

1 **Molecular cloning of IGF-1 and IGF-1 receptor and their expression pattern in the**
2 **Chilean flounder (*Paralichthys adspersus*)**

3

4 Sebastian Escobar^{1a}, Eduardo Fuentes¹, Erika Poblete¹, Juan A. Valdés¹, Ariel E.
5 Reyes^{2,4}, Marco Álvarez³, Alfredo Molina^{1,4,*}.

6

7 ¹Universidad Andres Bello, Facultad de Ciencias Biológicas, Laboratorio de
8 Biotecnología Molecular. Av. República 217, Santiago, Chile.

9 ²Universidad Andres Bello, Facultad de Ciencias Biológicas, Laboratorio de Biología
10 del Desarrollo. Av. República 217, Santiago, Chile.

11 ³Universidad Andres Bello, Facultad de Ciencias Biológicas, Laboratorio de Biología
12 Celular y Molecular. Av. República 217, Santiago, Chile.

13 ⁴Millennium Institute for Fundamental and Applied Biology, Santiago, Chile

14

15 ^a Present address: Consejo Superior de Investigaciones Científicas (CSIC), Instituto de
16 Acuicultura de Torre de la Sal (IATS), Ribera de Cabanes 12595 Castellón, Spain

17

18

19

20

21

22 *Corresponding author. Tel.: +56 2661 8319; fax: +56 2661 8415.

23 *E-mail address:* amolina@unab.cl (A. Molina).

24

25

26 **Abstract**

27 Insulin-like growth factor-1 and insulin-like growth factor-1 receptor (IGF-1 and
28 IGF-1R) play main roles in vertebrate growth and development. In fish, beside
29 contributing to somatic growth, both molecules exhibit pleiotropic functions. We
30 isolated complete cDNAs sequences encoding for both IGF-1 and IGF-1R in the
31 Chilean flounder by using RT-PCR and rapid amplification of cDNAs ends (RACE)
32 techniques. In addition, we analyzed gene expression in pre-metamorphic larvae and
33 different organs of adult fish through whole mount *in situ* hybridization and RT-PCR,
34 respectively. The IGF-1 cDNA sequence displays an open reading frame of 558
35 nucleotides, encoding a 185 amino acid preproIGF-1. Moreover, IGF-1R contains an
36 open reading frame spanning 4,239 nucleotides, rendering a 702 amino acid subunit
37 alpha and a 676 amino acid subunit beta. The deduced mature IGF-1 and IGF-1R
38 exhibited high sequence identities with their corresponding orthologs in fishes, specially
39 those domains involved in biological activity. RT-PCR showed expression of IGF-1 and
40 IGF-1R transcripts in all studied tissues, consistent with their pleiotropic functions.
41 Furthermore, we observed a strong IGF-1 expression in notochord in larvae of 9 days
42 post fertilization. Similarly, IGF-1R transcripts were observed in larvae of 9 days post
43 fertilization, in territories such as notochord, somites and head. Interestingly, both
44 mRNAs were detected in territories such as notochord, an embryonic midline structure
45 essential for the pattern of surrounding tissues as nervous system and mesoderm. Our
46 results suggest that IGF-1 and its receptor could have an important role in the
47 development of the nervous system, muscle and bone-related structures during larval
48 stages. The present data contributes to the knowledge of the insulin-like growth factor
49 signaling components in an emergent and new commercial important marine fish.

50 **Keywords:** IGF-1, IGF-1R, notochord, somites, Chilean flounder.

51 **Introduction**

52 The insulin-like growth factor signaling system plays an important role in
53 promoting the embryonic growth and development in vertebrates (Moriyama et al.,
54 2000). This pathway involves the coordinated function of two ligands, two cell surface
55 receptors and at least six high affinity binding proteins (Moriyama et al., 2000; Wood et
56 al., 2005). The biological effects of the IGF system are mediated mainly by the
57 interaction of IGF-1 ligand with IGF-1 receptor (IGF-1R) modulated through IGF
58 binding proteins (IGFBPs) (Riedemann and Macaulay, 2006). IGF-1 is synthesized as a
59 pre-pro-hormone, which undergoes at least two processing events: cleavage of the
60 signal peptide and the C-terminal peptide (Etherton, 2004; Le Roith et al., 2001). The
61 mature IGF-1 is a single chain polypeptide composed of 70 amino acids, which contains
62 domains A and B separated by a C domain, and a D carboxy domain (Humbel, 1990).
63 The IGF-1 receptor is synthesized as a single chain pre-pro-receptor, with a 30 residues
64 signal peptides that is co-translationally cleaved and a 1,337 amino acids pro-receptor
65 that is processed at a tetrabasic cleavage site to generate alpha and beta subunits
66 (LeRoith et al., 1995). The mature IGF-1R is comprised by two alpha subunits and two
67 beta subunits linked by disulphide bonds, forming $\alpha_2\beta_2$ heterotetramers. The alpha
68 subunits contain an extracellular ligand-binding domain and the beta subunits are
69 composed of a single transmembrane domain and a highly conserved intracellular TK
70 domain (LeRoith et al., 1995).

71 At a cellular level the IGF-1 acts in an autocrine/paracrine manner to control
72 physiological processes such as protein synthesis, cell proliferation, differentiation, and
73 apoptosis (Jones and Clemmons, 1995). Almost all biological actions of IGF-1 are
74 mediated by the type 1 IGF-1 receptor (IGF-1R) (Whitehead et al., 2000). Once
75 activated, IGF-1R undergoes a conformational change leading to autophosphorylation

76 of tyrosine residues that serve as recruiting sites for cytoplasmic proteins, including
77 insulin receptor substrate proteins (IRS). IRS molecules are associated with IGF-1R at
78 the cell surface, creating a scaffold for downstream molecules, such as phosphoinositide
79 3-kinase (PI3K) (Glass, 2005). Once IGF-1 signal transduction activates intracellular
80 PI3K activity, it results in an increased phosphorylation and activation of the
81 Akt/mTOR regulated pathways which increase protein synthesis and suppress protein
82 degradation (Rommel et al., 2001). Other described signaling transduction pathways
83 activated by IGF-1 are the MAP kinases MEK-ERK involved in cellular proliferation
84 and differentiation (Li and Johnson, 2006). In vertebrates, the insulin-like growth factor
85 system has been shown to be unique among growth factors, playing an important role in
86 the early patterning and muscle development: the mRNA microinjection of a non
87 functional IGF-1 receptor in zebrafish induced small sized embryos with the absence of
88 notochord and abnormal somites, on the other hand the over expression of IGF-1
89 resulted in a greatly expanded development of anterior structures (Eivers et al., 2004).

90 Over the past decade, IGF-1R and IGF-1 cDNAs partial and complete sequences
91 have been isolated from several vertebrates, including teleost such as coho salmon
92 (*Oncorhynchus kisutch*), rainbow trout (*Oncorhynchus mykiss*), turbot (*Psetta maxima*),
93 among others (Elies et al., 1999; Chan et al., 1997; Greene and Chen, 1999; Wood et al.,
94 2005). Additionally, two distinct IGF-1R genes named *igf-1ra* and *igf-1rb* were
95 described in zebrafish (*Danio rerio*) indicating a probable gene duplication during
96 teleost evolution (Maures et al., 2002).

97 Chilean flounder (*Paralichthys adspersus*) is a marine fish widely distributed
98 throughout the Chilean coast, which is raising a high economic value. However, slow
99 growth rate has been recognized as a major problem in marine fish, increasing the final
100 production cost. Thus, the viability to farm these species requires new knowledge in

101 order to develop new strategies to improve fish growth (Delgado et al., 2008). In this
102 regard, considering the relevant function of the IGF-1 signaling pathways in promoting
103 growth and skeletal muscle development in fish, we describe here as a first
104 approximation, the isolation and characterization of the full length IGF-1R and IGF-1
105 cDNAs, which include the codifying regions and the 5' and 3' untranslated regions of
106 both transcripts. Additionally, we studied their mRNA expression in pre-metamorphic
107 larvae and in different tissues of adult fish.

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126 **Materials and Methods:**

127 **Fish:** Chilean flounder fish (*Paralichthys adspersus*) were collected from the Centro de
128 Investigaciones Marinas de Quintay (CIMARQ) (V Region, Valparaíso, Chile). The fish
129 were maintained under natural temperature and photoperiod conditions corresponding to
130 geographic localization of CIMARQ (33°13'S 71°38'W) and were feed twice daily with
131 turbot pellet (Biomar, Chile). Adult fish (36 month old) were sacrificed through an
132 overdose of anesthetic (3-aminobenzoic acid ethyl ester) (300 mg/L). The kidney, gills,
133 intestine, gonads, spleen, liver, stomach, brain, white muscle, esophagus and red muscle
134 tissues were collected, directly frozen in liquid nitrogen and stored at -80°C.

135 Larvae were obtained after *in vitro* fertilization of eggs by male broodstock
136 sperm. Embryos were maintained under intensive-culture conditions in conic larval
137 culture tanks at 19°C \pm 2°C. Larvae at pre-metamorphic stages were collected, fixed in
138 4% paraformaldehyde in PBS for 2h at 4°C, dehydrated in methanol and stored at -
139 20°C.

