

# Notch signaling coordinates ommatidial rotation in the *Drosophila* eye via transcriptional regulation of the EGF-Receptor ligand Argos

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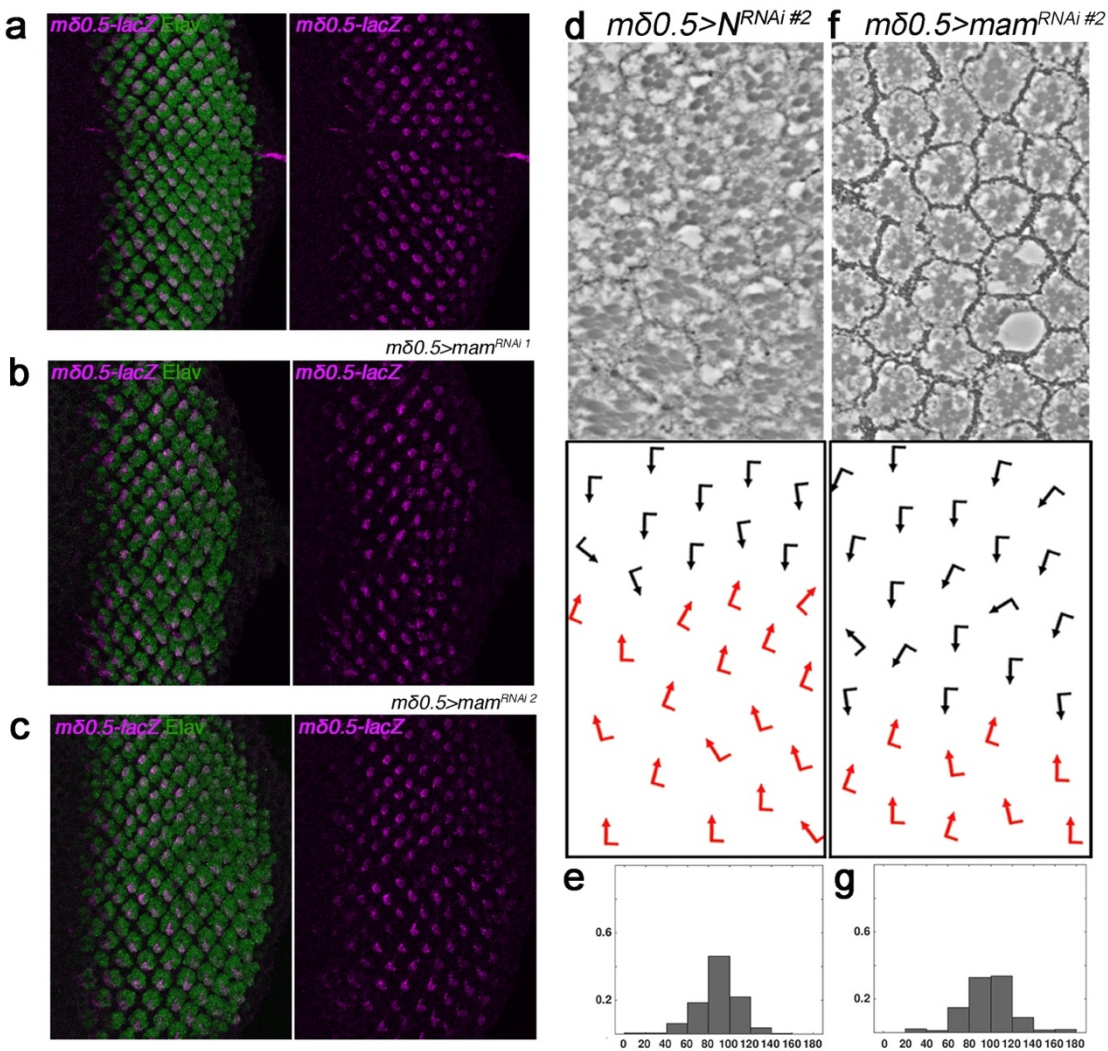
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## Key Words:

