1	Biomass and primary production of a 8-11 m depth meadow $versus < 3$ m depth
2	meadows of the seagrass Cymodocea nodosa (Ucria) Ascherson
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18 Abstract

20	Current knowledge about the abundance, growth, and primary production of the
21	seagrass <i>Cymodocea nodosa</i> (Ucria) Ascherson is biased towards shallow (depth $< 3 \text{ m}$ )
22	meadows although this species also forms extensive meadows at larger depths along the
23	coastlines. The biomass and primary production of a C. nodosa meadow located at a
24	depth of 8-11 m was estimated at the time of maximum annual vegetative development
25	(summer) using reconstruction techniques, and compared with those available from
26	shallow meadows of this species. A depth-referenced data base of values at the time of
27	maximum annual development was compiled to that end. The vegetative development
28	of <i>C. nodosa</i> at 8-11 m depth was not different from that achieved by shallow (depth <
29	3 m) meadows of this species. Only shoot density, which decreased from 1637 shoots
30	$m^{-2}$ to 605 shoots $m^{-2}$ , and the annual rate of elongation of the horizontal rhizome,
31	which increased from 23 cm apex <sup>-1</sup> year <sup>-1</sup> to 71 cm apex <sup>-1</sup> year <sup>-1</sup> , were different as depth
32	increased from $< 3$ m to 8-11 m. Depth was a poor predictor of the vegetative
33	development and primary production of C. nodosa. The biomass of rhizomes and roots
34	decreased with depth (g D.W. m <sup>-2</sup> = 480 ( $\pm$ 53, SE) - 32 ( $\pm$ 15, SE) depth (in m), R <sup>2</sup> =
35	0.12, $F = 4.65$ , d.f. = 35, $P = 0.0381$ ) which made total biomass of the meadow to show a
36	trend of decrease with depth but the variance of biomass data explained by depth was
37	low. The annual rate of elongation of the horizontal rhizome showed a significant
38	positive relationship with depth (cm apex <sup>-1</sup> year <sup>-1</sup> = $18 (\pm 5.1, SE) + 5.0 (\pm 1.33, SE)$
39	depth (in m); $R^2 = 0.50$ , F= 14.07, d.f. = 14, P = 0.0021). As shoot size and growth did
40	not change significantly with depth, the reduction of shoot density should drive any
41	changes of biomass and productivity of C. nodosa as depth increases. The processes by
42	which this reduction of <i>C. nodosa</i> abundance with depth occur remain to be elucidated.

- 43
- 44 Keywords: Seagrasses, Cymodocea nodosa, depth gradient, biomass, shoot density,
- 45 primary production, Mediterranean Sea

46 1. Introduction

47

48	Light availability sets the maximum depth of seagrass colonization (Duarte,
49	1991). Shoot density of seagrass meadows decreases as depth increases (Romero, 1989;
50	West, 1990; Romero et al., 1998; Krause-Jensen et al., 2000) while seagrass biomass
51	usually reaches a maximum at intermediate depths and decreases towards the depth
52	limit (Duarte, 1991; Krause-Jensen et al., 2000). Strong correlations between light
53	attenuation and the decline of shoot density and seagrass biomass with depth (Duarte,
54	1991; Krause-Jensen et al., 2000) suggest that light availability is the main factor
55	driving the depth distribution of seagrasses. In situ manipulation of light availability and
56	transplant experiments have shown that the leaf growth and biomass of the seagrass
57	Zostera marina L. at the deep limit of distribution is dependent on light availability
58	(Dennison and Alberte, 1985; Dennison and Alberte, 1986). The decline of shoot
59	density with depth is considered a plant response to reduce self-shading when light
60	availability is low (Olesen and Sand-Jensen, 1993;Krause-Jensen et al., 2000).
61	Additionally, the size of Z. marina shoots increases with depth, and more biomass is
62	allocated to leaves than to rhizomes and roots (Krause-Jensen et al., 2000). Z. marina
63	plants grown under an experimental light gradient showed that rhizome growth declined
64	faster than leaf growth as light was reduced (Olesen and Sand-Jensen, 1993). These
65	changes of biomass allocation are interpreted as a plant response to reduce respiration
66	costs and maintain growth under low light availability (Olesen and Sand-Jensen, 1993;
67	Krause-Jensen et al., 2000).
68	

69 Shoot density, leaf biomass, and productivity of the Mediterranean seagrass
70 *Posidonia oceanica* (L.) Delile decrease from the shallow to the deep limit of

distribution of this species (Pirc, 1984;Romero, 1989; Romero et al., 1998; Olesen et al.,
2002; Gobert et al., 2003). Shoot size has been shown to remain fairly constant (Pirc,
1984; Olesen et al., 2002) or decrease (Gobert et al., 2003) as depth increases, but the
number of leaves per shoot and the growth rate of individual shoots do not change with
depth (Pirc, 1984; Romero, 1989; Olesen et al., 2002; Gobert et al., 2003). Hence, the
decline of leaf biomass and productivity of *P. oceanica* meadows with depth seems to
be driven by the decline of shoot density mainly.

