

Overlap of Genetic Risk between Interstitial Lung Abnormalities and Idiopathic Pulmonary Fibrosis

Brian D. Hobbs^{1,2*}, Rachel K. Putman^{2*}, Tetsuro Araki^{3,4}, Mizuki Nishino^{3,4}, Gunnar Gudmundsson⁵, Vilundur Gudnason^{6,7}, Gudny Eirisdottir⁷, Nuno Rodrigues Zilhao Nogueira⁷, Josée Dupuis^{8,9}, Hanfei Xu⁸, George T. O'Connor^{9,10}, Ani Manichaikul^{11,12}, Jennifer Nguyen¹¹, Anna J. Podolanczuk¹³, Purnema Madahar¹³, Jerome I. Rotter^{14,15,16}, David J. Lederer^{13,17}, R. Graham Barr^{13,17}, Stephen S. Rich^{11,12}, Elizabeth J. Ampleford¹⁸, Victor E. Ortega¹⁸, Stephen P. Peters¹⁸, Wanda K. O'Neal¹⁹, John D. Newell, Jr.^{20,21}, Eugene R. Bleeker²², Deborah A. Meyers²², Richard J. Allen²³, Justin M. Oldham²⁴, Shwu-Fan Ma²⁵, Imre Noth²⁵, R. Gisli Jenkins²⁶, Toby M. Maher^{27,28†}, Richard B. Hubbard^{26,29}, Louise V. Wain³⁰, Tasha E. Fingerlin^{31,32}, David A. Schwartz^{32,33,34}, George R. Washko^{2,4}, Ivan O. Rosas², Edwin K. Silverman^{1,2}, Hiroto Hatabu^{3,4}, Michael H. Cho^{1,2*}, and Gary M. Hunninghake^{2,4*}; for the COPDGene Investigators, ECLIPSE Investigators, SPIROMICS Research Group, and UK ILD Consortium

¹Channing Division of Network Medicine, ²Division of Pulmonary and Critical Care Medicine, ³Department of Radiology, and ⁴Center for Pulmonary Functional Imaging, Brigham and Women's Hospital, Boston, Massachusetts; ⁵Department of Respiratory Medicine, Landspítali University Hospital, and ⁶Faculty of Medicine, University of Iceland, Reykjavik, Iceland; ⁷Icelandic Heart Association, Kopavogur, Iceland; ⁸Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts; ⁹NHLBI Framingham Heart Study, Framingham, Massachusetts; ¹⁰Pulmonary Center, Department of Medicine, Boston University, Boston, Massachusetts; ¹¹Center for Public Health Genomics, ¹²Department of Public Health Sciences, and ²⁵Division of Pulmonary and Critical Care Medicine, University of Virginia, Charlottesville, Virginia; ¹³Department of Medicine, College of Physicians and Surgeons, and ¹⁷Department of Epidemiology, Mailman School of Public Health, Columbia University, New York, New York; ¹⁴Institute for Translational Genomics and Population Sciences, Los Angeles Biomedical Research Institute, and ¹⁵Division of Genomic Outcomes, Department of Pediatrics and ¹⁶Department of Medicine, Harbor-UCLA Medical Center, Torrance, California; ¹⁸Department of Internal Medicine, Center for Precision Medicine, Wake Forest School of Medicine, Winston-Salem, North Carolina; ¹⁹Marsico Lung Institute, School of Medicine, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina; ²⁰Division of Cardiovascular and Pulmonary Imaging, Department of Radiology, University of Iowa Carver College of Medicine, Iowa City, Iowa; ²¹Department of Radiology, University of Washington, Seattle, Washington; ²²Division of Genetics, Genomics and Precision Medicine, Department of Medicine, University of Arizona, Tucson, Arizona; ²³Department of Health Sciences, University of Leicester, Leicester, United Kingdom; ²⁴Department of Internal Medicine, University of California Davis, Davis, California; ²⁶National Institute for Health Research, Biomedical Research Centre, Respiratory Research Unit, School of Medicine, and ²⁹Division of Epidemiology and Public Health, University of Nottingham, Nottingham, United Kingdom; ²⁷National Institute for Health Research, Respiratory Biomedical Research Unit, Royal Brompton Hospital, London, United Kingdom; ²⁸Fibrosis Research Group, Inflammation, Repair and Development Section, National Heart and Lung Institute, Imperial College, London, United Kingdom; ³⁰National Institute for Health Research, Leicester Respiratory Biomedical Research Centre, Glenfield Hospital, Leicester, United Kingdom; ³¹Center for Genes, Environment and Health, National Jewish Health, Denver, Colorado; and ³²Department of Biostatistics and Informatics, ³³Department of Medicine, School of Medicine, and ³⁴Department of Immunology, School of Medicine, University of Colorado Denver, Aurora, Colorado

ORCID IDs: 0000-0001-9564-0745 (B.D.H.); 0000-0002-8027-7450 (R.K.P.); 0000-0002-9559-1485 (A.J.P.); 0000-0001-5258-0228 (D.J.L.).

(Received in original form March 1, 2019; accepted in final form July 17, 2019)

A list of SPIROMICS investigators may be found before the beginning of the REFERENCES.

*These authors contributed equally to this work.

†T.M.M. is Associate Editor of *AJRCCM*. His participation complies with American Thoracic Society requirements for recusal from review and decisions for authored works.

Author Contributions: Study design: V.G., G.T.O'C., D.J.L., R.G.B., E.R.B., D.A.M., I.N., R.G.J., L.V.W., D.A.S., E.K.S., M.H.C., and G.M.H. Acquisition, analysis, or interpretation of the data: B.D.H., R.K.P., T.A., M.N., G.G., V.G., G.E., N.R.Z.N., J.D., H.X., G.T.O'C., A.M., J.N., A.J.P., P.M., J.I.R., D.J.L., R.G.B., S.S.R., E.J.A., V.E.O., S.P.P., W.K.O'N., J.D.N., E.R.B., D.A.M., R.J.A., J.M.O., S.-F.M., I.N., R.G.J., T.M.M., R.B.H., L.V.W., T.E.F., D.A.S., G.R.W., I.O.R., E.K.S., H.H., M.H.C., and G.M.H. Critical revision of the manuscript for important intellectual content: B.D.H., R.K.P., T.A., M.N., G.G., V.G., G.E., N.R.Z.N., J.D., H.X., G.T.O'C., A.M., J.N., A.J.P., P.M., J.I.R., D.J.L., R.G.B., S.S.R., E.J.A., V.E.O., S.P.P., W.K.O'N., J.D.N., E.R.B., D.A.M., R.J.A., J.M.O., S.-F.M., I.N., R.G.J., T.M.M., R.B.H., L.V.W., T.E.F., D.A.S., G.R.W., I.O.R., E.K.S., H.H., M.H.C., and G.M.H. Statistical analysis: B.D.H., R.K.P., N.R.Z.N., G.E., V.G., J.D., H.X., A.M., J.N., E.J.A., V.E.O., R.J.A., I.N., R.G.J., L.V.W., T.E.F., D.A.S., M.H.C., and G.M.H. Obtained funding: V.G., G.T.O'C., D.J.L., R.G.B., E.R.B., D.A.M., I.N., R.G.J., L.V.W., D.A.S., E.K.S., M.H.C., and G.M.H.

Correspondence and requests for reprints should be addressed to Gary M. Hunninghake, M.D., M.P.H., Pulmonary and Critical Care Division, Department of Medicine, Brigham and Women's Hospital, 75 Francis Street, Boston, MA 02115. E-mail: ghunninghake@bwh.harvard.edu.

This article has an online supplement, which is accessible from this issue's table of contents at www.atsjournals.org.

Am J Respir Crit Care Med Vol 200, Iss 11, pp 1402–1413, Dec 1, 2019

Copyright © 2019 by the American Thoracic Society

Originally Published in Press as DOI: 10.1164/rccm.201903-0511OC on July 24, 2019

Internet address: www.atsjournals.org

Abstract

Rationale: Interstitial lung abnormalities (ILAs) are associated with the highest genetic risk locus for idiopathic pulmonary fibrosis (IPF); however, the extent to which there are unique associations among individuals with ILAs or additional overlap with IPF is not known.

Objectives: To perform a genome-wide association study (GWAS) of ILAs.

Methods: ILAs and a subpleural-predominant subtype were assessed on chest computed tomography (CT) scans in the AGES (Age Gene/Environment Susceptibility), COPDGene (Genetic Epidemiology of Chronic Obstructive Pulmonary Disease [COPD]), Framingham Heart, ECLIPSE (Evaluation of COPD Longitudinally to Identify Predictive Surrogate End-points), MESA (Multi-Ethnic Study of Atherosclerosis), and SPIROMICS (Subpopulations and Intermediate Outcome Measures in COPD Study) studies. We performed a GWAS of ILAs in each cohort and combined the results using a meta-analysis. We assessed for overlapping associations in independent GWASs of IPF.

Measurements and Main Results: Genome-wide genotyping data were available for 1,699 individuals with ILAs and 10,274 control subjects. The *MUC5B* (mucin 5B) promoter variant rs35705950 was significantly associated with both ILAs ($P = 2.6 \times 10^{-27}$) and subpleural ILAs ($P = 1.6 \times 10^{-29}$). We discovered novel genome-wide associations near *IPO11* (rs6886640, $P = 3.8 \times 10^{-8}$) and *FCF1P3* (rs73199442, $P = 4.8 \times 10^{-8}$) with ILAs, and near *HTRE1* (rs7744971, $P = 4.2 \times 10^{-8}$) with subpleural-predominant ILAs. These novel associations were not associated with IPF. Among 12 previously reported IPF GWAS loci, five (*DPP9*, *DSP*, *FAM13A*, *IVD*, and *MUC5B*) were significantly associated ($P < 0.05/12$) with ILAs.

Conclusions: In a GWAS of ILAs in six studies, we confirmed the association with a *MUC5B* promoter variant and found strong evidence for an effect of previously described IPF loci; however, novel ILA associations were not associated with IPF. These findings highlight common genetically driven biologic pathways between ILAs and IPF, and also suggest distinct ones.

