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1 **Title**

2 Characterization of lignin-derived products from various lignocellulosics as treated by
3 semi-flow hot-compressed water

4

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21

22 **Abstract**

23

24 To elucidate the decomposition behaviors of lignin from the different taxonomic groups,
25 five different lignocellulosics were treated by hot-compressed water (230°C/10MPa/15
26 min) to fractionate the water-soluble portion, precipitates and insoluble residues. The
27 lignin-derived products in each fraction were, then, characterized and compared. As a
28 result, the delignification from monocotyledons such as nipa palm (*Nypa fruticans*) frond,
29 rice (*Oryza sativa*) straw and corn (*Zea mays*) cob were more extensive compared to that
30 from woods such as Japanese cedar (*Cryptomeria japonica*) and Japanese beech (*Fagus*
31 *crenata*), being gymnosperm and dicotyledon in angiosperm, respectively. The water-
32 soluble portion contained lignin monomers like coniferyl alcohol and phenolic acids,
33 while the precipitates were found to consist of higher molecular weight lignin with high
34 ether type linkages. In contrast, the lignin in the insoluble residues was rich in condensed
35 type structures. Therefore, in all five lignocellulosics, ether type linkages were
36 preferentially cleaved and condensed type lignin had resistance to hot-compressed water.
37 Furthermore, regarding monocotyledons, the lignin-carbohydrate complexes were
38 cleaved and the higher molecular weight lignin were eluted compared to woods. These
39 differences would facilitate the delignification in monocotyledons. Such information
40 provides a clue as to efficient utilization of various lignocellulosics.

41

42 (193 words)

43

44 1. Introduction

45

46 Due to the depletion of fossil fuel and the environmental issues of global warming, the
47 utilization of lignocellulosics as renewable resources has been examined towards the
48 biofuel and biochemical production. The cell wall of lignocellulosics consists mainly of
49 cellulose, hemicelluloses and lignin, and these components enforce a sophisticated
50 structure, having a resistance to water and organic solvents.

51 For the conversion of the lignocellulosics into value-added chemicals, a variety of
52 different decomposition methods of lignocellulosics have been explored. Hydrothermal
53 treatments have a high hydrolysis ability and dissolve even the hydrophobic lignin
54 because of the highly ionic products and low dielectric constant of water [1, 2]. Thus,
55 various kinds of hydrothermal treatments of lignocellulosics have been explored [3, 4],
56 and among hydrothermal treatments, a hot-compressed water treatment was conducted
57 under relatively milder condition, but its potential for lignocellulosics decomposition was
58 still quite high [5]. Such a hot-compressed water treatment can be categorized into batch,
59 flow and semi-flow types. Among them, semi-flow system can minimize degradations
60 because of the rapid removal of the degraded products from the reaction vessel [6, 7].

61 With such an advantage, our research group developed a semi-flow hot-
62 compressed water treatment, and achieved the high yields of saccharides from cellulose
63 and hemicelluloses [8–10]. With respect to the lignin, its decomposition behaviors of
64 woods such as Japanese cedar and Japanese beech were discussed through the
65 characterization of their lignin-derived products as treated by hot-compressed water [11,

66 12].

67 In contrast, non-woody lignocellulosics such as palm, rice and corn are also
68 expected as renewable resources and they are categorized as monocotyledons in
69 angiosperms, while woods such as Japanese cedar and Japanese beech as one of
70 gymnosperms and dicotyledons in angiosperms, respectively [13]. Among these
71 lignocellulosics, there are several differences existing in the lignin structures, for instance,
72 the lignin component unit structures and the association with phenolic acids [14].
73 Accordingly, their decomposition behaviors of lignin would be different one another.
74 Thus, in this study, five lignocellulosics, which are categorized into different taxonomical
75 groups, were treated with semi-flow hot-compressed water (230°C/10MPa). To facilitate
76 comparisons on the lignin decomposition behaviors, the characterization of the lignin-
77 derived products were conducted and compared among five lignocellulosics.

78

79

80 **2. Materials and methods**

81

82 **2.1 Samples and chemicals**

83

84 The sapwood of Japanese cedar (*Cryptomeria japonica*) as one of gymnosperms, the
85 sapwood of Japanese beech (*Fagus crenata*) as one of dicotyledons in angiosperms, the
86 nipa palm (*Nypa fruticans*) frond, the rice (*Oryza sativa*) straw and the corn (*Zea mays*)
87 cob as monocotyledons in angiosperms were selected in this study. Flour of each
88 lignocellulosics passing through an 18-mesh screen was extracted with acetone using a
89 Soxhlet apparatus and dried at 105°C for 24 h before the experiments. All chemicals used
90 in this study were of reagent grade without any purification.

