#### Elsevier Editorial System(tm) for Plant Science Manuscript Draft

Manuscript Number: PSL-D-13-00566R1

Title: Rubisco catalytic properties optimised for present and future climatic conditions

Article Type: SI: Photosynthesis and climate change

Keywords: Climate change; crops productivity; photosynthesis; Rubisco; water stress.

Corresponding Author: Dr. Jeroni Galmés,

Corresponding Author's Institution: Universitat de les Illes Balears

First Author: Jeroni Galmés

Order of Authors: Jeroni Galmés; Miquel À Conesa; Antonio Díaz-Espejo ; Antoni Mir ; Juan Alejandro Perdomo; Ülo Niinemets ; Jaume Flexas

Abstract: Because of its catalytic inefficiencies, Rubisco is the most obvious target for improvement to enhance the photosynthetic capacity of plants. Two hypotheses are tested in the present work. Firstly, that existing Rubiscos have optimal kinetic properties to maximize photosynthetic carbon assimilation in existing higher plants. Secondly, that current knowledge allows proposal of changes to kinetic properties to make Rubiscos more suited to changed conditions in chloroplasts that are likely to occur with climate change. The catalytic mechanism of Rubisco results in higher catalytic rates of carboxylation being associated with decreased affinity for CO2, so that selection for different environments involves a trade-off between these two properties. The simulations performed in this study confirm that the optimality of Rubisco kinetics depends on the species and the environmental conditions. In particular, environmental drivers affecting the CO2 availability for carboxylation (Cc) or directly shifting the photosynthetic limitations between Rubisco and RuBP regeneration determine to what extend Rubisco kinetics are optimally suited to maximize CO2 assimilation rate. In general, modeled values for optimal kinetic reflect the predominant environmental conditions currently encountered by the species in the field. Under future climatic conditions, photosynthetic CO2 assimilation will be limited by RuBP-regeneration, especially in absence of water stress, the largest rise in [CO2] and lowest increases in temperature. Under these conditions, the model predicts that optimal Rubisco should have high Sc/o and low kcatc.

**Title:** Rubisco catalytic properties optimised for present and future climatic conditions

**Authors:** Galmés J<sup>1\*</sup>, Conesa MÀ<sup>1</sup>, Díaz-Espejo A<sup>2</sup>, Mir A<sup>3</sup>, Perdomo JA<sup>1</sup>, Niinemets Ü<sup>4</sup>, Flexas J<sup>1</sup>

<sup>1</sup>Research Group in Plant Biology under Mediterranean Conditions, Department of Biology, Universitat de les Illes Balears; Carretera de Valldemossa km 7.5; 07122 Palma, Illes Balears, Spain

2 Irrigation and Crop Ecophysiology Group, Instituto de Recursos Naturales y Agrobiología de Sevilla (IRNAS, CSIC). Avenida Reina Mercedes 10, 41012 Sevilla, Spain

<sup>3</sup>Computational Biology and Bioinformatics Research Group, Department of Mathematics and Computer Science, Universitat de les Illes Balears; Carretera de Valldemossa km 7.5; 07122 Palma, Illes Balears, Spain

<sup>4</sup>Institute of Agricultural and Environmental Sciences, Estonian University of Life Sciences, Kreutzwaldi 1, Tartu 51014, Estonia

\*Corresponding author: Jeroni Galmés

**e-mail** jeroni.galmes@uib.cat / **fax** 34 / 971 / 173184 / **tel** 34 / 971 / 259720

Research Group in Plant Biology under Mediterranean Conditions, Department of Biology, Universitat de les Illes Balears. Carretera de Valldemossa Km 7.5, 07122 Palma, Illes Balears, Spain.

**Preprint submitted to Elsevier** 

### **Abstract**

Because of its catalytic inefficiencies, Rubisco is the most obvious target for improvement to enhance the photosynthetic capacity of plants. Two hypotheses are tested in the present work. Firstly, that existing Rubiscos have optimal kinetic properties to maximize photosynthetic carbon assimilation in existing higher plants. Secondly, that current knowledge allows proposal of changes to kinetic properties to make Rubiscos more suited to changed conditions in chloroplasts that are likely to occur with climate change. The catalytic mechanism of Rubisco results in higher catalytic rates of carboxylation being associated with decreased affinity for CO<sub>2</sub>, so that selection for different environments involves a trade-off between these two properties. The simulations performed in this study confirm that the optimality of Rubisco kinetics depends on the species and the environmental conditions. In particular, environmental drivers affecting the CO<sub>2</sub> availability for carboxylation  $(C_c)$  or directly shifting the photosynthetic limitations between Rubisco and RuBP regeneration determine to what extend Rubisco kinetics are optimally suited to maximize CO<sub>2</sub> assimilation rate. In general, modeled values for optimal kinetic reflect the predominant environmental conditions currently encountered by the species in the field. Under future climatic conditions, photosynthetic CO<sub>2</sub> assimilation will be limited by RuBP-regeneration, especially in absence of water stress, the largest rise in [CO<sub>2</sub>] and lowest increases in temperature. Under these conditions, the model predicts that optimal Rubisco should have high  $S_{c/o}$  and low  $k_{cat}^{c}$ .

### Keywords

Climate change, crops productivity, photosynthesis, Rubisco, water stress.

## 1. Introduction

The enzyme Rubisco catalyses the assimilation of CO<sub>2</sub> by the carboxylation of RuBP in the Calvin Cycle, and is therefore the most obvious target to improve the photosynthetic capacity of crops. Rubisco displays catalytic inefficiencies, including slow catalysis and imperfect discrimination between CO<sub>2</sub> and O<sub>2</sub> [1]. These inefficiencies not only limit the rate of CO<sub>2</sub> fixation, but also compromise the capacity of crops to use resources optimally, particularly water and nitrogen [1, 2]. In principle, overcoming these limitations could be successfully accomplished by molecular interventions in the genes coding for both the large (LSu) and the small (SSu) subunits of Rubisco [1, 3 - 5]. Among these interventions, replacement of crop Rubiscos by other versions of the enzyme with better catalytic performance have recently been tested and provide a powerful and promising approach [6 - 8]. However, the success – in terms of photosynthetic improvement – of the Rubisco replacement strategy will depend on the discovery of more efficient and compatible versions of the enzyme. As an example, [9] illustrated how Rubiscos from red algae would perform better than tobacco native Rubisco if they could be transferred into the chloroplast of tobacco.

A number of surveys have reported substantial variability among species in the kinetic parameters of Rubisco, the relative specificity for  $CO_2/O_2$  ( $S_{C/O}$ ), the Michaelis-Menten constants for  $CO_2$  ( $K_c$ ) and  $O_2$  ( $K_o$ ), and the maximum turnover of carboxylation ( $k_{cat}^c$ ) [5, 10 - 15]. Among the potential forces driving evolution of Rubisco catalysis, the availability of  $CO_2$  for ribulose-1,5-biphosphate (RuBP) carboxylation has been proposed as a major factor, explaining some of the differences among distant phylogenetic groups, between  $C_3$  and  $C_4$  species and among  $C_3$  plants [12, 16, 17]. Thus, those environments where plants have evolved under high temperatures and low soil water availability should be prioritized in the search for better versions of the enzyme [18]. Under these conditions, the  $CO_2/O_2$  concentration at the site of carboxylation is decreased due to lower leaf conductances in response to water scarcity and lower  $CO_2/O_2$  solubility ratios as

temperature increases [19]. It has been demonstrated that Rubisco has evolved towards higher  $S_{c/o}$  under these conditions, reducing RuBP oxygenation and favoring the carboxylase reaction [13]. Nevertheless, comparisons of the scarce data on Rubisco kinetics – particularly data describing the full set of kinetic parameters on the same species – indicate a strong, negative correlation between  $S_{c/o}$  and  $k_{cat}^{c}$ . Therefore, the suggestion that Rubiscos with high  $S_{c/o}$  from extreme environments would allow higher  $CO_2$  assimilation rates when transferred into the chloroplast of a crop plant grown under non-stressed conditions, and perhaps even in dense canopies where light limits photosynthesis, is doubtful. The maximum leaf conductances typically measured under optimal growth conditions provide maximum  $CO_2$  availability at the site of carboxylation [2]. Under these conditions, a Rubisco with a higher  $k_{cat}^{c}$ , although at the expenses of a lower  $S_{c/o}$ , may provide the higher benefits [20, 21].

The significant correlations between  $k_{\rm cat}^{\ c}$  and  $S_{\rm c/o}$  – and  $K_{\rm o}$  – have led to the opinion that Rubisco is nearly perfectly adapted for differing CO<sub>2</sub>/O<sub>2</sub> concentrations and thermal conditions in the chloroplast environment [14, 20, 22]. Structural and mechanistic constraints have not allowed the development – either naturally or artificially – of *the perfect* Rubisco with increased specificity to CO<sub>2</sub> and a high rate of carboxylation. Natural evolution has resulted instead in Rubiscos in which there is a compromise between CO<sub>2</sub>/O<sub>2</sub> affinities and the maximum rate of catalytic turnover dependent on the habitat and climate. This tradeoff is evident from a close inspection of published data which shows a significant scatter in Rubisco kinetics values and consequently in carboxylase catalytic efficiency ( $k_{\rm cat}^{\ c}/K_{\rm c}$ ) [5, 14, 22]. Much less is known about the oxygenase kinetic parameters, although some correlation between oxygenase and carboxylase catalytic constants has been observed [23].

Maximum agricultural yields are obtained by growing crops with non-limiting resources by extensive irrigation and fertilization practices. However, the environmental footprint of intensive agriculture and the predictions for higher global temperatures and lower water availability for most

current cropping areas demand novel solutions towards a more sustainable agriculture. It is widely believed that a more efficient Rubisco would provide not only more photoassimilates for plant growth, but importantly do this at a lower cost of water and nutrients [24, 25]. However, given the tradeoff between  $k_{\text{cat}}^{\text{c}}$  and  $S_{\text{c/o}}$ , it is not immediately evident what *more efficient* means in environmental and physiological contexts. Within this context, the present paper aims at the following: i) to model how temperature, and  $CO_2$  and  $O_2$  concentration affect Rubisco kinetics; ii) to test whether current Rubiscos are optimally suited for the present conditions in the chloroplast of higher plants; iii) to estimate the potential impact of climate change on the optimality of Rubisco kinetics and to identify targets of improvement. We argue that it is necessary to consider the tradeoff between specific activity and specificity to define rigorous criteria for engineering plants with *more efficient* Rubisco.