Keywords: genetics; genome-wide association study; interstitial lung abnormalities; idiopathic pulmonary fibrosis; SNP

Idiopathic pulmonary fibrosis (IPF), the most common and severe form of interstitial lung disease (ILD) (1), is a disorder of lung scarring that is reported

to affect one in 200 adults over the age of 65 (2), and results in a high rate of mortality (3, 4). A strong genetic basis for PF has been demonstrated in studies of

familial aggregation (5), as well as in genome-wide linkage and association studies that have provided replicable evidence for associations between

B.D.H. is supported by NIH grant K08 HL136928. R.K.P. is supported by NIH grant K08 HL140087. M.N. is supported by NIH grant R01 CA203636. G.G. is supported by the Oddur Olafsson Fund, project grant 141513-051 from the Icelandic Research Fund, and Landspítali Scientific Fund grants A-2015-030, A-2016-023, A-2017-030, A-2018-022, and A-2018-025. G.G. is supported by National Institute on Aging (NIA) grant 27120120022C and project grant 141513-051 from the Icelandic Research Fund. G.T.O'C. is supported by NIH grant OT2 OD026553. A.M. is supported by NIH grant R01 HL131565. A.J.P. is supported by NIH grant K23 HL140199. J.I.R. is supported by NIH grants R01 HL142302, R01 EY009052, R01EY023704, and P30 DK063491. D.J.L. is supported by NIH grants K24 HL131937, R01 HL103676, and R01 HL137234. R.G.B. is supported by NIH grants R01 HL077612, R01 HL093081, R01 HL121270, and R01 HL142028. S.S.R. is supported by NIH grants U01 HL120393, DP3 DK111906, and P01 HL136275. W.K.O'N. is supported by NIH grants R01 HL117843 and U24 HL141762. V.E.O. is supported by NIH grants K08 HL118128 and R01 HL142992. E.R.B. is supported by NIH grants UG1 HL1390534 and U01 HL109164. D.A.M. is supported by NIH grants R01 NR013700 and U01 HL109164. R.J.A. is supported by an Action Pulmonary Fibrosis Research Council grant G0901226. T.M.M. is supported by a National Institute of Health Research (NIHR) Clinician Scientist Fellowship (CS-2013-13-017). L.V.W. holds a GlaxoSmithKline/British Lung Foundation Chair in Respiratory Research. The research was partially supported by the NIHR Leicester Biomedical Research Centre; the views expressed are those of the author(s) and not necessarily those of the National Health Service, the NIHR, or the Department of Health. T.E.F. is supported by NIH grants R01 HL114587, R01 HL097163, and P01 HL132821. D.A.S. is supported by NIH grants P01 HL092870, R01 HL097163, R33 HL120770, R33 CA182360, and UH3 HL123442. G.R.W. is supported by NIH grants R01 HL116473 and R01 HL122464. I.O.R. is supported by NIH grants U01 HL133232 and R01 HL130974. E.K.S. is supported by NIH grants U01 HL089856, R01 HL113264, R01 HL137927, R01 HL133135, and P01 HL114501. M.H.C. is supported by NIH grants R01 HL135142, R01 HL113264, and R01 HL137927. G.M.H. is supported by NIH grants R01 HL111024, R01 HL130974, R01 135142, and project grant 141513-051 from the Icelandic Research Fund. The Framingham Heart Study is supported by NIH contracts N01-HC-25195 and HHSN268201500001. COPDGene is supported by NIH grants U01 HL089897 and U01 HL089856. The COPDGene project is also supported by the COPD Foundation through contributions made to an Industry Advisory Board comprised of AstraZeneca, Boehringer Ingelheim, GlaxoSmithKline, Novartis, Pfizer, Siemens, and Sunovion. The Age, Gene/Environment Susceptibility-Reykjavik Study was supported by NIA grant 27120120022C, NIH contracts N01-AG-1-2100 and HHSN27120120022C, the NIA Intramural Research Program, Hjartavernd (the Icelandic Heart Association), and the Althingi (the Icelandic Parliament). MESA and the MESA SHARe project are conducted and supported by the NHLBI in collaboration with MESA investigators. Support for MESA is provided by contracts HHSN2682015000031, N01-HC-95159, N01-HC-95160, N01-HC-95161, N01-HC-95162, N01-HC-95163, N01-HC-95164, N01-HC-95165, N01-HC-95166, N01-HC-95167, N01-HC-95168, N01-HC-95169, UL1-TR-000040, UL1-TR-001079, and UL1-TR-001420, and by National Center for Advancing Translational Sciences grant ULTR001881 and National Institute of Diabetes and Digestive and Kidney Disease Diabetes Research Center grant DK063491 to the Southern California Diabetes Endocrinology Research Center. SPIROMICS (Subpopulations and Intermediate Outcome Measures in COPD Study) is funded by contracts from the NHLBI (HHSN268200900013C, HSN268200900014C, HHSN268200900015C, HHSN268200900016C, HHSN268200900017C, HHSN268200900018C, HHSN268200900019C, and HHSN268200900020C) and a grant from the NIH/NHLBI (U01 HL137880), and supplemented by contributions made through the Foundation for the NIH and the COPD Foundation from AstraZeneca/MedImmune, Bayer, Bellerophon Therapeutics, Boehringer-Ingelheim Pharmaceuticals Inc., Chiesi Farmaceutici, Forest Research Institute Inc., GlaxoSmithKline, Grifols Therapeutics Inc., Ikaria Inc., Novartis Pharmaceuticals Corporation, Nycomed GmbH, ProterixBio, Regeneron Pharmaceuticals Inc., Sanofi, Sunovion, Takeda Pharmaceutical Co., Theravance Biopharma, and Mylan.

At a Glance Commentary

Scientific Knowledge on the

Subject: Individuals with interstitial lung abnormalities (ILAs) exhibit a clinical syndrome similar to that observed in patients with idiopathic pulmonary fibrosis (IPF), including physiologic decrements, radiologic progression, accelerated lung function decline, and an increased risk of death. ILAs are associated with the most common and highest genetic risk locus in IPF, the *MUC5B* (*mucin 5B*) promoter polymorphism rs35705950. However, the extent to which there is additional overlap in the genetic risk of ILAs and IPF is not known.

What This Study Adds to the Field:

In a genome-wide association study of ILAs, we confirmed findings at the *MUC5B* locus and identified three novel loci for ILAs and subpleural-predominant ILAs. These novel loci were not associated with IPF. Additionally, among 12 distinct, previously identified IPF GWAS loci, we identified 11 directionally consistent associations with ILAs, of which 7 were at least nominally significant and 5 (near *DPP9*, *DSP*, *FAM13A*, *IVD*, and *MUC5B*) were significantly associated after adjustment for multiple testing.

common genetic variants and IPF (6–11). Most consistently, IPF has been associated with increased copies of a common variant (rs35705950) in the promoter of the *MUC5B* (*mucin 5B*) gene (6, 8–11), a finding that may explain up to 30% of the risk of the disease (6).

Due to the severity of physiologic decrements and the high rate of mortality at the time of diagnosis, investigators have recently made efforts to identify IPF and other forms of PF in their earliest stages (12–14). These efforts include chest computed tomography (CT) image characterization of research participants who have not received a diagnosis but have classified imaging features that are suggestive not only of IPF specifically (15–17) but also of the broader set of ILDs (termed interstitial lung abnormalities

[ILAs]) (13). Evidence supporting a correlation between some research participants with ILAs who have not received a diagnosis and patients with IPF include a shared association with increased copies of the *MUC5B* promoter variant (12, 18). However, the extent to which research participants with ILAs who have not received a diagnosis and patients with IPF share common or unique genetic etiologies remains unclear.

We hypothesized that comparisons between research participants with and without ILAs would identify findings of genetic association shared with those identified in patients with IPF and, based on the diversity of ILA phenotypes (18) and ILDs in general (19), unique associations. To test this hypothesis, we genotyped common genome-wide single-nucleotide variants and imputed additional genotypes from reference panels, and then tested for association with visually assessed ILAs and the subpleural-predominant ILA subtype in populations of research participants from six unique cohorts. Based on the results, we performed further comparisons to examine the overlap of top genetic associations in research participants with ILAs (and subpleural-predominant ILAs) with genetic associations previously reported in patients with IPF (8). Some of the results of this study have been previously reported in the form of an abstract (20).

Methods

Study Population

The protocols for participant enrollment and phenotyping in the FHS (Framingham Heart Study) and the AGES (Age Gene/Environment Susceptibility)-Reykjavik, COPDGene (Genetic Epidemiology of COPD), ECLIPSE (Evaluation of COPD Longitudinally to Identify Predictive Surrogate End-points), MESA (Multi-Ethnic Study of Atherosclerosis)-Lung Study, and SPIROMICS (Subpopulations and Intermediate Outcome Measures in COPD Study) studies have been described previously (12–14, 21–23). Approval from the appropriate ethical/regulatory bodies was obtained for each cohort, and informed consent was obtained from all individuals. More detailed cohort information,

including cohort-specific methods, can be found in the online supplement.

ILA Characterization

ILA was characterized in all of the cohorts by visual assessments of chest CT scans. In FHS, AGES, COPDGene, and ECLIPSE, CT scans were evaluated for ILAs via a sequential reading method by up to three readers (radiologists and pulmonologists) who were blind to all participant-specific information, as previously described (24). ILAs in these cohorts were defined as nondependent changes affecting >5% of any lung zone. The abnormalities included ground-glass or reticular abnormalities, diffuse centrilobular nodularity, multiple nonemphysematous cysts, traction bronchiectasis, and honeycombing. Chest CT scans with either focal or unilateral ground-glass attenuation, focal or unilateral reticulation, or patchy ground-glass abnormalities were indeterminate for ILAs (12, 13). In MESA, ILAs were assessed by a radiologist using the above criteria, as previously described (25). In SPIROMICS, ILAs were classified as present or absent according to the presence or absence of bilateral, nondependent, peripheral (but not necessarily subpleural) ground-glass and/or reticular opacities and/or honeycombing (*see* the online supplement for further details).