91

92

93 **2.2 Characterization of lignocellulosics**

94

95 The chemical composition in each lignocellulosics was evaluated using the method of
96 Rabemanolontsoa et al. [15].

97 The contents of the phenolic acids, which are attached to lignin in
98 monocotyledons, were determined by the alkali extraction with 0.5M NaOH aqueous
99 solution [16]. The extracted portion was acidified and extracted with ethyl acetate. The
100 ethyl acetate-soluble portion was then dehydrated and evaporated under vacuum. The
101 obtained products were trimethylsilyl derivatized followed by gas chromatography-mass

102 spectrometry (GC-MS: GCMS-QP2010 Ultra, Shimadzu Co., Kyoto, Japan) analysis.
103 The capillary column was a CP Sil 8CB (Agilent Technology Inc., California, United
104 States). The temperature program was 1.0 min at 40°C, 5.0°C/min to 300°C, and 8.0 min
105 at 300°C. Helium carrier gas was used at a flow-rate of 1.5 mL/min. Injector and detector
106 temperatures were both 230°C.

107 For the analysis of lignin structure, the alkaline nitrobenzene oxidation was
108 performed according to the ordinary method [17] and the total yields of vanillin,
109 syringaldehyde, and *p*-hydroxybenzaldehyde were determined by GC (GC 2014,
110 Shimadzu Co., Kyoto, Japan). The vanillin can be produced from ferulic acid, and the *p*-
111 hydroxybenzaldehyde can be from *p*-coumaric acid and 4-hydroxybenzoic acid. Thus, the
112 yields of phenolic acids-derived vanillin and *p*-hydroxybenzaldehyde were subtracted
113 from the original yields of the alkaline nitrobenzene oxidation products to obtain the
114 actual yields of products only from the core lignin.

115

116

117 2.3 Hot-compressed water treatment

118

119 Approximately 0.5 g of oven-dried extractive-free lignocellulosics was placed in the
120 reaction vessel for which the hot-compressed water treatment was performed as described
121 in a previous paper [18]. The treatment condition was 230°C/10 MPa for 15 min, in which
122 most of the hemicelluloses and part of lignin can be decomposed [18].

123 After the treatment, the hot-compressed water-insoluble residue left over in the

124 reaction vessel was separated from the hot-compressed water-soluble portion, and dried
125 at 105°C for 12 h to measure its oven-dried weight. The separated hot-compressed water-
126 soluble portion was kept standing for 12 h under ambient condition. During this period,
127 it was separated to be water-soluble portion and precipitates which were retrieved by
128 filtration with a Millipore membrane filter (pore size: 0.45 μm). For the water-soluble
129 portion, the lignin-derived products were obtained by extraction with ethyl acetate.

130

131

132 2.4 Analytical methods of lignin-derived products

133

134 The lignin-derived products in three fractions, water-soluble portion, precipitates and
135 insoluble residue, were characterized. The content of lignin-derived products in each
136 fraction was examined as the combined yields of Klason lignin and acid-soluble lignin
137 [19]. Molecular weight distribution of lignin-derived products in the water-soluble
138 portion and precipitates was evaluated by gel permeation chromatography (GPC) analysis,
139 which was performed with LC-10A (Shimadzu, Kyoto, Japan) as described in the
140 previous study [12]. Before the GPC analysis, the precipitates were acetylated [20]. The
141 alkaline nitrobenzene oxidation analysis was conducted for the precipitates and insoluble
142 residues by means of the same method described above. Monomeric lignin-derived
143 compounds in water-soluble portion were determined and quantified by GC-MS analysis,
144 which was performed with same equipment and same methods as described above.

145

146 3. Results and discussion

147

148 3.1 Characterization of lignocellulosics

149

150 Fig. 1 shows the chemical composition of five different lignocellulosics used in this study.

151 As for Japanese cedar, one of the gymnosperm softwoods, the cellulose content is the

152 highest, and the lignin is secondly the highest to be 32.0wt%, followed by hemicellulose

153 with others such as protein, extractives and ash. For Japanese beech, one of the hardwoods

154 (dicotyledon in angiosperms), the chemical composition is relatively similar to that of

155 Japanese cedar, however the lignin is relatively lower in content. In contrast, regarding

156 nipa frond, rice straw and corn cob, monocotyledons in angiosperms, the lignin is lower

157 to be around 18~20wt%, with the protein and ash being much higher compared with

158 woods (Japanese cedar and Japanese beech).

