



 **Peer Reviewed**

Title:

Stomatal conductance is the main limitation to photosynthesis in sugar beet plants treated with Zn excess

Author:

[Sagardoy, Ruth](#), EEAD-CSIC, Zaragoza, Spain
[Flexas, Jaume](#), Univ. Illes Balears, Mallorca, Spain
[Ribas-Carbó, Miquel](#), Univ. Illes Balears, Mallorca, Spain
[Morales, Fermín](#), EEAD-CSIC, Zaragoza, Spain
[Abadia, Javier](#), EEAD-CSIC, Zaragoza, Spain

Publication Date:

04-24-2009

Publication Info:

The Proceedings of the International Plant Nutrition Colloquium XVI, Department of Plant Sciences, UC Davis, UC Davis

Permalink:

<http://escholarship.org/uc/item/6h53f5j5>

Keywords:

photosynthesis, stomata, sugar beet, zinc toxicity

Abstract:

The effects of high Zn concentrations in growth and photosynthetic parameters of sugar beet (*Beta vulgaris* L.) plants grown in hydroponics were investigated. Zinc toxicity (100 and 300 μ M) resulted in large reductions in biomass accumulation (>50%) and photosynthetic rates (40-50%). It was known that high Zn concentrations usually lead to decreases in net photosynthesis, but the effects of excess Zn on each of the possible factors limiting photosynthesis, including photochemistry, stomatal conductance and mesophyll conductance, had not been studied in detail until now. Leaf photochemistry was not affected by Zn, and reduced photosynthesis was due mostly to marked decreases in stomatal conductance, whereas mesophyll conductance decreased to a lower extent. Stomata were study using scanning electron microscopy (SEM) and large structural differences were observed in stomatal frequency, size and shape between 300 μ M Zn and control plants. In high Zn plants stomatal density was lower than in the controls, and stomata were smaller and had a more rounded shape than those present in control plants. The presence of these morphological changes could be associated to the reductions in photosynthetic rates under excess Zn.



Introduction

In low pH soils Zn availability is generally high, and Zn concentrations within the plant could become toxic (Broadley *et al.*, 2007). Excess Zn causes changes in root growth and morphology (Vaillant *et al.*, 2005), reductions in growth (Sagardoy *et al.*, 2009) and decreases in stomatal aperture (Sharma *et al.*, 1995; Sagardoy *et al.*, 2009), whereas photochemistry is not markedly affected (Sagardoy *et al.*, 2009). An increased mesophyll resistance to CO₂ diffusion with Zn toxicity has been also suggested to occur (Van Assche *et al.*, 1980; Prasad and Strzalka, 1999); however, these studies did not use an adequate method to estimate mesophyll diffusion conductance to CO₂ (g_m), but assessed instead a parameter which reflects the combination of g_m and carboxylation activity.

The aim of this work was to investigate the changes in photosynthetic characteristics under excess Zn in the model plant sugar beet (*Beta vulgaris* L.), considering the potential photosynthetic limiting factors mentioned above.

Materials and Methods

Plant material and growth conditions

Sugar beet (*Beta vulgaris* L. Orbis) seeds were germinated and grown in vermiculite for 3 weeks in a growth chamber at 25°C and a PPF of 600 μmol m⁻² s⁻¹ with a 16 h light: 8 h dark regime. Afterwards, plants were moved to glasshouse and transplanted to 20-L plastic buckets containing half-Hoagland nutrient solution with 45 μm Fe. Treatments were 1.20 (control), 100 and 300 μM ZnSO₄. Plants were used for measurements 10-14 days after imposing the Zn treatments.

Growth parameters and mineral nutrient analysis

Plants were collected and separated in roots and shoots at 14 days. Fresh (FW) and dry weights (DW) of each fraction were measured, and water content (WC) was determined as (fresh weight – dry weight) / fresh weight. Leaf areas were measured with a AM-100 leaf area meter (ADC, Herts, U.K.). Eight plants per treatment were measured. Zinc was determined by FAAS (Igartua *et al.*, 2000).

Gas-exchange and chlorophyll fluorescence measurements

Leaf gas exchange and chlorophyll fluorescence parameters were measured simultaneously using an open gas exchange system (Li-6400; Li-Cor, Inc., Lincoln, NE, USA) with an integrated fluorescence chamber head (Li-6400-40 leaf chamber fluorometer; Li-Cor, Inc.), according to Flexas *et al.* (2007). These measurements were carried out at days 10 to 13 after the start of the treatments, at 1500 μmol m⁻² s⁻¹ with 10% blue light. Cuvette CO₂ concentration (C_a) was set at 400 μmol CO₂ mol⁻¹ air and vapor pressure deficit was kept at 2.0 ± 0.2 kPa.