140

141 **RT-PCR and cDNA cloning:** Total RNA was isolated from liver using Trizol reagent
142 following the manufacturer protocols (Invitrogen, Carlsbad, CA, USA). A total of 5 μ g
143 of RNA previously treated with DNase I (1U/ μ L) was used for RNA first-strand cDNA
144 using M-MLV reverse transcriptase (Invitrogen, Carlsbad, CA, USA). PCR primers for
145 cloning IGF-1 receptor and IGF-1 cDNA (table 1) were designed from a consensus
146 analysis of conserved coding regions of known IGF-1 receptor and IGF-1 ligand
147 sequences from Japanese flounder, turbot and zebrafish. The PCR reaction, containing
148 the cDNA template, 10 μ L of 10X PCR buffer, 200 μ M of each dNTP, 1 μ g of the
149 forward and reverse primers, and 2.5U of Taq DNA polymerase (Promega, Madison,
150 WI, USA), was carried out in a final volume of 50 μ L. The IGF-1R and IGF-1 PCR

151 products were cloned into the pGEM-T easy vector (Promega, Madison, WI, USA) and
152 the clones were completely sequenced and assembled in only one sequence. The IGF-
153 1R and IGF-1 full-length 5'-terminal region, including the transcription start site, was
154 completed using the First Choice RLM-RACE kit (Ambion, Austin, TX, USA)
155 according to the manufacturer's instructions. Briefly, a RT-PCR with an adapter primer
156 and the IGF-1R gene specific primers (5IGF-1ROP 5'-
157 GACAGACAGCATCAGACCCCAAACA-3', 5IGF-1RIP 5'-
158 TGCCAGTCACAGGATACTTG-3') or the IGF-1 gene specific primers (5IGF-1OP 5'-
159 AAAAGCCTCTCTCTCCACACAC-3', 5IGF-1IP 5'-
160 TCTCTCCACACACAAACTGCAG-3') using a CIP/TAP mRNA as a template in a
161 nested reaction. The IGF-1R 3'-region was obtained using the gene-specific primers
162 (3IGF-1ROP 5' - ACCCAGGTCCTACCCCACTCAA -3', 3IGF-1RIP 5'-
163 TTCTCCCTTCGGGGAAAT GAGTTT -3'). The IGF-1 3'-region was obtained using
164 the gene-specific primers (3IGF-1OP 5' -ACCTGGAGATGTACTGTGCAC-3', 3IGF-
165 1IP 5'-CAAGACTAGCAAGGCAGCTC-3').

166

167 **Sequence analysis:** Amino acid sequences translated from the cDNA sequence were
168 compared with sequences in the GeneBank public database, by using the NCBI-BLAST
169 application (<http://www.ncbi.nlm.nih.gov/blast>). Multiple aminoacid sequences
170 alignment for the IGF-1 receptor and IGF-1 were performed using clustalW (Thompson
171 et al., 1994).

172

173 **Tissue expression and distribution of IGF-1 receptor and IGF-1 by RT-PCR**
174 **analysis:** Total RNA was extracted from different tissues (kidney, gills, intestine,
175 gonads, spleen, liver, stomach, brain, esophagus, white muscle and red muscle).

176 Reverse transcription reaction was performed using 1 µg of total RNA previously treated
177 with DNase I. For IGF-1R and IGF-1, gene-specific primers were designed to amplify a
178 320 (forward: 5'-GCGGGAATTCGATTGCCTTT -3', reverse: 5'-
179 ATCACGAGGGCGTAGTTGTA-3') and 454 pb (forward: 5'-
180 GTCTAGCGCTCTTTCCTTTCAGTG-3', reverse: 5'-
181 TTTTGTCTTGTCTGGTCGCTGTGC-3') fragments respectively. For normalization
182 purposes, gene-specific primers (forward: 5'-AGGGAAATCGTGCGTGACAT-3',
183 reverse: 5'-TCAGGCAGCTCATAGCTCTT-3') were used to amplify a β-actin 116 bp
184 fragment as constitutive gene expression control.

185

186 **Whole-mount in situ hybridization:** A 1,604 bp fragment corresponding to the IGF-
187 1R coding sequence was amplified by PCR using gene-specific primers (5'-
188 GCCTTCCAGAACATCACAGAG-3', 5'-TTGAACTCCTTCATGACGGAGG-3')
189 and the same cDNA described before as a template. This fragments were cloned into
190 pGEM-T Easy Vector System (Promega, Madison, WI, USA) originating the
191 pL3F4R1604, which was linearized with *NdeI* or *SpHI* restriction enzymes to
192 synthesized sense (control) and antisense riboprobes DIG-UTP-labeled (Roche
193 Diagnostics, Mannheim, Germany) using SP6 and T7 RNA polymerases (Promega,
194 Madison, WI, USA) respectively. A 454 bp fragment corresponding to the IGF-1 coding
195 sequence was amplified by PCR using gene-specific primers (5'-
196 CCTCTCCACTACTGCTGTGTGTC-3', 5'-ATGTCTGTGTGGCGTTGTGCAC-3')
197 and the same cDNA described before as a template. This fragments were cloned into
198 pGEM-T Easy Vector System (Promega, Madison, WI, USA) originating the
199 pLeigfI454, which was linearized with *NdeI* or *NcoI* restriction enzymes to synthesized
200 sense (control) and antisense riboprobes DIG-UTP-labeled (Roche Diagnostics,

201 Mannheim, Germany) using SP6 and T7 RNA polymerases (Promega, Madison, WI,
202 USA) respectively. The riboprobes were purified using mini Quick Spin Columns
203 (Roche Diagnostics, Mannheim, Germany) to eliminate unincorporated labeled
204 nucleotides. Whole mount in situ hybridization were performed according to Fuentes et
205 al., (2008). Briefly, after bleaching treatment, embryos and larvae were pre-hybridized
206 overnight at 60°C in hybridization buffer and then incubated overnight at 65°C in
207 hybridization buffer including 50 ng of sense or antisense IGF-1R and IGF-1
208 riboprobes. After hybridization, embryos and larvae were washed in a solution with
209 decreasing formamide concentration in 2X SSC, followed by two wash-steps with SSC
210 0.2X for 30 min at 65 °C. Larvae were incubated for 4 h in a blocking buffer at room
211 temperature. For immunodetection, samples were incubated overnight at 4°C with anti-
212 digoxigenin-AP antibody (Roche Diagnostics, Mannheim, Germany). After washes with
213 PBT to eliminate non-bounded antibodies and three additional washes with AP-buffer,
214 stains were performed with NBT/BCIP (75 mg/mL and 50 mg/mL, respectively)
215 (Promega, Madison, WI, USA) for 6 h at 37 °C. The experiment was performed four
216 times using n=15 individuals from each developmental stages. After in situ
217 hybridization, observed in a Leica MZ12.5 stereomicroscope and photographed with a
218 Leica DF300 camera.

219

220

221

222

223

224

225

226 **Results**

227 **Cloning and characterization of the IGF-1R and IGF-1 cDNA:** The complete
228 Chilean flounder IGF-1R and IGF-1 cDNA sequences (cfIGF-1R and cfIGF-1) were
229 obtained using RT-PCR coupled to RACE method.

230 The length of the complete IGF-1R cDNA sequence was 5,033 bp, which
231 includes a 622 bp 5'-untranslated region (UTR), an open reading frame (ORF) of 4,239
232 bp and a 172 bp 3'-UTR. The ORF encodes a putative protein of 1,412 amino acid
233 residues (Fig. 1A). The sequence analysis reveals that the Chilean flounder cDNA IGF-
234 1 receptor is organized into several major domains including a signal peptide sequence
235 of 30 amino acids, an extracellular alpha subunit of 702 amino acids and an intracellular
236 beta subunit of 676 amino acids (Fig. 1A).

237 The length of the complete IGF-1 cDNA sequence is 980 bp, which includes a
238 152 bp 5'-untranslated region (UTR), an open reading frame (ORF) of 558 bp and a 270
239 bp 3'-UTR, encoding a putative protein of 185 amino acid residues (Fig. 1B). The
240 comparative IGF-1 sequence analysis with other species reveals that the ligand deduced
241 protein is subdivided into six structural domains, including a signal peptide sequence of
242 44 amino acids, the B domain of 29 amino acids, the C domain of 10 amino acids, the A
243 domain of 21 amino acids, the D domain of 8 amino acids and the E domain of 73
244 amino acids (Fig. 1B).

245 Amino acid sequence alignment of cfIGF-1R (FJ438475.1) with different
246 vertebrates orthologs, including mammalian (human, X04434.1 and rat, AF056187.1),
247 birds (chicken, AJ223164.1), amphibians (*Xenopus*, AF055980.1) and fish (zebrafish,
248 AF400275.1 and Japanese flounder, AB065098.1) was performed. The cfIGF-1R
249 sequence was found to have 61%, 61%, 63%, 62% of identity with IGF-1R of human,
250 rat, chicken and *Xenopus*, respectively. Higher degrees of identity were found with

251 other fish IGF-1R sequences, including a 74% with zebrafish and as much as 97% with
252 another fish belonging to the *Paralichthys* genus, the Japanese flounder (Fig. 2A, Fig.
253 3A).

254 A potential proteolytic cleavage sequence R-X-R-R was conserved in all the
255 species compared. The cysteine-rich domain, into the alpha subunit of the cfIGF-1R,
256 contains 24 cysteine residues, which were also observed in all vertebrates IGF-1Rs. In
257 the beta subunit, an IRS-1 and IRS-2 binding site (NPEY and GVLY), a potential ATP
258 binding site (G-X-G-X-X-G-21-X-K), an autophosphorylation motif (YETDYY) and
259 seven tyrosine residues in the tyrosine kinase domain, were highly conserved in all the
260 studied species. A lesser conserved region was found in the carboxyl-terminal, where
261 large insertions were observed in the teleosts IGF-1R compared with those from higher
262 vertebrates (Fig. 2A, Fig. 3A).

