Natural sesquiterpenoids

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This review covers the isolation, structural determination, synthesis and chemical and microbiological transformations of natural sesquiterpenoids. 421 references are cited.

- 1 Introduction
- 2 Farnesane
- 3 Monocyclofarnesane
- 4 Bicyclofarnesane
- 5 Bisabolane, sesquicarane, heliannane group, majapolane and parvifolane
- 6 Sesquicamphane, cyclosesquicamphane, campherenane, fumagillane and petasitane
- 7 Trichothecane, cyclotrichothecane, cuparane, laurane, cyclolaurane and herbertane
- 8 Chamigrane, perforane, perforetane and widdrane
- 9 Carotane, acorane, cedrane, zizaane and anislactone group
- 10 Cadinane, cubebane, oplopane and helminthosporane
- 11 Himachalane, longifolane and longipinane

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- 12 Caryophyllane, modhephane, isocomane, silphinane, presilphiperfolane, silphiperfolane, pethybrane, botryane and quadrane
- 13 Humulane, hirsutane, lactarane, capnellane, protoilludane, illudane, illudalane, marasmane, pentalenane, tremulane and africanane
- 14 Germacrane
- 15 Elemane
- 16 Eudesmane, axane, cycloaxane, iphionane, isoiphionane and lindenane
- 17 Vetisperane and spiroaxane
- 18 Eremophilane and bakkane
- 19 Guaiane, pseudoguaiane, xanthane and patchoulane
- 20 Aromadendrane, bicyclogermacrane, aristolane, sinulariolane and valerenane
- 21 Cyclomyltaylane
- 22 Salvionane
- 23 Miscellaneous sesquiterpenoids
- 24 References

1 Introduction

Sesquiterpenes have been used as chemosystematic markers in the subtribe Hypochaeridinae (Lactuceae tribe) of the Asteraceae family.¹ The chemical constituents in *Achillea*² and *Inula*³ species have been reviewed. These genera are very rich in sesquiterpenes. Infrared spectroscopy has been utilised as a tool for chemotaxonomic studies within the *Achillea millefolium* group.⁴ The chemical and biogenetic relationships among *Aristolochia* species from southeastern Brazil have been studied.⁵ Twenty four sesquiterpene lactones with different skeleta have been investigated as IL-8 expression inhibitors in HeLa cells.⁶ It has been shown that two pockets in the active site of maize sesquiterpene synthase (TPS4) are responsible for 14 different sesquiterpenes produced by this enzyme.⁷

2 Farnesane

The biological activity, source organism and country of origin of the linear furano- and pyrrolo-terpenoids have been reviewed.⁸ Amaranthanolidols A–D 1–4 and amarantholidosides I–VII 5–11 are new nerolidol derivatives, which have been obtained from the weed *Amaranthus retroflexus*.^{9,10} Five new sesquiterpene lactones 12–16, and the known antheindurolides A and B, have been found in an extract from the aerial part of *Anthemis arvensis*.¹¹ In this



work the structure originally proposed of antheindurolide B has been revised to 17. Four novel sesquiterpene peroxides 18–21 have been obtained from a Formosan soft coral of the *Sinularia* genus.¹²



The biosynthesis of *trans-(S)*-nerolidol in fruits of *Fragaria* x *ananassa* (strawberry) has been studied using deuterated compounds. This work showed that this sesquiterpene is exclusively synthesised *via* the cytosolic mevalonic acid pathway.¹³ It has been shown that an *Arabidopsis thaliana* methyl transferase is capable of methylating farnesoic acid.¹⁴ 12-Fluoro-farnesylphosphonophosphate has been synthesised as a potential inhibitor of sesquiterpene cyclase.¹⁵ In an interesting review, C. D. Poulter has considered farnesyl diphosphate synthase to be "a paradigm for understanding structure and function relationships in *E*-polyprenyl diphosphate synthases".¹⁶

The first total synthesis of 6-oxodendrolasinolide has been accomplished.¹⁷ Caulerpenyne, taxifolial A, dihydrorhipocephalin and furocaulerpin have been synthesised for the first time in racemic- and enantio-enriched forms. The *in vitro* inhibition of the polymerisation of microtubules by these sesquiterpenes has also been investigated.¹⁸ 2-Methyl-1,4-benzoquinone has been used as



9 $R_1 = H R_2 = Glc$





starting material in the synthesis of two triprenylated toluquinones and two toluhydroquinone derivatives, which had been isolated from a marine-derived *Penicillium* fungus.¹⁹ A study of the superacid cyclisation of certain aliphatic sesquiterpene derivatives in ionic liquids has been carried out.²⁰ Several sesquiterpenoids, isolated from *Ferula fukanensis*, have been shown to inhibit nitric oxide production by a murine macrophage-like cell line.²¹

3 Monocyclofarnesane

An α -ionone derivative **22** has been found in the aerial parts of *Salsola tetrandra*.²² Bridelionosides A–F **23–28** and debilosides A–C **29–31** are megastigmane glucosides, which have been isolated from *Bridelia glauca*²³ and *Equisetum debile*,²⁴ respectively. Other compounds of this type, macarangiosides A–D **32–35**, obtained from the leaves of *Macaranga tanarius*, showed potent DPPH radical-scavenging activity.²⁵ A study of the components of *Youngia japonica*²⁶ led to the isolation of the glucoside **36**. An inamoside derivative, named cuneatoside, has been found in an extract from the leaves and branches of *Erythroxylum cuneatum*,²⁷ whilst vignoside **37**, a glucoside that inhibits the growth of human stomach cancer, has been obtained from *Vigna angularis* (adzuki bean).²⁸ Several glycosides of blumenol C, 13-hydroxy-blumenol C and 13-*nor*-5-carboxy-blumenol C accumulate when





roots of *Ornithogalum umbellatum* are colonised by an arbuscular mycorrhizal fungus.²⁹ A carotenoid dioxygenase from melon, that cleaves carotenoids generating ionone derivatives, has been characterised and cloned.³⁰

Two new megastigmane derivatives, **38** and **39**, have been obtained from the red alga *Gymnogongrus flabelliformis*.³¹ Another red alga *Laurencia luzonensis* contains five novel sesquiterpenes **40–44**, two of which **40** and **41** possess a new rearranged snyderane skeleton.³²

Racemic α -ionone has been used as starting material for syntheses of the enantiomeric forms of α - and γ -damascone³³ and of 3,4-didehydroionone stereoisomers.³⁴ Enantioselective syntheses of (+)-ricciocarpin A³⁵ and striatenic acid³⁶ have been achieved. A total synthesis of the *trinor*-cyclonerolidane sesquiterpene (–)-chokol A has been reported.³⁷



A 9-*cis*-epoxycarotenoid dioxygenase inhibitor has been used in the elucidation of the mechanisms of action of abscisic acid.³⁸ A concise and enantioselective synthesis of this plant hormone, and of a new analogue, has been reported.³⁹ (+)-AHI4 is a new inhibitor of ABA 8'-hydrolase, which has been synthesised with an α -axial hydroxyl group instead of the geminal methyls at C-6' characteristic of (+)-AHI1, (1'S,2'S)–(+)-6-*nor*-2'3'-dihydro-4'deoxy-ABA.⁴⁰ Bicyclic analogues of abscisic acid have been shown to have enhanced biological activity in plants.⁴¹

4 Bicyclofarnesane

Nebularic acids A **45**, nebularic acid B **46**, nebularilactone A **47** and nebularilactone B **48** are four new drimane sesquiterpenes, which have been found in the fungus *Lepista nebularis*.⁴² The fruiting bodies of *Stereum ostrea* contain the new sesquiterpene methoxylaricinolic acid.⁴³



The novel *dinor*-sesquiterpene japonicumin D **49** has been obtained from *Lycopodium japonicum*.⁴⁴ A phytochemical analysis of the tree *Pleodendron costaricense*⁴⁵ led to the isolation of the drimane derivative parritadial **50**. The fungicidal sesquiterpene

paxidal **51** has been found in a foliage extract from a New Zealand shrub, *Pseudowintera axillaris*, using a bioactivity-directed separation. This sesquiterpene possesses fungicidal activity against the plant pathogen *Phytophthora infestans*.⁴⁶ The structures of cinnamolid-3β-ol hemihydrate and 3β-hydroxycinnamolide acetate, two components of *Warburgia ugandensis*, have been determined by X-ray analysis.⁴⁷ Other compounds from this plant have been evaluated as inhibitors of 12(*S*)-HETE and of leukotriene metabolism.⁴⁸ The dimeric and trimeric drimane sesquiterpenes cinnafragrins A–C **52–54** have been found in an extract from the medicinal plant *Cinnamosma fragrans*.⁴⁹ In this work, the structure of capsicodendrin has been revised to a mixture of the C-12′ epimers **55**.



Sespendole **56** is a novel bioactive indolosesquiterpene, which has been isolated from the culture broth of the fungus *Pseudobotrytis terrestris* strain FKA-25.⁵⁰⁻⁵² The structures of dasyscyphins



A-C and niveulone have been determined as 57-59 and 60, respectively. These meroterpenoids have been obtained from the ascomycete Dasyscyphus niveus.53-55 Other compounds of this type with cytotoxic properties, tropolactones A-D 61-64, have been isolated from a marine derived fungus Aspergillus sp., which was obtained from an unidentified sponge, collected at Manele Bay in Hawaii.56 Chromatography of an extract from an unidentified species of the Dysidea genus afforded the bioactive sesquiterpene 65, which has been named avinosol.57 Three new species of this genus have been characterised by their sesquiterpenoid content. Thus, sesquiterpene-hydroquinones are the main components of Dysidea reformensis and Dysidea cachui, while the furanosesquiterpene dendrolansin was obtained from Dysidea uriae.58 The reactivity and biological activity of the marine sesquiterpene hydroquinone avarol, and related sesquiterpenes, have been reviewed.59

The biotransformation of (-)-Ambrox® by cell suspension cultures of kiwifruit, Actinidia deliciosa, has been investigated.⁶⁰ Stereoselective syntheses of 9-epi-Ambrox[®],⁶¹ and Superambrox[®],⁶² and racemic syntheses of hyphodermin B⁶³ and pallescensin A,64 have been reported. Drimane dienes with functional groups at C-6 have been synthesised by photodegradation of larixol derivatives.⁶⁵ (\pm) -Wiedendiol B and a siphonodictyal B derivative have been prepared using a new approach to the synthesis of sesquiterpene arenes.⁶⁶ The diterpene (+)-manool has been used as starting material for the preparation of the sesquiterpene hydroquinone (+)-ent-chromazonarol.⁶⁷ Another diterpene, sclareol oxide, has been employed in the synthesis of ent-thallusin,68 whilst a bioinspired strategy has been used in the synthesis of hongoquercin A and rhododaurichromanic acid A.⁶⁹ A highly diastereoselective formal synthesis of puupehedione and 8-epi-puupehedione has been described.⁷⁰ A short sequence of reactions has been used in the preparation of bicyclic sesquiterpene units linked to aromatic structures, which has



permitted formal syntheses of zonarol, zonarone, puupehedione and umbrosone.⁷¹ A rapid method for the construction of the A, B and C rings of terreulactone A has been reported.⁷²

The drimane sesquiterpenes fetidone A **66** and fetidone B **67** have been found in an extract from assafetida. This gum resin also contains known sesquiterpene coumarin ethers, one of which, (8-acetoxy)-5-hydroxyumbelliprenin, was showed to be a potent NF- κ B inhibitor,⁷³ while conferone **68**, isolated from *Ferula schtschurowskiana*, enhanced the cytotoxicity of vinblastine in MDCK-MDR1 cells.⁷⁴ The sesquiterpene coumarin saradaferin **69** has been obtained from *Ferula asafoetida*.⁷⁵ Other new compounds of this type, isofeterin **70**, lehmannolol **71** and sinkianone **72**, have also found in extracts from the roots of *Ferula sinkiangensis* and *Ferula teterrima*.⁷⁶



5 Bisabolane, sesquicarane, heliannane group, majapolane and parvifolane

The male-produced sex pheromones from the Brazilian rice stalk stink bug, Tibraca limbativentris, has been characterised as isomers of 1'S-zingiberenol.⁷⁷ Eight new bisabolane sesquiterpenes 73-80 have been isolated from the roots of Ligularia cymbulifera.⁷⁸ Another compound of this type 81 has been found in an extract from Achillea clavennae.79 The aerial parts of Lippia dulcis80 contain six novel bisabolane sesquiterpenes, which have been named peroxylippidulcines A-C 82-84, peroxyepilippidulcine B 85, epilippidulcine B 86 and epilippidulcine C 87. The bisabolane derivatives boivinianin A 88 and boivinianin B 89 have been identified as components of the stem bark of Cipadessa boiviniana (Meliaceae).⁸¹ Another three new sesquiterpenes 90-92 have been isolated from the roots of Leontopodium longifolium.82,83 Baccharisketone 93 is another new bisabolane derivative, which has been obtained from the leaves of Baccharis dracunculifolia.⁸⁴ The active principle of an extract from the roots of Ostericum koreanum⁸⁵ which affects adults of the Dermatophagoides genus, has been identified as the known sesquiterpene bisabolangelone **94**.

,OH OH ÓΩ⊢ όон 81 82 R₄ OН **83** $R_1 = \beta - H R_2 = \beta - OOH$ $R_1 = \beta - H$ $R_2 = \alpha - OOH$ 84 $R_1 = \alpha - H$ $R_2 = \beta - OOH$ $R_1 = \alpha - H$ $R_2 = \beta - OH$ 86 $R_1 = \alpha - H$ $R_2 = \alpha - O H$ 87 88 89 R = α -H and β -H A species of sponge of the Myrmekioderma genus,⁸⁶ collected

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79 $R_1 = H$ $R_2 = Ang$ $R_3 = OAng$

 $R_1 = Ang R_2 = H R_3 = CI$

AngC

 R_2O

80

OAng

OAng

OAnd

OAng

юн

Ο̈́H 77 $R_1 = \beta$ -OAng $R_2 = \beta$ -OH,H

78 $R_1 = \alpha$ -OAng $R_2 = O$

73 $R_1 = \alpha$ -OH $R_2 = \alpha$ -OAng

74 $R_1 = R_2 = \beta$ -OAng

75 $R_1 = \beta$ -OAng $R_2 = H$

A species of sponge of the *hymerioarma* genus, concerct at the Vanuatu coast, contains the bisabolane sesquiterpenes **95– 97**. Three other novel halogenated β -bisabolene derivatives **98–100** have been isolated from the red alga *Laurencia scoparia*,⁸⁷ whilst the aldingenins B–D **101–103** have been obtained from another species of this genus *Laurencia aldingensis*.⁸⁸ The sesquiterpenoids **104–107** have been found in an extract from the sea hare *Aplysia dactylomela*.⁸⁹ β -Bisabonol has been synthesised using a ringclosing olefin metathesis reaction in the key step.⁹⁰



Heliespirone B **108** and heliespirone C **109** are two sesquiterpenes with two novel spiro heterocyclic skeleta, which have been isolated from *Helianthus annuus*.⁹¹ The structures of heliannuols G and H, also obtained from this species, have been revised to **110** and **111**, respectively, as a consequence of enantiocontrolled total syntheses.⁹² This plant also contains heliannuol C, whose synthesis has also been accomplished.⁹³ A total synthesis of sesquicarene **112** has been described.⁹⁴ This sesquiterpene had been isolated from *Schisandra chinensis*.⁹⁵ The absolute configurations of majapolene



B **113** and acetylmajapolene B **114** have been determined by vibrational circular dichroism. These sesquiterpenes have been found in an extract from a *Laurencia* sp., collected in Malaysia.⁹⁶ The first enantioselective synthesis of (–)-parvifoline and (–)-curcuquinone has been reported.⁹⁷



6 Sesquicamphane, cyclosesquicamphane, campherenane, fumagillane and petasitane

Syntheses of two (*E*)-*endo*-bergamoten-12-oic acid derivatives have been reported.⁹⁸ Three new campherenane, **115**, **116** and **119**, and three new santalane sesquiterpenes **121–123**, with bioactive properties, have been isolated from *Santalum album* of Indian origin.⁹⁹ In this work, the absolute structure of **117**, **118** and **120** has also been established. These compounds had been obtained from commercial sandalwood.¹⁰⁰ In this plant, *Santalum album*, a quantitative chemotaxonomic approach has been used to study the biosynthetic relationships between the sesquiterpenes isolated from the heartwood.¹⁰¹ The effects of inhalation of East Indian



sandalwood essential oils and its main compound, α-santalol, on human physiological parameters, have been measured in healthy volunteers.¹⁰² Concise, enantio- and diastereo-selective total syntheses of fumagillol, RK-805, FR65814, ovalicin and 8demethylovalicin have been accomplished.¹⁰³ The rare sesquiterpene hydrocarbons petasitene **124** and albene **125** have been identified as components of a root oil obtained from *Artemisia vulgaris* of Serbian origin.¹⁰⁴



7 Trichothecane, cyclotrichothecane, cuparane, laurane, cyclolaurane and herbertane

Two highly cytotoxic compounds, 16-hydroxyverrucarin A **126** and verrucarin X **127**, have been isolated from fermentations of the fungus *Myrothecium roridum*.¹⁰⁵ Another three macrocyclic trichothecenes, 12'-hydroxyroridin E, roridin Q and 2',3'-deoxyroritoxin D, have been obtained from two strains of the marine-derived fungus *Myrothecium roridum* TUF 98F42, whilst roridin R **128**, and the known roridin A, roridin H and isororidin E, have been found in *Myrothecium* sp TUF 02F6.¹⁰⁶ Other strains of *Myrothecium roridum*, IFB-E009 and IFB-E012, which were isolated as endophytic fungi from the Chinese medicinal plants