159 The contents of phenolic acids, which are attached to lignin in monocotyledons,

160 were determined as given in Table 1. For the Japanese cedar and Japanese beech, phenolic

161 acids were not detected. In contrast, regarding the monocotyledons, the ferulic acid, *p*-

162 coumaric acid and *p*-hydroxybenzoic acid were obtained. The ferulic acid is known to

163 connect hemicelluloses with lignin, known as lignin-carbohydrate complex (LCC) in the

164 cell wall of monocotyledons [21]. In particular, the corn cob contained the highest amount

165 of ferulic acid to be 0.77wt% on lignocellulosics-basis (= 4.3wt% on lignin-basis). For

166 the *p*-coumaric acid, it was reported that *p*-coumaric acid is associated to the lignin of

167 monocotyledons [22]. In a similar manner to the ferulic acid, the corn cob contained the

168 highest amount of *p*-coumaric acid to be 1.5wt% on lignocellulosics-basis (= 8.3wt% on
 169 lignin-basis). Furthermore, the nipa frond contained a high amount of *p*-hydroxybenzoic
 170 acid, which was 0.78wt% on lignocellulosics-basis (= 4.0wt% on lignin-basis). In the
 171 palm species, it is reported that the *p*-hydroxybenzoic acid is associated with lignin [23].

172 In addition, to examine the lignin component units in these lignocellulosics, the
 173 alkaline nitrobenzene oxidation was conducted. As the decomposed products, vanillin
 174 (VA) derived from the guaiacyl (G) lignin, syringaldehyde (SA) from the syringyl (S)
 175 lignin and *p*-hydroxybenzaldehyde (HA) from the *p*-hydroxyphenyl (H) lignin were
 176 obtained. Although the detailed yields of each product will be discussed later in Fig. 5,
 177 the molar ratio of decomposed products to VA is shown in Table 1.

178 For Japanese cedar, VA was the main product with a slight amount of HA,
 179 indicating, as well known, that lignin in Japanese cedar is mainly composed of G lignin
 180 with a small amount of H lignin. In case of Japanese beech, on the other hand, both VA
 181 and SA were resulted and the ratio of SA to VA was quite high. Regarding three
 182 monocotyledons, all of VA, SA and HA were obtained. The nipa frond had a high ratio of
 183 SA to VA and a quite low of HA to VA. In contrast, the rice straw and corn cob had a high
 184 ratio of HA to VA.

185

186

187 3.2 Yields of lignin-derived products in each fraction

188

189 Fig. 2 shows the yields of the water-soluble portion, the precipitates and the insoluble

190 residue for five different lignocellulosics as treated by semi-flow hot-compressed water.
191 After treatment, the hot-compressed water-soluble portion was kept standing for 12 h
192 under ambient condition, during which it was separated to be the water-soluble portion
193 and the precipitates. In case of Japanese cedar and Japanese beech, the yields of insoluble
194 residues are relatively higher and the yields of hot-compressed water-soluble portion
195 (water-soluble portion + precipitates) are low. In contrast, from monocotyledons, around
196 70wt% was decomposed and solubilized into hot-compressed water. The precipitates
197 obtained were 5~10wt% on lignocellulosics-basis for all five lignocellulosics.

198 Fig. 3 shows the yields of lignin-derived products in water-soluble portion,
199 precipitates and insoluble residue from five lignocellulosics as treated by hot-compressed
200 water. For Japanese cedar, 43wt% of lignin was decomposed and eluted in hot-
201 compressed water with more than half of lignin remaining as insoluble residue. For
202 Japanese beech, more than half of lignin was decomposed and eluted in hot-compressed
203 water, indicating that the delignification is preferentially occurred from Japanese beech
204 compared to Japanese cedar. In contrast, the delignification from monocotyledons was
205 much more preferentially occurred to be around 80wt%. Significant differences were
206 observed between the delignification from woods (Japanese cedar and Japanese beech)
207 and monocotyledons. Among the hot-compressed water-soluble lignin, 20~30wt% of
208 lignin was retrieved as the precipitated lignin.

