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1	Genetically predicted circulating protein biomarkers and ovarian cancer risk
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- 53 <u>Abstract</u>
- 54

55 Objective

57 Most women with epithelial ovarian cancer (EOC) are diagnosed after the disease has metastasized 58 and survival in this group remains poor. Circulating proteins associated with the risk of developing 59 EOC have the potential to serve as biomarkers for early detection and diagnosis. We integrated large-60 scale genomic and proteomic data to identify novel plasma proteins associated with EOC risk.

- 61
- 62 Methods
- 63

We used the germline genetic variants most strongly associated (*P*<1.5×10⁻¹¹) with plasma levels of
1,329 proteins in 3,301 healthy individuals from the INTERVAL study to predict circulating levels of
these proteins in 22,406 EOC cases and 40,941 controls from the Ovarian Cancer Association
Consortium (OCAC). Association testing was performed by weighting the beta coefficients and
standard errors for EOC risk from the OCAC study by the inverse of the beta coefficients from
INTERVAL.

- 70
- 71 Results

72

We identified 26 proteins whose genetically predicted circulating levels were associated with EOC
risk at false discovery rate<0.05. The 26 proteins included MFAP2, SEMG2, DLK1, and NTNG1 and a
group of 22 proteins whose plasma levels were predicted by variants at chromosome 9q34.2. All 26
protein association signals identified were driven by association with the high-grade serous
histotype that comprised 58% of the EOC cases in OCAC. Regional genomic plots confirmed overlap

- 78 of the genetic association signal underlying both plasma protein level and EOC risk for the 26
- 79 proteins. Pathway analysis identified enrichment of seven biological pathways among the 26
- 80 proteins (*P*_{adjusted}<0.05), highlighting roles for Focal Adhesion-PI3K-Akt-mTOR and Notch signaling.
- 8182 Conclusion
- 83

The identified proteins further illuminate the etiology of EOC and represent promising new EOC
biomarkers for targeted validation by studies involving direct measurement of plasma proteins in EOC
patient cohorts.

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88 <u>Keywords</u>

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90 epithelial ovarian cancer, risk, circulating proteins, circulating biomarkers, genome-wide association
91 study

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104	Re	search Highlights
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106	٠	This study analyzed 667 germline genetic variants known to be associated with circulating
107		(plasma) levels of 1,329 proteins
108		
109	•	These variants were used to predict plasma protein levels in 22,406 epithelial ovarian cancer
110		cases and 40,941 controls
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112	•	Genetically predicted levels of 26 proteins were associated with all invasive epithelial ovarian
112	•	cancer rick
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114		The identified protoing wave envished for the Feed Adhesian DI2K Alt prTOD signaling Natch
115	•	The identified proteins were enriched for the Focal Adhesion-PISK-Akt-III OR signaling, Notch
110		signaling, and other pathways
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118	•	The identified proteins have the potential to serve as circulating biomarkers particularly for high-
119		grade serous epithelial ovarian cancer risk
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155 Introduction

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157 Ovarian cancer is the most common cause of death from gynecological malignancy in the United States and accounted for an estimated 295,000 incident cases and 184,000 deaths globally in 2018 158 [1,2]. Despite advances in treatment, survival rates in ovarian cancer continue to remain low, in 159 160 part, due to the late detection of most cases [3]. Nearly four decades after its discovery [4], 161 circulating levels of the protein cancer antigen 125 (CA-125) continue to be used to screen women at 162 high risk of developing ovarian cancer, such as those with a hereditary cancer syndrome, and 163 women with abnormal findings on examination and/or ultrasound. However, CA-125 has limited 164 sensitivity and specificity in these settings [5]. Furthermore, screening asymptomatic women for CA-165 125 level, despite the use of serial measurements and algorithmic approaches to the interpretation 166 of these levels [6] – and even in combination with transvaginal ultrasound – does not reduce ovarian 167 cancer mortality and is not recommended by the US Preventive Services Task Force [7]. Human 168 epididymis secretory protein E4 (HE4) has been developed in recent years as a blood-based protein 169 biomarker for the diagnosis of ovarian carcinoma [8], and the combination of CA-125 and HE4 is a 170 more accurate predictor of ovarian malignancy than either biomarker alone [9]. However, there 171 remains an urgent unmet need to identify novel circulating protein biomarkers that will be more 172 useful for the early detection of this aggressive disease.

173

Studies in search of new plasma protein biomarkers in ovarian cancer have been restricted to small sample sizes and evaluated limited protein panels [10,11]. In the current study, we adopted a different approach to the identification of circulating protein biomarkers of ovarian cancer risk using large-scale data from two genome-wide association studies (GWAS). The first data set was a GWAS of healthy blood donors in the INTERVAL study that has identified robust associations between inherited genetic variants and plasma protein levels [12]. The second data set was the the largest and latest published GWAS meta-analysis from the Ovarian Cancer Association Consortium

181	(OCAC) [13]. While epithelial ovarian cancer (EOC) accounts for approximately 90% of all ovarian
182	cancer cases, EOC itself is a diverse entity with distinct histological subtypes: high-grade serous (the
183	most common and lethal histotype), low-grade serous, clear cell, mucinous, endometrioid, and low
184	malignant potential (serous or mucinous) tumors. The OCAC GWAS included associations with all
185	invasive and histotype-specific EOC susceptibility. We used the inherited genetic variants robustly
186	associated with plasma protein levels in the INTERVAL GWAS to predict these levels in the OCAC
187	GWAS where plasma protein levels have not actually been measured but the variants have been
188	genotyped. Such predictions are likely to suffer from less selection bias and confounding because
189	the genetic variants on which they are based are randomly allocated at gametogenesis and fixed
190	after conception. Our study design enabled a comprehensive appraisal of the role of the levels of
191	over 1,300 plasma proteins in more than 22,000 ovarian cancer cases and over 40,000 controls.
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193	Methods
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195	Circulating (plasma) protein data set
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197	We used effect size estimates (beta coefficients) from genome-wide association analyses linking 667
197 198	We used effect size estimates (beta coefficients) from genome-wide association analyses linking 667 single nucleotide polymorphisms (SNPs) to the circulating (plasma) levels of 1,329 proteins in 3,301
197 198 199	We used effect size estimates (beta coefficients) from genome-wide association analyses linking 667 single nucleotide polymorphisms (SNPs) to the circulating (plasma) levels of 1,329 proteins in 3,301 healthy participants from the INTERVAL study [12], a bioresource of blood donors in England who
197 198 199 200	We used effect size estimates (beta coefficients) from genome-wide association analyses linking 667 single nucleotide polymorphisms (SNPs) to the circulating (plasma) levels of 1,329 proteins in 3,301 healthy participants from the INTERVAL study [12], a bioresource of blood donors in England who were recruited into a multi-center randomized trial of blood donation frequency [14]. Each of these
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197 198 199 200 201 202 203 204	We used effect size estimates (beta coefficients) from genome-wide association analyses linking 667 single nucleotide polymorphisms (SNPs) to the circulating (plasma) levels of 1,329 proteins in 3,301 healthy participants from the INTERVAL study [12], a bioresource of blood donors in England who were recruited into a multi-center randomized trial of blood donation frequency [14]. Each of these SNPs was associated with at least one of the plasma proteins at genome-wide significance (defined as $P < 1.5 \times 10^{-11}$ in the INTERVAL analysis [12]) and was the SNP most strongly associated with the circulating levels of that protein. Five hundred and eight-five SNPs were associated with the levels of only one circulating protein each while 82 SNPs were associated with multiple proteins (ranging
197 198 199 200 201 202 203 204 205	We used effect size estimates (beta coefficients) from genome-wide association analyses linking 667 single nucleotide polymorphisms (SNPs) to the circulating (plasma) levels of 1,329 proteins in 3,301 healthy participants from the INTERVAL study [12], a bioresource of blood donors in England who were recruited into a multi-center randomized trial of blood donation frequency [14]. Each of these SNPs was associated with at least one of the plasma proteins at genome-wide significance (defined as $P < 1.5 \times 10^{-11}$ in the INTERVAL analysis [12]) and was the SNP most strongly associated with the circulating levels of that protein. Five hundred and eight-five SNPs were associated with the levels of only one circulating protein each while 82 SNPs were associated with multiple proteins (ranging from 36 SNPs that were associated with two proteins each to one SNP that was associated with 95

proteins; Table S1). We restricted analysis to SNPs that had minor allele frequency (MAF) > 1% and

