

Cellular Immunity of Patients with Lung Cancer and Other Lung Diseases II. Analysis of interleukin-2 production

Saeko FUJIWARA¹⁾, Mitoshi AKIYAMA^{*2)}, Kyoko KOBUKE^{2,3)},
Masayuki HAKODA²⁾, Michiko YAMADA¹⁾, Hideo SASAKI¹⁾,
Kazuo NERIISHI¹⁾, Nobuoki KOHNO²⁾, Yutaka HOSODA¹⁾,
Masatoshi TAKAISHI²⁾ and Michio YAMAKIDO³⁾

1) *Department of Clinical Studies, Radiation Effects Research Foundation***, 5-2 Hijiyama Koen,
Minami-ku, Hiroshima 732, Japan

2) *Department of Radiobiology, Radiation Effects Research Foundation, Hiroshima 732, Japan*

3) *The Second Department of Internal Medicine, Hiroshima University School of Medicine, 1-2-3,
Kasumi, Minami-ku, Hiroshima 734, Japan*

(Received March 23, 1987)

Key words: Lung cancer, Interleukin-2

ABSTRACT

Interleukin-2 (IL-2) production by stimulated peripheral blood lymphocytes (PBL) from patients with lung cancer and noncancerous respiratory diseases was determined. The results are as follows:

- 1) Neither sex nor age difference was observed for IL-2 production among healthy people.
- 2) IL-2 production showed a positive correlation with the Leu-3a/Leu-2a ratio and a negative correlation with the percentage of HLA-DR⁺ cells.
- 3) IL-2 production of patients with lung cancer and noncancerous respiratory diseases did not differ from that of healthy persons.
- 4) No difference in IL-2 production was found in relation to the clinical stage of lung cancer, but subjects with low IL-2 production were mostly observed in the advanced stage group (Stage IV).

IL-2 is produced by helper T cells¹²⁾ and induces responder T cells to proliferate and differentiate into effector T cells with killer or helper activity¹⁴⁾. Recently, IL-2 has been employed in antitumor immunotherapy, in which killer T cells produced and expanded in *in vitro* with IL-2²⁾ are injected.

However, measurement of IL-2 production in various diseases has just started, and few results have yet been obtained. Thus, we measured the

IL-2 production of peripheral blood lymphocytes (PBL) from lung cancer patients and evaluated and analysed the clinical relationship of the results.

MATERIALS AND METHODS

Sample

The blood samples were obtained from 55 subjects with untreated lung cancer, 53 subjects with benign respiratory diseases and six subjects

* Correspondence to: Mitoshi Akiyama, M.D., Department of Radiobiology, Radiation Effects Research Foundation, 5-2 Hijiyama Park, Minami-ku, Hiroshima 732, Japan

** The Radiation Effects Research Foundation (formerly ABCC) was established in April 1975 as a private non-profit Japanese Foundation, supported equally by the Government of Japan through the Ministry of Health and Welfare, and the Government of the United States through the National Academy of Sciences under contract with the Department of Energy.

with sarcoidosis. All of these individuals were inpatients of either Hiroshima University attached Hospital, Hiroshima Citizens Hospital, Hiroshima Red Cross Hospital, Hiroshima Prefectural Hospital, or Yoshijima Hospital. In addition, 109 healthy RERF staff members were included in these studies.

Methods

After peripheral blood lymphocytes (PBL) were isolated, reactivity to mitogens [phytohemagglutinin (PHA), pokeweed mitogen (PWM), and concanavalin A (ConA)], percentages of lymphocyte subsets using monoclonal antibodies (anti-Leu-1, anti-Leu-2a, anti-Leu-3a, anti-HLA-DR, anti-Leu-11a, and anti-Leu-7), and IL-2 production were measured. Methods of measurement other than that of IL-2 production were described³⁾.