Trachelospermum jasminoides and *Artemisia annua*, respectively, contain the new cyclotrichothecane macrolides myrothecines A– C **129–131**. The absolute configuration of mytoxin B was also established in this study.¹⁰⁷ Another cyclotrichothecane derivative miophytocen C **132**, together with 3-hydroxyroridin E and 13′-acetyltrichoverrin B, were isolated from a saltwater culture of *Myrothecium verrucaria*, separated from a *Spongia* sp., collected in Hawaii.¹⁰⁸



GC–MS techniques have been used in the analysis of trichothecene mycotoxins in commercial corn harvested in Brazil.¹⁰⁹ The synthesis of four mycotoxins of this type labelled with carbon-13 and their application as internal standards in stable isotope dilution assays for their determination in foods has been described.¹¹⁰ The loss of toxicity of deoxynivalenol in extruded cereal-based products has been confirmed using different types of analysis.¹¹¹

Two new pathways for the enzyme-catalysed formation of trichodiene **133** have been proposed, one of which, a proton-transfer pathway, appears to be much more energetically favourable than the hydride transfer pathway usually proposed.¹¹² The structures of two new bazzanene derivatives have been determined as **134** and **135**. These sesquiterpenes have been isolated from a New Zealand liverwort, *Frullania falciloba*.¹¹³



A new sesquiterpene, 3,7-dihydroxydihydrolaurene 136, has been obtained from the red alga Laurencia obtusa, whilst a dimeric sesquiterpene of the cyclolaurane type 137 was isolated from Laurencia microcladia. These algae had been collected from the coastal rocks of Serifos in the Aegean Sea.¹¹⁴ Another species of this genus, Laurencia okamurai,¹¹⁵ contains a new laurane derivative, 3β-hydroxyaplysin 138, and two novel rearranged sesquiterpenes, laurokamurene A 139 and laurokamurene B 140. The structure of the laurokamurene A was identical to a metabolite which had been obtained from Laurencia microcladia (see our last review¹¹⁶). The structure assigned to HM-3, a sesquiterpene which had been isolated from the phytopathogenic fungus Helicobasidium mompa, has been found to be incorrect by total synthesis of the proposed structure.117 In this work the first synthesis of HM-3 141 and HM-4 142 has also been carried out. An enantioselective synthesis of (–)- α -herbertenol has been achieved.¹¹⁸ (±)- β -Cuparenone has been synthesised in a short and efficient procedure.¹¹⁹ A. Srikrishna et al. have carried out racemic total syntheses of herbertenediol,¹²⁰ grimaldone, epigrimaldone, a-cuparenones¹²¹ and lagopodin A,¹²² and a racemic formal synthesis of cuparene and herbertene.¹²³



8 Chamigrane, perforane, perforetane and widdrane

The thalloid liverwort, *Reboulia hemisphaerica*, contains two new *ent*- β -chamigrane derivatives **143** and **144**. The biotransformation of the latter by the fungus *Aspergillus niger* has also been described in this work.¹²⁴ A gas chromatographic method has been developed to quantify the chamigrane sesquiterpene elatol, at the surface and within the thallus, of 70 specimens of the red alga *Laurencia obtusa*.¹²⁵ This species also contains the new sesquiterpenes perforenol B **145** and the perforetane derivative



146.¹¹⁴ Total syntheses of racemic α -chamigrene, β -chamigrene and laurencenone C have been described.¹²⁶



The structure elucidation and biomimetic synthesis of liphagal **147** has been carried out. This selective inhibitor of PI3 kinase has been obtained from the sponge *Aka coralliphaga*.¹²⁷ The antifungal activity of widdrol and its microbiological transformation by the fungi *Colletotrichum gloeosporioides* and *Botrytis cinerea* have been investigated.¹²⁸

9 Carotane, acorane, cedrane, zizaane and anislactone group

A formal synthesis of (\pm) - α -cedrane and (\pm) - β -cedrane has been reported.¹²⁹ After 25 years of the alquene-arene *meta*photocycloaddition reaction, which was developed in a synthesis of α -cedrene, a review has appeared on the application of this reaction to the synthesis of other natural products.¹³⁰ D. E. Cane *et al.* have described the molecular cloning and characterisation of a new sesquiterpene synthase from *Streptomyces coelicolor*. This enzyme catalyses the cyclisation of farnesyl diphosphate to a novel sesquiterpene, *epi*-isozizaene **148**, which has not been isolated as a natural product, but which had been chemically prepared by acid treatment of (+)-zizaene.¹³¹



Three new secoprezizaane sesquiterpene lactones **149–151** have been obtained from *Illicium micranthum*.¹³² Racemic syntheses of jiadifenin,¹³³ merrilactone A¹³⁴ and 11-*O*-debenzoyltashironin¹³⁵ have been accomplished.



10 Cadinane, cubebane, oplopane and helminthosporane

The new sesquiterpene **152** has been isolated from an aqueous extract of *Malva silvestris*.¹³⁶ A phytochemical study of *Commiphora myrrha*¹³⁷ led to the isolation of six new cadinane derivatives, which have been named myrracadinols A–C **153–155** and myrracalamenes A–C **156–158**. The sesquiterpene **159** and the norsesquiterpene eupatorone **160** have been obtained from the flowers of *Eupatorium adenoforum*.¹³⁸ Other authors have isolated another novel sesquiterpenoid **161** from the leaves of this species.¹³⁹ Two new cadinane derivatives **162** and **163** have been found in an extract from the bark of *Jatropha neopauciflora*,¹⁴⁰ while the aerial



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parts of Pseudoelephantopus spicatus¹⁴¹ has been shown to contain a new lactone, named spicatocanolide A 164. Bioassay-guided studies of an ethanol extract from Swartzia polyphylla afforded T-cadinol as the metabolite responsible for the larvicidal and antimycobacterial activity of this Peruvian plant.142 Pubinernoid C 165 is a new cadinane sesquiterpene, which has been obtained from Schisandra pubescens.143 Three novel dehydroabietane diterpenes sugikorojins G-I 166-168, which incorporate a cadinane sesquiterpene in their structure, have been isolated from Cryptomeria *japonica*,^{144,145} whilst a diterpene-sesquiterpene ether **169** has been obtained from Calocedrus macrolepis, and named calocedimer B.¹⁴⁶ Chromatography of an extract from *Phomopsis cassiae*, an endophytic fungus associated with Cassia spectabilis, afforded five new cadinane derivatives 170–174.¹⁴⁷ A patent describing the cytotoxic activity of the dimers parviflorene H and parviflorene I has appeared.¹⁴⁸ Another cadinane sesquiterpene, amentotaxone 175, has been isolated from Amentotaxus formosana.¹⁴⁹ The cloning and functional characterisation of a cis-muuroladiene synthase from Mentha piperita has been described.150





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The structures of 4α , 5α , 10β -trihydroxycadinane **176** and its C-4 epimer **177** had been assigned to two sesquiterpenes, which had been isolated from *Jasonia candicans*¹⁵¹ and *Taiwania cryp-tomerioides*,¹⁵² respectively. Total syntheses of these compounds have now been carried out, indicating that both sesquiterpenes are identical, and possess the second of these structures.¹⁵³ The first total synthesis of xenitorin B and xenitorin C has been achieved, allowing the determination of their absolute configuration.¹⁵⁴ Syntheses of 7,8-dihydroxycalamenene and mansonone C have been achieved.¹⁵⁵ An alternative procedure for the synthesis of mansonone F and of a biflorin precursor has been described.¹⁵⁶ The structure of isocalamendiol has been applied to the determination of the absolute configuration of (–)-3-hydroxy- α -calacorene **178**.¹⁵⁸



6-Methoxygossypol and 6,6'-dimethoxygossypol have been obtained from the seed and root bark from a cotton variety growing on St. Vincent island.¹⁵⁹ The ratios of (+)-gossypol and (–)gossypol in leaves, stems and roots from selected accessions of *Gossypium hirsutum* have been determined.¹⁶⁰ The effects of these compounds, and of their racemic forms, on the survival and development of *Helicoverpa zea* larvae have been evaluated.¹⁶¹ The peroxidative coupling of hemigossypol to (+)-gossypol and (–)-gossypol in extracts of cottonseed has been investigated.¹⁶² Aquatidial **179** is a new bis-norsesquiterpene, which has been isolated from the roots of *Pachira aquatica*. This compound is probably formed by the dimerisation of a cadinane derivative, isohemygossypolone, which was also obtained from this plant.¹⁶³



The biosynthesis of artemisinin and its regulation in Artemisia annua have been reviewed.¹⁶⁴ The mechanism and stereochemistry of the cyclisation of farnesyl diphosphate into amorpha-4,11-diene by amorpha-4,11-diene synthase, a key enzyme in artemisinin biosynthesis, have been studied.^{165,166} A cDNA clone, encoding a cytochrome P450, catalyses the oxidation of amorpha-4,11diene, artemisinic alcohol and artemisinic aldehyde, which are intermediates in the biosynthesis of artemisinin.¹⁶⁷ The effects of overexpression of the endogenous farnesyl diphosphate synthase on the artemisinin content in Artemisia annua have been investigated.¹⁶⁸ Density functional theory calculations have been applied to a theoretical study of the reductive decomposition of this antimalarial sesquiterpene.¹⁶⁹ The production of artemisinic acid in an engineered strain of the yeast Saccharomyces cereviasae has been accomplished by J. D. Keasling et al. These authors first manipulated the FPP biosynthetic pathway to increase the production of this intermediate by the yeast, then introduced a gene, which transforms FPP to amorpha-4,11-diene, and finally introduced a cytochrome P450 that converts this diene into artemisinic acid.¹⁷⁰ The fungus Streptomyces griseus has been used in the biotransformation of a novel ketone-derivative of artemisinin.171

A paper describing a comparative assessment of the established and emerging technologies for the extraction of artemisinin has appeared.¹⁷² The accumulation of this antimalarial sesquiterpene during the development and senescence of the leaves of *Artemisia annua* has been studied.¹⁷³ A simple route to access new 10fluoro-artemisinin derivatives has been reported.¹⁷⁴ The reaction





mechanism of 12 antimalarial artemisinin derivatives with two competitive pathways has been studied by means of quantum chemical calculations.¹⁷⁵ A new highly active antimalarial drug of the artemisinin class, named artemisone **180**, has been described,¹⁷⁶ whilst new orally active derivatives of artemisinin, with high efficacy against multidrug-resistant malaria in mice, have been developed.¹⁷⁷ The anti-angiogenic activity of several deoxoartemisinin derivatives has been reported,¹⁷⁸ whilst the antiviral effect of artemisinin against the bovine viral diarrhoea virus has been investigated.¹⁷⁹ Artemisinin and extracts of the aerial parts of *Artemisia annua* have been evaluated as anti-insect products.¹⁸⁰

A stereoselective synthesis of (-)-cubebol has been accomplished.¹⁸¹ Other authors have reported the synthesis of this sesquiterpene and of the hydrocarbon (-)- α -cubebene.⁹⁴ New oplopane sesquiterpenes **181–186** have been isolated from *Ligularia narynensis*.^{182,183} Another compound of this type **187** has been found in an extract from the brown alga *Dictyopteris divaricata*.¹⁸⁴ New helminthosporal analogues with plant growth regulatory properties have been synthesised.¹⁸⁵

11 Himachalane, longifolane and longipinane

Four known himachalane sesquiterpenes have been identified in volatiles collected from the eggplant flea beetle, *Epitrix fuscula*.¹⁸⁶ An enantioselective total synthesis of (+)-β-himachalene has been achieved.¹⁸⁷ The conformations of longifolene¹⁸⁸ and of a longipinene derivative¹⁸⁹ have been studied by NMR and molecular mechanics calculations. The microbiological transformation of (+)-cycloisolongifol-5β-ol by the fungus *Cunninghamella elegans* has been investigated.¹⁹⁰

12 Caryophyllane, modhephane, isocomane, silphinane, presilphiperfolane, silphiperfolane, pethybrane, botryane and quadrane

The caryophyllane derivative **188** has been found in extracts from the Formosan soft coral *Sinularia gibberosa*.¹⁹¹ (–)- β -Caryophyllene has been identified as a gender-specific sesquiterpene emitted by the multicoloured Asian lady beetle, *Harmonia axyridis*.¹⁹² Known caryophyllane sesquiterpenes have been isolated from *Aframomun arundinaceum*,¹⁹³ *Pulicaria prostrata*¹⁹⁴ and *Zingiber nimmonii*.¹⁹⁵ The biotransformation of (–)-caryophyllene oxide by cell suspension cultures of *Catharanthus roseus*¹⁹⁶ and by several species of fungi¹⁹⁷ have been described. The transannular cyclisation of epoxycaryophyllenes catalysed by Ti^{III} has been investigated.¹⁹⁸ The absolute configuration of birkenal has been determined by chemical correlation with a caryophyllene derivative.¹⁹⁹

Three new sesquiterpenes 6-hydroxypunctaporonin A **189**, 6hydroxypunctaporonin B **190** and 6-hydroxypunctaporonin E **191** have been obtained from cultures of the fungicolous fungus *Pestalotiopsis disseminata*.²⁰⁰



The acid-catalysed rearrangement of (-)-modhephene **192** and (-)-isocomene **193** to a (-)-triquinane **194** has been studied.²⁰¹ The silphinene derivative **195** has been found in cultures of an isolate of the fungus *Penicillium griseofulvum*. This metabolite is a probable precursor of other sesquiterpenes, which were also obtained from this species and named penifulvins A-E **196–200**,



Nat. Prod. Rep., 2007, 24, 1350-1381 | 1361

and which possess a new carbon skeleton.^{202,203} The uncommon sesquiterpene hydrocarbons silphiperfol-5-ene **201**, modhephene **192**, pethybrene **202**, presilphiperfol-7-ene **203**, silphin-1-ene **204** and isocomene have been identified as components of a root oil obtained from *Artemisia vulgaris*.¹⁰⁴



The role of botrydienediol **205** in the biodegradation of the sesquiterpenoid phytotoxin botrydial **206** by *Botrytis cinerea* has been investigated.²⁰⁴ The absolute configurations of quadrone **207**, suberosenone, suberosanone and suberosenol A acetate have been determined *via* density functional theory calculations of optical rotation, electronic circular dichroism and vibrational circular dichroism.²⁰⁵ A patent describing the use of a sesquiterpene ketone, isolated from *Subergonia suberosa*, as an anticancer drug has appeared.²⁰⁶



13 Humulane, hirsutane, lactarane, capnellane, protoilludane, illudane, illudalane, marasmane, pentalenane, tremulane and africanane

The humulane sesquiterpenes mitissimols A–C **208–210**, and a mixture of mitissimol A oleate and mitissimol B linoleate, have been isolated from the fruiting bodies of *Lactarius mitissimus*.²⁰⁷ Another compound of this type, 10ξ-acetoxyhumula-3,7-diene,

has been obtained from the roots of *Casearia multinervosa*.²⁰⁸ The Sharpless asymmetric epoxidation of (+)-zerumbol has been studied.²⁰⁹ The structures of hirsutenols D–F have determined as **211–213**. These sesquiterpenes have been found in an extract of the culture broth of *Stereum hirsutum*.²¹⁰ Chemoenzymatic syntheses of (+)-hirsutic acid and (–)-complicatic acid have been achieved.²¹¹ Rapraesentins D–F **214–216** and 1,2-dehydrolactarolide A **217** are new plant growth regulators, which have been isolated from the fungi *Lactarius repraesentaneus*²¹² and *Lactarius vellereus*,²¹³ respectively. A group of lactarene derivatives have been tested as antiviral, cytotoxic, antiproliferative and immunotropic agents.²¹⁴



The basidiomycete *Ripartites metrodii* contains three new sesquiterpenes, riparols A–C **218–220** with an illudane, illudalane and protoilludane skeleton, respectively.²¹⁵ The russujaponols A–F **221–226** are six novel illudane sesquiterpenes, which have been obtained from the fruiting body of *Russula japonica*.²¹⁶ Chromatography of an extract from the cultured mycelia of the fungus *Echinodontium japonicum*²¹⁷ afforded two illudalane sesquiterpenes, which have been named echinolactone C **227** and echinolactone D **228**. An approach to the synthesis of (–)-coprinolone, a protoilludane derivative, has been reported.²¹⁸ The synthesis and biological evaluation of alcyopterosin A **229**



and other illudalane derivatives, as anticancer agents, has been described. $^{\rm 219}$

The synthesis of the tricyclic core of the 5 α -capnellenols has been reported.²²⁰ A total synthesis of the sesquiterpene dichomitol has revealed that the structure assigned to this compound was erroneous.²²¹ Two new marasmane sesquiterpenes, **230** and **231**, have been isolated from *Russula foetens*.²²²

The existence in *Streptomyces avermitilis* of a gene cluster for the biosynthesis of pentalenolactone **232** has been reported.²²³ The molecular cloning and assignment of the biochemical functions of PtlH and PtlI, a non-heme iron dioxygenase²²⁴ and a cytochrome P450,²²⁵ respectively, have been described. The first, in this fungus, converts 1-deoxypentalenic acid to 11β-hydroxy-1deoxypentalenic acid, whilst the second transforms pentalenene to pentalen-13-al. Quantum chemical computation studies have been carried out with the aim of ascertaining possible polycyclisation pathways of the farnesyl cation into pentalenene.²²⁶ An enantiospecific approach towards the synthesis of pentalenolactone has been described.²²⁷



Enantioselective syntheses of tremulenediol A 233 and tremulenolide A 234 have been accomplished.²²⁸ The new africanane alcohol 235 has been isolated from the essential oil of *Lippia integrifolia*.²²⁹



14 Germacrane

The new sesquiterpene **236** has been found in an extract from the roots of *Thapsia nitida*.²³⁰ The known sesquiterpenes germacrane D and kunzeanol have been isolated from the essential oil of *Brickellia veronicaefolia*²³¹ and from the leaf surface of *Solanum tuberosum*,²³² respectively. Other known germacrane derivatives have been obtained from the marine octocoral *Muricea austera*.²³³ The cloning, expression, purification and characterisation of recombinant germacrane A and germacrene D synthases from *Artemisia annua*²³⁴ and *Zingiber officinale*,²³⁵ respectively, have been described. The pheromone periplanone C has been synthesised as its racemate.²³⁶

The new germacrane lactones which have been isolated from natural sources during 2006 are listed in Table 1. The structures **237–259** represent the new germacranolides, while the structures **260–262** have been assigned to the new heliangolides and **263–264** to the melampolides. No new *cis,cis*-germacranolides have been obtained this year.