Based on prior data on the genetics of ILAs (18), and to provide consistency of subtyping across cohorts, an additional ILA subset was created that excluded ILAs limited to participants with centrilobular nodules alone (13) and included all participants with ILAs with predominantly subpleural imaging findings (subpleural predominant).

Genotyping and Imputation

Details regarding genotyping in each cohort can be found in the online supplement. The genotypes of individuals from European-ancestry studies were imputed via the Michigan Imputation Server using minimac3 with the Haplotype Reference Consortium (HRC v1.1) reference panel (26). Individuals of African American ancestry in COPDGene and MESA were imputed using the 1000 Genomes Project (Phase 3, version 5) (27). The *MUC5B* promoter polymorphism (rs35705950) was poorly imputed (imputation $R^2 < 0.5$) in

the AGES study, so instead we used direct genotyping data, which were available for 3,209 individuals.

Genome-Wide Association Study and Meta-analysis

Given the case-control imbalance, we performed Firth bias-corrected logistic regression (28, 29) in each ancestry subset of each study, adjusting for age, sex, pack-years of smoking, and ancestry-based principal components as appropriate for each study. In the FHS, to allow application of the Firth bias-corrected logistic regression, we selected a subset of unrelated participants for analysis, preferentially choosing ILA cases. Summary statistics from individual studies, including chromosome and position (hg19), effect allele and other allele oriented to the + strand, effect allele frequency, and imputation quality were uploaded to a secure site at the Brigham and Women's Hospital/Channing Division of Network Medicine.

The summary statistics from each study were assessed using EasyQC (30) version 10.1. Quality control assessments included allele frequency comparisons with either a Haplotype Reference Consortium or 1000 Genomes reference panel, SE versus sample size checking, and quantile-quantile plot visualization. Variants with an imputation quality metric of <0.5 , a minor allele count of <10 (using the effective sample size or the number of cases adjusted for imputation quality when appropriate), were set to missing. Variant names were all normalized to hg19 chromosome and position. Only the highest-frequency alternate allele was retained for multiallelic variants.

After completing the summary statistical quality control process, we performed an inverse-variance-weighted, fixed-effects meta-analysis in METAL (version 2011-03-25) (31, 32) for both the ILA and subpleural-predominant ILA analyses. In a set of secondary analyses, we performed a meta-analysis restricted to ILA and subpleural ILA results from European-ancestry subpopulations, and a smoking-stratified (ever-smokers compared with never-smokers) meta-analysis of our genome-wide significant variants. Only variants that were present in at least half of the cohort subpopulations in each meta-analysis were further evaluated. Genome-wide significance for all associations was considered to be $P < 5 \times 10^{-8}$. To identify distinct results at each locus in the

European-ancestry subpopulations, we used genome-wide complex trait analysis—conditional and joint analysis (33, 34) on all results with $P < 5 \times 10^{-6}$, using the default distance of 10 Mb. COPDGene non-Hispanic whites (the largest representative population) were the reference population for the genome-wide complex trait analysis—conditional and joint analysis.

Overlap of ILA Genetic Loci with IPF, High-Attenuation Areas, Smoking Behaviors, and Connective Tissue Disease

We evaluated the overlap of top ILA-associated genetic variants with IPF in two ways: 1) lookup of IPF genome-wide association study (GWAS) loci from the National Human Genome Research Institute-European Bioinformatics Institute (NHGRI-EBI) GWAS Catalog (35) (downloaded June 4, 2018) in our ILA GWAS results; 2) lookup of our top ILA-associated variants ($P < 5e-7$ with either ILAs or subpleural ILAs in a European-ancestry subpopulation) with IPF in a recent European-ancestry IPF GWAS and meta-analysis (*see* online supplement) (11). In the lookup of prior IPF GWAS variants in our results, we restricted the lookup to 12 distinct IPF GWAS loci (reported results are for the variant that demonstrated the greatest statistical significance at each locus). Significance for association of IPF GWAS variants with ILAs or subpleural-predominant ILAs was set to $P < 0.05/12$. Additional analyses using logistic regression conditioning on the *MUC5B* promoter polymorphism (rs35705950) were done to assess for independence of the multiple SNPs previously identified at the 11p15 locus. To evaluate the overlap between ILAs and high-attenuation areas (HAAs), which have been associated with early or subclinical ILD and future ILAs (14, 25), we performed a lookup of the previously reported genome-wide significant variants associated with HAAs (36). To evaluate the overlap between ILAs and smoking behaviors, we performed a lookup of the previously reported genome-wide significant variants associated with smoking behaviors (37). To assess a potential overlap of the genetic susceptibility to ILAs (and subpleural-predominant ILAs) with connective tissue disease (CTD)-associated ILD and sarcoidosis, we searched the NHGRI-EBI GWAS Catalog (35) for genome-wide

significant SNPs in European-ancestry association studies related to rheumatoid arthritis, sarcoidosis, systemic lupus erythematosus, inflammatory myopathies, and systemic sclerosis. We assessed the P value for association of the genome-wide significant rheumatologic disease SNPs with ILAs and subpleural-predominant ILAs. We used Bonferroni P values as a significance threshold to correct for multiple testing (number of genome-wide significant SNPs) within each unique trait.

Expression Quantitative Trait Loci Lookups of Top GWAS Variants

We assessed whether our significant ILA genetic risk variants were expression quantitative trait loci (eQTLs) in the lung and blood using multiple available datasets, including GTEx lung and blood eQTLs (38, 39), Westra and colleagues' blood eQTLs (40), Hao and colleagues' lung eQTLs (41), Jansen and colleagues' Netherlands Study of Depression and Anxiety/Netherlands Twin Register conditional blood eQTLs (42), and eQTL Consortium blood eQTL meta-analysis data (43). Only *cis*-eQTLs were assessed, and significant associations were determined using the adjusted P values reported in each available eQTL dataset.

Results

We performed a GWAS and meta-analysis of 1,699 participants with ILAs and 10,274 control subjects in six cohorts, and the subjects in each study were stratified into subpopulations according to European, African, and Hispanic ancestry. We performed a secondary GWAS and meta-analysis using the subset of 1,287 subpleural-predominant ILA cases (Figure 1). The baseline characteristics of each cohort and subpopulation, stratified by ILA status, are included in Table 1 (for the characteristics of participants with ILAs limited to the subpleural-predominant subtype, *see* Table E1 in the online supplement). Similar to what was found in prior studies, participants with ILAs tended to be older (15) and generally had greater exposure to tobacco smoke than those without ILAs.

Genome-Wide Association

We identified three genome-wide significant variants associated with ILAs, including one

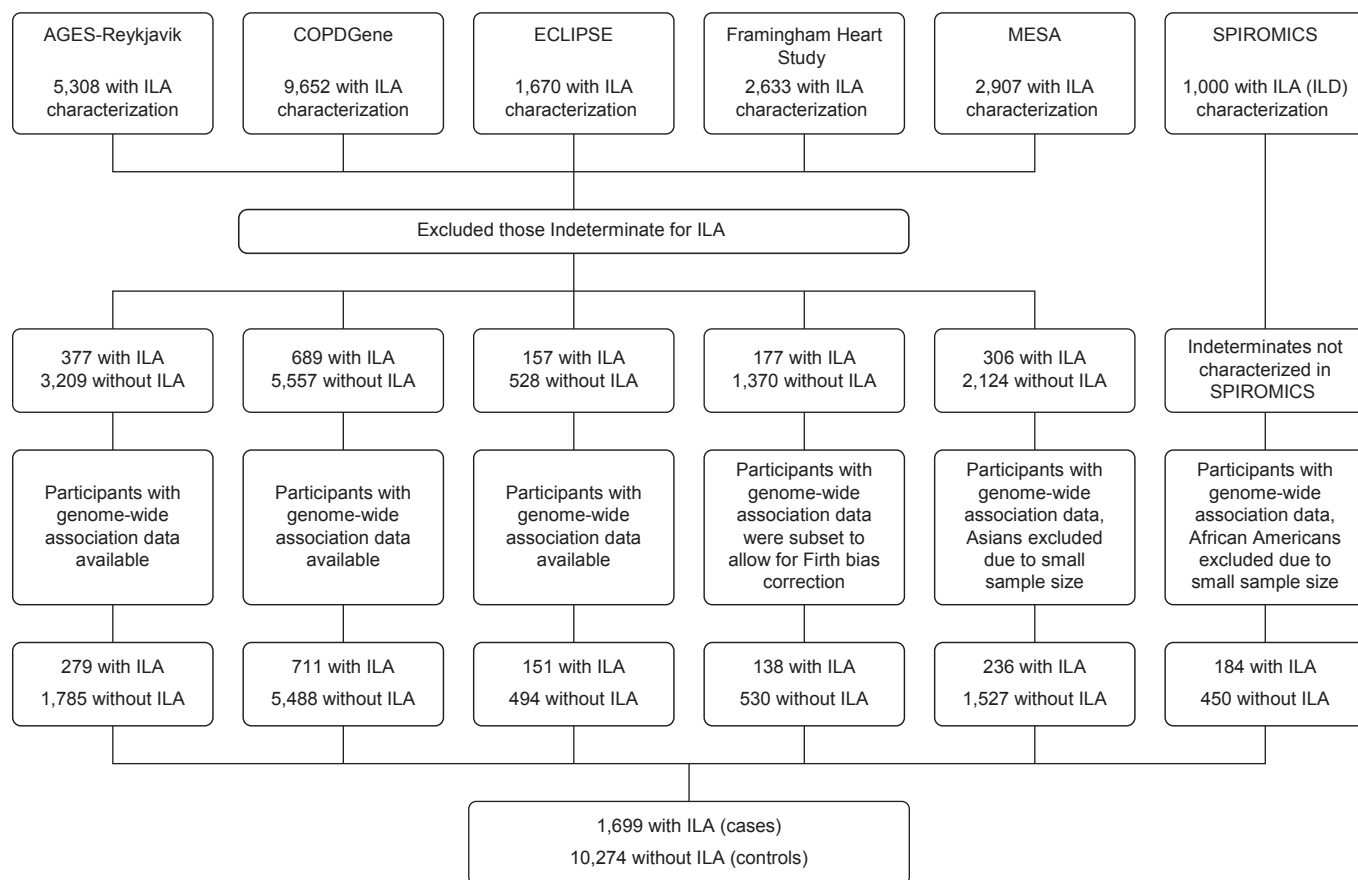


Figure 1. Flowchart depicting the participants included and excluded from the genome-wide association analysis by cohort and interstitial lung abnormality (ILA) status. AGES = Age Gene/Environment Susceptibility; COPD = chronic obstructive pulmonary disease; COPDGene = Genetic Epidemiology of COPD; ECLIPSE = Evaluation of COPD Longitudinally to Identify Predictive Surrogate End-Points; ILD = interstitial lung disease; MESA = Multi-Ethnic Study of Atherosclerosis; SPIROMICS = Subpopulations and Intermediate Outcome Measures in COPD Study.

