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Abrupt transitions between macrobenthic faunal assemblages across seagrass bed margins

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Abrupt transitions between macrobenthic faunal assemblages across seagrass bed margins

Abstract

The nature of the transition from one contrasting macrobenthic assemblage to another across interfaces between intertidal seagrass and unvegetated sand was investigated in the subtropical Moreton Bay Marine Park, eastern Australia, via six two-dimensional core lattices. The same pattern of transition was manifested in each lattice. Macrofaunal abundance, species density (both observed and estimated total) and assemblage composition did not vary with distance away from the interface within the 0.75 m wide marginal bands of each habitat type. Neither were there significant differences in assemblage metrics or composition between the marginal and non-edge regions of either habitat. There were, however, very marked differences in assemblage composition, abundance and species density across the 25 cm wide strip on either side of the actual interface, the interacting assemblages reacting symmetrically. All these differences therefore took place over an ecotone distance of only 0.5 m at most. Spatial trends in assemblage metrics across the boundary zone were captured accurately by second and third order polynomial regression models. It also appeared that edge effects on individual species within the seagrass were a variable local response not a consistent effect of closeness to the bare sand.

Keywords

bed, seagrass, across, assemblages, margins, faunal, abrupt, macrobenthic, between, transitions

Disciplines

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13				
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21 not vary within the 0.75 m wide marginal bands of each habitat type with

1 distance away from the interface. Neither were there significant differences 2 in assemblage metrics or composition between the marginal and non-edge regions of either habitat. There were, however, very marked differences 3 4 across the 25 cm wide strip on either side of the actual interface, the 5 interacting assemblages reacting symmetrically. All these differences in 6 composition, abundance and species density therefore took place over an 7 ecotone distance of only 0.5 m at most. Spatial trends in assemblage 8 metrics across the boundary zone were captured accurately by second and 9 third order polynomial regression models. It also appeared that edge effects 10 on individual species within the seagrass were a variable local response not 11 a consistent effect of closeness to the bare sand.

12

13 Key words

Benthos, biodiversity, ecotone, edge effects, sandflat, seagrass

16 **1. INTRODUCTION**

Most natural environments, including coastal marine ones (Eyre et al., 2011), are mosaics of different habitat patches and the transitional zones between these patches are increasingly being recognised as ecologically important, not least because landscape structure affects habitat quality (Levin et al., 2001; Zajac et al., 2003; Ries et al., 2004; etc.). The quality of seagrass habitats is of particular current concern because of their recent worldwide decline and fragmentation (Lotze et al., 2006; Waycott et al., 2009;

Fourgurean et al., 2012), because of concerns that the resultant denuded 1 2 habitat may support lesser animal abundance and biodiversity (e.g. Pillay et 3 al., 2010), and because seagrass beds provide economically important 4 ecosystem services, including not only physically protective habitat for the 5 juveniles of commercially-significant fish and crustaceans but also food for 6 those juveniles in the form of the smaller benthic macrofauna that 7 seagrasses support in abundance (Duarte, 2000; Unsworth and Cullen, 2010; Coles et al., 2011; Barbier et al., 2011). 8

9 Throughout much of the world, sheltered intertidal and shallow 10 subtidal sandflats occur in one of two characteristically alternative patch 11 states: one being seagrass beds, the other unvegetated sediment. These two 12 alternatives are dynamic and interchangeable, with each member of the pair 13 expanding into territory held by the other in one area or another of shared 14 sandflats (e.g. Yamakita et al., 2005; Berkenbusch et al., 2007; Carr et al., 15 2010). Although not always the case in vegetated versus unvegetated 16 comparisons, especially in high latitude areas subject only to Arenicola 17bioturbation (see e.g. Asmus and Asmus, 2000; Polte et al., 2005), wherever 18 the bare sediment is structured by the bioturbation of burrowing 19 thalassinidean crustaceans (Pillay and Branch, 2011) benthic macrofaunal 20 assemblages supported by the two alternative habitat types may be very 21 different, as, for example, in the Nanozostera capensis versus Callichirus 22 kraussi system in Langebaan, South Africa (Siebert and Branch, 2007) and 23 in the Nanozostera muelleri versus Trypaea australiensis system in Moreton 24 Bay, Australia (Barnes and Barnes, 2012). At these localities, as well as at

others (Boström and Bonsdorff, 1997), areas of bare sediment are dominated
by burrowing species whilst the seagrass supports a large epifaunal
component. Such faunal contrasts can often be linked to the marked
change in nature and stability of the substratum induced by the contrasting
ecosystem bioengineers (Berkenbusch and Rowden, 2007; Siebert and
Branch, 2007; etc.).

7 Some aspects of the boundary zone between seagrass and adjacent 8 habitat types have received detailed attention. In particular, edge effects 9 within blocks of seagrass have been extensively studied in both the Atlantic 10 and the Pacific (Bologna and Heck, 2002; Tanner, 2005, 2006; Warry et al., 11 2009; Murphy et al., 2010; etc), with, somewhat paradoxically, it often being 12 reported that densities of seagrass-associated animals were higher on the 13 margins of the bed than nearer its centre. Equivalent work on edge effects in 14 unvegetated sediment is much rarer, however. The studies by van Houte-15 Howes et al. (2004) and Tanner (2005) are two of the very few that have 16 extended right across an interface to examine both two habitat types. Van 17Houte-Howes et al. (2004) explored the boundary between mid-intertidal 18 unvegetated sand and Nanozostera muelleri in North Island, New Zealand, 19 and they reported evidence that distinctive assemblages were present at the 20 edges of the seagrass (at least under conditions of abundant seagrass 21 shoots). These authors investigated the transitional zone via a series of sampling points relatively far apart and spanning the relatively large total 22 23 distance of 100 m (at -50, -10, -1, +1 and +50 m, where zero is the interface 24 itself). Tanner (2005), from the perspective of the abundance of certain

seagrass-associated species in fragmented systems, narrowed the zone of
investigation to the 2 m on either side of the bare sand/seagrass boundary
and showed marked changes in population density of various seagrass
polychaetes, crustaceans and bivalves across this zone in a *N. muelleri*meadow at LWS in South Australia.

