussed sensitivities, the influence of the CO₂ cross sensitivity cannot be reduced by minimising the variability of the causing agent. Thus, precise determination of the CO₂ cross sensitivity is vital.

To vary the CO₂ amount we used the mixing device described above with an Ascarite (Sigma Aldrich, USA) filled cartridge. Ascarite has shown to be suitable to remove CO₂ without altering the mole fractions of the other investigated greenhouse gases (Glatzel-Mattheier, 1997). The H₂O that is produced by Ascarite during the CO₂ uptake is removed by the drying system. In order to use the results of the CO₂ cross-sensitivity experiment for $\delta^{13}\text{CO}_2$ as well, it is crucial to remove the CO₂ entirely in the Ascarite branch of the mixing device to avoid isotope fractionation. This was verified by taking aliquot flask samples for each CO₂ level and analysing $\delta^{13}\text{CO}_2$ by mass spectrometry. For our experimental conditions with CO₂ amounts as high as 800 $\mu\text{mol mol}^{-1}$ and a flow rate of up to 1 slpm, 80 g of Ascarite are sufficient to completely remove CO₂ from the Ascarite branch over the 10 h duration of the experiment.

In total seven cross-sensitivity experiments were conducted. The five initial experiments spanned approximately the ambient CO₂ range (340 to 440 $\mu\text{mol mol}^{-1}$). The later experiment used spiked CO₂ concentrations in order to investigate a wider CO₂ concentration range. Figure 5 displays results for both ranges of the CO₂ cross sensitivity for the worst-case example of N₂O. Both experiments are in accordance with each other, although they were performed several months apart. However, the wider CO₂ range reveals further details about the shape of the CO₂ cross sensitivity for N₂O. The wide range of the CO₂ cross sensitivity can be adequately described by a cubic relation, with a correlation coefficient $r^2 > 0.99$. The typical range of unpolluted ambient air (370 to 420 $\mu\text{mol mol}^{-1}$) can be approximated linearly with less than 0.05 nmol mol^{-1} N₂O deviation. However, for larger CO₂ values the deviation between the two approxima-

tions becomes substantial. The sensitivity of N₂O to CO₂ can be reduced to be negligible by selection of a narrower spectral range for N₂O analysis excluding severe overlap with CO₂, as described in Griffith et al. (2012).

The CO₂ cross sensitivities for CO and $\delta^{13}\text{CO}_2$ in the wider range of CO₂ are also not linear. Similarly to the case for N₂O, the cross sensitivities for both species can be approximated linearly in the unpolluted CO₂ range. For CO the CO₂ cross sensitivity starts deviating significantly from a linear relation for CO₂ values above 500 $\mu\text{mol mol}^{-1}$. In Table 1 the magnitude of the CO₂ cross sensitivities are summarised for the linearly approximated unpolluted CO₂ range. $\delta^{13}\text{CO}_2$ actually depends inversely on CO₂ and is dealt with explicitly in Griffith et al. (2012). For N₂O and $\delta^{13}\text{CO}_2$ the effect of the CO₂ cross sensitivity is by far larger than the ILC targets, whereas for CO as well as for CH₄ the CO₂ cross sensitivity is smaller. The Heidelberg FTIR data post-processing includes the non linear CO₂ cross sensitivity corrections for all species also in the linear range.

4 Instrument response function

As the raw *absolute* mole fraction determination of the in situ FTIR analyser differs from the internationally accepted WMO scales by up to a few percent, depending on species (Griffith et al., 2010, 2012), calibration of the FTIR analyser with internationally accepted standard reference material is necessary to achieve the required accuracy and comparability for ambient air monitoring. The shape of the instrument response function (IRF) determines the number of required calibration standards. To determine the IRF of the in situ FTIR analyser we analysed the IUP-Heidelberg set of primary laboratory standards calibrated by the WMO Central Calibration Laboratories (CCL). We have 13 laboratory standards, which have been calibrated for CO₂ on the X2007 mole fraction scale. Seven of these cylinders have also been calibrated for N₂O and five for CH₄ by NOAA/ESRL. NOAA/ESRL acts as the WMO-GAW Central Calibration Laboratory (CCL) for all these trace gases. For $\delta^{13}\text{CO}_2$ we received four reference standards that were calibrated on the VBDP/j-RAS06 scale by the Max Planck Institute for Biogeochemistry (MPI-BGC) in Jena, being the WMO-GAW Central Calibration Laboratory for stable isotopes in CO₂. The same cylinders were calibrated for the CO mole fractions by the MPI-BGC GasLab on the NOAA/ESRL 2004 scale. The calibrated range for each species is given in Fig. 6 and its caption.