at 11p15, at the known *MUC5B* promoter polymorphism, rs35705950 (odds ratio [OR], 1.97; 95% confidence interval [CI], 1.74–2.22; $P = 2.6 \times 10^{-27}$), as well as two novel loci: rs6886640 at 5q12, near *IPO11* (OR, 1.28; 95% CI, 1.18–1.41; $P = 3.8 \times 10^{-8}$), and rs73199442 at 3q13, near the long noncoding RNA *FCFIP3* (OR, 1.68; 95% CI, 1.39–2.02; $P = 4.8 \times 10^{-8}$) (Table 2 and Figures 2, E1, and E2). In the subpleural-predominant ILA analysis, in addition to the association with the *MUC5B* variant rs35705950 (OR, 2.22; 95% CI, 1.93–2.55; $P = 1.6 \times 10^{-29}$), we identified a novel genetic association at the 6q15 locus with rs7744971, near *HTR1E* (OR, 1.32; 95% CI, 95% CI 1.19–1.45; $P = 4.2 \times 10^{-8}$) (Table 2 and Figure 2). The ILA risk variant at 3q13.1 (rs73199442) was missing in the African- and Hispanic-ancestry subpopulations (due to low minor allele frequency), but it showed a consistent direction of effect in all European-ancestry

subpopulations (see forest plots in Figure 2). Similar results were noted in meta-analyses of ILAs and subpleural-predominant ILAs limited to individuals of European ancestry; however, the 5q12 locus was not significantly associated with ILAs, and the 6q15 locus had genome-wide significance in association with both ILAs and subpleural-predominant ILAs (Tables E2 and E3). For each variant that demonstrated genome-wide significance, we tested for genotype-by-smoking (ever-smokers compared with never-smokers) interactions. There was no evidence of a significant interaction between smoking status and any of the four genome-wide significant variants (Table E4 and Figure E3). To assess whether these novel ILA risk loci overlapped with IPF, we attempted to replicate our genome-wide significant associations with ILA and subpleural ILA associations in a European-ancestry GWAS and meta-analysis of 2,668 patients with

IPF and 8,591 control subjects (11) (see the online supplement). Aside from the known overlap at rs35705950 (IPF P value = 1.2×10^{-203}), none of our top ILA loci were significantly associated with IPF (Table 2).

Assessment of Replication for Prior IPF, HAAs, Smoking Behaviors, and CTD Genetic Loci

We examined the overlap of ILA and subpleural-predominant ILA genetic associations with 12 previously reported, distinct IPF GWAS loci from the NHGRI-EBI GWAS Catalog. There was a substantial enrichment of the 12 IPF GWAS loci in our ILA association results. Five SNPs near *DPP9*, *DSP*, *FAM13A*, *IVD*, and *MUC5B* were significantly associated ($P < 4.2 \times 10^{-3}$) with ILAs, and two additional SNPs at *MAPT* and *LRR34* were nominally significant ($P < 0.05$, but did not meet the

Table 1. Baseline Characteristics of the Participants, Stratified by Interstitial Lung Abnormality Status, in Each Cohort

	AGES-Reykjavik		Non-Hispanic Whites		African Americans		ECLIPSE		Framingham Heart Study		Non-Hispanic Whites		Hispanics		African Americans		SPIROMICS	
	No ILA (n = 1,785; 86%)	ILA (n = 279; 14%)	No ILA (n = 3,771; 88%)	ILA (n = 497; 12%)	No ILA (n = 1,717; 89%)	ILA (n = 214; 11%)	No ILA (n = 484; 77%)	ILA (n = 151; 23%)	No ILA (n = 530; 79%)	ILA (n = 138; 21%)	No ILA (n = 675; 85%)	ILA (n = 116; 15%)	No ILA (n = 385; 88%)	ILA (n = 54; 12%)	No ILA (n = 467; 88%)	ILA (n = 66; 12%)	No ILA (n = 450; 71%)	ILA (n = 184; 29%)
Age, yr, mean (SD)	76 (5)	77 (5)	61 (9)	64 (9)	54 (7)	55 (8)	62 (7)	64 (8)	58 (11)	71 (11)	69 (9)	75 (10)	67 (9)	74 (9)	69 (9)	74 (8)	65 (8)	68 (8)
Sex, F, n (%)	1,078 (60)	125 (45)	1,814 (48)	226 (45)	701 (41)	116 (54)	186 (34)	40 (26)	278 (56)	67 (49)	344 (51)	60 (52)	209 (54)	24 (54)	247 (53)	41 (62)	206 (46)	83 (45)
Body mass index, mean (SD)	27 (4)	27 (5)	29 (5)	29 (5)	29 (7)	29 (7)	27 (6)	26 (5)	28 (5)	28 (5)	28 (5)	28 (6)	30 (6)	30 (6)	30 (6)	27 (5)	27 (5)	29 (5)
Pack-years smoking, mean (SD)	2 (0-26)	21 (0-51)	40 (29-56)	45 (34-63)	34 (22-46)	35 (24-47)	45 (33-62)	43 (29-61)	0 (0-12)	8 (0-23)	14 (3-34)	15 (2-33)	3 (0-11)	13 (2-28)	11 (3-24)	18 (6-33)	41 (30-60)	50 (37-65)
Smoking status, n (%)																		
Current	211 (12)	46 (16)	1,426 (38)	263 (53)	1,377 (80)	178 (83)	189 (38)	69 (46)	33 (6)	11 (8)	43 (6)	8 (7)	20 (5)	6 (11)	50 (11)	8 (12)	126 (28)	59 (32)
Former	756 (42)	162 (58)	2,345 (62)	234 (47)	340 (20)	36 (17)	305 (62)	82 (54)	238 (45)	74 (54)	322 (48)	66 (59)	158 (41)	25 (46)	206 (44)	38 (58)	294 (65)	119 (65)
Never	818 (46)	71 (26)	—	—	—	—	—	—	259 (49)	53 (38)	310 (46)	40 (35)	207 (54)	23 (43)	211 (45)	20 (30)	30 (7)	6 (3)
History of COPD, n (%)	—	—	1,527 (40)	171 (34)	380 (22)	57 (27)	494 (100)	151 (100)	46 (9)	18 (13)	129 (22)	21 (21)	62 (17)	11 (23)	98 (24)	14 (25)	290 (64)	116 (63)

Definition of abbreviations: AGES = Age Gene/Environment Susceptibility; COPD = chronic obstructive pulmonary disease (defined as FEV₁/FVC ratio <70 on spirometry); COPDGene = Genetic Epidemiology of COPD; ECLIPSE = Evaluation of COPD Longitudinally to Identify Predictive Surrogate End-Points; ILA = interstitial lung abnormalities; IQ = interquartile interval; MESA = Multi-Ethnic Study of Atherosclerosis; SPIROMICS = Subpopulations and Intermediate Outcome Measures in COPD Study.
Missing data: MESA non-Hispanic whites COPD status: no ILA—76, ILA—17; MESA African Americans COPD status: no ILA—58, ILA—9; Framingham Heart Study body mass index—1; AGES-Reykjavik body mass index—1.

Table 2. Genome-Wide Significant Variants Associated with Interstitial Lung Abnormalities and Subpleural-Predominant Interstitial Lung Abnormalities, and Replication in an Idiopathic Pulmonary Fibrosis Cohort

Chromosome/ Location	Position	rsID	Risk Allele	Risk Allele Frequency	Nearest Gene	ILA vs. No ILA		Subpleural ILA vs. No ILA		Replication in IPF Cohort	
						Odds Ratio* (95% CI)	P Value	Odds Ratio (95% CI)	P Value	Odds Ratio (95% CI)	P Value
3q13	106571023	rs73199442	T	0.06	<i>FCF1P3</i>	1.68 (1.39–2.02)	5×10^{-8}	1.61 (1.31–1.99)	7×10^{-6}	0.98 (0.85–1.12)	0.73
5q12	62172476	rs6886640	G	0.62	<i>IPO11</i>	1.28 (1.18–1.41)	4×10^{-8}	1.27 (1.14–1.40)	8×10^{-6}	1.06 (0.99–1.14)	0.11
6q15	87737841	rs7744971	G	0.28	<i>HTR1E</i>	1.26 (1.16–1.37)	1×10^{-7}	1.32 (1.19–1.45)	4×10^{-8}	1.01 (0.94–1.09)	0.75
11p15	1241221	rs35705950	T	0.11	<i>MUC5B</i>	1.97 (1.74–2.22)	3×10^{-27}	2.22 (1.93–2.55)	2×10^{-29}	4.84 (4.37–5.36)	1×10^{-203}

Definition of abbreviations: CI = confidence interval; ILA = interstitial lung abnormality; IPF = idiopathic pulmonary fibrosis.

*Odds ratios are per copy of the risk allele.

threshold for significance after adjustment for multiple testing) in association with ILAs (Tables 3 and E5). All but one of the 12 IPF GWAS SNPs had a consistent direction of risk effect in IPF and ILAs. Despite the smaller sample size in the subpleural-predominant ILA association analysis, IPF genetic risk loci were generally more strongly associated (larger ORs and smaller *P* values) with the risk of subpleural ILAs than with the risk of ILAs.