6 In order to examine in detail the actual transition from seagrass faunal 7 assemblage to that of the adjacent unvegetated sediment, the present study 8 was designed to investigate the precise interface between the two. Further, 9 this was conducted at a locality for which recent data on the contrasting 10 faunal assemblages of the seagrass and unvegetated sediment away from the 11 interfaces were also available (Barnes and Barnes, 2012), permitting each 12 marginal habitat zone to be placed in context. Three specific questions were 13 addressed. (i) Do faunal transitions between seagrass and bare sediment 14 take the form of sharp ecotones or of more gradual ecoclines (sensu Attrill 15 and Rundle, 2002; Yarrow and Marín, 2007)? (ii) Do the two interacting 16 faunal assemblages react symmetrically to habitat change across the 17interface? (iii) What is the effect of the transition on the magnitude and 18 spatial variation of local benthic abundance and biodiversity?

19

20 2. METHODS

21 2.1. STUDY AREA, SAMPLE COLLECTION AND PROCESSING

Macrofaunal sampling was conducted over a period of 11 weeks during
the 2012 austral spring along the sheltered Rainbow Channel western coast

1 of North Stradbroke, a large (27,400 ha) sand-dune barrier island in the 2 relatively pristine Eastern Banks region of the oligohaline, sub-tropical 3 Moreton Bay Marine Park, Queensland (Dennison and Abal, 1999). The 4 Eastern Banks contain the majority of the Bay's 190 km² of seagrass 5 (Roelfsema et al., 2009) and support its greatest numbers of animal 6 species, including many southern outliers of the tropical Great Barrier Reef 7 fauna (Davie and Hooper, 1998). Here, for example, the seagrass is the 8 refuge and feeding ground for juveniles of many species of penaeid prawns 9 and fish as well as for green turtle and dugong (Weng, 1990; Davie et al., 10 2011). In the specific area sampled, the beds are predominantly of the 11 dwarf-eelgrass Nanozostera muelleri capricorni (= Zostera capricorni = 12 Zosterella capricorni) with some Halodule uninervis (especially at lower shore 13 levels) and Halophila ovalis, and the seagrass extends from the sublittoral 14 right up into the mangrove zone amongst the seaward pneumatophores of 15 Avicennia marina, i.e. over an intertidal vertical height of half the tidal range 16 (i.e. over c. 1 m) and in places for a distance of >500 m. Typically, the 17seagrass plants are of the smaller morphological forms characteristic of 18 shallow areas (Young and Kirkman, 1975). Also within the seagrass zone 19 are unvegetated areas of the fine- to medium-grained quartz sand that 20 comprises the island (Laycock, 1978), often occurring as a series of large 21 patches from about LWN to MSL. Such areas are structured by two 22 bioturbating decapods, the thalassinidean Trypaea australiensis and soldier 23 crab Mictyris longicarpus, both dependent on sedimentary diatoms 24 (Spilmont et al., 2009). The same sand, with or without a surface coating of 25 mud, underlies the upper-shore mangroves.

1 Data on the macrobenthic assemblages were collected from three sites 2 over a distance of 2.5 km centred on the Moreton Bay Research Station at Dunwich — (a) Deanbilla, (b) Polka, and (c) Yerrol (Fig. 1) — roughly in the 3 4 middle of a virtually uninterrupted 25+ km long belt of seagrass. Precise 5 sampling sites were located where the margins of the seagrass beds 6 appeared neither to be advancing nor retreating to avoid possible 7 complications resulting from temporally transitional states. Advancing 8 seagrass was identified by lines of young plants extending out from the bed 9 and retreating seagrass by the occurrence of dead root-rhizome mats 10 beneath the surface of the adjacent bare sand. As advocated by Fortin 11 (1994), in all cases data were collected from two-dimensional lattices across 12 and along the bare sand/seagrass interface (see Fig. 1). Six replicate 13 lattices were worked, each comprising 60 core samples in the form of 6 14 transects ('rows') worked parallel to the interface, at +0.75, +0.5, +0.25, -15 0.25, -0.5 and -0.75 m from it (where '+' indicates seagrass and '-' indicates 16 unvegetated sand), and 10 core samples ('columns') taken at 1 m intervals 17along each transect; this interval allowed for the high level of small-scale 18 spatial variability previously seen in such systems (Winberg et al., 2007; 19 Barnes and Ellwood, 2012). Interfaces were sampled at three intertidal 20 heights, near LWS, LWN and MSL, each replicated at two different sites 21 (LWS and LWN at both Deanbilla and Polka; MSL at Deanbilla and Yerrol). 22 At LWS and LWN the interfaces were between seagrass and unvegetated 23 open sandflat, and near MSL they were between seagrass and the sand 24 adjacent to the seaward zone of Avicennia pneumatophores. As noted at other localities (e.g. Bryan et al., 2007; Carr et al., 2010), the boundary 25