Measurement of IL-2 production

One million lymphocytes from subjects were suspended in 1 ml of test medium consisting of RPMI-1640 with 4 mM HEPES (hydroxyethylpiperazine N'-ethane sulfonic acid), 1% penicillin/streptomycin, 1% L-glutamine, 1% PHA-M (DIFCO Co.), and 2% heat-inactivated fresh human pooled AB serum. After incubation for 24 hours at 37°C in a 5% CO₂ incubator, the supernatant was harvested and stored at -80°C until assay. Culture supernatant was serially diluted twofold in Click's medium and added to wells of microplates in aliquots of 0.1 ml each. To each well, the IL-2-dependent mouse cytotoxic T cell line (CTL-2) cells were added at the rate of 4×10^3 cells/well and cultured at 37°C in a 5% CO₂ incubator for 24 hr. Four hours before harvesting, 0.4 μ Ci of ³H-thymidine (³H-TdR) was added. Cultures were harvested onto glass fiber filter strips, and ³H-TdR incorporation was determined.

Units of IL-2 production were determined by probit analysis⁴⁾. ³H-TdR incorporation data were plotted against log₂ dilutions of each individual IL-2 sample. The control rat IL-2 profile that intercepted the 50% activity was arbitrarily assigned a value of 1 unit, and the IL-2 production of the test sample expressed as units of activity relative to the control rat IL-2 (Fig. 1).

Method of Analysis

After the patients and their normal controls were closely matched by age and sex for com-

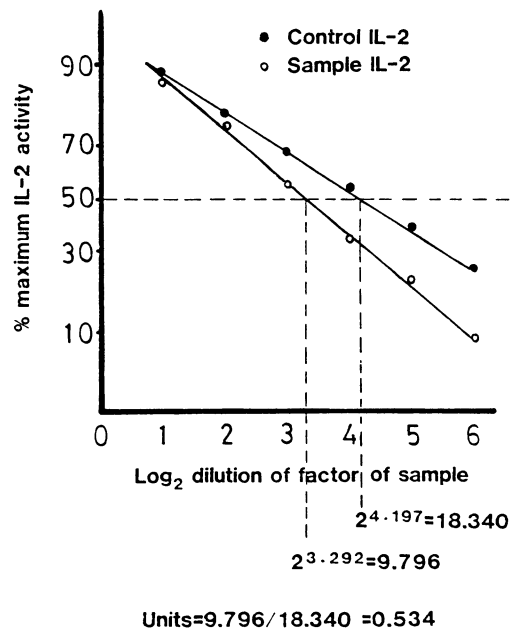


Fig. 1. Determination of units of IL-2 activity

parative analysis, the analysis was made based on 34 lung cancer patients with a mean age of 55.1 ± 6.9 years, 42 normal controls with a mean age of 52.9 ± 6.0 years, 53 noncancerous respiratory disease patients with a mean age of 53.0 ± 16.5 years, and six sarcoidosis patients.

The relationships of IL-2 production to age, sex, reactivity to mitogens, and the percentages of lymphocyte subsets were studied using correlation coefficient in 109 normal controls.

Student's T test were used for comparative analysis on the patients and their normal controls. The differences were considered individually and were judged to be significant when the p-value was 0.05 or less. However, to take into account the number of comparisons made, a difference should be considered statistically significant when the p-value is p/n or less ($p=0.05$, n = the number of tests for that table), and this was done.

RESULTS

1) Relationship of IL-2 production to age, reactivity to mitogens, and the percentages of lymphocyte subsets in healthy persons

As indicated in Table 1, IL-2 production showed a positive correlation with the Leu-3a-to-Leu-2a ratio ($p < 0.05$) and a negative corre-

Table 1. Correlation coefficients between IL-2 production and several immunological parameters in healthy persons (n=109)

	Age dependent	Mitogen response			Lymphocyte subsets					
		PHA	Con A	PWM	Leu-1	Leu-2a	Leu-3a	Leu-3a/ Leu-2a	HLA-DR	Leu-7
IL-2 (units)	0.146	0.097	0.054	-0.037	-0.055	-0.142	0.080	0.213*	-0.193*	0.006

Student's t test; p-values are two-sided and considered for each coefficient individually. * p<0.05

Table 2. Interleukin-2 production in healthy men and women

	Sex	Age			
		Total	<29	30 - 49	50 +
IL-2 (units)	Men	1.00 ± 0.65 (n=63)	0.79 ± 0.53 (n=7)	1.00 ± 0.61 (n=26)	1.04 ± 0.71 (n=30)
	Women	0.91 ± 0.51 (n=46)	0.90 ± 0.45 (n=19)	0.87 ± 0.47 (n=16)	1.00 ± 0.69 (n=11)

Numbers are mean ± standard deviation units.

