Chapter 10

MECHANISMS AND BIOMARKERS OF EXERCISE-INDUCED BRONCHOCONSTRICTION

AUTHORS AND DEGREES:

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Dr Anderson is the inventor of the mannitol test used for the diagnosis of bronchial hyperresponsiveness and receives a share of the royalties from the sale of AridolTM and OsmohaleTM paid to Royal Prince Alfred Hospital by Pharmaxis Ltd.

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KEY WORDS

Hyperpnea; water loss; osmolarity; epithelium; mast cells; eosinophils; eicosanoids; sensory nerves

KEY POINTS

- The conditioning of inhaled air during exercise-hyperpnea initiates osmotic
 and vascular events that lead, in susceptible individuals, to airway narrowing.
- A loss of physical barrier intergrity and impairment in signaling and secretory functions of the airway epithelium increases the susceptibility to EIB.
- Airway smooth muscle contraction and mucin release in individuals with EIB
 are mediated predominantly by release of inflammatory mediators with
 associated activation of neural pathways.
- ullet Cysteinyl leukotrienes and prostagland in D_2 are the primary inflammatory mediators released into the airways from mast cells and eosinophils during EIB.

SYNOPSIS

Exercise is a common trigger of bronchoconstriction. In recent years, there has been increased understanding of the pathophysiology of exercise-induced bronchoconstriction (EIB). While evaporative water loss and thermal changes have been recognized stimuli for EIB for almost 50 years, accumulating evidence points toward a pivotal role for the airway epithelium in orchestrating the inflammatory response linked to EIB. Overproduction of inflammatory mediators, relative underproduction of protective lipid mediators, and infiltration of the airways with eosinophils and mast cells are all established contributors to EIB. Sensory nerve activation and regional airway closure maybe important in EIB, but further research is warranted.

(100 words)

INTRODUCTION

The underlying basis for exercise-induced bronchoconstriction (EIB) is becoming increasingly understood. Initial work starting in the 1970s revealed the major determinants of EIB in susceptible individuals. The aim of this review is: to examine the respective roles of evaporative water loss and thermal changes as stimuli to EIB; to provide evidence that loss of physical barrier functions of the epithelium during exercise-induced hyperpnea is associated with the development of bronchoconstriction in susceptible individuals; to discuss the central role of leukocyte activation and the associated generation of lipid mediators and release of neuropetides that sustain bronchoconstriction during EIB; and to consider the role that regional airway closure may play in the development of EIB.

CONDITIONING THE AIR INSPIRED DURING EXERCISE

Under most conditions of exercise the air inspired needs to be heated and humidified to body conditions [37°C, 100% relative humidity (RH), or 44 mg H₂O/L] before it enters the alveoli. As a result, heat and water are lost from the airway surface during inspiration. The number of generations involved in conditioning depends on the level of ventilation reached and sustained during exercise, and the temperature and water content of the inspired air.(1)

Heat loss as a stimulus to airway narrowing

Cooling of the airways, from heat lost through vaporisation of water and from heating the inspired air, was initially identified as a potential stimulus for EIB.(2) The proposal was subsequently extended to include a rewarming of the airways after exercise.(3) (Figure 1) This hypothesis suggested that cooling initiated

vasoconstriction during exercise, followed by a reactive (or rebound) hyperaemia at the end of exercise.(3) These vascular events are most significant when air of sub zero temperature is inspired during intense exercise of 4 minutes or more (4) and when the smaller airways are recruited into the conditioning process, but are unlikely to occur in temperate or hot environments. These vascular events may be relevant to anyone performing vigorous exercise in cold conditions – although people with asthma may have a more rapid and exaggerated vascular response (5) –, and may serve to amplify airway narrowing in individuals with asthma with EIB.(6)

Water loss as a stimulus to airway narrowing

It is now well recognised that cooling and rewarming of the airways are not prerequisites for EIB. In individuals with asthma, EIB occurs when hot dry air is inhaled during exercise (7-9) and when the airways are not cooled below their temperature at rest.(10)

The concentration of water in the inspired air is an important determinant of EIB (11), with the rate of water loss during exercise relating directly to the severity of the airway response in an individual.(12) Importantly, the stimulus to EIB acts at the surface of the airways, because EIB is prevented simply by inhaling fully humidified air at body temperature.(11, 13) A loss of water by evaporation, in humidifying large volumes of air in a short time, is thought to cause a transient increase in concentration of ions (e.g., Na⁺, CI, Ca⁺⁺) in the airway surface liquid (ASL).(14) (Figure 1) This osmotic stimulus occurs after intense exercise when dry air of any temperature (cold or hot) is inspired, providing the duration of exercise is sufficient.

Because the surface area of the first 10 generations of airways is 742 cm² and contains less than 0.7 ml of ASL (15, 16), only a small loss of water would increase osmolarity above the normal value of ~320-340 mosm. A mathematical model, based on *in vivo* observations (17), predicted that breathing air of 26°C and 35% RH at 60 L/min would result in a cumulative loss of 0.44 mL *per* minute from the first 12 generations of airways.(18, 19) It is likely that this rate of water loss exceeds the rate of replacement across the epithelium, leading to a transient increase in osmolarity of the ASL.

Although methodological limitations render the accurate measurement of changes in ASL osmolarity difficult (20), there is direct evidence for evaporative water loss from the airways involved in conditioning the air. Mucociliary clearance rate was shown to be reduced during hyperpnea with dry air (but not humid air).(21) This reduction, however, occurred more markedly in people with asthma compared with healthy individuals, suggesting that the rate of water movement across the epithelium towards the lumen is slower in populations prone to EIB.(21)

An increase in ion concentration of the ASL during exercise would act as an osmotic stimulus for transport of water to the airway surface from the epithelial cells and other cells on, or near the airway surface. Because the basolateral membrane is less permeable to water than the apical membrane, it was proposed that epithelial cells, as well as other cells, would have a reduction in volume. (22) It is the regulatory volume increase following cell volume decrease that is thought to provide the signal for mediator release. The source of water to replace the epithelial cell volume is the submucosa and small changes in its osmolarity could be the signal to the increase in

bronchial blood flow that occurs when breathing dry air.(23-25) The osmotic stimulus to EIB is further supported by evidence that airway narrowing can be provoked in asthmatics by inhalation of hyperosmolar saline.(26) Further, the airway response to hypertonic saline is related to mast cell number (assessed by brush biopsy).(27) Mediators released in response to osmotic challenge with dry powder mannitol (28) are the same as exercise (29) and blocked by the same drugs.(30, 31)

The transient increase in osmolarity of ASL during exercise creates an environment known to be favourable for release of mediators from cells in, or near the airway surface (e.g., epithelial cells, sensory nerves, mast cells and eosinophils). While some mediators [prostaglandin E₂ (PGE₂)] may induce bronchodilation during exercise, other mediators [neurokinins, PGD₂, cysteinyl leukotrienes (cyst-LTs)] provoke airway narrowing by acting, either directly on bronchial smooth muscle or, *via* sensory nerves. (Figure 2) (29, 31)

Temperature as a modulator of time course and severity of EIB

The severity of EIB is similar in the same individual over a wide range of inspired air temperatures, from cool (9°C) to hot (36°C), (Figure 3) providing that the inspired water content is low and remains the same. (7-9) This is consistent with the relatively constant value measured for water content of expired air (29-33 mg/L), when breathing air at 22-40°C that contains <13 mg H₂O/L.(24) The maximum airway response is thought to occur when generations 10 to 12 are recruited, and this may take 8 minutes when the inspired air is warm. It is here, in the non-cartilaginous airways, that the density of mast cells is high in individuals with asthma.(32) This may explain why warm dry air can still cause a similar fall in expiratory flow to cool

air.(8) The cooler the inspired air, the faster the required generations will be recruited, and the earlier the response will occur.