All FTIR measurements of these calibration cylinders were corrected for residual and cross sensitivities as described in the previous sections; measurements were performed on three consecutive days. The sensitivity-corrected FTIR mole fractions plotted against the assigned cylinder reference values show no significant curvature, but the applied linear fits have a significant non-zero intercept. In Fig. 6 the residuals to the linear fit confirm that the assumption

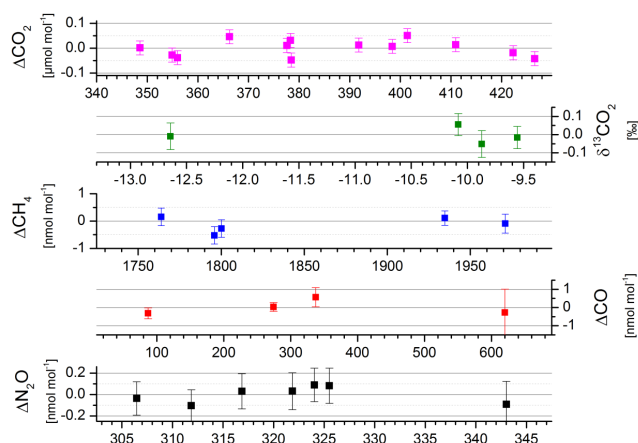


Fig. 6. Linearity of the FTIR analyser. For each species the residual to the linear fit of the externally assigned mole fractions against sensitivity-corrected FTIR mole fractions are shown. The investigated mole fraction ranges are defined by the spread of the secondary laboratory cylinders used for the Heidelberg GC system and are as follows: CO_2 : 348 to 426 $\mu\text{mol mol}^{-1}$; $\delta^{13}\text{CO}_2$: -12.65 to -9.55‰ ; CO : 90 to 620 nmol mol^{-1} ; CH_4 : 1757 to 1970 nmol mol^{-1} and N_2O : 307 to 343 nmol mol^{-1} .

of linear instrument response functions is justified for all species. The error bars in Fig. 6 depict the combined error of the FTIR measurement and the error of the calibration cylinder assigned mole fractions. The standard deviations of the residuals are 0.03 $\mu\text{mol mol}^{-1}$ for CO_2 , 0.04 ‰ for $\delta^{13}\text{CO}_2$, 0.4 nmol mol^{-1} for CO , 0.4 nmol mol^{-1} for CH_4 and 0.08 nmol mol^{-1} for N_2O . From these results we can conclude that the IRFs can, in accordance with the WMO ILC targets, be approximated linearly over the investigated range for all tracers. However, the non-zero intercepts of the regression functions prevent a simple one-point calibration, leading to a minimum of two calibration standards for the FTIR. For high-accuracy demands, as in atmospheric background monitoring programs, the usage of three calibration standards to define the IRF is advised. A three-point calibration reduces the sensitivity to individual outliers and instrument noise in the calibration measurements. In addition, the assumption of linearity for the IRF, and potential long-term changes, can routinely be tested using a goodness of the linear fit criteria.

5 Temporal stability of the FTIR analyser and calibration frequency

In order to assess the precision of the in situ FTIR analyser, measurement repeatability as well as reproducibility on different time scales has to be determined. In the following, we will use the terminology related to the GAW glossary of QA/QC (Klausen and Scheel, 2007). The issue of measurement stability is vital for all long-term monitoring efforts and

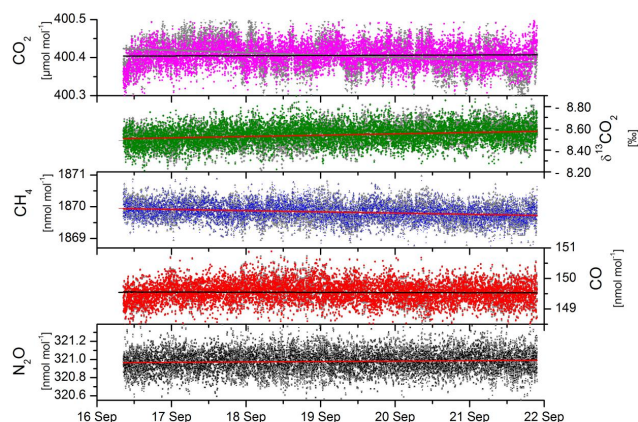


Fig. 7. Short-term stability of the FTIR components during September 2011. Minute-by-minute cylinder measurements over the course of 6 days. In grey the dry-air mole fractions are given (but are only visible in the case of CO_2). The coloured symbols show the sensitivity corrected values, with a linear fit to detect drifts.

determines the required calibration frequency to reach a certain level of reproducibility. In the following, we will subdivide the stability requirement into two parts: short term and long term. *Short term* handles drifts and other artefacts on a time scale from hours to days, while *long term* covers weeks to months.

5.1 Short-term stability

To quantify short-term drifts of the FTIR analyser, a target gas was continuously flushed through the cell at the standard flow rate of 1 slpm for 6 days, comprising a weekend as well as weekdays, to cover different laboratory conditions. Absorption spectra with an averaging time of 1 min were recorded and analysed offline to avoid downtime. The respective time series of the dry-air mole fractions, calculated according to Eq. (2), are shown in Fig. 7 in grey. The sensitivity corrected results are shown as well (coloured symbols).

