Sexual selection explains more functional variation in the mammalian major histocompatibility complex than parasitism

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Understanding drivers of genetic diversity at the major histocompatibility complex (MHC) is vitally important for predicting how vertebrate immune defencemightrespondtofutureselectionpressuresandforpreservingimmunogenetic diversity in declining populations. Parasite-mediated selection is believed to be the major selective force generating MHC polymorphism, and whileMHC-basedmatingpreferencesalsoexistformultiplespecies including humans, the general importance of matechoice is debated. To investigate the contributions of parasitism and sexual selection in explaining among-species variation in MHC diversity, we applied comparative methods and metaanalysis across 112 mammal species, including carnivores, bats, primates, rodentsandungulates. Wetested whether MHC diversity increased with parasiterichness and relative testes size (as an indicator of the potential for mate choice), while controlling for phylogenetic autocorrelation, neutral mutation rate and confounding ecological variables. We found that MHC nucleotide diversityincreasedwithparasiterichnessforbatsandungulatesbutdecreased withparasiterichnessforcarnivores. By contrast, nucleotided iversity increased with relative testes size for all taxa. This study provides support for both parasite-mediated and sexual selection in shaping functional MHC polymorphism across mammals, and importantly, suggests that sexual selection couldhaveamoregeneralrolethanpreviouslythought.

Subject Areas:

evolution, molecular biology, ecology

Keywords:

major histocompatibility complex, mate choice, parasite-mediated selection, parasite richness, phylogenetic comparative methods, meta-analysis

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1. Introduction

Asignificant fraction of the mammal genome is dedicated to immune defence, and immune genes are well known for their genetic variability [1,2]. Parasites havelong been viewed as a major selective force in shaping host genetic diversity [3,4], and the rate of adaptive evolution for genesthat interact most directly with pathogens can be exceptionally high [5]. Sexual selection can also influence immunogenetic variation; in particular, the 'good genes' hypothesis for resistance to parasites has been invoked to explain why some animals hold mating preferences in the absence of direct benefits of being choosy [6]. Thus, direct effects of parasites on host fitness, combined with sexual selection for mates that might confer beneficial genes to progeny, are the two most likely forces shaping immunogenetic diversity in animals.

Themajorhistocompatibilitycomplex(MHC) is an ideal candidate for identifying factors that determine immune genediversity, because it plays a crucial role in immune defence for virtually all vertebrates and can mediate mate choice in a variety of species, including humans [7,8]. The MHC encodes gly coproteins that bind to foreign antigens and present them to T-cells, initiating an immune response [9]. There are two major groups of MHC genes: class I responds to intracellular pathogens and class II interacts with extracellular

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pathogens [10]. In particular, the class IIDR Blocus has been extensively studied because of its high allelic diversity, and both diversity and specifical leles at this locus predict parasite resistance in an imals [7]. The DR Bexon 2 region encodes the functionally important antigen-binding sites (ABS) that recognize pathogen peptides, with evidence of intense positive selection at codons along the sequence [10]. Because different ABS bind to different pathogen proteins, multiple alleles are required to confer resistance to diverse pathogen strains and species [9].

Past work showed that even endangered species (which otherwiseharbourextremelylowdiversitybasedonselectively neutralloci)candisplayhighMHCgenotypicdiversity[11–13] withsuchobservationsattributedtostrongbalancingselection operating on MHCloci [3,7]. Despite the potential for universally strong selection on MHC genes across vertebrate taxa, species do differ in their levels of MHC variability [14]. Some of this variation can be explained by differential parasite pressure across species [15], but mechanisms underlying among-species variation in MHC have rarely been studied in acomparative sense (but see [15–17]).

Theorysuggeststhatdisassortativemating could also preserve allelic diversity across MHC loci [18], and numerous species, including rodents, fishes, birds and humans can discern MHC genotypes based on olfactory and other cues [19–21], and prefer scents of mates with complementary or dissimilar MHC genotypes [22,23]. A key challenge facing researchers studying matechoice and MHC is that ecological and demographic factors influence the opportunity for and benefits of being choosy [24]. As a result, most studies showing MHC-based mating preferences within species are based on laboratory or captive experiments, and studies conducted on wild populations have shown mixed results [25,26].