between N. m. capricorni beds and blocks of unvegetated sand can be very 1 2 sharp, and the sites near LWS and LWN had such sharp boundaries (Fig. 3 2). The seagrass/ pneumatophore-field interface near MSL, however, was 4 mostly more diffuse, broken up into small patches, and with an extensive 5 overlap of up to 20 m, although in places the seagrass had its upper limit 6 as a sharp boundary immediately below the pneumatophore-zone edge and 7 it was in these areas that the two upper-shore lattices were located. With 8 one exception, all lattices were located in areas where large continuous 9 blocks (>0.5 ha) of the two habitat types met to avoid the potentially 10 confounding variable of patch size (Bowden et al., 2001; Mills and 11 Berkenbusch, 2009). However, as previously (Barnes and Barnes, 2012), 12 the core samples near MSL at Yerrol had to be located on the margin of a 13 patch of seagrass isolated from the lower shore bed by an intervening 14 stretch of bare sand. Coordinates of each lattice were taken by means of a 15 hand-held GPS+GLONASS unit (with a stated accuracy of 3 m). To 16 compare transitional zone assemblages and those from within the main 17blocks of the relevant habitat types, equivalent data on the nature of faunal 18 assemblages at each locality >4 m away from any boundary zone were 19 extracted from the earlier database of Barnes and Barnes (2012). These 20 data were from samples taken 12 months previously, also from transects of 21 10 replicate cores.

All core samples were of 54 cm² area and 10 cm depth on the basis that (a) most benthic macrofauna in seagrass is known to occur in the top few mm of sediment (e.g. 98% in the top 5 mm in the study by Klumpp and

Kwak, 2005), and (b) earlier work (Barnes and Ellwood, 2012) had 1 2 suggested that 10 replicates of that surface area would yield an acceptable 3 standard error <17.5% of the arithmetic mean in estimation of local 4 macrofaunal abundance. This sampling procedure collects the smaller and 5 most numerous members of the macrofauna that constitute the large majority both of invertebrate biodiversity (Albano et al., 2011) and of the 6 7 food of the commercially-significant juvenile nekton (O'Brien, 1994), but not 8 the scarcer megafauna or the deeply-burrowing species that include the 9 ecosystem bioengineers Trypaea (that burrows down to depths of 1m) and 10 *Mictyris* (that vacates its burrow during the first part of low tide), although juveniles of both species were frequently obtained. Trampling adversely 11 12 impinges on seagrass systems (e.g. Eckrich and Holmquist, 2000) and 13 hence great care was taken to cause minimal disturbance to the site, 14 particularly to areas to be sampled on future occasions.

15 Collection and treatment of these core samples essentially followed the 16 same standard procedure as earlier studies of macrofaunal assemblages 17associated with Nanozostera beds both within the North Stradbroke 18 intertidal (e.g. Barnes and Barnes, 2011, 2012) and elsewhere (e.g. Barnes 19 and Ellwood, 2011; Barnes, 2013). Cores were collected at low tide, soon 20 after tidal ebb from the sites, and these were gently sieved through 710 µm 21 mesh on site. All sieved samples were immediately transported to a local laboratory, where each was placed in a 30 x 25 cm white tray in which the 22 23 living fauna was located by visual inspection and was counted; the time 24 interval between collection and analysis of any given batch of samples never

1 exceeding 24 hours. This continued until no further animal could be seen 2 during a 3-minute period. Animals were identified to species level wherever 3 possible, with nomenclature as listed in the World Register of Marine 4 Species (WoRMS, www.marinespecies.org). Several taxa, however, although 5 relatively important in Moreton Bay, have not yet been investigated 6 systematically in southern Queensland (Davie and Phillips, 2008) and their 7 identification to named species poses 'enormous problems' (Davie et al., 8 2011, p. 8). Such animals were treated as morphospecies, an operationally 9 appropriate procedure to detect spatial patterns of numbers of species and 10 their differential abundance (Dethier and Schoch, 2006; Albano et al., 11 2011). The number of seagrass shoots in each sample was also recorded, 12 but sessile macrofauna attached to the seagrass leaves was not.

13 2.2. STATISTICAL ANALYSES

14 Raw data on numbers of each component species in the various sampling horizons at different distances from the actual interface were 15 16 subjected to similarity analysis and various assemblage metrics were 17derived and compared across the same distances, both univariately by ANOVA and via spatial analyses. Assessment of faunal similarity and of 18 19 univariate changes in biodiversity metrics were carried out via *EstimateS* 20 8.2.0 (Colwell, 2011; Colwell et al., 2012). Given the high proportion of rare 21 species in the local seagrass fauna [29% of the 184 species obtained from 22 the boundary zone were represented by single individuals, and a further 23 14% by doubletons], estimates of total numbers of species (S_{max}) per unit 24 area (i.e. of species density sensu Gotelli and Colwell, 2001) were derived

1 from the observed numbers of species (S_{obs}) and their differential 2 abundance. Different estimators of true species density yield different 3 results, hence each value of S_{max} was obtained by taking the mean of three disparate EstimateS measures: Chao-2, Michaelis Menten Means, and 4 5 Abundance-based Coverage, using, where appropriate, the bias-correction 6 formula for Chao-2 and an upper abundance limit of 5 for infrequent 7 species for the Abundance-based Coverage estimator. To permit 8 quantitative comparison (Jost, 2006), both α and γ species diversities were 9 assessed as the linear 'effective number of species' by the reciprocal of the 10 Hill (1973) N_2 . Also because of the high probability of absence of species 11 present at low population density from any given sample, changes in 12 quantitative faunal composition (β -diversity) across adjacent transitional 13 horizons were estimated by the Chao et al. (2005; 2006) abundance-based 14 Sørensen (Bray-Curtis) (dis)similarity statistic, as modified to correct for 15 undersampling bias, again with a set upper abundance limit per sample for 16 rare species of 5. This modification substantially reduces the negative bias 17of traditional similarity indices, especially when samples from species-rich 18 assemblages are likely to be incomplete (Chao et al., 2005). Estimated 19 numbers of shared species between different assemblages were likewise 20 obtained from Chao et al.'s (2000) coverage-based estimator. Univariate 21 comparison of these assemblage metrics at various distances from the 22 interface, i.e. species density, total faunal abundance and dissimilarity, 23 used one-way or two-way ANOVA after ln[x+1] transformation of data, 24 followed by post-hoc Tukey HSD tests where applicable.