209

210

211 3.3 Structural analysis of lignin-derived products

212

213 Fig. 4 demonstrates GPC chromatograms of water-soluble portion and precipitates from
214 five lignocellulosics, detected at a wavelength of 280 nm. In order to evaluate the
215 molecular weight distribution of their lignin-derived products, the elution times of
216 polystyrene standards (MW: 162, 580, 1270, 5000) were displayed. The higher molecular
217 weight standards show the shorter elution time in GPC chromatograms.

218 As a result, the lignin-derived products in water-soluble portion contain
219 monomeric and oligomeric products up to the molecular weight of 1,000~2,000. In
220 contrast, the molecular weight of the lignin in the precipitates are higher than those in
221 water-soluble portion for all lignocellulosics. Among five different lignocellulosics, the
222 lignin-derived products in the precipitates from monocotyledons start to elute 2~3 min
223 faster than those from woods, indicating that the lignin-derived products from
224 monocotyledons are composed of higher molecular weight products than those from
225 woods.

226 In order to compare the relative proportion of ether linkages of lignin-derived
227 products, the alkaline nitrobenzene oxidation was conducted, to achieve its yields as
228 shown in Fig. 5. In case of Japanese cedar, the yields of vanillin (VA) from lignin in the
229 precipitates are higher than those from original wood flour. This result indicates that the
230 lignin in the precipitates maintains ether linkages. For the insoluble residue, the yield of
231 VA is relatively low, thus the lignin in the insoluble residue is rich in condensed type. For
232 Japanese beech, both VA and syringaldehyde (SA) are obtained. The total yields from
233 precipitates are quite high and those from insoluble residue are low in a similar manner

234 to Japanese cedar. Regarding the SA/VA ratio, the precipitates (2.6) is higher than the
235 original wood flour (2.2) and the insoluble residue is much lower (1.3). This could indicate
236 the precipitated lignin is rich in S lignin, but the residual lignin is in G lignin. As for the
237 monocotyledons, all of VA, SA and *p*-hydroxybenzaldehyde (HA) are obtained from all
238 fractions. The total yields from precipitates are almost similar to those from the original
239 wood flour, while the total yields from insoluble residues are lower as in woods.
240 Regarding the ratio of VA, SA and HA, the main products from the precipitates are SA
241 and the yields of VA and HA are relatively low. In contrast, as to the insoluble residues,
242 the ratio of VA and HA are relatively high.

243 Consequently, for all lignocellulosics, the lignin-derived products in precipitates
244 maintained the ether type linkages, whereas those in the insoluble residue were rich in
245 condensed type lignin. Except for Japanese cedar, being composed of only G lignin, the
246 lignin component units of precipitates contained more methoxyl group compared to those
247 of insoluble residues.

248

249

250 3.4 Lignin-derived products in water-soluble portion

251

252 From the GPC analysis, it was found that the lignin-derived products in water-soluble
253 portion contained monomers (Fig. 4). Thus, GC-MS analysis was performed to identify
254 the lignin-derived monomers in the water-soluble portion, and the yields of main products
255 was summarized in Table 2. These compounds were identified and quantified by

256 comparing the retention time and the mass fragmentation patterns with their model
257 compounds. As a result, the coniferyl alcohol, sinapyl alcohol and *p*-coumaryl alcohol,
258 which are the monomer precursors of lignification, were the main monomeric products.
259 The obtained monomers corresponded well to the lignin component units detected by the
260 alkaline nitrobenzene oxidation (Table 1). As for the woods, vanillin and syringaldehyde
261 were also obtained, while those aldehydes were not detected from monocotyledons.

262 In the water-soluble portion from monocotyledons, the phenolic acids were
263 obtained. Given that these phenolic acids were not obtained from woods, the obtained
264 acids would not be derived from the core lignin but from the phenolic acids attached to
265 lignin. In particular, the *p*-coumaric acid and *p*-hydroxybenzoic acid were obtained with
266 quite high yields.

267 With respect to the *p*-coumaric acid in corn cob, it was reported by 2D-NMR
268 analysis for the corn stover that the *p*-coumaric acid is attached to the lignin at γ position
269 of propane side-chain [22]. Although the detailed structures of *p*-coumaric acid in the
270 corn cob are not clarified yet, it seems to be quite possible that the high yield of *p*-
271 coumaric acid would be attached to the end position of lignin. Such a linkage would be
272 readily cleaved and resulted in a quite high yield.