This short-term stability test was conducted in September 2011 for the intermediate setup. Thus, the sample properties in the cell remained fairly constant over the course of the experiment, as expressed by their 1σ standard deviations: $\pm 0.01\text{ °C}$ for cell temperature, $\pm 0.01\text{ hPa}$ for cell pressure, $\pm 0.2\text{ }\mu\text{mol mol}^{-1}$ for moisture level, and $\pm 0.01\text{ slpm}$ for sample flow rate. The Heidelberg laboratory temperature oscillated between 23.2 and 25.1 °C for this period; however, the influence on cell temperature was small. The generally very stable sample properties in the cell lead to only small corrections of the residual sensitivities (Fig. 7). Only for CO_2 was a clear correlation to cell temperature found. The RTS corrections thus improve the CO_2 standard deviation of the 1 min measurements from 0.037 to 0.029 $\mu\text{mol mol}^{-1}$ – for all other species the sensitivity corrections are marginal.

The initial 6 h of the test show a small settling-in effect for CO₂ as well as for CO. Most likely this can be attributed to adjustments of the pressure regulator of the cylinder and the cell parameters. $\delta^{13}\text{CO}_2$ and CH₄ show a slight trend over the 6 days, persisting even after the sensitivity correction with 0.02‰ day^{-1} for $\delta^{13}\text{CO}_2$ and $-0.04\text{ nmol mol}^{-1}\text{ day}^{-1}$ for CH₄. The drift in $\delta^{13}\text{CO}_2$ might be related to a fractionation effect in the fast-emptying high-pressure cylinder; however, this cannot be proven since no pre- and post-mass-spectrometer measurements have been performed on this cylinder.

The same data set from September 2011 (Fig. 7) can be used to determine the repeatability of the FTIR measurements using Allan variance analysis (Werle et al., 1993). In Table 2, the 1σ repeatability for 2.5 and 10 min averaging time is given. An averaging time of 2.5 min is used in standard operating conditions for the Heidelberg in situ FTIR (3 min measurements including 30 s online analysis time). The 10 min repeatability is given for reasons of comparability to the earlier results from Griffith et al. (2010, 2012). Similar to the findings by Griffith et al. (2012), the repeatability of all species except for CO₂ initially improves with the square root of averaging time for at least 30 min. For longer integration intervals the repeatability still improves, but at a slightly lower rate. For sensitivity-corrected CO₂, the repeatability improves steadily up to an averaging time of 15 h, but only by one third of the square root of time. Detailed Allan variance plots based on our data can be found as Fig. 4 in Griffith et al. (2012).

We determined the reproducibility for measurements averaged over 3 min, as given in Table 2, by pooling three 1 min spectra and calculated their 1σ standard deviation. This reproducibility includes any potential changes or arbitrary drifts in the spectrometer or any sensor over the investigated 6-day period. The comparison of the repeatability and reproducibility of the 3 min averages in Table 2 emphasises the remarkably good short-term stability of the in situ FTIR.

5.2 Long-term stability of the in situ FTIR analyser

The long-term stability of the instrument response function (IRF) determines the calibration frequency as well as the calibration strategy. If the IRF is absolutely stable in time, interpolation between repeated calibrations does not improve measurement accuracy. This is due to the intrinsic uncertainty of each calibration measurement, which is then passed on to the measurements and increases their noise. If, on the other hand, the temporal changes of the IRF are larger than the instrument's repeatability, regular calibration improves the accuracy substantially. In the latter case the question concerning the required calibration frequency arises. In Sect. 5.1 we showed that, compared to the noise, for CO₂, CO and N₂O no systematic change of residual pressure and temperature sensitivity-corrected data occurred over the time scale of up to 6 days. Here we will investigate the longer time scales,

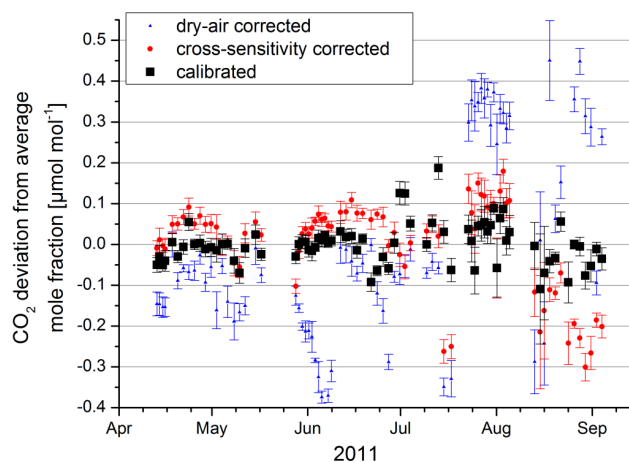


Fig. 8. CO₂ target gas record in its different data processing stages.

based on the data set we collected during the ICOS Demonstration Experiment. During these field campaigns, 24- and 48-hourly calibrations were performed with two standard cylinders, covering a suitable mole fraction range for all five components. In between the calibrations, a so-called target or surveillance tank was measured for quality control. In terms of sensitivity correction and calibration, the target cylinder was analysed in a similar way as any unknown sample from a cylinder. In the next section we will revisit the influence of the residual and cross sensitivities discussed in section 3 on the example of the CO₂ long-term target record.