Here, we use a comparative approach to investigate the relative influence of the two proposed main selective forces on MHC polymorphism: parasite-mediated selection and sexual selection. Our analysis focuses on mammals from five orders, as mammals have been relatively well studied for MHC variation, parasites and infectious diseases, and traits associated with sexual selection and reproductive skew. Key questions motivating this study were: (i) how does MHC diversity vary across mammal groups? (ii) are measures of allelic and nucleotide diversity elevated in species with higher potential for mate choice? (iii) is MHC diversity also elevated in species with greater parasite richness? and (iv) is there an association between parasite richness and degree of sexual selection, and how might this interaction shape the relationship with MHC polymorphism? We estimated the potential for sexual selection using relative testessizeasaproxyforcompetitionamongmalestoproduce offspring, as past work showed this measure is greater in species with promisc uous or polygynous (as opposed to mon-species with promisc uous or polygynous), and the promisc uous or polygynous (as opposed to mon-species with promisc uous or polygynous).ogamous)matingsystems[27-29].Toinferselectionpressures exerted by diverse parasite communities, we augmented existing data on parasites and pathogens (including viruses, bacteria, protozoa, helminths and arthropods) from free-living mammal populations [30-32]. We used phylogenetically informedanalysestotestkeypredictorsofimmunegenesdiversity across species, controlling for the potential effects of host phylogeny, ecological traits and uneven sampling effort. We also used meta-analyses to compare effect sizes across taxa and to better support the generality of the findings. To our knowledge, this work represents the first test of the importance

of sexual selection for explaining immunogenetic variation acrossawiderangeofmammals.

2. Material and methods

(a) Major histocompatibility complex data

Sequence data for 112 mammal species were compiled from GenBankusingG ENEIOUS v.5.6.3. Wefirstperformedapreliminary searchwiththekeyterm'MHCclassIIDRB', recoveredallmammal sequences and retained sequences including exon 2 of the DRB locus. We also searched on Web of Science and Biosis using each previously identified species Latin binomial and MHC as key terms. Sequences from subspecies were combined at the species level, and we followed the taxonomy of Wilson & Reeder [33]. Primate taxonomy followed the nomenclature from the Global Mammal Parasite Database [30] and the dataset from Garamszegi & Nunn [15] to correspond with parasite data. For each species, we recorded the number of animals sampled at the DRB locus because more alleles tend to be discovered as more individuals are sequenced.

Sequences were grouped according to Order (Carnivora, Chiroptera, Primate, Rodentia and Artiodactyla), imported into MEGA v.5[34]andalignedbyMUSCLE[35].Becausesequencesdiffered in length, we trimmed all exon 2 sequences to 171bp to estimate substitution rates. We removed pseudogenes and alleles with nucleotide insertions or deletions that might represent nonfunctioning alleles. We also removed DRB6 alleles from primates, as this locus is thought to be non-functional [36]. We checked for duplicates within species and removed non-unique sequences. Final numbers of sequences were recorded as numbers of alleles per species. For analyses of allelic richness, we used residuals from a regression analysis of log (number of animals sampled) to control for uneven sampling across species.

Rates of selection for functional variation is a biologically important measure of diversity, especially for sites that encode proteins responsible for binding to foreign peptides (ABS [10]). To estimate substitution rates, we used the most commonly used method [37] with correction for multiple substitutions at thesamesite[38].M EGA v.5wasusedtocomputewithin-species averagesforaminoacidchangingnon-synonymoussubstitutions (dN) at 15 ABS based on [39]. We repeated this process for synonymous substitutions (d S) at ABS to provide a baseline for neutral substitution rates. We avoided using the ratio d N: dS at ABS as in [15], as correlations with ratios may be more difficult to interpret, being influenced by both the nominator and denominator. However, we also ran all analyses using this ratio, and results were generally consistent (see the electronic supplementary material, figures S1 and S2, though power to detect significant associations was reduced owing to some species having no synonymous substitutions at the ABS). We considered using alternative ABS sites determined using the consensus of codon-based maximum-likelihood methods [40] applied to each main order sequence set (e.g. carnivore, bat, primate, rodent and ungulate). However, these predicted sites were strongly and significantly correlated with the 15 peptide-binding region residues determined based on protein crystallography (Pearson's correlation ranged from r(29) = 0.49, p < 0.01, primates, to r(12) = 0.93, p < 0.001, rodents). In addition, these 15 sites are known to be involved with antigen binding and have been shown to be under positive selection across a diverse set of taxa (carnivores, rodents and primates [41]; bats [42]). Therefore, we focus analyses on these documented 15 ABS sites, though analyses with putative taxonspecific ABS sites show over all consistent results (see the electronic specific ABS sites show over all consistent results).supplementary material, figure S3). The number of MHC alleles varies by allelic lineage [43] and by the number of duplicated DRBloci [44]. Because most species in our dataset are non-model organisms and because no information is available on the specific