We assessed the top 21 loci reported in a GWAS of several HAA phenotypes (36) in our ILA and subpleural GWAS results, and found no overlap of genetic loci between HAAs and ILAs; however, the direction of effect between the risk of HAAs and risk of ILAs and subpleural ILAs was generally consistent (Table E6).

To evaluate the overlap in genetic susceptibility to ILAs and smoking behaviors, we performed a lookup of our four genome-wide significant ILA variants in a recent GWAS of four smoking behaviors: smoking initiation, age of smoking initiation, smoking cessation, and cigarettes per day (37). The *P* value was >0.05 for association of the top four ILA SNPs with any smoking behavior (Table E7). We also assessed the genome-wide significant loci reported in the smoking GWAS (37). After correction for multiple testing, there was no significant overlap of smoking behavior SNPs with ILAs or subpleural-predominant ILAs (Table E8).

In a search for CTD-associated ILD and sarcoidosis-associated SNPs in the NHGRI-EBI GWAS Catalog, we found 357 SNPs associated with 17 traits reported in 39 publications. No SNPs were associated with ILAs. Only one CTD SNP (rs13389408, intronic to *STAT4* on chromosome 2) met the threshold for Bonferroni significance in association with subpleural ILAs (OR, 1.3; 95% CI, 1.1–1.5;

$P = 9.7 \times 10^{-4}$). The SNP rs13389408 was discovered in a meta-analysis of “systemic seropositive rheumatic diseases” (including systemic sclerosis, systemic lupus erythematosus, and idiopathic inflammatory myopathies) (44).

Logistic Regression Conditioning on *MUC5B* at 11p15

Our conditional analysis did not identify any conditionally distinct signals at 3q13, 5q12, or 6p15. However, a previous GWAS of IPF reported variant associations in *TOLLIP* (rs5743894, rs5743890, and rs111521887) that—despite proximity to *MUC5B* in the 11p15 region—were reported to be independent of the rs35705950 association with IPF due to minimal linkage disequilibrium ($R^2 < 0.2$) (9). In the COPDGene study non-Hispanic white and African American participants, we performed a meta-analysis of the association of previously reported *TOLLIP* SNPs with ILAs and subpleural ILAs, and conditioned each *TOLLIP* SNP association on the *MUC5B* rs35705950 genotype. When each *TOLLIP* SNP association was adjusted for the rs35705950 genotype, the *TOLLIP* SNPs’ effect sizes and strengths of association were diminished (Table E9). These data suggest that the *TOLLIP* SNP associations with ILAs and subpleural-predominant ILAs in COPDGene are not independent of *MUC5B* rs35705950.

eQTL Assessments for Identified Loci

We sought to determine whether our four genome-wide significant ILA- and subpleural ILA-associated variants have been reported as lung or blood eQTLs. The ILA risk variants at 5q12 (rs6886640) and 3q13 (rs73199442) were not reported as lung or blood eQTLs in any of the examined data. The *MUC5B* promoter polymorphism (rs35705950) T allele was associated with increased expression of

MUC5B in the lung (*q* value = 3.99×10^{-9}) in the GTEx lung eQTL data, but not in Hao and colleagues’ lung eQTL data (41). Furthermore, the rs35705950 T allele was associated with decreased expression of *CD151* in blood in the eQTL Consortium *cis*-eQTL data (43). The subpleural ILA 6q15 variant rs7744971 risk allele (G) was significantly associated with decreased blood expression of *AKIRIN2*, *SLC35A1*, *C6orf164*, and *RP1-102H19.6*, and increased blood expression of *ZNF292* in the eQTL Consortium *cis*-eQTL data (43). In the Netherlands Study of Depression and Anxiety/Netherlands Twin Register Conditional eQTL Catalog (42) the rs7744971 G allele was also associated with decreased *AKIRIN2* expression as well as with decreased *C6orf162* expression in blood.

Discussion

Our study, which presents the first GWAS of visually assessed ILAs, included 1,699 participants with ILAs and 10,274 control subjects, and has several notable findings. First, we provide the most comprehensive data to date demonstrating the links between genetic association findings in patients with IPF and research participants with ILAs. For example, these findings provide at least nominal, and directionally consistent, evidence for an association between ILAs and most of the common genetic variants that have previously been demonstrated to be associated with IPF (7–11). In addition, our results provide genome-wide significant evidence for association with two new genetic risk loci for ILAs overall (3q13 and 5q12) and one loci for subpleural-predominant ILAs specifically (6q15). These new loci do not show evidence of association with IPF, and although these data could represent

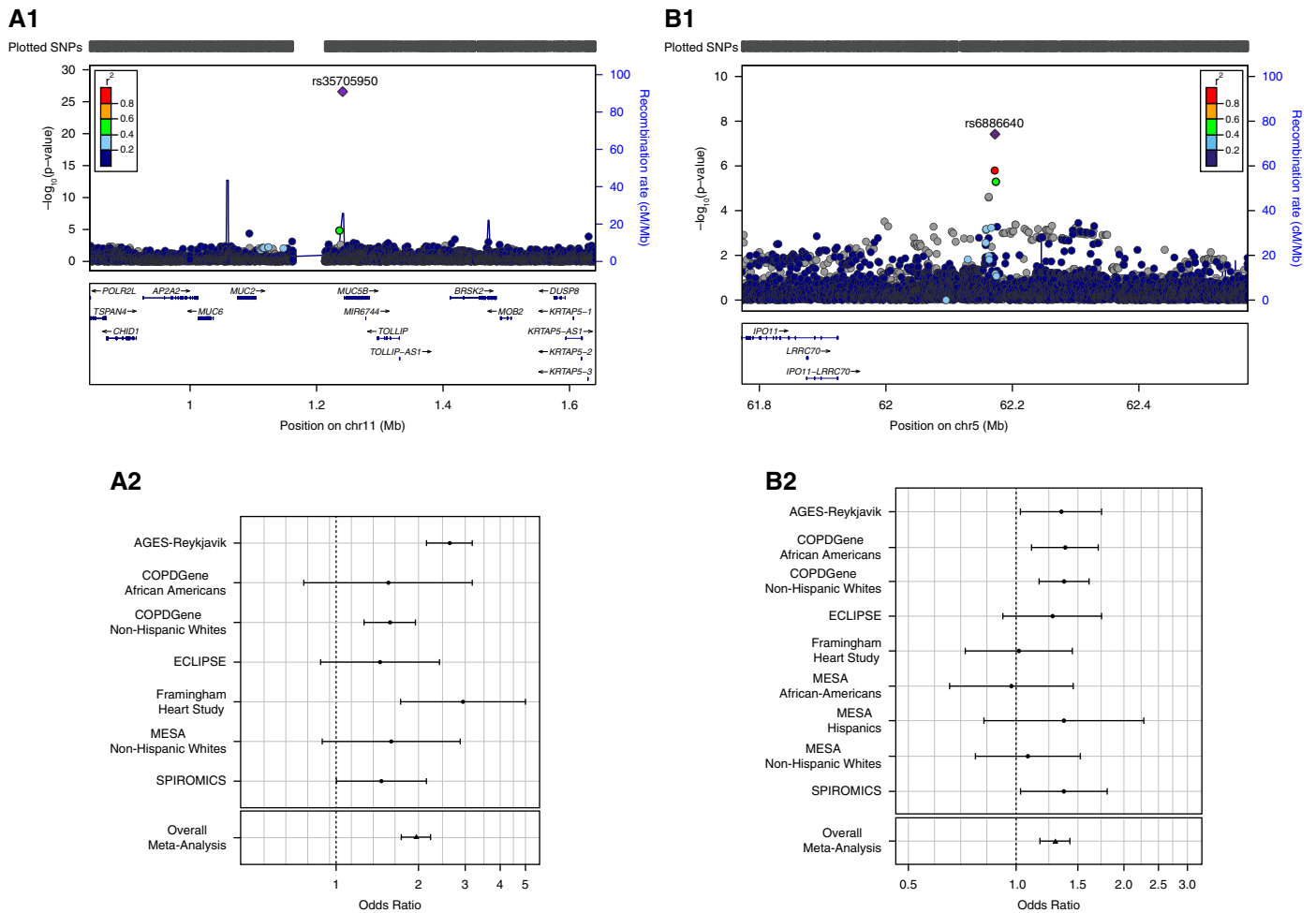


Figure 2. Locus zoom and forest plots for the genome-wide significant loci associated with interstitial lung abnormalities (ILAs) and subpleural-predominant ILAs. (A) Comparison of participants with and without ILAs. A1 is a locus zoom plot demonstrating the genome-wide significant association at rs35705950 (nearest gene *MUC5B*); A2 is a forest plot demonstrating the results in each individual cohort and the overall meta-analysis, with the x-axis on the log odds scale. (B) Comparison of participants with and without ILAs. B1 is a locus zoom plot demonstrating the genome-wide significant association at rs6886640 (nearest gene *IPO11*); B2 is a forest plot demonstrating the results in each individual cohort and the overall meta-analysis, with the x-axis on the log odds scale. (C) Comparison of participants with and without ILAs. C1 is a locus zoom plot demonstrating the genome-wide significant association at rs73199442 (nearest gene *FCF1P3*); C2 is a forest plot demonstrating the results in each individual cohort and the overall meta-analysis, with the x-axis on the log odds scale. (D) Comparison of participants with subpleural-predominant ILAs and those without ILAs. D1 is a locus zoom plot demonstrating the genome-wide significant association at rs7744971 (nearest gene *HTR1E*); D2 is a forest plot demonstrating the results in each individual cohort and the overall meta-analysis, with the x-axis on the log odds scale. AGES = Age Gene/Environment Susceptibility; COPD = chronic obstructive pulmonary disease; COPDGene = Genetic Epidemiology of COPD; ECLIPSE = Evaluation of COPD Longitudinally to Identify Predictive Surrogate End-points; MESA = Multi-Ethnic Study of Atherosclerosis; SPIROMICS = Subpopulations and Intermediate Outcome Measures in COPD Study.

false-positive associations, they may also be consistent with some cases of ILA representing early stages of diverse forms of ILD, associated with genetic risk factors distinct from IPF.