273 With regards to the *p*-hydroxybenzoic acid, it is reported to be attached to the
274 lignin at γ position of its sidechain for the frond of oil palm (*Elaeis guineensis*) [23].
275 Therefore, in case of nipa palm frond, it seems quite possible that *p*-hydroxybenzoic acid
276 would attach to the end position of lignin.

277 The ferulic acid was also found from monocotyledons. The majority of the ferulic

278 acid is known to be both esterified and etherified to lignin and hemicellulose [24, 25], to
279 form the lignin-carbohydrates complex (LCC) [21]. Thus, it would be probable that
280 ferulic acid obtained in the water-soluble portion would be from LCC linkages cleaved
281 during hot-compressed water treatment.

282

283

284 3.5 Lignin-derived products in precipitates

285

286 For all five different lignocellulosics, the lignin-derived products in the precipitates are
287 relatively higher in molecular weights, and their structures rich in ether linkages from the
288 analysis of the alkaline nitrobenzene oxidation. Regarding the precipitated lignin from
289 Japanese beech, their structures were quite similar to the milled wood lignin, considered
290 to be alike native lignin. Thus, the precipitated lignin seems to maintain the original lignin
291 structures [18]. Yamauchi et al. speculated for its reason that some micropores are
292 produced in the cell wall because of the decomposition of hemicelluloses and the cleavage
293 of lignin ether linkages at the beginning of hot-compressed water treatments. The
294 produced micropores would have then facilitated the elution of the lignin cluster, in which
295 ether type linkages are maintained to some extent [11]. Hydrophobic lignin could be
296 eluted in hot-compressed water due to the low dielectric constant of hot-compressed water,
297 whereas some large molecules cluster precipitated after hot-compressed water returned to
298 the ordinary water under ambient condition. The similar phenomena would be occurred
299 in the monocotyledons, because the lignin-derived products in the precipitates from

300 monocotyledons were composed of products with relatively higher molecular weight than
301 those from woods.

302 The ferulic acid contents of precipitates were quite low to be less than 0.1wt% on
303 precipitates basis. This indicates that the LCC structures in the cell walls would be mostly
304 cleaved during the hot-compressed water treatment. Under this condition, both
305 hemicelluloses and lignin in the cell wall were decomposed and eluted out from the cell
306 wall into hot-compressed water [8, 9]. To verify this behavior, further experiments on
307 their LCC should be performed.

308

309

310 3.6 Lignin-derived products in insoluble residue

311

312 About half of lignin from woods and about 20wt% of lignin from monocotyledons were
313 recovered as the insoluble residue. For all five different lignocellulosics, the lignin-
314 derived products were rich in condensed type lignin. Given that the ether type linkages
315 are easily cleaved by hot-compressed water treatment, but that the condensed type
316 linkages are difficult to be cleaved [26], the condensed lignin had a resistance to hot-
317 compressed water and the insoluble residues were rich in condensed type linkages.
318 Furthermore, for all lignocellulosics, the lignin in the insoluble residues were composed
319 of lower methoxyl components units compared to that in the precipitates.

320 Considering that the phenylpropane units with lower methoxyl content have
321 higher possibility to form condensed type linkages at 3 or 5 position of aromatic ring,

322 these results are reasonable. Thus, in terms of the component units, H lignin is thought to
323 be the most tolerant structure. However, the corn cob and the rice straw, which are
324 composed of high ratio of H lignin (Table 1), showed the extensive delignification
325 compared to other lignocellulosics, indicating that the decomposition of lignin would be
326 affected not only by the lignin component units but also by other lignin structures like
327 LCCs as described above.

328

329

330 **4. Concluding remarks**

331

332 Various lignocellulosics, belonging to the different taxonomic groups, were treated with
333 hot-compressed water (230°C/10MPa/15 min), and the obtained lignin-derived products
334 in each fraction were characterized. As a result, the delignification from monocotyledons
335 such as nipa palm frond, rice straw and corn cob was more extensive than that from woods
336 such as Japanese cedar and Japanese beech. For all lignocellulosics, the lignin of insoluble
337 residues were rich in condensed linkages, indicating that the decomposition of lignin due
338 to the cleavage of ether linkages. The residual lignin consisted of lower methoxyl content
339 in phenylpropane units, compared to those in the precipitates. Considering that the lignin
340 with lower methoxyl content can readily have condensed type linkages at 3 and/or 5
341 position of aromatic ring, the lower methoxyl content units reveal resistance to hot-
342 compressed water, remaining as insoluble residue. It is interesting, however, that the corn
343 cob and rice straw, which consisted of high ratio of H lignin, showed the extensive