263 Amino acid sequence alignment of the cfIGF-1 (EU017533.1) with those from
264 several other species, including human (M27544.1), rat (NM_001082479.1), chicken
265 (M32791.1), *Xenopus* (M29857.1), zebrafish (BC114262.1) and Japanese flounder
266 (AJ010602.1) was performed. The cfIGF-1 sequence was found to be 60%, 65%, 69%,
267 66%, 73%, 97% homologous to the IGF-1 of human, rat, chicken, *Xenopus*, zebrafish
268 and Japanese flounder, respectively (Fig. 2B, Fig 3B).

269 The comparison between the known IGF-1 sequences, reveal higher conserved
270 regions at both B and A domains while the C, D and E domains differ significantly (Fig.
271 3B). The predicted IGF-1 protein is highly rich in charged amino acid residues and
272 conserves the cysteine residues responsible for maintenance of tertiary structure.
273 Moreover, Chilean flounder IGF-1 contains conserved residues involved in the
274 interaction between IGF-1 with IGF-1 receptor and IGF-1 binding protein (Fig. 2B).

275 **Tissue distribution of IGF-1R and IGF-1 mRNA:** IGF-1R and IGF-1 mRNA
276 RT-PCR experiments were performed to study the expression of mRNA in different
277 tissues of adult fish using β -actin as a constitutive expression control (Fig. 4). The IGF-
278 1R and IGF-1 transcripts were detected in all investigated tissues. High levels of IGF-
279 1R mRNA expression were observed in the testis, intestine, liver, stomach, brain,
280 muscle and oesophagus. The IGF-1R mRNA expression detected in the kidney, gills
281 and spleen was at low levels. Moreover, high IGF-1 mRNA expression was observed in
282 the gills, liver, testis, intestine, spleen, white muscle and brain. Lower IGF-1 mRNA
283 expression levels were detected in the kidney, stomach, oesophagus and red muscle.

284 In addition, we studied the expression pattern of IGF-1R and IGF-1 mRNA in
285 embryos using whole mount *in situ* hybridization in Chilean flounder larvae from 9.0
286 days post-fertilization (dpf) (Fig. 5). The cfIGF-1R mRNA was detected at 9 dpf, larvae
287 exhibited IGF-1R mRNA expression in the somites, notochord and cartilaginous tissues
288 in the head. Additionally, cfIGF-1 mRNA was detected only in the notochord. Sense
289 probe was included as a negative control in all *in situ* hybridization experiments. No
290 signal was detected, showing that RNA hybridization was specific.

291
292
293
294
295
296
297

298 **Discussion**

299 In this work we reported the complete cDNA sequence of the IGF-1 and IGF-1
300 receptor from the flat fish the Chilean flounder (*Paralichthys adspersus*), an emergent
301 species for aquaculture. These results complement reported data of myostatin and the
302 growth hormone receptor (GHR) (Fuentes et al., 2008; Delgado et al., 2008), all
303 important genes for growth and development in vertebrates (Dayton and White, 2008).
304 The Chilean flounder pre-pro-IGF-1 consists of 185 amino acid residues, which
305 contains a signal peptide and B, C, A, D, and E domains. The cfIGF-1R cDNA consists
306 of 1412 amino acid residues, and like other vertebrates contains a signal peptide and
307 cysteine, trans membrane, juxtamembrane, tyrosine kinase and carboxy domains (Jones
308 and Clemmons, 1995; LeRoith et al., 1995). A multiple alignment of the deduced amino
309 acid sequence of cfIGF-1 and cfIGF-1R with other vertebrate sequences was performed
310 indicating a high degree of conservation of these proteins during vertebrate evolution
311 (Elies et al., 1999; Nakao et al., 2002).

312 The deduced protein sequence of cfIGF-1 shares an overall identity of 60 to 65%
313 with mammalian IGF-1 sequences, and 73 to 97% with other teleost IGF-1 sequences.
314 Among mature IGF-1, the B and A domains of the peptides are well conserved, while
315 the C, D and E domains differ significantly (Moriyama, 2000). The importance of this
316 high sequence identity of B and A domains in different species can be attributed to the
317 functional roles of these regions, which are involved in the binding of IGF-1 with its
318 receptor such as Arg21 and the Phe23-Tyr24-Phe25 motif in the B domain and Tyr58
319 (cf_Tyr56) in the A domain (Bayne et al., 1990; Zhang et al., 1994). The amino acidic
320 residues involved in IGF binding with IGFBP: Glu3, Thr4, Glu9, Gln15, and Phe16 in
321 the B domain and Phe47 (cf_Phe45) and Ser49 (cf_Ser47) in the A domain are highly
322 conserved (Clemmons et al., 1992; Magee et al., 1999). The C domain also contains

323 some conserved residues important in IGF-1R and/or IGFBP binding, such as Tyr31,
324 Arg36 and Arg37. Additionally, the cfIGF-1 sequence showed six conserved cysteine
325 residues CysB6, CysB18, CysA6, CysA7, CysA11, and CysA20 which are also located
326 at the same positions as mammals IGF-1 which is responsible for maintenance of
327 tertiary structure (Hober et al., 1992).

328 The deduced protein sequence of cfIGF-1R shares an overall identity of 61%
329 with mammalian IGF-1R sequences, and 68 to 97% with other teleost IGF-1 sequences.
330 Among mature IGF-1R, the cysteine, juxtamembrane and tyrosine kinase domains are
331 well conserved, while the C terminal domain differs significantly. Most of the
332 conserved regions are known to be critical for IGF-1R biological activity, such as the
333 ligand-binding motif, tyrosine kinase domain, ATP-binding site, and IRS binding site.
334 The ligand binding motif is located between the amino acidic residues Cys148 and
335 Cys302, showing a variable conservation, however, 24 cystein residues have a high
336 conservation degree in number and position (Jones and Clemmons, 1995). The tyrosine
337 kinase domain located between the amino acids residues Arg1003 (Cf_Lys1002) and
338 Phe1259 (Cf_Phe1256) contain the tyrosine cluster (Tyr1131, Tyr1135 and Tyr1136)
339 required for the receptor autophosphorilation and the ATP binding sequences (G-X-G-
340 X-X-G-21-X-K) (Gronborg et al., 1993). This domain is absolutely necessary for the
341 biological receptor activity in vertebrates. The juxtamembrane domain contains the
342 Insulin Receptor Substrate (IRS) binding site (NPEY and GVLY) with a high degree of
343 conservation (Moriyama et al., 2000). In contrast, the carboxy-terminus of the receptors
344 is the most divergent region; teleosts allocate insertions in the carboxyl-terminus and
345 this suggests that the function played by this region may differ between mammals
346 and fishes (Kuang et al., 2005). Moreover, isoforms of IGF-1R in teleost fish have been
347 reported: two subtypes of IGF-1R cDNAs were found to be coded by distinct genes in

348 zebrafish and the Japanese flounder (Maures et al., 2002; Nakao et al., 2002).
349 Additionally two partial cDNAs have been identified in coho salmon (*Oncorhynchus*
350 *kisutch*) and rainbow trout (*Oncorhynchus mykiss*), and it has been suggested that the
351 IGF-1Rs are encoded by two genes in these species (Chan et al., 1997; Greene and Chen
352 1999). Sequence comparison analysis revealed that the IGF-1R cDNA obtained in this
353 study belongs to the IGF-1Ra. Until now we have been unable to isolate the cDNA
354 sequence of the IGF-1Rb in Chilean flounder.

355 Insulin-like growth factor-1 is a mitogenic peptide that circulates in plasma and
356 acts in many tissues through the endocrine, paracrine and autocrine mechanisms
357 (Humbel, 1990). Although IGF-1 is secreted predominantly in the liver in response to
358 growth hormone, it is also produced in essentially all tissues. The biological responses
359 of IGF-1 are mainly mediated by the binding and activation the IGF-1 receptor. The
360 IGF-1R and IGF-1 mRNA expression were found in a wide variety of tissues; the tissue
361 mRNA expression patterns reported in this study are consistent with previous reported
362 data from other teleosts and the known pleiotropic role of the receptor and ligand (Duan
363 et al., 1993; Reinecke et al., 1997; Maures et al., 2002; Tse et al., 2002; Queenie et al.,
364 2003; Radaelli et al., 2003a; Radaelli et al., 2003b; Duval et al., 2002; Clay et al., 2005;
365 Patruno et al., 2006). RT-PCR revealed expression of IGF-1R and IGF-1 mRNA in the
366 kidney, gills, intestine, testis, spleen, liver, stomach, brain, oesophagus and skeletal
367 muscle. The expression of IGF-1 and IGF-1R in the brain suggests the IGF-1
368 involvement in central nervous system development (Kuang et al., 2005; Greene et al.,
369 1999; Ayaso et al., 2002). The presence of high levels of IGF-1R and IGF-1 mRNA in
370 the gonad supports their important roles such as regulators in hormone synthesis and
371 secretion, germ cell proliferation and differentiation (Weber et al., 2000; Hammond et
372 al., 1991). The presence of IGF-1R and IGF-1 in the gills and intestine is according to

373 their described roles in osmoregulation in fishes, and seawater adaptability (Sakamoto
374 and Hirano., 1991; Datuin et al., 2001; Ng et al., 2001). Moreover, the high expression
375 levels of IGF-1R and IGF-1 detected in skeletal muscle agree with their participation in
376 skeletal muscle satellite cell proliferation and differentiation mediated by the signaling
377 pathways Ras-MEK-ERK (Li and Johnson, 2006; Castillo et al., 2006) and their
378 hypertrophic role mediated by the signaling pathways PI3K-Akt-mTOR during fish
379 growth (Rommel et al., 2001; Castillo et al., 2006).