5.2.1 Influence of the CO₂ residual and cross sensitivities on long-term records

Figure 8 displays the deviations of the CO₂ target measurement from the mean mole fraction over the different evaluation stages from the raw FTIR measurements to the finally calibrated data. Each target gas value consists of the average and the standard deviation of five sequentially recorded 3 min spectra. The dry-air mole fractions as calculated by MALT are shown in blue; they exhibit step changes as well as gradual changes. The 1σ standard deviation for the CO₂ target gas is $0.25\text{ µmol mol}^{-1}$ with a peak to peak variability of 0.8 µmol mol^{-1} over this 5-month measurement period. The prominent changes in the dry-air mole fraction can be related to changes in cell temperature and pressure, caused by changing laboratory conditions and/or a degradation of the flow controller that was additionally installed during the ICOS Demonstration Experiment. Correction for residual and cross sensitivities determined in Sect. 3 improves the reproducibility of the 24- to 48-hourly values as well as the 1σ scatter over the whole period by a factor of two to only $0.11\text{ µmol mol}^{-1}$ (red dots in Fig. 8). The residual and cross sensitivity correction takes care of most of the pronounced step changes; however, some outliers are persistent (e.g. in mid-July) and can thus not be explained by a change in one of our

Table 2. Repeatability (1σ) for 2.5 and 10 min averaging times deduced from Allan-Variance analysis. Reproducibility (1σ) for 3 min measurements of a single tank over a 6-day period and target gas reproducibility over the course of several months (compare Fig. 9). Total uncertainty for a single ambient air measurement (see Sect. 6).

Species	Repeatability (1σ)		Reproducibility (1σ)			Total uncertainty incl. errors in sensitivities (1σ)
	2.5 min	10 min	Single tank 6 days, 3 min	Target: Intermediate setup	Target: IUP setup	Single measurement
CO ₂ [$\mu\text{mol mol}^{-1}$]	0.018	0.012	0.023	0.043	0.016	0.032
$\delta^{13}\text{CO}_2$ [‰]	0.05	0.03	0.06	0.062	0.034	0.07
CH ₄ [nmol mol ⁻¹]	0.2	0.1	0.2	0.21	0.12	0.25
CO [nmol mol ⁻¹]	0.2	0.1	0.2	0.26	0.11	0.22
N ₂ O [nmol mol ⁻¹]	0.07	0.04	0.07	0.042	0.042	0.084

investigated residual and cross sensitivities. The variability in the sensitivity-corrected target gas measurements is still larger than the observed short-term repeatability expressed by the error bars and investigated in Sect. 3. In addition, the CO₂ variability still shows long-term changes and outliers. Thus, applying regular, e.g. daily, calibration might help to reduce the CO₂ variability further. By using a linear interpolation between the 24- to 48-hourly calibration measurements to determine the IRF, the 1σ standard deviation of the CO₂ target measurements is reduced to $\pm 0.05 \mu\text{mol mol}^{-1}$, as shown by black squares in Fig. 8.

Although the calibrated measurements are generally satisfying the WMO compatibility goal of $\pm 0.1 \mu\text{mol mol}^{-1}$ for CO₂ measurements in the Northern Hemisphere, individual measurement periods show much more scatter (e.g. in July). Outliers in the target gas record (July 2011) are caused by bad calibration measurements. The impact of a single calibration measurement depends largely on the strategy chosen to derive the IRF. We therefore investigated different calibration strategies such as (a) averaged IRF, (b) interpolated IRF between smoothed (e.g. moving weekly median) calibration measurements, and (c) interpolated IRF between neighbouring calibration measurements. Based on the repeatability of the target cylinder measurements, strategy (c) yielded the best results, although its sensitivity to single bad calibration measurements is largest. Careful selection of calibration outliers is thus recommended to minimise artefacts in the time series. For all following results we used calibration strategy (c), which is the linear interpolation of the IRF between neighbouring calibration measurements.