344 delignification compared to other lignocellulosics. Regarding the lignin-derive products
345 from monocotyledons, the LCC were cleaved and higher molecular weight lignin was
346 eluted into hot-compressed water, compared to that from the woods. These differences
347 would have facilitated the delignification from monocotyledons. Based on these lines of
348 evidence, it was found that there are several differences on the delignification and the
349 decomposition behaviors of lignin between the woods (gymnosperm softwood and
350 angiosperm dicotyledonous hardwood) and monocotyledons as treated by hot-
351 compressed water.

352

353

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355

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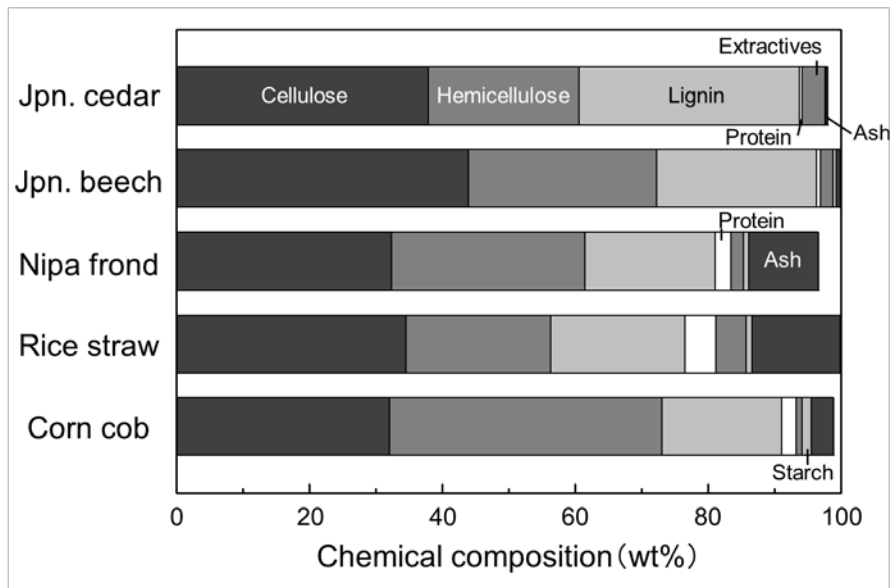
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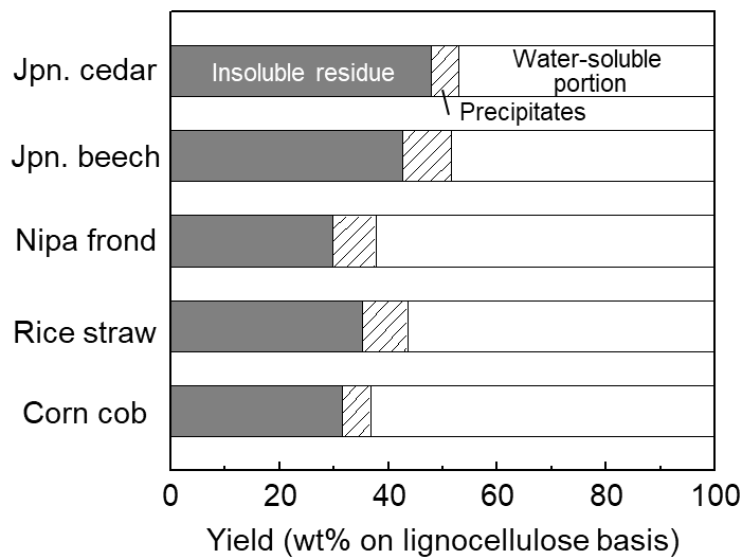
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Fig. 1 Chemical composition of lignocellulosics as determined by the method [13]