The structure **257** given to vernolide C²⁴⁷ proved to be the same as that reported for vernchinilide A (see our last review¹¹⁶). The full paper on the isolation of the lactones eupanilonides A and D from *Eupatorium lindleyanum* has appeared.²⁵¹ Known germacranolides have been obtained from *Camchaya calcarea*²⁵² and *Ichthiothere terminalis*.²⁵³ It has been shown that the sesquiterpene lactone parthenolide induces apoptosis of human acute myelogenous leukaemia stem cells,²⁵⁴ and of B-chronic limphocytic

Table 1 Sources of germacrane lactones

Source	Germacranolides	Ref.
Achillea asplenifolia	237–242	237
Achillea collina	243	238
Dimerostemma brasilianum	244	239
Eupatorium chinense	245	240
Eupatorium kiirunense	246	241
Glechoma longituba	247	242
Helianthus annuus	248-251	243
Inula britannica	252	244
Mulgedium tataricum	253, 254	245
Pseudoelephantopus spicatus	255	141
Sarcandra glabra	256	246
Vernonia cinerea	257, 258, See text	247
Vernonia triflosculosa	259	248
Heliangolides		
Eupatorium kiirunense	260, 261	241
Eupatorium lindlevanum	262	249
Melampolides		
Siegesbeckia orientalis	263, 264	250
0	·	

-OAc ċн

236

AngO

240

HO

R₁O



237 R = Valⁱ 238 R = Mebu



•OTig





сно

246 Eupakirunsin I

















251 R = Et







256 R = Glc-(6'-1")-Api

255 Spicatolide C



Vernolides C and D

257 R = COPrⁱ(2'-OH,3'-CI)





leukaemia cells.255 The synthesis and antiviral activity of a series of sesquiterpene lactones, structurally analogous to parthenolide, have been reported.256 The lactone cnicin has been shown to be a potent and irreversible inhibitor of the antibacterial target enzyme MurA.257 The anionic and photochemical behaviour of the hallerin derivative 265 have been studied.²⁵⁸ Several 13-amino costunolide derivatives have been synthesised and evaluated as anticancer agents.259



15 Elemane

Zinagrandinolides A–C **266–268** are three new sesquiterpene lactones, which have been isolated from *Zinnia grandiflora*.²⁶⁰ Two sesquiterpene glycosides sarcaglaboside C **269** and sarcaglaboside D **270** have been found in an extract from *Sarcandra glabra*.²⁴⁶ A known furanoelemene sesquiterpene has been obtained from the marine octocoral *Muricea austera*.²³³ The reaction of elemol with acetic acid–perchloric acid has been studied.²⁶¹



16 Eudesmane, axane, cycloaxane, iphionane, isoiphionane and lindenane

The eudesmane sesquiterpenes, isolated from the Asteraceae family, have been reviewed.²⁶² The essential oils from the liverwort *Plagiochila bifaria*²⁶³ contain the new *ent*-eudesmenones **271–273**. Three eudesmane sesquiterpenes **274**, **276** and **278** have been obtained from another liverwort *Lepidozia fauriana*.²⁶⁴ In this work, the structures of another two metabolites have been revised to **275** and **277**. These compounds had been obtained

from *Bazzania tridens*²⁶⁵ and *Lepidozia vitrea*,²⁶⁶ respectively. The new eudesmane derivative **279** has been found in an extract from a marine *Streptomyces* species.²⁶⁷



The new hydroperoxide **280** has been isolated from the aerial parts of *Aster spathulifolius*.²⁶⁸ Two cytotoxic sesquiterpenes, **281** and **282**, with a eudesmane and an axane skeleton, respectively, have been found in *Jatropha neopauciflora*.²⁶⁹ This plant also contains two cycloaxane derivatives, **283** and **284**.¹⁴⁰ A phytochemical study of a variety of *Juniperus polycarpus*²⁷⁰ afforded two new eudesmane sesquiterpenes, **285** and **286**. Other compounds of this type, pterodontriol E **287** and pterodontriol F **288**, have been obtained from the aerial parts of *Laggera pterodonta*,²⁷¹ whilst plenoxide **289** has been isolated from *Schisandra plena*.²⁷² Another two new sesquiterpenes, **290** and **291**, have been obtained from





Sambucus williamsii,²⁷³ and Youngia japonica,²⁶ respectively. The isolation of the new metabolites **292** and **293** from *Atractylodes macrocephala*²⁷⁴ and **294** from *Chloranthus henryi*²⁷⁵ has been reported. The structure of the sesquiterpene eudesma-5,12-dien-13-oic acid has been determined by X-ray analysis. This compound has been obtained from *Laggera pterodonta*.²⁷⁶ The eudesmane glycosides **295** and **296** have been found in *Fissistigma pallens*²⁷⁷ and *Pteris multifida*,²⁷⁸ respectively, whilst an acetylated eudesmane glucoside **297** has been isolated from *Apodytes dimidiate*.²⁷⁹ The antibacterial, antioxidant and α -amylase inhibitory activity of 3-oxo-selina-4,11(13)-dien-12-oic acid have been evaluated. This sesquiterpene has been obtained from *Varthemia iphionoides*.^{280,281}



The molecular cloning, expression and mechanistic study on the germacradienol/geosmin synthase from *Streptomyces avermitilis* have been described.²⁸² Moreover, it has been shown that a germacradienol/germacrane D synthase from *Streptomyces coelicolor* converts farnesyl diphosphate into geosmin.²⁸³ The first total syntheses of (\pm)-corymbolone²⁸⁴ and of 6β-cinnamoyloxy-1 α -hydroxy-5,10-bis-*epi*-eudesm-4-en-3-one²⁸⁵ have been reported. The insect antifeedant activity of (+)-pterocarpol has been evaluated.²⁸⁶

Several dihydroagarofuran derivatives have been obtained from Austroplenckia populnea,²⁸⁷ Celastrus angulatus,²⁸⁸ Celastrus orbiculatus,²⁸⁹ Celastrus paniculatus,²⁹⁰ Euonymus nanoides,²⁹¹ Microtropis fokienensis,²⁹² Pleurostylia opposita,²⁹³ Reissantia buchananii²⁹⁴ and Tripterygium wilfordii.²⁹⁵ Insights into the molecular mechanism of action of this type of sesquiterpenes, as specific non-transported inhibitors of human P-glycoprotein, have been described.²⁹⁶

The new eudesmanolides which have been obtained from different species are listed in Table 2. Their structures have been shown to be **298–316**.





298 Atractylenolide V

299 Eupakirunsin H









Table 2 Sources of eudesmanolides

Source	Eudesmanolides	Ref.
Atractylodes macrocephala	298	274
Eupatorium kiirunense	299	241
Eupatorium lindleyanum	300	249
Inula britannica	301, 302	244
Lantuca indica	303	297
Sarcandra glabra	304, 305	246
Sonchus uliginosus	306, 307, See text	298
Thapsia nitida	308-316	230



Six new lactones have been obtained from *Sonchus uliginosus*.²⁹⁸ Two are given in Table 2, but the other four had previously been isolated from *Sonchus transcarpicus*. Their structures were reported in our last review.¹¹⁶ Biotransformations of the lactones α -santonin and 6β -santonin by the fungus *Abisidia coerulea* and by cell cultures of the plants *Asparagus officinalis* have been carried out.²⁹⁹ The structure and biological activity of several chloro derivatives of α -santonin have been reported.³⁰⁰ An enantioselective total synthesis of the antileukaemic secoeudesmane lactones (–)-eriolanin and (–)-eriolangin has been accomplished.³⁰¹ The acid catalysed isomerisation of several alantolactone derivatives has been studied.³⁰²

A rearranged eudesmane sesquiterpenoid 317, with a novel bicyclic ring system, has been obtained from Jasonia montana.³⁰³ This compound was named jasomontanone, but unfortunately a norsesquiterpene, also isolated from this species, had previously been given the same name.³⁰⁴ This plant also contains the sesquiterpene 11,15-dihydroxy-isoiphionane-4-one 318, which has been named montanone.305 A previous version of this work had been published, and considered in our last review.¹¹⁶ We think that the metabolite 317 is identical with montanone, because the physical and spectroscopic data reported for both compounds, 317 and 318, are very similar. In consequence one of these structures must be erroneous, although the NMR data are more in accordance with the structure 317. The isoiphionane sesquiterpene isofaurinone 319 has been found in the essential oil of Anthemis aciphylla.³⁰⁶ A formal synthesis of the iphionane derivative 320 has been achieved.³⁰⁷ The lindelanolide **321** has been isolated from *Sarcandra glabra*,²⁴⁶ while a lindenane dimer, chloromultilide A **322**, has been obtained from *Chloranthus multistachys*.³⁰⁸



17 Vetisperane and spiroaxane

The new bioactive spirovetivane sesquiterpene **323** has been obtained from a Vietnamese agarwood.³⁰⁹ A racemic total synthesis of α -vetisperane, hinesol and β -vetivone has been described,³¹⁰ whilst an enantiospecific total synthesis of (+)-solanascone, (+)-dehydrosolanascone and (+)-anhydro- β -rotunol has been accomplished.³¹¹ (+)-2 β -Hydroxysolanascone, the aglycone of the phytoalexin isolated from flue-cured tobacco leaves, has been synthesised.³¹² The stereochemistry and the deuterium isotope effects associated with the cyclisation and rearrangements of (*E*,*E*)-farnesyl diphosphate, catalysed by hyoscyamus premnaspirodiene synthase (HPS), to give premnaspirodiene **324** have been described.³¹³



A new pair of diastereomeric spiroaxene derivatives, **325** and **326**, has been isolated from both sponges *Amorphinopsis foetida* and *Axinyssa aplysinoides*, collected at Papua New Guinea and Vanuatu, respectively. The first sponge also contains the known metabolite **327**, whose stereochemistry has now been revised, whilst the second contains the spiroaxene **328**, whose stereochemistry has also been corrected, and its new diastereomer **329**.³¹⁴ Another compound of this type, 3-oxo-axisonitrile-3 **330**, has been found in a Chinese sponge of the *Acanthella* genus.³¹⁵









333



334 $R_1 = \beta$ -OAng $R_2 = Me$ **335** $R_1 = \alpha$ -OH $R_2 = H$



336

337 R = Ang

0

338 R = Ac 339 R = H

340 R = Ang(4'-OH)



341



342 R = Glc(6'-SO₃K)



afforded the new sesquiterpenes angulifolide **346** and angulifolins A–C **347–349**. The structures of peribysin C and peribysin D have been revised to **350** and **352**, respectively.³²⁶ These compounds had been isolated from a strain of *Periconia byssoides*, which was originally separated from the sea hare *Aplysia kurodai*.³²⁷ Another strain of *Periconia* sp.,³²⁸ isolated from the same sea hare, contains a further two new eremophilane derivatives, which have been named peribysin H **351** and peribysin I **353**.

The new eremophilanolides which have been isolated from different species are listed in Table 3. Their structures have been shown to be **354–381**.

The stereochemistry and the deuterium isotope effects associated with the cyclisation and rearrangements of (E,E)-farnesyl

18 Eremophilane and bakkane

Pleodendione 331 and remophilanetriol 332 are two new eremophilane sesquiterpenes, which have been obtained from Pleodendron costaricense⁴⁵ and Rehmannia glutinosa,³¹⁶ respectively. The genus Ligularia is a good source of eremophilane derivatives. The sesquiterpenes 333 and 334 have been isolated from Ligularia hodgsonii³¹⁷ and Ligularia myriocephala,³¹⁸ respectively, whilst the compounds 335-336 and virgaurenones A-D 337-340 have been found in the varieties oligocephala³¹⁹ and virgaurea,³²⁰ respectively, of Ligularia virgaurea. Chromatography of an extract from the roots and aerial parts of Senecio mairetianus321 afforded the eremophilane derivative 341. The Japanese butterbur (Petasitis japonicus) contains fukinoside A 342, a novel sesquiterpene glycoside sulfate with anti-allergic activity.322 Two chemotypes of Petasites hybridus are known; one produces petasin 343 whilst the other contains furanopetasin 344, but only the first is useful for phytopharmaceutical preparations. Experimental crossings of these chemotypes have been performed to study the genetic basis of the occurrence of these sesquiterpenes.323

The furanoeremophilane **345** has been obtained from *Senecio* kaschkarovii.³²⁴ A phytochemical study of *Roldana angulifolia*³²⁵



diphosphate, catalysed by tobacco epiaristolochene synthase (TEAS), to give the corresponding hydrocarbon have been investigated.³¹³ GC-MS methods have been used to make a detailed analysis of other sesquiterpenes formed in this cyclisation.³³⁸ It has previously been shown that the side chain of the phenylalanine-178 of aristolochene synthase promotes the conversion of a eudesmane cation to aristolochene. Now, it has been reported that this catalytic function is mainly due to the large size of this chain, which facilitates the hydride shift from C-2 to C-3.³³⁹

Table 3 Sources of eremophilanolides

Source	Eremophilanolides	Ref.
Ligularia hodgsonii Ligularia lapathifolia Ligularia muliensis Ligularia myriocephala Ligularia platyglossa Ligularia virgaurea var. oligocephala Ligularia virgaurea var. virgaurea Senecio burtonii Senecio mairetianus	354-356 357-359 360, 362 363-366 367 368, 369 370, 371 372 373-380	317 329, 330 331, 332 318, 333 334 335 320 336 321
Senecio poepigii	381	337

The photosensitised oxidation of several furanoeremophilane sesquiterpenes has been investigated by two different groups.^{340,341} (–)-Carvone has been used as starting material in a synthesis of (–)-aristolochene,³⁴² whilst a racemic synthesis of 6-hydroxyeuryopsin, 1,10-epoxy-6-hydroxyeuryopsin, toluccanolide A and toluccanolide C has been reported.³⁴³ The structures of several compounds produced in the basic hydrolysis of four eremophilane esters, which were isolated from *Robinsonecio gerberifolius*, have been assigned.³⁴⁴

A synthesis of (\pm) -bakkenolide A has been described,³⁴⁵ whilst an approach to the diastereoselective construction of the *nor*bakkane skeleton has been reported.³⁴⁶

19 Guaiane, pseudoguaiane, xanthane and patchoulane

The new azulene pigments **382** and **383–384** have been obtained from the fruiting bodies of *Lactarius deliciosus*³⁴⁷ and *Lactarius hatsudake*,³⁴⁸ respectively. Another compound of this type **385**, a known bioactive chamazulene derivative, has been found to be formed from the lactone matricin by treatment with artificial gastric fluids.³⁴⁹ The essential oil of *Ajania fruticulosa* contains the bisazulene **386**, whose structure was determined by X-ray analysis.³⁵⁰





363 $R_1 = OH$ $R_2 = \beta - OMe$ $R_3 = \beta - OH$ **364** $R_1 = OH$ $R_2 = \alpha - OMe$ $R_3 = \alpha - OH$ **365** $R_1 = OAng$ $R_2 = \beta - OH$ $R_3 = \beta - OH$ **366** $R_1 = OAng$ $R_2 = \alpha - OH$ $R_3 = \alpha - OH$



372 R = Meacr(4'-OH)



367



Ligulides C_1 and C_2 368 R = α -Me 369 R = β -Me



Virgaurenolides A and B

370 R = Ang **371** R = Ac



373 R = H374 R = OH



375 $R_1 = Me$ $R_2 = \alpha$ -OMe **376** $R_1 = Me$ $R_2 = \alpha$ -OH **377** $R_1 = Me$ $R_2 = \alpha$ -H **378** $R_1 = H$ $R_2 = \alpha$ -H **379** $R_1 = H$ $R_2 = \alpha$ -OH **380** $R_1 = H$ $R_2 = \beta$ -OH



381



386



Pubinernoid B **387** is a new guaiane derivative, which has been obtained from *Schisandra pubescens*.¹⁴³ A phytochemical study of the aerial parts of *Crysothamnus viscidiflorus* afforded a sesquiterpene **388**, which was named chrysothol.³⁵¹ The authors have indicated that the spectroscopic data of this compound were identical with those reported for buchariol, isolated from *Salvia bucharica*;³⁵² consequently the structure of the latter must be corrected. In addition, it is also shown in this work that the structure given to another sesquiterpene, obtained from *Fagonia boveana*,³⁵³ is erroneous. The structures **389** or **390** were proposed. The exudates of the aerial parts of *Balsamorhiza sagittata* and *Balsamorhiza macrophilla*³⁵⁴ contain the new guaiane derivative **391**. A study of a methanolic extract from the fruits of *Torilis*

*japonica*³⁵⁵ afforded the cytotoxic sesquiterpenes torilin **392**, 1 α -hydroxy-torilin **393** and the new metabolite 1 β -hydroxy-torilin **394**. It has also been shown that torilin may be an ideal antiarrhythmic drug for atrial fibrillation.³⁵⁶ The bioactive guaiane derivatives **395** and **396** have been identified as constituents of a soft coral, *Sinularia* sp.³⁵⁷

A total synthesis of the cytotoxic guaiapyridine sesquiterpene alkaloid (+)-cananodine **397** has been accomplished.³⁵⁸ The norsesquiterpene (-)-clavukerin A has been synthesised using a bioinspired procedure.³⁵⁹ The first total synthesis of (+)-alismoxide and (+)-4-*epi*-alismoxide has been achieved.³⁶⁰ In this work the structure of a natural guaienediol, isolated from *Silphium*



perfoliatum, has also been corrected to that of the known sesquiterpene (–)-alismoxide. The microbiological transformation of (–)guaiol by the fungus *Eurotium rubrum* has been investigated.³⁶¹ Synthetic approaches to hydroazulenes and guaianes have been reported.³⁶²

Many new guaianolides have been isolated during the period covered by this review (see Table 4). The novel guaian-6a,12-olides, *i.e.* **398–441**, are listed in Table 5 and other new guaianolides are represented by the 6β ,12-lactones **442–449**, the guaian-8,12-olides **450–451** and the dimers **452–457**.