1 Spatial analyses were executed using a combination of GeoDa 2 software and the Geostatistical Analyst tools within ArcGIS10. Spatial 3 statistical techniques employed were cross-scale trend analysis, calculation 4 of the global Moran's I statistic at both local and regional scales, and 5 calculation of a semivariogram and associated semivariogram surface for 6 each lattice. Trend analysis was undertaken to ascertain whether 7 transitions in the four assemblage metrics could accurately and 8 consistently be captured in a simple spatial model. Trends were explored at 9 both local (lattice) and regional (all sites) scales by plotting values of the 10 metrics along an axis representing distance from the bare sand/seagrass 11 interface. Local scale analysis examined mean values from all sites along 12 the transition axis, whilst regional scale analysis retained the relative 13 geographical positions of each site. A series of models was developed 14 (linear, first, second and third order polynomial) to express the observed 15 changes in each metric at both regional and local scales. The accuracy of 16 each model was expressed as the percentage of variation of the observed 17distribution explained by the model (R^2). To assess cross-scale consistency 18 in models, the most accurate model type was recorded at both local and regional scales. 19

To assess the strength of spatial relationships between the macrofaunal assemblages, the global Moran's *I* statistic was calculated on each of the four metrics as a measure of spatial autocorrelation between point sample locations. This was calculated for specified distance classes as the cross product for a given assemblage metric at points across a

1 defined neighbourhood such that only the pairs of points within a specified 2 distance class of each point location were taken into account (Hamylton, 3 2013). This yields a large positive I in the presence of positive spatial 4 dependence, a large negative *I* in the presence of negative spatial 5 dependence, and values close to zero with a random assemblage 6 distribution. Spatial autocorrelation was also assessed at the local scale by 7 generating a semivariogram for each lattice. This calculated Geary's C 8 statistic on the total number of individuals sampled to assess similarity for 9 every possible combination of pairs of sample points across each lattice grid 10 (Hamylton, 2013). Geary's C measures correspondence for a given distance 11 class on the basis of the squared difference of a particular characteristic 12 between pairs of point locations, and ranged from 0 to some unspecified 13 value larger than 1. Values of Geary's C were then plotted against the 14 distance between points to generate a semivariogram exploring the 15 magnitude and spatial configuration of locally-measured autocorrelation 16 plotted across the map. The distance at which Geary's C levelled off and no 17further increase in the statistic was observed as the distance between point 18 pairs increased, the sill point, was recorded for each semivariogram. This 19 corresponds to a given distance range beyond which there was no spatial 20 autocorrelation. Finally, a semivariogram surface was generated to map 21 the value of the Geary's *C* across each lattice as a localized measure of 22 assemblage metric similarity in geographical space. To account for the 23 anisotropic effect across the tidal axis, a search direction of 90° was used to 24 define the neighbourhood of each point. The surface was generated using 25 an interpolation technique that combined values in the semivariogram plot

into bins based on the direction and distance between pairs of point sample
 locations. Binned values were then averaged and smoothed to produce the
 semivariogram surface.

4

5 3. RESULTS

6 Faunal assemblages of the adjacent 0.75 m wide boundary zones on 7 either side of the interface between seagrass and unvegetated sand at each 8 lattice showed markedly different abundance and species density (all one-9 way ANOVA $F_{1,34}$ >60; *P* << 0.0001). Overall, the marginal strip of seagrass 10 supported an observed total of 168 species (estimated true total 220) at an overall abundance of 2,430 ind. m^{-2} and a γ -diversity of 16.0. The abutting 11 12 marginal strip of sand, in contrast, supported only 61 observed species 13 (estimated true total 78), 750 ind. m^{-2} , and a γ -diversity of 11.6. Even 14 though contiguous, the two assemblages shared only 24% of their observed total fauna and only 44% of their higher-level taxa (at the level of 15 suborder/superfamily). The relative richness of the seagrass assemblage 16 17was not a consequence of its greater abundance (Gotelli and Colwell, 2001) 18 since the number of species per macrobenthic individual within the seagrass 19 was always considerably larger than that in the unvegetated sand (at a ratio 20 of 1 : >1.5) (Fig. 3a). Per unit area the ratio was close to 1 : 2.7. The 21 seagrass-edge assemblage was dominated numerically by the rissooid 22 Calopia imitata (434 m⁻²), grapsoid Enigmaplax littoralis (300 m⁻²), 23 phoxocephaloid *Limnoporeia* sp. (191 m⁻²), buccinoid *Nassarius burchardi* (109 m⁻²) and apseudoid *Longiflagrum caeruleus* (100 m⁻²), which were the 24

1 only species to contribute $\geq 4\%$ of the total individuals. In the unvegetated 2 sand the same percentage was contributed by each of the haustorioid 3 Urohaustorius mertungi (174 m⁻²), grapsoid Mictyris longicarpus (65 m⁻²), 4 galeommatoid Mysella vitrea (61 m⁻²), cerithioid Finella fabrica (47 m⁻²), 5 veneroid Eumarcia fumigata (42 m⁻²), phyllodocidan and scolecidan Goniada 6 Ptripartita (39 m⁻²), Leitoscoloplos Pnormalis (34 m⁻²) and Nephtys spp (33 m⁻ 7 ²), oedicerotoid *Doowia dexterae* (32 m⁻²) and spionidan *Spio pacifica* (31 m⁻²). 8 The rank-abundance curves of both assemblages, but particularly of that in 9 the seagrass, were of the concave form (Fig. 3b) characteristic of a number of 10 benthic marine systems (Hughes, 1984; and see Hubbell, 1997).