EPITHELIAL DYSFUNCTION IN EIB

The airway epithelium is the first structural barrier to the inhaled environment in the airway mucosa. Under normal circumstances, the epithelium forms a highly regulated and tight barrier that limits penetration of inhaled allergens, pathogens, pollutants and other toxic compounds into the internal lung environment. Exercise hyperpnea can cause acute disruption of the airway epithelium (33-35). Shedding of epithelial cells into the airway lumen after exercise challenge has been shown in individuals with asthma with EIB (36). A (partial) loss of physical integrity of the epithelial barrier is likely to disrupt airway homeostasis at a number of levels. Disruption of ASL balance (resulting in malfunction of the mucociliary escalator), plasma exudation (resulting in edema), and attraction and activation of inflammatory cells in response to injury can all compromise airway homeostasis. Instrinsic dysfunction of the airway epithelium – as found in individuals with asthma (37) – and/or extreme levels of ventilation – as developed by endurance athletes (38) – are likely to increase the susceptibility to /extent of airway epithelial injury and, thereby, explain the high prevalence of EIB reported in both these populations (39, 40).

Breaking through the epithelial barrier

As minute ventilation increases during exercise, so does the load of foreign substances entering the airways. Disruption of epithelial tight junctions and/or gaps into the epithelium facilitates the penetration of foreign substances, enhancing the potential for toxic, immune, and inflammatory responses, with ensuing

bronchoconstriction in susceptible individuals. During the pollen season, the severity of airway narrowing after outdoor exercise is increased in individuals with asthma with birch pollen allergy (41). The cellular infiltration and activation of mast cells and T cells (and possibly eosinophils) observed during natural allergen exposure in atopic individuals with asthma (42) would render the airway smooth muscle more responsive to the stress of exercise. Further, common air pollutants and irritants (such as ozone or chlorine derivatives) have the potential *per se* to acutely disrupt the lung epithelial barrier (43, 44). Inhalation of noxious airborne agents during exercise hyperpnea could therefore aggravate the damage to the epithelium, and increase the risk/severity of EIB. In support of this concept, a high prevalence of EIB has consistently been reported in athletes who train and compete in polluted environments. (1)

Disturbance of ASL

Apart from being a barrier, the airway epithelium plays other roles, including maintenance of ASL levels (45). Intact epithelial cells sense and autoregulate ASL height by active ion transport (46). A damaged airway epithelium would loose its ability to tightly regulate water movements, potentiating the osmotic effects of exercise hyperpnea and, potentially increasing the severity of EIB. In individuals with asthma with EIB, the extent of airway injury (as assessed by the percentage of columnar epithelial cells in sputum) at baseline has been shown to correlate with the severity of the airway narrowing after exercise (29).

Mucus dysfunction

Abnormal water movement, as a consequence of airway epithelial injury, also leads to alterations in rheological properties of mucus, with subsequent impairment of

mucociliary clearance and plug formation. A thicker and more elastic mucus is harder to clear from the airways and provides an environment for microbial growth, leading to infection and inflammation (47). In children with asthma (a population prone to EIB), viral infections of the airways are frequent and commonly associated with disease exacerbations (48). Further, in elite athletes with asthma and/or EIB, an increase respiratory infection susceptibility has recently been reported (49).

Mucus plugs could also directly contribute to lumenal obstruction in EIB. In individuals with asthma with EIB, exercise was found to initiate the release of MUC5AC (i.e., the predominant gel-forming mucin of goblet cells) (50). Moreover, an hyper-activity of the mucus-secreting goblets has been reported in both, individuals with asthma with EIB (50) and elite athletes (51). Therefore, alterations in mucosal rheology secondary to epithelial dysfunction are likely to contribute to airflow obstruction and post-exercise symptoms (such as cough and mucus hypersecretion).

Epithelial damage-restitution

Restitution of the epithelium upon injury is associated with many tissue responses including, amongst others, plasma exudation (52). Plasma exudate in the airway lumen could not only contribute to EIB through mucosal/submucosal edema, but may also exacerbate some of the events described previoulsy (i.e., shedding of epithelium, impairment of mucociliary transport and mucus plug formation) (53). Further, plasma exudate may cause airway inflammation and constriction due to its content of powerful mediators (52). In the context of elite sport, repeated plasma exudation has been proposed, *via* release of a wide range of chemoattractant factors and plasma proteins (such as cytokines and growth factors), to contribute to the development of EIB (54). In elite athletes, late onset (>25 yr) of EIB is common (55) and a change of

the contractile properties of the airway smooth muscle is likely to result from combined effects of inflammatory and physical environmental stimuli. In asthma, due to pre-existing inflamed airways, the potent plasma-derived proteins-peptides released in response to hyperpnea-induced epithelial injury could activate an inflammatory cascade, with ensuing bronchoconstriction.

LEUCOCYTE ACTIVATION AS A FUNDAMENTAL MECHANISM OF EIB The acute response

There is strong evidence that the sustained bronchoconstriction following exercise and dry air hyperpnea challenges is due to the release of leukocyte-derived inflammatory mediators in the airways. Similar evidence indicates that hyperosmolar aerosols lead to the release of leukocyte-derived mediators. (28) Specifically, leukocyte-derived eicosanoids including the leukotrienes (LT) and prostaglandins (PG) play a pivotal role. Following exercise challenge in asthmatics with EIB, there is sustained release of cysteinyl LTs (Cys-LTs, LTs C₄, D₄ and E₄) and PGD₂ into the conducting airways, corresponding with the development of acute bronchoconstriction (29, 31, 56). The release of these mediators is attenuated under conditions that reduce the severity of EIB (29, 31, 56, 57). Pharmacological inhibitors of this pathway unequivocally demonstrate that the release of these mediators play a causative role in the pathogenesis of EIB. (29, 31, 58) Inhibition of the key initial enzyme in LT biosynthesis, 5-lipoxygenase (5-LO) with either a 5-LO inhibitor or through inhibition of the 5-LO activating protein (FLAP)(59), significantly reduces the severity of EIB (60). Since LT receptor antagonists (LTRAs) that block the Cys-LT₁ receptor similarly reduce the severity of EIB (29, 60), LTD₄ is likely to mediate bronchoconstriction (as LTD₄ is the primary agonist for the CysLT₁ receptor).

However, the inhibition of EIB by LT modifiers is incomplete, implicating other bronchoconstrictive eicosanoids [such as PGD₂ and 15*S*-Hydroxyeicosatetranoic Acid (15*S*-HETE)], and/or the reduction in bronchoprotective mediators (such as PGE₂). PGD₂ binds to two major receptors; DP1 and DP2, or CRTH2. There has been intense interest in development of CRTH2 inhibitors in chronic asthma because this receptor serves as a key regulator of leukocyte activation, but the results of such inhibitors for treatment of stable asthma have been modest (61). With respect to EIB, the DP1 receptor may serve as the key receptor mediating bronchoconstriction and cough, as the DP1 receptor serves as a key activator of capsaicin sensitive sensory neurons (see below)(62).

Although mediators that initiate bronchoconstriction directly are largely attributed to products derived by leukocytes, the epithelium can directly release mediators, such as 15S-HETE, and can serve as a source of the initial products of eicosanoid metabolism, leading to leukocyte-derived mediators by transcellular metabolism. In some cases, these alterations in eicosanoid synthesis can lead to shunting of arachidonic acid (AA) away from the production of PGE₂ and to an imbalance between the proinflammatory mediators, such as Cys-LTs relative to PGE₂ (63). In addition to shunting of AA away from PGE₂ production, there are other alterations in the epithelium under the influence of IL-13 that reduce the production of PGE₂ (64).