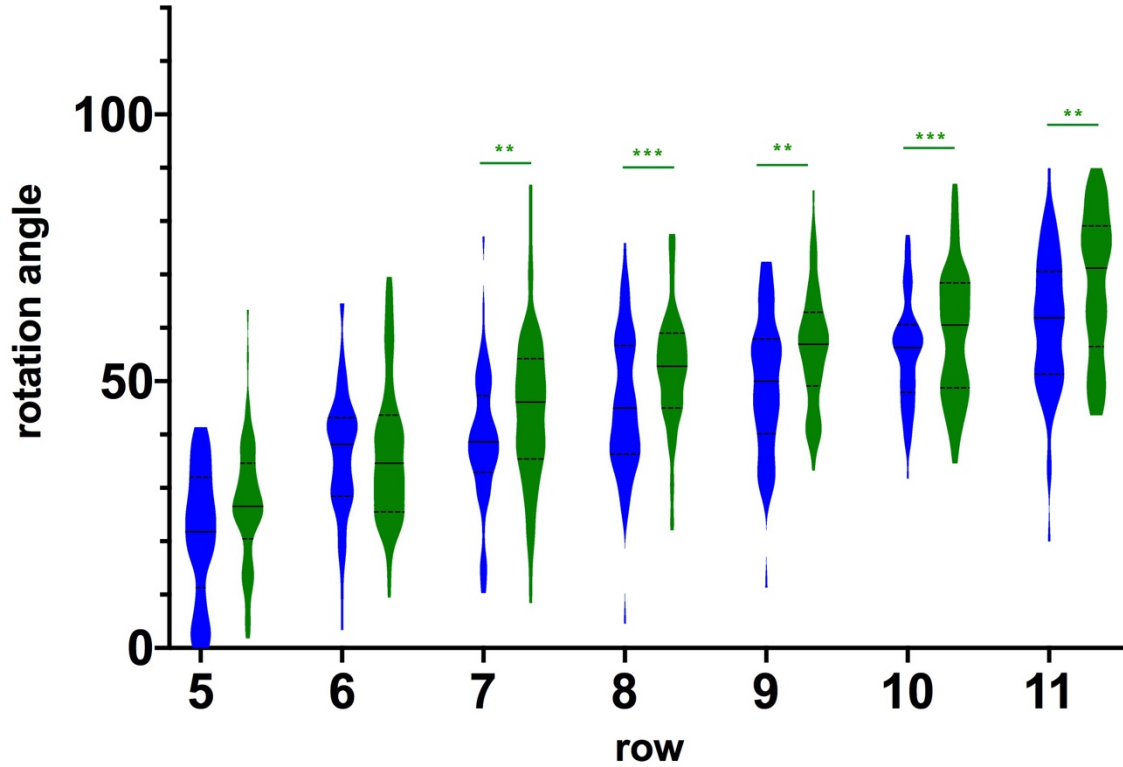
Notch Signaling, Morphogenesis, Ommatidial Rotation, Photoreceptor Specification, *Drosophila*