5.2.2 Long-term reproducibility for all species

Figure 9 comprises the sensitivity-corrected and calibrated target gas measurements for all components. In September 2011 the FTIR system was modified to the intermediate setup. The long-term reproducibility of the ICOS Demonstration Experiment setup can be judged based on the 1σ standard deviations up to September 2011: $\pm 0.06 \mu\text{mol}$

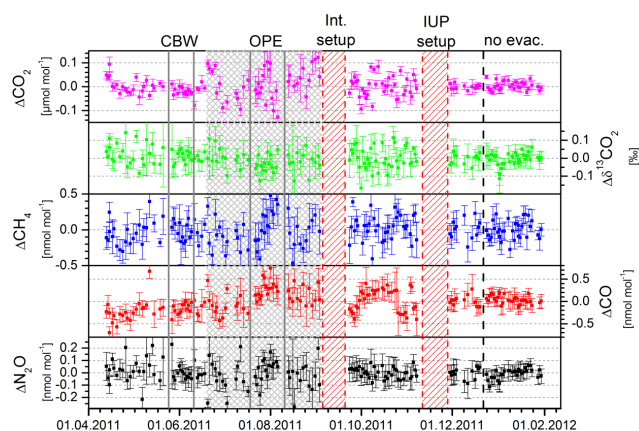


Fig. 9. Long-term stability of the FTIR analyser based on the 24- to 48-hourly target gas measurements. Vertical grey lines denote location changes of the instrument to Cabauw, the Netherlands (CBW), and Houdelaincourt, France (OPE). The grey shaded area highlights a period with less-stable cell pressure. Red shaded areas mark instrument down times due to modifications and/or laser failure. In September 2011 the internal mass flow controller was installed, and at the end of November 2011 the electronic pressure controller (EPC). The dashed black line indicates the change in standard operation conditions – from then on no evacuation was performed during sample exchange.

mol^{-1} for CO₂, ± 0.05 ‰ for $\delta^{13}\text{CO}_2$, $\pm 0.45 \text{ nmol mol}^{-1}$ for CO, $\pm 0.28 \text{ nmol mol}^{-1}$ for CH₄ and $\pm 0.1 \text{ nmol mol}^{-1}$ for N₂O. The 1σ standard deviations are thus close to or within the ILC targets for all trace gases (WMO, 2011). For $\delta^{13}\text{CO}_2$ the reproducibility of ± 0.05 ‰ is acceptable, keeping in mind that the in situ FTIR analyser is one of the first instruments delivering continuous $\delta^{13}\text{CO}_2$ measurements. The WMO-GAW requested target compatibility for $\delta^{13}\text{CO}_2$ is ± 0.01 ‰, and yet only met by very few mass spectrometer laboratories (Huang et al., 2011).

No significant drift was observed for any species. The small step change in CO is not explained by any of the investigated sensitivities. Re-calibration of the working standards

as well as the target tank at the Max Planck Institute for Biogeochemistry confirmed that neither the target cylinder nor the calibration cylinders drifted in any of the investigated species.

For CO₂ it is obvious that the performance of the in situ FTIR analyser declined after middle of June 2011. Checking the FTIR parameters revealed that from middle of June onwards, the standard deviation of the sample pressure increased from better than ± 2 hPa to more than ± 7 hPa. This increase was most likely caused by a progressive degradation of the additionally installed external mass flow controller (blue MFC in Fig. 1). In Sect. 3.3 we have shown that the largest pressure sensitivity corrections are found for CO₂; consequently, we observe the largest deviation between the short and long-term reproducibility for CO₂. The decrease in reproducibility for CO is caused by the step change on 15 June (compare Fig. 9) and for N₂O by several outliers.

The first red shaded area in Fig. 9 marks the re-building of the instrument to the intermediate setup. The reproducibility derived with this instrument configuration is given in Table 2. The largest improvement compared to the ICOS Demonstration Experiment setup, a factor of 2, was achieved for N₂O, improving the in situ FTIR analyser precision to well below the ILC target. The performance for CO₂, $\delta^{13}\text{CO}_2$ and CH₄ remains at a comparably good level. The drift, which was observed for CO, is caused by a drifting calibration gas cylinder being used during this time. Uncalibrated results suggest that the CO reproducibility improved as well.

To decouple sample pressure and flow, and to ease sample handling, an additional electronic pressure controller (EPC) (El-Press, Bronkhorst, The Netherlands) was installed at the inlet of the cell in the IUP setup. In Fig. 9 the target gas measurements performed with this new configuration are shown after the second red shaded area. The reproducibility of the sensitivity-corrected and -calibrated target gas results is also summarised in Table 2. The introduction of the additional EPC improved the reproducibility of all components except for N₂O by approximately a factor of two. The dashed line in Fig. 9 in mid-December 2011 indicates a change in our standard operating conditions. From there on we skipped the evacuation step during sample exchange and used the flushing approach (refer to Appendix A). This was done in order to investigate the benefits of not disrupting the temperature and moisture equilibrium in the cell by the evacuation. In Fig. 9 no significant difference between both sample change-over strategies is visible. Thus, after two months we changed back to our SOC to save calibration gas and to shorten the calibration measurement interval. This finding depends on the temperature sensor type used. It will most likely not be valid for the slow-responding RTD temperature sensor.

