

Modelling MC1R rare variants: A structural evaluation of variants detected in a Mediterranean case-control study.

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TO THE EDITOR

Many diseases are caused by single amino acid substitutions on key genes due to loss of stability, protein misfolding and changes in the interaction properties (Goldberg, 2003; Honig and Nicholls, 1995). Protein structures can be used to evaluate the effect of missense mutations on disease associated proteins, especially on highly polymorphic genes, where functional studies are likely to be incomplete.

Considering the involvement of the highly polymorphic melanocortin-1 receptor gene (*MC1R*) in melanoma predisposition, we investigated the role of the rare variants (minor allele frequency <1%) found in this gene in the Spanish population. A structural analysis was performed by evaluating the amino acid substitutions in the context of a three-dimensional model and comparing their effects with available functional data.

The *MC1R* gene, which is located on chromosome 16q24.3 and codes for a melanocyte-stimulating hormone receptor with seven transmembrane helices, is to date, the main genetic contributor to sporadic melanoma predisposition (Kennedy *et al.*, 2001). It has already been associated to melanoma in different Caucasian populations (Fernandez *et al.*, 2007; Ibarrola-Villava *et al.*, 2010; Ibarrola-Villava *et al.*, 2012). It is highly polymorphic, with more than 100 non-conservative reported variants (Gerstenblith *et al.*, 2007). Most of these changes are relatively rare, but their frequencies vary among populations, and it appears that they are more commonly found in South European populations than in those of North European descent (Gerstenblith *et al.*, 2007; Williams *et al.*, 2011).

Of the 1170 individuals sequenced (710 non-related melanoma cases and 460 cancer-free controls), 697 (59.6%) carried at least one *MC1R* variant, with 78 (11.2%) of them actually carrying a rare variant. Overall, 6.7% (78 of 1170 individuals) of the Spanish population carry an *MC1R* rare variant. We detected a total of 41 different *MC1R* variants. Among these, 31 variants appear rarely in Spain and variant Y298H appeared for the first time in our population. We have obtained statistically significant individual associations with melanoma for six

variants: V60L, S83P, R151C, I155T, R160W and D294H (Supplementary Table S1). The highest OR was estimated for I155T and R160W (OR: 3.81; *p-value*=0.003 and OR: 3.83; *p-value*=0.001 respectively). The estimated OR associated with carrying at least one non-synonymous variant was 1.79 (*p-value* = 4.93×10^{-11}). However, the OR for carrying only one rare variant was 0.97 (*p-value* = 0.93). There are many common non-synonymous *MC1R* variants associated with a red hair colour (RHC) phenotype: D84E, R142H, R151C, I155T, R160W and D294H. The association of other common variants such as V60L, V92M and R163Q with the RHC phenotype has not been clearly established, and therefore these variants have been considered non-red hair colour (NRHC) (Wong and Rees, 2005). Almost all carriers of rare changes in the Spanish population tend to have fair skin colour. However, only 40% of them also harboured an RHC or NRHC variant. Thus, rare variants might slightly modulate skin pigmentation.

Different studies have demonstrated the functional consequences of some of these *MC1R* changes. Thus, mutations in positions 84, 142, 151, 160 and 294 have been previously reported to alter protein function (Dessinioti *et al.*, 2011; Garcia-Borron *et al.*, 2005; Newton *et al.*, 2007; Ringholm *et al.*, 2004; Sanchez-Laorden *et al.*, 2009). In addition, previous functional studies have validated some of the 3D model prediction results. Position M128 appears to have no effect according to *in silico* predictions; however, the 3D model suggests it could be a putative folding disrupter, later confirmed in a functional study (Perez Oliva *et al.*, 2009).

Due to the importance of *MC1R* variants in pigmentation and melanoma predisposition we have undertaken a comprehensive three-dimensional structure modelling of 29 non-synonymous changes detected in our population in order to further explain putative implications of rare variants (Table 1 and Figure 1). After evaluating all changes according to *MC1R* structure, variant location and nature of the amino acid changes, 25 variants seemed to have structural implications and cluster in three different regions that may explain their differences in functionality: the GTPase/PKC signalling region, an alleged α -MSH-binding extracellular region, and a novel central core protein region. The first and better-known region harbours some

of the common RHC (R142H, R151C, I155T and R160W) and NRHC (R163Q) variants. Although these variants do not seem to bring about structural changes under the 3D-model, they are located in an intracellular region thought to be a GTPase and PKC interaction area that initiates the receptor downstream signalling pathway and internalization of the receptor which confirms previously reported functionality (Dessinioti *et al.*, 2011; Garcia-Borron *et al.*, 2005; Newton *et al.*, 2007). Interestingly, side chains of residues I155, T157, R160 and R163 define a common solvent accessible area, a PKC binding domain comprising ¹⁵⁷TLPR¹⁶⁰ and ¹⁶⁰ARR¹⁶³ (Sanchez-Laorden *et al.*, 2009), and therefore, variants in these positions could explain impairment of PKC activity. Rare variants R142C, R213W and T308M are located near this region.

The second important region includes the rest of the RHC (D84E and D294H) and NRHC (V60L and V92M) variants, plus other rare variants (S83P, G89R, M128T and Y298H). Variants located in this central region may be involved in protein folding processes, and therefore may be important in the maintenance of the receptor's integrity.

Finally, we propose a third new functional region which includes S41F, P268R, T272M, K278E, N281S and A285V rare variants. All except S41F are located in the extracellular end helical tips and though there is no evident structural implication, we believe this domain may be involved in α -MSH union (Perez Oliva *et al.*, 2009).