380 The spatial expression of IGF-1R and/or IGF-1 in the notochord and somites has
381 been described in other fish such as zebrafish, tilapia (*Oreochromis niloticus*), gilthead
382 seabream (*Sparus aurata*) and rainbow trout (*Oncorhynchus mykiss*) (Devoto et al.,
383 1996; Rescan et al., 2001; Radaelli et al., 2003b; Funkenstein et al., 1997; Maures et al.,
384 2002; Eivers et al., 2004; Berishvili et al., 2006). In zebrafish it has been shown that
385 IGF-1R and its ligand IGF-1 mRNA are expressed during early development, from
386 blastula (2 hpf) to the larval stage (96 hpf) showing that IGF-1R is maternally expressed
387 (Ayaso et al., 2002; Maures et al., 2002). We examined the mRNA expression pattern of
388 Chilean flounder IGF-1R and IGF-1 in larvae at pre-metamorphic stage through whole
389 mount *in situ* hybridization. We observed that Chilean flounder of 9 dpf, where larvae
390 begins to feed themselves, showed IGF-1R expression in the somites, head and
391 notochord, whereas IGF-1 expression was found in the notochord. We previously
392 described the expression pattern of the growth hormone receptor (GHR) and myostatin
393 genes during larvae development of Chilean flounder (Fuentes et al., 2008; Delgado et
394 al., 2008). Interestingly, we showed that GHR was expressed in similar territories as
395 IGF-1R, such as the somites, which give rise to muscle and the axial skeleton, and the
396 notochord, an essential structure for the proper formation of the nervous system and
397 mesoderm, the last of which gives rise to the somites later in embryo development

398 (Richardson et al., 1998; Stemple et al., 1996). Furthermore, ligands just as myostatin
399 and IGF-1 mRNAs, were mostly found in the notochord. Taken together, these
400 observations suggest there could be a synchronization between positive and negative
401 growth signals originated in the notochord that plays a crucial role in the control of
402 somite development.

403 In summary, the complete cDNA sequence of the IGF-1R and IGF-1 were
404 cloned from the Chilean flounder fish. The protein sequence includes all the structural
405 domains and motifs responsible for the interaction between ligand- receptor and IGF-1
406 mediated signal transduction. Indeed, our results contribute to the knowledge of the
407 IGF-1 system in the larvae and juvenile stages, both of which are crucial periods for
408 developing successful farming of the Chilean flounder.

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423 **Acknowledgements**

424

425 This work was supported by Grants N° 1090416 from FONDECYT and 15-03/28-
426 04/13-06I from UNAB Research Fund to A.M., FONDECYT N°1095128 to A.E.R., and
427 the Millennium Institute for Fundamental and Basic Biology (MIFAB). We thank Juan
428 Manuel Estrada for technical assistance and animal care in the Centro de Investigación
429 Marina de Quintay (CIMARQ) and Ashley VanCott from The University of Nevada,
430 Reno (USA) for improving and correcting the English of the manuscript

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448 **References**

449

450 Ayaso, E., Nolan, C.M., Byrnes, L., 2002. Zebrafish insulin-like growth factor-I
451 receptor: molecular cloning and developmental expression. *Mol. Cell. Endocrinol.*
452 191, 137-148.

453

454 Bayne, M.L., Applebaum, J., Chicchi, G.G., Miller, R.E., Cascieri, M.A., 1990. The
455 roles of tyrosines 24, 31, and 60 in the high affinity binding of insulin-like growth
456 factor-I to the type 1 insulin-like growth factor receptor. *J. Biol. Chem.* 265, 15648-
457 15652.

458

459 Berishvili, G., Shved, N., Eppler, E., Clota, F., Baroiller, J.F., Reinecke, M., 2006.
460 Organ-specific expression of IGF-I during early development of bony fish as
461 revealed in the tilapia, *Oreochromis niloticus*, by *in situ* hybridization and
462 immunohistochemistry: indication for the particular importance of local IGF-I. *Cell*
463 *Tissue Res.* 325, 287-301.

464

465 Castillo, J., Johnsen-Ammendrup, I., Codina, M., Navarro, I., Gutierrez, J., 2006.
466 IGF-I and insulin receptor signal transduction in trout muscle cells. *Am. J. Physiol.*
467 *Regul. Integr. Comp. Physiol.* 290, R1683-1690.

468

469 Chan, S.J., Plisetskaya, E.M., Urbinati, E., Jin, Y., Steiner, D.F., 1997. Expression
470 of multiple insulin and insulin-like growth factor receptor genes in salmon gill
471 cartilage. *Proc. Natl. Acad. Sci. U.S.A.* 94, 12446-12451.

472

473 Clay, L.A., Wang, S.Y., Wolters, W.R, Peterson, B.C., Waldbieser, G.C., 2005.
474 Molecular characterization of the insulin-like growth factor-I (IGF-I) gene in
475 channel catfish (*Ictalurus punctatus*). Biochim. Biophys. Acta 1731, 139-148.
476
477 Clemmons, D.R., Dehoff, M.L., Busby, W.H., Bayne, M.L., Cascieri, M.A., 1992.
478 Competition for binding to insulin-like growth factor (IGF) binding protein-2, 3, 4,
479 and 5 by the IGFs and IGF analogs. Endocrinology 131, 890-895.
480
481 Datuin, J.P., Ng, K.P., Hayes, T.B., Bern, H.A., 2001. Effects of glucocorticoids on
482 cartilage growth and response to IGF-I in the tilapia (*O. mossambicus*). Gen.Comp.
483 Endocrinol. 121, 289-294.
484
485 Dayton, W.R., White, M.E., 2008. Cellular and molecular regulation of muscle
486 growth and development in meat animals. J. Anim. Sci. 86, E217-E225.
487
488 Delgado, I., Fuentes, E., Escobar, S., Navarro, C., Corbeaux, T., Reyes, A.E., Vera,
489 M.I., Alvarez, M., Molina, A., 2008. Temporal and spatial expression pattern of the
490 myostatin gene during larval and juvenile stages of the Chilean flounder
491 (*Paralichthys adspersus*). Comp. Biochem. Physiol. B Biochem. Mol. Biol. 151,
492 197-202.
493
494 Devoto, S.H., Melancon, E., Eisen, J.S., Westerfield, M., 1996. Identification of
495 separate slow and fast muscle precursor cell *in vivo*, prior to somite formation.
496 Development 122, 3371-3380.
497

498 Duan, C., Duguay, S., Plisetskaya, E., 1993. Insulin-like growth factor I (IGF-I)
499 mRNA expression in coho salmon, *Oncorhynchus kisutch*: tissue distribution and
500 effects of growth hormone/prolactin family peptides. *Fish Physiol. Biochem.* 11,
501 371–379.

502

503 Duval, H., Rousseau, K., Elies, G., Le Bail, P., Dufour, S., Boeuf, G., Boujard, D.,
504 2002. Cloning, characterization, and comparative activity of turbot. IGF-I and IGF-
505 II. *Gen.Comp. Endocrinol.* 126, 269–278.

506

507 Eivers, E., McCarthy, K., Glynn, C., Nolan, C.M., Byrnes, L., 2004. Insulin-like
508 growth factor (IGF) signalling is required for early dorso-anterior development of
509 the zebrafish embryo. *Int. J. Dev. Biol.* 48, 1131-1140.

510

511 Elies, G., Duval, H., Bonnac, G., Wolff, J., Boeuf, G., Boujard, D., 1999. Insulin
512 and insulin-like growth factor-1 receptors in an evolved fish, the turbot: cDNA
513 cloning and mRNA expression. *Mol. Cell. Endocrinol.* 158, 173-185.

514

515 Etherton, T.D., 2004. Somatotropic function: the somatomedin hypothesis revisited.
516 *J. Anim. Sci.* 82, E239–E244.

517

518 Fuentes, E., Poblete, E., Reyes, A.E., Vera, M.I., Alvarez, M., Molina, A., 2008.
519 Dynamic expression pattern of the growth hormone receptor during early
520 development of the Chilean flounder. *Comp. Biochem. Physiol. B Biochem. Mol.*
521 *Biol.* 150, 93-102.