11 One-way ANOVA comparisons of the abundance of the dominant 12 species in the marginal and in the non-edge areas of their respective habitat 13 types did not show any significant differences in unvegetated areas of sand, except in respect of *Finella* which was more abundant in the marginal sand 14 15 zone than in the main sandflat (ANOVA $F_{1,22} = 4.9$; P = 0.04). In the seagrass 16 beds, several animals (including Calopia, Limnoporeia, Longiflagrum and the 17paratanaoid Leptochelia) showed greater abundance at the seagrass edge at 18 some sites but were less abundant marginally at others. Tanaids, for 19 example, were five times more abundant near some edges, but only half as 20 dense at others. Overall, the 25 peracaridan species present were more 21 abundant at the edge at half the sites but were less abundant at the other half. The most numerous seagrass inhabitant, Calopia, was also more 22 23 abundant near edges at two sites, but was less so at two others. On average, 24 the locality's 33 rissooid and other microgastropod species were less

abundant at the edge at two-thirds of the sites and overall were only half as abundant marginally, although the effect was not statistically significant. Only in respect of the rissooid *Pseudoliotia* was there a significant difference overall (ANOVA $F_{1,22}$ = 13.1; P = 0.001), this microgastropod being less abundant near the seagrass margins.

6 Regardless of site and of shore height, all six lattices displayed the 7 same pattern of distribution of both assemblage dissimilarity between 8 adjacent sampling horizons and changes in overall abundance and species 9 density across the boundary zone. Variation in quantitative assemblage 10 composition between adjacent sampling horizons (i.e. β -diversity) is shown in 11 Table 1. There was effectively no change between horizons within each of 12 the two marginal habitat blocks and no interactions with shore height (twoway ANOVA all $F_{2,2,4}$ <2.5; P >0.1), but a marked and highly significant 13 14 increase in dissimilarity occurred across the 25 cm wide strip on either side of the actual interface. The mean value of the dissimilarity statistic within 15 16 each habitat block was 0.10 (SE 0.02) and between the main body of the 17habitat type and its margin was 0.19 (SE 0.04), with in each case no 18 difference between the values within seagrass and within the sand (within 19 marginal zone ANOVA $F_{1,22} = 0.22$, P > 0.6; marginal vs non-marginal ANOVA 20 $F_{1,10} = 0.21$, P > 0.6). On either side of the interface the mean dissimilarity 21 statistic rose to 0.56 (SE 0.04).

Variation across the boundary zone in faunal abundance, observed
and estimated true species density, and levels of α-diversity is shown in
Figure 4a-c. Two-way ANOVA tests of these features in the seagrass

1 marginal horizons and in the unvegetated sand margins indicated no 2 significant effect of distance of the sampling horizon from the interface on 3 any assemblage metric, and no significant interaction between this distance and position on the shore (All seagrass $F_{3,2,6} < 1.2$, P > 0.3. All unvegetated 4 sand $F_{3,2,6}$, except α -diversity, ≤ 0.6 , P > 0.6; α -diversity F = 3.3, P = 0.06). In 5 each case, this included comparisons between the marginal zone and the 6 7 non-edge area of the habitat concerned. As previously noted at these sites (Barnes and Barnes, 2012), values of the $1/N_2 \alpha$ -diversity metric were very 8 9 variable (see Fig. 4c).

10 In marked contrast to the constancy within each of the marginal habitat zones, however, and as seen above in assemblage composition (Table 11 12 1), there were highly significant differences in faunal abundance and 13 observed and estimated true species density (though not α -diversity) between 14 adjacent horizons across the 25 cm wide strip on either side of the actual 15 interface (Table 2), where a very sharp change in seagrass abundance also 16 occurred (Fig. 4d). Each of the four seagrass horizons was significantly 17different from each of the four unvegetated-sand horizons in both faunal 18 abundance and species density (all Tukey HSD test statistics ≥ 6 ; $P \leq 0.003$). 19 Therefore, effectively all the dramatic change in composition, abundance and 20 species density between the faunal assemblages of seagrass and of adjacent 21unvegetated sand took place over the very small distance of 0.5 m at most. 22 Categorising the component assemblage species as either 'seagrass 23 specialist', 'unvegetated-sand specialist' or 'generalist', on the basis of 24 whether their abundance in the two habitat types was greater or less than a

ratio of 1 : 2.5, and removing all rare species (those with ≤5 individuals in
total) from the analysis, the two assemblages reacted symmetrically to the
interface (Fig. 5). Total abundance of the generalist category (principally *Nephtys, Goniada, Spio* and *Mysella*) was unaffected by the transition. As a
result of the difference in overall abundance, however, these four taxa were
significant elements in the fauna of the unvegetated sand (see above) but not
in that of the seagrass.