In the final IUP setup short and long-term reproducibility are similar for all components (see Table 2). These results demonstrate that the long-term stability of the in situ FTIR analyser is suitable for background air monitoring if mea-

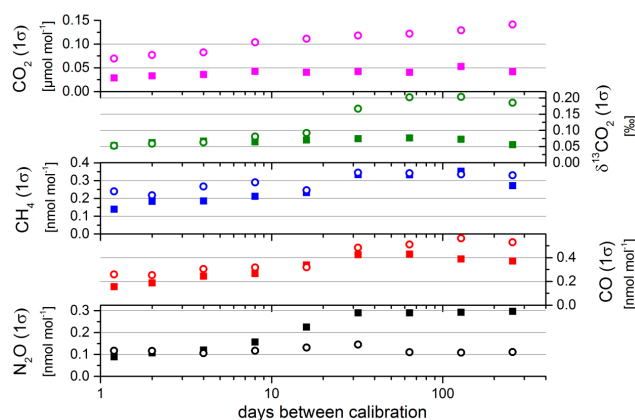


Fig. 10. Calibration frequency test. 1σ standard deviation of the target gas measurements as a function of days between calibrations. Open symbols represent less-stable instrument conditions with sample pressure variations (1σ) of more than ± 7 hPa. Filled symbols relate to stable conditions with sample pressure variations (1σ) of less than ± 2 hPa.

surements are calibrated and properly corrected for residual and cross sensitivities.

5.3 Calibration frequency

The target gas record can further be used to estimate the influence of the calibration frequency on the reproducibility. In Fig. 9 we used the highest calibration frequency of 24- to 48-hourly calibrations with two standard gases. With the target gas data set, we can investigate the influence of stepwise prolonged calibration intervals on the 1σ reproducibility of the results. Since, especially for CO₂, the performance of the FTIR analyser can be separated into two periods – stable and less-stable conditions (compare grey shaded area in Fig. 9) – we will evaluate the calibration frequency test for both stability conditions separately. The less-stable period, without proper control of the additional MFC introduced in the ICOS Demonstration Experiment setup, the instrument configuration is comparable to the initial UoW setup.

In each panel of Fig. 10 the relation between 1σ reproducibility and calibration frequency is plotted. Closed symbols represent stable instrument conditions with a cell pressure variability of less than ± 2 hPa, open symbols show less-stable conditions; here, the cell pressure variability was four times larger. Under stable instrument conditions the 1σ target reproducibility of CO₂ was better than $\pm 0.03 \mu\text{mol mol}^{-1}$ for 24- to 48-hourly calibrations. Prolonging the calibration frequency even to 2 or 3 months changes the reproducibility only slightly to $0.05 \mu\text{mol mol}^{-1}$. During less-stable instrument conditions, the reproducibility improves steadily with increasing calibration frequency; however, even 24- to 48-hourly calibrations do not result in the reproducibility of the stable conditions. It seems that under less-stable conditions, substantial variations occur on sub-daily time scales,

for example during the campaign in OPE the laboratory temperature changed by 10 °C within 1 day.

The other trace species, apart from N₂O, show a similar behaviour to that of CO₂, although the difference between stable and less-stable instrument conditions is not as pronounced. For the highest calibration frequency, the achieved reproducibility is on par or even better than the short-term reproducibility determined in Sect. 5.1. This is caused by the fact that the investigated target measurements as well as the calibration measurements are averaged values of five individual spectra reducing the scatter by a factor of $\sqrt{5}$. This implies that for a single 3 min ambient air measurement the reproducibility is a factor of $\sqrt{5}$ larger than the one shown for the target measurements. In contrast to the IRF of other gases, the IRF for N₂O has two large-step changes during the stable instrument conditions (not shown) leading to the inverse results in the calibration frequency investigation. The recorded sample and instrument properties give no evidence that could explain the step changes in the N₂O IRF.

Increasing the calibration frequency from weekly to 24- or 48-hourly improves the reproducibility on average by 20 % for all gases. Depending on the specific accuracy needs, weekly calibrations should be sufficient and beneficial for calibration gas consumption as well.

6 Error assessment for ambient air measurement

Different sources of error contribute to the total error of an individual ambient air measurement. These error contributions are partly systematic and partly random and can often not easily be separated. Obvious error contribution arises from (a) measurement repeatability, (b) uncertainties in residual and cross-sensitivity corrections, (c) uncertainties introduced by the two-point calibration, and (d) uncertainty of the assigned mole fraction of the calibration cylinders themselves.

These error sources are not independent from each other. For example, the uncertainties originating from (a) and (b) (repeatability and the residual and cross-sensitivity corrections) feed directly into the uncertainty introduced by the regular calibration. Thus, a combined uncertainty comprising (a), (b) and (c) can be derived from the reproducibility of the target measurements as listed in Table 2. However, the target measurement reproducibility is derived from averaged values and has thus to be multiplied by a factor of $\sqrt{5}$ (since there are 5 target measurements). For ambient air measurements the uncertainties of the CO₂ cross sensitivities must be added since this error component is not present in the target measurements. The total uncertainties are thus given in the last column of Table 2. The contribution of the uncertainties of the assigned values of the calibration cylinders is an additional, systematic error component that has to be accounted for separately.