522

523 Funkenstein, B., Almuly, R., Chan, S.J., 1997. Localization of IGF-I and IGF-I
524 receptor mRNA in *Sparus aurata* larvae. Gen. Comp. Endocrinol. 107, 291-303.
525

526 Glass, D.J., 2005. Skeletal muscle hypertrophy and atrophy signaling pathways. Int.
527 J. Biochem. Cell Biol. 37, 1974–1984.
528

529 Greene, M.W., Chen, T.T., 1999. Characterization of teleost insulin receptor family
530 members. II. Developmental expression of insulin-like growth factor type I receptor
531 messenger RNAs in rainbow trout. Gen. Comp. Endocrinol. 115, 270-281.
532

533 Gronborg, M., Wulff, B. S., Rasmussen, J. S., Kjeldsen, T., Gammeltoft, S., 1993.
534 Structure-function relationship of the insulin-like growth factor-I receptor tyrosine
535 kinase. J. Biol. Chem. 258, 23435-23440.
536

537 Hammond, J.M., Mondschein, S.E., Samaras, S.E., Canning, S.F., 1991. The
538 ovarian insulin-like growth factors, a local amplification mechanism for
539 steroidogenesis and hormone action. J. Steroid Biochem. Mol. Biol. 40, 411– 416.
540

541 Hober, S., Forsberg, G., Palm, G., Hartmanis, M., Nilsson B., 1992. Disulfide
542 exchange folding of insulin-like growth factor-I. Biochemistry 31, 1749-1756.
543

544 Humbel, R.E., 1990. Insulin-like growth factors I and II. Eur. J. Biochem. 190, 445-
545 462.
546

547 Jones, J.I., Clemmons, D.R., 1995. Insulin-like growth factors and their binding
548 proteins: biological actions. *Endocr. Rev.* 16, 3-34.
549

550 Kuang, Y.M., Li, W.S., Lin, H.R., 2005. Molecular cloning and mRNA Profile of
551 Insulin-like Growth Factor Type 1 receptor in orange-spotted Grouper, *Epinephelus*
552 *coioides*. *Acta Biochim. Biophys. Sin.* 37, 327-334.
553

554 LeRoith, D., Werner, H., Beitner-Johnson, D., Roberts, C.T., 1995. Molecular and
555 cellular aspects of the insulin-like growth factor I receptor. *Endocr. Rev.* 16, 143-
556 163.
557

558 LeRoith, D., Scavo, L., Butler, A., 2001. What is the role of circulating IGF-1?
559 *Trends Endocrinol. Metab.* 12, 48–52.
560

561 Li, J., Johnson, S.E., 2006. ERK2 is required for efficient terminal differentiation of
562 skeletal myoblasts. *Biochem. Biophys. Res. Commun.* 345, 1425-1433.
563

564 Magee, B.A., Shooter, G.K., Wallace, J.C., Francis, G.L., 1999. Insulin-like growth
565 factor I and its binding proteins: a study of the binding interface using B-domain
566 analogues. *Biochemistry* 38, 15863-15870.
567

568 Maures, T., Chan, S.J., Xu, B., Sun, H., Ding, J., Duan, C., 2002. Structural,
569 biochemical, and expression analysis of two distinct insulin-like growth factor I
570 receptors and their ligands in zebrafish. *Endocrinology* 143, 1858-1871.
571

572 Moriyama, S., Ayson, F.G., Kawauchi, H., 2000. Growth regulation by insulin-like
573 growth factor-I in fish. *Biosci. Biotechnol. Biochem.* 64, 1553-1562.
574

575 Nakao, N., Tanaka, M., Higashimoto, Y., Nakashima, K., 2002. Molecular cloning,
576 identification and characterization of four distinct receptor subtypes for insulin and
577 IGF-I in Japanese flounder, *Paralichthys olivaceus*. *J. Endocrinol.* 173, 365-375.
578

579 Ng, K.P., Datuin, J.P., Bern, H.A., 2001. Effects of estrogens in vitro and in vivo on
580 cartilage growth in the tilapia (*O. mossambicus*). *Gen.Comp. Endocrinol.* 121, 295-
581 304.
582

583 Patruno, M., Maccatrozzo, L., Funkenstein, B., Radaelli G., 2006. Cloning and
584 expression of insulin-like growth factors I and II in the shi drum (*Umbrina cirrosa*).
585 *Comp. Biochem. Physiol., B.* 144, 137–151.
586

587 Queenie, P., Vonga, K., Chana, M., Leungb, K., Cheng C., 2003. Common carp
588 insulin-like growth factor-I gene: complete nucleotide sequence and functional
589 characterization of the 5V-flanking region. *Gene* 322, 145–156.
590

591 Radaelli, G., Patruno, M., Maccatrozzo, L., Funkenstein B., 2003a. Expression and
592 cellular localization of insulin-like growth factor-II protein and mRNA in *Sparus*
593 *aurata* during development. *J. Endocrinol.* 178, 285–299.
594

595 Radaelli, G., Domeneghini, C., Arrighi, S., Bosi, G., Patruno, M., Funkenstein B.,
596 2003b. Localization of IGF-I, IGF-I receptor, and IGFBP-2 in developing *Umbrina*
597 *cirrosa* (Pisces: Osteichthyes). *Gen. Comp. Endocrinol.* 130, 232-244.
598
599 Reinecke, M., Schmid, A., Ermatinger, R., Loffing-Cueni D., 1997. Insulin-Like
600 Growth Factor I in the Teleost *Oreochromis mossambicus*, the Tilapia: Gene
601 Sequence, Tissue Expression, and Cellular Localization. *Endocrinology* 138, 3613-
602 3619.
603
604 Rescan, P.Y., Collet, B., Ralliere, C., Cauty, C., Delalande, J.M., Goldspink, G.,
605 Fauconneau, B., 2001. Red and White Muscle development in the trout (*O.mykiss*)
606 as shown by *in situ* hybridization of fast and slow myosin heavy chain transcripts. *J.*
607 *Exp. Biol.* 204, 2097-2101.
608
609 Richardson, M.K., Allen, S.P., Wright, G.M., Raynaud, A., Hanken J., 1998. Somite
610 number and vertebrate evolution. *Development* 125, 151-160.
611
612 Riedemann, J., Macaulay, V.M., 2006. IGF1R signalling and its inhibition. *Endocr.*
613 *Relat. Cancer* 13, S33-S43.
614
615 Rommel, C., Bodine, S.C., Clarke, B.A., Rossman, R., Nunez, L., Stitt, T.N.,
616 Yancopoulos, G.D., Glass, D.J., 2001. Mediation of IGF-1-induced skeletal
617 myotube hypertrophy by PI(3)K/Akt/mTOR and PI(3)K/Akt/GSK3 pathways. *Nat.*
618 *Cell Biol.* 3, 1009–1013.
619

620 Sakamoto, T., Hirano, T., 1991. Growth hormone receptors in the liver and
621 osmoregulatory organs of rainbow trout: characterization and dynamics during
622 adaptation to seawater. *J. Endocrinol.* 130, 425-433.

623

624 Stemple, D., Solnica-Krezel, L., Zwartkruis, F., Neuhauss, C.F., Schier, A.F.,
625 Malicki, J., Stainier, D.Y.R., Abdelilah, S., Rangini, Z., Mountcastle-Shah, E.,
626 Driever, W., 1996. Mutations affecting development of the notochord in zebrafish.
627 *Development* 123, 117-128.

628

629 Thompson, J.D., Higgins, D.G., Gibson, T.J., 1994. CLUSTAL W: improving the
630 sensitivity of progressive 416 multiple sequence alignment through sequence
631 weighting, position-specific gap penalties and weight 417 matrix choice. *Nucleic
632 Acids Res.* 22, 4673-4680.

633

634 Tse, M.C.L., Vong, Q.P., Cheng, C.H.K., Chan K.M., 2002. PCR-cloning and gene
635 expression studies in common carp (*Cyprinus carpio*) insulin-like growth factor-II.
636 *Biochim. Biophys. Acta* 1575, 63–74.

637

638 Weber, G.M., Sullivan, C.V., 2000. Effects of insulin-like growth factor-I on in
639 vitro final oocyte maturation and ovarian steroidogenesis in striped bass, *Morone
640 saxatilis*. *Biol. Reprod.* 63, 1049–1057

641

642 Wood, A.W., Duan, C., Bern, H.A., 2005. Insulin-like growth factor signaling in
643 fish. *Int. Rev. Cytol.* 243, 215-285.

644

645 Whitehead, J. P., Clark, S.F., Urso, B., James, D. E., 2000. Signalling through the
646 insulin receptor. *Curr. Opin. Cell. Biol.* 12, 222-228.

647

648 Zhang, W., Gustafson, T.A., Rutter, W.J., Johnson, J.D., 1994. Positively charged
649 side chains in the insulin-like growth factor-1 C and D-regions determine receptor
650 binding specificity. *J. Biol. Chem.* 269, 10609-10613.

651

652

653

654

655

656

657

658

659

660

661

662

663

664

665

666

667

668

669

670 **Figure legends**

671 Table 1. Primer sequences used in IGF-1R and IGF-1 cDNA cloning. W = A/T, R =
672 G/A, Y= T/C, S= G/C, K = G/T.

673

674 Figure 1.A) Chilean flounder IGF-1R cDNA complete sequence (GenBank number
675 FJ438475). Deduced amino acid sequence is indicated below the nucleotide sequence.
676 Start and stop codon are underlined. 5' and 3' UTR are indicated in small letters. The
677 signal peptide sequence is indicated by cursive letters. Alfa subunit and beta subunit are
678 underlined and boxed respectively. Transmembrane domain is indicated in bold
679 character. B) Chilean flounder IGF-1 cDNA complete sequence (GenBank number
680 EU017533). Deduced aminoacidic sequence is indicated below the nucleotide sequence.
681 Start and stop codon are underlined. 5' and 3' UTR are indicated in small letters. The
682 signal sequence is indicated by cursive letters. B and C domains are underlined and
683 bolded respectively. A, D and E domains are shown by the boxed, gray boxed and black
684 boxed areas respectively.