Mast cells and eosinophils are strongly implicated as the cellular sources of Cys-LTs and other eicosanoids (such as PGD₂) in EIB. Following exercise challenge, histamine and the mast cell protease tryptase are released into the airways, demonstrating mast cell degranulation (29). The eosinophil product eosinophilic

cationic protein (ECP) is released into the airways following challenge, and the amount of ECP release varies with the severity of the EIB under different experimental conditions (56). In an analogous situation, pharmacological inhibitors administered prior to mannitol challenge revealed that histamine and possibly PGD₂ are responsible for the early onset of bronchoconstriction, (30, 31, 65), while the release of CysLTs is responsible for sustained bronchoconstriction (30, 66).

The cellular and molecular phenotype associated with EIB

The susceptibility to develop bronchoconstriction in response to hyperpnea challenge varies widely among subjects with established asthma and among individuals without a prior diagnosis of asthma. Thus, the presence of EIB following a specific challenge tests represents clinically recognizable phenotype of asthma. Recent work to understand the biological basis of this syndrome has revealed a consistent biological endotype related to cellular inflammation, particularly the density or mast cells in the airway epithelium (67, 68). Quantitative morphometry of airway biopsies, using design-based stereology, showed that the density of mast cells localized to the airway epithelium is significantly higher in subjects with EIB relative to individuals without asthma and those with asthma who do not have EIB (Figure 4 A&B) (69). Genomewide expression studies of airway epithelial samples have also revealed high expression of the mast cell genes tryptase and carboxypeptidase A3 (CPA3), but low expression of chymase (Figure 4 C&D) (69, 70). This unique intraepithelial mast cell phenotype is also a biomarker of the "Th2 high" molecular phenotype of asthma (71, 72) that is IL-13 mediated (73, 74). The association between airway eosinophilia and the severity of EIB also provides evidence of this association with "type-2" inflammation (36, 75). Further, modifications of airway proteins that reflect

eosinophil activation are associated with the severity of EIB (76). Individuals with EIB have higher levels of Cys-LTs in induced sputum (36) and in exhaled breath condensate (EBC) (77). Higher levels of Cys-LTs in this group likely reflects the ability of mast cells and eosinophils to direct the eicosanoid pathway towards the synthesis of Cys-LTs as these cells are the predominant source of LTC4 synthase (LTC₄S), the key enzyme directing this pathway towards Cys-LT formation in the airways (78). The rate-limiting step in eicosanoid formation occurs at the initial release of AA by the hydrolysis of the sn-2 position of membrane phospholipids by a family of phospholipase A2 (PLA2) enzymes. Although the cytosolic PLA2s are known to serve this regulatory function, recent work has revealed that secreted PLA₂ (sPLA₂) activity is increased in asthma and is predominantly derived from sPLA₂ groups IIA and X (i.e., sPLA₂-IIA and sPLA₂-X) (63, 79). Further work has revealed that the level of sPLA₂-X is elevated in the airways of subjects with asthma, and tends to be higher in subjects with EIB (Figure 5) (80). In addition, sPLA2-X is posttranslationally activated by transglutaminase 2 (TGM2), an enzyme that is overexpressed in the airways of patients with EIB (70). As the key source of sPLA₂-X is the epithelium, and the enzyme can serve as an activator of myeloid cells, such as eosinophils, to generate the production of Cys-LTs (81), these results raise the possibility that sPLA2-X serves as a critical initiator of eicosanoid production in response to exercise-hyperpnea.

Other markers of airway inflammation have also been associated with EIB, including an increase in the fraction of exhaled nitric oxide (F_{ENO}) (82), especially in subjects with atopy (83). The levels of 8-isoprostanes, non-enzymatic products of phospholipid oxidation, are increased in EBC of asthmatics with EIB, and correlated with the

severity of EIB (84). A reduction in the level of the protective eicosanoid lipoxin A4 has also been described in EIB (85).

In summary, patients who are susceptible to EIB have epithelial shedding, overproduction of inflammatory mediators, such as Cys-LTs, relative underproduction of protective lipid mediators, and infiltration of the airways with eosinophils and mast cells (Figure 2).

Refractoriness to repeated exercise

A refractory period after EIB for ~2 hours occurs in 50 to 60% of people with asthma. This refractoriness is not due to a reduction in heat or water loss from the airways with repeated exercise. Many theories have been suggested to explain refractoriness.(86) The most compelling involve release of mediators during exercise and their sustained presence after exercise. One theory relates to the bronchodilator effect of PGE2 and the second relates to development of tolerance to the mediators of bronchoconstriction, particularly to the cyst-LTs. The finding that non-steroidal antiinflammatory agent indomethacin could prevent refractoriness to EIB was explained on the basis of blocking production of PGE₂ during initial challenge.(87) It is also known that refractoriness to exercise occurs after challenge with LTD₄.(88) Increased levels of PGE₂ has recently been reported during the refractory period following hyperpnea of dry air.(89) It has been proposed that repeated stimulation of the cyst-LT receptors lead to their internalisation on the bronchial smooth muscle that would explain refractoriness. When mediators were measured in response to repeated challenge with inhaled mannitol, the subjects most refractory to the second challenge had significantly higher levels of mediators following challenge. (90) Both

prostaglandins and leukotrienes work through G protein coupled receptors and the events proposed to explain refractoriness are illustrated in Figure 6.(86)

NEUROGENIC FACTORS AND EIB

Sensory nerve activation

The airways are innervated by network of sensory nerves that sense mechanical, nociceptive and inflammatory signals through afferent C and A fibers. These sensory neurons send signals centrally, leading to a coordinated response with the efferent parasympathetic nervous system (see below), as well as through a local reflex called retrograde axonal transmission. It is becoming increasingly clear that airway hyperresponsiveness (AHR) is mediated through alterations in both the sensory and autonomic nervous system pathways in the airways (91). This neural plasticity in asthma is the result of changes in the composition of the different types of sensory neurons, the production of neurokinins, such as neurokinin A (NKA) and substance P (SP), and the density of transient receptor potential (TRP) channels, such as TRP vanilloid receptor 1 (TRPV1) which serves as the receptor for capsaicin. These changes are the consequence of chronic inflammation and the generation of neurotropins, such as nerve growth factor (92). In murine models, the sensory neurons are required for the development of AHR (93). The precise anatomical changes in the sensory nervous system of the airways in human asthma are not known in detail; however, the threshold for activation of the sensory nerves by the TRPV1 agonist capsaicin is lowered in asthma (94).

In animal models of EIB, it is well established that the sensory nerves mediate bronchoconstriction in response to a period of dry air hyperpnea, and that the generation of leukocyte-derived eicosanoids plays the dominant role in activating this neural pathway. In a guinea pig model of hyperpnea-induced bronchoconstriction (HIB), inhibition of Cys-LT signaling, with either a 5-LO inhibitor or a LTRA, inhibited HIB and the release of neurokinins, while a neurokinin 2 receptor antagonist inhibited HIB, but not the release of leukotrienes, suggesting that leukotrienes cause bronchoconstriction via sensory nerves during HIB (95). In a dog model, a combination neurokinin 1 and 2 receptor antagonist inhibited HIB and the generation of LTs that are known in this model to cause HIB (96). Sensory nerves are activated directly by osmotic stimuli, but several eicosanoids can either directly activate or alter the activation threshold of sensory nerves (97). A recent study demonstrated that PGD₂, which is predominantly derived from mast cells, initiates sensory nerve activation through the DP1 receptor, regulating airway tone (62). Although the function of neurokinins during EIB has not been fully tested, there is evidence that the levels of NKA and Cys-LTs in the airways are correlated with the amount of bronchoconstriction following exercise challenge and with the release of MUC5AC (50). Overall, these findings suggest that in humans, bronchoconstriction and mucus release following exercise challenge is the consequence of sensory nerve activation that is initiated by the release of eicosanoids from airway leukocytes, such as mast cells.