## 2. Methods and theory

#### 2.1. Optimal Rubisco

According to the biochemical model of leaf photosynthesis [26], the net assimilation rate (A) is determined by the minimum of the RuBP-saturated  $(A_c)$  and RuBP-limited  $(A_j)$  CO<sub>2</sub> assimilation rates:

(1)  $A = \min(A_c, A_i),$ 

(2) 
$$A_c = \frac{V_{c \max} (C_c - \Gamma^*)}{C_c + K_c (1 + O_i / K_o)} - R_d$$

(3) 
$$A_j = \frac{(C_c - \Gamma^*)J/4}{C_c - 2\Gamma^*} - R_d$$
,

where  $V_{\rm cmax}$  is the maximal carboxylation rate,  $C_{\rm c}$  is the CO<sub>2</sub> concentration at the site of carboxylation in the chloroplast stroma,  $K_{\rm c}$  is the Michaelis-Menten constant for CO<sub>2</sub> and  $K_{\rm o}$  is that for O<sub>2</sub>,  $\Gamma^*$  is the CO<sub>2</sub> compensation point in the absence of mitochondrial respiration, J is the

 $CO_2$ -saturated electron transport rate of the thylakoid reactions which ultimately supply the necessary energy in the form of ATP and NADPH for the regeneration of RuBP, and  $R_d$  is the mitochondrial respiration in the light. The response of J to the photosynthetic photon flux density (PPFD) was calculated from the non-rectangular hyperbola proposed by Bernacchi et al. [27].

In equation (2),  $V_{\text{cmax}}$  was obtained from in vitro values as:

$$(4) V_{\text{cmax}} = k_{\text{cat}}^{\text{c}} \cdot E \cdot Act$$

 $k_{\text{cat}}^{\text{c}}$  is the Rubisco maximum turnover rate of carboxylation, E is the total quantity of catalytic sites and Act is the percentage of activation of Rubisco sites, i.e. its carbamylation state.

 $\Gamma^*$  was obtained from the Rubisco specificity factor,  $S_{c/o}$ , as:

(5) 
$$\Gamma^* = 0.5 \ O/\ S_{c/o}$$

Because the Rubisco kinetic parameters are interrelated [5, 28], we can express the equations (2) and (3) as determined by a unique kinetic parameter, similarly to the approach followed by Savir et al. [14].  $k_{cat}^c$  is the most reported parameter, probably due to its relative ease of measurement. Thus, equations (2) and (3) were reformulated by substituting  $I^*$ ,  $K_c$  and  $K_o$  by the statistical functions relating them to  $k_{cat}^c$ . To obtain the relationships of  $I^*$ ,  $K_c$  and  $K_o$  vs.  $k_{cat}^c$  at 25°C we used the data compiled in Savir et al. [14], excluding the photosynthetic bacteria. The regression coefficients were highly significant (P < 0.001), with  $r^2 > 0.75$  (Table S1). Because the different kinetic parameters present distinct sensitivities to temperature [29], the functions relating  $k_{cat}^c$  with the remaining parameters are dependent on temperature. Hence, to interrelate the Rubisco kinetic parameters at temperatures other than 25°C, values of  $k_{cat}^c$ ,  $I^*$ ,  $K_c$  and  $K_o$  were converted to the desired temperature using the equations described in Bernacchi et al. [29] for tobacco with the *in vitro* species-specific data measured at 25°C as the reference (i.e., the scaling constant c). Thereafter, regression analyses were again run between  $k_{cat}^c$  and  $I^*$ ,  $K_c$  and  $K_o$  (Table 1S). The analysis therefore assumes that the Rubisco kinetic parameters of the species included in the present study presented the same temperature dependency as that for Rubisco from *Nicotiana* 

*tabacum*, and that the equations hold for the whole range of temperatures from 15°C to 38°C considered in the present study [30].

# 2.2. Optimization of Rubisco kinetics at varying temperature, and CO2 and O2 concentration

A first aim of the study was to model how temperature and [CO<sub>2</sub>] and [O<sub>2</sub>] affect Rubisco kinetics. For this, the optimal value for  $k_{\text{cat}}^{\text{c}}$  (i.e. the one maximizing the net CO<sub>2</sub> assimilation rate) was modeled at varying  $C_{\text{c}}$  and given values for J,  $R_{\text{d}}$ , E and Act by solving equations (1), (2) and (3) expressed in terms of  $k_{\text{cat}}^{\text{c}}$  as the unique parameter, using Mathematica 9 software (Wolfram Research, Champaign, IL, USA). This analysis was performed at three different temperatures (15, 25 and 35 °C) and [O<sub>2</sub>] (1000, 170000 and 210000  $\mu$ mol mol<sup>-1</sup>). The optimal values for  $S_{\text{c/o}}$  ( $\Gamma$ \*),  $K_{\text{c}}$  and  $K_{\text{o}}$  were thereafter obtained from the functions relating them with  $k_{\text{cat}}^{\text{c}}$ , at the desired temperature (Table 1S).

#### 2.3. Optimization of Rubisco kinetics under present conditions

The second aim of the study was to test whether current Rubiscos are optimally suited for the present environmental conditions. By using the same equations, the net  $CO_2$  assimilation rate was calculated at a range of varying  $k_{cat}^c$  values for *Limonium gibertii*, *Nicotiana tabacum* and *Triticum aestivum*, under different environmental conditions (25°C-well watered, 25°C-water stress, 38°C-well watered and 38°C-water stress). *L. gibertii* was selected as being one of the most attractive species in terms of Rubisco traits due to its high specificity [13, 31]. *N. tabacum* is the species where most of attempts to bioengineer Rubisco have been made [32], while *T. aestivum* is one of the most important crops worldwide supporting a significant part of the human caloric intake. Values for the model input parameters:  $C_c$ , J,  $R_d$ , E and Rubisco activation state under the different environmental conditions are shown in Table 1, and were taken from Galmés et al.

(unpublished), Galmés et al. [33] and Perdomo et al. (unpublished), for *L. gibertii*, *N. tabacum* and *T. aestivum*, respectively. The growth conditions were similar for the three original experiments. In all cases, the plants were grown for at least one month at a photosynthetic photon flux density (PPFD) > 600 μmol m<sup>-2</sup> s<sup>-1</sup>, with a photoperiod of 12 h day/12 h night, air relative humidity above 40%, and varying air temperature (25 °C and 38 °C) and soil water availability – field capacity (well watered) and 40% field capacity (moderate water stress). For the different experiments, the model input parameters were measured following the procedures described in Galmés et al. [33]. The modeled optimal values of Rubisco kinetics were then compared to the *in vitro* values obtained from Galmés et al. (unpublished), Whitney et al. [5] and Savir et al. [14] for *L. gibertii*, *N. tabacum* and *T. aestivum*, respectively (see Table 2). Finally, deviation from optimality was calculated for each kinetic parameter as: (optimal value – *in vitro* value)/*in vitro* value.

## 2.4. Optimization of Rubisco kinetics under future conditions

The third aim of the present study was to evaluate the potential effects of climate change on current optimization of Rubisco kinetics in wheat. For this, eight different scenarios of change were considered, each one representing a specific combination of the three main drivers of climate change: atmospheric  $CO_2$  rise (550 or 800  $\mu$ mol mol<sup>-1</sup>), temperature increase (2°C or 5°C increase with respect to 25°C) and water availability (well watered and moderate water stress). The intensities of change in atmospheric  $[CO_2]$  ( $C_a$ ) and temperature were selected according to IPCC predictions [34]. Atmospheric  $[CO_2]$  is projected to continue rising to at least 550 ppm by 2050, and by 2100 the expected increase in air temperature is between 2 and 5°C, depending on the region and the applied model [35].

The impact of each climate change scenario on the stomatal conductance  $(g_s)$  was quantified in relation to the current  $g_s$  measured in wheat under optimal conditions (Perdomo et al. unpublished) as follows: 20% and 50% decrease at  $C_a$  of 550 and 800 ppm, respectively [36],

1.5% increase per °C increase in air temperature according to Evans and von Caemmerer [37], and 50% decrease under moderate water stress according to Galmés et al. [38]. Thereafter, considering the leaf mesophyll conductance  $(g_m)$  measured in the same experiment, a  $g_m/g_s$  ratio of 0.39 was obtained at 25°C. We are aware these values for the  $g_m/g_s$  ratio are lower than typical reported values for a range of species [39], but given the natural variability in this ratio we preferred to use values recently measured on wheat in our laboratory. The  $g_m/g_s$  ratio was changed at 27°C (0.44) and 30°C (0.52) assuming the response to temperature of  $g_m$  published for tobacco in Walker et al. [40].

The concentration of Rubisco catalytic sites and its activation state were also taken from 25°C-well watered plants (Perdomo et al. unpublished) and assumed to be constant for all climate change scenarios (Table 2S). According to a recent review, at the expected increase in  $C_a$ , temperature and severity of water stress, the changes in Rubisco concentration and activation will be minor [41]. By using *in vitro* published data for wheat Rubisco at 25°C [14],  $V_{cmax}$  was calculated at 25°C applying equation (4). A constant ratio  $J_{max}/V_{cmax}$  of 1.5 has been measured in wheat in our laboratory (Perdomo et al. unpublished), which matches very well with other estimates recently published in wheat [42].  $V_{cmax}$  and  $J_{max}$  temperature responses were assumed to be similar to those obtained for tobacco by Walker et al. [40]. Oxygen concentration was taken as 210000  $\mu$ mol mol<sup>-1</sup> and PPFD as 1500  $\mu$ mol mol<sup>-2</sup> s<sup>-1</sup>.

Finally, we ended up with two unknowns, A and  $C_c$ , and two equations, (2) or (3) for A and Fick's law for  $C_c$ :

(6) 
$$C_c = C_a - \frac{A}{g_s} - \frac{A}{g_m}$$

The resultant quadratic equation for Rubisco-limited photosynthesis (2) was solved according to von Caemmerer and Evans [43] and Ethier and Livingston [44], and that for RuBP-limited photosynthesis (3) solved according to Niinemets et al. [45].

All analyses were performed considering A to be first strictly limited by  $A_c$ , and then also including  $A_i$  limitation.

#### 3. Results and Discussion

# 3.1. Optimization of Rubisco kinetics at varying temperature, and $CO_2$ and $O_2$ concentration

Past surveys compiling data on Rubisco kinetics and specific activity are in agreement in that the maximum turnover rate of carboxylation ( $k_{cat}^c$ ) and the affinity for CO<sub>2</sub> (i.e. the inverse of the Michaelis-Menten constant for CO<sub>2</sub>,  $K_c$ ) or the relative specificity for CO<sub>2</sub> and O<sub>2</sub> ( $S_{c/o}$ ) are inversely related [5, 14, 22, 28, 46, 47]. This suggests constant values for the carboxylase catalytic efficiency ( $k_{cat}^c/K_c$ ) at given temperature and substrate concentration [48], and implies that the rate of carboxylation cannot be improved without losing affinity for CO<sub>2</sub> [5, 20, 49]. A direct consequence of limited variability in  $k_{cat}^c/K_c$  is that each particular Rubisco has been forced to evolve towards optimizing either the velocity or the affinity to CO<sub>2</sub>. Although conclusive proofs about the environmental factors driving the evolution of Rubisco have not been provided, there is some evidence that the concentration of the two gaseous substrates, CO<sub>2</sub> and O<sub>2</sub>, and long-term temperature environment are playing a decisive role [13, 17, 22, 50]. It is therefore relevant to explore how these key environmental factors modulate Rubisco performance.