8 Figure 6 illustrates a plot of the four assemblage metrics at the local 9 and regional scales. Spatial trends in the metrics were captured accurately 10 at both scales (local R^2 = 0.87-0.93; regional R^2 = 0.71-0.74) using second 11 and third order polynomial regression models, although local-scale models 12 consistently performed better than regional-scale ones. In terms of cross-13 scale comparisons, the highest performing model type remained consistent 14 transitioning from the local to the regional scale. The metric modelled with 15 the highest accuracy at each scale was α species diversity.

16 Global Moran's *I* statistics calculated for all sites together and for each 17individual lattice are shown in Table 3. Three of the four metrics (faunal 18 abundance and the two species density measures) exhibited significant 19 positive spatial autocorrelation of moderate strength (I = 0.16 to 0.44), 20 although α species diversity mostly showed weaker levels of autocorrelation 21 that approximated a randomly distributed community (I = -0.18 to 0.11). 22 The semivariogram surfaces (Fig. 7) indicate a symmetrical pattern of 23 autocorrelation about the bare-sand/seagrass interface at all sites. With the 24 exception of the Polka LWS site that showed by far the smallest maximum

sill points (30 cm) and most uniform distribution of spatial autocorrelation
values, lattices displayed low levels of autocorrelation close to the interface
(i.e. sill points of *c*. 30 cm – less than the distance apart of sampling points
across the interface) and higher levels within each of the two abutting
habitat types (i.e. sill points of *c*. 40-75 cm, *c.f.* the interval between
sampling points of 25 cm there) (Fig. 7).

7 Since the potential effect of positive spatial autocorrelation on the 8 reliability of the ANOVAs above will be to increase the chance of Type I errors 9 (Legendre, 1993; Dale and Fortin, 2002), its only effective occurrence in the 10 data nested within each of the abutting habitat types will result in it having 11 no adverse influence on our statistical conclusions. No relevant null 12 hypotheses involving within-habitat comparisons were rejected, and the 13 rejected cross-habitat null hypotheses were all based on data lacking 14 significant spatial autocorrelation.

15

16 **4. DISCUSSION**

17 Although the precise faunal assemblages present in the seagrass and 18 bare sand near Dunwich vary significantly from location to location and 19 those in the seagrass also vary up the shore (Barnes and Barnes, 2012), the 20 same three features of interfaces between these two habitat types stand out 21 regardless of site and shore height, i.e. occurred in all six lattices. First, the 22 faunal assemblages of the marginal zone of both seagrass and unvegetated 23 sand in each lattice not only did not vary with closeness to the actual

1 interface but were not significantly different from those present locally away 2 from the edges of their respective habitat types. Secondly, as in the main 3 habitat areas (Barnes and Barnes, 2012), the assemblages immediately on 4 either side of the interface were very different, with the seagrass supporting 5 both greater animal numbers (>3 x) and species densities (>2.5 x). Only a 6 few generalist species were common to both (some 10% of the total). All 7 marginal seagrass horizons were significantly different from all marginal 8 sandflat ones in composition, faunal abundance and species density. Hence 9 there must be a very abrupt change from one assemblage type to the other 10 across the interface. Thirdly, that this was accomplished over a distance of 11 only 0.5 m at most is unexpected; although Warry et al. (2009) have shown 12 that 0.5 m is a large distance for seagrass harpacticoids and other 13 meiofauna. Local ecological changes over small distances are commonplace 14 in seagrass systems (Thrush et al., 2001; Barnes and Ellwood, 2012) but the 15 changes taking place over 0.5 m here are far from local, occurring at all 16 shore heights investigated and at all localities. Attrill and Rundle (2002, p. 17929) define an ecotone (as distinct from an ecocline) as 'an area of relatively 18 rapid change, producing a narrow ecological zone between two different and 19 relatively homogeneous community types': the boundary zone and 20 assemblage types studied here exactly fit this definition. Although Hansen 21 and di Castri (1992) consider that ecotones can range in width from 22 distances measured in centimetres to those measured in kilometres, the 23 dimension of this ecotone must be one of the smallest ever recorded in 24 association with a major change from one ecosystem bioengineer to another 25 in what is otherwise essentially the same substratum type. Walker et al.

1 (2003), for example, considered an ecotone width of 4 m to be a 'sharp' 2 boundary, and to Kingston and Morris (2000) one less than 20 m is 'abrupt'. 3 Unlike most other transitional zones (Rand et al., 2006; Kark, 2013), that 4 between seagrass and unvegetated sand on Stradbroke certainly does not 5 appear to support mixed faunal assemblages enriched by elements from both 6 juxtaposed habitat types and to display peak local species density and 7 faunal abundance. Mean species diversity was indeed largest in this region 8 (Fig. 6d), although here this indicates a more even allocation of total 9 individuals between the few species present rather than increased numbers 10 of faunal elements (Magurran, 2004; Barnes and Barnes, 2012).

11 There are a number of potential reasons why the Stradbroke 12 seagrass/sandflat ecotone might be of such particularly small width. The 13 interfaces chosen for study were deliberately those that appeared stable, and 14 therefore those in actively advancing or retreating boundary zones may well 15 be more extensive. Much may also depend on the precise habitat provided 16 by the marginal seagrass areas (van Houte-Howes et al., 2004; Tuya et al., 172011). At the sites investigated, there was no diminution in the number of 18 seagrass shoots per unit area as the interface was approached (see Figure 19 4d), and maintenance of a relatively constant habitat framework — water velocities excepted (see below) - right up to the interface itself may well aid 20 21 continuation of relatively constant assemblage characteristics until that 22 point. Thirdly, the two assemblages are dominated by organisms of 23 contrasting biology: the seagrass fauna being largely epifaunal and that of 24 the unvegetated sand being mainly infaunal (Barnes and Barnes, 2012). It is