DRB lineage or genecopy number, we could not include these variables in our analysis. We compiled the number of recorded DRB locifore a chof 61 mammal species for follow-up analyses.

(b) Parasite data

Hostsexposedtoamorediverseparasiteassemblagecouldexperience selection for greater genetic diversity for resistance [16]. Because MHC class II genes recognize extracellular parasites, theremight bestronger relationships between parasites with prominent extracellular stages (such as helminths, arthropods and some microbes) and measures of DRB diversity. We compiled parasiter ichness data for each host species using the Global MammalParasiteDatabase(www.mammalparasites.org),themostcomprehensivecollectionofpublishedrecordsofparasiticorganismsfrom free-livingmammals[30]. For each host species, we recorded parasite richness as the total number of viruses, bacteria, protozoa, helminths and parasitic arthropods, as defined at the species-levelbasedoncurrenttaxonomicschemes. Wealsorecordedseparately thenumbers of helminths (thought to have strong coevolutionary relationships with their hosts), microparasites (viruses, bacteria and protozoa) and macroparasites (helminths and arthropods) to test whether some groups were more strongly associated with MHC class II diversity than others. We could not examine all parasitesubgroups(e.g.virusesandbacteria)individuallyowingtolow numbersforsomehosttaxa.

Parasiterichnessestimates dependstrongly on research effort [45]; better-studied host species tend to have more parasites reported to infect them. We therefore controlled for uneven sampling effort among hosts using the total number of citations for each host species (using Latin binomials and common taxonomic variants) from Web of Science as an indicator of scientific effort perhost. Following previous studies [15,31,46,47], we used citation counts to control for research effort instead of the cumulative number of individual hosts sampled across all studies, as some studies did not publish the number of an imals sampled, and other studies had high sample sizes despite testing for only a single parasite. We used residuals from a regression analysis of log (parasite richness) against log (citation count) ($R^2 = 0.45, \, F_{1.96} = 78.29, \, p < 0.001$) to estimate corrected parasite richness perhost.

(c) Estimates of sexual selection and ecological traits

Genetic mating system, and specifically the potential for female matechoice(asfemalestendtobethechoosiersex[48]), is expected to influence the strength of sexual selection on the MHC. Females inmonogamous or polygy nous mating systems are likely to be moreconstrained in their choice for mates that can provide direct or indirectbenefits[49]. By contrast, females in promiscuous or polyand rous mating systems are expected to have greater opportunity toselectamong potential mates. Relative testes size (testes mass/ bodymass)wasusedasaproxyforfemalepromiscuityandopportunities formate choice, as this measure has been shown to predict sexualselectionandmatingsystemacrossmammals(e.g.primates [27]; rodents [28]; carnivores [29]) and it is available for a large number of species. We compiled testes mass data and male body massfromtheliterature(seetheelectronicsupplementarymaterial, datasetS1). Insomeinstances, only testes length, circumference or volumemeasurementswereavailable, and in those cases, we converted these data to mass using the method of [27]. We then used the residuals from a regression analysis of log (testes mass) againstlog(malebodymass)to obtain relative testes size perspecies and performed this separately for each taxonomic group.