Student's t-test; comparison made between men and women; p-value are two-sided and considered for each coefficient individually. No significant differences were found.

Table 3. IL-2 production by disease

	n	IL-2 (unit)	
Normal	42	1.11 ± 0.71	
Lung cancer	34	1.31 ± 0.95	NS
Noncancerous respiratory disease	53	1.31 ± 0.84	NS
Sarcoidosis	6	0.74 ± 0.38	NS

Student t-test; comparison made between normal and disease; NS: no significant difference.

lation with the percentage of HLA-DR⁺ cells (p<0.05). It showed no correlation with other parameters.

2) Sex differences in IL-2 production

Healthy persons were divided into three age-groups, under 30, 30-49, and 50 or over. No sex difference was observed for IL-2 production (Table 2).

3) IL-2 production in disease

Table 4. IL-2 production in lung cancer patients by clinical stage

	Normal	Clinical stage of lung cancer patients			
		I	II	III	IV
IL-2 (units)	1.11 ± 0.71 (n=42)	1.18 ± 0.86 (n=20) NS	1.38 ± 0.70 (n=6) NS	1.30 ± 0.90 (n=15) NS	0.97 ± 0.95 (n=14) NS

Numbers are mean ± S.D. units
Student t-test; see Table 3

Table 5. Cases with low IL-2 production (those with IL-2 levels less than 1SD below the mean of the normal cases)

Case	Clinical stage	TNM classification	Histology	IL-2 production (units)
1	IV	T ₃ N ₂ M ₁	Small cell	0.022
2	IV	T ₃ N ₂ M ₂	Squamous cell	0.145
3	IV	T ₃ N ₁ M ₁	Adenocarcinoma	0.192
4	IV	T ₂ N ₁ M ₁	Unknown	0.263
5	III	T ₂ N ₂ M ₀	Adenocarcinoma	0.294
6	III	T ₃ N ₁ M ₀	Squamous	0.369

Table 6. Cases with high IL-2 production (those with IL-2 levels greater than 1SD above the mean of the normal cases)

Case	Clinical stage	TNM Classification	Histology	IL-2 production (units)
1	III	T ₂ N ₂ M ₀	Squamous cell	3.53
2	IV	T ₂ N ₂ M ₁	Adenocarcinoma	3.42
3	Ia	T ₁ N ₀ M ₀	Squamous cell	3.38
4	Ia	T ₄ N ₀ M ₀	Adenocarcinoma	2.73
5	Ia	T ₁ N ₀ M ₀	Adenocarcinoma	2.60
6	IV	T ₂ N ₁ M ₁	Adenocarcinoma	2.49
7	III	T ₁ N ₂ M ₀	Small cell	2.28
8	III	T ₃ N ₂ M ₀	Squamous cell	2.22
9	II	T ₂ N ₁ M ₀	Adenocarcinoma	2.14
10	II	T ₂ N ₁ M ₀	Unknown	2.02
11	III	T ₁ N ₂ M ₀	Adenocarcinoma	1.94
12	Ia	T ₁ N ₀ M ₀	Small cell	1.92
13	II	T ₁ N ₂ M ₀	Adenocarcinoma	1.86

Table 7. Comparison of lymphocyte subsets between the low IL-2 production group and the high IL-2 production group

	Lymphocyte subsets				
	Leu-1	Leu-2a	Leu-3a	Leu-3a/Leu-2a	HLA - DR
Normal (n=51)	63.5 ± 9.8	22.7 ± 6.8	41.4 ± 9.0	1.96 ± 0.86	15.4 ± 6.2
Lung cancer (Untreated)					
Total (n=51)	57.3 ± 13.5	20.6 ± 7.5	38.2 ± 10.8	2.12 ± 0.89	16.2 ± 9.4
Low IL-2 group (n=6)	57.3 ± 9.3	23.2 ± 5.9	34.2 ± 8.6*	1.57 ± 0.66*	20.1 ± 8.1*
High IL-2 group (n=12)	60.3 ± 13.9	19.4 ± 5.2	45.1 ± 8.1*	2.49 ± 0.84*	12.4 ± 3.9*

Numbers are mean ± SD percentage

Student t-test; comparison made between the low IL-2 production group and the high IL-2 production group;

* = p<0.05

IL-2 production of normal persons and patients with lung cancer, noncancerous respiratory diseases, and sarcoidosis was determined (Table 3).