The lactone hololeucin **429**, isolated from *Centaurea hololeuca*, represents the first example of a natural cyclic carbonate.³⁶⁴ Known guaianolides have been obtained from *Centaurea helenioides*,³⁷⁵ *Centaurea ptosimopappa*,³⁷⁶ *Centaurea scabiosa*,³⁷⁷ *Chondrilla juncea*,³⁷⁸ *Ixeris dentata*,³⁷⁹ *Rhaponticum pulchrum*³⁸⁰ and *Tarax-acum bessarabicum*³⁸¹ The structures of a guaianolide³⁸² **458** and of a tetrabromoderivative of cyclopropyldihydroargablin³⁸³ have been

Table 4 Sources of guaianolides

Source	Guaianolides	Ref.
Achillea asplenifolia	398, 405–408, 442, 443	237
Achillea clavennae	436–439	79
Achillea collina	399-402, 404, 409, 414, 444, 452	238
Amphoricarpos neumaveri	403, 431–434, 440	363
Balsamorhiza sagittata	411, 412	354
Centaurea hololeuca	429	364
Cichorium endivia	416	365
Crepis cameroonica	422	366
Eupatorium chinense	417-419, 425, 426, 441	240
Helianthus annuus	415	243
Inula britannica	453-456	367
Ixeris chinensis	424, 435	368
Ixeris sonchifolia	410	369
Laurus nobilis	423	370
Mulgedium tataricum	413	245
Salvia nubicola	450, 451, 457	371
Tanacetum fruticulosum	427, 428	372
Thansia garganica	445	373
Thansia nitida	446-448	230
Vernonia cinerea	449	374
Youngia japonica	420, 421, 430	26



determined by X-ray analysis. A conformational analysis of several 11,13-oxetane lactones has been carried out.³⁸⁴ The absolute configuration of the guaianolide **459** has been unambiguously determined by comparison of the calculated and experimental vibrational circular dichroism (VCD) spectra.³⁸⁵

The influence of the cultivation site on the guaianolide composition of *Cichorium intybus* has been studied.³⁸⁶ Some of the lactones isolated from *Eupatorium capillifolium* showed potent inhibitory effects on the growth of HeLa cells,³⁸⁷ whilst cynaropicrin has been shown to be a potent and irreversible inhibitor of the antibacterial target enzyme MurA.²⁵⁷ The biotransformation of dehydrocostuslactone by the fungus *Mucor polymorphosporus* has been investigated.³⁸⁸ An enantioselective synthesis of a 7,11dihydroxyguaianolide, possessing the stereochemistry of thapsigargin, has been described.³⁸⁹ A study of a thapsigargin derivative has revealed the importance of the length and flexibility of the side chain attached to O-8 in the activity of this sesquiterpene lactone, and its analogues, towards prostate cancer cells.³⁹⁰

Five novel pseudoguaianolides, dichrocepholides A–E **460–464**, have been isolated from *Dichrocephala integrifolia*.³⁹¹ Another two new compounds of this type, deacetyltetraneurin A **465** and hysterone E **466**, have been found in an extract from the flowers of *Parthenium hysterophorus*.³⁹² An enantioselective synthesis of (+)-8-*epi*-xanthatin has been reported.³⁹³

Table 5 Novel guaian-6α,12-olides



Name	Structure	Position of double bond(s)	Substituents and configurations	Ref.
Solidaginolide A isomer	398		1a.2a-epoxy. 4a.5a-epoxy. 8a-OAng, 10B-OH, 11a	237
Solidaginolide A isomer	399		$1\alpha.2\alpha$ -epoxy, $4\alpha.5\alpha$ -epoxy, 8α -OTig, $10B$ -OH, 11α	238
10- <i>eni</i> -Ezoartemin derivative	400		16.26 -epoxy, 36.46 -epoxy, 8α -OTig, 106 -OH, 11α	238
10- <i>eni</i> -Solidaginolide A	401		$1\alpha.2\alpha$ -epoxy. $3\alpha.4\alpha$ -epoxy. 8α -OTig. 10B-OH, 11 α	238
10- <i>eni</i> -Solidaginolide A deriv.	402		$1\alpha.2\alpha$ -epoxy. $3\alpha.4\alpha$ -epoxy. 8α -OAng. 10B-OH. 11 α	238
Amphoricarpolide deriv.	403		4α , 10α , 14 -epoxy, 11α -OH, 13 -OH	363
Artabsinolide A deriv.	404	1-5	2-oxo. 4α -OH. 8α -OTig. 10B-OH. 11 α	238
Tannunolide B derivative	405	2-3, 4-5, 10-1	8α -OAc (or -OAng). 11 α	237
Tannunolide C derivative	406	2-3, 4-15, 10-1	8α-OAng. 11α	237
Matricin derivative	407	2-3, 10-1	4α-OMe, 8α-OAng, 11α	237
Matricin derivative	408	2-3, 10-1	4β-OH (or -OMe), 8α-OAng, 11α	237
Tanaparthin derivative	409	2–3	2α.4α-dioxo, 8α-OCOPri (or -OTig), 10β-OH, 11α	238
Ixerin Z_{A}	410	3-4, 10-1, 11-13	2-oxo, 3-OGlc(6'-p-AcOBz)	369
Zubergenin derivative	411	3-4, 10-1, 11-13	8α-OAc, 9β-OEpang	354
Zubergenin derivative	412	3-4, 10-1, 11-13	5α-OH, 8α-OAc, 9β-OAng	354
Lactucin derivative	413	3-4, 10-1, 11-13	2-oxo, 8α-OR ₁ , 15-OH	245
5α-Hydroxymatricarin	414	3-4, 10-1	2-oxo, 5α-OH, 8α-OAc, 11α	238
Annuolide H	415	3-4, 11-13	2α-OH, 8β-OAng, 10α-OH	243
Hieracin derivative	416	3–4	2-oxo, 8α-OH, 10β-OMe, 11α, 15-OH	365
Eupatochinilide V	417	4-5, 10,1, 11-13	3β-OH, 8β-OAng, 14-oxo	240
Eupatochinilide VI	418	4-5, 10,1, 11-13	3β-OH, 8α-OAng, 14-oxo	240
Eupatochinilide VII	419	4-5, 10,1, 11-13	3β-OH, 8β-OEpang, 14-oxo	240
Zulazanin C derivative	420	4-15, 10-14, 11-13	3β -OGlc(3'-OR ₁)	26
Zulazanin C derivative	421	4-15, 10-14, 11-13	3β -OGlc(3'-OR ₁), 8α -OH	26
8-Desacylcynaropicrin	422	4-15, 10-14, 11-13	3β-ОН, 9β-ОН	366
Zaluzanin C derivative	423	4–15, 10–14, 11–13	3β-Cl	370
Chinensiolide E	424	4–15, 11–13	3-OR ₂ , 10α-OH	368
Eupatochinilide II	425	10–1, 11–13	3α,4α-epoxy, 8β-OEpang, 14-OH	240
Eupatochinilide IV	426	10–1, 11–13	3β-Cl, 4α-OH, 8β-OAng, 14-OH	240
Carlaolide A	427	10–1, 11–13	3β,4β-epoxy, 8α-OCOPri	372
Carlaolide B	428	10–1, 11–13	3β,4β-epoxy, 8α-OAng	372
Hololeucin	429	10–14, 11–13	3β,4β-carbonate, 8α-OMeacr(4'-OH), 15-OH	364
Zaluzanin C derivative	430	10–14, 11–13	3 -oxo, 4α , 8α -OR ₁	26
Amphoricarpolide deriv.	431	10–14, 11–13	3β-OH, 4α, 9β-OH, 15-OAc	363
Amphoricarpolide deriv.	432	10–14, 11–13	4α, 15-OAc	363
Amphoricarpolide deriv.	433	10–14	4α, 11α-OH, 13-OH, 15-OH (or -OAc)	363
Amphoricarpolide deriv.	434	10–14	4α, 11α-OH, 13-Cl, 15-OAc	363
Chinensiolide D	435	11–13	3-οχο, 4β, 10α-ΟΗ	368
9α-Acetoxyartecanin	436	11–13	1β,2β-ероху, 3β,4β-ероху, 9α-ОАс, 10α-ОН	79
A chlorohydrin	437	11–13	1β,2β-epoxy, 3α-Cl, 4β-OH, 10α-OH	79
A chlorohydrin	438	11–13	1β,2β-ероху, 3α-Cl, 4β-OH, 9α-OAc, 10α-OH	79
9α-Acetoxyandalucin	439	11–13	1α,2α-epoxy, 3α-OH, 4β-Cl, 9α-OAc, 10α-OH	79
Amphoricarpolide deriv.	440	11-13	4a, 10a,14-epoxy, 15-OH	363
Eupatochinilide III	441	11-13	3α,4α-epoxy, 8β-OTig(4'-OH), 10α,14-epoxy	240

 $R_1 = p - CO - CH_2 - (C_6H_4)OH R_2 = Glc(2' - OR_1, 6' - OR_1).$

The diverse sesquiterpene profile of *Pogostemon cablin* has been correlated with the existence of a limited number of sesquiterpene synthases in this species.³⁹⁴ Cyperotundone and α -cyperone have been obtained from dried tubers of *Cyperus rotundus*. The first of these ketones was oxidised with selenium dioxide to give 4-patchoulene-2,3-dione, which showed plant growth regulatory properties.³⁹⁵ A new synthetic route to the patchoulol skeleton has been devised. The compounds obtained have permitted novel insights into the structural requirements of patchouli odorants.³⁹⁶

20 Aromadendrane, bicyclogermacrane, aristolane, sinulariolane and valerenane

The Formosan soft coral *Clavularia inflata* var. *luzoniana*³⁹⁷ contains three new aromadendrane sesquiterpenoids **467–469**. The known sesquiterpene ledol has been obtained from the leaf surface of *Solanum tuberosum*.²³² A synthesis of (\pm) -epiglobulol has been reported.³⁹⁸

Kissoones A-C **470-472** and the compound **473** are four rearranged bicyclogermacrane derivatives, which have been isolated





Inulanolide C 455



Inulanolide D 456



Bisnubenolide 457



from the roots of *Valeriana fauriei*.^{399,400} The biotransformations of the sesquiterpenes (+)-aristol-1(10)-ene and plagiochilide by three microorganisms, *Chlorella fusca* var. *vacuolata*, a *Mucor* species and *Aspergillus niger*, have been investigated.⁴⁰¹ Two novel valerenane sesquiterpenes, caulerpal A **474** and caulerpal B **475**, have been found in the Chinese green alga *Caulerpa taxifolia*.⁴⁰² The structure of another valerenane derivative sinularianin B has been determined as **476**. This sesquiterpenoid has been isolated from a Formosan soft coral of the *Sinularia* genus.⁴⁰³ Other known sesquiterpenes of this type valerenal, valerenic acid and acetylvalerenolic acid have been obtained by a bioassay-guided fractionation of an extract from *Valeriana officinalis*. The two acids showed inhibitory activity against NF-κB in HeLa cells.⁴⁰⁴

21 Cyclomyltaylane

Five novel cyclomyltaylanoids **477–481** have been found in an extract from the Malagasy liverwort *Bazzania madagassa*.⁴⁰⁵ Another liverwort, *Reboulia hemisphaerica*, collected in Japan, contains three new cyclomyltaylane derivatives **482–484** and the previously known cyclomyltaylan-5 α -ol **485**. The absolute configuration of the latter has now been determined by application of the modified





461 R = α-OH
462 R = β-OH



464 R = β-OH





Mosher's method to one of the metabolites **486**, which was also obtained in this work by biotransformation of 485 with the fungus Aspergillus niger.¹²⁴

484

485







479 $R_1 = Me R_2 = H R_3 = Ac$ **480** $R_1 = CH_2OAc$ $R_2 = H$ $R_3 = Ac$



482 R = H 483 R = OAc



481 $R_1 = CH_2OAc R_2 = Ac R_3 = H$

 $R_1 = \beta$ -OH $R_2 = H$

 $R_1 = \alpha$ -OH $R_2 = H$

486 $R_1 = \alpha$ -OH $R_2 = OH$

Salvionane

22

A new "isodaucane" (salvionane) derivative 487 has been isolated from the bark of Jatropha neopauciflora.140

Miscellaneous sesquiterpenoids 23

The full paper with the structure of stereumone A has appeared.⁴⁰⁶ Vibralactone 488 has been obtained from cultures of the basidiomycete Boreostereum vibrans. This C13 derivative inhibits pancreatic lipase.⁴⁰⁷ The total synthesis of the fungal metabolites (+)massarinolin B and (+)-4-epi-massarinolin B has been achieved.408



Naupliolide 489, a sesquiterpene with a new carbon framework, has been isolated from Nauplius graveolens. The authors suggest that this metabolite may be formed from asteriscunolide C, also isolated from this plant.⁴⁰⁹ The novel lactone 490 has been found in the study of an extract from Laurus nobilis.370 An enantioselective synthesis of the (1S, 5R)-enantiomer of litseaverticillol A and litseaverticillol B has been accomplished.410 On the other hand, successful biomimetic syntheses of the litseaverticillol family of sesquiterpenes have been achieved, using singlet oxygen chemistry.411 In this work the structure of litseaverticillol E has been revised to 491. A total synthesis of racemic salsolene oxide 492 has been reported.412



Five minor sesquiterpenes 493-497, have been isolated from the brown alga Dictyopteris divaricata. These compounds have two new different carbon frameworks, which may be derived from a cadinane.¹⁸⁴ The Formosan soft coral Clavularia inflata var. luzoniana³⁹⁷ contains the sesquiterpenoid 498, which possesses a new ring system. Three sesquiterpenes with a novel carbon skeleton, paralemnanone 499, isoparalemnanone, 500 and paralemnanol 501, have been isolated from another soft coral,

*Paralemnalia thyrsoides.*⁴¹³ The sesquiterpene sinularianin A **502**, which also has a new carbon framework named sinulariolane, has been isolated from a coral of the *Sinularia* genus.⁴⁰³ The Okinawan marine sponge *Dysidea chlorea* contains four new sesquiterpenes, haterumadysins A–D **503–506**. These compounds inhibit the division of fertilised sea urchin eggs.⁴¹⁴ Spirofragilin **507** has been found in another species of this genus, *Dysidea fragilis*, which was collected in the South China Sea,⁴¹⁵ whilst *O*-methyl nakafuran-8 lactone **508** has been obtained from a





493

494 $R_1 = \alpha - H$ $R_2 = \beta - OH$ **495** $R_1 = \beta - H$ $R_2 = \alpha - OH$



496









499 R = β-OH **500** R = α-OH





502



503



Hainan sponge of the same genus.⁴¹⁶ Enantiospecific syntheses of 2-pupukeanone⁴¹⁷ and several allopupukeanones⁴¹⁸ have been achieved. Two new cytotoxic sesquiterpenoids, metachromin J **509** and metachromin K **510**, have been isolated from a marine sponge of the *Spongia* genus.⁴¹⁹



The germination of the seeds of some parasitic plants depends on the secretion of strigolactones. The occurrence, isolation, biosynthesis and the mode of action of these carotenoid-derived metabolites have been reviewed.^{420,421}

24 References

- 1 C. Zidorn, Biochem. Syst. Ecol., 2006, 34, 144.
- 2 X. T. Si, M. L. Zhang, Q. W. Shi and H. Kiyota, *Chem. Biodiversity*, 2006, **3**, 1163.
- 3 X. T. Si, M. L. Zhang, Q. W. Shi and H. Kiyota, *Chem. Biodiversity*, 2006, **3**, 371.
- 4 I. Werner, S. Glasl and G. Reznicek, Chem. Biodiversity, 2006, 3, 27.
- 5 K. L. Silva-Brandao, V. N. Solferini and J. R. Trigo, *Biochem. Syst. Ecol.*, 2006, 34, 291.
- 6 M. T. Lindenmeyer, A. Hrenn, C. Kern, V. Castro, R. Murillo, S. Müller, S. Laufer, J. Schulte-Mönting, B. Siedle and I. Merfort, *Bioorg. Med. Chem.*, 2006, 14, 2487.
- 7 T. G. Köllner, P. E. O'Maille, N. Gatto, W. Boland, J. Gershenzon and J. Degenhardt, Arch. Biochem. Biophys., 2006, 448, 83.
- 8 Y. Liu, S. Zhang and P. J. M. Abreu, Nat. Prod. Rep., 2006, 23, 630.
- 9 B. D'Abrosca, P. De Maria, M. DellaGreca, A. Fiorentino, A. Golino, A. Izzo and P. Monaco, *Tetrahedron*, 2006, **62**, 640.
- 10 A. Fiorentino, M. DellaGreca, B. D'Abrosca, A. Golino, S. Pacifico, A. Izzo and P. Monaco, *Tetrahedron*, 2006, **62**, 8952.
- 11 I. Vuckovic, L. Vujisic, V. Vajs, V. Tesevic, S. Macura, P. Janackovic and S. Milosavljevic, *Biochem. Syst. Ecol.*, 2006, 34, 303.
- 12 C. H. Chao, C. H. Hsieh, S. P. Chen, C. K. Lu, C. F. Dai, Y. C. Wu and J. H. Sheu, *Tetrahedron Lett.*, 2006, 47, 2175.
- 13 D. Hampel, A. Mosandl and M. Wüst, J. Agric. Food Chem., 2006, 54, 1473.
- 14 Y. Yang, J. S. Yuan, J. Ross, J. P. Noel, E. Pichersky and F. Chen, Arch. Biochem. Biophys., 2006, 448, 123.