78

79 *Cymodocea nodosa* (Ucria) Ascherson is a common seagrass species in the 80 Mediterranean Sea and the North-Atlantic coast of Africa, including the Canary Islands 81 (den Hartog, 1970), which shows an ample depth distribution from the intertidal 82 (Vermaat et al., 1993) to depths of 33-35 m (Drew, 1978; Reyes et al., 1995a; Canals 83 and Ballesteros, 1997). It can be found on a wide range of substrata too, from coarse 84 sand to muddy sediments (Peduzzi and Vukovič, 1990; Pavón-Salas et al., 2000), and 85 forms extensive meadows in shallow, sheltered places such as lagoons (Terrados and Ros, 1992; Ribera et al., 1997; Agostini et al., 2003), bays (Pérez and Camp, 1986; 86 87 Pérez and Romero, 1994) or harbor areas (Reyes et al., 1995b). Most part of the 88 knowledge about the magnitude and seasonality of biomass, growth and primary 89 production of C. nodosa has been produced in shallow (depth < 3 m) meadows (Caye 90 and Meinesz, 1985; Pérez et al., 1991; van Lent et al., 1991; Terrados and Ros, 1992; 91 Vermaat et al., 1993; Pérez and Romero, 1994; Pérez et al., 1994; Sfriso and Ghetti, 92 1998; Cancemi et al., 2002; Guidetti et al., 2002; Agostini et al., 2003). Logistical 93 reasons (i.e., shelter from wave action, diving time) might be behind this bias in 94 knowledge for C. nodosa also forms extensive meadows at larger depths along the 95 coastlines (Reyes et al., 1995a; Calvín et al., 1999).

07	The form analysis of the regestering development of Crimede each nodes a plana
97	The few analysis of the vegetative development of Cymodocea houosa along
98	depth gradients (from 2 to 6 m: Peduzzi and Vukovič, 1990; from 0.4 to 3.8 m: Olesen
99	et al., 2002) show that shoot density and leaf biomass decrease with depth, while the
100	rhizome plus roots to shoot biomass ratio either decreases or does not change, and shoot
101	size remains fairly constant.
102	
103	In this study we estimated plant size, shoot density, biomass and primary
104	production of Cymodocea nodosa growing at depths of 8 m and 11 m, a common
105	location of the meadows formed by this seagrass species in the Mediterranean, and
106	compared the results obtained with those available from shallow (depth $< 3m$ ) meadows
107	of this species to evaluate if the vegetative development of deep C. nodosa meadows is
108	different from that of shallow meadows. By increasing the depth range from which
109	knowledge about the vegetative development of this species is available we could
110	evaluate if depth-related changes of plant size, shoot density, biomass and primary
111	production of C. nodosa were consistent with those observed in other seagrass species.
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114	2. Methods
115	
116	The study was performed at Sant Pol beach, Sant Feliu de Guixols, NE Spain
117	(41° 47.227'N, 3° 03.206'E), where Cymodocea nodosa forms a spatially homogeneous
118	meadow (100 % cover of the substratum) between depths of 7 m and 11 m. The
119	meadow is patchy at a depth of 18.5 m, and isolated plants can be found at a depth of 21

120 m. C. nodosa is not present at depths < 7m. The main sampling station was located at a