Foreachmammalspecies, we also compiled data on two variables that could strongly influence parasitism, mating behaviour, and/or MHC diversity and evolution. First, effective population size ($N_{\rm e}$) can impact genetic diversity by affecting the realized mutation rate, strength of selection and the amount of genetic drift experienced by a population [50]. $N_{\rm e}$ can also influence

parasiterichness measures as larger populations can theoretically retainmore parasites than smaller populations [51,52]. In addition, species with greater population density might har bour more parasites with density-dependent transmission [53], and species with largergeographicalrangesizescouldencounteragreaterdiversity ofparasites[31,32]. Weused census population size as a proxy for effective population size, with the caveat that the ratio of approximately 0.1 on average [54]. Population size was estimated by multiplying average population density (individuals per km from the PanTHERIA database [55] by the species geographical range size (km²) extracted from spatial data provided by the 2010 International Union for Conservation of Nature Red List (http://www.iucnredlist.org/technical-documents/spatial-data#mammals, last accessed on 6 July 2012). Previous studies have shownthatthismeasureofpopulationsizeisasignificantpredictor ofboth N_e [15,56] and parasiterichness [52]. Second, body massis knowntoscalewithmanylife-historytraits, including population size,reproductiverate,evolutionaryrate[57]andparasitediversity [31,32,52], and thus was included as a covariate. Bodymass (g) data were extracted from PanTHERIA [55], or when not available, the primary literature. Our full comparative dataset can be found in theelectronic supplementary material, \$1. Variables were log transformedorsquarerootarcsintransformed(d S ratedata;[58])when necessary to meet normality assumptions, and no predictor variables showed strong collinearity (r < 0.07)[59].

(d) Comparative analyses

We tested whether log MHC all elic diversity and rates of positiveselection(d N)atfunctionalsites(astwoseparatedependentvariables) were associated with parasite richness and estimates of sexual selection. Because closely related species are more likely to share genetic and life-history traits [60], we used phylogenetic least-squares (PGLS) regressions that handle phylogenetic structurethroughavariance-covariancematrix[61]. Weassessed the importance of each predictor through a stepwise model selection procedure, in which the full models included the following: corrected parasite richness, relative testes size, log body mass, log population size, and the interaction between corrected parasite richness and relative testes size, while also including d asacovariatetocontrolforunderlyingsubstitution rate. We then simplified the starting model using Akaikein formation criteria correctedforsmallersamplesizes(AICc)following[62]andremoved variables that did not improve model fit (Δ AICc > 4). To avoid problems associated with missing values in AIC-based model commodel commodel and the commodel commodel commodel and the commodel commodelparison where sample size changes as terms are removed, we removedanyspecies from our dataset that did not have complete coverageforallofthepredictorvariablesofinterestinthestarting fullmodel. Totestforassociations between relative testessize (asa responsevariable) and corrected parasiterichness (predictor variable), we ran PGLS models controlling for the effects of taxonomicgroup, logpopulationsize and malebody mass.

The PGLS regression was conducted using the caper package in R[63] using Pagel's λ to adjust models for the amount of phylogenetic signal observed in each variable. The phylogeny was constructed using the mammalian supertree [64], and polytomies were randomly resolved (by adding branches of length equal to zero) using the multi2di function in the ape R package [65]. Species in the dataset but missing from the supertree assumed the names of the closest relatives (i.e. Papio cynocephalus was changed to Papiohamadryas, and Zalophus wollebaeki was changed to Zalophus californianus).

To test the robustness of our results, we estimated taxon-specificeffectsizes in the form of correlation coefficients [66] and ranmeta-analyses to compare the overall effect sizes for predictor variables across taxa in explaining MHC variation. We estimated grand classwise effect sizes after considering differences in sample size among orders, and also examined differences among

0.77 (0.20, 1.33)

0.22(-0.32, 0.77)