- 15 Y. L. Chen and H. T. Chiu, J. Chin. Chem. Soc., 2006, 53, 1161.
- 16 C. D. Poulter, Phytochem. Rev., 2006, 5, 17.
- 17 Y. Li, T. Zhang and Y. L. Li, Chin. Chem. Lett., 2006, 17, 147.
- 18 L. Commeiras, J. Bourdron, S. Douillard, P. Barbier, N. Vanthuyne, V. Peyrot and J. L. Parrain, *Synthesis*, 2006, 166.
- 19 B. A. Scheepers, R. Klein and M. T. Davies-Coleman, *Tetrahedron Lett.*, 2006, 47, 8243.
- 20 M. Grin'ko, V. Kul'chitskii, N. Ungur and P. F. Vlad, Chem. Nat. Compd. (Engl. Transl.), 2006, 42, 439.
- 21 T. Motai and S. Kitanaka, J. Nat. Med., 2006, 60, 54.
- 22 M. H. Oueslati, H. B. Jannet, Z. Mighri, J. Chriaa and P. M. Abreu, J. Nat. Prod., 2006, 69, 1366.
- 23 E. Sueyoshi, H. Liu, K. Matsunami, H. Otsuka, T. Shinzato, M. Aramoto and Y. Takeda, *Phytochemistry*, 2006, 67, 2483.
- 24 X. H. Xu, C. H. Tan, S. H. Jiang and D. Y. Zhu, *Helv. Chim. Acta*, 2006, 89, 1422.
- 25 K. Matsunami, I. Takamori, T. Shinzato, M. Aramoto, K. Kondo, H. Otsuka and Y. Takeda, *Chem. Pharm. Bull.*, 2006, 54, 1403.
- 26 W. Chen, Q. Liu, J. Wang, J. Zhou, D. Meng, J. Zuo, X. Zhu and W. Zhao, *Planta Med.*, 2006, **72**, 143.
- 27 T. Kanchanapoom, A. Sirikatitham, H. Otsuka and S. Ruchirawat, J. Asian Nat. Prod. Res., 2006, 8, 747.
- 28 T. Itoh, Y. Itoh, H. Hibasami, H. Hiroshige, H. Katsuzaki, K. Imai, Y. Furuichi and T. Komiya, *Nippon Shokuhin Kagaku Kogaku Kaishi*, 2005, **52**, 319, (*Chem. Abstr.*, 2006, **144**, 127875).
- 29 W. Schliemann, J. Schmidt, M. Nimtz, V. Wray, T. Fester and D. Strack, *Phytochemistry*, 2006, 67, 1196.
- 30 M. Ibdah, Y. Azulay, V. Portnoy, B. Wasserman, E. Bar, A. Meir, Y. Burger, J. Hirschberg, A. A. Schaffer, N. Katzir, Y. Tadmor and E. Lewinsohn, *Phytochemistry*, 2006, 67, 1579.
- 31 Z. H. Yuan, L. J. Han, X. Fan, M. Ma, D. Y. Shi and J. G. Shi, *Chin. Chem. Lett.*, 2006, **17**, 1205.
- 32 D. S. Makhanu, M. Yokoyama, T. Miono, T. Maesato, M. Maedomari, P. Wisespongpand and M. Kuniyoshi, *Bull. Fac. Sci., Univ. Ryukyus*, 2006, **81**, 115, (*Chem. Abstr.*, 2006, **145**, 372473).
- 33 S. Serra and C. Fuganti, Tetrahedron: Asymmetry, 2006, 17, 1573.
- 34 S. Serra, C. Fuganti and E. Brenna, Helv. Chim. Acta, 2006, 89, 1110.
- 35 N. W. Jan and H. J. Liu, Org. Lett., 2006, 8, 151.
- 36 Y. Aubin, G. Audran and H. Monti, Tetrahedron Lett., 2006, 47, 3669.
- 37 U. Groth, C. Kesenheimer and P. Kreye, Synlett, 2006, 2223.
- 38 N. Kitahata, S. Y. Han, N. Noji, T. Saito, M. Kobayashi, T. Nakano, K. Kuchitsu, K. Shinozaki, S. Yoshida, S. Matsumoto, M. Tsujimoto and T. Asami, *Bioorg. Med. Chem.*, 2006, 14, 5555.
- 39 T. R. Smith, A. J. Clark, G. J. Clarkson, P. C. Taylor and A. Marsh, Org. Biomol. Chem., 2006, 4, 4186.
- 40 Y. Araki, A. Miyawaki, T. Miyashita, M. Mizutani, N. Hirai and Y. Todoroki, *Bioorg. Med. Chem. Lett.*, 2006, 16, 3302.
- 41 J. M. Nyangulu, K. M. Nelson, P. A. Rose, Y. Gai, M. Loewen, B. Lougheed, J. W. Quail, A. J. Cutler and S. R. Abrams, *Org. Biomol. Chem.*, 2006, 4, 1400.
- 42 H. V. K. Wangun, H. Doerfelt and C. Hertweck, *Eur. J. Org. Chem.*, 2006, 1643.
- 43 Y. H. Kim, B. S. Yun, I. J. Ryoo, J. P. Kim, H. Koshino and I. D. Yoo, J. Antibiot., 2006, 59, 432.
- 44 X. L. Li, Y. Zhao, X. Cheng, L. Tu, L. Y. Peng, G. Xu and Q. S. Zhao, *Helv. Chim. Acta*, 2006, **89**, 1467.
- 45 V. T. Amiguet, P. Petit, C. A. Ta, R. Nuñez, P. Sanchez-Vindas, L. Poveda-Alvarez, M. L. Smith, J. T. Arnason and T. Durst, J. Nat. Prod., 2006, 69, 1005.
- 46 N. J. Brennan, L. Larsen, S. D. Lorimer, N. B. Perry, E. L. Chapin, T. L. Werk, M. J. Henry and D. R. Hahn, *J. Agric. Food Chem.*, 2006, 54, 468.
- 47 Y. X. Zhang, S. H. Jackson, M. S. Rajab, F. R. Fronczek and S. F. Watkins, Acta Crystallogr., Sect. C: Cryst. Struct. Commun., 2006, 62, 0219.
- 48 A. A. Wube, S. Gibbons, K. Asres, B. Streit, M. Adams, R. Bauer and F. Bucar, *Planta Med.*, 2006, **72**, 754.
- 49 L. Harinantenaina and S. Takaoka, J. Nat. Prod., 2006, 69, 1193.
- 50 R. Uchida, Y. P. Kim, I. Namatame, H. Tomoda and S. Omura, J. Antibiot., 2006, 59, 93.
- 51 R. Uchida, H. Tomoda and S. Omura, J. Antibiot., 2006, 59, 298.
- 52 R. Uchida, Y. P. Kim, T. Nagamitsu, H. Tomoda and S. Omura, J. Antibiot., 2006, **59**, 338.
- 53 V. Rojas de la Parra, V. Mierau, T. Anke and O. Sterner, *Tetrahedron*, 2006, 62, 1828.

- 54 V. Mierau, V. Rojas de la Parra, O. Sterner and T. Anke, *J. Antibiot.*, 2006, **59**, 53.
- 55 V. Rojas de Parra, V. Mierau, T. Anke and O. Sterner, J. Antibiot., 2006, **59**, 57.
- 56 M. Cueto, J. B. MacMillan, P. R. Jensen and W. Fenical, *Phytochem-istry*, 2006, 67, 1826.
- 57 A. R. Diaz-Marrero, P. Austin, R. van Soest, T. Matainaho, C. D. Roskelley, M. Roberge and R. J. Andersen, Org. Lett., 2006, 8, 3749.
- 58 J. L. Carballo, E. Zubia and M. J. Ortega, *Biochem. Syst. Ecol.*, 2006, 34, 498.
- 59 D. Sladic and M. J. Gasic, *Molecules*, 2006, 11, 1.
- 60 A. Nasib, S. G. Musharraf, S. Hussain, S. Khan, S. Anjum, S. Ali, Atta-ur-Rahman and M. I. Choudhary, J. Nat. Prod., 2006, 69, 957.
- 61 H. J. Yang, B. G. Li, X. H. Cai, H. Y. Qi, Y. G. Luo and G. L. Zhang, J. Nat. Prod., 2006, 69, 1531.
- 62 C. Fehr and I. Farris, Angew. Chem., Int. Ed., 2006, 45, 6904.
- 63 L. C. Henderson, W. A. Loughlin, I. D. Jenkins, P. C. Healy and M. R. Campitelli, J. Org. Chem., 2006, 71, 2384.
- 64 J. S. Foot, A. T. Phillis, P. P. Sharp, A. C. Willis and M. G. Banwell, *Tetrahedron Lett.*, 2006, 47, 6817.
- 65 P. F. Vlad, A. C. Ciocarlan, M. N. Coltsa, C. Deleanu, O. Costan, Y. A. Simonov, V. Ch. Kravtsov, J. Lipkowski, T. Lis and A. de Groot, *Tetrahedron*, 2006, **62**, 8489.
- 66 A. Bernet and K. Seifert, Helv. Chim. Acta, 2006, 89, 784.
- 67 J. Villamizar, F. Plata, N. Canudas, E. Tropper, J. Fuentes and A. Orcajo, *Synth. Commun.*, 2006, 311.
- 68 X. Gao, Y. Matsuo and B. B. Snider, Org. Lett., 2006, 8, 2123.
- 69 A. V. Kurdyumov and R. P. Hsung, J. Am. Chem. Soc., 2006, 128, 6273.
- 70 A. Gansäuer, A. Rosales and J. Justicia, Synlett, 2006, 927.
- 71 A. Gansäuer, J. Justicia, A. Rosales, D. Worgull, B. Rinker, J. M. Cuerva and J. E. Oltra, *Eur. J. Org. Chem.*, 2006, 4115.
- 72 H. Liu, D. R. Siegel and S. J. Danishefsky, Org. Lett., 2006, 8, 423.
- 73 G. Appendino, L. Maxia, M. Bascope, P. J. Houghton, G. Sanchez-Duffhues, E. Muñoz and O. Sterner, J. Nat. Prod., 2006, 69, 1101.
- 74 C. Barthomeuf, M. Demeule, J. Grassi, A. Saidkhodjaev and R. Beliveau, *Planta Med.*, 2006, **72**, 634.
- 75 D. Bandyopadhyay, B. Basak, A. Chatterjee, T. K. Lai, A. Banerji, J. Banerji, A. Neuman and T. Prange, *Nat. Prod. Res.*, 2006, 20, 961.
- 76 J. R. Yang, Z. An, Z. H. Li, S. Jing and H. L. Qin, *Chem. Pharm. Bull.*, 2006, 54, 1595.
- 77 M. Borges, M. Birkett, J. R. Aldrich, J. E. Oliver, M. Chiba, Y. Murata, R. A. Laumann, J. A. Barrigossi, J. A. Pickett and M. C. B. Moraes, *J. Chem. Ecol.*, 2006, **32**, 2749.
- 78 C. M. Liu, D. Q. Fei, Q. H. Wu and K. Gao, J. Nat. Prod., 2006, 69, 695.
- 79 S. Trifunovic, V. Vajs, Z. Juranic, Z. Zizak, V. Tesevic, S. Macura and S. Milosavljevic, *Phytochemistry*, 2006, 67, 887.
- 80 M. Ono, T. Tsuro, H. Abe, M. Eto, M. Okawa, F. Abe, J. Kinjo, T. Ikeda and T. Nohara, J. Nat. Prod., 2006, 69, 1417.
- 81 D. A. Mulholland, K. McFarland and M. Randrianarivelojosia, *Biochem. Syst. Ecol.*, 2006, 34, 365.
- 82 J. X. Li, X. P. Yang and Z. J. Jia, Chin. Chem. Lett., 2006, 17, 776.
- 83 J. X. Li, C. J. Lin, X. P. Yang and Z. J. Jia, *Chem. Biodiversity*, 2006, 3, 783.
- 84 M. Fukuda, E. Ohkoshi, M. Makino and Y. Fujimoto, *Chem. Pharm. Bull.*, 2006, 54, 1465.
- 85 S. W. Kang, H. K. Kim, W. J. Lee and Y. J. Ahn, J. Agric. Food Chem., 2006, 54, 3547.
- 86 Y. Letourneux, J. M. Brunel, R. Fernández, M. Dherbomez and C. Debitus, *Heterocycl. Commun.*, 2005, 11, 291.
- 87 D. Davyt, R. Fernández, L. Suescun, A. W. Mombrú, J. Saldaña, L. Domínguez, M. T. Fujii and E. Manta, J. Nat. Prod., 2006, 69, 1113.
- 88 L. R. de Carvalho, M. T. Fujii, N. F. Roque and J. H. G. Lago, *Phytochemistry*, 2006, **67**, 1331.
- 89 I. Brito, T. Dias, A. R. Díaz-Marrero, J. Darias and M. Cueto, *Tetrahedron*, 2006, **62**, 9655.
- 90 D. Spitzner and J. Zepf, Nat. Prod. Res., 2006, 20, 99.
- 91 F. A. Macías, J. L. G. Galindo, R. M. Varela, A. Torres, J. M. G. Molinillo and F. R. Fronczek, Org. Lett., 2006, 8, 4513.
- 92 S. Morimoto, M. Shindo, M. Yoshida and K. Shishido, *Tetrahedron Lett.*, 2006, 47, 7353.
- 93 B. Biswas, P. K. Sen and R. V. Venkateswaran, *Tetrahedron Lett.*, 2006, 47, 4019.
- 94 A. Fürstner and P. Hannen, Chem.-Eur. J., 2006, 12, 3006.