121	depth of 11 m with additional samples collected at a depth of 8 m. Sampling was
122	performed in July 2002, when the vegetative development of C. nodosa is near its
123	annual maximum (Terrados and Ros, 1992; Pérez and Romero, 1994; Cebrián et al.,
124	1997; Marbà et al., 1996; Rismondo et al., 1997; Sfriso and Ghetti, 1998; Guidetti et al.,
125	2002; Agostini et al., 2003). The vertical attenuation coefficient for downward
126	irradiance $(K_d)$ was estimated by measuring scalar irradiance at noon just below the
127	surface of the sea and at the top of the leaf canopy at the depth of 11 m with a Li-Cor
128	spherical quantum sensor LI-193SB (Kirk, 1983; page 95). K <sub>d</sub> at Sant Pol beach in July
129	2002 was 0.095 m <sup>-1</sup> , a relatively low value for coastal waters (cf. Kirk, 1983; page 112).
130	
131	The abundance of Cymodocea nodosa was quantified as shoot density (number
132	of shoots m <sup>-2</sup> ) and biomass (g dry weight m <sup>-2</sup> ). Shoot and flower densities were
133	estimated by counting the number of shoots and flowers present in 25 samples collected
134	using a 10.2 cm in-diameter stainless-steel corer that was inserted in the sediment to a
135	depth of 40 cm. The biomass of C. nodosa in 10 of the corer samples was sorted into
136	leaves, rhizomes, roots, and flowers, and dried at 60°C during 48 hours to estimate dry
137	weight. Nitrogen in the leaves was determined using a Carlo-Erba NA-1500 CHN
138	analyzer, and phosphorus was determined spectrophotometrically (Parsons et al., 1984)
139	after wet oxidation with boiling $H_2SO_4$ . Seed abundance was estimated from the number
140	of seeds that were collected in the corer samples.
141	
142	Growth rates were estimated using reconstruction techniques (Duarte et al.,
143	1994) which are based on the estimation of the age of the shoots using the
144	"plastochrone" concept, the time elapsed between the appearance of two consecutive

145 structural modules (i.e., leaves) in plants (Erikson and Michelini, 1957). The age in

146 years of the shoots was estimated by counting the number of standing leaves and scars 147 left by fallen leaves on the vertical rhizome of each shoot and dividing the resultant 148 number by the average number of leaves produced by *C. nodosa* in one year. Such 149 average was estimated through the analysis of the sequence of lengths of the internodes 150 of large pieces of vertical rhizome (cf. Duarte et al., 1994) which were measured with a 151 stereomicroscope.

152

153 The length (cm) of the vertical rhizome of all the shoots collected in the corer 154 samples was measured, and the number of roots, root scars and flower scars present in 155 them was counted. These counts, when divided by the age of the shoots, provide an 156 estimate of the annual elongation of the vertical rhizome and the number of roots and 157 flowers produced by the shoots. Horizontal rhizome pieces of more than 30 cm in-158 length were collected haphazardly throughout the meadow by hand to estimate their 159 growth rate. The number of standing leaves and leaf scars (i.e. an age estimate) of all 160 the shoots in each rhizome piece collected was counted, as well as the number of 161 internodes and distance (cm) between them. The age difference between shoots in a 162 rhizome piece allowed us to calculate the average elongation and number of internodes 163 produced per leaf plastochrone. The annual rates of elongation and internode production 164 of the horizontal rhizome were estimated by multiplying those per-leaf-plastochrone 165 estimates by the average number of leaves produced during one year. A subsample of 166 10 fragments of both vertical and horizontal rhizomes was haphazardly chosen and the 167 number of internodes of each fragment was counted and its length measured before 168 drying to estimate average mass and length of vertical and horizontal rhizome 169 internodes. Similarly, three subsamples of 10 small and 10 large unbroken roots were 170 chosen and weighed after drying to estimate the average mass per root. The abundance

of apexes of horizontal rhizome was estimated by counting the number of them present
in 24 quadrats (1600 cm<sup>2</sup>) haphazardly placed within the meadow. Knowledge of the
age of each shoot allows to build the age distribution of the population of shoots (cf.
Duarte et al., 1994). Mean and median shoot age were estimated, and the resultant age
distributions were compared with those available in the literature (KolmogorovSmirnov two sample test, Sokal and Rohlf, 1981).

177

178 We searched the marine ecology an biology literature for studies describing the 179 vegetative development (plant size, shoot density, biomass, nutrient content of leaves, 180 growth, and productivity) of Cymodocea nodosa and compiled a depth-referenced data 181 base of values at the time of maximum annual development (listed in caption of Fig. 1). 182 To evaluate the differences of vegetative development between "shallow" and "deep" 183 meadows we compared the mean of the values obtained at the depths of 8 m and 11m at 184 our study site with the mean of the corresponding values reported by those compiled 185 studies which were performed at depths < 3 m (two-sample t test, two-tailed). Graphing 186 of compiled values against depth suggested that vegetative development was highly 187 variable at shallow meadows and that declines with depth of shoot density, biomass and 188 other variables describing vegetative development occurred at depths > 3 m. We 189 calculated the coefficient of variation of the mean (V\*, Sokal and Rohlf, 1981) of 190 selected variables describing the vegetative development of C. nodosa in shallow (depth 191 < 3 m) and deep (depth > 3 m) meadows to evaluate the significance of the differences 192 in variability of the vegetative development between them (two-tailed Z test for 193 difference between two coefficients of variation, Zar, 1999). Further, we used linear 194 regression to examine depth-associated changes of the vegetative development of C. 195 nodosa.