We define optimal Rubisco as Rubisco having a combination of kinetic traits yielding the maximum photosynthesis under given environmental conditions. According to the  $C_3$  photosynthesis model [26], the rate of  $CO_2$  assimilation (A) is the minimum of the ribulose-1,5-bisphosphate (RuBP)-saturated rate of photosynthesis ( $A_c$ ) and the RuBP-limited rate ( $A_j$ ). Consequently, it is important to consider both limitations in assessing the kinetic values for an optimal Rubisco.

At a constant concentration of Rubisco active sites (24  $\mu$ mol m<sup>-2</sup>), and strictly considering the  $A_c$ -limited rate of photosynthesis, increasing the concentration of  $CO_2$  in the chloroplastic stroma ( $C_c$ ) shifts optimality towards increased  $k_{cat}^c$ , and decreased  $S_{c/o}$  (Fig. 1), in agreement with previous analyses [49]. This fact explains the benefits of the catalytic modifications in  $C_4$  Rubiscos under saturating [ $CO_2$ ]. In particular, on average greater  $k_{cat}^c$  values in  $C_4$  than in  $C_3$  plants [8, 51]. The dependency of optimum  $k_{cat}^c$  on  $C_c$  is non-linear, being greater at lower  $C_c$ , and decreasing at higher  $C_c$  (Fig. 1). The inclusion of  $A_j$ -limitation in the analysis changes the response of the optimum  $k_{cat}^c$  on  $C_c$ . In the initial phase, at the  $A_c$ -limited region, optimum  $k_{cat}^c$  increases with  $C_c$  until a certain threshold at which  $A_c$ = $A_j$ . When the threshold  $C_c$  is exceeded,  $C_c$  becomes  $C_c$  in the initial phase with increasing  $C_c$  (Fig. 1). We note that the threshold  $C_c$  depends not only on Rubisco kinetics, but also on light intensity that modifies the rate of RuBP regeneration and on temperature effects on the capacity for photosynthetic electron transport (for explicit derivation of the condition of co-limitation see e.g. [52]).

Due to the tradeoff between  $k_{\text{cat}}^{\text{c}}$  and  $S_{\text{c/o}}$ , the response of optimum  $S_{\text{c/o}}$  on  $C_{\text{c}}$  was opposite to that for optimum  $k_{\text{cat}}^{\text{c}}$  (Fig. 1). For the  $A_{\text{j}}$ -limited rate of photosynthesis,  $S_{\text{c/o}}$  for an optimal Rubisco increased even at high  $C_{\text{c}}$ . This fact explains the predicted beneficial effects of replacing wheat Rubisco by that from *Limonium gibertii* that has a higher  $S_{\text{c/o}}$  [53].

Regardless of whether A is limited by Rubisco or RuBP regeneration, the relationship between the optimal  $k_{\rm cat}^{\ c}/K_c$  and  $C_c$  exhibited a biphasic response. Optimal  $k_{\rm cat}^{\ c}/K_c$  increased with  $C_c$  until a maximum value, after which further increases in  $C_c$  resulted in decreases in  $k_{\rm cat}^{\ c}/K_c$  (Fig. 1). Remarkably, the  $C_c$  value at which the maximum value of optimal  $k_{\rm cat}^{\ c}/K_c$  was attained, ca. 100  $\mu$ mol mol<sup>-1</sup> at 15 °C, 150  $\mu$ mol mol<sup>-1</sup> at 25 °C and 200  $\mu$ mol mol<sup>-1</sup> at 35°C, roughly coincides with the values typically reported for  $C_c$  in  $C_3$  species under non-stressful well-watered conditions and current ambient atmospheric  $CO_2$  concentrations. The reported  $C_c$  values range from 200  $\mu$ mol

mol<sup>-1</sup> in crops and herbaceous plants [37, 54 - 56] to less than 100  $\mu$ mol mol<sup>-1</sup> in sclerophyll leaves with high leaf mass per area [45, 56 - 60].

Rubisco kinetic parameters are all very sensitive to temperature [13, 61 - 63]. However, the relative effect of a given change in temperature is different for each kinetic parameter, due to their differential temperature dependence [29, 64]. This can be observed in Fig. 1, where the proportional effect of altered temperature was different in the response of optimal  $k_{\text{cat}}^{\text{c}}$ ,  $S_{\text{c/o}}$  and  $k_{\text{cat}}^{\text{c}}/K_{\text{c}}$  to varying  $C_{\text{c}}$  (Fig. 1). In the analysis considering  $A_{\text{j}}$  limitation (Fig. 1 right-side panels), an increase in temperature increased the threshold value of  $C_{\text{c}}$  where maximum and minimum values for optimal  $k_{\text{cat}}^{\text{c}}$  and  $S_{\text{c/o}}$  are obtained. This is because temperature affects the  $C_{\text{c}}$  at which the transition from Rubisco to RuBP regeneration limitation occurs [64 - 66].

Changes in the concentration of  $O_2$  also conditioned the above described responses of optimal  $k_{\rm cat}^{\ c}$  and  $S_{\rm c/o}$  to varying  $C_{\rm c}$  (Fig. 1S). As hypothesized [67], an atmosphere with very little molecular oxygen corresponding to the geological past when Rubisco evolved more than 2.5 billion years ago, would favor a Rubisco with increased  $k_{\rm cat}^{\ c}$ , and decreased  $S_{\rm c/o}$ , but only in the case when  $A_{\rm c}$  limits A. Under low  $[O_2]$  and for  $A_j$ -limited A, high  $C_{\rm c}$  would demand Rubiscos with lower  $k_{\rm cat}^{\ c}$  and higher  $S_{\rm c/o}$  (Fig. 1S). According to the biochemical model of photosynthesis [26], under  $A_j$  limitation, the unique kinetic parameter of Rubisco directly determining A is  $S_{\rm c/o}$  (or  $\Gamma^*$ ). Therefore, the demanded lower  $k_{\rm cat}^{\ c}$  is simply an indirect effect of the tradeoff between  $k_{\rm cat}^{\ c}$  and  $S_{\rm c/o}$ .

## 3.2. Rubisco is not optimal in current environmental conditions of $C_3$ plants

Empirical data of leaf gas-exchange, chlorophyll fluorescence and Rubisco biochemistry were compiled for the perennial semi-shrub *Limonium gibertii* adapted to stressful environments in shallow, salty soils in the Mediterranean, annual herb *Nicotiana tabacum* (tobacco) and annual grass *Triticum aestivum* (wheat) from experiments performed under similar environmental

conditions (Table 1). These data were used to model A over a range of values of  $k_{\text{cat}}^{\text{c}}$  to test whether Rubisco kinetics of these three species were optimal under the characteristic environmental conditions encountered in the field (Fig. 2).

First, we considered the predictions under well-watered non-stressed conditions at 25 °C and assumed RuBP-saturated rate of A. Under these conditions, the modeled optimal Rubisco had higher  $k_{\text{cat}}^{\text{c}}$  and  $K_{\text{c}}$  than reported in vitro values for three species (Table 2). Deviations from the actual and the modeled optimum values of  $k_{\text{cat}}^{\text{c}}$  under these conditions were higher in L. gibertii and wheat, and less in tobacco (Fig. 2). Water stress, by reducing  $C_c$ , decreases the optimal values of  $k_{\text{cat}}^{\text{c}}$  (Fig. 2), in agreement with the idea that increasing  $S_{\text{c/o}}$  would be particularly beneficial under semi-arid and arid conditions characterized by most severe limitation of photosynthesis by water availability [15, 52]. Under water stress at 25 °C, the modeled optimal Rubisco had a lower  $k_{\text{cat}}^{\text{c}}$  and  $K_{\text{c}}$  than the reported in vitro values for L. gibertii and tobacco, while modeled and in vitro values perfectly matched in wheat (Table 2). In wheat, increasing leaf temperature to 38 °C augmented the modeled optimal  $k_{\text{cat}}^{\text{c}}$ , but the correspondence with temperature-corrected in vitro values was maintained. Subsequent application of water stress to wheat grown at 38 °C decreased the optimal  $k_{\text{cat}}^{\text{c}}$  to values slightly lower than the *in vitro* ones (Table 2). Overall, evaluating the departure from optimality of all characteristics of Rubisco as the average deviation from optimality for individual traits,  $k_{\text{cat}}^{\ \ c}$ ,  $S_{\text{c/o}}(\Gamma^*)$  and  $K_{\text{c}}$  modeled under  $A_{\text{c}}$ -limited photosynthesis, Rubiscos of Limonium and wheat were better suited for conditions typically measured under water stress, while tobacco Rubisco lost optimality under water stress (Table 3). These results may be a consequence of the enzyme evolution under the predominant environmental conditions encountered by these species in the field. The rocky, saline and hot areas under Mediterranean climate inhabited by Limonium species necessitate a strict control of water loss by stomatal closure, thereby restricting the CO<sub>2</sub> availability for carboxylation [57]. On the other hand, wild wheat relatives and the first domesticated varieties of wheat were drought and salt tolerant [68, 69]. These results corroborate

the results of a past simulation study showing that current Rubisco kinetics of many  $C_3$  plants are better suited for low  $[CO_2]$  [49].

In well-watered plants grown at 25 °C and at moderately high light of 1500  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, A was limited by the rate of RuBP-regeneration ( $A_j$ ) in all three species (Table 2). Under these conditions, optimum  $k_{cat}^c$  is determined at the  $A_c$  to  $A_j$  co-limitation point. At this point, the values of optimum  $k_{cat}^c$  were lower than those modeled strictly under  $A_c$ -limitation (Fig. 2). The values of optimum  $k_{cat}^c$ ,  $K_c$  and  $I^*$  under  $A_c$ ,  $A_j$  co-limitation were lower than *in vitro* reported values (Table 2). Under water stress at 25 °C, the optimum  $k_{cat}^c$  corresponded to the  $A_c$ -limited rate in *Limonium* and tobacco, but still to  $A_c$ ,  $A_j$ -co-limited rate in wheat, with slight increase in the optimum values compared with the well-watered treatment (Fig. 2, Table 2). By increasing the temperature to 38 °C in wheat, A became  $A_c$  limited and optimal  $k_{cat}^c$  increased as explained above (Fig. 2). The overall optimality of Rubisco kinetics after considering  $A_j$ -limitation was improved only in *Limonium* at 25°C and well-watered conditions (Table 3).