had either been genotyped or imputed with quality score > 0.8 – both in the INTERVAL analytic
sample and in the OCAC data set (described below). These SNP-protein associations included 908
<i>trans</i> -associations where the top SNP associated with the protein was > 1 Mb away from the gene
encoding the protein and 421 <i>cis</i> -associations where the top SNP associated with the protein was < 1
Mb away from the gene encoding the protein. Plasma protein levels in the INTERVAL study were
quantified using an expanded aptamer-based multiplex protein assay called SOMAscan [12,15] and
germline genotypes were measured on Affymetrix Axiom UK Biobank array with imputation into a
combined combined 1000 Genomes Phase 3-UK10K reference panel. We used the same protein
names and identifiers, including UniProt and SOMAmer IDs (Table S1), as used in the original
INTERVAL genetic report [12] for consistency. That report contains additional details of sample and
genotype quality control, imputation, and association analysis in the INTERVAL study.
Epithelial ovarian cancer data set
Summary statistics (beta coefficients and standard errors) from a GWAS meta-analysis for EOC
susceptibility in women of European ancestry were obtained from OCAC [13]. The GWAS meta-
analysis included 22,406 invasive EOC cases overall and 40,941 controls and this "all invasive EOC"
case-control set was the focus of the primary analysis in the current study. EOC histotype-specific
summary statistics from the same GWAS meta-analysis were also evaluated for seven histological
subtypes as a secondary analysis in the current study. This included high-grade serous (13,037
cases), low-grade serous (1,012 cases), low malignant potential serous (1,954 cases), invasive
mucinous (1,417 cases), low malignant potential mucinous (1,149), clear cell (1,366), and
endometrioid (2,810 cases) EOC cases and 40,941 controls. Additional details of sample and
genotype quality control, imputation, and association analytic procedures for the OCAC GWAS meta-
analysis have been previously published [13].

233 Statistical analysis

234

235 We used the Wald ratio to estimate the effect of genetically predicted circulating protein levels on 236 ovarian cancer risk. The Wald estimator in this context is the ratio of the beta coefficient for a SNP 237 from the ovarian cancer GWAS meta-analysis to the beta coefficient for the same SNP from the 238 plasma protein genome-wide association analysis. The SNP most strongly associated with the 239 circulating level of each protein in the INTERVAL data set was used. The standard error of the Wald 240 estimator is the ratio of the standard error for the SNP from the ovarian cancer GWAS meta-analysis 241 to the absolute value of the beta coefficient for the SNP from the plasma protein genome-wide 242 association analysis. These analyses were performed using the R (version 3.6.2) statistical 243 computing language. *P*-values were calculated using the formula: 244 "pnorm(abs(Wald_estimator)/standard_error_of_Wald_estimator, lower.tail=FALSE) * 2" and the 245 multiple comparisons burden for testing 1,329 SNP-protein-ovarian cancer associations was 246 accounted for using false discovery rate (FDR) control by the method of Benjamini and Hochberg as 247 implemented in the "p.adjust" function. The Wald estimator allowed for incorporation of the beta 248 coefficient for the SNP from the plasma protein analysis and allowed easy inference of the direction 249 of the association (whether positive or inverse) between plasma protein level and ovarian cancer 250 risk. Therefore, we preferred the Wald estimator over directly testing for the genetic association 251 between the top plasma protein level-associated SNP and ovarian cancer risk (although in practice 252 both approaches provided almost identical P-values). As noted above, our primary analysis was for 253 all invasive EOC risk, given that this combined phenotype had the largest sample size, while in 254 secondary analyses we evaluated histotype-specific risk.

255

We followed up genetically predicted circulating levels of proteins that were found to be associated (FDR < 0.05) in our study with ovarian cancer risk to assess whether the top plasma protein level-associated SNP was part of the top ovarian cancer genetic association signal in the

259 same genomic region – a positional overlap that would reinforce the role of the SNP as a driver of 260 both circulating protein levels and ovarian cancer risk. We did this by visualizing ovarian cancer 261 genetic associations for all SNPs with MAF > 1% and imputation quality > 0.8 in the OCAC data set in 262 the 500 kb window centered on the top protein-associated SNP (i.e., +/- 250 kb on either side) using 263 two-way scatter plots generated in *Stata* (version 14, StataCorp LP, College Station, TX). For SNPs 264 with stronger P-values for association with ovarian cancer risk in OCAC as compared to the top 265 protein-associated SNP, the correlation between the stronger P-value SNPs and the top protein-266 associated SNP was calculated using the LDlink online tool and data from the 1000 Genomes 267 European ancestry populations [16]. If the same SNP association signal drives both plasma protein 268 level and ovarian cancer risk, we expected one of the following three scenarios to be true: (i) the top 269 protein-associated SNP is also the top ovarian cancer associated SNP or (ii) it is strongly correlated 270 $(r^2 > 0.9)$ with the top ovarian cancer associated SNP(s) or (iii) there are multiple independent ($r^2 < 1$ 271 0.01) genetic association signals in OCAC in the same region and the top protein-associated SNP is 272 one of these associations. A second follow-up analysis of proteins that achieved FDR < 0.05 in our 273 study involved mapping these to the genes encoding them and evaluating the genes for enrichment 274 of pathways (at P < 0.05 after adjustment for testing multiple pathways) using the Enrichr online tool 275 [17] and the "WikiPathways Human 2019" database [18] that contains annotations for 472 known 276 biological pathways. A final follow-up analysis involved searching for genome-wide significant 277 associations ($P < 5 \times 10^{-8}$) between the top plasma protein level-associated SNP for each of the 278 proteins that achieved FDR < 0.05 in our study and other diseases and traits in the published (i.e., 279 MEDLINE indexed) literature. This search was performed using the PhenoScanner (version 2) online 280 tool [19], querying published European-ancestry GWAS. The aim was to identify pleiotropic diseases 281 and traits that may provide an alternative explanation for the plasma protein-EOC risk associations 282 identified, stemming from their associations with the same top SNPs. Such pleiotropic diseases and 283 traits associated with the same SNPs may also be the cause or consequence of plasma protein level 284 changes that in turn are associated with EOC risk.

286 <u>Results</u>

288	Genetically predicted circulating levels of 26 proteins were associated with all invasive EOC risk at
289	FDR < 0.05 (13 positive and 13 inverse associations; Table 1 and Table S1). First, this included a
290	positive association between MFAP2 encoded by MFAP2 on chromosome 1 and all invasive EOC risk
291	(Microfibrillar-associated protein 2; P_{Wald} = 1.8 x 10 ⁻⁴ , FDR = 0.01). The top MFAP2 plasma protein-
292	associated SNP rs4920605 ($P_{OCAC-GWAS-all}$ = 1.82 x 10 ⁻⁴) in the INTERVAL study was ~8 kb from the
293	transcription start site (TSS) of <i>MFAP2</i> . There was only one SNP in the same region, rs143483351
294	($P_{OCAC-GWAS-all} = 1.76 \times 10^{-4}$), a multi-allelic variant 2 kb from rs4920605, with a slightly stronger
295	association with all invasive EOC risk (Fig. 1 (a) and Table S2). SNP rs143483351 could not be
296	evaluated in LDlink [16] for correlation with rs4920605 because it was a multi-allelic variant.
297	Second, our FDR < 0.05 results also included an inverse association between NTNG1 encoded by
298	NTNG1 on chromosome 1 (in a genomic region distinct from MFAP2) and all invasive EOC risk
299	(Netrin-G1; P_{Wald} = 4.9 x 10 ⁻⁴ , FDR = 0.03). The top NTNG1 plasma protein-associated SNP
300	rs115668827 ($P_{OCAC-GWAS-all}$ = 4.9 x 10 ⁻⁴) in the INTERVAL study was ~4 kb from the TSS of <i>NTNG1</i> .
301	There was only one SNP in the same region, rs11185086 ($P_{OCAC-GWAS-all} = 3.8 \times 10^{-4}$), 173 kb from
302	rs115668827, with a stronger association with all invasive EOC risk (Fig. 1 (b) and Table S2).
303	However, rs11185086 and rs115668827 represented independent signals in the same region ($r^2 = 7 \text{ x}$
304	10 ⁻⁴). Third, the list of 26 plasma proteins identified included positive associations between SEMG2
305	(Semenogelin-2) and ovarian cancer risk and DLK1 (Protein delta homolog 1) and ovarian cancer risk
306	(for both associations – P_{Wald} = 4.0 x 10 ⁻⁴ , FDR = 0.02). The top SEMG2 plasma protein-associated
307	SNP and the top DLK1 plasma protein-associated SNP in the INTERVAL study was the same SNP,
308	rs12881760 ($P_{OCAC-GWAS-all}$ = 3.96 x 10 ⁻⁴), which is ~16 kb from the TSS of <i>DLK1</i> on chromosome 14.
309	SEMG2 is encoded by SEMG2 on chromosome 20 and rs12881760 is associated with its circulating
310	level by acting in <i>trans</i> . SNP rs12881760 is part of a cluster of three SNPs that includes rs10144381

311 ($P_{OCAC-GWAS-all} = 3.50 \times 10^{-4}$) and rs12881545 ($P_{OCAC-GWAS-all} = 3.56 \times 10^{-4}$), which are within 3 kb of each 312 other and strongly correlated ($r^2 > 0.93$), and together mark the strongest association signal with all 313 invasive EOC risk in the *DLK1* region (Fig. 1 (c) and Table S2).