- 95 Y. Ohta and Y. Hirose, Tetrahedron Lett., 1968, 1251.
- 96 K. Monde, T. Taniguchi, N. Miura, C. S. Vairappan and M. Suzuki, *Chirality*, 2006, 18, 335.
- 97 S. P. Chaván, M. Thakkar, G. F. Jogdand and U. R. Kalkote, J. Org. Chem., 2006, 71, 8986.
- 98 F. A. Bermejo, A. Fernández-Mateos, A. Marcos-Escribano, R. Martín-Lago, L. Mateos-Burón, M. Rodríguez-López and R. Rubio-González, *Tetrahedron*, 2006, 62, 8933.
- 99 T. H. Kim, H. Ito, T. Hatano, J. Takayasu, H. Tokuda, H. Nishino, T. Machiguchi and T. Yoshida, *Tetrahedron*, 2006, 62, 6981.
- 100 T. Ochi, H. Shibata, T. Higuti, K. Kodama, T. Kusumi and Y. Takaishi, J. Nat. Prod., 2005, 68, 819.
- 101 C. G. Jones, E. L. Ghisalberti, J. A. Plummer and E. L. Barbour, *Phytochemistry*, 2006, 67, 2463.
- 102 E. Heuberger, T. Hongratanaworakit and G. Buchbauer, *Planta Med.*, 2006, **72**, 792.
- 103 J. Yamaguchi, M. Toyoshima, M. Shoji, H. Kakeya, H. Osada and Y. Hayashi, Angew. Chem., Int. Ed., 2006, 45, 789.
- 104 P. Blagojevic, N. Radulovic, R. Palic and G. Stojanovic, J. Agric. Food Chem., 2006, 54, 4780.
- 105 S. Schoettler, M. Bascope, O. Sterner and T. Anke, Z. Naturforsch., C: Biosci., 2006, 61, 309.
- 106 J. Xu, A. Takasaki, H. Kobayashi, T. Oda, J. Yamada, R. E. P. Mangindaan, K. Ukai, H. Nagai and M. Namikoshi, J. Antibiot., 2006, 59, 451.
- 107 L. Shen, R. H. Jiao, Y. H. Ye, X. T. Wang, C. Xu, Y. C. Song, H. L. Zhu and R. X. Tan, *Chem.–Eur. J.*, 2006, **12**, 5596.
- 108 T. Amagata, C. Rath, J. F. Rigot, N. Tarlov, K. Tenney, F. A. Valeriote and P. Crews, J. Med. Chem., 2003, 46, 4342.
- 109 T. V. Milanez and L. M. Valente-Soares, J. Braz. Chem. Soc., 2006, 17, 412.
- 110 S. Asam and M. Rychlik, J. Agric. Food Chem., 2006, 54, 6535.
- 111 Y. Cetin and L. B. Bullerman, J. Agric. Food Chem., 2006, 54, 1949.
- 112 Y. J. Hong and D. J. Tantillo, Org. Lett., 2006, 8, 4601.
- 113 F. Nagashima, M. Toyota and Y. Asakawa, *Chem. Pharm. Bull.*, 2006, 54, 1347.
- 114 M. Kladi, H. Xenaki, C. Vagias, P. Papazafiri and V. Roussis, *Tetrahedron*, 2006, **62**, 182.
- 115 S. C. Mao and Y. W. Guo, J. Nat. Prod., 2006, 69, 1209.
- 116 B. M. Fraga, Nat. Prod. Rev., 2006, 23, 943.
- 117 A. Srikrishna and P. C. Ravikumar, Tetrahedron, 2006, 62, 9393.
- 118 T. Matsuda, M. Shigeno, M. Makino and M. Murakami, Org. Lett., 2006, 8, 3379.
- 119 M. G. Kulkami, S. I. Davawala, M. P. Shinde, A. P. Dhondge, A. S. Borhade, S. W. Chavhan and D. D. Gaikwad, *Tetrahedron Lett.*, 2006, 47, 3027.
- 120 A. Srikrishna and G. Satyanarayana, Tetrahedron, 2006, 62, 2892.
- 121 A. Srikrishna and D. B. Ramachary, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2006, 45, 1216.
- 122 A. Srikrishna, B. V. Lakshmi and P. C. Ravikumar, *Tetrahedron Lett.*, 2006, 47, 1277.
- 123 A. Srikrishna and G. Satyanarayana, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2006, 45, 2465.
- 124 M. Furusawa, T. Hashimoto, Y. Noma and Y. Asakawa, *Chem. Pharm. Bull.*, 2006, 54, 996.
- 125 D. B. Sudatti, S. V. Rodrigues and R. C. Pereira, J. Chem. Ecol., 2006, 32, 835.
- 126 A. Srikrishna, B. V. Lakshmi and M. Mathews, *Tetrahedron Lett.*, 2006, 47, 2103.
- 127 F. Marion, D. E. Williams, B. O. Patrick, I. Hollander, R. Mallon, S. C. Kim, D. M. Roll, L. Feldberg, R. van Soest and R. J. Andersen, *Org. Lett.*, 2006, 8, 321.
- 128 Y. Ortíz-Nuñez, I. Spengler-Salabarria, I. G. Collado and R. Hernández-Galán, J. Agric. Food Chem., 2006, 54, 7517.
- 129 J. J. Crawford, W. J. Kerr, M. McLaughlin, A. J. Morrison, P. L. Pauson and G. J. Thurston, *Tetrahedron*, 2006, 62, 11360.
- 130 D. Chappell and A. T. Russell, Org. Biomol. Chem., 2006, 4, 4409.
- 131 X. Lin, R. Hopson and D. E. Cane, J. Am. Chem. Soc., 2006, 128, 6022.
- 132 X. J. Dong, X. D. Zhu, Y. F. Wang, Q. Wang, P. Ju and S. Luo, *Helv. Chim. Acta*, 2006, **89**, 983.
- 133 D. A. Carcache, Y. S. Cho, Z. Hua, Y. Tian, Y. M. Li and S. M. Danishefsky, J. Am. Chem. Soc., 2006, 128, 1016.
- 134 G. Mehta and S. R. Singh, Angew. Chem., Int. Ed., 2006, 45, 953.

- 135 S. P. Cook, A. Polara and S. J. Danishefsky, J. Am. Chem. Soc., 2006, 128, 16440.
- 136 F. Cutillo, B. D'Abrosca, M. DellaGreca, A. Fiorentino and A. Zarrelli, *Phytochemistry*, 2006, 67, 481.
- 137 F. Ahmed, M. Ali and O. Singh, Pharmazie, 2006, 61, 728
- 138 M. Z. Wang, Y. Y. Zhang, S. L. Li, X. H. Cai and X. D. Luo, *Helv. Chim. Acta*, 2006, 89, 3104.
- 139 L. He, J. Yang, A. C. Cao, Y. M. Liu, Y. An and J. G. Shi, *Chin. J. Chem.*, 2006, 24, 1375.
- 140 A. Garcia and G. Delgado, Helv. Chim. Acta, 2006, 89, 16.
- 141 H. H. Issa, S. M. Chang, Y. L. Yang, F. R. Chang and Y. C. Wu, *Chem. Pharm. Bull.*, 2006, 54, 1599.
- 142 R. Rojas, B. Bustamante, P. Ventosilla, I. Fernández, L. Caviedes, R. H. Gilman, O. Lock and G. B. Hammond, *Chem. Pharm. Bull.*, 2006, 54, 278.
- 143 S. X. Huang, J. Yang, W. L. Xiao, Y. L. Zhu, R. T. Li, L. M. Li, J. X. Pu, X. Li, S. H. Li and H. D. Sun, *Helv. Chim. Acta*, 2006, 89, 1169.
- 144 K. Yoshikawa, T. Tanaka, A. Umeyama and S. Arihara, *Chem. Pharm. Bull.*, 2006, 54, 315.
- 145 K. Yoshikawa, K. Suzuki, A. Umeyama and S. Arihara, *Chem. Pharm. Bull.*, 2006, 54, 574.
- 146 C. L. Hsieh, L. L. Shiu, M. H. Tseng, Y. Y. Shao and Y. H. Kuo, J. Nat. Prod., 2006, 69, 665.
- 147 G. H. Silva, H. Lopes-Teles, L. M. Zanardi, M. C. Marx-Young, M. Nogueira-Eberlin, R. Hadad, L. H. Pfenning, C. M. Costa-Neto, I. Castro-Gamboa, V. da Silva-Bolzani and A. R. Araújo, *Phytochemistry*, 2006, 67, 1964.
- 148 M. Ishibashi, K. Toume and T. Koyano, Jpn. Kokai Tokkyo Koho JP, 2006, 256983, (Chem. Abstr., 2006, 145, 331793).
- 149 H. L. Chen, L. W. Wang, H. J. Su, B. L. Wei, S. Z. Yang and C. N. Lin, Org. Lett., 2006, 8, 753.
- 150 I. M. Prosser, R. J. Adams, M. H. Beale, N. D. Hawkins, A. L. Phillips, J. A. Pickett and L. M. Field, *Phytochemistry*, 2006, 67, 1564.
- 151 A. A. Ahmed and A. A. Mahmoud, Tetrahedron, 1998, 54, 8141
- 152 Y. H. Kuo, C. F. Chyu and H. C. Lin, *Chem. Pharm. Bull.*, 2003, **51**, 986.
- 153 L. Fang, F. Bi, C. Zhang, G. Zheng and Y. Li, Synlett, 2006, 2655.
- 154 W. S. Chang, K. S. Shia, H. J. Liu and T. W. Ly, Org. Biomol. Chem., 2006, 4, 3751.
- 155 G. A. Kraus and I. Jeon, Org. Lett., 2006, 8, 5315.
- 156 P. S. Poon and A. K. Banerjee, Nat. Prod. Res., 2006, 20, 629.
- 157 D. Ruiz-Leon, P. Rivera, I. Brito, M. Rodriguez and V. Manriquez, Z. Kristallogr., New Cryst. Struct., 2005, 220, 355.
- 158 R. D. Stipanovic, L. S. Puckhaber, J. H. Reibenspies and H. J. Williams, *Phytochemistry*, 2006, 67, 1304.
- 159 M. K. Dowd and S. M. Pelitire, J. Agric. Food Chem., 2006, 54, 3265.
- 160 R. D. Stipanovic, L. S. Puckhaber and A. A. Bell, J. Agric. Food Chem., 2006, 54, 1633.
- 161 R. D. Stipanovic, J. D. Lopez, Jr., M. K. Dowd, L. S. Puckhaber and S. E. Duke, *J. Chem. Ecol.*, 2006, **32**, 959.
- 162 C. R. Benedict, J. Liu and R. D. Stipanovic, *Phytochemistry*, 2006, 67, 356.
- 163 V. E. Paula, M. R. Rocha, L. C. de A. Barbosa and O. W. Howarth, J. Braz. Chem. Soc., 2006, 17, 1443.
- 164 P. J. Weathers, S. Elkholy and K. K. Wobbe, In Vitro Cell. Dev. Biol.: Plant, 2006, 42, 309.
- 165 S. Picaud, P. Merck, X. He, O. Sterner, M. Brodelius, D. E. Cane and P. E. Brodelius, Arch. Biochem. Biophys., 2006, 448, 150.
- 166 S. H. Kim, K. Heo, Y. J. Chang, S. H. Park, S. K. Rhee and S. U. Kim, J. Nat. Prod., 2006, 69, 758.
- 167 K. H. Teoh, D. R. Polichuk, R. Devin, D. W. Reed, G. Nowak and P. S. Covello, *FEBS Lett.*, 2006, **580**, 1411.
- 168 J. L. Han, B. E. Liu, H. C. Ye, H. Wang, Z. Q. Li and G. F. Li, J. Integr. Plant Biol., 2006, 48, 482.
- 169 A. Gutterres-Taranto, J. W. de Mesquita-Carneiro and M. Teixeira de Araujo, *Bioorg. Med. Chem.*, 2006, 14, 1546.
- 170 D. K. Ro, E. M. Paradise, M. Ouellet, K. J. Fisher, K. L. Newman, J. M. Ndungu, K. A. Ho, R. A. Eachus, T. S. Ham, J. Kirby, M. C. Y. Chang, S. T. Whiters, Y. Shiba, R. Sarpong and J. D. Keasling, *Nature*, 2006, **440**, 940.
- 171 J. H. Liu, Y. G. Chen, B. Y. Yu and Y. J. Chen, *Bioorg. Med. Chem. Lett.*, 2006, 16, 1909.
- 172 A. A. Lapkin, P. K. Plucinski and M. Cutler, J. Nat. Prod., 2006, 69, 1653.

- 173 W. J. M. Lommen, E. Schenk, H. J. Bouwmeester and F. W. A. Verstappen, Planta Med., 2006, 72, 336.
- 174 C. Chollet, B. Crousse, M. Ourévitch and D. Bonnet-Delpon, J. Org. Chem., 2006, 71, 3082
- 175 S. Tonmunphean, V. Parasuk and S. Kokpol, Bioorg. Med. Chem., 2006. 14. 2082
- 176 R. K. Haynes, B. Fugmann, J. Stetter, K. Rieckmann, H. D. Heilmann, H. W. Chan, M. K. Cheung, W. L. Lam, H. N. Wong, S. L. Croft, L. Vivas, L. Rattray, L. Stewart, W. Peters, B. L. Robinson, M. D. Edstein, B. Kotecka, D. E. Kyle, B. Beckermann, M. Gerisch, M. Radtke, G. Schmuck, W. Steinke, U. Wollborn, K. Schmeer and A. Römer, Angew. Chem., Int. Ed., 2006, 45, 2082.
- 177 C. Singh, S. Chaudhary and S. K. Puri, J. Med. Chem., 2006, 49, 7227.
- 178 M. Jung, J. Tak, W. Y. Chung and K. K. Park, Bioorg. Med. Chem. Lett., 2006, 16, 1227.
- 179 M. R. Romero, M. A. Serrano, M. Vallejo, T. Efferth, M. Alvarez and J. J. G. Marin, Planta Med., 2006, 72, 1169.
- 180 M. E. Maggi, A. Mangeaud, M. C. Carpinella, C. G. Ferrayoli, G. R. Valladares and S. M. Palacios, J. Chem. Ecol., 2005, 31, 1527.
- 181 C. Fehr and J. Galindo, Angew. Chem., Int. Ed., 2006, 45, 2901.
- 182 X. Gao, T. Shen, W. D. Xie and Z. J. Jia, Chin. Chem. Lett., 2006, 17, 341.
- 183 X. Gao, C. J. Lin, W. D. Xie, T. Shen and Z. J. Jia, Helv. Chim. Acta, 2006, 89, 1387.
- 184 F. Song, X. Xu, S. Li, S. Wang, J. Zhao, Y. Yang, X. Fan, J. Shi and L. He, J. Nat. Prod., 2006, 69, 1261.
- 185 F. C. Chaves, L. C. A. Barbosa, A. J. Demuner and A. A. Silva, Z. Naturforsch., B: Chem. Sci., 2006, 61, 1287.
- 186 B. W. Zilkowski, R. J. Bartelt, A. A. Cossé and R. J. Petroski, J. Chem. Ecol., 2006, 32, 2543.
- 187 T. L. Ho and R. J. Chein, Helv. Chim. Acta, 2006, 89, 231.
- 188 G. Subramaniam, S. Karimi and D. Phillips, Magn. Reson. Chem., 2006, 44, 1118
- 189 C. M. Cerda-García-Rojas, D. Guerra-Ramírez, L. U. Román-Marín, J. D. Hernández-Hernández and P. Joseph-Nathan, J. Mol. Struct. (THEOCHEM), 2006, 789, 37.
- 190 M. I. Choudhary, W. Kausar, Z. A. Siddiqui and Attar-ur-Rahman, Z. Naturforsch., B: Chem. Sci., 2006, 61, 1035
- 191 S. P. Chen, C. H. Chao, H. C. Huang, Y. C. Wu, C. K. Lu, C. F. Dai and J. H. Sheu, Bull. Chem. Soc. Jpn., 2006, 79, 1547.
- 192 A. E. Brown, E. W. Riddick, J. R. Aldrich and W. E. Holmes, J. Chem. Ecol., 2006, 32, 2489.
- 193 H. K. Wabo, P. Tane and J. D. Connolly, Biochem. Syst. Ecol., 2006, 34 603
- 194 D. T. Sadyrbekov, G. A. Atazhanova, A. T. Kulyyasov, V. A. Raldugin, Yu. V. Gatilov, M. M. Shakirov, T. T. Edil'baeva, K. M. Turdybekov and S. M. Adekenov, Chem. Nat. Compd. (Engl. Transl.), 2006, 42, 41.
- 195 B. Sabubal, M. Dan, A. John, R. Kurup, N. S. Pradeep, R. K. Valsamma and V. George, Phytochemistry, 2006, 67, 2469.
- 196 M. I. Choudhary, Z. A. Siddiqui, S. Khan, S. G. Musharraff and Atta-ur-Rahman, Z. Naturforsch., B: Chem. Sci., 2006, 61, 197.
- 197 M. I. Choudhary, Z. A. Siddiqui, S. A. Newaz and Atta-ur-Rahman, J. Nat. Prod., 2006, 69, 1429.
- 198 A. F. Barrero, M. M. Herrador, J. F. Quílez del Moral, P. Arteaga, E. M. Sánchez, J. F. Arteaga and M. Piedra, Eur. J. Org. Chem., 2006, 3434.
- 199 D. V. Domrachev and A. V. Tkachev, Chem. Nat. Compd. (Engl. Transl.), 2006, 42, 304.
- 200 S. T. Deyrup, D. C. Swenson, J. B. Gloer and D. T. Wicklow, J. Nat. Prod., 2006, 69, 608.
- 201 P. Joseph-Nathan, B. Reyes-Trejo and M. S. Morales-Ríos, J. Org. Chem., 2006, 71, 4411.
- 202 S. H. Shim, D. C. Swenson, J. B. Gloer, P. F. Dowd and D. T. Wicklow, Org. Lett., 2006, 8, 1225.
- 203 S. H. Shim, J. B. Gloer and D. T. Wicklow, J. Nat. Prod., 2006, 69, 1601.
- 204 M. Daoubi, R. Durán-Patrón, R. Hernández-Galán, A. Benharref, J. R. Hanson and I. G. Collado, Tetrahedron, 2006, 62, 8256.
- 205 P. J. Stephens, D. M. McCann, F. J. Devlin and A. B. Smith, III, J. Nat. Prod., 2006, 69, 1055.
- 206 S. Qi, C. Zhang and X. Li, Faming Zhuanli Shenqing Gongkai Shuomingshu CN 1.837.206 (Chem. Abstr., 2006, 145, 486183).
- 207 D. Q. Luo, Y. Gao, J. M. Gao, F. Wang, X. L. Yang and J. K. Liu, J. Nat. Prod., 2006, 69, 1354.