This analysis and the simulation by Zhu et al. [49] focusing on  $C_3$  plants reveal that Rubisco kinetics are far from being optimal, and that there is room for improvement. Our results highlight that the optimality of Rubisco kinetics depends on the species and the environmental conditions. In fact, due to the inherent tradeoff between  $S_{c/o}$  and  $k_{cat}^c$ , being optimal under some circumstances, such as water limitations, results in non-optimality in well-watered conditions, especially if there is a cross-over to RuBP-limited photosynthesis. As this analysis further demonstrates, occurrence of RuBP-limitation itself is an important factor altering the Rubisco optimality. In fact, plants are exposed to varying light conditions during the day and only uppermost leaves in the canopy are exposed to full sunlight [45, 70, 71], implying that a large fraction of leaves in the canopy is RuBP-limited at any moment of time and all leaves are RuBP-limited at some moment during the day. In a typical canopy, light distribution is heterogeneous and  $V_{cmax}$  and  $J_{max}$  are distributed accordingly. However, the ratio of  $V_{cmax}$  to  $J_{max}$  does not differ much

between sites [72]. This drove to Chen et al. [73] to propose the coordination theory in which nitrogen inverted in the different photosynthetic pools of the leaves produce a co-limitation by  $A_c$  and  $A_j$  on a daily average. Taken all this, optimization of Rubisco needs consider RuBP-limited photosynthesis as well.

#### 3.3. Optimality of Rubisco kinetics will be affected by climate change

The optimality of Rubisco kinetics was tested under different scenarios of climate change, including varying conditions of atmospheric  $CO_2$ , temperature and water availability, and compared to modeled optimality under present conditions. Based on assumptions of how the environmental changes will impact stomatal  $(g_s)$  and mesophyll  $(g_m)$  conductances to  $CO_2$ , dark respiration rate  $(R_d)$ , photosynthetic electron transport rate (J) and Rubisco traits (see Material and Methods), a value for  $C_c$  corresponding to given set of environmental drivers (ambient  $CO_2$  concentration, light intensity, temperature) was obtained.

Irrespective of whether photosynthesis was RuBP-saturated or -limited, the overall optimality of Rubisco kinetics ( $k_{cat}^c$ ,  $I^*$  and  $K_c$ ) increased in relation to current performance, in three of the eight scenarios of change, decreased in three and did not change significantly in two (Table 4). Under moderately increased [CO<sub>2</sub>] of up to 550 µmol mol<sup>-1</sup>, shift of Rubisco performance towards increased optimality is expected under those scenarios with decreased water availability, regardless of the increase in temperature (Table 4). At a [CO<sub>2</sub>] of 800 µmol mol<sup>-1</sup>, however, more optimal performance compared to performance under current conditions will occur under water stress and severe increment in temperature (Table 4). A closer inspection reveals that the predicted change in optimal Rubisco parameters is driven by possible modifications in  $C_c$ . If climate change drivers lead to  $C_c$  increases, then optimality will decrease, and the reverse occurs if climate change leads to lower  $C_c$  that can be the case under more severe water limitation (Fig. 3).

This is in agreement with the hypothesis that the availability of  $CO_2$  is the main factor modulating Rubisco kinetics evolution [13, 17, 22, 50].

For  $A_c$ -limited photosynthesis, the modeled optimal values of  $k_{cat}^c$  as well as of  $\Gamma^*$ ,  $K_c$  and  $K_0$  increased under all scenarios of change, as compared to current values (Table 2S). However, a part of the increase in  $k_{cat}^c$  is directly caused by the increase in the leaf temperature. At the same predicted increase in temperature, the increase in optimal  $k_{cat}^c$  was lower under the conditions restricting  $CO_2$  availability for carboxylation, i.e. under water stress and modest increases in atmospheric  $CO_2$ . This again emphasizes the complex multivariate dependence of optimal characteristics of Rubisco.

At the calculated  $C_c$  values, photosynthesis is limited by RuBP-regeneration  $(A_i)$  for all scenarios of change, especially in those scenarios with no water stress, highest rise in the [CO<sub>2</sub>], and lowest increase in leaf temperature (Table 2S). In general, under  $A_c$ ,  $A_j$  co-limitation, values of optimal  $k_{\text{cat}}^{\ \ c}$  increased in all cases, as compared to current values, except the scenario predicting no increase in water stress, 800 µmol CO<sub>2</sub> mol<sup>-1</sup> and 2 °C increase in leaf temperature. Noticeably, this is the scenario with the highest predicted increase in  $C_c$  (Table 2S). Regardless of the increase in optimal  $k_{\text{cat}}^{\text{c}}$  relative to present situation, in absolute terms, optimal  $k_{\text{cat}}^{\text{c}}$  under all scenarios is lower than in vitro reported data (Table 2S). In consequence, under future climate, optimal Rubisco for  $A_i$ -limited A should have a higher  $S_{c/o}$ . This finding should be considered in next attempts to engineer Rubisco of crop plants for a higher photosynthetic capacity. Previous simulations ignoring changes in environmental conditions showed that a decrease, not an increase in  $S_{c/o}$  would enhance  $CO_2$  assimilation when photosynthesis was  $A_c$ -limited, but already pointed out that increasing  $S_{c/o}$  would maximize carbon gain under  $A_i$ -limitation [22, 52]. At the crop level, this trend for light-limited photosynthesis under future climate is supported by experimental evidence showing that canopies become denser with plants having higher leaf area indices at higher CO<sub>2</sub> [74, 75]. This may be even more important in wild plants growing under strong light

competition. Accordingly, interspecific differences in  $S_{c/o}$  [13] may strongly modify the competitive potential of different species and thereby alter species dominance and range in future. As demonstrated, the situation can be dramatically different when the severity of drought increases, as expected for a series of ecosystems, in particular in Mediterranean and other semi/arid ecosystems [76]. Such important aspects, including species differences in  $C_c$  [56], are not currently considered in modeling plant responses to future conditions. Of equal importance is that future modelling approaches should consider the effects of  $K_m$  for RuBP under dynamic conditions when light changes rapidly within seconds to minutes, leading to significant changes in the pool size of RuBP [77].

Optimization of Rubisco kinetics largely depends on the ratio between the maximum capacity for carboxylation ( $V_{cmax}$ ) and the capacity for regeneration of RuBP ( $J_{max}$ ). This is because the  $V_{\rm cmax}/J_{\rm max}$  ratio together with light intensity, determines the  $C_{\rm c}$  value at which the transition between the  $A_c$ - and  $A_i$ -limited rates occurs. In fact, it has been suggested that, ideally, a crop plant should express a high  $k_{\text{cat}}^{\text{c}}$ -Rubisco in the upper canopy leaves, exposed to full sunlight, higher temperature and water deficits (therefore  $A_c$ -limited), and a high  $S_{c/o}$ -Rubisco in the shaded lower canopy leaves ( $A_i$ -limited) [20, 49]. Some reports demonstrate that the expected increments in the photosynthetic rate due to higher [CO<sub>2</sub>] may be counteracted by limited nitrogen availability [78, 79]. Actually, under elevated CO<sub>2</sub>, redistribution of nitrogen allocation from Rubisco towards RuBP-regeneration has been documented with clear effects on  $V_{\text{cmax}}/J_{\text{max}}$  ratio [80, 81]. In view of reports compiled in a recent review [44], we have considered constant concentration of Rubisco active sites. However, changes in the activity of Rubisco Activase and in the concentration of Rubisco inhibitors would directly impact on the concentration of Rubisco active sites. Hence, the observed decreases in the Rubisco activation state at elevated [CO<sub>2</sub>] [82], the decreased activity of Rubisco activase under high temperatures [83], or the increased concentration of tight-binding of inhibitors reported in several species under severe drought [25, 84] would lead to lower  $V_{\rm cmax}$ .

Thus, a decrease in  $V_{\rm cmax}/J_{\rm max}$  could shift the limitation towards  $A_{\rm c}$ , and therefore, increase the relevancy of  $k_{\rm cat}^{\ \ c}$  over  $S_{\rm c/o}$ . In fact, this point is equally valid for acclimation to altered temperatures, where cross-over  $C_{\rm c}$  concentration might change due to different temperature relationships of  $J_{\rm max}$  and  $V_{\rm cmax}$  (Fig. 1) [85, 86].

## 4. Concluding remarks

Previous analyses of Rubisco optimization suggested that the kinetic parameters are nearly perfectly optimized [14, 22]. However, the present study shows that analysis of Rubisco optimization necessitates considering the predominant environmental conditions, which influence the [CO<sub>2</sub>] and temperature conditions at the site of carboxylation. When including physiologically relevant conditions of chloroplastic [CO<sub>2</sub>] and temperature, the analysis reveals that Rubisco is not that perfectly optimized, and that there is room for improvement. These results are in agreement with previous simulation by Zhu et al. [49], and the apparent controversy may depend on whether the analysis compares distant phylogenetic groups [14, 22] or is restricted to higher plants [49]. We also demonstrate that current Rubiscos will improve their performance under future climate if change drivers lead to decreased availability of CO<sub>2</sub> for carboxylation. This is likely to occur in future scenarios predicting drier soils, highest increase in temperature and lowest increment in atmospheric [CO<sub>2</sub>]. Overall, the present results, in line with previous surveys, suggest that the concentration of CO<sub>2</sub> and O<sub>2</sub> in the chloroplast and the leaf temperature have been playing a decisive role in shaping Rubisco evolution.

## 5. Acknowledgements

The study was financially supported by the Spanish Ministry of Science and Innovation (AGL2009-07999 and AGL2009-11310/AGR), the Estonian Ministry of Science and Education (institutional grant IUT-8-3) and the European Commission through the European Regional Fund

(the Center of Excellence in Environmental Adaptation). JAP received a pre-doctoral fellowship of Conselleria d'Educació, Cultura i Universitats (Govern de les Illes Balears). We would like to acknowledge Dr. Alfred J Keys (Rothamsted Research) and three ammoniums referees for their helpful comments on a previous version of the manuscript.

#### 6. References

- [1] M.A.J. Parry, P.J. Andralojc, J. Scales, M.E. Salvucci, A.E. Carmo-Silva, H. Alonso, S.M. Whitney, Rubisco activity and regulation as targets for crop improvement. J. Exp. Bot. 64 (2013) 717–730.
- [2] J. Flexas, J. Galmés, A. Gallé, J. Gulías, A. Pou, M. Ribas-Carbo, M. Tomàs, H. Medrano, Improving water use efficiency in grapevines: potential physiological targets for biotechnological improvement. Aust. J. Grape Wine Res. 16 (2010) 106–121.
- [3] I. Kanevski, P. Maliga, D.F. Rhoades, S. Gutteridge, Plastome engineering of ribulose-1,5-bisphosphate carboxylase/oxygenase in tobacco to form a sunflower large subunit and tobacco small subunit hybrid. Plant Physiol. 119 (1999) 133–142.
- [4] T.J. Andrews, S.M. Whitney, Manipulating ribulose bisphosphate carboxylase/oxygenase in the chloroplasts of higher plants. Arch. Biochem. Biophys. 414 (2003) 159–169.
- [5] S.M. Whitney, R.L. Houtz, H. Alonso, Advancing our understanding and capacity to engineer nature's CO<sub>2</sub>-sequestering enzyme, Rubisco. Plant Physiol. 155 (2011) 27–35.
- [6] R.E. Sharwood, S. von Caemmerer, P. Maliga, S.M. Whitney, The catalytic properties of hybrid Rubisco comprising tobacco small and sunflower large subunits mirror the kinetically equivalent source Rubiscos and can support tobacco growth. Plant Physiol. 146 (2008) 83–96.
- [7] C. Ishikawa, T. Hatanaka, S. Misoo, C. Miyake, H. Fukayama, Functional incorporation of sorghum small subunit increases the catalytic turnover rate of Rubisco in transgenic rice. Plant Physiol. 156 (2011) 1603–1611.
- [8] S.M. Whitney, R.E. Sharwood, D. Orr, S.J. White, H. Alonso, J. Galmés, Isoleucine 309 acts as a C<sub>4</sub> catalytic switch that increases ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) carboxylation rate in Flaveria. Proc. Natl. Acad. Sci. U.S.A. 108 (2011) 14688–14693.
- [9] S.M. Whitney, T.J. Andrews, Plastome-encoded bacterial ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) supports photosynthesis and growth in tobacco. Proc. Natl. Acad. Sci. U.S.A. 98 (2011) 14738–14743.