1 well established (Orth et al., 1984; van Houte-Howes et al., 2004; etc) that 2 seagrass root/rhizome systems inhibit burrowing, whilst the lack of a plant 3 cover can expose surface-dwelling species to heavy predation. Such 4 contrasting assemblages are not universal, however. Differences between 5 seagrass and adjacent sand assemblages, and hence changes across 6 seagrass/sand boundaries, in high-latitude, high-abundance but low 7 biodiversity regions such as the north-west Atlantic, for example, have in 8 contrast been shown to be less significant (Asmus and Asmus, 2000; Polte et 9 al., 2005; Barnes, 2010). Bray-Curtis similarity between seagrass (N. noltii) 10 and muddy-sand assemblages at a 52°N site that supported a combined 11 fauna of 33 species at overall densities of >50,000 ind. m^{-2} , for example, was 12 >0.8 (RSK Barnes, unpubl.); c.f. the equivalent Stradbroke value of 0.13. In 13 temperate N. m. capricorni systems at 36°-37°S in New Zealand, van Houte-14 Howes et al. (2004) also found little difference between overall faunal 15 abundance, species density and species diversity in the seagrass and in the 16 unvegetated sediment, and noted that differences in assemblage composition 17between the two were due mainly to differential abundance.

18 Thrush (1991) noted that spatial autocorrelation in variables that 19 capture the biodiversity of macrobenthic assemblages has the potential to 20 confound experimental designs and to affect the reliability of inferential 21 statistics that may be used to assess distribution patterns. For example, the 22 presence of spatial dependence can result in a violation of the statistical 23 assumption of independence of observations. Tobler's (1970) First Law of 24 Geography, that objects closer together tend to be more similar than those

1 further apart, has repeatedly been found to apply to point sample datasets of 2 ecological communities (Fortin and Dale, 205), although critically not here across the seagrass/bare-sand interface. Granted the considerable 3 4 differences between values of the studied assemblage metrics in seagrass 5 and in unvegetated sand, it is hence no surprise that moderate 6 autocorrelation was detected within each boundary zone in three of the four 7 metrics (i.e. in faunal abundance and both species density measures). In 8 contrast to the other metrics, however, α species diversity showed 9 remarkable and generally consistent resistance to autocorrelation at all sites. 10 This might be taken to suggest that $1/N_2$ could potentially be a good 11 resistant measure of biodiversity for benthic studies, albeit one strongly 12 biassed towards the evenness component. That this is not so, however, is 13 suggested by the very large degree of scatter in this metric seen in both this 14 (Fig. 4c) and the earlier study of the sites by Barnes and Barnes (2012). 15 Indeed, despite the very marked changes in habitat type, in assemblage 16 composition, in overall faunal abundance and in species density across the 17interface, the $1/N_2$ metric failed to detect any significant change in 18 biodiversity there. Its resistance could simply be a measure of its 19 insensitivity (see Magurran and Henderson, 2010).

It is also noteworthy that the results of this study do not show large straight-forward and consistent edge effects. Earlier work on the response of benthic animals to the edges of seagrass beds has produced a complicated picture (Ries and Sisk, 2004), reactions seemingly varying from group to group (Murphy et al., 2010) and from study to study (Tuya et al., 2011) or

not even occurring at all (Nakaoka & Toyohara, 2000). The most consistent 1 2 effect has been claimed to be the greater densities of peracaridan 3 crustaceans (including tanaids) found at edges than away from them 4 (Boström et al., 2006). Nevertheless, in Port Phillip Bay, Victoria, tanaids 5 increased in abundance away from the edge, and the effect in other peracaridans was dependent on size of patch investigated (Murphy et al., 6 7 2010). In the present study, tanaids were more abundant near the edge at 8 some sites but were less so at others, as indeed were peracaridan 9 crustaceans in general, and microgastropod species displayed the same 10 pattern although overall they were only half as abundant marginally. In the 11 Stradbroke intertidal, only one animal (the microgastropod Pseudoliotia) 12 showed a consistent reaction. Little previous work has concerned small, 13 fragile, epibenthic microgastropods such as *Calopia* and *Pseudoliotia* — 14 forms that dominate the Stradbroke seagrass patches away from edges 15 (Barnes and Barnes, 2011, 2012; Barnes and Ellwood, 2012). But as water 16 velocities are known to be higher near seagrass margins (Peterson et al., 172004; Murphy et al., 2010), it is perhaps not surprising that some species, 18 such as *Pseudoliotia*, should be less abundant and widespread under the 19 more turbulent conditions near the interface, maybe via effects on their 20 recruitment (Bologna and Heck, 2002; Matias et al., 2013) or because of 21 dislodgement (Tuya et al., 2011). In addition, small, fragile animals may be 22 at greater risk from predators when relatively exposed (Kark and van 23 Rensburg, 2006; Barnes, 2010). Generally, however, it would appear that 24 edge effects are a variable local response to specific habitat conditions within patch margins rather than a consistent effect of closeness to an interface
 with a different habitat type.

3 Winberg et al. (2007) have highlighted the value of examining multiple 4 measures of macrobenthic biodiversity across a variety of spatial scales. The 5 present study has examined spatial patterning in quantitative assemblage 6 composition and in four assemblage metrics at the local (lattice) and regional 7 (all sites) scales traversing spatial dimensions ranging over 5 orders of 8 magnitude from 0.25 m to 2.5 km. Significantly, the emergent assemblage 9 features captured in each of the four metrics could accurately and 10 consistently be characterised across scales as second and third order 11 polynomial regression models. Such models further our understanding of 12 how communities are organised in space (Peres-Neto and Legendre, 2010) 13 and can be used as a basis for predicting community composition at 14 unsampled locations within the spatial extent of the sample lattice, for 15 example, underpinning interpolation techniques such as kriging. As is the 16 case with all interpolation (as opposed to extrapolation) methods, predictive 17values should not be generated outside the geographical area within which 18 the model parameters were defined by the trend analysis.