## Supplementary Info

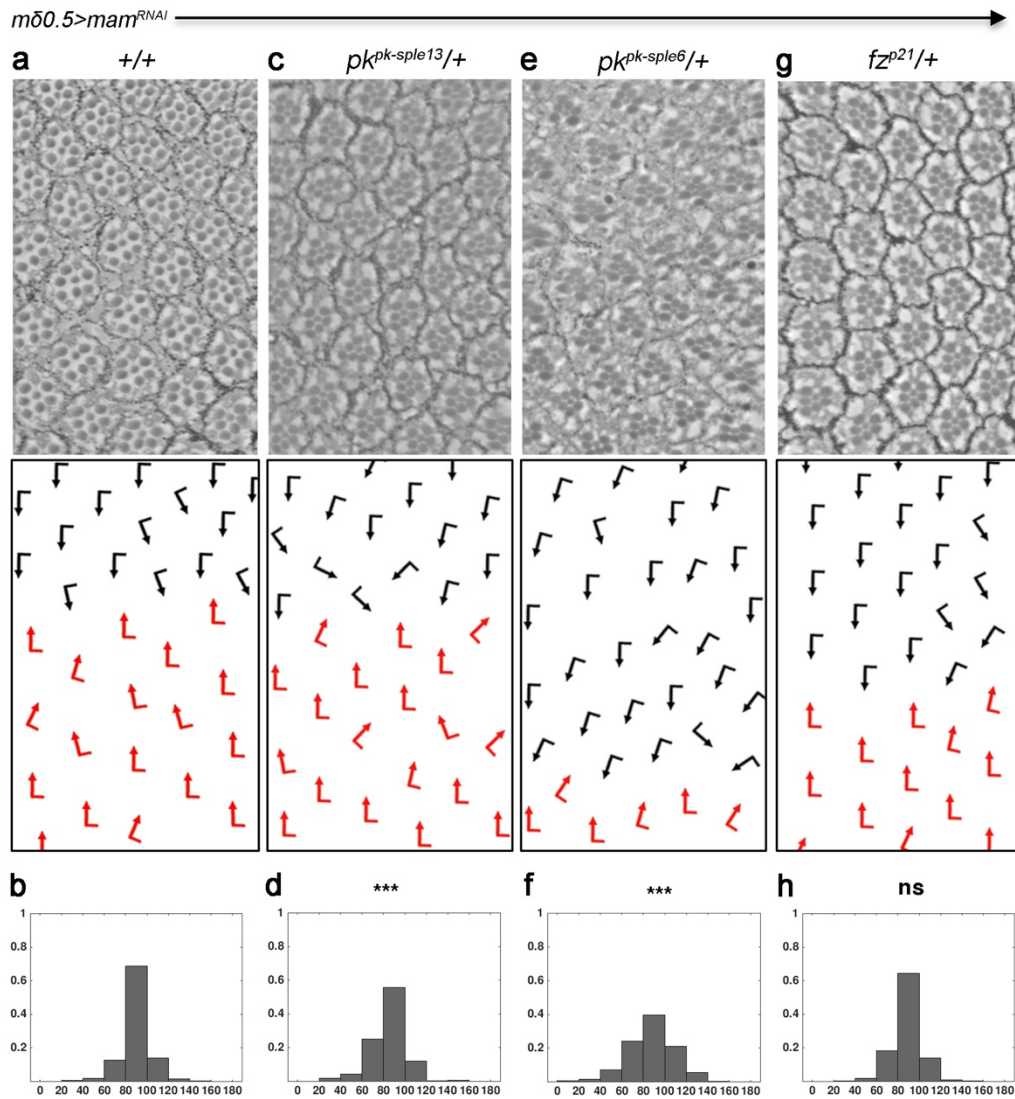


### Supplementary Figure S1. Perturbation of Notch signaling in the eye leads to misorientation of ommatidia.

(a-c) Third larval instar eye imaginal discs stained for *mδ0.5-lacZ* (magenta) and Elav (green) in wild type (a), *mδ0.5>mam<sup>RNAi</sup> #1* (b), and *mδ0.5>mam<sup>RNAi</sup> #2* (c). Note that *mδ0.5-lacZ* is specifically expressed at high levels in R4s in wild type and *mam* KD across eye discs. (d-g) Adult eye sections with ommatidial orientation schematics and orientation angle histograms for *mδ0.5>N<sup>RNAi</sup> #2* (BL7078) (d, e), and *mδ0.5>mam<sup>RNAi</sup> #2* (BL63601) (f, g), n>300, 3 eyes each.



**Supplementary Figure S2. Notch signaling is required in R3/R4 pairs to regulate OR.** Quantification of rotation angles observed in individual preclusters in rows 5-11 as plotted for control (wild type) in blue, and *mδ0.5>mam<sup>RNAi</sup>* (BL63601) in green. Statistical analyses were performed for each row between genotypes. Asterisks denote significance by chi-square test (\* p<0.05, \*\* p<0.005, \*\*\* p<0.0005).