685

686 Figure 2. Amino acid multiple sequence alignments of IGF-1R and IGF-1 in different
687 vertebrates species: A) GenBank numbers: human (*Homo sapiens*) GenBank:
688 X04434.1; rat (*Rattus norvegicus*) GenBank: AF056187.1; chicken (*Gallus gallus*)
689 GenBank: AJ223164.1; African clawed frog (*Xenopus laevis*) GenBank: AF055980.1;
690 zebrafish (*Danio rerio*) GenBank: AF400275.1; Japanese flounder (*Paralichthys*
691 *olivaceus*) GenBank: AB065098.1, Chilean flounder (*Paralichthys adspersus*)
692 GenBank: FJ438475.1. Conserved cysteine and tyrosine residues are indicated in black
693 boxed areas. The proteolytic cleavage sequence and potential ATP binding sites are
694 shaded. Potential IRS-2 and IRS-1 binding sites are in bold characters. Cystein,

695 transmembrane, juxtamembrane, tyrosine kinase and carboxy domains area indicated in
696 open boxed areas .B) GenBank accession numbers: human (*Homo sapiens*) GenBank:
697 M27544.1; rat (*Rattus norvegicus*) GenBank: NM_001082479.1; chicken (*Gallus*
698 *gallus*) GenBank: M32791.1; African clawed frog (*Xenopus laevis*) GenBank:
699 M29857.1; zebrafish (*Danio rerio*) GenBank: BC114262.1; Japanese flounder
700 (*Paralichthys olivaceus*) GenBank: AJ010602.1; Chilean flounder (*Paralichthys*
701 *adpersus*) GenBank: EU017533.1. Conserved cysteine residues are indicated by black
702 boxed areas. Conserved residues implicated in IGF-1R binding are indicated in a grey
703 boxed area. IGFBP binding is indicated in bold characters. B, C, A, D and E domains
704 are indicated in open boxed areas.

705

706 Fig.3. Comparison of the Chilean flounder IGF-1 and IGF-1R deduced domains with
707 Japanese flounder, turbot, carp, zebrafish, xenopus, chicken, rat and human IGF-1R and
708 IGF-1. A) GenBank accession numbers: human (*Homo sapiens*) GenBank: X04434.1;
709 rat (*Rattus norvegicus*) GenBank: AF056187.1; chicken (*Gallus gallus*) GenBank:
710 AJ223164.1; African clawed frog (*Xenopus laevis*) GenBank: AF055980.1; zebrafish
711 (*Danio rerio*) GenBank: AF400275.1 and BC163723.1; Carp (*Cyprinus carpio*)
712 GenBank: AY144591.1; turbot (*Psetta maxima*) GenBank: AJ224993.1; Japanese
713 flounder (*Paralichthys olivaceus*) GenBank: AB065098.1, Chilean flounder
714 (*Paralichthys adpersus*) GenBank: FJ438475.1. The top line drawing represents the
715 cDNA structure of human IGF-1R. The comparison between Chilean flounder and other
716 species were performed using 170, 43, 253 and 155 amino acid corresponding to the
717 cystein domain, juxtamembrane domain, tyrosine kinase domain and c-terminal domain,
718 respectively. B) GenBank accession numbers: human (*Homo sapiens*) GenBank:
719 M27544.1; rat (*Rattus norvegicus*) GenBank: NM_001082479.1; chicken (*Gallus*

720 *gallus*) GenBank: M32791.1; African clawed frog (*Xenopus laevis*) GenBank:
721 M29857.1; zebrafish (*Danio rerio*) GenBank: BC114262.1; Carp (*Cyprinus carpio*)
722 GenBank: BAA11878.1; turbot (*Psetta maxima*) GenBank: ACL14947.1; Japanese
723 flounder (*Paralichthys olivaceus*) GenBank: AJ010602.1; Chilean flounder
724 (*Paralichthys adspersus*) GenBank: EU017533.1. The top line drawing represents the
725 cDNA structure of human IGF-1. The comparison between Chilean flounder and other
726 species were performed using 29, 10, 21, 8 and 73 amino acid corresponding to B, C, A,
727 D and E domains, respectively.

728

729 Fig. 4. Qualitative mRNA distribution of the Chilean flounder IGF-1R and IGF-1 in
730 different tissues assessed by RT-PCR. β -actin was used as constitutive expression
731 control. (figure is representative of four independent experiments).

732

733 Fig. 5. Expression pattern of IGF-1 and IGF-1R mRNAs in Chilean flounder larvae.
734 Expression of IGF-1 and IGF-1R mRNA were analyzed at 9 dpf in Chilean flounder
735 larvae, through whole-mount *in situ* hybridization using sense and antisense probes. A,
736 larvae at 9 dpf show strong IGF-1 expression in the notochord (nc). B, higher
737 magnification shows the expression in the notochord (nc), but not in somites (s). C,
738 larvae at 9 dpf show IGF-1R expression in the notochord, somites and the head (h). D,
739 higher magnification shows expression in the notochord and somites. We did not detect
740 positive signals using the sense probe (insets in A and B, respectively). Pictures are
741 representative of four independent experiments. Abbreviations: h, head; nc, notochord;
742 s, somites.

743

<i>primer name</i>	<i>sequence</i>	<i>nucleotide position</i>
1F(sense)	5'-TGAGWTRACCAGCCTGAAGGAC-3'	1000-1021 igf1r
1R(antisense)	5'-CTTTRAGCAGTAGTTGTGYTG-3'	2582-2603 igf1r
2F(sense)	5'-CTTGTTTTGGRTCTGATGCTG-3'	654- 676 igf1r
2R(antisense)	5'-AGCAGAGCTCAGGGTTCTTCTC-3'	1070- 1091 igf1r
3F(sense)	5'-GCCWTTCCAGAACATCACAGAG-3'	2200- 2221 igf1r
3R(antisense)	5'-ATCAGYTCRAACAGCATGTCAG-3'	4326- 4347 igf1r
4F(sense)	5'-AGCCTCCGTCATGAAGGAGTTC-3'	3796- 3790 igf1r
4R(antisense)	5'-TTGAACTCCTTCATGACGGAGG-3'	3770- 3792 igf1r
5F(sense)	5'-GACCTTGCTCTCAAGTGCAC-3'	1476- 1495 igf1r
5R(antisense)	5'-TGTGACTCGTTGTCTCGTTAG-3'	2100- 2120 igf1r
6F(sense)	5'-AGACATGGATGACAACACCAAG-3'	2644-2665 igf1r
6R(antisense)	5'-TGAAGTGGAGCTTGGTCAGAC-3'	3291-3310 igf1r
7F(sense)	5'-GGATGTCACCAGAGTCTCTGA-3'	4146- 4166 igf1r
7R(antisense)	5'-TTGATGCTGTGATGATCTCCA-3'	4392- 4413 igf1r
1F(sense)	5'-CCTCTCCACTRCTGCTGVGTGTC-3'	233-254 igf1
1R(antisense)	5'-ATRTCTGTGTGSCGTTGTGCWC-3'	499-520 igf1