198 3. Results

200	Shoot density of Cymodocea nodosa at 8-11 m depth in Sant Pol beach in July
201	2002 was lower than the annual maximum shoot density reported in most of the
202	meadows of this species studied previously (Fig. 1a). Leaf, rhizome plus roots, and total
203	biomass of C. nodosa at the annual maximum at 8-11m depth were also at the low end
204	of the range of reported values (Fig. 1b). The annual maximum shoot density of C.
205	<i>nodosa</i> meadows at depth of 8-11 m was lower (two sample t-test, $P < 0.05$ ) than that of
206	meadows located at depths $< 3$ m (Table 1). The annual maximum of leaves, rhizome
207	plus roots, and total biomass tended to be also lower at 8-11 m depth than at depths $< 3$
208	m but the differences of the means were not significant (Table 1). The relative
209	allocation of biomass to leaves and rhizomes plus roots at 8-11 m depth was not
210	different than that of meadows located at depths $< 3$ m (Table 1, Fig. 1c).
211	
212	The average number of leaves of Cymodocea nodosa shoots at the time of
213	maximum vegetative development at 8-11 m depth was not different from that of
214	meadows located at depths $< 3$ m (Table 1, Fig. 1d). The same occurred with the
215	average shoot mass (Table 1, Fig. 1e).
216	
217	The annual average leaf plastochrone of Cymodocea nodosa at 8-11 m depth
218	was not different than that of meadows located at depths $< 3$ m (Table 1, Fig. 1f). The
219	annual leaf and total production of C. nodosa at 8-11 m depth were at low end of the
220	range of values published previously but was not different from the average values

221	estimated in meadows located at depths $< 3$ m (Table 1, Figs. 1g and 1h). The
222	elongation rate of the horizontal rhizome was, however, higher at 8-11 m depth than the
223	average value estimated in meadows located at depths $< 3 \text{ m}$ (Table 1). The nitrogen
224	and phosphorus content of C. nodosa leaves at 8-11 m depth was not different than the
225	average of values found in meadows located at depths $< 3$ m (Table 1).
226	
227	The coefficient of variation of vegetative features and production estimates of
228	Cymodocea nodosa was not different (two-tailed Z test for difference between two
229	coefficients of variation, $P > 0.05$ ) between meadows located at depths lower and higher
230	than 3 m (Table 2). Depth was a poor predictor of the vegetative development of $C$ .
231	nodosa for most of the linear regressions of the different variables used to describe the
232	vegetative development against depth were not significant neither considering the whole
233	range of data compiled nor only those data corresponding to depths $> 3$ m (Table 1).
234	Rhizome plus roots biomass decreased with depth (- $32 \pm 14.6$ (SE) g D.W. m <sup>-1</sup> , P <
235	0.05, Table 1) which made total biomass to show a marginally significant trend of
236	decrease with depth too (Fig. 1b, Table 1). The rate of elongation of the horizontal
237	rhizome increased with depth ( $P < 0.05$ , Fig. 2a).
238	
239	The average age of <i>Cymodocea nodosa</i> shoots varied between 2.5 years and 3.2
240	years while the median age varied between 2 years and 3 years in meadows located at
241	depths > 3 (Table 3), but were 1.7-1.8 years and 1 year only, respectively, in meadows

located at depths < 3 m. The comparison of the age distributions of living *C. nodosa* 

shoots obtained in this study (Fig. 3a) with those available in the literature showed that

- 244 most shoot age distributions were different (Table 3). The age distributions of dead
- shoots at 8 m and 11 m were different (Kolmogorov-Smirnov two sample test, D =0.186

246	$> D_{0.05} = 0.169$ , Fig. 3b), with average and median ages of 2.0 years and 2 years at the
247	depth of 8 m, and 1.7 years and 1 years, respectively, at the depth of 11 m.