- 208 A. Mosaddik and P. G. Waterman, Nat. Prod. Commun., 2006, 1, 601
- 209 T. Kitayama, A. Furuya, C. Moriyama, T. Masuda, S. Fushimi, Y. Yonekura, H. Kubo, Y. Kawai and S. Sawada, Tetrahedron: Asymmetry, 2006, 17, 2311.
- 210 N. H. Yoo, J. P. Kim, B. S. Yun, I. J. Ryoo, I. K. Lee, E. S. Yoon, H. Koshino and I. C. Yoo, J. Antibiot., 2006, 59, 110.
- 211 K. A. B. Austin, M. G. Banwell, G. J. Harfoot and A. C. Willis, Tetrahedron Lett., 2006, 47, 7381.
- 212 M. Kashiwabara, T. Kamo, H. Makabe, H. Shibata and M. Hirota, Biosci., Biotechnol., Biochem., 2006, 70, 1502.
- 213 T. Kamo, M. Matsue, M. Kahiwabara and M. Hirota, Biosci., Biotechnol., Biochem., 2006, 70, 2307.
- 214 E. Krawczyk, M. Kniotek, M. Nowaczyk, T. Dzieciatkowski, M. Przybylski, A. Majewska and M. Luczak, Planta Med., 2006, 72, 61.
- 215 D. Weber, G. Erosa, O. Sterner and T. Anke, Z. Naturforsch., C: Biosci., 2006, 61, 663.
- 216 K. Yoshikawa, A. Kaneko, Y. Matsumoto, H. Hama and S. Arihara, J. Nat. Prod., 2006, 69, 1267.
- 217 S. Suzuki, T. Murayama and Y. Shiono, Z. Naturforsch., B: Chem. Sci., 2006, 61, 1295.
- 218 A. L. Lawrence, H. A. Wegner, M. F. Jacobsen, R. M. Adlington and J. E. Baldwin, Tetrahedron Lett., 2006, 47, 8717.
- 219 L. M. Finkielsztein, A. M. Bruno, S. G. Renou and G. Y. Moltrasio-Iglesias, Bioorg. Med. Chem., 2006, 14, 1863.
- 220 W. Itano, T. Ohshima and M. Shibasaki, Synlett, 2006, 3053.
- 221 G. Mehta and K. Pallavi, Tetrahedron Lett., 2006, 47, 8355.
- 222 X. N. Wang, J. H. Shen, J. C. Du and J. K. Liu, J. Antibiot., 2006, 59, 669.
- 223 C. N. Tetzlaff, Z. You, D. E. Cane, S. Takamatsu, S. Omura and H. Ikeda, Biochemistry, 2006, 45, 6179.
- 224 Z. You, S. Omura, H. Ikeda and D. E. Cane, J. Am. Chem. Soc., 2006, 128. 6566.
- 225 R. Quaderer, S. Omura, H. Ikeda and D. E. Cane, J. Am. Chem. Soc., 2006, 128, 13036.
- 226 P. Gupta and D. J. Tantillo, J. Am. Chem. Soc., 2006, 128, 6172.
- 227 S. A. Testorao and R. A. Spanevello, Org. Lett., 2006, 8, 3793.
- 228 B. L. Ashfeld and S. F. Martin, Tetrahedron, 2006, 62, 10497.
- 229 A. C. Coronel, C. M. Cerda-García-Rojas, P. Joseph-Nathan and C. A. N. Catalan, Flavour Fragrance J., 2006, 21, 839.
- 230 J. J. Rubal, F. M. Guerra, F. J. Moreno-Dorado, Z. D. Jorge, G. M. Massanet, H. Sohoel, U. W. Smitt, K. Frydenvang, S. B. Christensen, C. Nielsen and M. Eriksson, J. Nat. Prod., 2006, 69, 1566.
- 231 B. Rivero, I. Rivero-Cruz, J. M. Rodríguez, C. M. Cerda-Garcia-Rojas and R. Mata, J. Nat. Prod., 2006, 69, 1172.
- 232 B. Szafranek, K. Chraprowska, D. Waligora, R. Palavinska, A. Banach and J. Szafranek, J. Agric. Food Chem., 2006, 54, 7729.
- 233 M. Gutiérrez, T. L. Capson, H. M. Guzmán, J. González, E. Ortega-Barria, E. Quiñoa and R. Riguera, J. Nat. Prod., 2006, 69, 1379.
- 234 C. M. Bertea, A. Voster, F. W. A. Verstappen, M. Maffei, J. Beekwilder and H. J. Bouwmeester, Arch. Biochem. Biophys., 2006, 448, 3.
- 235 S. Picaud, M. E. Olsson, M. Brodelius and P. E. Brodelius, Arch. Biochem. Biophys., 2006, 452, 17.
- 236 R. Matovic, A. Ivkovic, M. Manojlovic, Z. Tokiv-Vujosevic and R. N. Saicic, J. Org. Chem., 2006, 71, 9411.
- 237 M. N. Todorova, B. Mikhova, A. Trendafilova, A. Vitkova, H. Duddeck and M. Anchev, Biochem. Syst. Ecol., 2006, 34, 136.
- 238 A. Trendafilova, M. Todorova, B. Mikhova, A. Vitkova and H. Duddeck, Phytochemistry, 2006, 67, 764.
- 239 R. Stefani, K. Schorr, J. M. Tureta, W. Vichnewski, I. Merfort and F. B. Da Costa, Z. Naturforsch., C: Biosci., 2006, 61, 647.
- 240 S. P. Yang, J. G. Cheng, J. Huo, H. L. Jiang, K. X. Chen and J. M. Yue, Chin. J. Chem., 2005, 23, 1530.
- 241 Y. C. Shen, K. L. Lo, Y. H. Kuo and A. T. Khalil, Nat. Prod. Commun., 2006, 1, 531.
- 242 Q. J. Zhang, X. S. Yang, H. Y. Zhu, Y. Wang, X. J. Hao and B. A. Song, Chin. Chem. Lett., 2006, 17, 355.
- 243 F. A. Macías, A. Fernández, R. M. Varela, J. M. G. Molinillo, A. Torres and P. L. C. A. Alves, *J. Nat. Prod.*, 2006, **69**, 795. 244 H. Bai, C. S. Lai, K. He, Z. Zhou, L. Zhang, Z. Quan, N. Zhu, Q. Y.
- Zheng, M. H. Pan and C. T. Ho, J. Nat. Prod., 2006, 69, 531.
- 245 X. X. Wang, C. J. Lin and Z. J. Jia, Planta Med., 2006, 72, 764.
- 246 Y. Li, D. M. Zhang, J. B. Li, S. S. Yu, Y. Li and Y. M. Luo, J. Nat. Prod., 2006, 69, 616.

- 247 A. Chea, S. Hout, C. Long, L. Marcourt, R. Faure, N. Azar and R. Elias, *Chem. Pharm. Bull.*, 2006, **54**, 1437.
- 248 O. Kos, V. Castro, R. Murillo, L. Poveda and I. Merfort, *Phytochemistry*, 2006, 67, 62.
- 249 J. Huo, S. P. Yang, J. Ding and J. M. Yue, J. Integr. Plant Biol., 2006, 48, 473.
- 250 L. L. Wang and L. H. Hu, J. Integr. Plant Biol., 2006, 48, 991.
- 251 N. Y. Yang, L. J. Tian, S. H. Qian, J. A. Duan and P. Li, *Zhongguo Tianran Yaowu*, 2005, 3, 224, (*Chem. Abstr.*, 2006, 145, 79719).
- 252 N. Vongvanich, P. Kittakoop, P. Charoenchai, S. Intamas, K. Sriklung and Y. Thebtaranonth, *Planta Med.*, 2006, **72**, 1427.
- 253 R. Stefani and F. B. Da Costa, Biochem. Syst. Ecol., 2006, 34, 757.
- 254 M. L. Guzman, R. M. Rossi, L. Karnischky, X. Li, D. R. Peterson, D. S. Howard and C. T. Jordan, *Blood*, 2005, **105**, 4163.
- 255 A. J. Steele, D. T. Jones, K. Ganeshaguro, V. M. Duke, B. C. Yogashangary, J. M. North, M. W. Lowdell, P. D. Kottaridis, A. B. Mehta, A. G. Prentice, A. V. Hoffbrand and R. G. Wickremasinghe, *Leukemia*, 2006, **20**, 1073.
- 256 D. R. Hwang, Y. S. Wu, C. W. Chang, T. W. Lien, W. C. Chen, U. K. Tan, J. T. A. Hsu and H. P. Hsieh, *Bioorg. Med. Chem.*, 2006, 14, 83.
- 257 A. Bachelier, R. Mayer and C. D. Klein, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 5605.
- 258 G. Croce, M. Milanesio, D. Viterbo, M. Clericuzio, P. Ugliengo and G. Appendino, *Eur. J. Org. Chem.*, 2006, 3140.
- 259 S. K. Srivastava, A. Abraham, B. Bhat, M. Jaggi, A. T. Singh, V. K. Sanna, G. Singh, S. K. Agarwal, R. Mukherjee and A. C. Burman, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 4195.
- 260 B. P. Bashyal, S. P. McLaughlin and A. A. L. Gunatilaka, J. Nat. Prod., 2006, 69, 1820.
- 261 S. Wahidulla, M. B. Govenkar and S. K. Paknikar, *Helv. Chim. Acta*, 2006, **89**, 496.
- 262 Q. X. Wu, Y. P. Shi and Z. J. Zhia, Nat. Prod. Rep., 2006, 23, 699.
- 263 T. Hackl, W. A. König and H. Muhle, Phytochemistry, 2006, 67, 778.
- 264 H. S. Shy and C. L. Wu, J. Asian Nat. Prod. Res., 2006, 8, 723.
- 265 R. C. Chang and C. L. Wu, J. Chin. Chem. Soc., 1999, 46, 191.
- 266 Y. F. Shu, H. C. Wei and C. L. Wu, Phytochemistry, 1994, 37, 773.
- 267 S. J. Wu, S. Fotso, F. Li, S. Qin, G. Kelter, H. H. Fiebig and H. Laatsch, J. Antibiot., 2006, 59, 331.
- 268 S. O. Lee, S. Z. Choi, S. U. Choi, G. H. Kim, Y. C. Kim and K. R. Lee, *Arch. Pharm. Res.*, 2006, **29**, 845.
- 269 A. García and G. Delgado, J. Nat. Prod., 2006, 69, 1618.
- 270 M. Okasaka, Y. Takaishi, Y. Kashiwada, O. K. Kodzhimatov, O. Ashurmentov, A. J. Lin, L. M. Consentino and K. H. Lee, *Phytochemistry*, 2006, 67, 2635.
- 271 Y. B. Liu, W. Jia, W. Y. Gao, A. H. Zhao, Y. W. Zhang, Y. Takaishi and H. Q. Duan, *J. Asian Nat. Prod. Res.*, 2006, 8, 303.
- 272 R. T. Li, A. H. Zhao, Y. H. Sheng, Z. Na and H. D. Sun, *J. Asian Nat. Prod. Res.*, 2005, **7**, 847.
- 273 X. Yang, M. Wong, N. Wang, A. S. C. Chan and X. Yao, *Chem. Pharm. Bull.*, 2006, 54, 676.
- 274 H. S. Ding, M. Y. Liu, W. L. Chang and H. C. Lin, *Chin. Pharm. J.* (*Beijing, China*), 2005, **57**, 37.
- 275 B. Wu, S. He, X. D. Wu and Y. J. Pan, Planta Med., 2006, 72, 1334.
- 276 Y. Q. Xu, Y. D. Lu and Y. L. Quan, *Acta Crystallogr., Sect. E: Struct. Rep. Online*, 2006, **62**, 01844.
- 277 T. T. Thuy, T. V. Sung and N. T. Hao, *Pharmazie*, 2006, 61, 570.
- 278 H. Hu, Y. Jian, X. Zheng, J. Liu and H. Cao, Indian J. Chem., Sect.
- B: Org. Chem. Incl. Med. Chem., 2006, 45, 1274.
 279 L. Harinantenaima, E. Mananjarasoa and K. Yamasaki, Z. Naturforsch., B: Chem. Sci., 2006, 61, 113.
- 280 M. Al-Dabbas, F. Hashinaga, S. Abdelgalei, T. Suganuma, K. Akiyama and H. Ayashi, J. Ethnopharmacol., 2005, 97, 237.
- 281 M. M. Al-Dabbas, K. Kitahara, T. Suganuma, F. Hashimoto and K. Tadera, *Biosci.*, *Biotechnol.*, *Biochem.*, 2006, 70, 2178.
- 282 D. E. Cane, X. He, S. Kobayashi, S. Omura and H. Ikeda, J. Antibiot., 2006, 59, 471.
- 283 J. Jiang, X. He and D. E. Cane, J. Am. Chem. Soc., 2006, 128, 8128.
- 284 H. M. C. Ferraz, A. J. C. Souza, B. S. M. Tenius and G. G. Bianco,
- Tetrahedron, 2006, 62, 9232.
 285 C. Zhang, G. J. Zheng, J. C. Chen, L. J. Fang and Y. L. Li, Chin. Chem. Lett., 2006, 17, 1290.
- 286 M. Morimoto, H. Fukumoto, M. Hiratani, W. Chavasiri and K. Komai, *Biosci., Biotechnol., Biochem.*, 2006, 70, 1864.
- 287 J. R. de Souza, G. D. F. Silva, T. Miyakoshi and C. L. Chen, J. Nat. Prod., 2006, 69, 1225.

- 288 W. Mingan, W. Wenjun, Z. Jingbo, J. Zhiqing and Z. Wenming, *Nat. Prod. Res.*, 2006, **20**, 653.
- 289 Y. Q. Guo, X. Li, J. J. Lee, J. Xu, N. Li, D. L. Meng and J. H. Wang, J. Asian Nat. Prod. Res., 2006, 8, 739.
- 290 Y. Lu, S. Yang, Z. Zou, X. Luo, H. Chen and L. Xu, *Heterocycles*, 2006, 68, 1241.
- 291 H. Wang and X. Tian, Chem. Pharm. Bull., 2006, 54, 219.
- 292 J. J. Chen, T. H. Chou, C. Y. Duh and I. S. Chen, J. Nat. Prod., 2006, 69, 685.
- 293 E. L. Whitson, S. M. V. Mala, C. A. Veltri, T. S. Bugni, E. D. de Silva and C. M. Ireland, *J. Nat. Prod.*, 2006, **69**, 1833.
- 294 F. R. Chang, I. H. Chen, S. C. Liao, H. H. Issa, K. I. Hayashi, H. Nozaki, Y. C. Wu and K. H. Lee, *Planta Med.*, 2006, **72**, 89.
- 295 M. Horiuch, C. Murakami, N. Fukamiya, D. Yu, T. H. Chen, K. F. Bastow, D. C. Zhang, Y. Takaishi, Y. Imakura and K. H. Lee, *J. Nat. Prod.*, 2006, **69**, 1271.
- 296 F. Muñoz-Martinez, C. P. Reyes, A. Perez-Lomas, I. A. Jimenez, F. Gamarro and S. Castanys, *Biochim. Biophys. Acta*, 2006, **1758**, 98.
- 297 M. S. Fan, G. Ye and C. G. Huang, Asian J. Chem., 2006, 18, 1540.
- 298 Z. X. Zhang, W. D. Xie, P. L. Li, Y. P. Shi and Z. J. Jia, *Helv. Chim. Acta*, 2006, **89**, 2927.
- 299 L. Yang, J. Dai, J. I. Sakai and M. Ando, J. Asian Nat. Prod. Res., 2006, 8, 317.
- 300 S. A. Ivasenko, T. T. Edil'baeva, A. T. Kulyyasov, G. A. Atazhanova, A. I. Drab, K. M. Turdybekov, V. A. Raldugin and S. M. Adekenov, *Chem. Nat. Compd. (Engl. Transl.)*, 2006, **42**, 36.
- 301 J. Merten, A. Hennig, P. Schwab, R. Fröhlich, S. V. Tokalov, H. O. Gutzeit and P. Metz, *Eur. J. Org. Chem.*, 2006, 1144.
- 302 S. G. Klochkov, S. V. Afanas'eva and A. N. Pushin, Chem. Nat. Compd. (Engl. Transl.), 2006, 42, 400.
- 303 A. A. Mahmoud, Nat. Prod. Commun., 2006, 1, 15.
- 304 A. A. Ahmed and J. Jakupovic, Phytochemistry, 1990, 29, 3658.
- 305 A. A. El-Bassuony and A. M. Kabbash, *Saudi Pharm. J.*, 2006, 14, 126.
- 306 K. H. C. Baser, B. Demirci, G. Iscan, T. Hashimoto, F. Demirci, Y. Noma and Y. Azakawa, *Chem. Pharm. Bull.*, 2006, 54, 222.
- 307 A. Klein and M. Miesch, Synthesis, 2006, 2613.
- 308 S. P. Yang and J. M. Yue, Tetrahedron Lett., 2006, 47, 1129.
- 309 J. Y. Ueda, L. Imamura, Y. Tezuka, Q. L. Tran, M. Tsuda and S. Kadota, *Bioorg. Med. Chem.*, 2006, 14, 3571.
- 310 A. Nakazaki, T. Era, Y. Numada and S. Kobayashi, *Tetrahedron*, 2006, 62, 6264.
- 311 A. Srikrishna and S. S. V. Ramasastry, *Tetrahedron Lett.*, 2005, 46, 7373.
- 312 A. Srikrishna and S. S. V. Ramasastry, *Tetrahedron Lett.*, 2006, 47, 335.
- 313 D. J. Schenk, C. M. Starks, K. R. Manna, J. Chappell, J. P. Noel and R. M. Coates, Arch. Biochem. Biophys., 2006, 448, 31.
- 314 C. J. Wegerski, R. N. Sonnenschein, F. Cabriales, F. A. Valeriote, T. Matainaho and P. Crews, *Tetrahedron*, 2006, 62, 10393.
- 315 X. H. Yan, X. Z. Zhu, J. L. Yu, D. Z. Jin, Y. W. Guo, E. Mollo and G. Cimino, J. Asian Nat. Prod. Res., 2006, 8, 579.
- 316 H. Oh, Bull. Korean Chem. Soc., 2005, 26, 1303, (Chem. Abstr., 2006, 144, 366476).
- 317 J. Q. Xu, Y. S. Li, Y. M. Li, S. H. Jiang, C. H. Tan and D. Y. Zhu, *Planta Med.*, 2006, **72**, 567.
- 318 J. X. Liu, X. N. Wei and Y. P. Shi, Planta Med., 2006, 72, 175.
- 319 Y. Li and Y. P. Shi, Helv. Chim. Acta, 2006, 89, 870.
- 320 M. Tori, K. Honda, H. Nakamizo, Y. Okamoto, M. Sakaoku, S. Takaoka, X. Gong, Y. Shen, C. Kuroda and R. Hanai, *Tetrahedron*, 2006, **62**, 4988.
- 321 A. L. Pérez-Castorena, A. Arciniegas, S. L. Guzmán, J. L. Villaseñor and A. Romo de Vivar, J. Nat. Prod., 2006, 69, 1471.
- 322 M. Yoshikawa, T. Morikawa, J. Tanaka and H. Shimoda, *Heterocycles*, 2006, 68, 2335.
- 323 R. Chizzola, T. Langer and C. Franz, Planta Med., 2006, 72, 1254.
- 324 L. P. Zhang, L. P. Ma, P. Yang and D. L. Cheng, *Gaodeng Xuexiao Huaxue Xuebao*, 2006, **27**, 1286, (*Chem. Abstr.*, 2007, **146**, 4172).
- 325 A. Arciniegas, A. L. Pérez-Castorena, J. L. Villaseñor and A. Romo de Vivar, J. Nat. Prod., 2006, 69, 1826.
- 326 H. Koshimo, H. Satoh, T. Yamada and Y. Esumi, *Tetrahedron Lett.*, 2006, **47**, 4623.
- 327 T. Yamada, M. Iritani, K. Minoura, K. Kawai and A. Numata, Org. Biomol. Chem., 2004, 2, 2131.