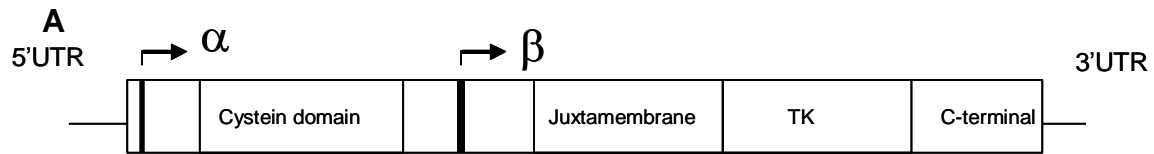
A

1 - cagcctcgagccaccaccctgtggatctggagcttattttctgccgccaccctaacctctctctctccacccttctctgccattct
91 - ccctgggcttactcaccatgttctgtggagtctgctgggtaataaaaccttcaogttatctttgacatataatattttcagttgtgtac
181 - caccagcccggtcgcaccgttaacgcgcactcgtgttttcaagagatggagactgaagatggggatccatctgtatccagatca
271 - aattactggcatgtgtgccacaccagatccaagatctctgtgactggcggcaaaacgggagactctattaatattttatgttctctt
361 - tttttctctccatgggaactccctgctataacaattctaaacatttttagattaaaaaataatattttattctaaatctaatcctgg
451 - tagacaagcaaccacgctttgcatgctaccatagagaagccactgggtacacacatttttttctcatttttaaattattgaag
541 - ctctatttttccagaggatttttctactttggctccctttgcttatttttaatttctgagcggagaatgatgttgaggaccATGAGATC
1 - M R S
631 - TGGCTACTGATGGGTAGCAGACCTTGTTTGGGGTCTGATGCTGTCTGCTCCACTATCTGCATATGGCCCAGTATGGAGAGATTTG
4 - G S L M G S T T L F W G L M L S V S T I C I W P T Y G E I C
721 - TGGTCCGGTATTGACATCCGAAATGACATCAGTGAAGTCAAGCGGCTGGAGAAGTGCACGGTGGTGGAGGGCTACCTGCAGATCCTCTCT
34 - G P G I D I R N D I S E F K R L E N C T V V E G Y L Q I L L
811 - CATCAATGACAACCAACAACATCCATCAAGAGGTTTTCCGCTCCCTCAGCTTCCCAAGTGACCATCAAGGACTTCTGCTGTCT
64 - I N D K K T N I N I H Q V F R S L S F P K L T I I T D Y L L C L
901 - GTTTCGCTCTCTGGCTTGGACAGCTCAGCATGCTCTTCCCAACCTGAGCATCATCCGTGGGGGAGCTTCTTACAACACGCCCT
94 - F R V S G L D S L S M L G C P T N L S I I R G R Q L G F Y N Y A L
991 - CGTGATCTATGAGATGACCAGCCTGAAGACATTTGCTGTACAACTGAAGAACATCACCAGGGGAGCAGGAGTGTGAGAAGAACCC
124 - V I Y E M T S L K D I G L Y N L K N I T R G A T R I E K N P
1081 - TGAGCTCTGCTACCTGACTCGGTGGACTGGTCTCTATTGATGGATGCGGAGTTCAACAACATTATCAATGGAAATAAGAAGGCTAAGGA
154 - E L C Y L D S V D A A Q S L L K D C T V I E G N L D I N I R
1171 - GTGCGACAATGTCTGCCAGGAATCATGGAGATAAACCCTTTGTAAGAGGACGTTGTTCAATGACAACACGACTACCGCTGCTGGAC
184 - C D N V C P G I M E D N P L C K R T L F N D N Y D Y R C W T
1261 - CTCTACCCAGTGCCAGAAGTTTGGCCAGAACACTGCAAAATATGCTGTACCGACAAGGGGAGTGTCCACAGCCAGCTGCTGGCCAC
214 - S T Q C Q K V C P E H C K Y A C T D K G E C C H S Q C L G T
1351 - CTGCACTGAAACCAACACGACATGGCTGTCCACCTGCCACTTCCACGAGGAGCCGCTGCGTGGCAGACTGTCCTCCGGGCAT
244 - C T P N I I D S V D A A Q S L L K D C T V I E G N L D I N I R
1441 - TTACAAGTTCGAGAGCTGGCGTGCATCACCATGGGACCTTGCTCTCAAGTGCACCTGCCTGTCGACCTCAGTTTGTTCATCCAGGGGG
274 - Y K F E S W R C I T M G P C S Q V H L P V D P G F V I H G G
1531 - AGAATGTATGATGAATGCCCTCTGGCTTACACGAAACGAGACTAATCGAATGTTTTCAGCGCCTGCAACGGACTTGGCAGCAAGT
304 - E C M H E C P S G F T R N E T N R M F C S A C N G L C D K V
1621 - CTGCACACCAACATCATGACTCTGTGGATGCTGCTCAGTCTCTGAAGGACTGCACCGTATCGAGGGCAATCTGGACATCAACATTCG
334 - C T P N I I D S V D A A Q S L L K D C T V I E G N L D I N I R
1711 - TCCGCGAAATAACATAGCTCTGAGCTGGAGAGCTTTCATGGATGTATCCAGACAGTGAAGGGCTATGTGAAGATTTCGACACTCCCACGC
364 - R G N N I A S E L E S F M G L I Q T V K G Y V K I R H S H A
1801 - GCTTGGCTCGTGTCTTCCCTCAAGAGCTGCGTTACATCAATGGCAGGAACCTATCGACAACATGTATTCTCTCCGCCATCAACAA
394 - L G S L S F L K S L R Y I N G Q E L I D N M Y S F S A I N N
1891 - CCAGCACTGGCAGTACCTGTGGGACTGGAGTCAACAACTGACTATTTCGAGCTGGAGCCTCTTCTTTCGCGGAAACCCCAACTCTG
424 - Q H L Q Y L W D W S Q H N L T I R A G R L F F R R N P K L C
1981 - CATGCTGAGATCCACACCATGTGGGAAAAGACGAAGATCACCAGCAAGCGGAGGAGGATTTCCGCAACAACGGTGAAGAGCCAG
454 - M S E I H T M W E K T K I T A K P E E G D F R N N G E R A S
2071 - CTGTGAAAGTCAACCTGACGTTCAAGACTAACGAGACAACGAGTCAAGTCAAGCTGACGTGGGAGGACTACCAGCCACGACTT
484 - C E S H T L T F K T N E T T S H M I K L T W E R Y Q P P D F
2161 - CGGAGACCTCATGACTTCACTGCTTCAAGGAGTGCCTTTCCAGAACATCACAGAGTTCGACGGACAGGACGGCTGCGGCTCAAA
514 - G D L I S F I V Y F K E S P F Q N I T E F D G Q G G S N
2251 - CAGCTGGCAGATGGTGGACGTGGATCTACCTCAGGATAAAACAGTGAACCAATGTAGTCTCCGACCTGAAGCCCTGGACCCAGTA
544 - S W H M V D V D L P Q D K T S E P N V S L P H L K P W T Q Y
2341 - TGCCATCTTCGTGAAGGCCATCACCTGCAGGTGGAGGACAACACACTCAGTGGGCGCAAGAGTGCATCATCTCCGACAGCCGCC
574 - A I F V K A I T L Q V E D K H I T G A K S D I I Y I R T R P
2431 - ATCGTCTCTTCTGTGCCATAAAGACGCCCGCGCTTTTGCACCTCCTCAACCAAGCTGGTGGTGAAGTGGTGCCTCCCTGTTCCCAA
604 - S P S V P K D A R A F A N S S T K L V V K W S P P V F P N
2521 - CGGCAACCTGACCTACTACCTGGTCCGTGGCAGCAGAGCCGAGGACAGGAGCTATACCAACAACACTACTGCTCCAAGAGAGCTGAA
634 - G N L T Y Y L V R W Q Q Q P E D A R E L Y Q H N Y C S K E E L K
2611 - GATCCCGATCAGGATTTACGCGACAGGGCTCACAGACATGGATGACACACCAAGCCCAAGTCAAACTTGGGGGGGAGGAGCAAGG
664 - I P A T R I S A T G L T D M D D N T K P H Q V K L G G G R E C A
2701 - GCCATGTGCCTTGCGAAGACCGCAGGAGGACCGGAGGAGGACCGCGTCTTCTGAAGATTTTGGAGACTTCTCTCCCAA
694 - P M C L A E D A E E K D R E K D D R V F L K I F E A N F L H N
2791 - TGCCATCTTCTGCGGACCTCCAGACCGTCCAGCAGAGATGTGTTCCGGCTGGCCAACGACACGCTGTTTCCAGCAGCGCCGGGAA
724 - A I F L P R P P P D R R R R D V F G V A N D T L F H D S A G K
2881 - GGGGAACACCCCTCGCGCGGCAACAGTACAGATGGCTTCTCTATTAAGAGTACCCTTTCATGGGAGCAAGAGCTCAGCAGA
754 - S N T T L G P G N S T D G V P P I K E Y P F M E D K S S A E
2971 - ATATTTAGACATCCCAACCTGCAGCCCTCACAGTCTACCCTTGACATCCACGCTGCAATGAGGAGTGGGCGCTGCGAGCCCGG
784 - Y L D I P N L Q P F T V Y R L D I H A C N E E V G R C S A G
3061 - AGCATTCTGCTTCTCCAGACCAACCTGCGGTCAAAGCAGACGACATCCCTGGAAAAGTATCTATGAGCGCAGTACAAGGTTGAGGG
814 - A F V F S Q T K P A V K A D D I P G K V I Y E R S D D K V E G
3151 - TTCTGTGTGCTGACTGGCCAGCCATCATGCCAACGAGCTCATCTCGATGTATGAGATCAAGTCCGTTTGGGAGTGGCCTGA
844 - S V V L H W P E P I M P N G L I L M Y E I K F R L G T E P E
3241 - GAAACACGAGTGTGTGCGCGCAGCACTACCCTGAGCAGAGGAGCTGCTGACCAACCTCAGTTTCAAGAACTACTCTGCCGTGT
874 - K H E C V S R Q H Y R E H R G A R L T N L S S G N Y S A R V
3331 - GCGCGCCACTTCTTAGCAGGGAACCGCTCCTGGACAGAGCGTGTTTTCTACGTGCTCCACCAACAGAGACGATGGTGTGCTT
904 - R A T S L A G N G S W T E S V F F Y V P P P K R D D G V A F
3421 - TTATTTGGTATCATAATTTCCATGATAGCAGCTCCTCCTTGTGCCAGCCTACCACCATTTCTTCTTTGTGAACAAAAGAGGAAACAG
934 - Y L V I I I P I A A C T L I A S L T T I L F F V N K K R N S
3511 - CGACAGACTGGGAAATGGAGTCTTATGCTCTGCTCAATCCAGAATACTTACGCGCTGCTGAGATGTACGTCGGGATGAGTGGGAGT
964 - D R L G N G V L Y A S V N P E Y F S A A E M Y V P D E W E V
3601 - AGCAGGAGAGAAGTACTATGCAAGGAGCTGGCCAGGCTTCTTCCGATGGTGTATGAAGTTTAGCCAAGAGTGTGGTCAAGGA
994 - A R E K I T M H K E L G Q G S F G M V Y E G L A K S V V K D
3691 - TGAGCCTGAGACCGGAGTAGCCATCAAGACGGTCAACAGAGTCCGCGAGCATGAGGAGCGCATCGAGTTTTTGGAGCAGGCCCTCCGTCAT
1024 - E P E T R V A I K T V N E S A S M R E R I E F L D E A S V M
3781 - GAAGGAGTTCAACTGCTCACCATGTGGTCCGGTCTGGTCTCTCAGGGTCAAGCCACTCTGGTCAATCATGAGGAGTGGTCAAGCAG
1054 - K E F N C H H V V R L L G V V S Q G Q P T L V I M E L M T R
3871 - TGGAGATCTCAAGACCCACTGCGCTCGCTCGCGAAGGAAACTCCACCACCCAGGCTTACCCCACTCAAAAATATCCAGATGGC
1084 - S D L K S H L R K E N S T T Q V L P P L K K M I Q M A
3961 - GGGGAAATCGCTGACGGCATGGCTACCTTAAACGCCAACAGTTTGTCCACAGAGTCTGGCAGCCAGGAACTGCATGGTGGCTGAGGA
1114 - S E I A D G M A Y L F N A N K F V H R D L A A R N C M V A E D