- [10] D.B. Jordan, W.L. Ogren, Species variation in the specificity of ribulose biphosphate carboxylase/oxygenase. Nature 291 (1981) 513–515.
- [11] A.J. Keys, Rubisco: Its Role in Photorespiration, Philos. Trans. R. Soc. B Biol. Sci. 313 (1986) 325–336.
- [12] J.A. Raven, Land plant biochemistry. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 355 (2000) 833–846.
- [13] J. Galmes, J. Flexas, A.J. Keys, J. Cifre, R. Mitchell, P.J. Madgwick, R. Haslam, H. Medrano, M.A.J. Parry, Rubisco specificity factor tends to be larger in plant species from drier habitats and in species with persistent leaves. Plant, Cell environ. 28 (2005) 571–579.
- [14] Y. Savir, E. Noor, R. Milo, T. Tlusty, Cross-species analysis traces adaptation of Rubisco toward optimality in a low-dimensional landscape. Proc. Natl. Acad. Sci. U.S.A. 107 (2010) 3475–3480.
- [15] I. Andersson, A. Backlund, Structure and function of Rubisco. Plant Physiol. Biochem. 46 (2008) 275–291.
- [16] E. Delgado, H. Medrano, A.J. Keys, M.A.J. Parry, Species variation in Rubisco specificity factor. J. Exp. Bot. 46 (1995) 1775–1777.
- [17] J.N. Young, R.E.M. Rickaby, M.V. Kapralov, D.A. Filatov, Adaptive signals in algal Rubisco reveal a history of ancient atmospheric carbon dioxide. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 367 (2012) 483–492.
- [18] C. Peterhansel, M. Niessen, R.M. Kebeish, Metabolic engineering towards the enhancement of photosynthesis. Photochem. Photobiol. 84 (2008) 1317–1323.
- [19] J. Galmés, H. Medrano, J. Flexas, Photosynthetic limitations in response to water stress and recovery in Mediterranean plants with different growth forms. New Phytol. 175 (2007) 81– 93.
- [20] S.P. Long, E.A. Ainsworth, A.D.B. Leakey, J. Nösberger, D.R. Ort, Food for thought: lower than expected crop yield stimulation with rising CO<sub>2</sub> concentrations. Science 312 (2006) 1918–1921.
- [21] E.A. Ainsworth, A. Rogers, A.D.B. Leakey, Targets for crop biotechnology in a future high-CO<sub>2</sub> and high-O<sub>3</sub> world. Plant Physiol. 147 (2008) 13–19.
- [22] G.G.B. Tcherkez, G.D. Farquhar, T.J. Andrews, Despite slow catalysis and confused substrate specificity, all ribulose bisphosphate carboxylases may be nearly perfectly optimized. Proc. Natl. Acad. Sci. U.S.A. 103 (2006) 7246–7251.
- [23] M.J. André, Modelling <sup>18</sup>O<sub>2</sub> and <sup>16</sup>O<sub>2</sub> unidirectional fluxes in plants: II. analysis of rubisco evolution. Biosystems. 103 (2011) 252–264.
- [24] R.J. Spreitzer, Questions about the complexity of chloroplast ribulose-1,5-bisphosphate carboxylase/oxygenase. Photosynth. Res. 60 (1999) 29–42.

- [25] M.A.J. Parry, P.J. Andralojc, S. Khan, P. Lea, A.J. Keys, Rubisco Activity: Effects of drought stress, Ann. Bot. 89 (2002) 833–839.
- [26] G.D. Farquhar, S. von Caemmerer, J.A. Berry, A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. Planta 149 (1980) 78–90.
- [27] C.J. Bernacchi, C. Pimentel, S.P. Long, In vivo temperature response functions of parameters required to model RuBP-limited photosynthesis, Plant, Cell Environ. 26 (2003) 1419–1430.
- [28] G. Bainbridge, P.J. Madgwick, S. Parmar, A.J. Keys, J.E. Pitts, M.A.J. Parry, Engineering Rubisco to change its catalytic properties. J. Exp. Bot. 46 (1995) 1269–1276.
- [29] C.J. Bernacchi, E.L. Singsaas, C. Pimentel, A.R. Portis, S.P. Long, Improved temperature response functions for models of Rubisco-limited photosynthesis. Plant, Cell Environ. 24 (2001) 253–259.
- [30] A. Diaz-Espejo, New challenges in modeling photosynthesis: temperature dependencies of Rubisco kinetics. Plant. Cell Environ.(2013).
- [31] M.A.J. Parry, P.J. Madgwick, J.F.C. Carvalho, P.J. Andralojc, Prospects for increasing photosynthesis by overcoming the limitations of Rubisco. J. Agric. Sci. 145 (2007) 31–43.
- [32] S.M. Whitney, R.E. Sharwood, Construction of a tobacco master line to improve Rubisco engineering in chloroplasts. J. Exp. Bot. 59 (2008) 1909–1921.
- [33] J. Galmés, H. Medrano, J. Flexas, Acclimation of Rubisco specificity factor to drought in tobacco: discrepancies between in vitro and in vivo estimations. J. Exp. Bot. 57 (2006) 3659–3667.
- [34] IPCC, Cambio climático 2007: Informe de síntesis. Contribución de los grupos de trabajo I, II y III al cuarto informe de evaluación del grupo Intergubernamental de expertos sobre el cambio climático. (2007).
- [35] IPCC, Working Group I Contribution to the IPCC Fifth Assessment Report Climate Change 2013: The Physical Science Basis Summary for Policymakers. (2013).
- [36] E.A. Ainsworth, A. Rogers, The response of photosynthesis and stomatal conductance to rising [CO<sub>2</sub>]: mechanisms and environmental interactions. Plant. Cell Environ. 30 (2007) 258–270.
- [37] J.R. Evans, S. von Caemmerer, Temperature response of carbon isotope discrimination and mesophyll conductance in tobacco. Plant. Cell Environ. 36 (2012) 745–756.
- [38] J. Galmés, J. Flexas, R. Savé, H. Medrano, Water relations and stomatal characteristics of Mediterranean plants with different growth forms and leaf habits: responses to water stress and recovery. Plant Soil 290 (2007) 139–155.
- [39] J. Flexas, Ü. Niinemets, A. Gallé, M.M. Barbour, M. Centritto, A. Diaz-Espejo, C. Douthe, J. Galmés, M. Ribas-Carbó, P.L. Rodriguez, F. Rosselló, R. Soolanayakanahally, M.

- Tòmas, I.J. Wright, G.D. Farquhar, H. Medrano, Diffusional conductances to CO<sub>2</sub> as a target for increasing photosynthesis and photosynthetic water-use efficiency. Photosynth. Res. (2013).
- [40] B. Walker, L.S. Ariza, S. Kaines, M.R. Badger, A.B. Cousins, Temperature response of invivo Rubisco kinetics and mesophyll conductance in *Arabidopsis thaliana*: comparisons to *Nicotiana tabacum*. Plant. Cell Environ. (2013).
- [41] J. Galmés, I. Aranjuelo, H. Medrano, J. Flexas, Variation in Rubisco content and activity under variable climatic factors. Photosynth. Res. (2013).
- [42] C. Zhu, L. Ziska, J. Zhu, Q. Zeng, Z. Xie, H. Tang, X. Jia, T. Hasegawa, The temporal and species dynamics of photosynthetic acclimation in flag leaves of rice (*Oryza sativa*) and wheat (*Triticum aestivum*) under elevated carbon dioxide. Physiol. Plant. 145 (2012) 395–405.
- [43] S. von Caemmerer, J. Evans, Determination of the average partial pressure of CO<sub>2</sub> in chloroplasts from leaves of several C<sub>3</sub> plants. Funct. Plant Biol. 18 (1991) 287–305.
- [44] G.J. Ethier, N.J. Livingston, On the need to incorporate sensitivity to CO<sub>2</sub> transfer conductance into the Farquhar-von Caemmerer-Berry leaf photosynthesis model, Plant, Cell Environ. 27 (2004) 137–153.
- [45] Ü. Niinemets, A. Díaz-Espejo, J. Flexas, J. Galmés, C.R. Warren, Importance of mesophyll diffusion conductance in estimation of plant photosynthesis in the field. J. Exp. Bot. 60 (2009) 2271–2282.
- [46] R.J. Spreitzer, M.E. Salvucci, Rubisco: structure, regulatory interactions, and possibilities for a better enzyme. Annu. Rev. Plant Biol. 53 (2002) 449–475.
- [47] S.P. Long, E.A. Ainsworth, A. Rogers, D.R. Ort, Rising atmospheric carbon dioxide: plants FACE the future. Annu. Rev. Plant Biol. 55 (2004) 591–628.
- [48] R. Eisenthal, M.J. Danson, D.W. Hough, Catalytic efficiency and  $k_{cat}/K_M$ : a useful comparator? Trends Biotechnol. 25 (2007) 247–249.
- [49] X. Zhu, A. Portis, S.P. Long, Would transformation of C<sub>3</sub> crop plants with foreign Rubisco increase productivity? A computational analysis extrapolating from kinetic properties to canopy. Plant. Cell Environ. 27 (2004) 155–165.
- [50] R.F. Sage, Variation in the k<sub>cat</sub> of Rubisco in C<sub>3</sub> and C<sub>4</sub> plants and some implications for photosynthetic performance at high and low temperature. J. Exp. Bot. 53 (2002) 609–620.
- [51] P.A. Christin, N. Salamin, A.M. Muasya, E.H. Roalson, F. Russier, G. Besnard, Evolutionary switch and genetic convergence on *rbcL* following the evolution of C<sub>4</sub> photosynthesis. Mol. Biol. Evol. 25 (2008) 2361–2368.
- [52] Ü. Niinemets, J.D. Tenhunen, A model separating leaf structural and physiological effects on carbon gain along light gradients for the shade-tolerant species *Acer saccharum*. Plant, Cell Environ. 20 (1997) 845–866.