7 Conclusions and outlook

The modified in situ FTIR spectrometer is well suited for GHG monitoring and fulfils the precision, accuracy and stability needs for CO₂, CO, CH₄ and N₂O measurements at background sites. The reproducibility for $\delta^{13}\text{C}\text{CO}_2$ does not formally fulfil the ILC targets set by WMO-GAW; however, 0.03 ‰ target reproducibility (0.07 ‰ single measurement) is still very good for a continuously measuring instrument. The final IUP setup of the in situ FTIR is additionally equipped with an internal EPC and MFC to control sample pressure and flow rate. To improve the temperature measurement the RTD sensor was replaced by a thermocouple that is located in the middle of the cell to improve the representativeness of the measurement. These improvements lead to a factor-of-two-better repeatability for all species. In the IUP setup the effects of the residual pressure sensitivity and flow rate-sensitivity are negligible. The same is true for the H₂O cross sensitivity since the residual moisture variations can be restricted to only a few $\mu\text{mol mol}^{-1}$ of H₂O. Reducing the causes of variability is always superior to any post-processing correction. However, for those parameters where this is not possible, e.g. CO₂ cross sensitivity, precise determination of these parameters is essential. Currently, the largest remaining issue is related to the ability of measuring the *true* sample temperature in the cell. Resolving this temperature problem might allow for measuring calibration and target gas cylinders in static mode, which would reduce the gas consumption by a factor of five. Measuring the sample temperature at multiple locations could be a first step towards improving the representativeness of the temperature measurement. The small Reynolds number in the cell (≈ 10 , see Appendix A) for our standard operating conditions constitutes a more general problem. Improved sample delivery to the cell that increases turbulence would help minimizing temperature gradients within the cell. We also recommend placing the Nafion drier inside the temperature-controlled enclosure since it is important for thermal sample pre-conditioning.

The instrument has proven to be acceptably linear for all components in the ambient concentration range (also for non-background conditions); however, the instrument response functions have a non-zero offset for all components, implying the need for at least two calibration gases. The in situ FTIR analyser is sufficiently stable to run with weekly calibrations only. In standard operation conditions, almost 30 L of air are needed for one cylinder measurement. Thus, a 50 L cylinder pressurised to 20 MPa lasts for more than four years, even if 10 to 20 % of the gas is remaining in the cylinder to avoid potential drifts of components such as CO₂ (Kitzis, 2009; Langenfelds et al., 2005). Although the lifetime of a FTIR calibration gas cylinder is thus longer than that for classical GC systems (typical lifetime ≈ 1 yr), it is shorter than the expected calibration gas cylinder lifetime for other optical techniques like quantum cascade lasers (QCL) or CRDS.

For the laser techniques the calibration cylinder lifetime is expected to be on the order of decades, depending on the calibration scheme (e.g. Winderlich et al., 2010). For conservative, high-accuracy applications, one may want to have a full year of overlap between two calibration gas generations, resulting in an effective calibration gas lifetime of three years. For high-accuracy applications we recommend the use of three calibration cylinders, spanning the range of expected ambient concentrations. The usage of three calibration gases instead of the minimally needed two reduces the sensitivity to individual outliers and instrument noise. In any case, we recommend daily target or surveillance gas measurements for quality control. The lifetime of a daily target gas is, unfortunately, limited to nine months only; thus, we recommend a second sub-target being measured on a two-weekly basis only. The sub-target can then also be used to inter-connect two to three standard gas generations.

Comparing the in situ FTIR to other optical state-of-the-art greenhouse gas analysers such as Cavity Ring Down Spectroscopy, Off-Axis Integrated Cavity Output Spectroscopy or quantum cascade lasers shows that the performance of the in situ FTIR is comparable to all other techniques for all five species. Currently, no other technique is able to provide measurements of CO₂, δ¹³C, CO, CH₄ and N₂O with a single instrument. The gas consumption and the sample change-over time of all other techniques is, however, smaller. This is beneficial for the conservation of calibration gas as well as fast sample exchanges, e.g. at atmospheric tower sites with multiple inlet heights measured with one single instrument. Residual and cross sensitivities are a general issue for all optical GHG analysers and are subject to ongoing investigation (e.g. Rella et al., 2013; Zellweger et al., 2012; Vogel et al., 2013; Chen et al., 2012). Providing a comprehensive comparison of the different techniques for all species is, however, beyond the scope of this paper. A comparison study of recent CO measurement techniques can be found by Zellweger et al. (2012), for the performance of other species and techniques, please refer to individual papers, e.g. Winderlich et al. (2010) for CO₂ and CH₄, McManus et al. (2008) for N₂O, and Vogel et al. (2012) for δ¹³C.