314

The remaining 22 of the 26 all invasive EOC risk-associated circulating proteins identified were 315 316 proxied by 10 correlated SNPs ($r^2 > 0.38$) spanning a ~10 kb interval on chromosome 9 (Table 1). Three 317 of these SNPs were the top SNP for one protein each, four for two proteins each, two for three proteins 318 each, and one SNP was the top SNP for five proteins in the INTERVAL data set (Table 1). Ten proteins 319 demonstrated a positive association and 12 showed an inverse association with all invasive EOC risk. 320 Twenty-one of the 22 proteins were encoded by genes > 1 Mb away from this chromosome 9 interval 321 (trans-associations) and most were in fact encoded by genes located on other chromosomes. The 322 only exception to this was the plasma protein BGAT (Histo-blood group ABO system transferase) 323 encoded by ABO and the top BGAT plasma level-associated SNP, rs505922, is ~2 kb from the TSS of 324 ABO. The ABO locus (chromosome 9q34.2) is a known genome-wide significant ($P < 5 \times 10^{-8}$) locus for 325 all invasive and high-grade serous ovarian cancer risk [13,20]. The ten protein level-associated SNPs 326 spanned the ABO locus and were among the top 50 all invasive EOC risk SNPs in the 500 kb region 327 (Table S2 and Fig. 1 (d)). The top all invasive EOC risk SNP in the region, rs587729126 ($P_{OCAC-GWAS-all}$ = 328 8.3 x 10^{-10}), was the top SNP in the INTERVAL study for association with circulating levels of FA20B 329 (Glycosaminoglycan xylosylkinase) and sICAM-2 (Intercellular adhesion molecule 2). This overlap of 330 top associations led to these two proteins emerging as the plasma proteins whose genetically 331 predicted levels were most strongly associated with all invasive EOC risk in our analysis (for both 332 associations – P_{Wald} = 8.1 x 10⁻¹⁰, FDR = 4.5 x 10⁻⁷; Table 1). The PhenoScanner search indicated that 333 eight of the ten protein level-associated SNPs that spanned the ABO locus were associated with 62 334 traits (Table S3). Overall, for all 26 proteins identified (associated with SNPs in the regions presented 335 in Fig. 1 and discussed above), we observed a clear overlap between the top circulating protein level-336 associated SNP and the top all invasive EOC risk association, lending further confidence to the

association between plasma protein levels predicted by these SNPs and disease risk. We did not identify any additional proteins at FDR < 0.05 in any of the histotype-specific analyses (Table S1). An inspection of the high-grade serous EOC results (Table S1) confirmed that all 26 FDR < 0.05 protein associations with all invasive EOC risk were driven by associations in the high-grade serous EOC sample, which contributed the largest number of cases to the all invasive EOC sample. Pathway enrichment analysis of the genes encoding the 26 proteins identified seven pathways at $P_{adjusted} < 0.05$ (Table 2).

344

345 Discussion

346

By combining genome-wide association data from 22,406 all invasive EOC cases and 40,941 controls and plasma proteome-wide genetic association data from 3,301 healthy individuals, we identified 26 proteins whose genetically inferred circulating levels were associated with EOC risk after false discovery rate control (FDR < 0.05). The combination of these data sets offered unprecedented scale to evaluate the role of over 1,300 plasma proteins in the development of EOC and identified circulating protein biomarkers with the potential for clinical translational in the early detection and diagnosis of EOC.

354

355 We observed that the top plasma protein level-associated SNP was either the top all invasive 356 EOC risk SNP in the 500 kb region centred on the SNP (Fig. 1 (a) and (d)) or it was the top SNP of one 357 of two independent ($r^2 < 0.01$) all invasive EOC risk associations in the region (Fig. 1 (b)) or it was part 358 of a cluster of highly correlated ($r^2 > 0.9$) SNPs that together marked the top all invasive EOC risk 359 association in the region (Fig. 1 (c)). This suggests that our results are unlikely to be due to linkage 360 disequilibrium contamination, i.e., regional genetic architecture where the top plasma protein level-361 associated SNP is weakly correlated with the top all invasive EOC risk SNP and results in a spurious 362 association underpinned by two distinct SNP signals (one for protein and another for EOC). While

363	the focus of our analysis was the use of SNPs associated with plasma protein levels to evaluate the
364	association between plasma protein levels and EOC risk and not the direct genetic association
365	between SNPs and EOC risk, we note that there were 667 unique SNPs used in the analysis (Table S1)
366	and the 13 unique SNPs underpinning the 26 proteins identified (Table 1) were all associated with all
367	invasive EOC risk at $P < 0.05/667$, which would be the conventional threshold for statistical
368	significance if this was a SNP-based association study of 667 SNPs. Ten of the 13 SNPs were
369	genome-wide significant ($P < 5 \times 10^{-8}$) as they are located at a previously reported all invasive and
370	high-grade serous EOC risk locus at or near ABO on chromosome 9q34.2 [13,20]. The three
371	remaining SNPs (spanning three distinct genomic regions; Fig. 1 (a), (b), and (c)) may well represent
372	as yet unidentified genetic susceptibility loci for all invasive EOC that we are presently
373	underpowered to detect at GWAS levels of significance ($P < 5 \times 10^{-8}$). Thus, loci known to be
374	associated with the plasma proteome may aid in the discovery of sub-threshold GWAS loci for
375	disease susceptibility in much the same way as previously demonstrated for other biological
376	information integrated into genetic association studies [21].

378 Pathway analysis highlighted that five of the 26 proteins whose genetically predicted plasma 379 levels were associated with all invasive EOC risk at FDR < 0.05 belonged to the "Focal Adhesion-PI3K-380 Akt-mTOR-signaling pathway (P_{pathway} (adjusted) = 0.006; Table 2), which was the maximum overlap seen 381 between any established biomolecular pathway and the 26 proteins. The genes encoding these 382 proteins were located across different chromosomes, but the SNPs associated most strongly with 383 their plasma levels were all located at the 9q34.2 locus. The PI3K-Akt-mTOR intracellular signaling 384 cascade is a major regulator of the cell cycle and has key roles in cellular quiescence, growth and 385 proliferation, and cancer cell survival and metastasis [22]. Somatic aberrations in this pathway are 386 found in the majority of high-grade serous ovarian tumors [23]. Another pathway identified at 387 $P_{\text{pathway (adjusted)}} < 0.05$ was Notch signaling and this association was driven, in turn, by associations 388 between genetically predicted circulating levels of two Notch proteins, MFAP2 (chromosome

389 1p36.13) and DLK1 (chromosome 14q32.2), and all invasive EOC risk. It is noteworthy that these two 390 plasma proteins associated with EOC risk at FDR < 0.05 were encoded by genes located on distinct 391 chromosomes but the genes/proteins were members of the same biological pathway. DLK1, a non-392 canonical Notch ligand, has a demonstrated role in promoting ovarian carcinogenesis via Notch 393 activation and epithelial-mesenchymal transition [24]. The microfibrillar-associated protein 2 394 (MFAP2), previously named microfibril-associated glycoprotein 1 (MAGP1), activates integrin 395 signaling and is a potential oncogene [25–27]. Another protein identified at FDR < 0.05, Netrin-G1 or 396 NTNG1, is involved in apoptosis and known to be dysregulated particularly in endocrine-related 397 tumors [28,29]. Further, the gene that encodes NTNG1 has been shown to be overexpressed in 398 malignant ovarian tumors [30].