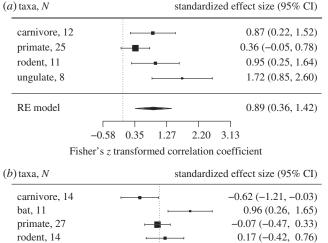
3. Results

Ourfinaldatasetcomprised112mammalspecies(26carnivores; 14chiropterans;37primates;16rodentsand19ungulates),2454 sequences and 2665 host-parasite species combinations (list of species with full trait and genetic datasets are provided in the electronic supplementary material, dataset S1 and S2). Wetestedforrelationshipsbetweenparasitism(usingtotalparasiterichness, helminthrichness, and micro-and macroparasite richness) and sexual selection (using relative testes size as an indicator of mating system) on the rate of positive selection (dN at ABS) and MHC allelic richness across mammals. Analyses controlled for mammal phylogeny, the rate of neutral substitutions(d S at ABS), measures of sampling effort and two hosttraitsknowntobeimportantforparasitismand/orgenetic diversity based on previous studies (body mass and population size; electronic supplementary material, tables S1-S3). All predictorandresponsevariablesexceptrelativeallelicrichness and parasite richness showed strong phylogenetic signal and were more similar among closer relatives (see the electronic supplementarymaterial,tableS2).

Multivariatemodelsshowedthatthestrengthofselection at ABS increased with relative testes size across all five mammalorderstestedhere(figure1 a and the electronic supplementary material, table S1). Effect sizes differed among orders with approximately 64% of the variability attributed to heterogeneity among the true effects [67] (p = 0.04, $I^2 = 63.84\%$), possibly owing to biological differences among mammal groups or the methodology used in different studies. When corrected parasite richness was included as a predictor in the full model with relative testes size, its main effect and interaction with relative testes size was non-significant. However, removing the relative testes size variable (and including 21 additional species that were missing testes data) showed that the strength of selection at ABS increased with total parasite richness (corrected for sampling effort) for bats and ungulates and decreased with parasite richness for carnivores (figure 1 b and the electronic supplementary material, table S1).

Tests using data from parasite subgroups, including microparasites, macroparasites and helminths, showed that selection on ABS decreased with helminth and macroparasite richnessforcarnivores, and also decreased with microparasite richness for primates (figure 2; electronic supplementary material, figure S2). Ratios of N: dS increased with macroparasite and microparasite richness for carnivores and ungulates (see the electronic supplementary material, figure S2). Neutral substitutionrate(d S at ABS) was a positive predictor of d N at ABS(seetheelectronicsupplementarymaterial, figureS4) but onlyinmodels without relative testes size (and larger sample sizes). Taxonomic group was also a significant predictor of allelicsubstitutionatABS,withbothd N andd S beinggreatestfor bats and primates and lowest for carnivores and ungulates (figure 3c).

Wetestedallelicrichnessasaseparatemeasureofselection on ABS. This measure differed among mammal taxonomic groups and increased with population size for ungulates (electronic supplementary material, tables S1 and S3e) but did not depend on measures of parasite richness or testes size.



0.22 Fisher's z transformed correlation coefficient

1.22

2.22

-1.78

Figure 1. Forest plots showing predictors of the rate of non-synonymous sub $stitutions (d\,\textit{N}\,) at \, ABS. PGLS models show the effect of ($ a) relative testes size and (b) parasite richness, run separately for each mammal group. The vertical dotted line is positioned at zero and error bars denote 95% Cls.N referstothenumber of species included; REmodel, random effects model. Heterogeneity test for relative testes size: Q = 8.433, p = 0.038, $I^2 = 63.84\%$; parasite richness: Q =17.408, p = 0.002, $I^2 = 79.06\%$.

Allelicrichnesswassignificantlylowerforungulatesthanany othermammalgroup(figure 3 b). Aposthoc ANOV Arevealed that ungulates also had significantly fewer duplicated DRB locithan other mammal orders, whereas primates had the most $(F_{1,39} = 8.359, p = 0.006; figure 3 a).$

Finally, we tested for a relationship between relative testes size and corrected parasiterichness, to ask whether the strengthof sexual selection might covary positively with parasite pressure. Across all orders, we found a weak negative association between relative testes size and total parasite richness (p = 0.09; electronic supplementary material, figure S5a) with homogeneous effect sizes across taxa (Q = 1.079, p = 0.18, $I^2 = 0.00\%$), although other components of parasite richness showed notrend (p > 0.1; electronic supplementary material, figureS5).

4. Discussion

ungulate, 15

RE model

Wefoundevidencethatpressurefromadiverseparasitefauna (represented by corrected parasite richness at the host species level) was associated with positive selection at the MHC DRB locus in bats and ungulates only. Species in these two groups that harboured greater parasite richness also showed higher rates of functionally significant evolutionary changewithintheMHC.Bycomparison, greater potential for sexual selection (represented by relative testes size as an indicatorofmatingsystem)predictedgreaterpositiveselectionon functionally important MHC sites across all five orders of mammalsexaminedhere.