Scanning electron microscopy (SEM-EMX)

The surface of fresh sugar beet leaves was visualized with an Hitachi S-3400 N microscope fitted with an EDX analyzer (Röntec XFlash Si(Li)) at the ICB-CSIC.

Results

Plant dry mass was progressively reduced with high Zn concentrations in the nutrient solution. Plants treated with high Zn contained less water and had smaller leaf areas than control plants. Zinc concentration increased significantly with 100 or 300 μM Zn in the nutrient solution, approximately 10-fold in shoots and 1.6-fold in roots (Table 1).

Zn treatment	1.2 μM	100 μM	300 μM
DW (g plant ⁻¹)	2.8 \pm 0.4 ^a	1.3 \pm 0.2 ^b	0.8 \pm 0.2 ^b
leaf area (cm ²)	136.7 \pm 5.5 ^a	48.3 \pm 5.2 ^b	29.8 \pm 3.0 ^c
WC (%)	96.8 \pm 0.2 ^a	91.1 \pm 0.1 ^b	87.6 \pm 0.3 ^c
Zn in shoots ($\mu\text{g g}^{-1}$ DW)	129.7 \pm 11.9 ^a	1223.7 \pm 64.5 ^b	1184.3 \pm 102.7 ^b
Zn in roots ($\mu\text{g g}^{-1}$ DW)	136.1 \pm 15.0 ^a	218.4 \pm 2.9 ^b	202.9 \pm 0.0 ^b

Table 1. Growth parameters of sugar beet plants grown with different Zn concentrations for 14 days. Data are means \pm SE of eight replicates. Data followed by the same letter are not significantly different (Duncan's test) at $p < 0.05$ level.

At normal CO₂ concentration (400 μM CO₂ mol⁻¹ air), leaves of plants grown with high concentrations of Zn showed marked decreases in photosynthetic rate when compared to control plants, whereas no significant differences were measured between the 100 and 300 μM Zn treatments (Table 2). Stomatal conductance was drastically reduced (by 70%), whereas mesophyll conductance was reduced less (by 44%). C_i and C_c decreased progressively (Table 2) with increased Zn. No differences were found for maximum carboxylation velocity (V_{max}) and the maximum rate of electron transport (J_{max}) (Table 2), although it was not possible to estimate J_{max} in the 300 μM treatment due to the lack of increase in photosynthetic rates at high CO₂ in these plants.

Zn treatment	1.2 μM	100 μM	300 μM
A _N ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$)	21.4 \pm 1.2 ^a	12.4 \pm 1.4 ^b	11.1 \pm 1.8 ^b
F _v /F _m	0.821 \pm 0.001 ^a	0.807 \pm 0.005 ^a	0.802 \pm 0.008 ^a
ETR ($\mu\text{mol e}^- \text{ m}^{-2} \text{ s}^{-1}$)	143.8 \pm 3.1 ^a	114.0 \pm 5.8 ^a	120.0 \pm 17.3 ^a
Φ_{PSII}	0.218 \pm 0.005 ^a	0.173 \pm 0.009 ^a	0.182 \pm 0.026 ^a
g _s (mol CO ₂ m ⁻² s ⁻¹)	0.231 \pm 0.033 ^a	0.070 \pm 0.014 ^b	0.055 \pm 0.010 ^b
g _m (mol CO ₂ m ⁻² s ⁻¹)	0.389 \pm 0.091 ^a	0.243 \pm 0.055 ^{ab}	0.204 \pm 0.048 ^b
V _{c, max} ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$)	104.4 \pm 4.5	102.8 \pm 5.0	126.5 \pm 19.1
J _{max} ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$)	128.7 \pm 6.7	122.6 \pm 1.0	-
C _i ($\mu\text{mol CO}_2 \text{ mol}^{-1}$ air)	286 \pm 8 ^a	200 \pm 11 ^b	176 \pm 17 ^b
C _c ($\mu\text{mol CO}_2 \text{ mol}^{-1}$ air)	221 \pm 19 ^a	143 \pm 14 ^b	115 \pm 9 ^b

Table 2. Photosynthetic parameters measured in intact plants grown in nutrient solution with different concentrations of Zn. Data are means \pm SE of five replicates. Data followed by the same letter are not significantly different (Duncan's test) at $p < 0.05$ level.

Leaf surface samples were scanned at 300 x and 3000 x. Zinc-stressed plants had a smoother surface, with smaller, less abundant stomata when compared to the controls (Fig. 1a, b). Stomata in the 300 μM Zn treated plants had a more rounded shape, with stomatal slits shorter than those found in the controls (Fig. 1c, d).