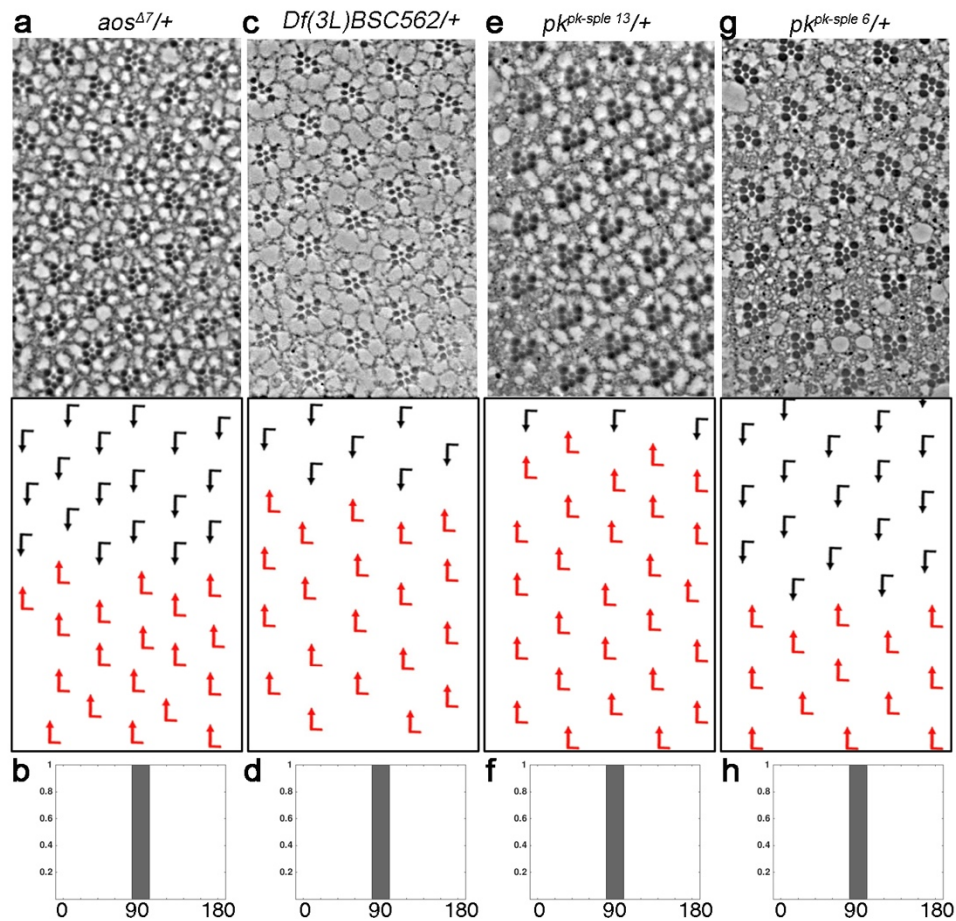
**Supplementary Figure S3. *mam* genetically interacts with *pk* to regulate OR.**

(a-h) Adult eye sections with ommatidial orientation schematics and ommatidial orientation angle histograms of eyes from *mδ0.5>mam<sup>RNAi</sup>* (BL28046) in the following genetic backgrounds: (a-b) +/+ (wt control); (c-d) *pk<sup>pk-sple13</sup>/+*, (e-f) *pk<sup>pk-sple6</sup>/+* and (g-h) *fz<sup>p21</sup>/+*. Asterisks denote significance by chi-square test (\*\*\*)  $p < 0.0005$ ). Note robust enhancement of the *mδ0.5>mam<sup>RNAi</sup>* rotation phenotype by both *pk<sup>-/+</sup>* genotypes.

<i>mδ-Gal4&gt;mam</i> <sup>RNAi BL28046</sup>	
<i>nmo</i> <sup>DB/+</sup>	<i>ns</i>
<i>aos</i> <sup>Δ7/+</sup>	<i>enhancer</i>
<i>Df(3L)BSC562/+</i>	<i>enhancer</i>
<i>egfrt</i> <sup>1/+</sup>	<i>ns</i>
<i>egfrt</i> <sup>2/+</sup>	<i>ns</i>
<i>spi</i> <sup>1/+</sup>	<i>ns</i>
<i>spi</i> <sup>2/+</sup>	<i>ns</i>
<i>shg</i> <sup>p34/+</sup>	<i>ns</i>
<i>sca</i> <sup>1/+</sup>	<i>ns</i>
<i>sca</i> <sup>2/+</sup>	<i>ns</i>

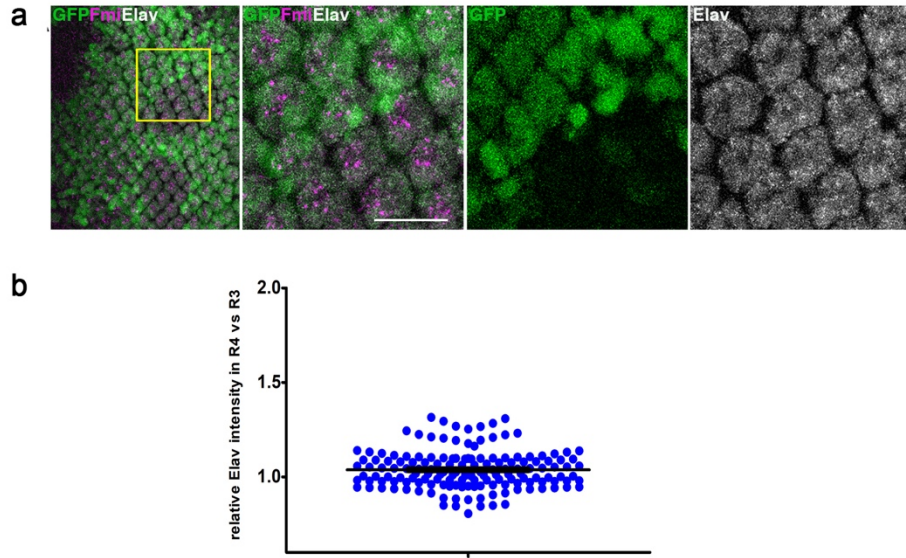
**Supplemental Table S1. Genetic interactions with ommatidial rotation associated genes that are not part of the PCP group.**