Veryfewstudieshaveconsideredpredictorsofcross-species variation in MHC polymorphism, and those conducted to datefocused on the relationship between parasite richness and MHC

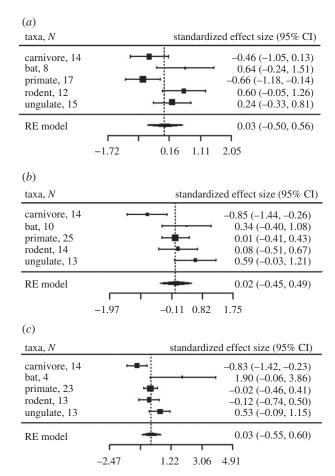


Figure 2. Forest plots showing predictors of the rate of non-synonymous substitutions at ABS. The results of PGLS models showing the effect of (a) microparasite richness, (b) macroparasite richness and (c) helminth richness, runseparately foreach host and parasite group. Nometa-analysis random effects model was found to be significant for the parasite predictor variables (p>0.1). The vertical dotted line is positioned at zero for Fisher's z transformed correlation coefficient. N refers to number of species; RE model, random effects model. Heterogeneity test for microparasite richness: Q=5.186, p=0.159, $I^2=43.24\%$; macroparasite richness: Q=6.506, p=0.089, $I^2=55.08\%$; helminth richness: Q=4.329, P=0.228, $I^2=0.00\%$.

allelic richness [15–17]. In spite of the extensive intraspecific empirical evidence for MHC–parasite associations (reviewed in [7,69]), there is surprisingly weak support for parasites driving variation in MHC diversity among species. One study found that allelic richness increased with helminth richness across 10 rodent species [16], and another study found that the rate of positive selection at ABS (but not allelic richness) was positively related to nematoderichness (but not to talparasite richness) across 27 primates [15]. Our study differed from prior studies by using genetic data from multiple populations perspecies, by including broader taxonomic groups in parasite richness estimates, by not delineating alleles by their functional lineages, and by not separately analysing nematode richness (for which in sufficient data were available from allorders).

The negative relationship between parasite richness and measures of the strength of selection on MHC in carnivores ran counter to our expectations. One reason for this pattern could be due to interactions between threat status and infectious disease risk, such that carnivores might be more vulnerable to population bottlenecks and genetic drift that concurrently reduce their genetic diversity and increases theirs usceptibility to parasitism. Manyhigh-profile threatened

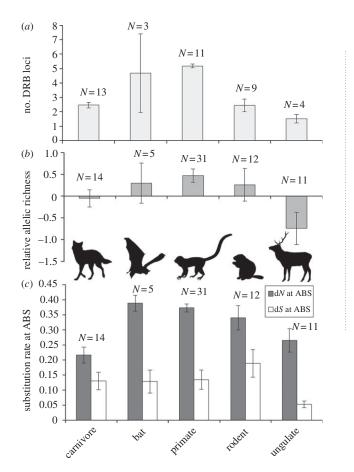


Figure 3. Genetic diversity at the MHC by mammal group. (a) The number of DRB loci, (b) relative allelic richness and (c) the rate of non-synonymous (dN) and synonymous substitutions (dS) at the ABS for carnivores, bats, primates, rodents and ungulates. Error bars denote 95% Cls. N refers to the number of species with data.

carnivores have depleted MHC diversity (wild dogs [70]; Ethiopianwolves[71]; cheetahs[72]) and have simultaneously experienced declines from introduced in fectious diseases such as rabies, canine distemperands are optic mange [73,74]; these carnivore species might be exceptionally well studied and better represented in our dataset. One way to examine this issue further might be to include estimates of effective population size in comparative analyses and to distinguish between native versus introduced parasites and pathogens. Analternative explanation is that different taxa are indifferent stages of the coevolutionary arms race. If parasites lead the game, evidence might support parasitis mass adriver of MHC polymorphism (leading to a positive relationship). However, if hosts lead the game, greater MHC diversity might reduce parasite pressure (leading to an egative relationship).