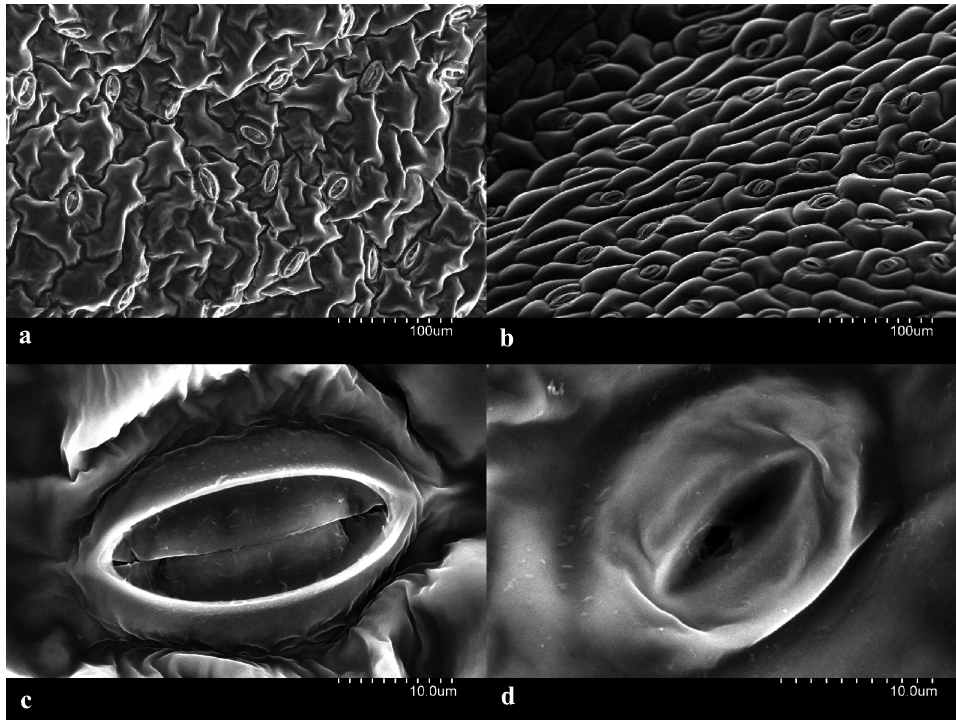


Fig. 1 Leaf surface (300 x) and stomata (3000 x) from control (a, c) and 300 μ M Zn grown sugar beet (b, d) observed with scanning electron microscopy (SEM).

Acknowledgements

Work supported by the Spanish Ministry of Science and Innovation (projects AGL2007-61948 and BFU2008-01072/BFI). R.S. was supported by an I3P-CSIC predoctoral fellowship.

References

- Broadley MR, White PJ, Hammond JP, Zelko I, Lux A. 2007. Zinc in plants. *The New Phytologist* 173: 677-702.
- Flexas J, Ortuño MF, Ribas-Carbó M, Díaz-Espejo A, Florez-Sarasa ID, Medrano H. 2007. Mesophyll conductance to CO₂ in *Arabidopsis thaliana*. *The New Phytologist* 175: 501-511.
- Igartua E, Grasa R, Sanz M, Abadía A, Abadía J. 2000. Prognosis of iron chlorosis from the mineral composition of flowers in peach. *Journal of Horticultural Science and Biotechnology* 75(1): 111-118.
- Prasad MNV, Strzalka K. 1999. Impact of heavy metals on photosynthesis. In: Prasad MNV, Hagemeyer J, eds. *Heavy Metal Stress in Plants: from Molecules to Ecosystems*. Springer, Berlin, 117-138.
- Sargadoy R, Morales F, López-Millán AF, Abadía A, Abadía J. 2009. Effects of zinc toxicity in sugar beet (*Beta vulgaris* L.) plants grown in hydroponics. *Plant Biology*, doi: 10.1111/j.1438-8677.2008.00153.x
- Sharma PN, Tripathi A, Bisht SS. 1995. Zinc requirement for stomatal opening in cauliflower. *Plant Physiology* 107: 751-756.
- Vaillant N, Monnet F, Hitmi A, Sallanon H, Coudret A. 2005. Comparative study of responses in four *Datura* species to a zinc stress. *Chemosphere* 59: 1005-1013.
- Van Assche F, Ceulemans R, Clijsters H. 1980. Zinc mediated effects on leaf CO₂ diffusion conductances and net photosynthesis in *Phaseolus vulgaris* L. *Photosynthesis Research* 1(3): 171-180.