1	Genome Sequence of Lactobacillus pentosus IG1, a strain
2	isolated from Spanish-style green olive fermentations
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26 Abstract

Lactobacillus pentosus is the most prevalent lactic acid bacteria in the Spanish-style
green olive fermentations. Here we present the draft genome sequence of *L. pentosus*IG1, a bacteriocin producer strain with biotechnological and probiotic properties
isolated from this food fermentations.

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32 Lactobacilli have been largely associated with food fermentations and, more recently, 33 they have been used as probiotics since they may promote health in man and animals 34 (6). Lactobacillus pentosus is the most frequent lactic acid bacteria isolated from 35 Spanish-style green olive fermentations, and has been successfully used as starter 36 cultures for these fermentations (8). In addition, certain strains of L. pentosus have 37 shown to possess probiotic effects, improving the mucosal immunity and the resistance 38 to bacterial infections (2, 5). This genome sequence will allow us to explore the 39 biotechnological and probiotics properties of Lactobacillus pentosus IG1, a bile-40 resistant strain displaying bacteriocin activity against a wide range of spoilage and 41 pathogen bacteria.

The whole genome of IG1 was sequenced using a paired-end strategy by pyrosequencing technology with the platform 454 Life Sciences, GS FLX Titanium system. The sequence data consisted of 8-kb paired-end libraries (481,661 reads totalling ~ 175, 5 Mb, ~ 45-fold coverage of the genome). The paired-end reads were assembled into 13 scaffolds (3,982,537 nt), of which the scaffold 1 (3,687,424 nt) corresponds to the entire chromosome, while scaffolds 2 (125,904 nt), 3 (77,782 nt), 4 (36,648 nt), 5 (21,059 nt), 6 (10,521 nt), 11 (2,654 nt) and 12 (2,509 nt) seem to belong

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49 to seven distinct plasmids, what is according to the plasmid profile obtained for *L*.
50 *pentosus* IG1.

The draft genome of *L. pentosus* IG1 includes 3,982,537 nt bases, being the biggest genome of lactobacilli described to date. The G+C mol% content of the chromosome (scaffold 1) was determined to be 44, 9%. There is a single predicted copy of the 16S and 23S rRNA, 3 copies of the 5S rRNA and 44 predicted tRNAs. The genome contains 3,133 predicted CDS, of which 2972 belong to scaffold 1.

56 Comparative genomic analysis indicated that the closest genome is that of 57 Lactobacillus plantarum (4, 9, 10). Putative functions could be assigned to 82% of the 58 encoded proteins, and the remaining 18% could not be placed (putative uncharacterized 59 proteins). Remarkably, this strain harbours sixteen putative two-component regulatory 60 systems, which may reflect a great ability of adaptation to the changing environmental 61 conditions. L. pentosus IG1 presents several genes that could encode putative probiotic 62 functions involved in adhesion (collagen-binding proteins, exopolysaccharides, 63 lipoteichoic acids), resistance to stress (mannose phosphotransferase systems, bile salt 64 hydrolases) and microbial competition (lux S-mediated production of AI-2 and AI-3-65 like molecules and bacteriocin production). The genes responsible for the bacteriocin 66 activity of L. pentosus IG1 are identical to those of the two-component class IIb 67 bacteriocin plantaricin S of L. plantarum LPCO10 (3). Finally, we found a locus that 68 resembles the gene cluster of plantaricins, which is typical of the L. plantarum species 69 (1, 7).

Nucleotide sequence accession number. The 13 scaffolds sequences of *Lactobacillus pentosus* IG1 were deposited in the EMBL database with accession numbers FR874848
 to FR874860.

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80	References
81	
82	1. Diep, D. B., D. Straume, M. Kjos, C. Torres, and I. F. Nes. 2009. An overview of
83	the mosaic bacteriocin pln loci from Lactobacillus plantarum. A review. Peptides
84	30: 1562-74.
85	
86	2. Izumo T., F. Izumi, I. Nakagawa, Y. Kitagawa, H. Shibata, and Y. Kiso. 2011.
87	Influence of Lactobacillus pentosus S-PT84 Ingestion on the Mucosal Immunity of
88	Healthy and Salmonella Typhimurium-Infected Mice". Biosci. Microflora, 30: 27-35.
89	
90	3. Jiménez-Díaz, R, J. L. Ruiz-Barba, D. P. Cathcart, H. Holo, I. F. Nes, K. H
91	Sletten, and P. J. Warner. 1995. Purification and partial amino acid sequence of
92	plantaricin S, a bacteriocin produced by Lactobacillus plantarum LPCO10, the activity
93	of which depends on the complementary action of two peptides. Appl. Environ.
94	Microbiol. 61: 4459-4463.
95	
96	4. Kleerebezem, M., et al. 2003. Complete genome sequence of Lactobacillus
97	plantarum WCFS1. Proc. Natl. Acad. Sci. U. S. A. 100:1990-1995.
98	

99	5. Kotani, Y., et al. 2010. Oral intake of <i>Lactobacillus pentosus</i> strain b240 accelerates
100	salivary immunoglobulin A secretion in the elderly: a randomized, placebo-controlled,
101	double-blind trial. Immun. Ageing 7:11.
102	
103	6. Lebeer, S., J. Vanderleyden, and S. C. J. De Keersmaecker. 2008. Genes and
104	molecules of lactobacilli supporting probiotic action. Microbiol. Mol. Biol. Rev.
105	72: 728-764.
106	
107	7. Maldonado, A., R. Jiménez-Díaz, J. L. Ruiz-Barba. 2004. Induction of Plantaricin
108	Production in Lactobacillus plantarum NC8 after Coculture with Specific Gram-Positive
109	Bacteria Is Mediated by an Autoinduction Mechanism. J. Bacteriol. 186: 1556-1564.
110	
111	8. Ruiz-Barba, J. L., D. P. Cahtcart, P. J. Warner, and R. Jiménez-Díaz. 1994. Use
112	of Lactobacillus plantarum LPCO10, a bacteriocin producer, as a starter culture in
113	Spanish-style green olive fermentations. Appl. Environ. Microbiol. 60:2059-2064.
114	
115	9. Wang, Y., et al. 2011. Complete Genome Sequence of the Probiotic Lactobacillus
116	plantarum ST-III. J. Bacteriol. 193:313-314.
117	
118	10. Zhang, Z. Y., et al. 2009. Complete genome sequence of Lactobacillus plantarum
119	JDM1. J. Bacteriol. 191:5020-5021.
120	
121	