249	The abundance of Cymodocea nodosa flowers at 8-11 m depth was not different
250	than that estimated in meadows located at depths $< 3$ m (Table 1) but likely mismatches
251	between peak flowering and sampling time make literature comparisons of flower
252	abundance difficult. The average and median age of shoots bearing a flower or flower
253	scars on the vertical rhizome were 3.4 years and 3 years at depth of 8 m, and 3.5 years
254	and 3 years, respectively, at depth of 11 m; the age distributions of these shoots were
255	not different at 8 m and 11 m (Kolmogorov-Smirnov two sample test, $D$ =0.142 $<$ $D_{0.05}$
256	= 0.290, Fig. 3c). We found flowers in shoots of only 5, 7 and 8 leaf plastochrones in
257	age. Seed abundance tended to be lower at 8-11 m depth than at meadows located at
258	depths $< 3$ m but the difference of the means was not significant (Table 1).
259	

260

261 Discussion

262

263 Our results show that the vegetative development of Cymodocea nodosa at 8-11 264 m depth is not different from that achieved by shallow (depth < 3 m) meadows of this 265 species. Only shoot density, which decreased by a factor of 2.7, and the annual rate of 266 elongation of the horizontal rhizome, which increased by a factor of 3, at 8-11 m depth 267 were different. Our results also show that depth is a poor predictor of the vegetative 268 development and primary production of C. nodosa for most of the linear regressions of 269 the variables describing the vegetative development of C. nodosa against depth were 270 non-significant.

272	The reduction of seagrass shoot density and biomass with depth has been
273	recognized before and attributed to the parallel decrease of light availability (Duarte,
274	1991). Further, the reduction of shoot density with depth has been considered the most
275	effective mechanism for seagrasses to acclimate to reductions of light availability as
276	depth increases (Olesen et al., 2002). A threshold light for rhizome branching to occur
277	has been proposed as a mechanism to explain the decrease of shoot density with depth
278	of Zostera noltii (Peralta et al., 2002) and Posidonia oceanica (Romero, 1989).
279	

280 Previous studies that have examined the vegetative development of Cymodocea 281 nodosa along depth gradients (Peduzzi and Vukovič, 1990; Olesen et al., 2002) show 282 that shoot density and leaf biomass decrease with depth. Given that the range of depths 283 examined by these studies is narrower (from 0.4 m to 6 m) than that examined by us 284 (from 0.3 m to 11 m) we expected to find stronger negative relationships of these 285 variables with depth than we did. The low predicting value that depth had to explain the 286 vegetative development and primary production of C. nodosa in our compiled data set might be due to wide spatial variation of water turbidity and light availability among the 287 sites included in it, and apparently higher  $K_d$  values at shallow sites (0.57 m<sup>-1</sup> to 0.35 m<sup>-1</sup>) 288 <sup>1</sup> in the 0.4-1.5 m depth range, cf. Duarte, 1991; Terrados and Ros, 1995; Guidetti et al., 289 2002) than at deep sites (0.19 m<sup>-1</sup> to 0.095 m<sup>-1</sup> in the 4-11 m depth range (Cancerni et 290 291 al., 2002; this study). Unfortunately, data about irradiance attenuation were not 292 available from most of the sites. Our results are consistent with previous studies on this 293 species (Peduzzi and Vukovič, 1990; Olesen et al., 2002) for neither shoot mass and the 294 number of leaves per shoot nor the average annual leaf plastochrone and productivity of 295 individual shoots changed significantly with depth. Shoot density, leaf biomass and

productivity of *Posidonia oceanica* decrease significantly with depth while shoot size
and growth remain fairly constant (Pirc, 1984; Romero, 1989; Romero et al., 1998;
Olesen et al., 2002; Gobert et al., 2003). In the case of *Zostera marina*, shoot density
decreases exponentially with depth while shoot size increases linearly; as a result
biomass peaks at intermediate depths (Krause-Jensen et al., 2000).

301

302 We found that the rhizome plus roots to shoot biomass ratio of *Cymodocea* 303 nodosa did not change with depth. However, the biomass of rhizomes plus roots 304 decreased significantly with depth driving a marginally significant reduction of total 305 biomass of the meadow which suggests that a slight change of the mass allocation 306 pattern might take place. Previous studies on this species are inconclusive for the 307 rhizome plus roots to shoot biomass ratio either did not change (Olesen et al., 2002) or 308 decreased (Peduzzi and Vukovič, 1990) as depth increased. The rhizome plus roots to 309 shoot biomass ratio of *Posidonia oceanica* does not seem to change with depth (Romero 310 et al., 1998; Olesen et al., 2002) but decreases in Zostera marina (Olesen and Sand-311 Jensen, 1993; Krause-Jensen et al., 2000). Preferential allocation of biomass to leaves as 312 depth increases should not be considered, therefore, a general response of seagrasses to 313 reduce respiration costs and maintain growth under low light availability. Further, our 314 results do not provide support to the hypothesis that the variability of seagrass 315 vegetative development is higher at shallow depths where physical disturbance and light 316 availability are high than at deep meadows where physical disturbance and light 317 availability are low (Krause-Jensen et al., 2000; Middelboe et al., 2003). 318