Multiple lines of evidence now demonstrate a shared genetic risk between some research participants with ILAs who have not received a diagnosis and patients with clinically diagnosed IPF. Similar to what was found in all prior GWASs of IPF

that included the gain-of-function *MUC5B* promoter variant (8–11), our study demonstrates that the *MUC5B* promoter variant rs35705950 has the most significant association with ILAs. In addition, among 12 loci that previously showed genome-wide evidence for association with IPF (7–11) in at least one study, we present evidence for directionally consistent associations in 11, and 5 of these were significant after adjustment for multiple

testing ($P < 0.05/12$). More specifically, this study provides support for the fact that common genetic variants in seven genomic regions are associated with early and/or mild stages of PF in addition to their known association with more advanced stages of disease. In support of the latter statement is the fact that most of these genetic association findings were stronger when the ILA phenotype was limited to those with subpleural

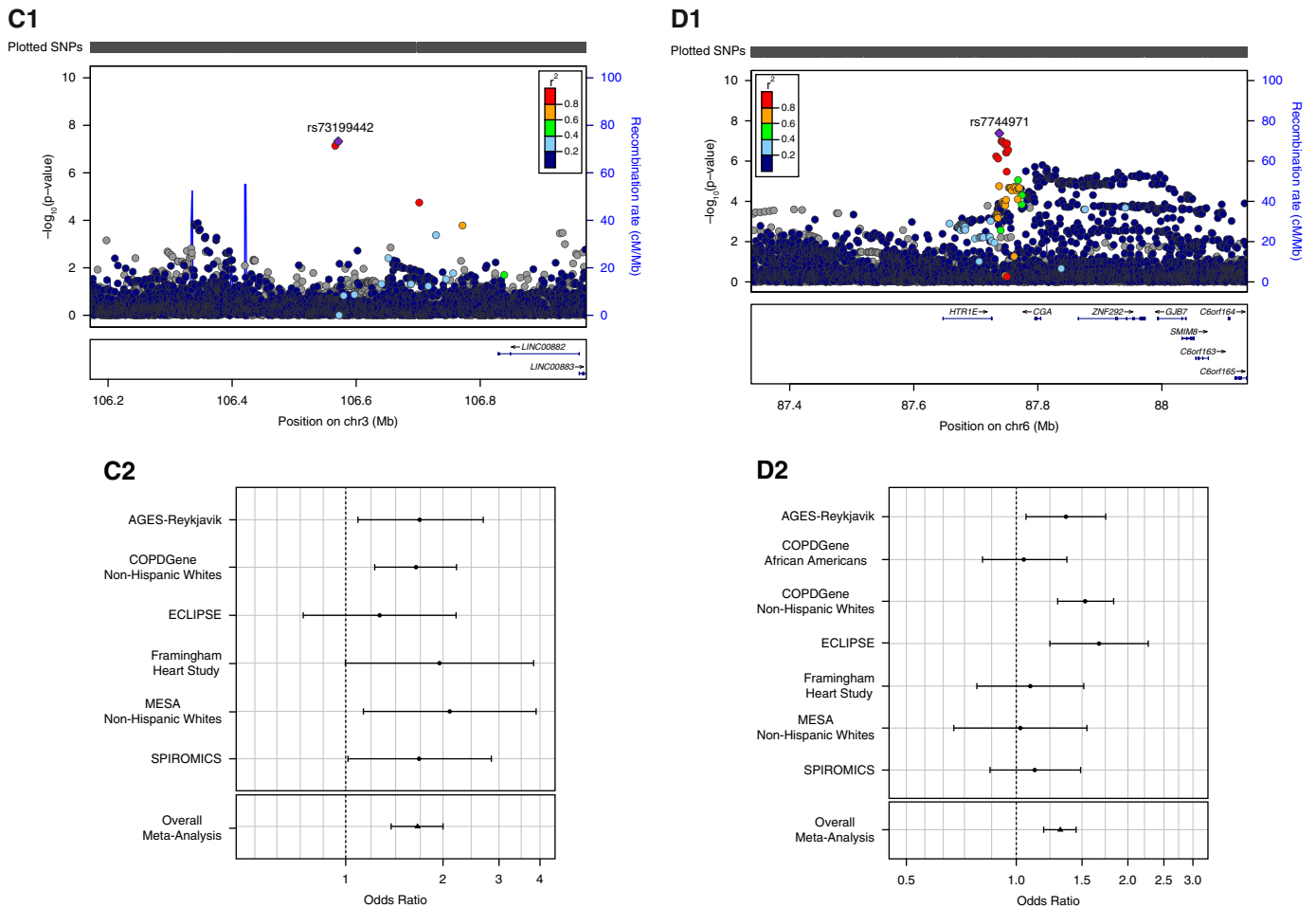


Figure 2. (Continued).

reticular involvement (an imaging phenotype we have previously demonstrated to be associated with subpleural fibrosis) (16).

Our study adds to the growing body of evidence that demonstrates that increasing copies of the gain-of-function minor allele of the *MUC5B* promoter variant (rs35705950) increase the risk of IPF (6, 8–11), ILA (12, 18), and other forms of PF (44), perhaps as the result of an increased expression *MUC5B* in the distal airspaces (6, 45–47). Additionally, our conditional logistic regression analyses are consistent with prior analyses (8) that demonstrated that there do not appear to be additional distinct 11p15 genetic variants associated with either ILA or IPF, after accounting for the effects of the *MUC5B* promoter genotype (rs35705950). Recent studies in mice demonstrated that *Muc5b* overexpression led to impaired mucociliary clearance and the persistence of PF in

response to bleomycin challenge, which could be mitigated by targeted mucolytic agents (48). Future studies will be needed to determine whether intervention in individuals with early stages of *MUC5B*-associated interstitial changes (12, 18) could help to prevent progression to more advanced forms of PF.

Although our study demonstrates that genetic risk factors are shared by patients with IPF and some research participants with ILAs, it also identifies genetic loci in some research participants with ILAs that are distinct. These results may be consistent with the fact that ILAs identify diverse imaging features associated with ILDs in general (24), and it is possible that genetic associations that reflect more diverse pathobiologic and clinical processes than are found in patients with IPF contribute to some of the interstitial imaging findings in these unique diseases (49). For example, given the fact that many of these cohorts

include large populations of smokers, some of these findings could be consistent with a genetic risk of developing smoking-related interstitial fibrosis (50). Although our results do not provide statistically significant evidence of a genotype-by-smoking interaction, the power for these analyses was limited, so this interaction should be evaluated in future studies. In addition, there is some evidence to suggest that the genetic risk of developing ILD in some CTDs may be distinct from the processes that lead to IPF (51–53). Until our findings can be tested in sufficiently large cohorts of these other important populations of individuals with ILD, they should be viewed as preliminary.

The association between the minor allele (G) of the SNP rs7744971 on chromosome 6q14 and subpleural ILAs deserves some mention, as the G allele of this SNP was previously demonstrated to be

Table 3. Association of 12 Previously Identified Idiopathic Pulmonary Fibrosis Genome-Wide Association Loci with Interstitial Lung Abnormalities and Subpleural Interstitial Lung Abnormalities

Chromosome/ Location	rsID	IPF Risk Allele	Nearest Gene	Studies	IPF Odds Ratio (95% CI)	ILA vs. No ILA		Subpleural ILA vs. No ILA		Direction of Effect Consistent with Prior Reports
						Odds Ratio (95% CI)	P Value	Odds Ratio (95% CI)	P Value	
4q22	rs2609255	G	<i>FAM13A</i>	(8)	1.29 (1.18–1.42)	1.18 (1.07–1.29)	5×10^{-4}	1.22 (1.09–1.35)	3×10^{-4}	Yes
6p24	rs2076295	G	<i>DSP</i>	(8, 11)	1.44 (1.35–1.54)	1.14 (1.05–1.2)	0.001	1.18 (1.08–1.29)	3×10^{-4}	Yes
11p15	rs35705950	T	<i>MUC5B</i>	(8)	2.43 (2.13–2.77)	1.97 (1.74–2.22)	3×10^{-27}	2.22 (1.93–2.55)	2×10^{-29}	Yes
15q15	rs2034650	A	<i>IVD</i>	(8)	1.30 (1.19–1.41)	1.08 (0.99–1.17)	0.07	1.15 (1.05–1.26)	0.003	Yes
19p13	rs12610495	G	<i>DPP9</i>	(8)	1.29 (1.18–1.41)	1.14 (1.03–1.26)	0.01	1.23 (1.10–1.37)	2×10^{-4}	Yes
3q26	rs6793295	C	<i>LRRC34</i>	(8)	1.30 (1.19–1.42)	1.06 (0.97–1.15)	0.20	1.12 (1.01–1.24)	0.03	Yes
17q21	rs1981997	G	<i>MAPT</i>	(8)	1.41 (1.28–1.56)	1.16 (1.03–1.30)	0.01	1.19 (1.05–1.36)	0.009	Yes
5p15	rs2736100	A	<i>TERT</i>	(7, 8)	2.11 (1.61–2.78)	1.03 (0.95–1.12)	0.44	1.06 (0.96–1.16)	0.23	Yes
10q24	rs11191865	A	<i>OBFC1</i>	(8)	1.25 (1.15–1.35)	1.03 (0.95–1.12)	0.46	1.03 (0.94–1.13)	0.56	Yes
13q34	rs1278769	G	<i>ATP11A</i>	(8)	1.27 (1.14–1.39)	1.04 (0.95–1.15)	0.37	1.04 (0.94–1.16)	0.45	Yes
15q25	rs62025270	A	<i>AKAP13</i>	(11)	1.27 (1.18–1.37)	1.09 (0.99–1.20)	0.08	1.07 (0.96–1.20)	0.23	Yes
7q22	rs4727443	C	<i>LOC100128334/ LOC105375423</i>	(8)	1.30 (1.19–1.41)	0.95 (0.87–1.03)	0.19	0.93 (0.84–1.02)	0.12	No

Definition of abbreviations: CI = confidence interval; ILA = interstitial lung abnormality; IPF = idiopathic pulmonary fibrosis. For each locus, only the SNP is reported.

an eQTL that is expected to result in decreased expression of the gene *AKIRIN2* (42). Basic research has indicated that Akirin-2, which is known to be expressed in the lung (54), is a critical factor in the innate immune system, which helps to regulate inflammatory gene transcription (55) and B-cell activation (56). Although Akirin-2 is required for embryonic development (57), selective knockdown of *akirin2* in myeloid cells in mice was shown to result in impaired inflammatory cytokine production in macrophages in response to Toll-like receptor stimulation (57). It is believed that some of the effect of Akirin-2 is mediated through its important function as a bridge between some transcription factors (e.g., NF- κ B [57] and Twist [58]), which have also been implicated in the development of pulmonary inflammation [59] and fibrosis [60]) and gene transcription. Future work to confirm the role of this variant in expression of *AKIRIN2*, and to test the role of Akirin-2 deficiency in some forms of ILD mediated by pulmonary inflammation may be warranted.