399

400 Larger genetic association studies of the circulating proteome as well as for EOC 401 susceptibility may identify additional candidate biomarkers for EOC. Moreover, such studies may 402 profile additional proteins (including CA-125, which was not profiled in the INTERVAL study) and 403 include individuals of non-European ancestries, offering new opportunities for plasma protein 404 biomarker discovery. The present study was unable to identify associations for EOC histotypes other 405 than for the most common high-grade serous hisotype and this is another area where larger sample 406 sizes might help. The INTERVAL and OCAC data sets used in this analysis were based on participants 407 of European ancestry and there is a compelling need for similar trans-ancestry analyses. Smaller 408 GWAS of ovarian cancer risk in women of African and East Asian ancestry have been reported by 409 OCAC but there is no circulating protein level GWAS comparable to the INTERVAL study as yet in a 410 cohort that is not of European ancestry [31,32]. A major strength of the current analysis was the 411 ability to appraise the roles of over 1,300 proteins. A vital next step in assessing the role of the 412 plasma proteome in EOC risk and validating our findings will involve directly measuring the 26 413 proteins shortlisted by our study in EOC case and control sample collections that have pre-diagnostic 414 and longitudinal follow-up biospecimens available such as the Prostate, Lung, Colorectal and Ovarian

415 (PLCO) Cancer Screening Trial [33]. The pleiotropic associations observed at the 9q34.2 locus where 416 eight of the ten SNPs associated with plasma protein levels were also associated with 62 other traits 417 leaves open the possibility that some of these traits, rather than the protein levels, may underlie the 418 association with EOC risk. Alternatively, some of these traits may lie up- or downstream of the 419 protein levels and mediate the association with EOC risk as part of the same causal pathway. 420 Further studies will be required to dissect these possibilities. For example, five of the eight SNPs at 421 the 9q34.2 (ABO) locus are associated with low density lipoprotein-cholesterol (LDL-C) levels with 422 the SNP alleles predicting lower LDL-C levels associating with reduced EOC risk (Table S3). This is 423 consistent with a recent analysis based on the OCAC data set which showed that lower LDL-C level 424 genetically predicted by SNPs in or near HMGCR, which encodes the enzyme inhibited by statins,

425 was associated with reduced EOC risk [34].

426

427 In conclusion, our integrative analysis of large-scale proteomic and genomic data sets 428 identified several associations between genetically predicted circulating protein levels and EOC risk 429 that were statistically significant after FDR control and biologically plausible. These plasma proteins 430 are candidate biomarkers with the potential for application in the early diagnosis of this aggressive 431 gynecological cancer. The associations shed new light on EOC biology and should inform a range of 432 follow-up laboratory-based studies and targeted biomarker validation projects wherein the 26 433 identified plasma proteins are directly tracked in incident EOC cases and controls over time. 434 435 **Author Contribution statement** 436

437 Conceptualization: PDPP, WZ, SPK. Data curation, Formal analysis, and Visualization: DPCC, GJ, XS,
438 SPK. Project administration and Resources: JMS, WZ, PDPP, OCAC. Funding acquisition: JMS, PDPP,
439 SPK. Supervision: PDPP, WZ, SPK. Writing - original draft: DPCC, GJ, XS. Writing - review & editing:
440 All authors.
441

- 442 **Conflict of Interest statement**
- 443
- 444 The authors have no conflict of interest to declare.
- 445

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452	OCAC is available in PMID: 28346442/PMCID: PMC5612337/DOI: 10.1038/ng.3826 and for									
453	INTERVAL is available in PMID: 29875488/PMCID: PMC6697541/DOI: 10.1038/s41586-018-0175-2.									
454										
455	Availability of data and code									
456										
457	The Ovarian Cancer Association Consortium (OCAC) data set can be downloaded from:									
458	ftp://ftp.ebi.ac.uk/pub/databases/gwas/summary_statistics/PhelanCM_28346442_GCST004462_									
459										
460	The genetic associations with circulating protein biomarkers were obtained from the INTERVAL study									
461	and can be downloaded from:									
462	https://static-content.springer.com/esm/art%3A10.1038%2Fs41586-018-0175-									
463	<u>2/MediaObjects/41586_2018_175_MOESM4_ESM.xlsx</u>									
464										
465	Code used to perform the analysis reported in this paper is available at:									
466	nttps://github.com/siddhartha-kar/circulating-proteins-and-ovarian-cancer									
467										
468	Other online tools used –									
469										
470	LDlink: <u>https://ldlink.nci.nih.gov</u>									
471	Forichy https://appa.pharpa.pac.m.adu/Enrichy									
472	Ennom: <u>https://amp.pharm.mssm.edu/Ennom</u>									
473	WikiPathways: <u>https://www.wikipathways.org</u>									
475 476	PhenoScapper: http://www.phenoscapper.medschl.cam.ac.uk/									
477	Thenosedinel. <u>http://www.prenosedinel.medsen.edin.de.uk/</u>									
478	References									
479										
480	[1] R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2020, CA Cancer J Clin, 70 (2020) 7–30.									
481	[2] F. Bray, J. Ferlay, I. Soeriomataram, R.L. Siegel, L.A. Torre, A. Jemal, Global cancer statistics 2018:									
482	GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA									
483	Cancer J Clin. 68 (2018) 394–424.									
484	[3] U.A. Matulonis, A.K. Sood, L. Fallowfield, B.F. Howitt, J. Sehouli, B.Y. Karlan, Ovarian cancer, Nat									
485	Rev Dis Primers. 2 (2016) 16061.									
486	[4] R.C. Bast, M. Feenev, H. Lazarus, L.M. Nadler, R.B. Colvin, R.C. Knapp, Reactivity of a monoclonal									
487	antibody with human ovarian carcinoma. J. Clin. Invest. 68 (1981) 1331–1337.									
488	[5] F.L. Moss, J. Hollingworth, T.M. Revnolds, The role of CA125 in clinical practice, J. Clin. Pathol. 58									
489	(2005) 308–312.									
490	[6] V. Dochez, H. Caillon, E. Vaucel, J. Dimet, N. Winer, G. Ducarme, Biomarkers and algorithms for									
491	diagnosis of ovarian cancer: CA125. HE4. RMI and ROMA, a review. J Ovarian Res. 12 (2019) 28.									
492	[7] US Preventive Services Task Force, D.C. Grossman, S.I. Curry, D.K. Owens, M.I. Barry, K.W.									
493	Davidson, C.A. Doubeni, J.W. Epling, A.R. Kemper, A.H. Krist, A.F. Kurth, C.S. Landefeld, C.M.									
494	Mangione, M.G. Phipps, M. Silverstein, M.A. Simon, CW. Tseng, Screening for Ovarian Cancer									
495	US Preventive Services Task Force Recommendation Statement, JAMA. 319 (2018) 588–594.									