- 328 T. Yamada, K. Minomura, R. Tanaka and A. Numata, J. Antibiot., 2006, **59**, 345.
- 329 D. Q. Fei, Y. F. Han, G. Wu and K. Gao, J. Asian Nat. Prod. Res., 2006, 8, 99.
- 330 Y. S. Li, S. S. Li, Z. T. Wang, S. D. Luo and D. Y. Zhu, Nat. Prod. Res., 2006, 20, 1241.
- 331 Q. H. Wu, H. X. Wang and K. Gao, Chin. Chem. Lett., 2006, 17, 215.
- 332 Q. H. Wu, C. M. Liu, Y. J. Chen and K. Gao, *Helv. Chim. Acta*, 2006, **89**, 915.
- 333 J. X. Liu, X. N. Wei, Y. P. Shi and R. H. Lu, *Chin. Chem. Lett.*, 2005, 16, 1618.
- 334 J. Q. Liu, C. F. Zhang, Z. T. Wang and M. Zhang, *Zhongguo Tianran Yaowu*, 2005, 3, 340, (*Chem. Abstr.*, 2006, 145, 434709).
- 335 Q. X. Wu, Q. Y. Wei and Y. P. Shi, Pharmazie, 2006, 61, 241.
- 336 J. C. Ndom, J. T. Mbafor, A. G. B. Azebaze, J. C. Vardamides, Z. Kakam, A. F. W. Kamdem, A. Deville, T. M. Ngando and Z. T. Fomum, *Phytochemistry*, 2006, 67, 838.
- 337 M. Reina, A. González-Coloma, D. Domínguez-Díaz, R. Cabrera, C. Giménez-Mariño, M. L. Rodríguez and L. Villarroel, *Nat. Prod. Res.*, 2006, 20, 13.
- 338 P. E. O'Maille, J. Chappell and J. P. Noel, Arch. Biochem. Biophys., 2006, 448, 73.
- 339 S. Forcat and R. K. Allemann, Org. Biomol. Chem., 2006, 4, 2563.
- 340 Y. S. Li, Z. T. Wang, S. D. Luo, S. S. Li and D. Y. Zhu, Nat. Prod. Res., 2006, 20, 724.
- 341 J. Iqbal, A. Gupta and A. Husain, ARKIVOC, 2006, 107.
- 342 G. Blay, L. Cardona, A. M. Collado, B. García and J. R. Pedro, J. Org. Chem., 2006, 71, 4929.
- 343 L. H. Mace, M. S. Shanmugham, J. D. White and M. G. B. Drew, Org. Biomol. Chem., 2006, 4, 1020.
- 344 A. Arciniegas, A. L. Pérez-Castorena, G. Cuevas, F. del Río-Portilla and A. Romo de Vivar, *Magn. Reson. Chem.*, 2006, 44, 30.
- 345 M. G. Constantino, K. T. de Oliveira, E. C. Polo, G. V. J. da Silva and T. J. Brocksom, *J. Org. Chem.*, 2006, **71**, 9880.
- 346 H. M. C. Ferraz, T. O. Vieira and L. F. Silva, Jr., Synthesis, 2006, 2748.
- 347 X. L. Yang, D. Q. Luo and J. K. Liu, Z. Naturforsch., B: Chem. Sci., 2006, 61, 1180.
- 348 L. Z. Fang, H. J. Shao, W. Q. Yang and J. K. Liu, *Helv. Chim. Acta*, 2006, **89**, 1463.
- 349 M. Ramadan, S. Goeters, B. Watzer, E. Krause, K. Lohmann, R. Bauer, B. Hempel and P. Imming, *J. Nat. Prod.*, 2006, **69**, 1041.
- 350 E. V. Tikhonova, G. A. Atazhanova, V. A. Raldugin, I. Yu Bagryanskaya, Yu. V. Gatilov, M. M. Shakirov and S. M. Adekenov, *Chem. Nat. Compd. (Engl. Transl.)*, 2006, **42**, 298.
- 351 A. A. Ahmed, M. E. F. Hegazy, N. M. Hassan, M. Wodjcinska, J. Karchesy, P. W. Pare and T. Mabry, *Phytochemistry*, 2006, 67, 1547.
- 352 V. U. Ahmed, M. Zahid, M. S. Ali, A. K. Jassbi, M. Abbas, Z. Ali and M. Z. Iqbal, *Phytochemistry*, 1999, **52**, 1319.
- 353 S. K. Gedara, O. B. Abdel-Halim, S. H. El-Sharkawy, O. M. Shalama, T. W. Shier and A. F. Halim, Z. Naturforsch., C: Biosci., 2003, 58, 23.
- 354 A. E. H. Mohamed, A. A. Ahmed, E. Wollenweber, B. Bohm and Y. Asakawa, *Chem. Pharm. Bull.*, 2006, **54**, 152.
- 355 H. W. Park, S. U. Choi, N. I. Baek, S. H. Kim, J. S. Eun, J. H. Yang and D. K. Kim, Arch. Pharm. Res., 2006, 29, 131.
- 356 Y. G. Kwak, D. K. Kim, T. Z. Ma, S. A. Park, H. Park, Y. H. Jung, D. J. Yoo and J. S. Eun, *Arch. Pharm. Res.*, 2006, **29**, 834.
- 357 G. W. Zhang, X. Q. Ma, J. Y. Su, K. Zhang, H. Kurihara, X. S. Yao and L. M. Zheng, *Nat. Prod. Res.*, 2006, **20**, 659.
- 358 D. Craig and G. D. Henry, Eur. J. Org. Chem., 2006, 3558.
- 359 G. Blay, B. García, E. Molina and J. R. Pedro, J. Nat. Prod., 2006, 69, 1234.
- 360 G. Blay, B. García, E. Molina and J. R. Pedro, J. Org. Chem., 2006, 71, 7866.
- 361 M. Miyazawa and A. Sugawara, Nat. Prod. Res., 2006, 20, 731.
- 362 B. Föhlisch, R. Flogaus, G. H. Henle, S. Sendelbach and S. Henkel, *Eur. J. Org. Chem.*, 2006, 2160.
- 363 I. Djordjevic, M. Jadranin, V. Vajs, N. Menkovic, V. Tesevic, S. Macura and S. Milosavljevic, Z. Naturforsch., B: Chem. Sci., 2006, 61, 1437.
- 364 S. Rosselli, A. Maggio, G. Bellone and M. Bruno, *Tetrahedron Lett.*, 2006, 47, 7047.
- 365 W. Kisiel and K. Michalska, Fitoterapia, 2006, 77, 354.
- 366 J. C. Ndom, J. T. Mbafor, J. D. Wansi, A. W. Kamdem, L. M. Meva'a, J. C. Vardamides, F. Toukam, D. Pegyemb, T. M. Ngando, H. Laatsch and Z. T. Fomum, *Nat. Prod. Res.*, 2006, **20**, 435.

- 367 H. Z. Jin, D. Lee, J. H. Lee, K. Lee, Y. S. Hong, D. H. Choung, Y. H. Kim and J. J. Lee, *Planta Med.*, 2006, **72**, 40.
- 368 S. Zhang, M. Zhao, L. Bai, T. Hasegawa, J. Wang, L. Wang, H. Xue, Q. Deng, F. Xing, Y. Bai, J. I. Sakai, J. Bai, R. Koyanagi, Y. Tsukomo, T. Kataoka, K. Nagai, K. Hirose and M. Ando, *J. Nat. Prod.*, 2006, 69, 1425.
- 369 W. F. He, B. B. Xu, J. C. Pan, J. C. Lu, S. J. Song and S. X. Xu, J. Asian Nat. Prod. Res., 2006, 8, 481.
- 370 S. Dall'Acqua, G. Viola, M. Giorgetti, M. C. Loi and G. Innocenti, *Chem. Pharm. Bull.*, 2006, 54, 1187.
- 371 M. S. Ali, W. Ahmed, A. F. Armstrong, S. A. Ibrahim, S. Ahmed and M. Parvez, *Chem. Pharm. Bull.*, 2006, 54, 1235.
- 372 A. Rustaiyan and S. Sedaghat, Acta Hortic., 2005, 677, (Chem. Abstr., 2007, 146, 41908).
- 373 H. Liu, K. G. Jensen, L. M. Tran, M. Chen, L. Zhai, C. E. Olsen, H. Søhoel, S. R. Denmeade, J. T. Isaacs and S. B. Christensen, *Phytochemistry*, 2006, 67, 2651.
- 374 X. Chen, Z. J. Zhan and J. M. Yue, Nat. Prod. Res., 2006, 20, 125.
- 375 N. Yayli, C. Baltaci, Y. Gok, E. Aydin and O. Ucuncu, *Turk. J. Chem.*, 2006, **30**, 229, (*Chem. Abstr.*, 2006, **145**, 24182).
- 376 C. Çelik, S. Rosselli, A. M. Maggio, R. A. Raccuglia, I. Uysal, W. Kisiel, K. Michalska and M. Bruno, *Biochem. Syst. Ecol.*, 2006, 34, 349.
- 377 E. A. Krasnov, V. A. Raldugin, T. V. Kadyrova and I. P. Kaminskii, Chem. Nat. Compd. (Engl. Transl.), 2006, 42, 495.
- 378 C. Zidorn, R. Spitaler, E. P. Ellmerer and H. Stuppner, *Biochem. Syst. Ecol.*, 2006, **34**, 900.
- 379 E. M. Ahn, M. H. Bang, M. C. Song, M. H. Park, H. Y. Kim, B. M. Kwon and N. I. Baek, Arch. Pharm. Res., 2006, 29, 937.
- 380 J. Cis, G. Nowak and W. Kisiel, Biochem. Syst. Ecol., 2006, 34, 862.
- 381 W. Kisiel and K. Michalska, Biochem. Syst. Ecol., 2006, 34, 356.
- 382 Y. Zhu, M. Yang and Z. J. Jia, Acta Crystallogr., Sect. E: Struct. Rep. Online, 2006, 62, o510.
- 383 R. I. Dzhalmakhanbetova, S. B. Akhmetova, V. A. Raldugin, Yu. V. Gatilov, G. A. Atazhanova and S. M. Adekenov, *Chem. Nat. Compd. (Engl. Transl.)*, 2006, 42, 310.
- 384 F. A. Macías, V. M. I. Viñolo, F. R. Fronczek, G. M. Massanet and J. M. G. Molinillo, *Tetrahedron*, 2006, 62, 7747.
- 385 S. Bercion, T. Buffeteau, L. Lespade and M. A. Couppe de K. Martin, J. Mol. Struct. (THEOCHEM), 2006, 791, 186.
- 386 J. G. Foster, W. M. Clapham, D. P. Belesky, M. Labreveux, M. H. Hall and M. A. Sanderson, J. Agric. Food Chem., 2006, 54, 1772.
- 387 S. K. Sadhu, K. Hirata, X. Li, T. Otsuki, T. Koyano, S. Preeprame, T. Kowithayakorn and M. Ishibashi, J. Nat. Med., 2006, 60, 325.
- 388 X. C. Ma, L. J. Wu and D. A. Guo, J. Asian Nat. Prod. Res., 2006, 8, 713.
- 389 F. L. Manzano, F. M. Guerra, F. J. Moreno-Dorado, Z. D. Jorge and G. M. Massanet, Org. Lett., 2006, 8, 2879.
- 390 H. Søhoel, A. M. L. Jensen, J. V. Møller, P. Nissen, S. R. Denmeade, J. T. Isaacs, C. E. Olsen and S. B. Christensen, *Bioorg. Med. Chem.*, 2006, 14, 2810.
- 391 T. Morikawa, O. B. Abdel-Halim, H. Matsuda, S. Ando, O. Muraoka and M. Yoshikawa, *Tetrahedron*, 2006, 62, 6435.
- 392 B. Das, G. Mahender, Y. K. Rao, C. Ramesh, K. Venkateswarlu, K. Ravikumar, M. Geethangili and Y. M. Tzeng, *Helv. Chim. Acta*, 2006, 89, 285.
- 393 D. A. Kummer, J. B. Brenneman and S. F. Martin, *Tetrahedron*, 2006, 62, 11437.
- 394 F. Deguerry, L. Pastore, S. Wu, A. Clark, J. Chappell and M. Schalk, Arch. Biochem. Biophys., 2006, 454, 123.
- 395 M. Morimoto and K. Komai, Weed Biol. Manage., 2005, 5, 203, (Chem. Abstr., 2006, 145, 485925).
- 396 P. Kraft, C. Weymuth and C. Nussbaumer, *Eur. J. Org. Chem.*, 2006, 1403.
- 397 S. K. Wang, M. J. Huang and C. Y. Duh, J. Nat. Prod., 2006, 69, 1411.
- 398 T. Oonishi, A. Taniuchi, M. Mori and Y. Sato, *Tetrahedron Lett.*, 2006, 47, 5617.
- 399 Y. Guo, J. Xu, Y. Li, T. Yamakuni and Y. Ohizumi, *Planta Med.*, 2006, 72, 373.
- 400 Y. Guo, J. Xu, Y. Li, R. Watanabe, Y. Oshima, T. Yamakuni and Y. Ohizumi, *Chem. Pharm. Bull.*, 2006, **54**, 123.
- 401 M. Furusawa, T. Hashimoto, Y. Noma and Y. Asakawa, *Chem. Pharm. Bull.*, 2006, 54, 861.
- 402 S. C. Mao, Y. W. Guo and X. Shen, *Bioorg. Med. Chem. Lett.*, 2006, 16, 2947.

- 403 C. H. Chao, C. H. Hsieh, S. P. Chen, C. K. Lu, C. F. Dai and J. H. Sheu, *Tetrahedron Lett.*, 2006, **47**, 5889.
- 404 N. J. Jacobo-Herrera, N. Vartiainen, P. Bremner, S. Gibbons, J. Koistinaho and M. Heinrich, *Phytother. Res.*, 2006, **20**, 917.
- 405 L. Harinantenaina, R. Kurata, S. Takaoka and Y. Asakawa, *Phytochemistry*, 2006, **67**, 2616.
- 406 G. H. Li, L. Li, M. Duan and K. Q. Zhang, *Chem. Biodiversity*, 2006, 3, 210.
- 407 D. Z. Liu, F. Wang, T. G. Liao, J. G. Tang, W. Steglich, H. J. Zhu and J. K. Liu, Org. Lett., 2006, 8, 5749.
- 408 M. Rodríguez-López and F. A. Bermejo, *Tetrahedron*, 2006, **62**, 8095.
- 409 M. Akssira, F. Mellouki, A. Salhi, H. Alilou, A. Saouf, F. El-Hanbali, J. F. Arteaga and A. F. Barrero, *Tetrahedron Lett.*, 2006, 47, 6719.
- 410 A. Morita, H. Kiyota and S. Kuwahara, *Biosci., Biotechnol., Biochem.*, 2006, **70**, 2564.
- 411 I. Margaros, T. Montagnon, M. Tofi, E. Pavlakos and G. Vassilikogiannakis, *Tetrahedron*, 2006, 62, 5308.

- 412 P. A. Wender, M. P. Croatt and B. Witulski, *Tetrahedron*, 2006, 62, 7505.
- 413 H. C. Huang, Z. H. Wen, C. H. Chao, A. F. Ahmed, M. Y. Chiang, Y. H. Kuo, C. H. Hsu and J. H. Sheu, *Tetrahedron Lett.*, 2006, 47, 8751.
- 414 K. Ueda, T. Kadekaru, E. R. O. Siwu, M. Kita and D. Uemura, *J. Nat. Prod.*, 2006, **69**, 1077.
- 415 Z. G. Yu, K. S. Bi, Y. W. Guo, E. Mollo and G. Cimino, J. Asian Nat. Prod. Res., 2006, 8, 467.
- 416 Z. Y. Shao, J. Li, C. J. Sim, J. Y. Li, Z. Y. Li, F. J. Nan and Y. W. Guo, J. Asian Nat. Prod. Res., 2006, 8, 223.
- 417 A. Srikrishna, P. R. Kumar and S. J. Gharpure, *Indian J. Chem., Sect.* B: Org. Chem. Incl. Med. Chem., 2006, **45**, 1909.
- 418 A. Srikrishna and G. Satyanarayana, *Tetrahedron Lett.*, 2006, **47**, 367.
- 419 Y. Takahashi, M. Tsuda, J. Fromont and J. Kobayashi, *Heterocycles*, 2006, **67**, 791.
- 420 A. J. Humphrey and M. H. Beale, Phytochemistry, 2006, 67, 636.
- 421 A. J. Humphrey, A. M. Galster and M. H. Beale, *Nat. Prod. Rep.*, 2006, **23**, 592.