Linear regression of the elongation rate of the horizontal rhizome of *Cymodocea nodosa* against depth predicted a rate of increase rate of 5 cm apex<sup>-1</sup> year<sup>-1</sup> per m depth,

321 which is consistent with the results of Olesen et al. (2002). However, Peduzzi and 322 Vukovič (1990) did not find any change of the rate of elongation the horizontal rhizome 323 of *C. nodosa* between depths of 2 m and 6 m. The rate of elongation of the horizontal 324 rhizome of *Posidonia oceanica* did not change between depths of 0.7 m and 15.6 m 325 (Olesen et al., 2002). It has been suggested that the elongation of the horizontal rhizome 326 of seagrasses might depend on shoot density (Caye and Meinesz, 1985; Marbà and 327 Duarte, 1998), likely through a self-shading constraint. Shoot density, however, 328 explained little of the variability of the rate of elongation of the horizontal rhizome data included in our depth-referenced data base (Fig. 2b;  $R^2 = 0.15$ , F = 2.34, d, f. = 13, P = 0.15 329 330 0.1496).

331

332 The age distributions of Cymodocea nodosa shoots show exponential reductions 333 of shoot density since the  $\leq 1$  year class at shallow meadows (cf. Pérez, 1989; Terrados 334 and Ros, 1992) and since the 2 year class at deep meadows (Reyes et al., 1995b, and 335 this study). These results suggest that shoot turnover decreases with depth. Indeed, the 336 maximum age of shoots at 8-11 m depth was 11-12 years but only 4 to 8 years at 337 shallow meadows (Pérez, 1989; Terrados and Ros, 1992), and median shoot age was 338 lower at shallow than at deep meadows. The age distributions of dead shoots at 8-11 m 339 depth suggest, however, that most part of the shoots die within one year of being 340 produced.

341

In conclusion, the vegetative development of *Cymodo cea nodosa* at 8-11 m depth was not different from that achieved by shallow (depth < 3 m) meadows of this species. Only shoot density, which decreased by a factor of 2.7, and the annual rate of elongation of the horizontal rhizome, which increased by a factor of 3, were different as

346	depth increased from < 3 m to 8-11 m. Depth was a poor predictor of the vegetative
347	development and primary production of C. nodosa. As shoot size and growth did not
348	change significantly with depth, the reduction of shoot density should drive any changes
349	of biomass and productivity of C. nodosa as depth increases. The processes by which
350	this reduction of C. nodosa abundance with depth occur remain to be elucidated.
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352	
353	Acknowledgements
354	
355	This study was funded by the Specific Programme "Energy, Environment and
356	Sustainable Development" of the Fifth Framework Programme of the European Union
357	(contract number EVK3-CT-2000-00044, Monitoring and Managing of European
358	Seagrass Beds). We thank Jens Borum for the analysis of nitrogen and phosphorus
359	content of C. nodosa leaves, and the comments of the reviewers and Jan E. Vermaat
360	which improved the manuscript significantly.
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545 Figure captions

547	Figure 1	Shoot densit	v(a)	total hiomass	(h)	rhizome	nhis roots to	shoot ratio	(c)
J <del>4</del> /	riguit I.	. Shoot uch sh	y (a).	, what biomass	(U)	, IIIIZOIIIC	pius 100 ls il		(0),