- [8] R.G. Moore, D.S. McMeekin, A.K. Brown, P. DiSilvestro, M.C. Miller, W.J. Allard, W. Gajewski, R.
 Kurman, R.C. Bast, S.J. Skates, A novel multiple marker bioassay utilizing HE4 and CA125 for the
 prediction of ovarian cancer in patients with a pelvic mass, Gynecol Oncol. 112 (2009) 40–46.
- [9] R.G. Moore, A.K. Brown, M.C. Miller, S. Skates, W.J. Allard, T. Verch, M. Steinhoff, G. Messerlian,
 P. DiSilvestro, C.O. Granai, R.C. Bast, The use of multiple novel tumor biomarkers for the
 detection of ovarian carcinoma in patients with a pelvic mass, Gynecol Oncol. 108 (2008) 402–
 408.
- 503 [10] S. Enroth, M. Berggrund, M. Lycke, J. Broberg, M. Lundberg, E. Assarsson, M. Olovsson, K.
 504 Stålberg, K. Sundfeldt, U. Gyllensten, High throughput proteomics identifies a high-accuracy 11
 505 plasma protein biomarker signature for ovarian cancer, Commun Biol. 2 (2019) 221.
- 506 [11] S. Enroth, M. Berggrund, M. Lycke, M. Lundberg, E. Assarsson, M. Olovsson, K. Stålberg, K.
 507 Sundfeldt, U. Gyllensten, A two-step strategy for identification of plasma protein biomarkers
 508 for endometrial and ovarian cancer, Clin Proteomics. 15 (2018) 38.
- [12] B.B. Sun, J.C. Maranville, J.E. Peters, D. Stacey, J.R. Staley, J. Blackshaw, S. Burgess, T. Jiang, E.
 Paige, P. Surendran, C. Oliver-Williams, M.A. Kamat, B.P. Prins, S.K. Wilcox, E.S. Zimmerman, A.
 Chi, N. Bansal, S.L. Spain, A.M. Wood, N.W. Morrell, J.R. Bradley, N. Janjic, D.J. Roberts, W.H.
 Ouwehand, J.A. Todd, N. Soranzo, K. Suhre, D.S. Paul, C.S. Fox, R.M. Plenge, J. Danesh, H. Runz,
 A.S. Butterworth, Genomic atlas of the human plasma proteome, Nature. 558 (2018) 73–79.
- 514 [13] C.M. Phelan, K.B. Kuchenbaecker, J.P. Tyrer, S.P. Kar, K. Lawrenson, S.J. Winham, J. Dennis, A. 515 Pirie, M.J. Riggan, G. Chornokur, M.A. Earp, P.C. Lyra, J.M. Lee, S. Coetzee, J. Beesley, L. 516 McGuffog, P. Soucy, E. Dicks, A. Lee, D. Barrowdale, J. Lecarpentier, G. Leslie, C.M. Aalfs, K.K.H. 517 Aben, M. Adams, J. Adlard, I.L. Andrulis, H. Anton-Culver, N. Antonenkova, AOCS study group, 518 G. Aravantinos, N. Arnold, B.K. Arun, B. Arver, J. Azzollini, J. Balmaña, S.N. Banerjee, L. 519 Barjhoux, R.B. Barkardottir, Y. Bean, M.W. Beckmann, A. Beeghly-Fadiel, J. Benitez, M. 520 Bermisheva, M.Q. Bernardini, M.J. Birrer, L. Bjorge, A. Black, K. Blankstein, M.J. Blok, C. 521 Bodelon, N. Bogdanova, A. Bojesen, B. Bonanni, Å. Borg, A.R. Bradbury, J.D. Brenton, C. 522 Brewer, L. Brinton, P. Broberg, A. Brooks-Wilson, F. Bruinsma, J. Brunet, B. Buecher, R. Butzow, 523 S.S. Buys, T. Caldes, M.A. Caligo, I. Campbell, R. Cannioto, M.E. Carney, T. Cescon, S.B. Chan, J. 524 Chang-Claude, S. Chanock, X.Q. Chen, Y.-E. Chiew, J. Chiquette, W.K. Chung, K.B.M. Claes, T. 525 Conner, L.S. Cook, J. Cook, D.W. Cramer, J.M. Cunningham, A.A. D'Aloisio, M.B. Daly, F. 526 Damiola, S.D. Damirovna, A. Dansonka-Mieszkowska, F. Dao, R. Davidson, A. DeFazio, C. 527 Delnatte, K.F. Doheny, O. Diez, Y.C. Ding, J.A. Doherty, S.M. Domchek, C.M. Dorfling, T. Dörk, L. 528 Dossus, M. Duran, M. Dürst, B. Dworniczak, D. Eccles, T. Edwards, R. Eeles, U. Eilber, B. 529 Ejlertsen, A.B. Ekici, S. Ellis, M. Elvira, EMBRACE Study, K.H. Eng, C. Engel, D.G. Evans, P.A. 530 Fasching, S. Ferguson, S.F. Ferrer, J.M. Flanagan, Z.C. Fogarty, R.T. Fortner, F. Fostira, W.D. 531 Foulkes, G. Fountzilas, B.L. Fridley, T.M. Friebel, E. Friedman, D. Frost, P.A. Ganz, J. Garber, M.J. 532 García, V. Garcia-Barberan, A. Gehrig, GEMO Study Collaborators, A. Gentry-Maharaj, A.-M. 533 Gerdes, G.G. Giles, R. Glasspool, G. Glendon, A.K. Godwin, D.E. Goldgar, T. Goranova, M. Gore, 534 M.H. Greene, J. Gronwald, S. Gruber, E. Hahnen, C.A. Haiman, N. Håkansson, U. Hamann, 535 T.V.O. Hansen, P.A. Harrington, H.R. Harris, J. Hauke, HEBON Study, A. Hein, A. Henderson, 536 M.A.T. Hildebrandt, P. Hillemanns, S. Hodgson, C.K. Høgdall, E. Høgdall, F.B.L. Hogervorst, H. 537 Holland, M.J. Hooning, K. Hosking, R.-Y. Huang, P.J. Hulick, J. Hung, D.J. Hunter, D.G. Huntsman, 538 T. Huzarski, E.N. Imyanitov, C. Isaacs, E.S. Iversen, L. Izatt, A. Izquierdo, A. Jakubowska, P. 539 James, R. Janavicius, M. Jernetz, A. Jensen, U.B. Jensen, E.M. John, S. Johnatty, M.E. Jones, P. 540 Kannisto, B.Y. Karlan, A. Karnezis, K. Kast, KConFab Investigators, C.J. Kennedy, E. 541 Khusnutdinova, L.A. Kiemeney, J.I. Kiiski, S.-W. Kim, S.K. Kjaer, M. Köbel, R.K. Kopperud, T.A. 542 Kruse, J. Kupryjanczyk, A. Kwong, Y. Laitman, D. Lambrechts, N. Larrañaga, M.C. Larson, C. 543 Lazaro, N.D. Le, L. Le Marchand, J.W. Lee, S.B. Lele, A. Leminen, D. Leroux, J. Lester, F. Lesueur, 544 D.A. Levine, D. Liang, C. Liebrich, J. Lilyquist, L. Lipworth, J. Lissowska, K.H. Lu, J. Lubinński, C. 545 Luccarini, L. Lundvall, P.L. Mai, G. Mendoza-Fandiño, S. Manoukian, L.F.A.G. Massuger, T. May, 546 S. Mazoyer, J.N. McAlpine, V. McGuire, J.R. McLaughlin, I. McNeish, H. Meijers-Heijboer, A.

547 Meindl, U. Menon, A.R. Mensenkamp, M.A. Merritt, R.L. Milne, G. Mitchell, F. Modugno, J. 548 Moes-Sosnowska, M. Moffitt, M. Montagna, K.B. Moysich, A.M. Mulligan, J. Musinsky, K.L. 549 Nathanson, L. Nedergaard, R.B. Ness, S.L. Neuhausen, H. Nevanlinna, D. Niederacher, R.L. 550 Nussbaum, K. Odunsi, E. Olah, O.I. Olopade, H. Olsson, C. Olswold, D.M. O'Malley, K.-R. Ong, 551 N.C. Onland-Moret, OPAL study group, N. Orr, S. Orsulic, A. Osorio, D. Palli, L. Papi, T.-W. Park-552 Simon, J. Paul, C.L. Pearce, I.S. Pedersen, P.H.M. Peeters, B. Peissel, A. Peixoto, T. Pejovic, L.M. 553 Pelttari, J.B. Permuth, P. Peterlongo, L. Pezzani, G. Pfeiler, K.-A. Phillips, M. Piedmonte, M.C. 554 Pike, A.M. Piskorz, S.R. Poblete, T. Pocza, E.M. Poole, B. Poppe, M.E. Porteous, F. Prieur, D. 555 Prokofyeva, E. Pugh, M.A. Pujana, P. Pujol, P. Radice, J. Rantala, C. Rappaport-Fuerhauser, G. 556 Rennert, K. Rhiem, P. Rice, A. Richardson, M. Robson, G.C. Rodriguez, C. Rodríguez-Antona, J. 557 Romm, M.A. Rookus, M.A. Rossing, J.H. Rothstein, A. Rudolph, I.B. Runnebaum, H.B. Salvesen, 558 D.P. Sandler, M.J. Schoemaker, L. Senter, V.W. Setiawan, G. Severi, P. Sharma, T. Shelford, N. 559 Siddiqui, L.E. Side, W. Sieh, C.F. Singer, H. Sobol, H. Song, M.C. Southey, A.B. Spurdle, Z. Stadler, 560 D. Steinemann, D. Stoppa-Lyonnet, L.E. Sucheston-Campbell, G. Sukiennicki, R. Sutphen, C. 561 Sutter, A.J. Swerdlow, C.I. Szabo, L. Szafron, Y.Y. Tan, J.A. Taylor, M.-K. Tea, M.R. Teixeira, S.-H. 562 Teo, K.L. Terry, P.J. Thompson, L.C.V. Thomsen, D.L. Thull, L. Tihomirova, A.V. Tinker, M. 563 Tischkowitz, S. Tognazzo, A.E. Toland, A. Tone, B. Trabert, R.C. Travis, A. Trichopoulou, N. Tung, 564 S.S. Tworoger, A.M. van Altena, D. Van Den Berg, A.H. van der Hout, R.B. van der Luijt, M. Van 565 Heetvelde, E. Van Nieuwenhuysen, E.J. van Rensburg, A. Vanderstichele, R. Varon-Mateeva, A. 566 Vega, D.V. Edwards, I. Vergote, R.A. Vierkant, J. Vijai, A. Vratimos, L. Walker, C. Walsh, D. 567 Wand, S. Wang-Gohrke, B. Wappenschmidt, P.M. Webb, C.R. Weinberg, J.N. Weitzel, N. 568 Wentzensen, A.S. Whittemore, J.T. Wijnen, L.R. Wilkens, A. Wolk, M. Woo, X. Wu, A.H. Wu, H. 569 Yang, D. Yannoukakos, A. Ziogas, K.K. Zorn, S.A. Narod, D.F. Easton, C.I. Amos, J.M. Schildkraut, 570 S.J. Ramus, L. Ottini, M.T. Goodman, S.K. Park, L.E. Kelemen, H.A. Risch, M. Thomassen, K. Offit, 571 J. Simard, R.K. Schmutzler, D. Hazelett, A.N. Monteiro, F.J. Couch, A. Berchuck, G. Chenevix-572 Trench, E.L. Goode, T.A. Sellers, S.A. Gayther, A.C. Antoniou, P.D.P. Pharoah, Identification of 573 12 new susceptibility loci for different histotypes of epithelial ovarian cancer, Nat. Genet. 49 574 (2017) 680-691.