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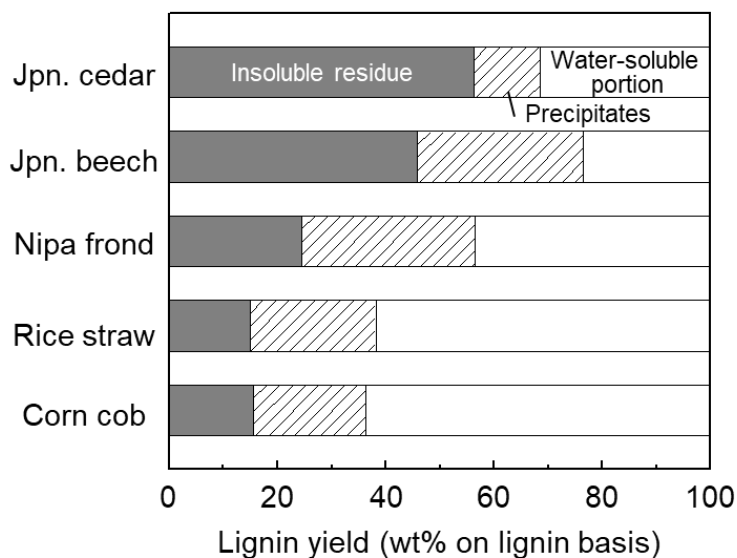
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Fig. 2 Yields of water-soluble portion, precipitates and insoluble residue from five different

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lignocellulosics as treated by semi-flow hot-compressed water

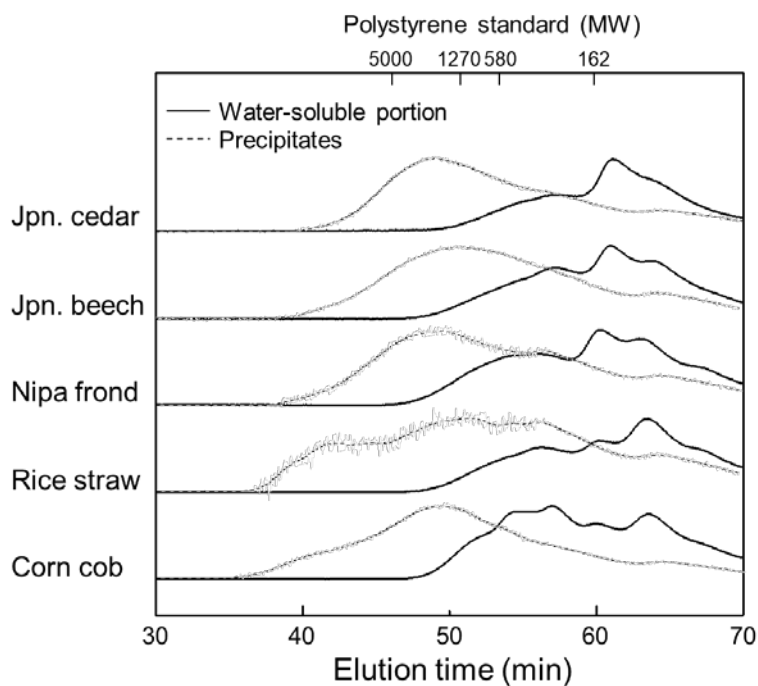
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436 Fig. 3 Yields of lignin in water-soluble portion, precipitates and insoluble residue from five
437 different lignocellulosics as treated by semi-flow hot-compressed water