- [53] M.A.J. Parry, M.J. Hawkesford, An integrated approach to crop genetic improvement. J. Integr. Plant Biol. 54 (2012) 250–259.
- [54] J. Flexas, M.F. Ortuño, M. Ribas-Carbó, A. Diaz-Espejo, I.D. Flórez-Sarasa, H. Medrano, Mesophyll conductance to CO<sub>2</sub> in *Arabidopsis thaliana*. New Phytol. 175 (2007) 501–511.
- [55] J. Galmés, M.À. Conesa, J.M. Ochogavía, J.A. Perdomo, D.M. Francis, M. Ribas-Carbó, R. Savé, J. Flexas, H. Medrano, J. Cifre, Physiological and morphological adaptations in relation to water use efficiency in Mediterranean accessions of *Solanum lycopersicum*. Plant. Cell Environ. 34 (2011) 245–260.
- [56] Ü. Niinemets, J. Flexas, J. Peñuelas, Evergreens favored by higher responsiveness to increased CO<sub>2</sub>, Trends Ecol. Evol. 26 (2011) 136–142.
- [57] J. Galmés, M. Ribas-Carbó, H. Medrano, J. Flexas, Rubisco activity in Mediterranean species is regulated by the chloroplastic CO<sub>2</sub> concentration under water stress. J. Exp. Bot. 62 (2011) 653–665.
- [58] J.M. Warren, R.J. Norby, S.D. Wullschleger, Elevated CO<sub>2</sub> enhances leaf senescence during extreme drought in a temperate forest. Tree Physiol. 31 (2011) 117–130.
- [59] J.J. Peguero-Pina, J. Flexas, J. Galmés, Ü. Niinemets, D. Sancho-Knapik, G. Barredo, D. Villarroya, E. Gil-Pelegrín, Leaf anatomical properties in relation to differences in mesophyll conductance to CO<sub>2</sub> and photosynthesis in two related Mediterranean Abies species. Plant. Cell Environ. 35 (2012) 2121–2129.
- [60] F. Hassiotou, M. Ludwig, M. Renton, E.J. Veneklaas, J.R. Evans, Influence of leaf dry mass per area, CO<sub>2</sub>, and irradiance on mesophyll conductance in sclerophylls. J. Exp. Bot. 60 (2009) 2303–2314.
- [61] N.P. Hall, A.J. Keys, Temperature dependence of the enzymic carboxylation and oxygenation of ribulose 1,5-bisphosphate in relation to effects of temperature on photosynthesis. Plant Physiol. 72 (1983) 945–948.
- [62] A. Brooks, G.D. Farquhar, Effect of temperature on the CO<sub>2</sub>/O<sub>2</sub> specificity of ribulose-1,5-bisphosphate carboxylase/oxygenase and the rate of respiration in the light. Planta 165 (1985) 397–406.
- [63] R.F. Sage, Y.P. Cen, M. Li, The activation state of Rubisco directly limits photosynthesis at low CO<sub>2</sub> and low O<sub>2</sub> partial pressures. Photosynth. Res. 71 (2002) 241–50.
- [64] S. von Caemmerer, Biochemical models of leaf photosynthesis, CSIRO, Collingwood, 2000.
- [65] K. Hikosaka, K. Ishikawa, A. Borjigidai, O. Muller, Y. Onoda, Temperature acclimation of photosynthesis: mechanisms involved in the changes in temperature dependence of photosynthetic rate. J. Exp. Bot. 57 (2006) 291–302.

- [66] W. Yamori, S. von Caemmerer, Effect of Rubisco activase deficiency on the temperature response of CO<sub>2</sub> assimilation rate and Rubisco activation state: insights from transgenic tobacco with reduced amounts of Rubisco activase. Plant Physiol. 151 (2009) 2073–2082.
- [67] E.G. Nisbet, R.E.R. Nisbet, Methane, oxygen, photosynthesis, rubisco and the regulation of the air through time. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 363 (2008) 2745–2754.
- [68] T.D. Colmer, T.J. Flowers, R. Munns, Use of wild relatives to improve salt tolerance in wheat. J. Exp. Bot. 57 (2006) 1059–1078.
- [69] D. Placido, M. Campbell, J. Folsom, X. Cui, G. Kruger, P. Baenziger, H. Walia, Introgression of novel traits from a wild wheat relative improves drought adaptation in wheat. Plant Physiol. 161 (2013) 1806–1819.
- [70] Ü. Niinemets, Photosynthesis and resource distribution through plant canopies. Plant. Cell Environ. 30 (2007) 1052–1071.
- [71] F. Valladares, J. Garcia-Plazaola, F. Morales, Ü. Niinemets, Photosynthetic responses to radiation, in: J. Flexas, F. Loreto, H. Medrano (Eds.), Terrestrial photosynthesis in a changing environment. A molecular, physiological and ecological approach, Cambridge: University Press, Cambridge, 2012, pp. 239–256.
- [72] J.R. Evans, H. Poorter, Photosynthetic acclimation of plants to growth irradiance: the relative importance of specific leaf area and nitrogen partitioning in maximizing carbon gain. Plant. Cell Environ. 24 (2001) 755–767.
- [73] J.L. Chen, J.F. Reynolds, P.C. Harley, J.D. Tenhunen, Coordination theory of leaf nitrogen distribution in a canopy. Oecologia 93 (1993) 63–69.
- [74] F. Ewert, Modelling plant responses to elevated CO<sub>2</sub>: how important is leaf area index? Ann. Bot. 93 (2004) 619–627.
- [75] R.J. Norby, E.H. Delucia, B. Gielen, C. Calfapietra, C.P. Giardina, J.S. King, J. Ledford, H.R. McCarthy, D.J.P. Moore, R. Ceulemans, P. De Angelis, A.C. Finzi, D.F. Karnosky, M.E. Kubiske, M. L6ukac, K.S. Pregitzer, G.E. Scarascia-Mugnozza, W.H. Schlesinger, R. Oren, Forest response to elevated CO<sub>2</sub> is conserved across a broad range of productivity. Proc. Natl. Acad. Sci. U. S. A. 102 (2005) 18052–18056.
- [76] J. Christensen, B. Hewitson, A. Busuioc, A. Chen, X. Gao, I. Held, R. Jones, R. Kolli, W. Kwon, R. Laprise, Regional climate projections, in: S. Solomon, D. Qin, M. Manning, Z. Chen, M. Marquis, K. Averyt, M. Tignor, H. Miller (Eds.), Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, Cambridge: University Press, Cambridge and New York, 2007, pp. 847–940.
- [77] J. Kobza, J.R. Seemann, Regulation of ribulose-1,5- bisphosphate carboxylase activity in response to diurnal changes in irradiance. Plant Physiol. 89 (1989) 918–924.

- [78] N.P.R. Anten, T. Hirose, Y. Onoda, T. Kinugasa, H.Y. Kim, M. Okada, K. Kobayashi, Elevated CO<sub>2</sub> and nitrogen availability have interactive effects on canopy carbon gain in rice. New Phytol. 161 (2004) 459–471.
- [79] S. Seneweera, A. Makino, N. Hirotsu, R. Norton, Y. Suzuki, New insight into photosynthetic acclimation to elevated CO<sub>2</sub>: The role of leaf nitrogen and ribulose-1,5-bisphosphate carboxylase/oxygenase content in rice leaves. Environ. Exp. Bot. 71 (2011) 128–136.
- [80] E.A. Ainsworth, A. Rogers, The response of photosynthesis and stomatal conductance to rising [CO<sub>2</sub>]: mechanisms and environmental interactions. Plant. Cell Environ. 30 (2007) 258–270.
- [81] I. Aranjuelo, L. Cabrera-Bosquet, J.L. Araus, S. Nogués, Carbon and nitrogen partitioning during the post-anthesis period is conditioned by N fertilisation and sink strength in three cereals. Plant Biol. 15 (2013) 135–143.
- [82] Y. Cen, R. Sage, The regulation of Rubisco activity in response to variation in temperature and atmospheric CO<sub>2</sub> partial pressure in sweet potato. Plant Physiol. 139 (2005) 979–990.
- [83] R.F. Sage, D.A. Way, D.S. Kubien, Rubisco, Rubisco activase, and global climate change, J. Exp. Bot. 59 (2008) 1581–1595.
- [84] A.E. Carmo-Silva, A.J. Keys, P.J. Andralojc, S.J. Powers, M.C. Arrabaça, M.A.J. Parry, Rubisco activities, properties, and regulation in three different C<sub>4</sub> grasses under drought. J. Exp. Bot. 61(2010) 2355–2366.
- [85] K. Hikosaka, A. Murakami, T. Hirose, Balancing carboxylation and regeneration of ribulose-1,5- bisphosphate in leaf photosynthesis: temperature acclimation of an evergreen tree, *Quercus myrsinaefolia*. Plant, Cell Environ. 22 (1999) 841–849.
- [86] O. Muller, K. Hikosaka, T. Hirose, Seasonal changes in light and temperature affect the balance between light harvesting and light utilisation components of photosynthesis in an evergreen understory shrub. Oecologia 143 (2005) 501–508.

**Table 1.** Leaf photosynthetic parameters for *Limonium gibertii*, *Nicotiana tabacum* and *Triticum aestivum* under different conditions of water availability (WW: well-watered; WS: water stress) and leaf temperature. Values for the net CO<sub>2</sub> assimilation rate (A), stomatal conductance ( $g_s$ ), mesophyll conductance ( $g_m$ ), CO<sub>2</sub> concentration in the chloroplast ( $C_c$ ), rate of electron transport (J), mitochondrial respiration in the light ( $R_d$ ), concentration of Rubisco sites (E) and Rubisco activation state consisted in experimental data from Galmés et al. (unpublished), Galmés et al. [33] and Perdomo et al. (unpublished) for L. *gibertii*, N. *tabacum* and T. *aestivum*, respectively. Photosynthetic photon flux density and oxygen concentration during measurements were constant at 1500 μmol m<sup>-2</sup> s<sup>-1</sup> and 210000 μmol mol<sup>-1</sup>, respectively. Growth conditions were similar for the three original sources, and the plants were acclimated (i.e. grown for at the least one month) to the experimental conditions.