- 575 [14] E. Di Angelantonio, S.G. Thompson, S. Kaptoge, C. Moore, M. Walker, J. Armitage, W.H.
 576 Ouwehand, D.J. Roberts, J. Danesh, INTERVAL Trial Group, Efficiency and safety of varying the
 577 frequency of whole blood donation (INTERVAL): a randomised trial of 45 000 donors, Lancet.
 578 390 (2017) 2360–2371.
- [15] J.C. Rohloff, A.D. Gelinas, T.C. Jarvis, U.A. Ochsner, D.J. Schneider, L. Gold, N. Janjic, Nucleic Acid
 Ligands With Protein-like Side Chains: Modified Aptamers and Their Use as Diagnostic and
 Therapeutic Agents, Mol Ther Nucleic Acids. 3 (2014) e201.
- 582 [16] M.J. Machiela, S.J. Chanock, LDlink: a web-based application for exploring population-specific
 583 haplotype structure and linking correlated alleles of possible functional variants,
 584 Bioinformatics. 31 (2015) 3555–3557.
- [17] M.V. Kuleshov, M.R. Jones, A.D. Rouillard, N.F. Fernandez, Q. Duan, Z. Wang, S. Koplev, S.L.
 Jenkins, K.M. Jagodnik, A. Lachmann, M.G. McDermott, C.D. Monteiro, G.W. Gundersen, A.
 Ma'ayan, Enrichr: a comprehensive gene set enrichment analysis web server 2016 update,
 Nucleic Acids Res. 44 (2016) W90-97.
- [18] D.N. Slenter, M. Kutmon, K. Hanspers, A. Riutta, J. Windsor, N. Nunes, J. Mélius, E. Cirillo, S.L.
 Coort, D. Digles, F. Ehrhart, P. Giesbertz, M. Kalafati, M. Martens, R. Miller, K. Nishida, L.
 Rieswijk, A. Waagmeester, L.M.T. Eijssen, C.T. Evelo, A.R. Pico, E.L. Willighagen, WikiPathways:
 a multifaceted pathway database bridging metabolomics to other omics research, Nucleic
 Acids Res. 46 (2018) D661–D667.

[19] M.A. Kamat, J.A. Blackshaw, R. Young, P. Surendran, S. Burgess, J. Danesh, A.S. Butterworth, J.R. Staley, PhenoScanner V2: an expanded tool for searching human genotype-phenotype associations, Bioinformatics. 35 (2019) 4851–4853.

597 [20] K.B. Kuchenbaecker, S.J. Ramus, J. Tyrer, A. Lee, H.C. Shen, J. Beesley, K. Lawrenson, L. 598 McGuffog, S. Healey, J.M. Lee, T.J. Spindler, Y.G. Lin, T. Pejovic, Y. Bean, Q. Li, S. Coetzee, D. 599 Hazelett, A. Miron, M. Southey, M.B. Terry, D.E. Goldgar, S.S. Buys, R. Janavicius, C.M. Dorfling, 600 E.J. van Rensburg, S.L. Neuhausen, Y.C. Ding, T.V.O. Hansen, L. Jønson, A.-M. Gerdes, B. 601 Ejlertsen, D. Barrowdale, J. Dennis, J. Benitez, A. Osorio, M.J. Garcia, I. Komenaka, J.N. Weitzel, 602 P. Ganschow, P. Peterlongo, L. Bernard, A. Viel, B. Bonanni, B. Peissel, S. Manoukian, P. Radice, 603 L. Papi, L. Ottini, F. Fostira, I. Konstantopoulou, J. Garber, D. Frost, J. Perkins, R. Platte, S. Ellis, 604 EMBRACE, A.K. Godwin, R.K. Schmutzler, A. Meindl, C. Engel, C. Sutter, O.M. Sinilnikova, GEMO 605 Study Collaborators, F. Damiola, S. Mazoyer, D. Stoppa-Lyonnet, K. Claes, K. De Leeneer, J. Kirk, 606 G.C. Rodriguez, M. Piedmonte, D.M. O'Malley, M. de la Hoya, T. Caldes, K. Aittomäki, H. 607 Nevanlinna, J.M. Collée, M.A. Rookus, J.C. Oosterwijk, Breast Cancer Family Registry, L. 608 Tihomirova, N. Tung, U. Hamann, C. Isaccs, M. Tischkowitz, E.N. Imyanitov, M.A. Caligo, I.G. 609 Campbell, F.B.L. Hogervorst, HEBON, E. Olah, O. Diez, I. Blanco, J. Brunet, C. Lazaro, M.A. 610 Pujana, A. Jakubowska, J. Gronwald, J. Lubinski, G. Sukiennicki, R.B. Barkardottir, M. Plante, J. 611 Simard, P. Soucy, M. Montagna, S. Tognazzo, M.R. Teixeira, KConFab Investigators, V.S. 612 Pankratz, X. Wang, N. Lindor, C.I. Szabo, N. Kauff, J. Vijai, C.A. Aghajanian, G. Pfeiler, A. Berger, 613 C.F. Singer, M.-K. Tea, C.M. Phelan, M.H. Greene, P.L. Mai, G. Rennert, A.M. Mulligan, S. 614 Tchatchou, I.L. Andrulis, G. Glendon, A.E. Toland, U.B. Jensen, T.A. Kruse, M. Thomassen, A. 615 Bojesen, J. Zidan, E. Friedman, Y. Laitman, M. Soller, A. Liljegren, B. Arver, Z. Einbeigi, M. 616 Stenmark-Askmalm, O.I. Olopade, R.L. Nussbaum, T.R. Rebbeck, K.L. Nathanson, S.M. 617 Domchek, K.H. Lu, B.Y. Karlan, C. Walsh, J. Lester, Australian Cancer Study (Ovarian Cancer 618 Investigators), Australian Ovarian Cancer Study Group, A. Hein, A.B. Ekici, M.W. Beckmann, 619 P.A. Fasching, D. Lambrechts, E. Van Nieuwenhuysen, I. Vergote, S. Lambrechts, E. Dicks, J.A. 620 Doherty, K.G. Wicklund, M.A. Rossing, A. Rudolph, J. Chang-Claude, S. Wang-Gohrke, U. Eilber, 621 K.B. Moysich, K. Odunsi, L. Sucheston, S. Lele, L.R. Wilkens, M.T. Goodman, P.J. Thompson, Y.B. 622 Shvetsov, I.B. Runnebaum, M. Dürst, P. Hillemanns, T. Dörk, N. Antonenkova, N. Bogdanova, A. 623 Leminen, L.M. Pelttari, R. Butzow, F. Modugno, J.L. Kelley, R.P. Edwards, R.B. Ness, A. du Bois, 624 F. Heitz, I. Schwaab, P. Harter, K. Matsuo, S. Hosono, S. Orsulic, A. Jensen, S.K. Kjaer, E. Hogdall, 625 H.N. Hasmad, M.A.N. Azmi, S.-H. Teo, Y.-L. Woo, B.L. Fridley, E.L. Goode, J.M. Cunningham, R.A. 626 Vierkant, F. Bruinsma, G.G. Giles, D. Liang, M.A.T. Hildebrandt, X. Wu, D.A. Levine, M. Bisogna, 627 A. Berchuck, E.S. Iversen, J.M. Schildkraut, P. Concannon, R.P. Weber, D.W. Cramer, K.L. Terry, 628 E.M. Poole, S.S. Tworoger, E.V. Bandera, I. Orlow, S.H. Olson, C. Krakstad, H.B. Salvesen, I.L. 629 Tangen, L. Bjorge, A.M. van Altena, K.K.H. Aben, L.A. Kiemeney, L.F.A.G. Massuger, M. Kellar, A. 630 Brooks-Wilson, L.E. Kelemen, L.S. Cook, N.D. Le, C. Cybulski, H. Yang, J. Lissowska, L.A. Brinton, 631 N. Wentzensen, C. Hogdall, L. Lundvall, L. Nedergaard, H. Baker, H. Song, D. Eccles, I. McNeish, 632 J. Paul, K. Carty, N. Siddiqui, R. Glasspool, A.S. Whittemore, J.H. Rothstein, V. McGuire, W. Sieh, 633 B.-T. Ji, W. Zheng, X.-O. Shu, Y.-T. Gao, B. Rosen, H.A. Risch, J.R. McLaughlin, S.A. Narod, A.N. 634 Monteiro, A. Chen, H.-Y. Lin, J. Permuth-Wey, T.A. Sellers, Y.-Y. Tsai, Z. Chen, A. Ziogas, H. 635 Anton-Culver, A. Gentry-Maharaj, U. Menon, P. Harrington, A.W. Lee, A.H. Wu, C.L. Pearce, G. 636 Coetzee, M.C. Pike, A. Dansonka-Mieszkowska, A. Timorek, I.K. Rzepecka, J. Kupryjanczyk, M. 637 Freedman, H. Noushmehr, D.F. Easton, K. Offit, F.J. Couch, S. Gayther, P.P. Pharoah, A.C. 638 Antoniou, G. Chenevix-Trench, Consortium of Investigators of Modifiers of BRCA1 and BRCA2, 639 Identification of six new susceptibility loci for invasive epithelial ovarian cancer, Nat. Genet. 47 640 (2015) 164-171. 641 [21] X. Wang, N.R. Tucker, G. Rizki, R. Mills, P.H. Krijger, E. de Wit, V. Subramanian, E. Bartell, X.-X. 642 Nguyen, J. Ye, J. Leyton-Mange, E.V. Dolmatova, P. van der Harst, W. de Laat, P.T. Ellinor, C. 643 Newton-Cheh, D.J. Milan, M. Kellis, L.A. Boyer, Discovery and validation of sub-threshold