THE ROLE OF REGIONAL ALTERATIONS IN THE AIRWAYS AND THE DEVELOPMENT OF EIB

In model systems, particularly in animal models with either airway injury or inflamed airways, the mechanism leading to AHR includes regional altertions in airway diameter that are heterogenous, leading to regional airway closure that contributes to

the development of airflow obstruction (98, 99). With the use of hyperpolarized helium imaging to assess regional ventilation, it is apparent that the development of airflow obstruction after exercise represents heterogeneous effects on airway caliber, ranging from conducting airway narrowing to frank airway closure (100). Treatment to reduce the severity of EIB reduces these heterogeneous regional defects in ventilation (101). These findings suggest that the stimulus for EIB causes bronchoconstriction in regions of the airways that are specifically susceptible to this stimulus, most likely because of alterations in the airway wall related to aiway inflammation. Such regional alterations in ventilation are also known to be present at baseline in subjects with asthma, and are associated with the severity of asthma (102). There is evidence that areas of regional heterogeneity in ventilation tend to persist over time, further suggesting that these regions of the airways have heightened susceptibility to airway narrowing (103). Additional studies are needed to fully understand the nature of these regional alterations in airway structure and airway dysfunction in humans.

SUMMARY/FUTURE CONSIDERATIONS

Water evaporation from the airways and thermal changes during exercise-hyperpnea are well established stimuli to EIB. In recents years, it has become increasingly clear that a dysfunctional airway epithelium – characterised by a loss of physical barrier integrity and impairment in signaling and secretory functions – renders individuals with asthma, as well as elite athletes susceptible to EIB. In these two populations, lipid mediator release (cys-LTs and PGD₂) has repeatedly been observed in response to exercise challenge (or its surrogates), highlighting the inflammatory nature of the condition. Additional work is warranted to define the exact role of sensory nerves and

of regional alterations in ventilation in the pathogenesis of EIB. As EIB represents a precise asthma phenotype that can be characterized clinically, and is associated with an increasingly well defined biological endotype, these findings should prove useful to target therapy for this important aspect of asthma.

(4,132 words)

REFERENCES

- 1. Rundell KW, Anderson SD, Sue-Chu M, Bougualt V, Boulet L-P. Air quality and temperature effects on exercise-induced bronchoconstriction. Comprehensive Physiology 2015;5:579-610.
- 2. Deal EC, McFadden ER, Ingram RH, Strauss RH, Jaeger JJ. Role of respiratory heat exchange in production of exercise-induced asthma. J Appl Physiol:Respirat Environ Exercise Physiol. 1979;46:467-75.
- 3. McFadden ER, Lenner KA, Strohl KP. Postexertional airway rewarming and thermally induced asthma. J Clin Invest. 1986;78(1):18-25.
- 4. McFadden ER, Gilbert IA. Exercise-induced asthma as a vascular phenomenon. Exercise-induced Asthma. 1999;130:115-35.
- 5. Gilbert IA, Fouke JM, McFadden ER. Heat and water flux in the intrathoracic airways and exercise-induced asthma. J Appl Physiol. 1987;63(4):1681-91.
- 6. Anderson SD, Holzer K. Exercise-induced asthma: Is it the right diagnosis in elite athletes? J Allergy Clin Immunol. 2000;106(3):419-28.
- 7. Anderson SD, Schoeffel RE, Black JL, Daviskas E. Airway cooling as the stimulus to exercise-induced asthma a re-evaluation. Eur J Respir Dis. 1985;67(1):20-30.
- 8. Hahn A, Anderson SD, Morton AR, Black JL, Fitch KD. A re-interpretation of the effect of temperature and water content of the inspired air in exercise-induced asthma. Am Rev Respir Dis. 1984;130(4):575-9.
- 9. Eschenbacher WL, Moore TB, Lorenzen TJ, Weg JG, Gross KB. Pulmonary responses of asthmatic and normal subjects to different temperature and humidity conditions in an environmental chamber. Lung. 1992;170(1):51-62.
- 10. Zawadski DK, Lenner KA, McFadden ER. Comparison of intraairway temperatures in normal and asthmatic subjects after hyperpnea with hot, cold, and ambient air. Am Rev Respir Dis. 1988;138(6):1553-8.
- 11. Strauss RH, McFadden ER, Ingram RH, Deal EC, Jaeger JJ, Stearns D. Influence of heat and humidity on the airway obstruction induced by exercise in asthma. J Clin Invest. 1978;61(2):433-40.
- 12. Anderson SD, Schoeffel RE, Follet R, Perry CP, Daviskas E, Kendall M. Sensitivity to heat and water loss at rest and during exercise in asthmatic patients. Eur J Respir Dis. 1982;63(5):459-71.
- 13. Anderson SD, Daviskas E, Schoeffel RE, Unger SF. Prevention of severe exercise-induced asthma with hot humid air. Lancet. 1979;2(8143):629.
- 14. Anderson SD. Is there a unifying hypothesis for exercise-induced asthma? J Allergy Clin Immunol. 1984;73(5 Part 2):660-5.
- 15. Anderson SD, Daviskas E, Smith CM. Exercise-induced asthma: a difference in opinion regarding the stimulus. Allergy Proc. 1989;10(3):215-26.
- 16. Anderson SD, Daviskas E. Pathophysiology of exercise-induced asthma: role of respiratory water loss. In: Weiler J, editor. Allergic and Respiratory Disease in Sports Medicine. New York: Marcel Dekker; 1997. p. 87-114.
- 17. McFadden ER, Jr., Pichurko BM, Bowman HF, Ingenito E, Burns S, Dowling N, et al. Thermal mapping of the airways in humans. J Appl Physiol. 1985;58(2):564-70.
- 18. Daviskas E, Gonda I, Anderson SD. Local airway heat and water vapour losses. Respiration Physiology. 1991;84(1):115-32.
- 19. Daviskas E, Gonda I, Anderson SD. Mathematical modelling of the heat and water transport in the human respiratory tract. J Appl Physiol. 1990;69(1):362-72.