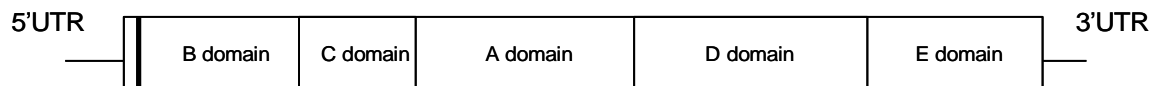
4051 - CTTTCATCGTGAAGATTGGAGATTTTGGCATGACCAGAGACATATATGAGACAGATTACTACCGCAAAGGTGGTAAGGGTCTGCTCCCTGT
 1144 - F I V K I G D F G M T R D I Y E T D Y Y R K G G K G L L P V
 4141 - CCGCTGGATGTCACCAGAGTCTCTGAAGGATGGAGTCTTTACTACTAACTCTGATGTTTGGTCGTTTGGGGTGTACTGTGGAGATTTT
 1174 - R W M S P E S L K D G V F T T N S D V W S F G V V L W E I S
 4231 - CACCCTGGCTGAGCAGCGTACCAGGGTCTGTCCAATGAGCAGGTGCTCCGCTTTGTTCATGGAGGGAGACTGTGGAGAAACCAAAA
 1204 - T L A E Q P Y Q G L S N E Q V L R F V M E G G L L E K P Q N
 4321 - CTGTCCTGACATGCTGTTTGTAGCTGATGCGAATGTGTGGCAGTACAATCTAAAATGCGTCCATCCTTTGTGGAGATCATCAGCAGCAT
 1234 - C P D M L F E L M R M C W Q Y N P K M R P S F V E I I S S I
 4411 - CAAAGATGAAGTGGATTCTCCCTTCAGGGAAATGGGTTTCTTCTACAGTGAAGAACAAGCCGCTGACCCGAGGAGCTGGACATGGA
 1264 - K D E L D S P F R E M G F F Y S E K N K P P D T E E L D M E
 4501 - GGTAGAAAACATGGAGAACATTCCTACTGGACCCTGCATCCACCAGGCAGCCCTCTGCTGCCGCCGCCCTCGTCGGGGTGCACGGGAGG
 1294 - V E N M E N I P L D P A S T R Q P S A A A A P S S G C T G G
 4591 - AACCGCGCCCTCTGCGCAGCAGTTATCCCCATGCAAGGCCGAGTACTCTTTACTGGGACTGTGTCTCCCTCCTCCCGGGCC
 1324 - T P P P S A Q Q L S P M Q G P S T P L L G P V S P S S P G E
 4681 - GGTGCTCAGCCTTGGCGTCTCCGGCCAAAGCTTTGGACAAGCACTCAGGACATGTCTCGGCCAACGGCCCTGTGGTGGTGTCTCGGCC
 1354 - V A S A L A S P G Q A L D K H S G H V S A N G P V V V L R E
 4771 - CAACCTTGATGAGATGCAACCTTATGCACACATGAACGGGGCAGAAAGGACGAACGGGCATTACCCTGCCCCAGTCGTCGGCTGCTG
 1384 - N L D E M Q P Y A H M N G G R K D E R A L P L P Q S S A C *
 4861 - Atatcagaccagtcctcattctcctcttaacacaggtaaaccaaaccctcctcctcatctcttaaggaggatgggtgaggtaagaatga
 4951 - cttgcttttctactgtgtgccagtagatggtctctccactcctttaaggtcaaactcaaatgtaaaactgaaaaaaaaa

B

1 - tcgaggatccgaacactcggttttgctggtttgatgaaatgaaatgaaatagtggtttgtatatatgagcatgagctgctgctggtgct
 91 - gctgctgctgctgctgctgctgcttctcttctcgcgggctttgaactgcccagagaccggtgggATGTCTAGCGCTCTTTCTTTTCAGTGGC
 1 - M S S A L S F Q W H
 181 - ATTTATGTGATGTCTTCAAGAGTGGATGTGCTGTATCTTGTAGCCACACCCTCTCACTACTGCTGTGTCTCACCTCACCCTGACTCCGA
 11 - L C D V F K S A M C C I S C S H T L S L L L C V L T L T P T
 271 - CGGCAACAGGGGGGGACCGGAGACCCCTGTGCGGGGGGAGCTGGTCGACACGCTGCAGTTTGTGTGGAGAGAGAGGCTTTTATTTCA
 41 - A T G A G P E T L C G A E L V D T L Q F V C G E R G F Y F S
 361 - GTAAACCAACAGGTTATGGCCCCAATGCACGGCGGTACGCGGCATTGTGGACAGTGTCTTCCAAGCTGTGAGCTGCGGCACCTGG
 71 - K P T G Y G P N A R R S R E I V D E C C F Q S C E L R H L E
 451 - AGATGTACTGTGACCTGCAAGACTAGCAATGCCCTGCGCTCTGTGCGTGCACAACGCCACACAGACATGCCGAGAGCACCCTAAGGTTA
 101 - M Y C A P A K T S N A A R S V R A Q R H T D M P R A P K V S
 541 - GTACCGCAGGGCACAAGTGGACAAGGGCAGAGCGTAGGACAGCAGCAGCCAGACAAGACAAAAACAAGAGAGACCTTTACCGG
 131 - T A G H K V D K G T E R R T A Q Q P D K T K N K K R P L P G
 631 - GACATAGTCACTACAAGCTTTGCTTTTCATGCGCCAAAGCCAGCTGCTTACATTTTGTGTAGGAATTGTATGTGAATGatgttaacctg
 161 - H S H S Q A L I L F M R Q S Q L I T F C V G I V C E *
 721 - ttcagaggattgataccactcacaatctgttcatcttagtataaaactacaaccagcaaacatgtatgttatcattcactcggattgat
 811 - gcaacatgttttttagtatactcatagtatctatgagttggtcagactaaatctggttgtgtgtggataaaagcagatcaaatcactgagct
 901 - tcaacacattatgtctttaatacaacaaaacttaaaaaaaaaaacctatagttggagtcgtatataatcoggatccg



	<i>cysteine domain</i>	<i>juxtamembrane domain</i>	<i>Tirosine_kinase domain</i>	<i>C_terminal domain</i>	<i>Whole protein</i>
Ch_flounder/j_flounder	95	97	96	94	97
Ch_flounder/turbot	63	95	91	38	67
Ch_flounder/carp	72	95	92	63	76
Ch_flounder/zebrafish	69	92	92	66	74
IGFR1a					
Ch_flounder/zebrafish	58	92	92	50	68
IGFR1b					
Ch_flounder/xenopus	50	93	90	48	62
Ch_flounder/chicken	52	90	91	47	63
Ch_flounder/rat	48	86	91	42	61
Ch_flounder/human	49	88	91	41	61

B

	<i>B domain</i>	<i>C domain</i>	<i>A domain</i>	<i>D domain</i>	<i>E domain</i>	<i>Whole protein</i>
Ch_flounder/j_flounder	97	100	95	88	73	97
Ch_flounder/turbot	100	100	91	88	49	96
Ch_flounder/carp	97	58	95	25	27	74
Ch_flounder/zebrafish	83	67	95	38	27	73
Ch_flounder/xenopus	97	67	86	50	26	66
Ch_flounder/chicken	93	42	91	38	26	69
Ch_flounder/rat	93	42	86	38	25	65
Ch_flounder/human	86	42	86	38	23	60

Figure 4

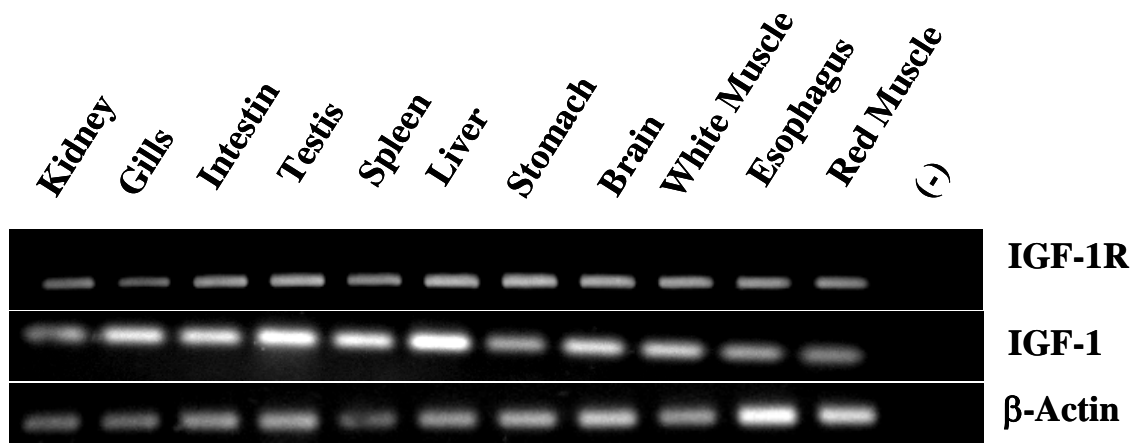


Figure 5
[Click here to download high resolution image](#)