644 genome-wide association study loci using epigenomic signatures, Elife. 5 (2016).

645 [22] C. Porta, C. Paglino, A. Mosca, Targeting PI3K/Akt/mTOR Signaling in Cancer, Front Oncol. 4
646 (2014) 64.

- [23] M.K. Ediriweera, K.H. Tennekoon, S.R. Samarakoon, Role of the PI3K/AKT/mTOR signaling
 pathway in ovarian cancer: Biological and therapeutic significance, Semin. Cancer Biol. 59
 (2019) 147–160.
- [24] C.-C. Huang, S.-H. Cheng, C.-H. Wu, W.-Y. Li, J.-S. Wang, M.-L. Kung, T.-H. Chu, S.-T. Huang, C.-T.
 Feng, S.-C. Huang, M.-H. Tai, Delta-like 1 homologue promotes tumorigenesis and epithelialmesenchymal transition of ovarian high-grade serous carcinoma through activation of Notch
 signaling, Oncogene. 38 (2019) 3201–3215.
- [25] L.-W. Yao, L.-L. Wu, L.-H. Zhang, W. Zhou, L. Wu, K. He, J.-C. Ren, Y.-C. Deng, D.-M. Yang, J.
 Wang, G.-G. Mu, M. Xu, J. Zhou, G.-A. Xiang, Q.-S. Ding, Y.-N. Yang, H.-G. Yu, MFAP2 is
 overexpressed in gastric cancer and promotes motility via the MFAP2/integrin α5β1/FAK/ERK
 pathway, Oncogenesis. 9 (2020) 17. https://doi.org/10.1038/s41389-020-0198-z.
- [26] X. Gong, T. Dong, M. Niu, X. Liang, S. Sun, Y. Zhang, Y. Li, D. Li, IncRNA LCPAT1 Upregulation
 Promotes Breast Cancer Progression via Enhancing MFAP2 Transcription, Mol Ther Nucleic
 Acids. 21 (2020) 804–813.
- [27] M. Wu, Y. Ding, X. Jiang, Y. Chen, N. Wu, L. Li, H. Wang, Y. Huang, N. Xu, L. Teng, Overexpressed
 MAGP1 Is Associated With a Poor Prognosis and Promotes Cell Migration and Invasion in
 Gastric Cancer, Front Oncol. 9 (2019) 1544.
- 664 [28] H. Arakawa, Netrin-1 and its receptors in tumorigenesis, Nat. Rev. Cancer. 4 (2004) 978–987.
- [29] W. Hao, M. Yu, J. Lin, B. Liu, H. Xing, J. Yang, D. Sun, F. Chen, M. Jiang, C. Tang, X. Zhang, Y. Zhao,
 Y. Zhu, The pan-cancer landscape of netrin family reveals potential oncogenic biomarkers, Sci
 Rep. 10 (2020) 5224.
- [30] A.D. Papanastasiou, G. Pampalakis, D. Katsaros, G. Sotiropoulou, Netrin-1 overexpression is
 predictive of ovarian malignancies, Oncotarget. 2 (2011) 363–367.
- [31] A. Manichaikul, L.C. Peres, X.-Q. Wang, M.E. Barnard, D. Chyn, X. Sheng, Z. Du, J. Tyrer, J. Dennis,
 A.G. Schwartz, M.L. Cote, E. Peters, P.G. Moorman, M. Bondy, J.S. Barnholtz-Sloan, P. Terry,
 A.J. Alberg, E.V. Bandera, E. Funkhouser, A.H. Wu, C.L. Pearce, M. Pike, V.W. Setiawan, C.A.
 Haiman, African American Breast Cancer Consortium (AABC), African Ancestry Prostate Cancer
 Consortium (AAPC), J.R. Palmer, L. LeMarchand, L.R. Wilkens, A. Berchuck, J.A. Doherty, F.
 Modugno, R. Ness, K. Moysich, B.Y. Karlan, A.S. Whittemore, V. McGuire, W. Sieh, K.
- Lawrenson, S. Gayther, T.A. Sellers, P. Pharoah, J.M. Schildkraut, African American Cancer
 Epidemiology Study (AACES) and the Ovarian Cancer Association Consortium (OCAC),
- 678 Identification of novel epithelial ovarian cancer loci in women of African ancestry, Int J Cancer.
 679 146 (2020) 2987–2998.
- 680 [32] K. Lawrenson, F. Song, D.J. Hazelett, S.P. Kar, J. Tyrer, C.M. Phelan, R.I. Corona, N.I. Rodríguez-681 Malavé, J.-H. Seo, E. Adler, S.G. Coetzee, F. Segato, M.A.S. Fonseca, C.I. Amos, M.E. Carney, G. 682 Chenevix-Trench, J. Choi, J.A. Doherty, W. Jia, G.J. Jin, B.-G. Kim, N.D. Le, J. Lee, L. Li, B.K. Lim, 683 N.A. Adenan, M. Mizuno, B. Park, C.L. Pearce, K. Shan, Y. Shi, X.-O. Shu, W. Sieh, Australian 684 Ovarian Cancer Study Group, P.J. Thompson, L.R. Wilkens, Q. Wei, Y.L. Woo, L. Yan, B.Y. Karlan, 685 M.L. Freedman, H. Noushmehr, E.L. Goode, A. Berchuck, T.A. Sellers, S.-H. Teo, W. Zheng, K. 686 Matsuo, S. Park, K. Chen, P.D.P. Pharoah, S.A. Gayther, M.T. Goodman, Genome-wide 687 association studies identify susceptibility loci for epithelial ovarian cancer in east Asian women, 688 Gynecol. Oncol. 153 (2019) 343-355.
- [33] C.S. Zhu, W.-Y. Huang, P.F. Pinsky, C.D. Berg, M. Sherman, K.J. Yu, D.M. Carrick, A. Black, R.
 Hoover, P. Lenz, C. Williams, L. Hawkins, M. Chaloux, S. Yurgalevitch, S. Mathew, A. Miller, V.
 Olivo, A. Khan, S.M. Pretzel, D. Multerer, P. Beckmann, K.G. Broski, N.D. Freedman, The
 Prostate, Lung, Colorectal and Ovarian Cancer (PLCO) Screening Trial Pathology Tissue
 Resource, Cancer Epidemiol. Biomarkers Prev. 25 (2016) 1635–1642.
- [34] J. Yarmolinsky, C.J. Bull, E.E. Vincent, J. Robinson, A. Walther, G.D. Smith, S.J. Lewis, C.L. Relton,
 R.M. Martin, Association Between Genetically Proxied Inhibition of HMG-CoA Reductase and
 Epithelial Ovarian Cancer, JAMA. 323 (2020) 646–655.
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- 698 Table Legends

Table 1. Associations identified between genetically predicted circulating (plasma) protein levels and
 all invasive epithelial ovarian cancer risk.