In contrast to the taxon-specific evidence of parasite-mediated selection, we found that relative testes size, as an indicator of sperm competition and the potential for sexual selection to operate at the species level [27], was positively associated with the rate of evolution at ABS across all mammal groups in our study. This finding provides evidence that species with high potential for mate choice tend to have higher MHC nucleotide diversity at functionally important sites. There are several non-exclusive explanations for this result. First, species with greater relative testes size and sperm competition might have faster reproductive rates, increasing the speed of selection for new variants. Indeed, Sommer *etal*. [75] foundhigher levels of MHC variation in a

fast-reproducing and promiscuous rodent relative to a monogamous and slower reproducing relative, and hypothesized that slow reproduction might constrain MHC polymorphism.A second hypothesis is that greater sperm competition indicates greater promiscuity and increased exposure to sexually orsociallytransmitteddiseases, which could enhance selection on immune defences [76,77]. As a third possibility, females with more potential mates might select genetically complementary or non-related mates and by doing so, serve to increaseMHCvariability. A fourthhypothesis is that relative testes size is correlated with androgen levels [78], which can suppress immune function or mediate male behaviour and increase exposure to and selection by parasites [79,80]. Our finding of a weak negative relationship between parasiterichness and relative testes size, however, is not consistent with thishypothesis. Importantly, each of these mechanisms predicts that greater promiscuity will lead to greater genetic diversity, are sultalready observed for MHC and neutral genetic diversity across passerine birds [81]. Our analysis did not support an interactive effect between parasite richness and relative testes size, a smight be expected if both high parasite pressureand the potential formate choice were necessary to drive highMHCdiversity.

Our study strongly supported taxonomic group as an important predictor for MHC allelic and sequence diversity. Specifically, ungulates had significantly lower allelic richness than any other order, possibly owing to fewer duplicated DRBloci(figure2 a). Primates, incomparison, had significantly greaterallelic richness and more duplicated DRBloci. Average nucleotide divergence (π) is positively associated with the number of duplicated DRBloci in rodents [44] and this could be an important mechanism providing baseline genetic variation. Life-history traits or ecological conditions that affect the likelihood of MHC geneduplication events might therefore help predict MHC polymorphism in natural populations.

Overall, our study extends previous comparative work on MHC evolution by showing that both parasite-mediated selection and sexual selection can operate as independent forces maintaining differences in MHC diversity across mammal species. Evidence that parasites served as agents of selection was only found for bats and ungulates, but support for sexual selection was universal across mammal groups tested here. Potential explanations for this pattern include greaters election on immune genes driven by higher pressurefrom socially or sexually transmitted disease, and greater opportunities formate choice leading to faster rates of substitution. Importantly, our analyses emphasize that comparativestudies can contribute to knowledge on MHC ecology and evolution. We expect results of this study will encourage more work on the influence of sexual selection on MHC variabilityinwildpopulations, with great relevance for conservation genetics and predicting species responses to future diseaserisk.

Acknowledgements. Wethank B. Hanand C. Nunn for access to parasite data, J. Wares, J. Carroll, J. Moore, M. Yabsley, V. Ezenwa, J. Rushmore, D. Streicker, the Altizer Laboratory group, M. MacManes and an anonymous reviewer for discussion and comments that greatly improved the previous manuscript. J. C. W., L. Z. G. and S. A. designed the research; J. C. W., S. G. M., S. H. and P. R. S. compiled data; J. C. W. and L. Z. G. analysed data; and J. C. W., L. Z. G., S. H., P. R. S. and S. A. wrote the paper.

Funding statement. The Anthro Treeworkshop (supported by NSF grant nos. BCS-0923791 and EF-0905606) provided guidance on phylogenetic comparative analyses. The Odum School of Ecology, National Geographic Society, Animal Behavior Society, Association for Women in Science and the Philosophical Society provided financial support. J.C.W. was supported by a Graduate School Assistantship from the University of Georgia and a T-32 Training grant from the National Institutes of Health. L.Z.G. was supported by the 'Plan Nacional' program of the Spanish government (ref. no. CGL2012-40026 and CGL2012-38262). S.A. was supported by funding from the NSF (DEB-0643831).

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