Appendix A

Sample exchange and thermodynamic equilibrium in the cell

Measurement accuracy depends on the instrument precision as well as on the complete exchange of the gas sample in the cell without memory effects. Thus, the cell has to be either evacuated or flushed for a sufficiently long time to completely remove the previous sample. We will briefly discuss both approaches:

- a. Flushing: the mean exchange time τ of a well-mixed volume is given by the ratio of volume to flow rate. The

influence of the preceding sample decreases according to $\exp(-t/\tau)$, where t is the flushing time. Thus, we need more than $t = 8 \tau$ to reduce the memory effect to less than 0.03 %, which in the case of a 100 μmol mol⁻¹ change in CO₂ between the two samples results in a 0.03 μmol mol⁻¹ memory effect. With the given cell volume and the standard flow rate of 1 slpm, a flushing time of 28 min is thus theoretically required. An example of this change-over method is illustrated in Fig. A1 (black squares). In this example the equilibrium value, which is defined as the average mole fraction measured from minute 30 to 45 is reached after 21 min, with a mole fraction difference between the two samples of 140 μmol mol⁻¹.

- b. Evacuation: to reach comparably small memory effects as under (a), the cell and the inlet system need to be evacuated to < 0.3 hPa, which, with the available equipment, is not reached within 30 min. Thus, we decided to use a two step evacuation procedure: the cell is first evacuated to 10 hPa, then filled with the new sample to 500 hPa, evacuated a second time down to 10 hPa and subsequently filled to the desired cell pressure of 1100 hPa. This stepwise change-over is much faster than a one-step evacuation and ensures that the memory effect is less than 0.02 %. The complete sample change-over procedure requires 7 L of gas and takes 8 min, including 1 min stabilisation time after the final pressure and flow settings are reached. An example of the two-step evacuation procedure is also shown in Fig. A1 (blue triangles).

Using the two-step evacuation procedure for sample exchange, close-to-equilibrium values are already observed 12 min after a sample change-over. As evacuation and re-filling affects the temperature of the cell, the first two measurements (taken in minutes: 6 to 9 and 9 to 12) have to be discarded.

However, exchanging the sample entirely, and thus avoiding memory effects, is not necessarily sufficient to avoid transient settling-in effects after sample change-over. The thermodynamic properties of the sample, such as temperature and pressure, need to reach their equilibrium conditions as well. Sample temperature is especially crucial since, in contrast to pressure, we cannot assume that temperature is homogeneously distributed within the cell. This may have several causes: (a) different sample temperatures when entering the cell; (b) the spectrometer heats one end of the cell, causing a small but significant temperature gradient along the cell; (c) the Reynolds number of the cell calculated for our SOC is only about 10, and thus mixing is far from turbulent, leading to a persistence of any temperature gradient. For a tube geometry the Reynolds number can be calculated as $Re = (u_m d)/\nu$, with u_m : mean velocity of the gas ($1 \times 10^{-3} \text{ m s}^{-1}$) in the cell, d : diameter of the cell (0.15 m) and ν : kinematic viscosity of air ($1.5 \times 10^{-5} \text{ m}^2 \text{ s}^{-1}$).

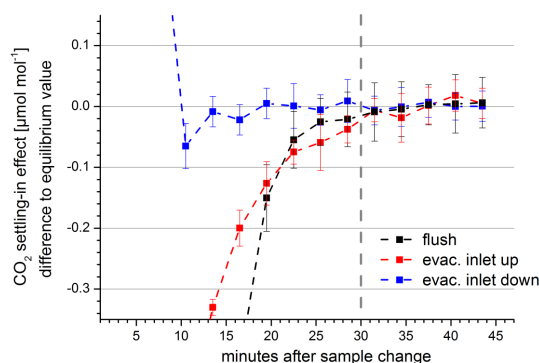


Fig. A1. Comparison of the different sample change-over methods for CO₂. The settling-in effects are shown as relative derivation from the equilibrium value defined by the average of the last five measurements. Sample exchange by flushing only is shown in black. Sample exchange via evacuation, as used in the standard operation conditions, is in blue with standard cell orientation and in red with the cell turned upside down. Measurements are shown centred in their 3 min interval.

The impact of thermodynamic disequilibrium is shown in Fig. A1 by the red symbols. These measurements have been performed using the same cylinder, regulator and evacuation procedures as for the data indicated in blue; however, the cell was rotated by 180°. Turning the cell upside down changed the position of the temperature sensor as well as that of the 1/4" (6.35 mm) dip tube that delivers the sample into the cell. For the measurements plotted in red, the dip tube was at the top of the cell, and the temperature sensor was at the bottom. We can assume that the memory effect caused by incomplete sample exchange is similar for both positions since the same two-step evacuation approach was used, but due to the different location of the sample inlet and/or the temperature sensor the sample apparently takes longer until it reaches its thermodynamic equilibrium. Since this settling-in effect is seen for all components, it seems likely that it might be related to temperature. The measured sample temperature between the two cell positions changed by 0.5 °C; higher temperatures were observed with the temperature sensor in the upper position. Thus, we can conclude that temperature is not homogeneously distributed in the cell, and that we are not able to measure the true mean sample temperature with one temperature sensor only. However, as long as the temperature distribution in the cell is stable under standard operation conditions and is reached for both ambient air and cylinder measurements, the calibration of the instrument will compensate for the error in temperature measurement.

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