	L. gi	bertii	N. tal	расит	T. aestivum				
	25°C-WW	25°C-WS	25°C-WW	25°C-WS	25°C-WW	25°C-WS	38°C-WS	38°C-WS	
$A  (\mu \text{mol m}^{-2}  \text{s}^{-1})$	23.5	10.8	23.8	8.9	23.8	14.5	17.8	13.03	
$g_{\rm s}  ({\rm mol}  {\rm m}^{-2}  {\rm s}^{-1})$	0.306	0.095	0.335	0.049	0.520	0.094	0.436	0.219	
$g_{\rm m}  ({\rm mol} \; {\rm m}^{-2} \; {\rm s}^{-1})$	0.354	0.099	0.404	0.089	0.203	0.037	0.258	0.139	
$C_{\rm c}$ (µmol mol <sup>-1</sup> )	167.1	80.1	206.4	92.2	181.4	109.1	217.8	172.1	
J (µmol m <sup>-2</sup> s <sup>-1</sup> )	132.3	157.6	175.6	184.3	130.6	144.4	278.3	274.1	
$R_{\rm d}$ (µmol m <sup>-2</sup> s <sup>-1</sup> )	2.1	1.1	1.2	1.2	0.4	0.9	1.3	0.4	
$E  (\mu \text{mol m}^{-2})$	64.0	81.7	75.5	77.5	62.9	72.1	90.4	84.0	
Activation state (%)	75	65	77	82	80	80	50	50	

**Table 2.** Current and optimal values of the net CO<sub>2</sub> assimilation rate (A) and *in vitro* Rubisco kinetics for *Limonium gibertii*, *Nicotiana tabacum* and *Triticum aestivum* under different conditions of water availability (WW: well-watered; WS: water stress) and leaf temperature. To obtain the optimal values, the photosynthetic rate was considered to be limited strictly by Rubisco ( $A_c$  limitation) or co-limited by Rubisco and RuBP-regeneration capacity ( $A_c$ ,  $A_j$  co-limitation). Values for A were experimentally measured (same as in Table 1). Current values at 25°C of the maximum turnover rate of carboxylation ( $k_{cat}^c$ ), the CO<sub>2</sub> compensation point in the absence of mitochondrial respiration ( $I^*$ ), and the Michaelis-Menten constants of Rubisco for CO<sub>2</sub> ( $K_c$ ) and O<sub>2</sub> ( $K_o$ ) were obtained from Galmés et al. (unpublished), Whitney et al.[5], and Savir et al. [14] for L gibertii, N tabacum and I aestivum, respectively. Values of in vitro Rubisco kinetics for N tabacum at 38°C were obtained after applying temperature equations from Bernacchi et al. [29], but using measured data at 25°C as the reference.

	L. gib	ertii	N. tab	асит	T. aestivum				
	25°C-WW	25°C-WS	25°C-WW	25°C-WS	25°C-WW	25°C-WS	38°C-WW	38°C-WS	
Current values									
$A  (\mu \text{mol m}^{-2}  \text{s}^{-1})$	23.5	10.8	23.8	8.9	23.8	14.5	17.8	13.03	
$k_{\rm cat}^{}$ (s <sup>-1</sup> )	2.5		3.2		2.5		5.5		
$\Gamma$ • ( $\mu$ mol mol <sup>-1</sup> )	35.2		48.2		43.9		66.3		
$K_{\rm c}$ (µmol mol <sup>-1</sup> )	273		339		443		1737		
$K_{\rm o}$ (µmol mol <sup>-1</sup> )	3416	571	233468		622014		928102		
		Opti	mal values u	$nder A_c lim$	itation				
$A  (\mu \text{mol m}^{-2}  \text{s}^{-1})$	21.2	8.5	33.8	12.5	26.5	15.3	19.9	14.0	
$k_{\rm cat}^{\ \ c} ({\rm s}^{-1})$	3.3	1.9	3.6	2.2	3.4	2.6	5.9	5.2	
$\Gamma$ • (µmol mol <sup>-1</sup> )	45.6	31.0	47.8	35.0	46.5	38.9	61.2	55.9	

,	•	1			•			
$K_{\rm c}$ (µmol mol <sup>-1</sup> )	422	264	465	293	438	330	1364	1218
$K_{\rm o}$ (µmol mol <sup>-1</sup> )	376046	333502	385614	342472	379868.0	353048	701605	681460
		Optimal	values und	$\operatorname{er} A_{\operatorname{c}}, A_{\operatorname{i}} \operatorname{co}$	limitation			
$A \text{ (}\mu\text{mol m}^{-2}\text{ s}^{-1}\text{)}$	18.0	8.5	27.8	12.5	21.1	14.9	19.9	14.0
$k_{\rm cat}^{}$ (s <sup>-1</sup> )	1.8	1.9	1.8	2.2	1.6	2.0	5.9	5.2
$\Gamma$ • ( $\mu$ mol mol <sup>-1</sup> )	29.6	31.0	30.2	35.0	26.7	32.5	61.2	55.9
$K_{\rm c}$ (µmol mol <sup>-1</sup> )	255.9	264	259.5	293	240.5	274.3	1364	1218
$K_{\rm o}$ (µmol mol <sup>-1</sup> )	330688	333502	331884	342472	325442	336659	701605	681460

**Table 3.** Rubisco kinetics optimality in *Limonium gibertii*, *Nicotiana tabacum* and *Triticum aestivum* under different conditions of water availability (WW: well-watered; WS: water stress) and leaf temperature. Rubisco kinetics optimality was calculated as the average of the optimality for the maximum turnover rate of carboxylation ( $k_{cat}^{c}$ ), the CO<sub>2</sub> compensation point in the absence of mitochondrial respiration ( $I^{*}$ ) and the Michaelis-Menten constant of Rubisco for CO<sub>2</sub> ( $K_c$ ). The photosynthetic rate was considered to be limited strictly by Rubisco ( $A_c$  limitation) or co-limited by Rubisco and RuBP-regeneration capacity ( $A_c$ ,  $A_j$  co-limitation). The input values of the model are shown in Table 1.

		A <sub>c</sub> limitation	$A_{\rm c}, A_{\rm j}$ co-limitation		
I aibantii	25°C-WW	37.9	17.3		
L. gibertii	25°C-WS	13.6	13.6		
N. tabacum	25°C-WW	16.7	34.6		
	25°C-WS	24.1	24.1		
T. aestivum	25°C-WW	14.4	40.4		
	25°C-WS	13.0	28.1		
	38°C-WW	12.5	12.5		
	38°C-WS	17.0	17.0		

**Table 4.** Rubisco kinetics optimality in *Triticum aestivum* under different scenarios of climate change, at varying atmospheric  $CO_2$  concentration  $(C_a)$ , leaf temperature  $(T_{leaf})$  and water status (WW: well-watered; WS: water stress). The photosynthetic rate was considered to be limited strictly by Rubisco  $(A_c \text{ limitation})$  or co-limited by Rubisco and RuBP-regeneration capacity  $(A_c, A_j \text{ co-limitation})$ . The input values of the model are shown in Table 2S. The equations describing the correlation among Rubisco kinetic parameters at the different temperatures are shown in Table 1S.

Climate change scenario	$C_{\rm a}$	$T_{\text{leaf}}$	Water status	$A_{\rm c}$ limitation	$A_{\rm c}, A_{\rm j}$ co-limitation
Current climate	400	25	WW	24.9	44.6
1	550	27	WW	30.1	47.6
2	550	27	WS	14.0	34.5
3	800	27	WW	44.6	55.5
4	800	27	WS	24.9	44.1
5	550	30	WW	21.8	44.5
6	550	30	WS	12.0	29.2
7	800	30	WW	37.2	53.8
8	800	30	WS	17.3	40.4

**Figure 1.** Optimal values for the maximum turnover rate of carboxylation  $(k_{cat}^{c})$ , specificity factor  $(S_{c/o})$  and carboxylation catalytic efficiency  $(k_{cat}^{c}/K_c)$  at a range of CO<sub>2</sub> concentrations in the chloroplast  $(C_c)$  at 15°C (empty circles), 25°C (filled circles) and 35°C (empty triangles). The photosynthetic rate was considered to be limited strictly by Rubisco  $(A_c)$  limited, left graphs) or co-limited by Rubisco and RuBP-regeneration capacity  $(A_c, A_j)$  co-limited, right graphs). The rates of electron transport (J) were 42, 74 and 106 μmol m<sup>-2</sup> s<sup>-1</sup> at 15°C, 25°C and 35°C, respectively. The rates of mitochondrial respiration in the light  $(R_d)$  were 0.5, 1.0 and 1.8 μmol m<sup>-2</sup> s<sup>-1</sup> at 15°C, 25°C and 35°C, respectively. The concentration of Rubisco catalytic sites (30 μmol m<sup>-2</sup>) and its activation state (80%) were considered invariable at the three temperatures.

**Figure 2.** Net CO<sub>2</sub> assimilation rate (A) at a range of values for the maximum turnover rate of carboxylation ( $k_{cat}^{c}$ ) for *Limonium gibertii*, *Nicotiana tabacum* and *Triticum aestivum*, under non-stress conditions (25°C-well-watereted, filled circles), 25°C-water stress (empty circles), 38°C-well-watered (filled triangles) and 38°C-water stress (empty triangles). The photosynthetic rate was considered to be limited strictly by Rubisco ( $A_{c}$  limited, left graphs) or co-limited by Rubisco and RuBP-regeneration capacity ( $A_{c}$ ,  $A_{j}$  co-limited, right graphs). The dotted line indicates the actual  $k_{cat}^{c}$  value for the three species. The input values of the model are shown in Table 1. The equations describing the correlation among Rubisco kinetic parameters at the different temperatures are shown in Table 1S.

**Figure 3.** The relationship between the predicted change in optimal Rubisco in T. aestivum under the different scenarios of climate change (indicated in Table 4) and the chloroplastic  $CO_2$  concentration ( $C_c$ ) estimated for the same scenarios. Values of the

predicted change in optimal Rubisco are normalized to the current values (set at 1), <1 means improvement of Rubisco optimality, >1 means decrement of Rubisco optimality. The photosynthetic rate was considered to be limited strictly by Rubisco ( $A_c$  limitation, filled circles and solid line) or co-limited by Rubisco and RuBP-regeneration capacity ( $A_c$ ,  $A_j$  co-limitation, empty circles, dashed line).

Figure 1.

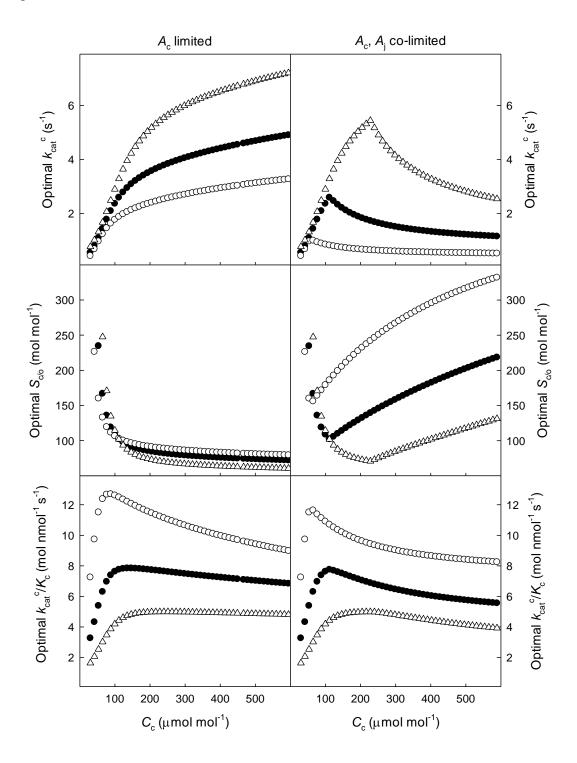


Figure 2.