- 20. Davis MS, Daviskas E, Anderson SD, Kotaru C, Hejal RB, Finigan JH, et al. Airway surface fluid desiccation during isocapnic hyperpnea. J Appl Physiol. 2003;94(6):2545-7.
- 21. Daviskas E, Anderson SD, Gonda I, Chan HK, Cook P, Fulton R. Changes in mucociliary clearance during and after isocapnic hyperventilation in asthmatic and healthy subjects. Eur Respir J. 1995;8(5):742-51.
- 22. Anderson SD, Daviskas E. The mechanism of exercise-induced asthma is J Allergy Clin Immunol. 2000;106(3):453-9.
- 23. Anderson SD, Daviskas E. The airway microvasculature and exercise-induced asthma. Thorax. 1992;47(9):748-52.
- 24. Anderson SD, Daviskas E. Airway drying and exercise induced asthma. In: McFadden ER, editor. Exercise Induced Asthma *Lung Biology in Health and Disease*. New York: Marcel Dekker; 1999. p. 77-113.
- 25. Agostoni P, Arena V, Doria E, Susini G. Inspired gas relative humidity affects systemic to pulmonary bronchial blood flow in humans. Chest. 1990;97(6):1377-80.
- 26. Smith CM, Anderson SD. A comparison between the airway response to isocapnic hyperventilation and hypertonic saline in subjects with asthma. Eur Respir J. 1989;2(1):36-43.
- 27. Gibson PG, Saltos N, Borgas T. Airway mast cells and eosinophils correlate with clinical severity and airway hyperresponsiveness in corticosteroid-treated asthma. J Allergy Clin Immunol. 2000;105(4):752-9.
- 28. Brannan JD, Gulliksson M, Anderson SD, Chew N, Kumlin M. Evidence of mast cell activation and leukotriene release after mannitol inhalation. Eur Respir J. 2003;22(3):491-6.
- 29. Hallstrand TS, Moody MW, Wurfel MM, Schwartz LB, Henderson WR, Aitken ML. Inflammatory basis of exercise-induced bronchoconstriction. Am J Respir Crit Care Med. 2005;172(6):679-86.
- 30. Brannan JD, Gulliksson M, Anderson SD, Chew N, Seale JP, Kumlin M. Inhibition of mast cell PGD₂ release protects against mannitol-induced airway narrowing. Eur Respir J. 2006;27(5):944-50.
- 31. Kippelen P, Larsson J, Anderson SD, Brannan JD, Dahlen B, Dahlen SE. Effect of sodium cromoglycate on mast cell mediators during hyperpnea in athletes. Med Sci Sports Exerc. 2010;42(10):1853-60.
- 32. Carroll NG, Mutavdzic S, James AL. Distribution and degranulation of airway mast cells in normal and asthmatic subjects. Eur Respir J. 2002;19(5):879-85.
- 33. Chimenti L, Morici G, Paternò A, Santagata R, Bonanno A, Profita M, et al. Bronchial epithelial damage after a half-marathon in nonasthmatic amateur runners. Am J Physiol Lung Cell Mol Physiol. 2010;298(6):L857-62.
- 34. Bolger C, Tufvesson E, Sue-Chu M, Devereux G, Ayres JG, Bjermer L, et al. Hyperpnea-induced bronchoconstriction and urinary CC16 levels in athletes. Med Sci Sports Exerc. 2011;43(7):1207-13.
- 35. Bolger C, Tufvesson E, Anderson SD, Devereux G, Ayres JG, Bjermer L, et al. Effect of inspired air conditions on exercise-induced bronchoconstriction and urinary CC16 levels in athletes. J Appl Physiol (1985). 2011;111(4):1059-65.
- 36. Hallstrand TS, Moody MW, Aitken ML, Henderson WR. Airway immunopathology of asthma with exercise-induced bronchoconstriction. J Allergy Clin Immunol. 2005;116(3):586-93.
- 37. Holgate ST. Epithelium dysfunction in asthma. J Allergy Clin Immunol. 2007;120(6):1233-44; quiz 45-6.

- 38. Anderson SD, Kippelen P. Airway injury as a mechanism for exercise-induced bronchoconstriction in elite athletes. J Allergy Clin Immunol. 2008;122(2):225-35; quiz 36-7.
- 39. Carlsen KH, Anderson SD, Bjermer L, Bonini S, Brusasco V, Canonica W, et al. Exercise-induced asthma, respiratory and allergic disorders in elite athletes: epidemiology, mechanisms and diagnosis: part I of the report from the Joint Task Force of the European Respiratory Society (ERS) and the European Academy of Allergy and Clinical Immunology (EAACI) in cooperation with GA2LEN. Allergy. 2008;63(4):387-403.
- 40. Cabral AL, Conceição GM, Fonseca-Guedes CH, Martins MA. Exercise-induced bronchospasm in children: effects of asthma severity. Am J Respir Crit Care Med. 1999;159(6):1819-23.
- 41. Karjalainen J, Lindqvist A, Laitinen LA. Seasonal variability of exercise-induced asthma especially outdoors. Effect of birch pollen allergy. Clin Exp Allergy. 1989;19(3):273-8.
- 42. Djukanović R, Feather I, Gratziou C, Walls A, Peroni D, Bradding P, et al. Effect of natural allergen exposure during the grass pollen season on airways inflammatory cells and asthma symptoms. Thorax. 1996;51(6):575-81.
- 43. Carbonnelle S, Francaux M, Doyle I, Dumont X, de Burbure C, Morel G, et al. Changes in serum pneumoproteins caused by short-term exposures to nitrogen trichloride in indoor chlorinated swimming pools. Biomarkers. 2002;7(6):464-78.
- 44. Krishna MT, Springall D, Meng QH, Withers N, Macleod D, Biscione G, et al. Effects of ozone on epithelium and sensory nerves in the bronchial mucosa of healthy humans. Am J Respir Crit Care Med. 1997;156(3 Pt 1):943-50.
- 45. Matsui H, Davis CW, Tarran R, Boucher RC. Osmotic water permeabilities of cultured, well-differentiated normal and cystic fibrosis airway epithelia. J Clin Invest. 2000;105(10):1419-27.
- 46. Tarran R. Regulation of airway surface liquid volume and mucus transport by active ion transport. Proc Am Thorac Soc. 2004;1(1):42-6.
- 47. Fahy JV, Dickey BF. Airway mucus function and dysfunction. N Engl J Med. 2010;363(23):2233-47.
- 48. Olenec JP, Kim WK, Lee WM, Vang F, Pappas TE, Salazar LE, et al. Weekly monitoring of children with asthma for infections and illness during common cold seasons. J Allergy Clin Immunol. 2010;125(5):1001-6.e1.
- 49. Bonini M, Gramiccioni C, Fioretti D, Ruckert B, Rinaldi M, Akdis C, et al. Asthma, allergy and the Olympics: a 12-year survey in elite athletes. Curr Opin Allergy Clin Immunol. 2015;15(2):184-92.
- 50. Hallstrand TS, Debley JS, Farin FM, Henderson WR. Role of MUC5AC in the pathogenesis of exercise-induced bronchoconstriction. J Allergy Clin Immunol. 2007;119(5):1092-8.
- 51. Bougault V, Loubaki L, Joubert P, Turmel J, Couture C, Laviolette M, et al. Airway remodeling and inflammation in competitive swimmers training in indoor chlorinated swimming pools. J Allergy Clin Immunol. 2012;129(2):351-8.e1.
- 52. Persson CG, Erjefält JS, Andersson M, Greiff L, Svensson C. Extravasation, lamina propria flooding and lumenal entry of bulk plasma exudate in mucosal defence, inflammation and repair. Pulm Pharmacol. 1996;9(3):129-39.
- 53. Persson CG. Plasma exudation and asthma. Lung. 1988;166(1):1-23.
- 54. Anderson SD, Kippelen P. Exercise-induced bronchoconstriction: pathogenesis. Curr Allergy Asthma Rep. 2005;5(2):116-22.