- 548 number of standing leaves per shoot (d), and shoot mass (e) at the annual maximum of
- 549 vegetative development of *Cymodocea nodosa*, and annual average leaf plastochrone
- 550 interval (f), leaf (g), and total production (h) against depth. Triangles represent the 8-11
- m depth meadow studied. Data sources: Pirc et al., 1983; Caye and Meinesz, 1985;
- 552 Pérez and Camp, 1986; Terrados, 1986; Duarte and Sand Jensen, 1990; Peduzzi and
- 553 Vukovič, 1990; Buia and Mazzella, 1991; Terrados, 1991; Terrados and Ros, 1992;
- 554 Terrados, 1993; Terrados and Ros, 1993; Pérez and Romero, 1994; Pérez et al.,
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- 558 Cancemi et al., 2002; Guidetti et al., 2002; Olesen et al., 2002; Agostini et al., 2003;
- 559 Cunha and Duarte, 2005.
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- 561 Figure 2. Linear regression of the annual rate of horizontal rhizome elongation against
- depth (a) and shoot density (b) of *Cymodocea nodosa*. Triangles as in Fig. 1
- 563
- 564 Figure 3. Age distribution of living shoots (a), dead shoots (b), and of shoots that bore a
- flower and/or a flower scar in the vertical rhizome (c) of *Cymodocea nodosa* at the
- 566 depths of 8 and 11 m at the study site.
- 567







1 Table 1. Vegetative features and production estimates of Cymodocea nodosa at depths of 8 m and 11 m in Sant Pol beach (NE Spain),

2 corresponding values reported previously of shallow (depth < 3 m) meadows of this species (see caption Fig. 1 for data sources), and results of

3 two-sample t-tests evaluating the significance of the difference between them, and of the linear regressions against depth considering the whole

4 range of depth data compiled and only those depths > 3 m. Bold face indicates significant differences.

Variable	This study Depth: 8 m Mean ± SE	This study Depth: 11 m Mean ± SE	Shallow meadows Depth < 3 m Mean ± SE, n	t-value, d.f., P-value	Linear regression whole depth range R <sup>2</sup> , F-value, P-value	Linear regression depths > 3 m R <sup>2</sup> , F-value, P-value
Annual maximum shoot density, shoots m <sup>2</sup>	623 ± 49	588 ± 38	1637 ± 106.5, 30	2.50, 30, 0.0179	0.06, 2.29, 0.1391	0.18, 1.35, 0.2892
Leaf biomass, g DW m <sup>-2</sup>	46 ± 7.7	33±3.6	188±33.1,29	1.18, 29, 0.2457	0.03, 1.17, 0.2852	0.07, 0.58, 0.4685
Rhizome and root biomass, $g DW m^2$	$186 \pm 36.7$	$116 \pm 16.3$	$454 \pm 48.3, 28$	1.68, 28, 0.1043	0.12, 4.65, 0.0381	0.07, 0.56, 0.4794
Annual maximum total biomass,	$235 \pm 44.9$	$150 \pm 20.6$	664±81.7,25	1.64, 25, 0.1141	0.09, 4.13, 0.0504	0.08, 0.67, 0.4382
g DW m <sup>-2</sup>						
Rhizome and roots to shoot ratio	5.6 ± 2.11	$3.8 \pm 0.66$	$3.1 \pm 0.37, 28$	1.15, 28, 0.2594	0.01, 0.35, 0.5565	0.09, 0.68, 0.4369
Abundance of flowers, flowers m <sup>-2</sup>	$243\pm 60$	$153 \pm 27$	124±33.7,7	1.15, 7, 0.2876	0.03, 0.37, 0.5537	0.03, 0.10, 0.7767
Abundance of seeds, seeds m <sup>-2</sup>	$7\pm5$	$65 \pm 29$	243 ± 62.8, 7	1.80, 7, 0.1148	0.04, 0.95, 0.3389	0.18, 2.63, 0.1307
Number of leaves per shoot	$4.2 \pm 0.1$	$3.6 \pm 0.1$	$3.72 \pm 0.265, 9$	0.32, 9, 0.7539	0.01, 0.10, 0.7514	0.00, 0.00, 0.9422
Shoot mass, mg DW shoot-1	$73.8 \pm 10.2$	58.3 ± 3.1	97 ± 12.4, 25	0.71, 25, 0.4867	0.02, 0.68, 0.4156	0.48, 2.77, 0.1944
Leaf plastochrone, days	38.5	40.1	31.4±1.82, 15	1.58, 15, 0.1339	0.07, 1.52, 0.2321	0.21, 1.34, 0.2989
Elongation of horizontal rhizome, cm apex <sup>-1</sup> year <sup>-1</sup>	88.2±19.1	54.7± 8.0	23.3±3.9,12	4.40, 12, 0.0009	0.50, 14.07, 0.0021	0.37, 1.17, 0.3928
Leaf production, g DWm <sup>-2</sup> year <sup>-1</sup>	$104 \pm 114.3$	87±93.4	594±187.3, 10	1.21, 10, 0.2551	0.07, 1.23, 0.2850	0.14, 0.81, 0.4083
Total production, g DWm <sup>-2</sup> year <sup>-1</sup>	$275 \pm 192.9$	169 ± 128.3	998±432.3, 5	1.20, 5, 0.2844	0.18, 1.58, 0.2489	0.50, 2.01, 0.2919
Leaf N, % of dry weight	$2.00 \pm 0.040$	$2.06 \pm 0.045$	$1.99 \pm 0.089, 22$	0.04, 22, 0.9677	0.05, 1.31, 0.2635	0.54, 3.57, 0.1551
Leaf P, % of dry weight	$0.206 \pm 0.0016$	$0.149 \pm 0.0016$	$0.138 \pm 0.0115, 21$	1.04, 21, 0.3085	0.11, 2.82, 0.1074	0.91, 10.80, 0.1880