Note that besides *aos*, the other Egfr-signaling associated genes did not interact with *mδ-Gal4>mamRNAi*. Similarly, *E-cad/shg* and *sca*, which have been linked to ommatidial rotation do not affect the *mam*-RNAi mediated phenotype. For all genotypes, *n*>300 from at least 3 independent eyes.

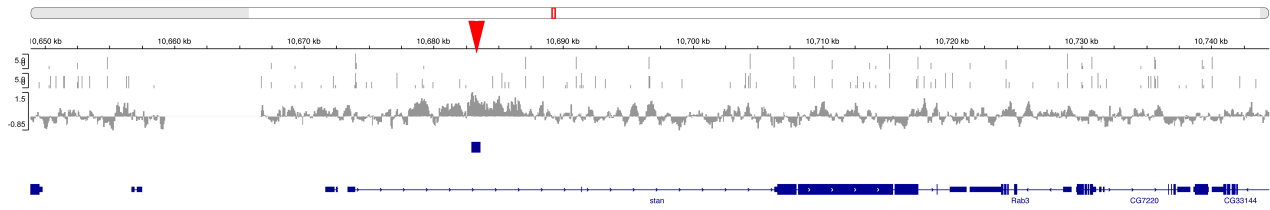


**Supplementary Figure S4. Heterozygous mutations in *aos* and *pk* that interact with the Notch-signaling genotypes do not show dominant effects in an otherwise wild-type background.**

(a-h) Adult eye sections with ommatidial orientation schematics and ommatidial orientation angle histograms of eyes from the following genetic backgrounds: (a-b) *aos*<sup>Δ7/+</sup>, (c-d) *DF(3L)BSC562/+*, (e-f) *pk*<sup>*pk-sple*13/+</sup>, and (g-h) *pk*<sup>*pk-sple*6/+</sup>. Note that all eyes have wild-type appearance in these heterozygous backgrounds.



**Supplementary Figure S5. Notch signaling does not affect Elav levels in R4.** (a) Third larval instar eye imaginal discs mosaic for  $m\delta 0.5 > mam^{RNAi}$  (BL28046; marked by absence of GFP/green) stained for Fmi (magenta) and Elav (gray). Note that the level of Elav is indistinguishable in the wild-type (GFP, green) vs the mutant clonal area (black). See Figure 4g in main text for quantification and comparison to *aos* expression. Scale bar is 10  $\mu\text{m}$ . (b) Quantification of relative intensity of Elav in R4 vs. R3 in wild type, note that the ratio is basically 1, meaning that there is no differential expression between the two cells.



**Supplementary Figure S6. Occupation by Su(H) in genomic sequences of the *fmi/stan* locus.**

ChIP enrichment for Su(H)-occupancy at the *stan/fmi* locus in CNS samples ( $\alpha$ -Su(H) enrichment relative to input, scale  $\log_2$ )<sup>60</sup>. Blue bar indicates potential region of significant enrichment. Gray bars indicate the positions of Su(H)-binding motifs; bar height represents the motif-score (scale 0-5); upper graph indicates motif conservation across 12 *Drosophila* species. Gene regions are depicted in dark blue. Although some enrichment is detected (small blue bar above large intron in gene schematic), there are no conserved binding site in that region (red arrowhead), suggesting that it is not meaningful. These data are consistent with the notion that Notch-Su(H) signaling is not a transcriptional regulator of *fmi/stan*. Furthermore, there was no ChIP enrichment detected associated with genomic regions of the *pk* locus.