Rare variants located in positions 41, 128, 89, 213 and 308 are described to alter protein function (Ozola *et al.*, 2013; Perez Oliva *et al.*, 2009; Sanchez-Laorden *et al.*, 2007). M128T and R213W showed markedly loss of function and displayed reduced cell surface expression and agonist binding affinity while variants S41F and G89R showed almost complete loss of function with no detectable cell surface expression and complete inability to activate the cAMP pathway. Finally, residue T308 is described to be implicated in protein phosphorylation. Therefore, the ensuing outcomes of this *MCIR* variant modelling are highly consistent with data from previously published functional *MCIR* studies.

To sum up, changes S41F, S83P, G89R, R142C, M128T, R213W, P268R, Y298H and T308M (Figure 1b, highlighted in deep blue) are predicted to have structural consequences, while variants T272M, K278E, N279K, N281S and A285V (Figure 1b highlighted in cyan) may have mild or no structural implications, but may be potentially important for protein recognition due to their near extracellular location. SNP3d *in silico* predictions represent a useful tool to evaluate amino acid change effects in a protein structure. However, they can only discriminate between damaging or benign effect. Our model combines folding effects, nature of the amino acid substitution and location of both individual variants and clusters.. The use of 3D models may constitute a robust and exploratory prediction method, as it combines folding and visualization of the amino acid changes, and therefore could be a promising tool in order to assess further functional *MC1R* variant studies. To our knowledge, this is the first time that a large number of rare *MC1R* variants are evaluated by taking into account the structural effects of the resulting residue changes.

CONFLICT OF INTEREST

The authors state no conflict of interest.

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TABLES

Table 1. *In silico* analysis of *MC1R* variants

Mutation	SNPs3d	Polyphen	Pub Funct data	3D-Model				Predict
				Possible effects of mutation	Folding	Surface exposed	Region ^a	
V60L (r)	Benign	●	F	Conserved change	No effect	Yes	Core	⊗
D84E (R)	Affect	●	F	Conserved charge larger residue	Affect	No	Core	⊗
V92M (r)	Benign	○	NF	Conserved change	No effect	No	-	⊗
R142H (R)	Affect	●	F	Cytosolic conserved	No effect	No	PKC/GTPase sig.	⊗
R151C (R)	Affect	●	F	Cytosolic non conserved	No effect	Yes	PKC/GTPase sig.	⊗
I155T (R)	Affect	●	F	Cytosolic non conserved	No effect	Yes	PKC/GTPase sig.	⊗
R160W (R)	Affect	○	F	Cytosolic conserved	No effect	Yes	PKC/GTPase sig.	⊗
R163Q (r)	Benign	○	F	Cytosolic conserved	No effect	Yes	PKC/GTPase sig.	⊗
D294H (R)	Affect	●	F	Charge change	Affect	No	Core	⊗
C35Y	Affect	●	F	Non conserved	No effect	Yes	-	⊗
S41C	Affect	○	-	Conserved change	No effect	No	MSH binding	⊗
S41F	Affect	○	F	Bulkier hydrophobic change	Affect	No	MSH binding	⊗
F45L	Affect	●	-	Conserved change	No effect	No	-	⊗
S83P	Affect	●	-	Induces distortion of helix 2	Affect	No	Core	⊗
G89R	Affect	○	F	Charged bulkier residue	Affect	Yes	Core	⊗
V92L	Benign	○	-	Conserved change	No effect	No	Core	⊗
T95M	Affect	○	-	Conserved change	No effect	No	-	⊗
V122M	Benign	○	-	Conserved change	No effect	Yes	-	⊗
M128T	Benign	○	F	Hydrophobic to small polar	Affect	No	Core	⊗
R142C	Affect	●	-	Non conserved	Affect	No	PKC/GTPase sig.	⊗
R213W	Affect	○	F	Bulky hydrophobic replacement	Affect	Yes	PKC/GTPase sig.	⊗
P268R	Affect	●	-	Charged bulkier residue	Affect	Yes	MSH binding	⊗
T272M	Affect	●	F	Change to large apolar	No effect	Yes	MSH binding	⊗
K278E	Benign	○	-	Charge change	No effect	Yes	MSH binding	⊗
N279K	Affect	●	-	Non conserved change	No effect	Yes	MSH binding	⊗
N281S	Affect	○	NF	Change to smaller residue	No effect	Yes	MSH binding	⊗
A285V	Benign	○	-	Bulkier residue	No effect	Yes	MSH binding	⊗
Y298H	Affect	●	-	Charge change	Affect	No	Core	⊗
T308M	Affect	●	-	Change to large apolar	Affect	Yes	PKC/GTPase sig.	⊗

SNPs3d (<http://www.snps3d.org/>) and Polyphen (<http://genetics.bwh.harvard.edu/pph2/>) online tools

Pub Funct data: Previous published functional data on different *MC1R* variants

3D-Model: three dimensional model variant evaluation according to *MC1R* structure and variant location

Predict: Global evaluation of prediction model

(R), Red hair colour variants; (r), Non red hair colour variants

F, functional data available; NF, non-functional data available, -, non described

● Probably damaging; ○ Possibly damaging; ○ Benign

⊗ Predictive effect; ⊗ Predictive No effect

^aLocation in the three putative regions described by our group, PKC/GTPase sig., GTPase signalling; (-) estimated neutral variants that do not correspond to any sub-region of the protein

FIGURE LEGENDS

Figure 1: MC1R three-dimensional model. Representation of 25 non-synonymous variants described in the Spanish population. a) Location of Red Hair Color (RHC) and Non Red Hair Color (NRHC) variants. Positions RHC are colored in red whereas NRHC are colored in orange. b) Location of potentially-functional rare variants according to their structure. Positions colored in deep blue and cyan. c) Magnification of receptor's core central region. d) Detail of important residues in PKC MC1R binding domain. Residues are colored according to atom nature. Oxygen in red and nitrogen in purple.