- 55. Fitch KD. An overview of asthma and airway hyper-responsiveness in Olympic athletes. Br J Sports Med. 2012;46(6):413-6.
- 56. Mickleborough TD, Lindley MR, Ray S. Dietary salt, airway inflammation, and diffusion capacity in exercise-induced asthma. Med Sci Sports Exerc. 2005;37(6):904-14.
- 57. Simpson AJ, Bood JR, Anderson SD, Romer LM, Dahlén B, Dahlén SE, et al. A standard, single dose of inhaled terbutaline attenuates hyperpnea-induced bronchoconstriction and mast cell activation in athletes. J Appl Physiol (1985). 2016;120(9):1011-7.
- 58. Kippelen P, Larsson J, Anderson SD, Brannan JD, Delin I, Dahlen B, et al. Acute effects of beclomethasone on hyperpnea-induced bronchoconstriction. Med Sci Sports Exerc. 2010;42(2):273-80.
- 59. Kent SE, Bentley JH, Miller D, Sterling R, Menendez R, Tarpay M, et al. The effect of GSK2190915, a 5-lipoxygenase-activating protein inhibitor, on exercise-induced bronchoconstriction. Allergy Asthma Proc. 2014;35(2):126-33.
- 60. Hallstrand TS, Henderson WR. Role of leukotrienes in exercise-induced bronchoconstriction. Curr Allergy Asthma Rep. 2009;9(1):18-25.
- 61. Farne H, Jackson DJ, Johnston SL. Are emerging PGD2 antagonists a promising therapy class for treating asthma? Expert Opin Emerg Drugs. 2016;21(4):359-64.
- 62. Maher SA, Birrell MA, Adcock JJ, Wortley MA, Dubuis ED, Bonvini SJ, et al. Prostaglandin D2 and the role of the DP1, DP2 and TP receptors in the control of airway reflex events. Eur Respir J. 2015;45(4):1108-18.
- 63. Hallstrand TS, Chi EY, Singer AG, Gelb MH, Henderson WR, Jr. Secreted phospholipase A2 group X overexpression in asthma and bronchial hyperresponsiveness. Am J Respir Crit Care Med. 2007;176(11):1072-8.
- 64. Trudeau J, Hu H, Chibana K, Chu HW, Westcott JY, Wenzel SE. Selective downregulation of prostaglandin E_2 -related pathways by the Th2 cytokine IL-13. J Allergy Clin Immunol. 2006;117(6):1446-54.
- 65. Kippelen P, Larsson J, Anderson SD, Brannan JD, Delin I, Dahlen B, et al. Acute effects of beclomethasone on hyperpnea-induced bronchoconstriction. Med Sci Sports Exerc. 2010;42(2):273-80.
- 66. Currie GP, Haggart K, Lee DK, Fowler SJ, Wilson AM, Brannan JD, et al. Effects of mediator antagonism on mannitol and adenosine monophosphate challenges. Clin Exp Allergy. 2003;33(6):783-8.
- 67. Lötvall J, Akdis CA, Bacharier LB, Bjermer L, Casale TB, Custovic A, et al. Asthma endotypes: a new approach to classification of disease entities within the asthma syndrome. J Allergy Clin Immunol. 2011;127(2):355-60.
- 68. Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. Nat Med. 2012;18(5):716-25.
- 69. Lai Y, Altemeier WA, Vandree J, Piliponsky AM, Johnson B, Appel CL, et al. Increased density of intraepithelial mast cells in patients with exercise-induced bronchoconstriction regulated through epithelially derived thymic stromal lymphopoietin and IL-33. J Allergy Clin Immunol. 2014;133(5):1448-55.
- 70. Hallstrand TS, Wurfel MM, Lai Y, Ni Z, Gelb MH, Altemeier WA, et al. Transglutaminase 2, a novel regulator of eicosanoid production in asthma revealed by genome-wide expression profiling of distinct asthma phenotypes. PLoS One. 2010;5(1):e8583.

- 71. Dougherty RH, Sidhu SS, Raman K, Solon M, Solberg OD, Caughey GH, et al. Accumulation of intraepithelial mast cells with a unique protease phenotype in T(H)2-high asthma. J Allergy Clin Immunol. 2010;125(5):1046-53 e8.
- 72. Woodruff PG, Boushey HA, Dolganov GM, Barker CS, Yang YH, Donnelly S, et al. Genome-wide profiling identifies epithelial cell genes associated with asthma and with treatment response to corticosteroids. Proc Natl Acad Sci U S A. 2007;104(40):15858-63.
- 73. Woodruff PG, Modrek B, Choy DF, Jia G, Abbas AR, Ellwanger A, et al. Thelper type 2-driven inflammation defines major subphenotypes of asthma. Am J Respir Crit Care Med. 2009;180(5):388-95.
- 74. Peters MC, Mekonnen ZK, Yuan S, Bhakta NR, Woodruff PG, Fahy JV. Measures of gene expression in sputum cells can identify TH2-high and TH2-low subtypes of asthma. J Allergy Clin Immunol. 2014;133(2):388-94.
- 75. Duong M, Subbarao P, Adelroth E, Obminski G, Strinich T, Inman M, et al. Sputum eosinophils and the response of exercise-induced bronchoconstriction to corticosteroid in asthma. Chest. 2008;133(2):404-11.
- 76. Jin H, Hallstrand TS, Daly DS, Matzke MM, Nair P, Bigelow DJ, et al. A halotyrosine antibody that detects increased protein modifications in asthma patients. J Immunol Methods. 2014;403(1-2):17-25.
- 77. Carraro S, Corradi M, Zanconato S, Alinovi R, Pasquale MF, Zacchello F, et al. Exhaled breath condensate cysteinyl leukotrienes are increased in children with exercise-induced bronchoconstriction. J Allergy Clin Immunol. 2005;115(4):764-70.
- 78. Cai Y, Bjermer L, Halstensen TS. Bronchial mast cells are the dominating LTC₄S-expressing cells in aspirin-tolerant asthma. Am J Respir Cell Mol Biol. 2003;29(6):683-93.
- 79. Hallstrand TS, Lai Y, Ni Z, Oslund RC, Henderson WR, Jr., Gelb MH, et al. Relationship between levels of secreted phospholipase A groups IIA and X in the airways and asthma severity. Clin Exp Allergy. 2011;41(6):801-10.
- 80. Hallstrand TS, Lai Y, Altemeier WA, Appel CL, Johnson B, Frevert CW, et al. Regulation and function of epithelial secreted phospholipase A2 group X in asthma. Am J Respir Crit Care Med. 2013;188(1):42-50.
- 81. Lai Y, Oslund RC, Bollinger JG, Henderson WR, Jr., Santana LF, Altemeier WA, et al. Eosinophil cysteinyl leukotriene synthesis mediated by exogenous secreted phospholipase A2 group X. J Biol Chem. 2010;285(53):41491-500.
- 82. Scollo M, Zanconato S, Ongaro R, Zaramella C, Zacchello F, Baraldi E. Exhaled nitric oxide and exercise-induced bronchoconstriction in asthmatic children. Am J Respir Crit Care Med. 2000;161(3 Pt 1):1047-50.
- 83. Malmberg LP, Pelkonen AS, Mattila PS, Hammaren-Malmi S, Makela MJ. Exhaled nitric oxide and exercise-induced bronchoconstriction in young wheezy children interactions with atopy. Pediatr Allergy Immunol. 2009;20(7):673-8.
- 84. Barreto M, Villa MP, Olita C, Martella S, Ciabattoni G, Montuschi P. 8-Isoprostane in exhaled breath condensate and exercise-induced bronchoconstriction in asthmatic children and adolescents. Chest. 2009;135(1):66-73.
- 85. Tahan F, Saraymen R, Gumus H. The role of lipoxin A4 in exercise-induced bronchoconstriction in asthma. J Asthma. 2008;45(2):161-4.
- 86. Larsson J, Anderson SD, Dahlen S-E, Dahlen B. Refractoriness to Exercise Challenge: A review of the mechanisms old and new. Immunol Allergy Clin N Am. 2013;33:329-45.