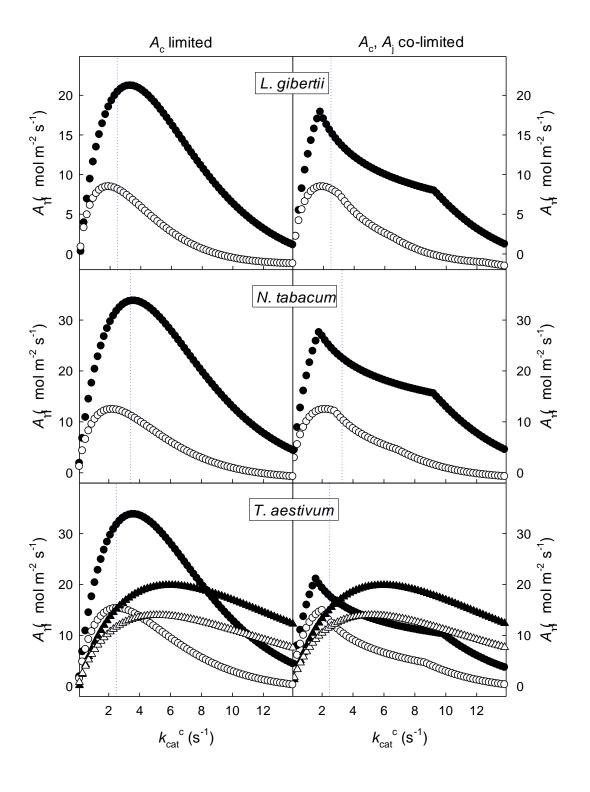
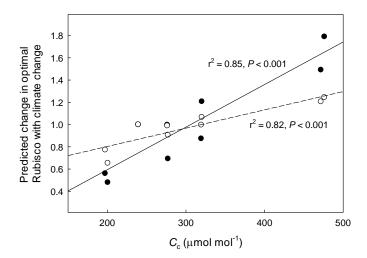


Figure 3.



## Table(s) supplemental

**Table 1S.** Equations describing the relationship between the maximum turnover rate of carboxylation ( $k_{\text{cat}}^{\text{c}}$ ) and the CO<sub>2</sub> compensation point in the absence of mitochondrial respiration ( $\Gamma^*$ ), and the Michaelis-Menten constants of Rubisco for CO<sub>2</sub> ( $K_c$ ) and O<sub>2</sub> ( $K_c$ ) at the different temperatures used in the present article.

$T_{leaf}$	$arGamma^*$	$K_{\rm c}$	$K_{ m o}$
	$22.39+18.40 \cdot \ln(k_{\text{cat}}^{\text{c}})$	$45.7 \cdot \exp(0.634 \cdot k_{\text{cat}}^{\text{c}})$	$K_{\rm c}/(0.0003 \cdot \exp(0.471 \cdot k_{\rm cat}^{\rm c}))$
25 °C	$14.64 + 25.93 \cdot \ln(k_{\text{cat}}^{\text{c}})$	$142.1 \cdot \exp(0.330 \cdot k_{\text{cat}}^{\text{c}})$	$K_{\rm c} / (0.0005 \cdot \exp(0.245 \cdot k_{\rm cat}^{\ \ c}))$
27 °C	$12.17 + 27.69 \cdot \ln(k_{\text{cat}}^{\text{c}})$	$176.7 \cdot \exp(0.292 \cdot k_{\text{cat}}^{\text{c}})$	$K_{\rm c} / (0.0006 \cdot \exp(0.216 \cdot k_{\rm cat}^{\ \ c}))$
30 °C	$7.78 + 30.51 \cdot \ln(k_{\text{cat}}^{\text{c}})$	$243.7 \cdot \exp(0.242 \cdot k_{\text{cat}}^{\text{c}})$	$K_{\rm c} / (0.0007 \cdot \exp(0.180 \cdot k_{\rm cat}^{\ \ c}))$
35 °C	$-1.61+35.72 \cdot \ln(k_{\text{cat}}^{\text{c}})$	$410.7 \cdot \exp(0.181 \cdot k_{\text{cat}}^{\text{c}})$	$K_{\rm c}/(0.0009 \cdot \exp(0.133 \cdot k_{\rm cat}^{\ \ c}))$
38 °C	$-8.62 + 39.17 \cdot \ln(k_{\text{cat}}^{\text{c}})$	$557.2 \cdot \exp(0.151 \cdot k_{\text{cat}}^{\text{c}})$	$K_{\rm c}/(0.0013 \cdot \exp(0.112 \cdot k_{\rm cat}^{\ \ c}))$
	$r^2 = 0.79, P < 0.001$	$r^2 = 0.87, P < 0.001$	$r^2 = 0.75, P < 0.001$

**Table 2S.** Modeled optimum values of Rubisco kinetics in *Triticum aestivum* under different scenarios of climate change, at varying atmospheric  $CO_2$  concentration ( $C_a$ ), leaf temperature ( $T_{leaf}$ ) and water status (WW: well-watered; WS: water stress). The predicted effects of climate change variables on the stomatal conductance ( $g_s$ ) were: 20% and 50% decrease at  $C_a$  of 550 and 800 μmol mol<sup>-1</sup>, respectively; 1.5% increase per °C increase; 50% decrease under WS. The photosynthetic rate was considered to be limited strictly by Rubisco ( $A_c$  limitation) or co-limited by Rubisco and RuBP-regeneration capacity ( $A_c$ ,  $A_j$  co-limitation). Values for  $g_s$ , rate of mitochondrial respiration in the light ( $R_d$ ), maximum turnover rate of carboxylation ( $k_{cat}^c$ ),  $CO_2$  compensation point in the absence of mitochondrial respiration ( $I^*$ ), Michaelis-Menten constants of Rubisco for  $CO_2$  ( $K_c$ ) and  $O_2$  ( $K_o$ ), concentration of Rubisco catalytic sites (E) and Rubisco activation state under current climate were obtained from experimental data as explained in Tables 1 and 2S. Photosynthetic photon flux density (PPFD) and oxygen concentration (O) during measurements were constant at 1500 μmol m<sup>-2</sup> s<sup>-1</sup> and 210000 μmol mol<sup>-1</sup>, respectively.

		Cl	imate chan	ge scenario	ı				
	Current climate	1	2	3	4	5	6	7	8
$C_{\rm a}$ (µmol mol <sup>-1</sup> )	400	550	550	800	800	550	550	800	800
$T_{\text{leaf}}$ (°C)	25	27	27	27	27	30	30	30	30
Water status	WW	WW	WS	WW	WS	WW	WS	WW	WS
			Input par	ameters					
$g_s  (\text{mol m}^{-2}  \text{s}^{-1})$	0.52	0.40	0.20	0.33	0.16	0.39	0.19	0.31	0.16
$g_{\rm m}  ({\rm mol}  {\rm m}^{-2}  {\rm s}^{-1})$	0.20	0.18	0.09	0.14	0.07	0.20	0.10	0.16	0.08
$C_{\rm c}$ (µmol mol <sup>-1</sup> )	239	320	197	476	276	319	200	472	277
$J  (\mu \mathrm{mol} \; \mathrm{m}^{-2} \; \mathrm{s}^{-1})$	137	149	149	149	149	164	164	164	164
$R_{\rm d}  (\mu { m mol} \; { m m}^{-2}  { m s}^{-1})$	0.37	0.42	0.42	0.42	0.42	0.50	0.50	0.50	0.50

$k_{\text{cat}}^{\text{c}}(\text{s}^{-1})$	2.5	2.8	2.8	2.8	2.8	3.4	3.4	3.4	3.4
$\Gamma$ • (µmol mol <sup>-1</sup> )	43.9	46.9	46.9	46.9	46.9	51.7	51.7	51.7	51.7
$K_{\rm c}$ (µmol mol <sup>-1</sup> )	443	551	551	551	551	760	760	760	760
$K_{\rm o}~(\mu { m mol}~{ m mol}^{-1})$	622014	663007	663007	663007	663007	728467	728467	728467	728467
E (μmol m <sup>-2</sup> )	62.9	62.9	62.9	62.9	62.9	62.9	62.9	62.9	62.9
Activation state (%)	80	80	80	80	80	80	80	80	80
$O$ ( $\mu$ mol mol <sup>-1</sup> )	210000	210000	210000	210000	210000	210000	210000	210000	210000
PPFD (μmol m <sup>-2</sup> s <sup>-1</sup> )	1500	1500	1500	1500	1500	1500	1500	1500	1500
		Modeled o	ptimum val	ues of Rubi	isco kinetic	S			
			$A_{\rm c}$ lim	itation					
A (μmol m <sup>-2</sup> s <sup>-1</sup> )	36	45.6	27	65.3	39.5	43.6	24.3	63.2	37.6
$k_{\text{cat}}^{\text{c}}(\text{s}^{-1})$	3.8	4.5	3.8	5.0	4.3	5.0	4.3	5.6	4.8
$\Gamma$ • (µmol mol <sup>-1</sup> )	49.1	53.6	49.2	56.6	52.4	57.0	52.1	60.3	55.7
$K_{\rm c}$ (µmol mol <sup>-1</sup> )	495	650	535	756	615	823	688	994	782
$K_{\rm o}$ (µmol mol <sup>-1</sup> )	391941	412637	392430	429059	406699	476338	454718	493434	470063
			$A_{\rm c}$ , $A_{\rm i}$ co-	limitation					
A (μmol m <sup>-2</sup> s <sup>-1</sup> )	25	29	23	33	28	31	24	35	29
$k_{\rm cat}^{}$ (s <sup>-1</sup> )	1.5	1.6	2.0	1.3	1.7	2.0	2.7	1.6	2.2
$\Gamma$ • (µmol mol <sup>-1</sup> )	24.5	24.4	31.6	19.3	26.4	28.9	37.7	22.5	31.4
$K_{\rm c}$ (µmol mol <sup>-1</sup> )	230	278	318	258	288	395	465	361	413
$K_{\rm o}$ (µmol mol <sup>-1</sup> )	321876	331167	342939	324723	334177	394348	411100	385212	398657

#### Figure 1 supplemental caption

**Figure 1S.** Optimal values for the maximum turnover rate of carboxylation  $(k_{cat}^{c})$ , specificity factor  $(S_{c/o})$  and carboxylation catalytic efficiency  $(k_{cat}^{c}/K_c)$  at a range of CO<sub>2</sub> concentrations in the chloroplast  $(C_c)$  and three different oxygen concentrations: 1000 (empty triangles), 170 000 (empty circles) and 210 000 μmol mol<sup>-1</sup> (filled circles). The photosynthetic rate was considered to be limited strictly by Rubisco  $(A_c$  limited, left graphs) or co-limited by Rubisco and RuBP-regeneration capacity  $(A_c, A_j$  co-limited, right graphs). The rate of electron transport (74 μmol m<sup>-2</sup> s<sup>-1</sup>), the rate of mitochondrial respiration in the light  $(1.0 \text{ μmol m}^{-2} \text{ s}^{-1})$ , and the concentration of Rubisco catalytic sites  $(30 \text{ μmol m}^{-2})$  and its activation state (80%) were considered invariable at the three oxygen concentrations.

Figure 1S.