No significant difference was observed in IL-2 production between the normal group and each disease group.

Neither did detailed analysis on the stage of lung cancer show any significant differences in IL-2 production (Table 4). However, the four out of six lung cancer subjects with IL-2 production less than 1 standard deviation lower than the mean of the normal group were in stage IV and the remaining two, in stage III (Table 5). On the other hand, four of the 13 lung cancer subjects with IL-2 production more than 1 standard deviation above the mean of the normal group were in stage I, three in stage II, four in stage III, and two in stage IV. No difference in the distribution was observed by stage (Table 6). Furthermore, the lymphocyte subset percentages were reviewed for six subjects with low IL-2

production and 12 subjects with high IL-2 production. As indicated in Table 7, the low IL-2 production group showed, compared with the high production group, a significantly lower percentage of Leu-3a⁺ cells (p<0.05) and Leu-3a-to-Leu-2a ratio (p<0.05) and a higher percentage of HLA-DR⁺ cells (p<0.05).

DISCUSSION

A number of reports have been published on the effect of aging on IL-2 production. Gillis et al⁵ and Yamakido et al¹³ reported that IL-2 production is impaired in aged people. However, we observed no relationship between age and IL-2 production. This disagreement is due to the age distribution. Gillis et al⁵ examined a group of person 67–81 years of age and Yamakido et al¹³, a group with a mean age of 68.9 ± 8.0 years, while we examined those under 61 years of age. Sex difference was not observed in IL-2 production, a finding which agrees with the result of Yamakido et al¹³.

Autoimmune diseases such as systemic lupus erythematosus (SLE)^{1,6)} and a certain T cell leukemia¹⁰⁾ are well known to show abnormal IL-2 production. Paganelli et al⁹⁾ reported reduced IL-2 production in children with serious immunodeficiency. With regard to IL-2 activity of cancer patients, Nakayama et al⁸⁾ reported that IL-2 activity was much lower in most patients with cancer who had metastasis than in those without metastasis. Similarly, Rey et al¹¹⁾ observed impaired IL-2 production in cancer patients. We did not observe any significant difference in IL-2 production between normal persons and lung cancer patients with or without metastasis. However, subjects with lower IL-2 production were predominantly observed in stage IV. These findings may indicate that the decrease of IL-2 production is related to the tumor progress in cancer patients.

Further, we observed that the percentage of Leu-3a⁺ cells was lower in the lower IL-2 production group than in the higher production group. Reduced IL-2 presumably is due to the paucity of IL-2 producing subsets.

No difference was observed in IL-2 production by peripheral blood lymphocytes between sarcoidosis patients and normal persons in the present study. Hunninghake et al⁷⁾ have reported the presence of large amount of IL-2 production by lung T cells from patients with sarcoidosis and high intensity alveolitis, while detectable amounts of IL-2 were not found in supernatant of blood T cells.

ACKNOWLEDGMENT

The authors are greatly indebted to Drs. Kazumasa Noumi, Tsutomu Inamizu of Hiroshima University Attached Hospital, Drs. Noritomo Senoo and Mitsuhiro Takano of Hiroshima Citizens Hospital, Dr. Masahiro Kuwabara of Hiroshima Prefectural Hospital, Dr. Tatsuya Yoshimi of Hiroshima Red Cross Hospital, and Dr. Hidemasa Maruishi of Yoshijima Hospital for their cooperation in the collection of patient blood and data. We are also grateful to Ms. Kyoko Ozaki, Yoshiko Watanabe, and Noriko Hoshiga of Immunology Laboratory for providing technical support of great value, and to Dr. Michael A Bean for his kind advice.