- 87. O'Byrne PM, Jones GL. The effect of indomethacin on exercise-induced bronchoconstriction and refractoriness after exercise. Am Rev Respir Dis. 1986;134(1):69-72.
- 88. Manning PJ, Watson RM, O'Byrne PM. Exercise-induced refractoriness in asthmatic subjects involves leukotriene and prostagland in interdependent mechanisms. Am Rev Respir Dis. 1993;148(4 Pt 1):950-4.
- 89. Bood JR, Sundblad BM, Delin I, Sjödin M, Larsson K, Anderson SD, et al. Urinary excretion of lipid mediators in response to repeated eucapnic voluntary hyperpnea in asthmatic subjects. J Appl Physiol (1985). 2015;119(3):272-9.
- 90. Larsson J, Perry CP, Anderson SD, Brannan JD, Dahlen S-E, Dahlen B. The occurrence of refractoriness and mast cell mediator release following mannitol-induced bronchoconstriction. J Appl Physiol. 2011;110:1029-35.
- 91. Zaccone EJ, Undem BJ. Airway Vagal Neuroplasticity Associated with Respiratory Viral Infections. Lung. 2016;194(1):25-9.
- 92. Ogawa H, Azuma M, Uehara H, Takahashi T, Nishioka Y, Sone S, et al. Nerve growth factor derived from bronchial epithelium after chronic mite antigen exposure contributes to airway hyperresponsiveness by inducing hyperinnervation, and is inhibited by in vivo siRNA. Clin Exp Allergy. 2012;42(3):460-70.
- 93. Trankner D, Hahne N, Sugino K, Hoon MA, Zuker C. Population of sensory neurons essential for asthmatic hyperreactivity of inflamed airways. Proc Natl Acad Sci U S A. 2014;111(31):11515-20.
- 94. Belvisi MG, Birrell MA, Khalid S, Wortley MA, Dockry R, Coote J, et al. Neurophenotypes in Airway Diseases. Insights from Translational Cough Studies. Am J Respir Crit Care Med. 2016;193(12):1364-72.
- 95. Lai YL, Lee SP. Mediators in hyperpnea-induced bronchoconstriction of guinea pigs. Naunyn Schmiedebergs Arch Pharmacol. 1999;360(5):597-602.
- 96. Freed AN, McCulloch S, Meyers T, Suzuki R. Neurokinins modulate hyperventilation-induced bronchoconstriction in canine peripheral airways. Am J Respir Crit Care Med. 2003;167(8):1102-8.
- 97. Taylor-Clark TE, Nassenstein C, Undem BJ. Leukotriene D4 increases the excitability of capsaicin-sensitive nasal sensory nerves to electrical and chemical stimuli. Br J Pharmacol. 2008;154(6):1359-68.
- 98. Lundblad LK, Thompson-Figueroa J, Allen GB, Rinaldi L, Norton RJ, Irvin CG, et al. Airway hyperresponsiveness in allergically inflamed mice: the role of airway closure. Am J Respir Crit Care Med. 2007;175(8):768-74.
- 99. Bates JH, Cojocaru A, Haverkamp HC, Rinaldi LM, Irvin CG. The synergistic interactions of allergic lung inflammation and intratracheal cationic protein. Am J Respir Crit Care Med. 2008;177(3):261-8.
- 100. Samee S, Altes T, Powers P, de Lange EE, Knight-Scott J, Rakes G, et al. Imaging the lungs in asthmatic patients by using hyperpolarized helium-3 magnetic resonance: assessment of response to methacholine and exercise challenge. J Allergy Clin Immunol. 2003;111(6):1205-11.
- 101. Kruger SJ, Niles DJ, Dardzinski B, Harman A, Jarjour NN, Ruddy M, et al. Hyperpolarized Helium-3 MRI of exercise-induced bronchoconstriction during challenge and therapy. J Magn Reson Imaging. 2014;39(5):1230-7.
- 102. de Lange EE, Altes TA, Patrie JT, Gaare JD, Knake JJ, Mugler JP, et al. Evaluation of asthma with hyperpolarized helium-3 MRI: correlation with clinical severity and spirometry. Chest. 2006;130(4):1055-62.

- 103. Niles DJ, Kruger SJ, Dardzinski BJ, Harman A, Jarjour NN, Ruddy M, et al. Exercise-induced bronchoconstriction: reproducibility of hyperpolarized 3He MR imaging. Radiology. 2013;266(2):618-25.
- 104. Hallstrand TS. New insights into pathogenesis of exercise-induced bronchoconstriction. Curr Opin Allergy Clin Immunol. 2012;12(1):42-8.

FIGURE CAPTIONS

Figure 1. Both airway cooling and mucosal dehydration occur in response to evaporative water loss from the airway surface. These events lead to exercise-induced bronchoconstriction. The figure is taken from Rundell et al. (1), and modified, with permission, from the original by Anderson and Daviskas (24).

Figure 2. Cells involved in exercise-induced bronchoconstriction. The epithelial cell layer is the source of prostagland E_2 , a bronchodilating prostagland in. The levels are lower in asthmatics. The mast cells are the source of prostagland in D_2 and cysteinyl leukotrienes and eosinophils are also a source of cysteinyl leukotrienes. These bronchoconstricting mediators likely act via sensory nerves to cause smooth muscle contraction and airway narrowing. This has been demonstrated indirectly by the increase in leukotrienes being related to the increase in mucin (MUC5A) released from goblet cells and the release of neurokin A. The bronchial smooth muscle may also be stimulated directly by some of these same mediators. The figure is taken from Rundell et al. (1), and modified, with permission, from the original by Hallstrand et al. (104).

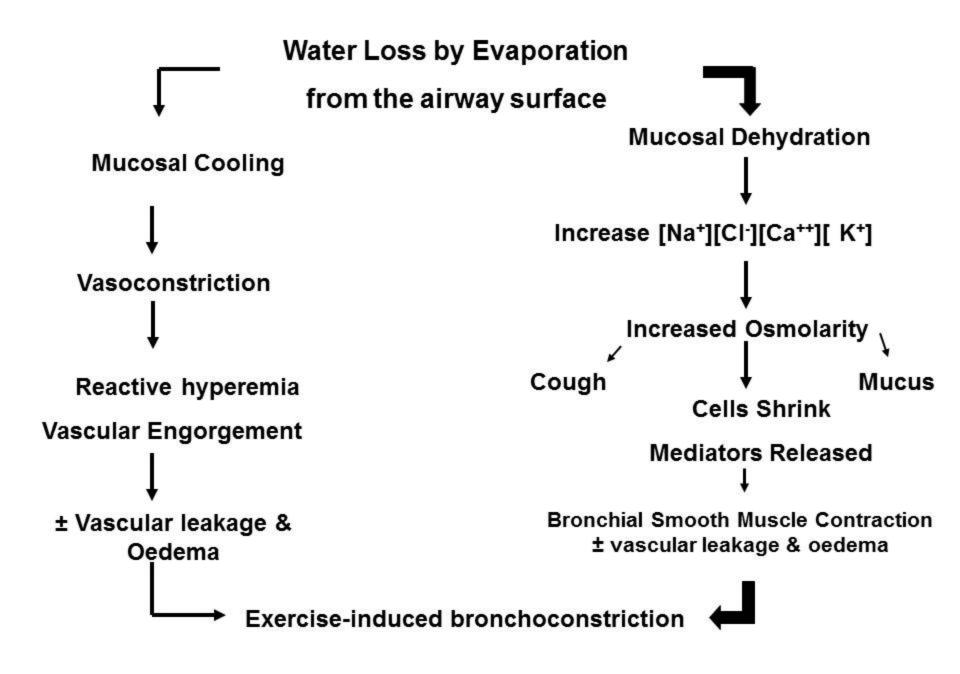
Figure 3. The severity of the bronchoconstriction after exercise is similar in the same individual over a wide range of inspired air temperatures. Panel A drawn from Deal et al., (2), with the reduction in forced expiratory volume in 1 sec (FEV₁) after exercise expressed as percentage of the pre-exercise level; Panel B drawn from Hahn et al. (8), with the reduction in peak expiratory flow rate (PEFR) expressed as percentage of the pre-exercise level.

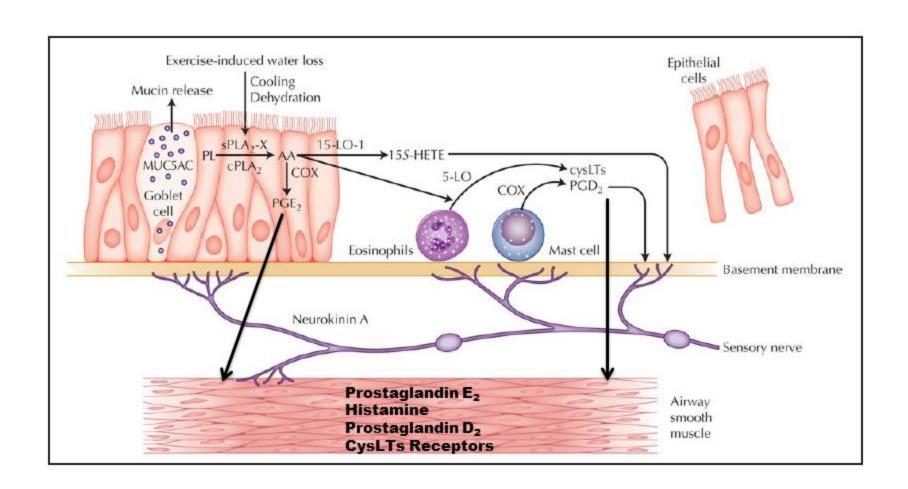
Figure 4. Mast cell infiltration of the airway epithelium in asthma. Quantitative morphometry of endobronchial biopsy samples and epithelial brush gene expression from individuals without asthma (Ctrl) compared with individuals with asthma and EIB (EIB+) and individuals with asthma without EIB (EIB-). Quantitative morphometry reveals that the number of mast cells per volume of airway epithelium (A) and the number of mast cells per area of the basal lamina (B) are substantially increased in individuals with EIB compared to asthmatics without EIB and normal controls. Epithelial brushings show a pattern of mast cell gene expression of tryptase (C) and CPA3 (D) with high expression of these genes in EIB, but lower expression of Chymase (data not shown). Adapted from Lai et al. (69), with permission.