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703 **Table 2.** Pathways enriched among the genes encoding the 26 all invasive epithelial ovarian cancer
704 risk-associated circulating protein biomarkers identified.

706 Figure Legends

Fig. 1. Regional genetic association plots. Genetic association with all invasive epithelial ovarian cancer risk (negative logarithm base 10 *P*-value) from the Ovarian Cancer Association Consortium study is plotted on the Y-axis and chromosomal position (build 37/hg 19) is plotted on the X-axis. SNPs are marked with blue dots or colored diamonds. SNPs marked with colored diamonds are the SNPs most strongly associated in the INTERVAL study with circulating (plasma) levels of the proteins named in the titles of the plots.

Tables

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Table 1. Associations ide	ntified between genetically prec	licted circulating (pla	sma) prote	ein levels and all li	ivasive epitne	lial ovarian cance	r fisk.					
Protein	Protein full name	Top SNP associated with plasma level of protein	Chr	Posª	Cis /trans ^b	Gene encoding protein	Gene mapped to SNP	OR	L95 %CL	U95 %CL	Р	FDR
FA20B	Glycosaminoglycan xylosylkinase	rs587729126	9	136138765	trans	FAM20B	ABO	1.47	1.30	1.66	8.1x10 ⁻¹⁰	4.5x10 ⁻⁷
sICAM-2	Intercellular adhesion molecule 2	rs587729126	9	136138765	trans	ICAM2	ABO	0.65	0.57	0.75	8.1x10 ⁻¹⁰	4.5x10 ⁻⁷
VEGF sR2	Vascular endothelial growth factor receptor 2	rs635634	9	136155000	trans	KDR	ABO	0.84	0.80	0.89	3.0x10 ⁻⁹	4.5x10 ⁻⁷
ISLR2	Immunoglobulin superfamily containing leucine-rich repeat protein 2	rs115478735	9	136149711	trans	ISLR2	ABO	0.80	0.75	0.86	2.8x10 ⁻⁹	4.5x10 ⁻⁷
Met	Hepatocyte growth factor receptor	rs635634	9	136155000	trans	MET	ABO	0.77	0.71	0.84	3.0x10 ⁻⁹	4.5x10 ⁻⁷
TPST2	Protein-tyrosine sulfotransferase 2	rs115478735	9	136149711	trans	TPST2	ABO	1.36	1.23	1.51	2.8x10 ⁻⁹	4.5x10 ⁻⁷
LIF sR	Leukemia inhibitory factor receptor	rs635634	9	136155000	trans	LIFR	ABO	0.72	0.64	0.80	3.0x10 ⁻⁹	4.5x10 ⁻⁷
Endoglin	Endoglin	rs635634	9	136155000	trans	ENG	ABO	0.66	0.57	0.76	3.0x10 ⁻⁹	4.5x10 ⁻⁷
IGF-I sR	Insulin-like growth factor 1 receptor	rs635634	9	136155000	trans	IGF1R	ABO	0.64	0.55	0.74	3.0x10 ⁻⁹	4.5x10 ⁻⁷
sE-Selectin	E-selectin	rs2519093	9	136141870	trans	SELE	ABO	0.92	0.89	0.95	4.2x10 ^{.9}	4.7x10 ⁻⁷
IL-3 Ra	Interleukin-3 receptor subunit alpha	rs2519093	9	136141870	trans	IL3RA	ABO	0.89	0.86	0.93	4.2x10 ⁻⁹	4.7x10 ⁻⁷
C1GLC	C1GALT1-specific chaperone 1	rs2519093	9	136141870	trans	C1GALT1C1	ABO	1.17	1.11	1.24	4.2x10 ⁻⁹	4.7x10 ⁻⁷
IR	Insulin receptor	rs507666	9	136149399	trans	INSR	ABO	0.85	0.80	0.90	8.4x10 ⁻⁹	8.6x10 ⁻⁷
QSOX2	Sulfhydryl oxidase 2	rs149092047	9	136139907	trans	QSOX2	ABO	1.09	1.06	1.13	1.2x10 ⁻⁷	1.0x10 ⁻⁵
FAM3D	Protein FAM3D	rs149092047	9	136139907	trans	FAM3D	ABO	1.12	1.07	1.16	1.2x10 ⁻⁷	1.0x10 ⁻⁵
GOLM1	Golgi membrane protein 1	rs149092047	9	136139907	trans	GOLM1	ABO	1.15	1.09	1.22	1.2x10 ⁻⁷	1.0x10 ⁻⁵
Desmoglein-2	Desmoglein-2	rs687621	9	136137065	trans	DSG2	ABO	1.39	1.23	1.57	1.7x10 ⁻⁷	1.3x10 ⁻⁵
ST4S6	Carbohydrate sulfotransferase 15	rs550057	9	136146597	trans	CHST15	ABO	0.79	0.72	0.86	2.0x10 ⁻⁷	1.4x10 ⁻⁵
Alkaline phosphatase, intestine	Intestinal-type alkaline phosphatase	rs550057	9	136146597	trans	ALPI	ABO	0.74	0.66	0.83	2.0x10 ⁻⁷	1.4x10 ⁻⁵
Coagulation Factor VIII	Coagulation Factor VIII	rs9411377	9	136145404	trans	F8	ABO	1.16	1.09	1.22	5.5x10 ⁻⁷	3.7x10 ⁻⁵
BGAT	Histo-blood group ABO system transferase	rs505922	9	136149229	cis	ABO	ABO	1.05	1.03	1.08	6.7x10 ⁻⁷	4.0x10 ⁻⁵
DC-SIGN	CD209 antigen	rs505922	9	136149229	trans	CD209	ABO	1.09	1.05	1.12	6.7x10 ⁻⁷	4.0x10 ⁻⁵
MFAP2	Microfibrillar-associated protein 2	rs4920605	1	17315425	cis	MFAP2	MFAP2	1.27	1.12	1.45	1.8x10 ⁻⁴	0.011
SEMG2	Semenogelin-2	rs12881760	14	101176335	trans	SEMG2	DLK1	1.10	1.04	1.15	4.0x10 ⁻⁴	0.021
DLK1	Protein delta homolog 1	rs12881760	14	101176335	cis	DLK1	DLK1	1.10	1.04	1.16	4.0x10 ⁻⁴	0.021
NTNG1	Netrin-G1	rs115668827	1	107678268	cis	NTNG1	NTNG1	0.89	0.84	0.95	4.9x10 ⁻⁴	0.025
a Build 37/h19 position. b Cis if the top SNP is < 1 Mb from the gene encoding protein and trans if the top SNP is > 1 Mb from the gene encoding the protein. c Odds ratio (OR), lower 95% confidence limit (U95%CL), upper 95% confidence limit (U95%CL), P-value, and false discovery rate (FDR) from the current study. OR is in terms of all invasive EOC risk per standard deviation increase in circulating (plasma) protein level.												

Table 2. Pathways enriched among the genes encoding the 26 all invasive epithelial ovarian cancer risk-associated circulating protein biomarkers identified.

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Pathway ^a	Overlap ^b	P-value	Adjusted P ^c	Genes ^d
Pathways Regulating Hippo Signaling WP4540	4/98	7.5 x 10⁻ ⁶	0.004	INSR; KDR; MET; IGF1R
Hippo-Merlin Signaling Dysregulation WP4541	4/120	1.7 x 10 ⁻⁵	0.004	INSR; KDR; MET; IGF1R
Focal Adhesion-PI3K-Akt-mTOR-signaling pathway WP3932	5/303	3.9 x 10 ⁻⁵	0.006	INSR; IL3RA; KDR; MET; IGF1R
PI3K-Akt Signaling Pathway WP4172	5/340	6.8 x 10 ⁻⁵	0.008	INSR; IL3RA; KDR; MET; IGF1R
Ras Signaling WP4223	4/184	8.9 x 10 ⁻⁵	0.008	INSR; KDR; MET; IGF1R
Ebola Virus Pathway on Host WP4217	3/129	6.1 x 10 ⁻⁴	0.041	CD209; ICAM2; IGF1R
Canonical and Non-canonical Notch signaling WP3845	2/27	5.6 x 10 ⁻⁴	0.044	MFAP2; DLK1

^a From the "WikiPathways 2019 Human" pathway database (with associated WP identifier number). ^b The number of genes out of the 26 genes evaluated/the total number of genes annotated to the pathway.

^c Adjusted for testing 472 pathways.
 ^d The genes (out of the 26 genes evaluated) that are annotated to the pathway.