In addition to the lack of statistically significant replication for the novel loci presented here, our study has some other limitations. First, even though our study included nearly all available cohorts in which ILA characterization and genetic testing have been performed, larger sample sizes may still be required to more adequately characterize the genetic risk of ILA. Second, we cannot exclude the possibility that the generalizability of our

findings may be limited by the omission of some participants from these analyses. Third, although we were able to demonstrate genetic associations with ILA and subpleural-predominant ILA, the sample size may have limited our ability to detect associations with additional specific radiologic features and patterns. In addition, we cannot rule out the possibility that intercohort differences in the methods used for ILA characterization or subclassification could have introduced phenotypic heterogeneity, thus influencing our power to detect genetic associations. Efforts to develop standards for ILA characterization across different research populations could help to minimize this concern. Finally, although our findings demonstrate that the genetic risk of ILA in research participants overlaps with the genetic risk of IPF, it is important to note that we do not know the extent to which the genetic risk of IPF is shared with the risk of other forms of PF (e.g., ILD in the setting of rheumatoid arthritis) (45) or could result in a milder or less progressive form of PF.

In conclusion, our study demonstrates that although the *MUC5B* promoter variant is the dominant variant that is common between ILAs and IPF, there are nominally significant associations between ILAs and the majority of genetic loci that were previously associated with IPF. In addition, our findings provide evidence of novel genetic loci associated with ILAs, but not with IPF. These findings provide further evidence that the *DPP9*, *DSP*, *FAM13A*,

IVD, and *MUC5B* loci may be important in the risk of both early and later stages of PF. Our findings also suggest that ILA characterization may help to identify the genetic risk of developing imaging abnormalities that may represent an early stage of other diverse forms of ILD. ■

Author disclosures are available with the text of this article at www.atsjournals.org.

Acknowledgment: The authors thank the SPIROMICS participants and participating physicians, investigators, and staff for making this research possible. More information about the study and how to access SPIROMICS data can be found at www.spiromics.org.

Current and former investigators of the SPIROMICS sites and reading centers:

Neil E. Alexis, Wayne H. Anderson, Mehrdad Arjomandi, Igor Barjaktarevic, R. Graham Barr, Lori A. Bateman, Surya P. Bhatt, Richard C. Boucher, Russell P. Bowler, Stephanie A. Christenson, Alejandro P. Comellas, Christopher B. Cooper, David J. Couper, Gerard J. Criner, Ronald G. Crystal, Jeffrey Curtis, Claire M. Doerschuk, Mark T. Dransfield, Brad Drummond, Christine M. Freeman, Craig Galban, MeiLan K. Han, Nadia N. Hansel, Annette T. Hastie, Eric A. Hoffman, Yvonne Huang, Robert J. Kaner, Richard E. Kanner, Eric C. Kleerup, Jerry A. Krishnan, Lisa M. LaVange, Stephen C. Lazarus, Fernando J. Martinez, Wendy C. Moore, John D. Newell, Jr., Robert Paine III, Laura Paulin, Cheryl Pirozzi, Nirupama Putcha, Elizabeth C. Oelsner, Sanjeev Raman, Stephen I. Rennard, Donald P. Tashkin, J. Michael Wells, Robert A. Wise, Prescott G. Woodruff, Lisa Postow, and Lisa Viviano.

References

- American Thoracic Society; European Respiratory Society. American Thoracic Society/European Respiratory Society International Multidisciplinary Consensus Classification of the Idiopathic Interstitial Pneumonias: this joint statement of the American Thoracic Society (ATS), and the European Respiratory Society (ERS) was adopted by the ATS board of directors, June 2001 and by the ERS Executive Committee, June 2001. *Am J Respir Crit Care Med* 2002; 165:277–304.
- Raghu G, Chen SY, Yeh WS, Maroni B, Li Q, Lee YC, et al. Idiopathic pulmonary fibrosis in US Medicare beneficiaries aged 65 years and older: incidence, prevalence, and survival, 2001–11. *Lancet Respir Med* 2014;2:566–572.
- Raghu G, Collard HR, Egan JJ, Martinez FJ, Behr J, Brown KK, et al.; ATS/ERS/JRS/ALAT Committee on Idiopathic Pulmonary Fibrosis. An official ATS/ERS/JRS/ALAT statement: idiopathic pulmonary fibrosis: evidence-based guidelines for diagnosis and management. *Am J Respir Crit Care Med* 2011;183:788–824.
- Raghu G, Rochweg B, Zhang Y, Garcia CA, Azuma A, Behr J, et al.; American Thoracic Society; European Respiratory Society; Japanese Respiratory Society; Latin American Thoracic Association. An official ATS/ERS/JRS/ALAT clinical practice guideline: treatment of idiopathic pulmonary fibrosis. An update of the 2011 clinical practice guideline. *Am J Respir Crit Care Med* 2015;192:e3–e19.
- Steele MP, Speer MC, Loyd JE, Brown KK, Herron A, Slifer SH, et al. Clinical and pathologic features of familial interstitial pneumonia. *Am J Respir Crit Care Med* 2005;172:1146–1152.
- Seibold MA, Wise AL, Speer MC, Steele MP, Brown KK, Loyd JE, et al. A common MUC5B promoter polymorphism and pulmonary fibrosis. *N Engl J Med* 2011;364:1503–1512.
- Mushiroda T, Wattanapokayakit S, Takahashi A, Nukiwa T, Kudoh S, Ogura T, et al.; Pirfenidone Clinical Study Group. A genome-wide association study identifies an association of a common variant in TERT with susceptibility to idiopathic pulmonary fibrosis. *J Med Genet* 2008;45:654–656.
- Fingerlin TE, Murphy E, Zhang W, Peljto AL, Brown KK, Steele MP, et al. Genome-wide association study identifies multiple susceptibility loci for pulmonary fibrosis. *Nat Genet* 2013;45:613–620.
- Noth I, Zhang Y, Ma SF, Flores C, Barber M, Huang Y, et al. Genetic variants associated with idiopathic pulmonary fibrosis susceptibility and mortality: a genome-wide association study. *Lancet Respir Med* 2013;1:309–317.
- Fingerlin TE, Zhang W, Yang IV, Ainsworth HC, Russell PH, Blumhagen RZ, et al. Genome-wide imputation study identifies novel HLA locus for pulmonary fibrosis and potential role for auto-immunity in fibrotic idiopathic interstitial pneumonia. *BMC Genet* 2016;17:74.
- Allen RJ, Porte J, Braybrooke R, Flores C, Fingerlin TE, Oldham JM, et al. Genetic variants associated with susceptibility to idiopathic pulmonary fibrosis in people of European ancestry: a genome-wide association study. *Lancet Respir Med* 2017;5:869–880.
- Hunninghake GM, Hatabu H, Okajima Y, Gao W, Dupuis J, Latourelle JC, et al. MUC5B promoter polymorphism and interstitial lung abnormalities. *N Engl J Med* 2013;368:2192–2200.
- Washko GR, Hunninghake GM, Fernandez IE, Nishino M, Okajima Y, Yamashiro T, et al.; COPDGene Investigators. Lung volumes and emphysema in smokers with interstitial lung abnormalities. *N Engl J Med* 2011;364:897–906.
- Lederer DJ, Enright PL, Kawut SM, Hoffman EA, Hunninghake G, van Beek EJ, et al. Cigarette smoking is associated with subclinical parenchymal lung disease: the Multi-Ethnic Study of Atherosclerosis (MESA)-lung study. *Am J Respir Crit Care Med* 2009; 180:407–414.
- Putman RK, Hatabu H, Araki T, Gudmundsson G, Gao W, Nishino M, et al. Evaluation of COPD Longitudinally to Identify Predictive Surrogate Endpoints (ECLIPSE) Investigators; COPDGene Investigators. Association between interstitial lung abnormalities and all-cause mortality. *JAMA* 2016;315:672–681.
- Miller ER, Putman RK, Vivero M, Hung Y, Araki T, Nishino M, et al. Interstitial lung abnormalities and histopathologic correlates in patients undergoing lung nodule resection [abstract]. *Am J Respir Crit Care Med* 2017;195:A1120.
- Podolanczuk AJ, Oelsner EC, Barr RG, Bernstein EJ, Hoffman EA, Easthausen IJ, et al. High-attenuation areas on chest computed tomography and clinical respiratory outcomes in community-dwelling adults. *Am J Respir Crit Care Med* 2017;196:1434–1442.
- Putman RK, Gudmundsson G, Araki T, Nishino M, Sigurdsson S, Gudmundsson EF, et al. The MUC5B promoter polymorphism is associated with specific interstitial lung abnormality subtypes. *Eur Respir J* 2017;50:1700537.
- American Thoracic Society; European Respiratory Society. American Thoracic Society/European Respiratory Society International Multidisciplinary Consensus Classification of the Idiopathic Interstitial Pneumonias: this joint statement of the American Thoracic Society (ATS), and the European Respiratory Society (ERS) was adopted by the ATS board of directors, June 2001 and by the ERS Executive Committee, June 2001. *Am J Respir Crit Care Med* 2002; 165:277–304.
- Putman RK, Hobbs BD, Araki T, Nishino M, Dupuis J, Sun F, et al. Genetic determinants of interstitial lung abnormalities [abstract]. *Am J Respir Crit Care Med* 2018;197:A6181.
- Hurst JR, Vestbo J, Anzueto A, Locantore N, Müllerova H, Tal-Singer R, et al.; Evaluation of COPD Longitudinally to Identify Predictive Surrogate Endpoints (ECLIPSE) Investigators. Susceptibility to exacerbation in chronic obstructive pulmonary disease. *N Engl J Med* 2010;363:1128–1138.
- Harris TB, Launer LJ, Eiriksdottir G, Kjartansson O, Jonsson PV, Sigurdsson G, et al. Age, gene/environment susceptibility-Reykjavik study: multidisciplinary applied phenomics. *Am J Epidemiol* 2007; 165:1076–1087.
- Couper D, LaVange LM, Han M, Barr RG, Bleecker E, Hoffman EA, et al.; SPIROMICS Research Group. Design of the Subpopulations and Intermediate Outcomes in COPD Study (SPIROMICS). *Thorax* 2014;69:491–494.
- Washko GR, Lynch DA, Matsuoka S, Ross JC, Umeoka S, Diaz A, et al. Identification of early interstitial lung disease in smokers from the COPDGene Study. *Acad Radiol* 2010;17:48–53.
- Podolanczuk AJ, Oelsner EC, Barr RG, Hoffman EA, Armstrong HF, Austin JH, et al. High attenuation areas on chest computed tomography in community-dwelling adults: the MESA study. *Eur Respir J* 2016;48:1442–1452.
- McCarthy S, Das S, Kretzschmar W, Delaneau O, Wood AR, Teumer A, et al.; Haplotype Reference Consortium. A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet* 2016;48: 1279–1283.
- Abecasis GR, Auton A, Brooks LD, DePristo MA, Durbin RM, Handsaker RE, et al.; 1000 Genomes Project Consortium. An integrated map of genetic variation from 1,092 human genomes. *Nature* 2012;491:56–65.
- Firth D. Bias reduction of maximum likelihood estimates. *Biometrika* 1993;80:27–38.
- Ma C, Blackwell T, Boehnke M, Scott LJ; GoT2D investigators. Recommended joint and meta-analysis strategies for case-control association testing of single low-count variants. *Genet Epidemiol* 2013;37:539–550.
- Winkler TW, Day FR, Croteau-Chonka DC, Wood AR, Locke AE, Mägi R, et al.; Genetic Investigation of Anthropometric Traits (GIANT) Consortium. Quality control and conduct of genome-wide association meta-analyses. *Nat Protoc* 2014;9:1192–1212.
- Abecasis G, Li Y, Willer C. METAL MetaAnalysis Helper. Version release 2011-03-25;2011. Available from: http://genome.sph.umich.edu/wiki/METAL_Program.
- Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 2010;26: 2190–2191.
- Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet* 2011;88:76–82.
- Yang J, Ferreira T, Morris AP, Medland SE, Madden PA, Heath AC, et al.; Genetic Investigation of Anthropometric Traits (GIANT) Consortium; DIAbetes Genetics Replication And Meta-analysis (DIAGRAM) Consortium. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. *Nat Genet* 2012;44:369–375, S1–3.