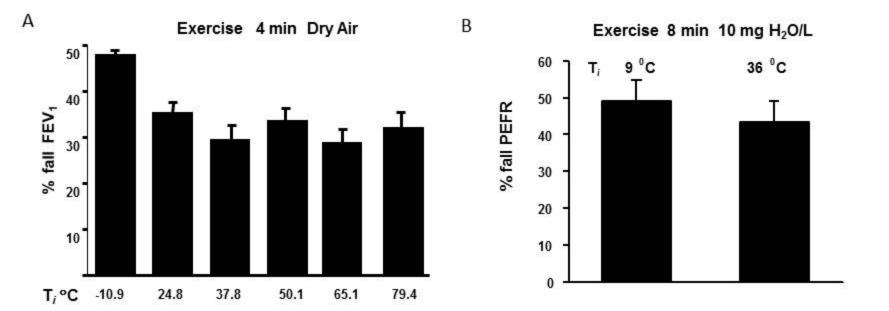
Figure 5. Secreted phospholipase A₂ (sPLA₂-X) protein levels in the airways in asthma. Quantitative assessment of sPLA₂-X protein levels in induced sputum supernatant and lysates of epithelial brushings from the lower airways in individuals without asthma (Ctrl) compared with individuals with asthma and EIB (EIB+) and individuals with asthma without EIB (EIB-). Induced sputum levels of sPLA₂-X were elevated in asthma, particularly in individuals with EIB (A). The levels of sPLA₂-X in the epithelium were elevated only in the asthma group with EIB (B). Adapted from Hallstrand et al. (80), with permission.

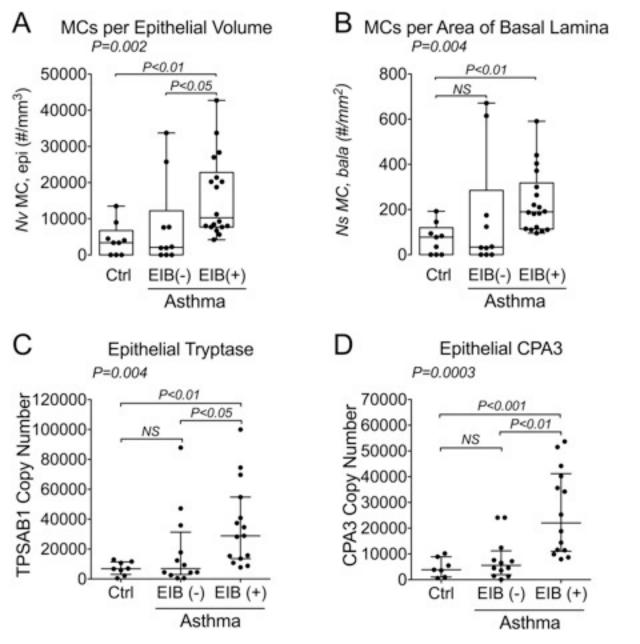
Figure 6. Desensitization of Cys-LT1 as a mechanism of refractoriness. With exercise, there is release of cysteinyl leukotrienes from mast cells that stimulate the GPCR Cys-LT1 causing contraction of the muscle and exercise-induced bronchoconstriction. The PGs generated during the same exercise could, via stimulation of the GPCRs DP1/EP1-4/TP, lead to crosstalk so that the Cys-LT1 is

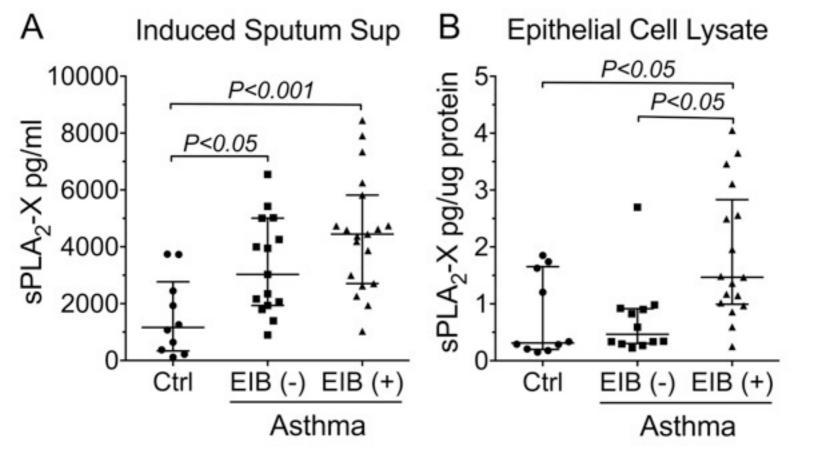
desensitized through phosphorylation and internalization. Refractoriness to a second exercise challenge within a short period occurs as a result. When the levels of mediators return to baseline values, the Cys-LT1 receptors are recycled back to the cell surface and the muscle becomes sensitive to further stimulation. When generation of PGs is inhibited by agents, such as indomethacin and flurbiprofen, then there is limited desensitization of the Cys-LT1 and, therefore, limited refractoriness. Cys-LT1, cysteinyl leukotriene receptor1; DP1, prostaglandin D receptor1; EP1-4, prostaglandin E receptor1-4; GPCR, G-protein coupled receptor; TP, thromboxane receptor. From Larsson et al. (86), with permission.

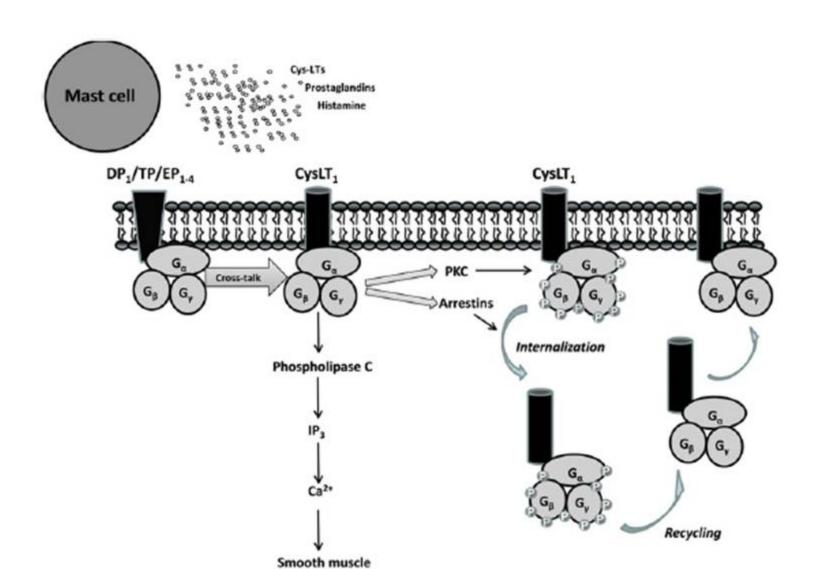












contraction