Overall, we conclude that faunal transitions across stable bare sand/seagrass boundaries in Moreton Bay take the form of very narrow ecotones indeed, across which the two assemblages react symmetrically, with the actual interface being the location of the entire change in faunal abundance and biodiversity observed between the contrasting patch types. However, this state seems unlikely to be a necessary response to changes in the prevailing habitat conditions *per se* and more likely to be a specific
consequence of the two low-latitude, low-abundance but high-biodiversity
faunal assemblages that are interacting here (or of the key members of those
assemblages), in that such marked quantitative and/or qualitative changes
in assemblages do not necessarily occur across intertidal bare
sediment/*Nanozostera* bed boundaries elsewhere, e.g. in lower latitudes.

7

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1 Table 1. Changes in assemblage composition across adjacent sampling 2 horizons on either side of seagrass/unvegetated-sand interfaces, as 3 estimated by the Chao/Sørensen abundance-based dissimilarity statistic, 4 corrected for undersampling bias (estimated complete similarity = 0; 5 estimated complete dissimilarity = 1). Positive horizon distances are in 6 seagrass, negative ones in unvegetated sand, the interface itself being zero. 7 The sharp change between +0.25 and -0.25 m from the interface (in bold) is 8 highly significant (post-hoc Tukey HSD statistic >12; *P* <0.0001). 9 10 Sampling horizons being compared (in metres from interface) 11 >+4 to +0.75 to +0.5 to +0.25 to -0.25 to -0.5 to -0.75 to 12 Site -0.5 -0.75 +0.75+0.5+0.25-0.25 >-4 13 14 Deanbilla MSL 0.03 0.03 0.08 0.70 0.01 0.19 0.43 15 Yerrol MSL 0.18 0.10 0.10 0.56 0.23 0.25 0.08 16 Polka LWN 0.34 0.00 0.18 0.50 0.12 0.04 0.04 17Deanbilla LWN 0.00 0.08 0.51 0.09 0.05 0.19 0.16 18 Polka LWS 0.16 0.00 0.04 0.43 0.07 0.02 0.09 Deanbilla LWS 0.00 0.18 19 0.35 0.28 0.20 0.67 0.23

1 Table 2. Summary of post-hoc Tukey HSD tests on various macrofaunal

2 assemblage metrics immediately on either side of the unvegetated sand/ seagrass

3 interface (i.e. 0.5 m apart). Significant effects are indicated in bold.

4			
5	Emergent property	test statistic	Р
6			
7	Faunal abundance	6.87	<0.001
8	Observed species density	9.62	<0.0001
9	Estimated true species density	6.49	<0.001
10	α-diversity	0.16	1
11			

12

Table 3. Global Moran's *I* statistic calculated on mean assemblage metrics at
 the local (individual sites) and regional (all sites) scales. Significant effects (at
 P <0.05) are indicated in bold.

Site	Abundance	Smax	S_{obs}	α-diversity
Deanbilla MSL	0.37	0.22	0.26	-0.18
Yerrol MSL	0.29	0.16	0.35	0.05
Polka LWN	0.21	0.41	0.44	0.11
Deanbilla LWN	0.37	0.29	0.37	0.04
Polka LWS	0.42	0.41	0.43	-0.2
Deanbilla LWS	0.26	0.29	0.37	-0.0009
All sites	0.44	0.38	0.43	0.26

1 Legends for Figures

2 1. Sampling locations, with inset showing the structure of the individual3 core lattices.

4 2. Interface between intertidal seagrass and unvegetated sandflat at Polka.
5 Foreground width some 2 m.

6 3. (A) Mao Tau numbers of macrofaunal species in marginal areas of
7 seagrass and of unvegetated sandflat in relation to (a) cumulative number of
8 individuals sampled (squares) [i.e. species richness] and (b) cumulative area
9 sampled, as number of cores (circles) [i.e. species density]. (Sample
10 accumulation orders randomised 50 times.) (B) Ranked species abundance
11 curves.

12 4. The distribution of assemblage metrics across the sampling horizons on 13 either side of the interface between unvegetated sand (-) and seagrass (+) in 14 the six lattices: (A) mean macrofaunal abundance (± SE); (B) mean observed 15 (S_{obs}) and estimated true (S_{max}) species density (± SE); (C) macrofaunal α 16 diversity values; and (D) mean number of seagrass shoots per core sample (± 17SE; n = 60). Values of macrofaunal abundance and species density away 18 from the transitional zone (>+4 and >-4) are shown for comparison in A and 19 В.

5. Percentage of maximum total abundance of seagrass specialist (n = 55),
bare-sand specialist (n = 10) and generalist (n = 8) macrofaunal species (as
judged by their local distributions) in each sampling horizon on either side of
the intertidal interface between unvegetated sand (-) and seagrass (+).

6. Spatial trend analysis plot of the assemblage metrics at the local (lattice)
and regional (all sites) scales. Mean values for each lattice are shown on the
left hand side and values are plotted in their geographical (*x*, *y*) locations on
the right hand side with the *z* axis depicting the value of the metric. The x
axis on both the local and regional plot is a proxy for distance from the
seagrass/bare-sand interface.

- 7 7. Semivariogram plots and surfaces constructed for the lattices at each site.8 Stated sill points for each lattice are the maximum values at that site.
- 9