35. Welter D, MacArthur J, Morales J, Burdett T, Hall P, Junkins H, *et al.* The NHGRI GWAS Catalog, a curated resource of SNP-trait associations. *Nucleic Acids Res* 2014;42:D1001–D1006.
36. Manichaikul A, Wang XQ, Sun L, Dupuis J, Borczuk AC, Nguyen JN, *et al.* Genome-wide association study of subclinical interstitial lung disease in MESA. *Respir Res* 2017;18:97.
37. Liu M, Jiang Y, Wedow R, Li Y, Brazel DM, Chen F, *et al.*; 23andMe Research Team; HUNT All-In Psychiatry. Association studies of up to 1.2 million individuals yield new insights into the genetic etiology of tobacco and alcohol use. *Nat Genet* 2019;51:237–244.
38. GTEx Consortium. The genotype-tissue expression (GTEx) project. *Nat Genet* 2013;45:580–585.
39. GTEx Consortium. Human genomics: the genotype-tissue expression (GTEx) pilot analysis: multitissue gene regulation in humans. *Science* 2015;348:648–660.
40. Westra HJ, Peters MJ, Esko T, Yaghootkar H, Schurmann C, Kettunen J, *et al.* Systematic identification of trans eQTLs as putative drivers of known disease associations. *Nat Genet* 2013;45:1238–1243.
41. Hao K, Bossé Y, Nickle DC, Paré PD, Postma DS, Laviolette M, *et al.* Lung eQTLs to help reveal the molecular underpinnings of asthma. *PLoS Genet* 2012;8:e1003029.
42. Jansen R, Hottenga JJ, Nivard MG, Abdellaoui A, Laport B, de Geus EJ, *et al.* Conditional eQTL analysis reveals allelic heterogeneity of gene expression. *Hum Mol Genet* 2017;26:1444–1451.
43. Vösa U, Claringbould A, Westra H-J, Bonder MJ, Deelen P, Zeng B, *et al.* Unraveling the polygenic architecture of complex traits using blood eQTL meta-analysis [preprint]. *bioRxiv* 2018;DOI: <https://doi.org/10.1101/447367>.
44. Acosta-Herrera M, Kerick M, González-Serna D, Wijmenga C, Franke A, Gregersen PK, *et al.*; Myositis Genetics Consortium; Scleroderma Genetics Consortium. Genome-wide meta-analysis reveals shared new *loci* in systemic seropositive rheumatic diseases. *Ann Rheum Dis* 2019;78:311–319.
45. Juge PA, Lee JS, Ebsstein E, Furukawa H, Dobrinskikh E, Gazal S, *et al.* MUC5B promoter variant and rheumatoid arthritis with interstitial lung disease. *N Engl J Med* 2018;379:2209–2219.
46. Kropski JA, Pritchett JM, Zoz DF, Crossno PF, Markin C, Garnett ET, *et al.* Extensive phenotyping of individuals at-risk for familial interstitial pneumonia reveals clues to the pathogenesis of interstitial lung disease. *Am J Respir Crit Care Med* 2014;191:417–426.
47. Nakano Y, Yang IV, Walts AD, Watson AM, Helling BA, Fletcher AA, *et al.* MUC5B promoter variant rs35705950 affects MUC5B expression in the distal airways in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2016;193:464–466.
48. Hancock LA, Hennessy CE, Solomon GM, Dobrinskikh E, Estrella A, Hara N, *et al.* Muc5b overexpression causes mucociliary dysfunction and enhances lung fibrosis in mice. *Nat Commun* 2018;9:5363.
49. Travis WD, Costabel U, Hansell DM, King TE Jr, Lynch DA, Nicholson AG, *et al.*; ATS/ERS Committee on Idiopathic Interstitial Pneumonias. An official American Thoracic Society/European Respiratory Society statement: update of the international multidisciplinary classification of the idiopathic interstitial pneumonias. *Am J Respir Crit Care Med* 2013;188:733–748.
50. Katzenstein AL. Smoking-related interstitial fibrosis (SRIF): pathologic findings and distinction from other chronic fibrosing lung diseases. *J Clin Pathol* 2013;66:882–887.
51. Johnson CRP, Lloyd T, McGready J, Horton M, Hall J, Mammen A, *et al.* MUC5B promoter variant is not associated with myositis-related interstitial lung disease [abstract]. *Chest* 2012;142:422A.
52. Peljto AL, Steele MP, Fingerlin TE, Hinchcliff ME, Murphy E, Podlasky S, *et al.* The pulmonary fibrosis-associated MUC5B promoter polymorphism does not influence the development of interstitial pneumonia in systemic sclerosis. *Chest* 2012;142:1584–1588.
53. Borie R, Crestani B, Dieude P, Nunes H, Allanore Y, Kannengiesser C, *et al.* The MUC5B variant is associated with idiopathic pulmonary fibrosis but not with systemic sclerosis interstitial lung disease in the European Caucasian population. *PLoS One* 2013;8:e70621.
54. Bosch PJ, Fuller LC, Weiner JA. An essential role for the nuclear protein Akirin2 in mouse limb interdigital tissue regression. *Sci Rep* 2018;8:12240.
55. Tartey S, Takeuchi O. Chromatin remodeling and transcriptional control in innate immunity: emergence of Akirin2 as a novel player. *Biomolecules* 2015;5:1618–1633.
56. Tartey S, Matsushita K, Imamura T, Wakabayashi A, Ori D, Mino T, *et al.* Essential function for the nuclear protein Akirin2 in B cell activation and humoral immune responses. *J Immunol* 2015;195:519–527.
57. Goto A, Matsushita K, Gesellchen V, El Chamy L, Kutenkeuler D, Takeuchi O, *et al.* Akirins are highly conserved nuclear proteins required for NF-kappaB-dependent gene expression in *Drosophila* and mice. *Nat Immunol* 2008;9:97–104.
58. Nowak SJ, Aihara H, Gonzalez K, Nibu Y, Baylies MK. Akirin links twist-regulated transcription with the Brahma chromatin remodeling complex during embryogenesis. *PLoS Genet* 2012;8:e1002547.
59. Christman JW, Sadikot RT, Blackwell TS. The role of nuclear factor-kappa B in pulmonary diseases. *Chest* 2000;117:1482–1487.
60. Tan J, Tedrow JR, Nouriaie M, Dutta JA, Miller DT, Li X, *et al.* Loss of Twist1 in the mesenchymal compartment promotes increased fibrosis in experimental lung injury by enhanced expression of CXCL12. *J Immunol* 2017;198:2269–2285.