- 1 Table 2. Coefficient of variation of vegetative features and production estimates of
- 2 *Cymodocea nodosa* in shallow (depth < 3 m) and deep (depth > 3 m) meadows, and Z
- 3 test to evaluate the significance of the difference between them. See caption Fig. 1 for
- 4 data sources.

Variable	Depth < 3 m V* $\pm$ SE	Depth > 3 m V* ± SE	Z value Critica $ Z_{0.05} _{(2)} = 1.960$
Annual maximum shoot density, shoots m <sup>2</sup>	35±5.1	55 ± 17.4	1.430
Leaf biomass, g DW m <sup>-2</sup>	94±20.3	61 ± 17.9	0.923
Rhizome and root biomass, g DW m <sup>-2</sup>	$56 \pm 9.5$	$77 \pm 26.6$	0.891
Annual maximum total biomass, g DW m <sup>-2</sup>	61 ± 11.3	67 ± 21.3	0.194
Rhizome and roots to shoot ratio	$62 \pm 11.0$	64 ± 19.9	0.013
Abundance of flowers, flowers m <sup>-2</sup>	69±25.3	$122 \pm 65.1$	0.864
Abundance of seeds, seeds m <sup>-2</sup>	66±23.5	$219 \pm 117.7$	0.960
Number of leaves per shoot	21 ± 5.1	$43 \pm 14.5$	1.739
Shoot mass, mg DW shoot-1	63 ± 12.0	22 ± 7.2	1.507
Leaf plastochrone, days	$22\pm4.2$	$19 \pm 5.3$	0.392
Elongation of horizontal rhizome, cm apex <sup>-1</sup> year <sup>-1</sup>	58±15.1	53 ± 22.7	0.217
Leaf production, g DWm <sup>-2</sup> year <sup>-1</sup>	$97 \pm 36.2$	$79\pm30.8$	0.362
Total production, g DWm <sup>-2</sup> year <sup>-1</sup>	91±45.5	$67 \pm 31.9$	0.381
Leaf N, % of dry weight	$21 \pm 3.2$	$17 \pm 5.5$	0.562
Leaf P, % of dry weight	$38\pm 6.6$	$55\pm27.6$	0.573

Table 3. Number of shoots aged, average and median shoot age (on main diagonal) of *Cymodocea nodosa* at the study location and other meadows of this species. D values and critical  $D_{0.05}$  values (below main diagonal) of Kolmogorov Smirnov two sample tests performed to evaluate the significance of the difference between the age distribution of *C. nodosa* shoots at each meadow. Bold face indicates significant differences in shoot age distribution.

	11 m, this study	8 m, this study	6 m, Reyes et al., 1995a	1 m, Pérez, 1989	0.5 m, Terrados and Ros, 1992
11 m, this study 8 m, this study 6 m, Reyes et al., 1995a 1 m, Pérez 1989 0.5 m, Terrados and Ros, 1992	588, 2.5, 2 $\mathbf{D} = 0.141 >> D_{0.05} = 0.078$ $\mathbf{D} = 0.156 >> D_{0.05} = 0.069$ $\mathbf{D} = 0.223 >> D_{0.05} = 0.064$ $\mathbf{D} = 0.224 >> D_{0.05} = 0.065$	623, 3.2, 3 $D = 0.054 < D_{0.05} = 0.068$ $D = 0.320 >> D_{0.05} = 0.062$ $D = 0.320 >> D_{0.05} = 0.064$	1112, 3.1, 3 $\mathbf{D} = 0.347 >> D_{0.05} = 0.051$ $\mathbf{D} = 0.339 >> D_{0.05} = 0.053$	1920, 1.7, 1 $\mathbf{D} = 0.056 > D_{0.05} = 0.046$	1641, 1.8, 1