REFERENCES

1. **Alcocer-Varela, J. and Alarcon-Segovia, D.** 1982. Decreased production of and response to interleukin-2 by cultured lymphocytes from patients with systemic lupus erythematosus. *J. Clin. Invest.* **69**: 1388–1392.
2. **Cheever, M.A., Greenberg, P.D. and Fefer, A.** 1981. Specific adoptive therapy of established leukemia with syngeneic lymphocytes sequentially immunized in vivo and in vitro and nonspecifically expanded by culture with interleukin 2. *J. Immunol.* **126**: 1318–1322.
3. **Fujiwara, S., Akiyama, M., Kobuke, K., Ichimaru, S., Takaishi, M., Inamizu, T., Kouno, N., Takami, S., Noumi, K., Kawamoto, M. and Yamakido, M.** 1986. Cellular immune competence of patients with lung cancer and other lung diseases. I. Analysis of peripheral blood lymphocyte subsets using monoclonal antibodies. *Hiroshima J. Med. Sci.* **35**: 227–236.
4. **Gillis, S., Ferm, M.M., Ou, W. and Smith, K.A.** 1978. T cell growth factor: Parameters of production and a quantitative microassay for activity. *J. Immunol.* **120**: 2027–2032.
5. **Gillis, S., Kozak, R., Durante, M. and Weksler, M.E.** 1981. Immunological studies of aging: Decreased production of and response to T cell growth factor by lymphocytes from aged humans. *J. Clin. Invest.* **67**: 937–942.
6. **Horwitz, D.A., Linker-Israeli, M., Bakke, A.C., Kitridou, R.C., Gendler, S. and Lemoine, C.M.** 1982. Defective interleukin-2 production in systemic lupus erythematosus: Relationship with T helper cell lymphopenia and depressed interleukin-1 activity. *Arthritis Rheum.* **25**: S28
7. **Hunninghake, G.W., Bedell, G.N., Zavala, D.C., Monick, M. and Brady, M.** 1983. Role of interleukin-2 release by lung T cells in active pulmonary sarcoidosis. *Am. Rev. Respir. Dis.* **128**: 634–638.
8. **Nakayama, E., Asano, S., Takuwa, N., Yokota, J. and Miwa, S.** 1983. Decreased TCGF activity in the culture medium of PHA stimulated peripheral mononuclear cells from patients with metastatic cancer. *Clin. Exp. Immunol.* **51**: 511–516.
9. **Paganelli, R., Aiuti, F., Beverley, P.C.L. and Levinsky, R.J.** 1983. Impaired production of interleukins in patients with cell-mediated immunodeficiencies. *Clin. Exp. Immunol.* **51**: 338–344.
10. **Poiesz, B.J., Ruscetti, F.W., Mier, J.W., Woods, A.M. and Gallo, R.C.** 1980. T-cell lines established from human T-lymphocytic neoplasias by direct response to T-cell growth factor. *Proc. Natl. Acad. Sci. USA.* **77**: 6815–6819.
11. **Rey, A., Klein, B., Zagury, D., Thierry, C. and Serrou, B.** 1983. Diminished interleukin-2 activity production in cancer patients bearing solid tumors and its relationship with natural killer cells. *Im-*

- munol. Letters **6**: 175–178.
12. **Shaw, J., Caplan, B., Paetkau, V., Pilarski, L.M., Delovitch, T.L. and McKenzie, I.F.** 1980. Cellular origins of co-stimulator (IL 2) and its activity in cytotoxic T lymphocyte response. *J. Immunol.* **124**: 2231–2239.
 13. **Yamakido, M., Yanagida, J., Ishioka, S., Matsuzaki, S., Hozawa, S., Akiyama, M., Kobuke, K., Inamizu, T. and Nishimoto, Y.** 1985. Interleukin-2 production and lymphocyte proliferation in aged and young humans. *Hiroshima J. Med. Sci.* **34**: 95–99.
 14. **Wagner, H., Hardt, C., Heeg, K., Rollinghoff, M. and Pfizenmaier, K.** 1980. T-cell-derived helper factor allows in vivo induction of cytotoxic T cells in nu/nu mice. *Nature* **284**: 278–280.