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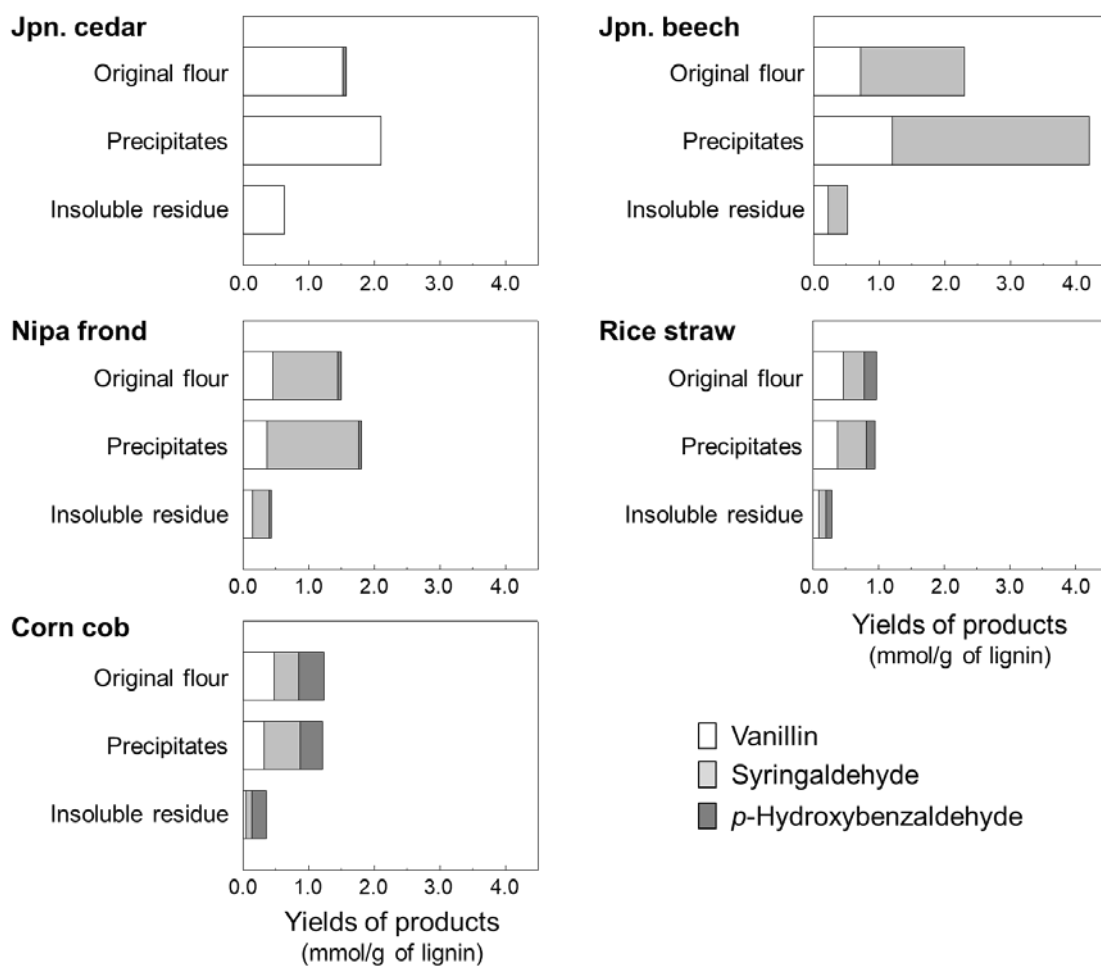
440 Fig. 4 GPC chromatographs of lignin-derived products in water-soluble portion and precipitates
441 from five different lignocellulosics as treated by hot-compressed water

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447 Fig. 5 Yields of alkaline nitrobenzene oxidation products of original flour, precipitates and

448 insoluble residue from five different lignocellulosics as treated by hot-compressed water

449 Table 1 Lignin content, phenolic acids content and alkaline nitrobenzene oxidation yields from five different lignocellulosics

Lignocellulosics	Lignin content (wt%)	Phenolic acids (wt%)			Nitrobenzene oxidation yields (Molar ratio to vanillin)		
		Ferulic acid	<i>p</i> -Coumaric acid	<i>p</i> -Hydroxy benzoic acid	VA*	SA*	HA*
Jpn. Cedar	32.0	-	-	-	1	0	0.05
Jpn. Beech	24.0	-	-	-	1	2.19	0
Nipa frond	19.6	0.13	0.03	0.78	1	2.15	0.10
Rice straw	20.2	0.55	0.79	-	1	0.67	0.41
Corn cob	18.0	0.77	1.50	-	1	0.80	0.79

 450 * VA: Vanillin, SA: Syringaldehyde, HA: *p*-Hydroxybenzaldehyde

451 Table 2 Monomeric compounds in water-soluble portion from five different lignocellulosics as treated by semi-flow hot-compressed water

Lignocellulosics	Lignin-derived monomers (wt%)				Phenolic acids (wt%)		
	Vanillin+ Syringaldehyde	Coniferyl alcohol	Sinapyl alcohol	<i>p</i> -Coumaryl alcohol	Ferulic acid	<i>p</i> -Coumaric acid	<i>p</i> -Hydroxy benzoic acid
Jpn. Cedar	0.06	0.81	-	-	0	-	-
Jpn. Beech	0.20	0.20	0.80	-	0	-	-
Nipa frond	-	0.13	0.26	0	0.01	0.05	0.40
Rice straw	-	0.19	0.01	0.010	0.06	0.18	-
Corn cob	-	0.05	0.14	0.003	0.02	0.52	-

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